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Pregnancies through oocyte donation. A mini review of pathways involved in placental dysfunction

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Pregnancies resulting from assisted reproductive techniques (ART) are increasingly prevalent worldwide. While most pregnancies conceived through in-vitro fertilization (IVF) progress without complications, mounting evidence suggests that these pregnancies are at a heightened risk of adverse perinatal outcomes. Specifically, IVF pregnancies involving oocyte donation have garnered attention due to numerous reports indicating an elevated risk profile for pregnancy-related complications within this subgroup of patients. The precise mechanisms contributing to this increased risk of complications remain incompletely understood. Nonetheless, it is likely that they are mediated by an abnormal immune response at the fetal-maternal interface. Additionally, these outcomes may be influenced by baseline patient characteristics, such as the etiology of infertility, absence of corpus luteum, and variations in endometrial preparation protocols, among other factors. This review aims to succinctly summarize the most widely accepted mechanisms that potentially contribute to the onset of placental dysfunction in pregnancies conceived through oocyte donation.

KEYWORDS

in-vitro, infertility, preeclampsia, perinatal outcome, immune tolerance

Introduction

Pregnancies through assisted reproductive techniques (ART) are on the rise worldwide; current estimates report nearly 3.2 million cycles per year, with Asia, Europe, and North America as the major contributors (1). The increasing numbers can be partially explained by lower cost and easier access to ART facilities (2, 3), a progressive delay in maternal age at first pregnancy (4, 5), and policymaking and social acceptance of non-traditional families (6, 7).

Even though most pregnancies through *in-vitro* fertilization (IVF) evolve without pregnancy-related complications (8, 9), there is growing evidence that these pregnancies are at higher risk of adverse perinatal outcomes such as preterm birth, preeclampsia, fetal growth restriction and stillbirth (10–15). Recently, the Society of Maternal & Fetal Medicine (SMFM) released a series of recommendations highlighting the need for proper study and management (16).

The exact mechanisms that lead to the increased risk of pregnancy complications are not fully understood, they are probably mediated by baseline characteristics such as maternal age and comorbidities, intrinsic factors of infertility and the interventions carried out during the fertilization process. This is reflected in how perinatal risks vary according to the fertilization method used, the endometrial preparation protocols, the presence of corpus luteum, the selected transfer method (frozen vs. fresh embryo transfer), and the origin of the selected oocyte (12, 14, 17–22).

Donated oocytes, perinatal outcomes, and placental dysfunction

Among ART, IVF pregnancies through oocyte donation (OD) represent roughly 5 to 7% of all embryo transfers (4, 23), also with an increasing trend over time. In the last few years, these pregnancies have gained attention as several reports have demonstrated a higher risk profile of pregnancy-related complications (24–26).

When compared against IVF conceived with autologous oocytes, pregnancies from OD have shown lower placental volumes at the first trimester (27), a different uterine perfusion profile across gestation (28, 29), higher rates of villitis of unknown etiology (VUE) (30) and also an increased risk of preeclampsia (14, 26) and placental related disease in the third trimester (14, 17, 31–33).

More recently a retrospective study conducted by our group (34), compared antenatal indicators of placental dysfunction between donated and autologous oocytes in the third trimester, demonstrating an abnormal growth velocity from the second trimester to delivery among ART gestations, especially in those conceived with donated oocytes. These findings support mechanisms related to progressive placental dysfunction, rather than abnormal placentation.

Mechanisms involved in placental dysfunction in OD

Placental dysfunction can manifest in different ways, such as preterm birth, fetal growth restriction, preeclampsia and stillbirth, among others (35–38). In line with the above, several mechanisms have been involved in the onset of placental dysfunction and preeclampsia (39) among pregnancies conceived through OD. In the following sections, these pathways are addressed.

Baseline characteristics and infertility etiology

Among infertile couples, several baseline characteristics could be related to a higher risk of placental dysfunction and preeclampsia. Among them maternal age is still one of the main factors related with IVF success (40, 41). While this is true for IVF with autologous oocytes, some studies have shown that pregnancy outcomes (i.e., cumulative live rate) among gestations conceived through OD depend mainly on donor age (42, 43). However, the former seems to not apply when it comes to the risk of placental related disease (44). In fact, several factors may interact as mediators for placental dysfunction; first, there is consistent evidence that women with advanced maternal age have more comorbidities, a higher risk of preeclampsia and present more complicated patterns of multimorbidity during pregnancy (45-47). Second, endometrial receptivity has been proposed to be negatively affected by age, potentially influencing implantation, placental function and pregnancy outcome (48, 49), yet further studies are needed. Finally, infertility etiology could also influence pregnancy outcomes; A diminished ovarian reserve, which is a common indication of ART with OD, has been proposed as an indicator of a reduced vascular capacity and has been independently associated with a higher risk of preeclampsia and placental malperfusion lesions (50, 51). Also, premature ovarian failure, recurrent pregnancy loss and idiopathic infertility have been related with several underlying autoimmune diseases (i.e., systemic lupus erythematosus and antiphospholipid syndrome) (52-54), all conditions highly related with placental dysfunction, preeclampsia and adverse pregnancy outcome (55, 56). Other conditions such as endometriosis have been related with a reduced oocyte yield and a dysregulated decidualization leading to a reduced fertilization rate and a higher risk of preeclampsia (57-61). Also, polycystic ovarian syndrome has been related with an increased oxidative stress and chronic inflammation leading to a higher risk of VUE and hypertensive disorder of pregnancy (62-67). Moreover, altered pathways in lipids and glucose metabolism have been proposed to lead to altered placental structure, villous overcrowding, and finally abnormal placental function (68-70). Although by themselves they do not constitute a frequent indication for OD, they may coexist and act as contributing factors to placental dysfunction.

Embryo transfer method, endometrial preparation protocols, and role of corpus luteum

Several publications demonstrate a different risk profile according to the selected ART protocol (71, 72). Overall, most evidence supports that frozen embryo transfer (FET) presents (among others) a lower risk of small for gestational age and perinatal mortality, but a higherrisk of preeclampsia and placental disease when compared with fresh embryo transfer (14, 71, 73). Regardless, in the last few years the use of FET has presented a progressive increase (23), in part due to a reduced risk of ovarian hyperstimulation syndrome and the expansion of the "freeze all" strategy (which facilitates single embryo transfer and allows time for preimplantation genetic testing).

Although the above refers to studies carried out mainly in IVF with autologous oocytes, when it comes to pregnancies through OD, pooled data report that nearly 40% of them come from FET (23). The former is relevant as it has been argued that the increased risk of preeclampsia found in FET could be linked with the selected protocol for embryo transfer, rather than the cryopreservation and freezing-thaving process itself (74, 75).

Briefly, commonly used protocols for embryo transfer could be summarized in, natural cycles, stimulated cycles, and programmed cycles. In the latter, there is no ovulation associated, therefore no corpus luteum (CL). This becomes relevant as programmed cycles are employed in OD and there is consistent evidence that CL produces not only progesterone and estrogen, but also Relaxin and VEGF. The last two have been found to be implicated in maternal renal and circulatory pregnancy-adaptation and are not replaced during programmed cycles (20, 21, 76). Also, impaired endometrial receptivity has been linked with placental dysfunction among IVF (77). Therefore, it is plausible that the absence of these factors could contribute to an abnormal uterine environment, a suboptimal endometrium support with impaired decidualization, and an insufficient maternal-pregnancy adaptation (78). Thus, leading to the higher risk of placental dysfunction found in pregnancies through OD.

