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Rapid and precise identification of cervicothoracic necrotizing fasciitis caused by *Prevotella* and *Streptococcus constellatus* by using Nanopore sequencing technology: a case report

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Introduction: Cervicothoracic necrotizing fasciitis (CNF) is one form of necrotizing soft-tissue infections, which could lead to patient demise during short course. Therefore, early recognition and immediate treatment contribute to promising prognosis of patients.

Case presentation: A 58-year-old diabetic patient presented with a sore throat and progressive irritation of the neck and chest for 4 days. The initial diagnosis was considered to be soft-tissue infection and the clinician gave empirical antiinfectious medication for expectant treatment. During the course of disease, surgical incision was performed to relieve suffocation and shortness of breath. The drainage fluids were detected with microbiological culture and molecular sequencing. Nanopore sequencing technology (NST) helped to identify the coinfection of *Streptococcus constellatus* and *Prevotella* spp., which was not recognized during the original period of 15 days. The precise identification of pathogen supported to guide the pharmacologic treatment with meropenem and linezolid. Ultimately, combined with the surgical observation and postsurgical pathological examination, the patient was diagnosed as CNF, which could be much more acute and serious than normal soft-tissue infections. The patient has been successfully treated with prompt antimicrobial medication and appropriate surgical debridement.

Conclusion: This case presented a CNF patient with type 2 diabetes, successfully recovered after prompt microbial detection, precise anti-infectious treatment, and appropriate surgical intervention. It highlights the importance of recognizing pathogen by applying rapid microbiological detection, including NST, in acute and serious infectious disease.

KEYWORDS

cervicothoracic necrotizing fasciitis (CNF), necrotizing soft-tissue infections (NSTIs), nanopore sequencing technology (NST), coinfection, case report

1 Introduction

Necrotizing soft-tissue infections (NSTIs) is a deep soft-tissue infection causing progressive destruction of muscle fascia and overlying subcutaneous fat (1), with the incidence of 0.3 to 15 cases per 100,000 population, but a mortality of over 20% (2). Cervicothoracic necrotizing fasciitis (CNF) is one form of NSTIs with relatively rare infections of head and neck, compared with other body areas reported, and could lead to patient demise during short course (3). Thus, early recognition and immediate treatment are key to favorable outcomes of CNF patients.

The definitive diagnosis of necrotizing fasciitis relies on the surgical evaluation, and pharmacologic treatment is recommended being guided with microbiological identification (4), especially Gram's staining (1, 5). Traditional microbiological detection includes Gram staining, culture and antimicrobial sensitivity information (1), which could be time-consuming (6) and coverage-restricted (7). Nanopore sequencing technology (NST) is an emerging platform for pathogenic molecular detection, which is a promising method for fast, precised, and broad-range microbial identification (8, 9). Therefore, NST might contribute to the improvement of prognosis in patients with necrotizing fasciitis.

Herein, we presented a successfully treated CNF patient with type 2 diabetes. Guided with NST, the clinician identified the coinfection of pathogenic *Prevotella* and *Streptococcus constellatus*. With the combination of surgical debridement and prompt antimicrobial treatment, the patient achieved clinical cure and discharged after 46-day treatment. To our knowledge, cases of CNF identified with bacterial coinfection by NST have been rarely reported. It suggests that NST could provide rapid and precised clue of anti-infectious medication, and be benefit on CNF diagnosis and treatment.

2 Case presentation

A 58-year-old male patient presented to the clinic with a chief complaint of sore throat for 4 days. The pain was particularly noticeable while swallowing, accompanied with a hoarse voice. Despite receiving anti-inflammatory treatment, the sore throat was not ameliorated significantly. As a result, the patient was admitted to the Otorhinolaryngologic Department (Day 0). Upon admission, the patient's vital signs were as follows: ear temperature of 37.4°C, heart rate of 102 bpm/min, blood pressure of 98/59 mmHg, and respiratory rate of 21 times/min. The clinician gave empirical treatment with ceftizoxime and levofloxacin (Figure 1A).