Developmental stage at embryo transfer (i.e., blastocyst-vs. cleavage-stage) has been proposed to influence perinatal outcomes (79, 80). To date, exploring the independent effect of developmental stage at the time of transfer and the impact of cryopreservation on the outcome of interest has been challenging. A recent network meta-analysis (81) demonstrates (with a very-low certainty of evidence) that frozenblastocyst transfer was associated with a reduction in the risk for LBW compared with both fresh-transfer modalities, and fresh-cleavage transfer may be associated with a reduction in the risk for perinatal death compared with frozen-blastocyst transfer. However, high-quality RCTs and individual participant data meta-analyses are still lacking.

Preimplantation genetic testing

Similar to the reported increase of pregnancies conceived through ART (1), the use of preimplantation genetic testing (PGT) has demonstrated a progressive increase over time (82, 83). In part due a higher risk of pregnancies with chromosomal abnormalities among patients with advanced maternal age and the possibility of testing for several inherited disorders among patients with recurrent pregnancy loss and recurrent implantation failure, among others (83). Most of PGT are conducted through trophectoderm biopsy, in which 5 to 10 trophectoderm cells are extracted as study samples (83-85). As placenta develops from the trophectoderm (86), there is some concern that the use of PGT could be related to defective placentation and the development of placental dysfunction (87, 88), thus increasing the risk of pregnancy complications such as hypertensive disorder of pregnancy and preeclampsia among others (89, 90). While initial meta-analyses showed that PGT pregnancies were associated with a higher risk of hypertensive disorder of pregnancy, their results were limited by a high sample heterogeneity (91, 92). A most recent systematic review and meta-analysis, restricted only to singletons from FET cycles, including 11.469 live births after PGT and 20.438 live births after IVF/ICSI (no-PGT), concludes that trophectoderm biopsy does not alter the risk of developing hypertensive disorders in subsequent pregnancies (84). Nonetheless, larger cohort studies and well-designed RCTs are still lacking.

Regardless of the above, the use of PGT could be considered at least as non-routine among pregnancies through OD. Since, it has been shown to report no benefit among fresh oocyte donation cycles recipients (93–95), and conflicting results have been reported for frozen oocyte donation cycles recipients (95, 96). Therefore, it seems reasonable not to consider PGT as a major contributing factor for placental dysfunction among OD pregnancies.

Immune tolerance breakdown

Normal placentation and pregnancy evolution requires the development of maternal immune tolerance to a semi-allogeneic fetus. To date, most accepted mechanisms involved in pregnancy immunomodulation and crosstalk between mother and fetus include; (i) a trophoblast with an overall poor antigenicity, mainly due to a lack of classic HLA-I and II antigens, with the exception of HLA-C, and the expression of nonclassical HLA molecules of class E and G (97, 98); (ii) a shift in the functional balance of T helper (Th) cells towards type-2 cells with a decline in cell-mediated Th1-type immunity (99); (iii) a change in the activity of uterine natural killer (uNK) cells from cytotoxic to regulatory, mainly producing chemokines, growth factors, cytokines and angiogenic factors, of relevance for the development of maternal–fetal interface (100); and (iv) a major proportion of macrophages with an anti-inflammatory, M2-like phenotype, involved in the dampening of immune reactions (98).

Several findings support the role of immunological dysfunction in the development of preeclampsia among spontaneous conception (39, 100). Pregnancy after OD is considered as a unique model to assess the immunologic pathways involved in placental dysfunction, as the fetus is an absolute allograft in contrast to semi-allograft fetus in natural conception.

In line with the above, it has been shown that among OD pregnancies, the degree of HLA mismatch between mother and fetus is correlated with a higher number of maternal decidual-activated $CD4^+$ Treg cells (101–104), a reduced number of tissue macrophages (105, 106), and the development of gestational hypertension and preeclampsia (107, 108). Furthermore, the risk of preeclampsia has been reported to be even higher among pregnancies conceived with double gamete donation (oocyte and sperm donation) (109), which could be attributed to an additive effect from the lack of paternal antigen-specific tolerance (97).

Also, genome-wide mRNA analysis in placentas from OD pregnancies have shown a reduced expression of thrombomodulin (110), several complement regulatory proteins (111), and altered immunoregulation by co-inhibitory pathways (112).

Moreover, several placental lesions are observed at different histologic levels in women with pregnancies conceived through OD, supporting an abnormal immune response. Of remark, (i) severe chronic deciduitis with dense fibrinoid deposition is a characteristic finding in OD pregnancy. Suggesting an important maternal alloimmune reaction resembling host versus graft disease at the human fetal–maternal interface (113). (ii) Also, a significantly increased prevalence of VUE is reported among pregnancies conceived through OD (30) which represent a manifestation of maternal anti-fetal rejection. (iii) Of remark, Schonkeren et al. (114) described a specific histologic lesion among uncomplicated OD pregnancies consistent on a diffuse inflammatory infiltrate involving the entire chorionic plate. In their study, preeclampsia occurs only in the group without the immunological lesion. Therefore, this lesion could reflect a protective immune mechanism towards the completely allogeneic fetus.

Other mechanisms

It is known that there are social determinants for placental insufficiency, being more prevalent among women from disfavoured socioeconomic status (115). The pathways operating these relationships are not fully understood, and epigenetic mechanisms may explain intergenerational transmission (116). A fraction of egg donations is non-altruistically motivated, making donors more likely to come from a more disadvantaged socioeconomic background, which could result in higher rates of perinatal complications in recipients.

Discussion

The development of ART and specifically the progress achieved in conceiving pregnancies through OD represent a significant opportunity for couples which under other conditions would not be able to achieve pregnancy. However, it should be acknowledged that there is consistent evidence of a higher risk profile among this subgroup of patients.

In this mini review, we intended to succinctly summarize the most widely accepted pathways linked with placental dysfunction. Overall, it could be stated that several non-exclusive physio pathological mechanisms are involved, rendering to these patients a cumulative higher risk of progressive placental dysfunction and preeclampsia.

It is our belief that the subgroup of OD pregnant patients requires further attention. First, already among infertile populations there are reports of higher morbidity & mortality (117, 118). Second, at the population level, there is a progressive and consistent trend of increasing numbers. Theoretically, this could lead to a worldwide higher frequency of preeclampsia. Third, at the individual level, the patient baseline characteristics plus the combination of the physio pathological mechanisms involved could potentially lead to more severe cases (119–121).

Regarding management of ART pregnancies, current recommendation from the Society of Maternal–Fetal Medicine (16) and the UK National Institute of Clinical Excellence (122), consider IVF as a moderate risk factor for preeclampsia and recommends low-dose aspirin and serial scanning only if an additional risk factor is found. However, these guidelines lump together all ART techniques as an overall category, without establishing differences between the mode of conception. Moreover, there are no clear recommendations regarding other surveillance tools, such as maternal and fetal Doppler assessment or angiogenic markers assessment, which arguably have shown moderate-to-good performance for the prediction of adverse perinatal outcomes among high-risk pregnancies (123, 124), and has been proposed as a tool to capture placental dysfunction secondary to pathophysiologic mechanisms other than early defective trophoblast invasion (125, 126).

There are still several research gaps and potential future developments in the field; for one side, there is a need for better characterization and a more complete risk-profile assessment of candidates for OD. In line with the above, identifying novel predictive factors to assess the risk for maternal serious complications may be of value (127). Also, evaluation for signs of immune tolerance breakdown, through the assessment of cellular subpopulations imbalance or its product (such as cytokines or chemokines) (128) and its correlation with known clinical signs of placental dysfunction (i.e., angiogenic markers or fetal & maternal Doppler), could also be explored. Moreover, HLA screening and matching could also be considered as a suitable tool attempting to decrease the reported immune tolerance disbalance (129).

Therapeutic interventions such as the use of some immunosuppressive agents have already shown some encouraging results enhancing outcomes among patients with recurrent pregnancy loss. Among them, hydroxychloroquine is a known anti-inflammatory and immune regulator drug commonly used in patients with autoantibodies disease. Its use during pregnancy has shown to improve the live birth rate in patients with persistent positive antiphospholipid antibodies and to reduce the risk of preeclampsia and fetal loss in mid and late pregnancy among patients with systemic lupus erythematosus (130–132). Also, when combined with prednisone, it has shown to improve outcomes of frozen embryo transfer in antinuclear antibody-positive patients undergoing IVF/ ICSI treatment (133). Moreover, its use has been reported as an effective therapeutic strategy in women with repeated implantation failure due cellular immune abnormalities, through a shift in Th2 responses (134). Therefore, hydroxychloroquine could be proposed as a potential treatment for immune tolerance imbalance among pregnancies through OD. However, there is still scarcity of highquality data that precludes further recommendations (135, 136). Finally, up to date and evidence based counselling about the related short and long-term risk should be offered to OD candidates, as in some cases the risk may be significant, and even overcome the benefits (137–141).

In conclusion, compelling evidence suggests the convergence of various additive factors associated with placental dysfunction in pregnancies conceived through oocyte donation. These factors encompass patient baseline characteristics, absence of corpus luteum, and dysfunction in pregnancy immune tolerance. Further research is imperative as this demographic constitutes a subgroup exhibiting the highest susceptibility to placental dysfunction, potentially necessitating a more vigilant follow-up – a practice not presently endorsed by existing guidelines.

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References

1. Adamson GD, Zegers-Hochschild F, Dyer S. Global fertility care with assisted reproductive technology. *Fertil Steril.* (2023) 120:473–82. doi: 10.1016/j. fertnstert.2023.01.013

2. Chiware TM, Vermeulen N, Blondeel K, Farquharson R, Kiarie J, Lundin K. IVF and other ART in low-and middle-income countries: a systematic landscape analysis. *Hum Reprod Update.* (2021) 27:213–28. doi: 10.1093/humupd/dmaa047

3. Imrie R, Ghosh S, Narvekar N, Vigneswaran K, Wang Y, Savvas M. Socioeconomic status and fertility treatment outcomes in high-income countries: a review of the current literature. *Hum Fertil.* (2023) 26:27–37. doi: 10.1080/14647273.2021.1957503

4. Adamson GD, Zegers-Hochschild F, Dyer S, Chambers G, de Mouzon J, Ishihara O, International Committee for Monitoring Assisted Reproductive Technology: World report on assisted reproductive technology, (2018). Available at:https://www.icmartivf.org/reports-publications/ (Accessed December 14, 2022)

5. OECD Family Database – OECD. Available at:https://www.oecd.org/els/family/ database.htm, (Accessed September 17, 2023)

6. Inhorn MC, Patrizio P. Infertility around the globe: new thinking on gender, reproductive technologies and global movements in the 21st century. *Hum Reprod Update*. (2015) 21:411–26. doi: 10.1093/humupd/dmv016

7. Fertility treatment: trends and figures/HFEA. (2019), Available at:https://www.hfea. gov.uk/about-us/publications/research-and-data/fertility-treatment-2019-trends-andfigures/#Section7 (Accessed October 17, 2023)

8. Cutting R. Single embryo transfer for all. *Best Pract Res Clin Obstet Gynaecol.* (2018) 53:30–7. doi: 10.1016/j.bpobgyn.2018.07.001

9. Shah JS, Vaughan DA, Leung A, Korkidakis A, Figueras F, Garcia D. Perinatal outcomes in singleton pregnancies after in vitro fertilization cycles over 24 years. *Fertil Steril.* (2021) 116:27–35. doi: 10.1016/j.fertnstert.2021.01.043

10. Qin J-B, Sheng X-Q, Wu D, Gao S-Y, You Y-P, Yang T-B. Worldwide prevalence of adverse pregnancy outcomes among singleton pregnancies after in vitro fertilization/ intracytoplasmic sperm injection: a systematic review and meta-analysis. *Arch Gynecol Obstet*, (2017) 295:285–301. doi: 10.1007/s00404-016-4250-3

11. Cavoretto P, Candiani M, Giorgione V, Inversetti A, Abu-Saba MM, Tiberio F. Risk of spontaneous preterm birth in singleton pregnancies conceived after IVF/ICSI treatment: meta-analysis of cohort studies. *Ultrasound Obstet Gynecol.* (2018) 51:43–53. doi: 10.1002/uog.18930

12. Bay B, Boie S, Kesmodel US. Risk of stillbirth in low-risk singleton term pregnancies following fertility treatment: a national cohort study. *BJOG*. (2019) 126:253-60. doi: 10.1111/1471-0528.15509

 Cavoretto PI, Giorgione V, Sotiriadis A, Viganò P, Papaleo E, Galdini A. IVF/ICSI treatment and the risk of iatrogenic preterm birth in singleton pregnancies: systematic review and meta-analysis of cohort studies. J Matern Fetal Neonatal Med. (2020) 35:1987–96. doi: 10.1080/14767058.2020.1771690

14. Chih HJ, Elias FTS, Gaudet L, Velez MP. Assisted reproductive technology and hypertensive disorders of pregnancy: systematic review and meta-analyses. *BMC Pregnancy Childbirth*. (2021) 21:449. doi: 10.1186/s12884-021-03938-8

15. Sarmon KG, Eliasen T, Knudsen UB, Bay B. Assisted reproductive technologies and the risk of stillbirth in singleton pregnancies: a systematic review and meta-analysis. *Fertil Steril.* (2021) 116:784–92. doi: 10.1016/j.fertnstert.2021.04.007

16. Society for Maternal-Fetal Medicine (SMFM)Ghidini A, Gandhi M, Mccoy J, Kuller JA. Publications Committee. Society for Maternal-Fetal Medicine Consult Series #60: management of pregnancies resulting from in vitro fertilization. *Am J Obstet Gynecol.* (2022) 226:B2–B12. doi: 10.1016/j.ajog.2021.11.001,

17. Johnson KM, Hacker MR, Resetkova N, O'Brien B, Modest AM. Risk of ischemic placental disease in fresh and frozen embryo transfer cycles. *Fertil Steril.* (2019) 111:714–21. doi: 10.1016/j.fertnstert.2018.11.043