At Day 4, the patient began experiencing progressive irritation in the neck and chest. The dermathemia and swelling of cervicothoracic skin was obvious, with a high skin temperature, obvious skin tenderness, and palpable skin fluctuation and crepitus. Following multidisciplinary consultation, the initial diagnosis was considered to be soft-tissue infection, and the medication were empirically changed as ceftriaxone and teicoplanin. With no significant improvement of the chief complaints, the patient was transferred to the Infectious Department for infectious control and further treatment (Day 6). Given the high level of inflammatory indicators and the unsatisfactory response to conventional antibiotics, the medication was switched to meropenem and linezolid. It was worth noting that the patient was diagnosed with type 2 diabetes mellitus (T2DM) during hospitalization. For the purpose of further diagnosis, the cervical and chest computed tomography (CT)-scan was performed at Day 9, which showed swelling and pneumatosis in the right parts of vocal cord, parapharyngeal space and cervicothoracic subcutaneous tissue (Figure 1B), with inflammatory changes. However, the disease suddenly progressed with the chief complaints being increased suffocation and shortness of breath in the afternoon of Day 9. To relieve discomfort, thoracentesis was performed, along with skin incision and effusion drainage. The drainage fluid was positively cultured with colonies of Streptococcus mitis, which was covered with the anti-infectious spectrum of present medication. Three days later (Day 12), the symptoms of swelling and pneumatosis exhibited significant amelioration, according to the CT-scan (Figure 1B, Day 12). For a comprehensive and accurate pathogen diagnosis, molecular sequencing with NST of drainage fluid was performed, which revealed positive detection for Prevotella intermedia, Prevotella enoeca and Streptococcus constellatus (Table 1). The NST result suggested polymicrobial infection, which was not recognized during the last 15-day treatment. Fortunately, the spectrum of anti-infectious drugs could cover the detected pathogen. The level of inflammation biomarkers, monitored in the duration of therapy, continued to decrease (Figure 1C), indicating the effectiveness of antiinfection treatment.

Despite the improvement of clinical indicators (Figure 1C) and imaging features, the patient still complained about the discomfort of cervicothoracic tension and pain. Thus, the diagnosis suspected to be CNF and thoracotomy debridement is recommended (1) and performed (Day 22). Extensive liquefaction of subcutaneous tissue and part of deep fascia was observed during the operation and the pathological examination was consistent with tissue abscess and necrosis (Figure 1D), which both confirmed the diagnosis of CNF. Laboratory Risk Indicator Score for Necrotizing Fasciitis (LRINEC) score is a tool to distinguish CNF from other soft tissue infections (10). We retrospectively calculated the LRINEC scores during the course of disease, which exhibited a gradual downward trend (Figure 1C). Moreover, after confirming the diagnosis of CNF, magnetic resonance imaging (MRI) was performed at Day 26. Results of MRI exhibited the shrinkage of soft-tissue lesion and amelioration of the CNF (Figure 1E). Evidences mentioned above further confirmed the effectiveness of our treatment. Ultimately, the patient recovered uneventfully from the surgery and discharged from hospital on Day 46.

3 Discussion

This case presented a diabetic patient diagnosed as CNF and successfully treated. With rapid detection of microbial sequencing based on NST, coinfection of *Prevotella* and *S. constellatus* was additionally identified, which contributed to the disease recognition and therapeutic success. Characterized with rapid progression and high mortality, CNF could exhibit improved prognosis with prompt recognition, appropriate surgical debridement, and precise antimicrobial medication. Molecular detection, including NST, could provide precise data on pathogenic microorganisms, which is essential for promptly adjusting antibiotic therapy for progressive diseases, such as CNF.

NSTIs is generally divided into four types, according to epidemiological features and causes (1, 11). Type I NSTIs is characterized by polymicrobial infection, usually includes both aerobic and anaerobic bacteria. Type II NSTIs is commonly monomicrobial infection, especially Gram-positive organisms, such



Process and diagnosis of CNF case. (A) Timeline of disease detection, diagnosis and treatment. (B) Image morphology of cervical and chest CT scan before (Day 9) and after (Day 12) thoracentesis incision. Disease progression at Day 9 and symptoms improvement after incision at Day 12. The subcutaneous emphysema was decreased significantly with bilateral pleural effusion (marked with red arrays). (C) Changing trends of clinical indicators and the LRINEC score retrospectively calculated during the course of disease. WBC, white blood cell; CRP, C-reactive protein. (D) Pathological examination exhibited tissue abscess and necrosis (marked with green arrays) with hematoxylin and eosin (H&E) staining (10x20). (E) Image morphology of cervical and chest MRI (Day 26). There is no significant subcutaneous emphysema 4 days after surgical debridement (marked with red arrays).