18. Bosdou JK, Anagnostis P, Goulis DG, Lainas GT, Tarlatzis BC, Grimbizis GF. Risk of gestational diabetes mellitus in women achieving singleton pregnancy spontaneously or after ART: a systematic review and meta-analysis. *Hum Reprod Update*. (2020) 26:514–44. doi: 10.1093/humupd/dmaa011

19. Matsuzaki S, Nagase Y, Takiuchi T, Kakigano A, Mimura K, Lee M. Antenatal diagnosis of placenta accreta spectrum after in vitro fertilization-embryo transfer: a systematic review and meta-analysis. *Sci Rep.* (2021) 11:9205. doi: 10.1038/ s41598-021-88551-7

20. Singh B, Reschke L, Segars J, Baker VL. Frozen-thawed embryo transfer: the potential importance of the corpus luteum in preventing obstetrical complications. *Fertil Steril.* (2020) 113:252–7. doi: 10.1016/j.fertnstert.2019.12.007

21. Conrad KP, von Versen-Höynck F, Baker VL. Potential role of the corpus luteum in maternal cardiovascular adaptation to pregnancy and preeclampsia risk. *Am J Obstet Gynecol.* (2022) 226:683–99. doi: 10.1016/j.ajog.2021.08.018

22. Niu Y, Suo L, Zhao D, Wang Y, Miao R, Zou J. Is artificial endometrial preparation more associated with early-onset or late-onset preeclampsia after frozen embryo transfer? *J Assist Reprod Genet*. (2023) 40:1045–54. doi: 10.1007/s10815-023-02785-0

23. Chambers GM, Dyer S, Zegers-Hochschild F, de Mouzon J, Ishihara O. International Committee for Monitoring Assisted Reproductive Technologies world

report: assisted reproductive technology, 2014†. Hum Reprod. (2021) 36:2921-34. doi: 10.1093/humrep/deab198

24. Savasi VM, Mandia L, Laoreti A, Cetin I. Maternal and fetal outcomes in oocyte donation pregnancies. *Hum Reprod Update*. (2016) 22:620–33. doi: 10.1093/humupd/dmw012

25. Moreno Sepulveda J, Checa MA. Risk of adverse perinatal outcomes after oocyte donation: a systematic review and meta-analysis. J Assist Reprod Genet. (2019) 36:2017–37. doi: 10.1007/s10815-019-01552-4

26. Keukens A, Van Wely M, Van Der Meulen C, Mochtar MH. Pre-eclampsia in pregnancies resulting from oocyte donation, natural conception or IVF: a systematic review and meta-analysis. *Hum Reprod.* (2022) 37:586–99. doi: 10.1093/humrep/deab267

27. Rizzo G, Aiello E, Pietrolucci ME, Arduini D. Placental volume and uterine artery Doppler evaluation at 11 + 0 to 13 + 6 weeks' gestation in pregnancies conceived with in-vitro fertilization: comparison between autologous and donor oocyte recipients: placental volume in IVF pregnancies. *Ultrasound Obstet Gynecol.* (2016) 47:726–31. doi: 10.1002/uog.14918

28. Inversetti A, Mandia L, Candiani M, Cetin I, Larcher A, Savasi V. Uterine artery Doppler pulsatility index at 11–38 weeks in ICSI pregnancies with egg donation. *J Perinat Med.* (2018) 46:21–7. doi: 10.1515/jpm-2016-0180

29. Cavoretto PI, Farina A, Miglio R, Zamagni G, Girardelli S, Vanni VS. Prospective longitudinal cohort study of uterine arteries Doppler in singleton pregnancies obtained by IVF/ICSI with oocyte donation or natural conception. *Hum Reprod.* (2020) 35:2428–38. doi: 10.1093/humrep/deaa235

30. Dancey S, Mery E, Esteves A, Oltean I, Hayawi L, Tang K. Placenta pathology in recipient versus donor oocyte derivation for in vitro fertilization in a setting of hypertensive disorders of pregnancy and IUGR. *Placenta*. (2021) 108:114–21. doi: 10.1016/j.placenta.2021.03.012

31. Modest AM, Johnson KM, Karumanchi SA, Resetkova N, Young BC, Fox MP. Risk of ischemic placental disease is increased following in vitro fertilization with oocyte donation: a retrospective cohort study. *J Assist Reprod Genet.* (2019) 36:1917–26. doi: 10.1007/s10815-019-01545-3

32. Johnson KM, Hacker MR, Thornton K, Young BC, Modest AM. Association between in vitro fertilization and ischemic placental disease by gestational age. *Fertil Steril.* (2020) 114:579–86. doi: 10.1016/j.fertnstert.2020.04.029

33. Modest AM, Smith LH, Toth TL, Collier A-RY, Hacker MR. Multifoetal gestations mediate the effect of in vitro fertilisation (IVF) on ischaemic placental disease in autologous oocyte IVF more than donor oocyte IVF. *Paediatr Perinat Epidemiol.* (2022) 36:181–9. doi: 10.1111/ppe.12857

34. Caradeux J, Ávila F, Vargas F, Fernández B, Winkler C, Mondión M. Fetal growth velocity according to the mode of assisted conception. *Fetal Diagn Ther.* (2023) 50:299–308. doi: 10.1159/000531451

35. Messerlian C, Maclagan L, Basso O. Infertility and the risk of adverse pregnancy outcomes: a systematic review and meta-analysis. *Hum Reprod.* (2013) 28:125–37. doi: 10.1093/humrep/des347

36. Bastek JA, Brown AG, Anton L, Srinivas SK, D'addio A. Biomarkers of inflammation and placental dysfunction are associated with subsequent preterm birth. *J Matern Fetal Neonatal Med.* (2011) 24:600–5. doi: 10.3109/14767058.2010.511340

37. Burton GJ, Jauniaux E. Pathophysiology of placental-derived fetal growth restriction. Am J Obstet Gynecol. (2018) 218:S745-61. doi: 10.1016/j.ajog.2017.11.577

38. Morgan TK. Role of the placenta in preterm birth: a review. Am J Perinatol. (2016) 33:258–66. doi: 10.1055/s-0035-1570379

39. Jung E, Romero R, Yeo L, Gomez-Lopez N, Chaemsaithong P, Jaovisidha A. The etiology of preeclampsia. *Am J Obstet Gynecol.* (2022) 226:S844–66. doi: 10.1016/j. ajog.2021.11.1356

40. Fertility treatment: preliminary trends and figures | HFEA. (2021), Available at:https://www.hfea.gov.uk/about-us/publications/research-and-data/fertility-treatment-2021-preliminary-trends-and-figures/ (Accessed October 17, 2023)

41. Seshadri S, Morris G, Serhal P, Saab W. Assisted conception in women of advanced maternal age. *Best Pract Res Clin Obstet Gynaecol.* (2021) 70:10–20. doi: 10.1016/j. bpobgyn.2020.06.012

42. Wang YA, Farquhar C, Sullivan EA. Donor age is a major determinant of success of oocyte donation/recipient programme. *Hum Reprod.* (2012) 27:118–25. doi: 10.1093/humrep/der359

43. Hogan RG, Wang AY, Li Z, Hammarberg K, Johnson L, Mol BW. Oocyte donor age has a significant impact on oocyte recipients' cumulative live-birth rate: a population-based cohort study. *Fertil Steril.* (2019) 112:724–30. doi: 10.1016/j. fertnstert.2019.05.012

44. Soares SR, Troncoso C, Bosch E, Serra V, Simón C, Remohí J. Age and uterine receptiveness: predicting the outcome of oocyte donation cycles. J Clin Endocrinol Metab. (2005) 90:4399–404. doi: 10.1210/jc.2004-2252

45. Li J, Li Y, Duan Y, Xiao X, Luo J, Luo M. Dose-response associations of maternal age with pregnancy complications and multimorbidity among nulliparas and multiparas:

a multicentric retrospective cohort study in southern China. J Glob Health. (2023) 13:4117. doi: 10.7189/jogh.13.04117

46. Lean SC, Derricott H, Jones RL, Heazell AEP. Advanced maternal age and adverse pregnancy outcomes: a systematic review and meta-analysis. *PLoS One.* (2017) 12:e0186287. doi: 10.1371/journal.pone.0186287

47. Sheen J-J, Wright JD, Goffman D, Kern-Goldberger AR, Booker W, Siddiq Z. Maternal age and risk for adverse outcomes. *Am J Obstet Gynecol.* (2018) 219:390. e1–390.e15. doi: 10.1016/j.ajog.2018.08.034

48. Pathare ADS, Loid M, Saare M, Gidlöf SB, Zamani Esteki M, Acharya G. Endometrial receptivity in women of advanced age: an underrated factor in infertility. *Hum Reprod.* (2023) 29:773–93. doi: 10.1093/humupd/dmad019

49. Neykova K, Tosto V, Giardina I, Tsibizova V, Vakrilov G. Endometrial receptivity and pregnancy outcome. *J Matern Fetal Neonatal Med.* (2022) 35:2591–605. doi: 10.1080/14767058.2020.1787977

50. Ganer Herman H, Volodarsky-Perel A, Ton Nu TN, Machado-Gedeon A, Cui Y, Shaul J. Diminished ovarian reserve is a risk factor for preeclampsia and placental malperfusion lesions. *Fertil Steril.* (2023) 119:794–801. doi: 10.1016/j. fertnstert.2023.01.029

51. Woldringh GH, Frunt MHA, Kremer J. Decreased ovarian reserve relates to preeclampsia in IVF/ICSI pregnancies. *Hum Reprod.* (2006) 21:2948–54. doi: 10.1093/ humrep/del155

52. Grossmann B, Saur S, Rall K, Pecher A-C, Hübner S, Henes J. Prevalence of autoimmune disease in women with premature ovarian failure. *Eur J Contracept Reprod Health Care.* (2020) 25:72–5. doi: 10.1080/13625187.2019.1702638

53. Deroux A, Dumestre-Perard C, Dunand-Faure C, Bouillet L, Hoffmann P. Female infertility and serum auto-antibodies: a systematic review. *Clin Rev Allergy Immunol.* (2017) 53:78–86. doi: 10.1007/s12016-016-8586-z

54. Del Porto F, Ferrero S, Cifani N, Sesti G, Proietta M. Antiphospholipid antibodies and idiopathic infertility. *Lupus.* (2022) 31:347–53. doi: 10.1177/09612033221076735

55. Society for Maternal-Fetal Medicine (SMFM)Silver R, Craigo S, Porter F, Osmundson SS, Kuller JA, et al. Society for maternal-fetal medicine consult series #64: systemic lupus erythematosus in pregnancy. *Am J Obstet Gynecol.* (2023) 228:B41–60. doi: 10.1016/j.ajog.2022.09.001,

56. Sammaritano LR. Antiphospholipid syndrome. *Best Pract Res Clin Rheumatol.* (2020) 34:101463. doi: 10.1016/j.berh.2019.101463

57. Drummond K, Danesh NM, Arseneault S, Rodrigues J, Tulandi T, Raina J. Association between endometriosis and risk of preeclampsia in women who conceived spontaneously: a systematic review and meta-analysis. *J Minim Invasive Gynecol.* (2023) 30:91–9. doi: 10.1016/j.jmig.2022.11.008

58. Farland LV, Prescott J, Sasamoto N, Tobias DK, Gaskins AJ, Stuart JJ. Endometriosis and risk of adverse pregnancy outcomes. *Obstet Gynecol.* (2019) 134:527–36. doi: 10.1097/AOG.00000000003410

59. Vercellini P, Viganò P, Bandini V, Buggio L, Berlanda N, Somigliana E. Association of endometriosis and adenomyosis with pregnancy and infertility. *Fertil Steril.* (2023) 119:727–40. doi: 10.1016/j.fertnstert.2023.03.018

60. Rabaglino MB, Conrad KP. Evidence for shared molecular pathways of dysregulated decidualization in preeclampsia and endometrial disorders revealed by microarray data integration. *FASEB J.* (2019) 33:11682–95. doi: 10.1096/fj.201900662R

61. Conrad KP, Rabaglino MB, Post Uiterweer ED. Emerging role for dysregulated decidualization in the genesis of preeclampsia. *Placenta*. (2017) 60:119–29. doi: 10.1016/j.placenta.2017.06.005

62. Pan H, Xian P, Yang D, Zhang C, Tang H, He X. Polycystic ovary syndrome is an independent risk factor for hypertensive disorders of pregnancy: a systematic review, meta-analysis, and meta-regression. *Endocrine*. (2021) 74:518–29. doi: 10.1007/s12020-021-02886-9

63. Sha T, Wang X, Cheng W, Yan Y. A meta-analysis of pregnancy-related outcomes and complications in women with polycystic ovary syndrome undergoing IVF. *Reprod Biomed Online*. (2019) 39:281–93. doi: 10.1016/j.rbmo.2019.03.203

64. Roos N, Kieler H, Sahlin L, Ekman-Ordeberg G, Falconer H. Risk of adverse pregnancy outcomes in women with polycystic ovary syndrome: population based cohort study. *BMJ*. (2011) 343:d6309. doi: 10.1136/bmj.d6309

65. Hochberg A, Mills G, Volodarsky-Perel A, Nu TNT, Machado-Gedeon A, Cui Y. The impact of polycystic ovary syndrome on placental histopathology patterns in invitro fertilization singleton live births. *Placenta*. (2023) 139:12–8. doi: 10.1016/j. placenta.2023.05.015

66. Schoots MH, Bourgonje MF, Bourgonje AR, Prins JR, van Hoorn EGM, Abdulle AE. Oxidative stress biomarkers in fetal growth restriction with and without preeclampsia. *Placenta*. (2021) 115:87–96. doi: 10.1016/j.placenta.2021.09.013

67. Chiarello DI, Abad C, Rojas D, Toledo F, Vázquez CM, Mate A. Oxidative stress: Normal pregnancy versus preeclampsia. *Biochim Biophys Acta Mol basis Dis.* (2020) 1866:165354. doi: 10.1016/j.bbadis.2018.12.005

68. Riesche L, Bartolomei MS. Assisted reproductive technologies and the placenta: clinical, morphological, and molecular outcomes. *Semin Reprod Med.* (2018) 36:240–8. doi: 10.1055/s-0038-1676640

69. Chen M, Wu L, Zhao J, Wu F, Davies MJ, Wittert GA. Altered glucose metabolism in mouse and humans conceived by IVF. *Diabetes*. (2014) 63:3189–98. doi: 10.2337/db14-0103

70. Vambergue A, Fajardy I. Consequences of gestational and pregestational diabetes on placental function and birth weight. *World J Diabetes.* (2011) 2:196–203. doi: 10.4239/wjd.v2.i11.196

71. Roque M, Haahr T, Geber S, Esteves SC, Humaidan P. Fresh versus elective frozen embryo transfer in IVF/ICSI cycles: a systematic review and meta-analysis of reproductive outcomes. *Hum Reprod.* (2019) 25:2–14. doi: 10.1093/humupd/dmy033

72. Sha T, Yin X, Cheng W, Massey IY. Pregnancy-related complications and perinatal outcomes resulting from transfer of cryopreserved versus fresh embryos in vitro fertilization: a meta-analysis. *Fertil Steril.* (2018) 109:330–342.e9. doi: 10.1016/j. fertnstert.2017.10.019

73. Sacha CR, Harris AL, James K, Basnet K, Freret TS, Yeh J. Placental pathology in live births conceived with in vitro fertilization after fresh and frozen embryo transfer. *Am J Obstet Gynecol.* (2020) 222:360.e1–360.e16. doi: 10.1016/j.ajog.2019.09.047

74. Blazquez A, García D, Vassena R, Figueras F, Rodriguez A. Risk of pre-eclampsia after fresh or frozen embryo transfer in patients undergoing oocyte donation. *Eur J Obstet Gynecol Reprod Biol.* (2018) 227:27–31. doi: 10.1016/j.ejogrb.2018.05.030

75. Ginström Ernstad E, Wennerholm U-B, Khatibi A, Petzold M, Bergh C. Neonatal and maternal outcome after frozen embryo transfer: increased risks in programmed cycles. *Am J Obstet Gynecol.* (2019) 221:126.e1–126.e18. doi: 10.1016/j.ajog.2019.03.010

76. Guo X, Yi H, Li TC, Wang Y, Wang H, Chen X. Role of vascular endothelial growth factor (VEGF) in human embryo implantation: clinical implications. *Biomol Ther.* (2021) 11:253. doi: 10.3390/biom11020253

77. Cottrell HN, Deepak V, Spencer JB, Sidell N, Rajakumar A. Effects of Supraphysiologic levels of estradiol on endometrial decidualization, sFlt1, and HOXA10 expression. *Reprod Sci.* (2019) 26:1626–32. doi: 10.1177/1933719119833485

78. Conrad KP. Evidence for corpus luteal and endometrial origins of adverse pregnancy outcomes in women conceiving with or without assisted reproduction. *Obstet Gynecol Clin N Am.* (2020) 47:163–81. doi: 10.1016/j.ogc.2019.10.011

79. Marconi N, Allen CP, Bhattacharya S, Maheshwari A. Obstetric and perinatal outcomes of singleton pregnancies after blastocyst-stage embryo transfer compared with those after cleavage-stage embryo transfer: a systematic review and cumulative metaanalysis. *Hum Reprod.* (2022) 28:255–81. doi: 10.1093/humupd/dmab042

80. Glujovsky D, Quinteiro Retamar AM, Alvarez Sedo CR, Ciapponi A, Cornelisse S. Cleavage-stage versus blastocyst-stage embryo transfer in assisted reproductive technology. *Cochrane Database Syst Rev.* (2022) 2022:CD002118. doi: 10.1002/14651858. CD002118.pub6

81. Siristatidis C, Papapanou M, Karageorgiou V, Martins WP, Bellos I, Teixeira DM. Congenital anomaly and perinatal outcome following blastocyst-vs cleavage-stage embryo transfer: systematic review and network meta-analysis. *Ultrasound Obstet Gynecol.* (2023) 61:12–25. doi: 10.1002/uog.26019

82. Theobald R, SenGupta S, Harper J. The status of preimplantation genetic testing in the UK and USA. *Hum Reprod.* (2020) 35:986–98. doi: 10.1093/humrep/deaa034

83. Spinella F, Bronet F, Carvalho F, Coonen E, De Rycke M, Rubio C. ESHRE PGT consortium data collection XXI: PGT analyses in 2018. *Hum Reprod Open.* (2023) 2023:hoad010. doi: 10.1093/hropen/hoad010

84. Mao D, Xu J, Sun L. Impact of trophectoderm biopsy for preimplantation genetic testing on obstetric and neonatal outcomes: a meta-analysis. *Am J Obstet Gynecol.* (2023) S0002-9378:544–6. doi: 10.1016/j.ajog.2023.08.010

85. Kokkali G, Traeger-Synodinos J, Vrettou C, Stavrou D, Jones GM, Cram DS. Blastocyst biopsy versus cleavage stage biopsy and blastocyst transfer for preimplantation genetic diagnosis of beta-thalassaemia: a pilot study. *Hum Reprod.* (2007) 22:1443–9. doi: 10.1093/humrep/del506

86. Turco MY, Moffett A. Development of the human placenta. *Development.* (2019) 146:dev163428. doi: 10.1242/dev.163428

87. Yao Q, Chen L, Liang Y, Sui L, Guo L, Zhou J. Blastomere removal from cleavagestage mouse embryos alters placental function, which is associated with placental oxidative stress and inflammation. *Sci Rep.* (2016) 6:25023. doi: 10.1038/srep25023

88. Sugawara A, Sato B, Bal E, Collier AC, Ward MA. Blastomere removal from cleavage-stage mouse embryos alters steroid metabolism during pregnancy. *Biol Reprod.* (2012) 87:1–9. doi: 10.1095/biolreprod.111.097444

89. Zhang WY, von Versen-Höynck F, Kapphahn KI, Fleischmann RR, Zhao Q, Baker VL. Maternal and neonatal outcomes associated with trophectoderm biopsy. *Fertil Steril.* (2019) 112:283–290.e2. doi: 10.1016/j.fertnstert.2019.03.033

90. Zheng W, Yang SH, Yang C, Ren BN, Sun SM, Liu YL. Perinatal outcomes of singleton live births after preimplantation genetic testing during single frozen-thawed blastocyst transfer cycles: a propensity score-matched study. *Fertil Steril.* (2022) 117:562–70. doi: 10.1016/j.fertnstert.2021.12.020

91. Hou W, Shi G, Ma Y, Liu Y, Lu M, Fan X. Impact of preimplantation genetic testing on obstetric and neonatal outcomes: a systematic review and meta-analysis. *Fertil Steril.* (2021) 116:990–1000. doi: 10.1016/j.fertnstert.2021.06.040

92. Zheng W, Yang C, Yang S, Sun S, Mu M, Rao M. Obstetric and neonatal outcomes of pregnancies resulting from preimplantation genetic testing: a

systematic review and meta-analysis. *Hum Reprod Update*. (2021) 27:989–1012. doi: 10.1093/humupd/dmab027

93. Doyle N, Gainty M, Eubanks A, Doyle J, Hayes H, Tucker M. Donor oocyte recipients do not benefit from preimplantation genetic testing for aneuploidy to improve pregnancy outcomes. *Hum Reprod.* (2020) 35:2548–55. doi: 10.1093/humrep/deaa219

94. Martello CL, Kulmann MIR, Donatti LM, Bos-Mikich A, Frantz N. Preimplantation genetic testing for aneuploidies does not increase success rates in fresh oocyte donation cycles: a paired cohort study. *J Assist Reprod Genet*. (2021) 38:2909–14. doi: 10.1007/s10815-021-02339-2

95. Roeca C, Johnson R, Carlson N, Polotsky AJ. Preimplantation genetic testing and chances of a healthy live birth amongst recipients of fresh donor oocytes in the United States. J Assist Reprod Genet. (2020) 37:2283–92. doi: 10.1007/s10815-020-01874-8

96. Peyser A, Brownridge S, Rausch M, Noyes N. The evolving landscape of donor egg treatment: success, women's choice, and anonymity. *J Assist Reprod Genet.* (2021) 38:2327–32. doi: 10.1007/s10815-021-02262-6

97. Saito S, Nakabayashi Y, Nakashima A, Shima T, Yoshino O. A new era in reproductive medicine: consequences of third-party oocyte donation for maternal and fetal health. *Semin Immunopathol.* (2016) 38:687–97. doi: 10.1007/s00281-016-0577-x

98. Krop J, Tian X, van der Hoorn M-L, Eikmans M. The mac is back: the role of macrophages in human healthy and complicated pregnancies. *Int J Mol Sci.* (2023) 24:5300. doi: 10.3390/ijms24065300

99. Scherjon S, Lashley L, van der Hoorn M-L, Claas F. Fetus specific T cell modulation during fertilization, implantation and pregnancy. *Placenta*. (2011) 32:S291–7. doi: 10.1016/j.placenta.2011.03.014

100. Yagel S, Cohen SM, Goldman-Wohl D. An integrated model of preeclampsia: a multifaceted syndrome of the maternal cardiovascular-placental-fetal array. *Am J Obstet Gynecol.* (2022) 226:S963–72. doi: 10.1016/j.ajog.2020.10.023

101. Nakabayashi Y, Nakashima A, Yoshino O, Shima T, Shiozaki A, Adachi T. Impairment of the accumulation of decidual T cells, NK cells, and monocytes, and the poor vascular remodeling of spiral arteries, were observed in occyte donation cases, regardless of the presence or absence of preeclampsia. *J Reprod Immunol.* (2016) 114:65–74. doi: 10.1016/j.jri.2015.07.005

102. Tilburgs T, Scherjon SA, van der Mast BJ, Haasnoot GW, Voort-Maarschalk V-VD. Fetal-maternal HLA-C mismatch is associated with decidual T cell activation and induction of functional T regulatory cells. *J Reprod Immunol.* (2009) 82:148–57. doi: 10.1016/j.jri.2009.05.003

103. Chernyshov VP, Tumanova LE, Sudoma IA, Bannikov VI. Th1 and Th2 in human IVF pregnancy with allogenic fetus. *Am J Reprod Immunol.* (2008) 59:352–8. doi: 10.1111/j.1600-0897.2007.00578.x

104. Tian X, Goemaere NNT, van der Meeren L, Yang J, Kapsenberg JM, Lashley LEELO. Inflammatory placental lesions are specifically observed in healthy oocyte donation pregnancies with extreme fetal-maternal incompatibility. *Placenta*. (2023) 143:100–9. doi: 10.1016/j.placenta.2023.10.005

105. Bürk MR, Troeger C, Brinkhaus R, Holzgreve W, Hahn S. Severely reduced presence of tissue macrophages in the basal plate of pre-eclamptic placentae. *Placenta*. (2001) 22:309–16. doi: 10.1053/plac.2001.0624

106. van der Hoorn M-LP, van Egmond A, Swings GMJS, van Beelen E, van der Keur C, Tirado-González I. Differential immunoregulation in successful oocyte donation pregnancies compared with naturally conceived pregnancies. *J Reprod Immunol.* (2014) 101–102:96–103. doi: 10.1016/j.jri.2013.08.002

107. van Bentem K, Bos M, van der Keur C, Brand-Schaaf SH, Haasnoot GW, Roelen DL. The development of preeclampsia in oocyte donation pregnancies is related to the number of fetal-maternal HLA class II mismatches. *J Reprod Immunol.* (2020) 137:103074. doi: 10.1016/j.jri.2019.103074

108. Lashley L, Haasnoot GW, Spruyt-Gerritse M, Claas FHJ. Selective advantage of HLA matching in successful uncomplicated oocyte donation pregnancies. *J Reprod Immunol.* (2015) 112:29–33. doi: 10.1016/j.jri.2015.05.006

109. Blazquez A, García D, Vassena R, Figueras F, Rodriguez A. Risk of preeclampsia in pregnancies resulting from double gamete donation and from oocyte donation alone. *Pregnancy Hypertens*. (2018) 13:133–7. doi: 10.1016/j.preghy.2018.06.010

110. Bos M, Baelde HJ, Bruijn JA, Bloemenkamp KWM, van der Hoorn M-LP, Turner RJ. Loss of placental thrombomodulin in oocyte donation pregnancies. *Fertil Steril.* (2017) 107:119–129.e5. doi: 10.1016/j.fertnstert.2016.10.005

111. Lashley L, Buurma A, Swings GMJS, Eikmans M, Anholts JDH, Bakker JA. Preeclampsia in autologous and oocyte donation pregnancy: is there a different pathophysiology? *J Reprod Immunol.* (2015) 109:17–23. doi: 10.1016/j.jri.2015.03.004

112. van Hof LJ, Dijkstra KL, van der Keur C, Eikmans M, Baelde HJ, Bos M. Decreased expression of ligands of placental immune checkpoint inhibitors in uncomplicated and preeclamptic oocyte donation pregnancies. *J Reprod Immunol.* (2020) 142:103194. doi: 10.1016/j.jri.2020.103194

113. Gundogan F, Bianchi DW, Scherjon SA, Roberts DJ. Placental pathology in egg donor pregnancies. *Fertil Steril*. (2010) 93:397–404. doi: 10.1016/j.fertnstert.2008.12.144

114. Schonkeren D, Swings G, Roberts D, Claas F, de Heer E, Scherjon S. Pregnancy close to the edge: an immunosuppressive infiltrate in the chorionic plate

of placentas from uncomplicated egg cell donation. *PLoS One*. (2012) 7:e32347. doi: 10.1371/journal.pone.0032347

115. Hirst JE, Villar J, Victora CG, Papageorghiou AT, Finkton D, Barros FC. The antepartum stillbirth syndrome: risk factors and pregnancy conditions identified from the INTERGROWTH-21st project. *BJOG*. (2018) 125:1145–53. doi: 10.1111/1471-0528.14463

116. McDade TW, Ryan CP, Jones MJ, Hoke MK, Borja J, Miller GE. Genome-wide analysis of DNA methylation in relation to socioeconomic status during development and early adulthood. *Am J Phys Anthropol.* (2019) 169:3–11. doi: 10.1002/ajpa.23800

117. Murugappan G, Li S, Lathi RB, Baker VL, Luke B, Eisenberg ML. Increased risk of severe maternal morbidity among infertile women: analysis of US claims data. *Am J Obstet Gynecol.* (2020) 223:404.e1–404.e20. doi: 10.1016/j.ajog.2020.02.027

118. Murugappan G, Li S, Alvero RJ, Luke B, Eisenberg ML. Association between infertility and all-cause mortality: analysis of US claims data. *Am J Obstet Gynecol.* (2021) 225:57.e1–57.e11. doi: 10.1016/j.ajog.2021.02.010

119. Masturzo B, Di Martino D, Prefumo F, Cavoretto P, Germano C, Gennarelli G. Higher rate of early-onset preeclampsia in pregnancies following oocyte donation according to increasing maternal age. *Arch Gynecol Obstet.* (2019) 300:861–7. doi: 10.1007/s00404-019-05291-w

120. Ervaala A, Laivuori H, Gissler M, Kere J, Kivinen K, Pouta A. Characteristics of preeclampsia in donor cell gestations. *Pregnancy Hypertens*. (2022) 27:59–61. doi: 10.1016/j.preghy.2021.12.005

121. Dai F, Lan Y, Pan S, Wang Y, Hua Y, Xiao W. Pregnancy outcomes and disease phenotype of hypertensive disorders of pregnancy in singleton pregnancies after in vitro fertilization: a retrospective analysis of 1130 cases. *BMC Pregnancy Childbirth*. (2023) 23:523. doi: 10.1186/s12884-023-05838-5

122. RCOG. The investigation and management of the small-for-gestational-age fetus. Green-top Guideline 31 (2013), Available at:https://www.rcog.org.uk/globalassets/documents/guidelines/gtg_31.pdf

123. Martinez-Portilla RJ, Caradeux J, Meler E, Lip-Sosa DL, Sotiriadis A, Figueras F. Third-trimester uterine artery Doppler for prediction of adverse outcome in late small-for-gestational-age fetuses: systematic review and meta-analysis. *Ultrasound Obstet Gynecol.* (2020) 55:575–85. doi: 10.1002/uog.21940

124. Conde-Agudelo A, Villar J, Kennedy SH, Papageorghiou AT. Predictive accuracy of cerebroplacental ratio for adverse perinatal and neurodevelopmental outcomes in suspected fetal growth restriction: systematic review and meta-analysis. *Ultrasound Obstet Gynecol.* (2018) 52:430–41. doi: 10.1002/uog.19117

125. Llurba E, Turan O, Kasdaglis T, Harman CR, Baschat AA. Emergence of lateonset placental dysfunction: relationship to the change in uterine artery blood flow resistance between the first and third trimesters. *Am J Perinatol.* (2013) 30:505–12. doi: 10.1055/s-0032-1329181

126. Binder J, Monaghan C, Thilaganathan B, Carta S, Khalil A. De-novo abnormal Uteroplacental circulation in the third trimester: pregnancy outcome and pathological implications. *Ultrasound Obstetr Gynecol.* (2017) 52:60–5. doi: 10.1002/uog.17564

127. Pimentel C, Solene D, Frédérique J, Guillaume B, Jean L, Maëla LL. What are the predictive factors for preeclampsia in oocyte recipients? *J Hum Reprod Sci.* (2019) 12:327–33. doi: 10.4103/jhrs.JHRS_43_19

128. Behnam Sani K, Sawitzki B. Immune monitoring as prerequisite for transplantation tolerance trials. *Clin Exp Immunol.* (2017) 189:158–70. doi: 10.1111/ cei.12988

129. Mangum DS, Caywood E. A clinician's guide to HLA matching in allogeneic hematopoietic stem cell transplant. *Hum Immunol.* (2022) 83:687–94. doi: 10.1016/j. humimm.2022.03.002

130. Ye S, Liu Y, Zhao X, Ma Y, Wang Y. Hydroxychloroquine improves pregnancy outcomes of women with positive antinuclear antibody spectrum test results. *Front Med.* (2023) 10:1113127. doi: 10.3389/fmed.2023.1113127

131. Duan J, Ma D, Wen X, Guo Q, Gao J, Zhang G. Hydroxychloroquine prophylaxis for preeclampsia, hypertension and prematurity in pregnant patients with systemic lupus erythematosus: a meta-analysis. *Lupus*. (2021) 30:1163–74. doi: 10.1177/09612033211007199

132. Tian Y, Xu J, Chen D, Yang C, Peng B. The additional use of hydroxychloroquine can improve the live birth rate in pregnant women with persistent positive antiphospholipid antibodies: a systematic review and meta-analysis. *J Gynecol Obstet Hum Reprod.* (2021) 50:102121. doi: 10.1016/j.jogoh.2021.102121

133. Gao R, Deng W, Meng C, Cheng K, Zeng X, Qin L. Combined treatment of prednisone and hydroxychloroquine may improve outcomes of frozen embryo transfer in antinuclear antibody-positive patients undergoing IVF/ICSI treatment. *Lupus*. (2021) 30:2213–20. doi: 10.1177/09612033211055816

134. Ghasemnejad-Berenji H, Ghaffari Novin M, Hajshafiha M, Nazarian H, Hashemi SM, Ilkhanizadeh B. Immunomodulatory effects of hydroxychloroquine on Th1/Th2 balance in women with repeated implantation failure. *Biomed Pharmacother*. (2018) 107:1277–85. doi: 10.1016/j.biopha.2018.08.027

135. Andreescu M. The impact of the use of immunosuppressive treatment after an embryo transfer in increasing the rate of live birth. *Front Med.* (2023) 10:67876. doi: 10.3389/fmed.2023.1167876

136. Mirzaei M, Amirajam S, Moghimi ES, Behzadi S, Rohani A, Zerangian N. The effects of hydroxychloroquine on pregnancy outcomes in infertile women: a systematic review and meta-analysis. *J Med Life*. (2023) 16:189–94. doi: 10.25122/jml-2022-0095

137. Schutte JM, Schuitemaker NWE, Steegers EAP, van Roosmalen J. Maternal death after oocyte donation at high maternal age: case report. *Reprod Health*. (2008) 5:12. doi: 10.1186/1742-4755-5-12

138. Korb D, Schmitz T, Seco A, Le Ray C, Santulli P, Goffinet F. Increased risk of severe maternal morbidity in women with twin pregnancies resulting from oocyte donation. *Hum Reprod.* (2020) 35:1922–32. doi: 10.1093/humrep/deaa108

139. Garcia Castro J, Rodríguez-Pardo J, Díaz de Terán J. Eclampsia-induced posterior reversible encephalopathy syndrome in a donor oocyte recipient. *J Family Reprod Health.* (2020) 14:269–72. doi: 10.18502/jfrh.v14i4.5211

140. Sadeghi MR. Do we have the right to challenge the rules of nature using science and technology tools? J Reprod Infertil. (2019) 20:199–200.

141. Le Ray C, Scherier S, Anselem O, Marszalek A, Tsatsaris V, Cabrol D. Association between oocyte donation and maternal and perinatal outcomes in women aged 43 years or older. *Hum Reprod.* (2012) 27:896–901. doi: 10.1093/humrep/der469