	pathogens				

Genus	Species	Number of detected reads		
Prevotella	Prevotella intermedia	773		
Prevotella	Prevotella enoeca	317		
Streptococcus	Streptococcus constellatus	252		

as group A Streptococcal (GAS) and methicillin-resistant *Staphylococcus aureus* (MRSA) (12). Infections caused by other pathogenic microbes, such as Gram-negative microbes and fungi, are classified as type III and Type IV NSTIs, respectively. The causative microorganism of this case was cultured as *S. mitis* only, and it suspected to be type II NSTIs. With Nanopore sequencing, pathogenic microorganisms were identified as *Prevotella* spp. and *Streptococcus* spp. Combined with elderly age and diabetes, the true classification should be type I NSTIs for this patient. Besides, clinical routine culture is usually aerobe-covered only and consuming 2 to 7 days. In contrast, Nanopore sequencing as an emerging platform, carries broader microbial coverage and shorter time of detection (9), which significantly contributes to type identification based on epidemiological features and causes, leading to a better understanding of NSTIs and control of disease progress.

Clinical manifestation plays a crucial role in the management of CNF. Unfortunately, during the initial phase, symptoms of CNF May be subtle or even absent. Combined with absence of specific laboratory indicators (13), those could lead to delayed treatment for CNF patients (14). CT scan and MRI are sufficient assessment for CNF. Nevertheless, surgical exploration remains the only definitive method for diagnosing NSTIs (15). Previous reported cases of CNF would emphasize the importance of surgical intervention on CNF treatment, especially for case with synchronous CNF and pharyngocutaneous fistula (16), patient with acute epiglottitis and abscess complicated with CNF (17), and surgical reconstruction of CNF (18). Combined with surgical debridement, control of hyperglycemia and source of infection would significantly affect the outcome of CNF (19). Thus, timely diagnosis and supportive therapies, including anti-infective drugs, surgical exploration and debridement, are crucial for improving patient outcomes. A case reported a patient initially treated as deep neck infection while later found descending necrotizing mediastinitis with CNF. The patient underwent repeat surgeries within the overall duration of 220 days (20). For this case, we used Nanopore sequencing for precise pathogen identification, supplement to the initial detection of culture and Gram staining. Results of NST supported the definitive antibiotics medication before and after the surgical debridement, fortunately achieving successful treatment of this high-risk case within 46 days.

Prevotella is a Gram-negative bacteria and *S. constellatus* is a Gram-positive coccobacillus, both of which are anaerobic and present in the oral microbiome of healthy people (21). There have been a few reported cases where certain strains of *Prevotella* and *S. constellatus* have caused opportunistic endogenous infections (22, 23), such as chronic infections, abscesses, and anaerobic pneumonia (21, 24). Although rarely reported, *Prevotella* and *S. constellatus* has been associated with NSTIs in recent case reports (25, 26), and found present in all NSTIs cases of fatality (27). For the patient of this case, considering the initial symptom of a sore throat and the presence of conditional pathogenic *Prevotella* and *S. constellatus* in the normal oral cavity, it is highly likely that the pathogenic bacteria originated

from the oral cavity. Furthermore, the identification of *Prevotella* should not be underestimated in NSTIs cases.

Pathogen detection is an important component in the diagnosis and treatment of infectious diseases. Nanopore platform is capable of direct RNA and DNA sequencing, providing a theoretical advantage in clinical diagnostics on both rapid species identification and antimicrobial resistance gene detection (9, 28). Cases of microbial identification and management of CNF patients were also previously reported, while the methodology was mainly utilized with culture (29, 30). In this particular case, traditional microbial culture took at least 72h to identify a positive coccus with unexplained subcutaneous emphysema, whereas NST successfully identified the presence of Prevotella and S. constellatus infections. Lydia-Ann J Ghuneim et al. has demonstrated that the final outcomes of antimicrobial therapy to polymicrobial infections are dictated by the complex microbial ecology, and the precised identification of ecological members May optimize antimicrobial treatment (31). Studies have proven that high-throughout sequencing, such as metagenomic next-generation sequencing, could improve detection rate for pathogen in polymicrobial bloodstream infections (32), polymicrobial brain abscess (33), and complex pleural infections (34). The results of this study provide supportive evidences for excellent performances of NST in pathogen detection. It highlights the usefulness of NST in identifying pathogens that are challenging to grow by using conventional methods. The prognosis could be significantly enhanced by early diagnosis using NST, identification of the causative pathogens, and targeted administration of antibiotic therapy.

4 Conclusion

In conclusion, this case presented an elderly high-risk CNF patient with type 2 diabetes, successfully recovered after prompt antibiotic treatment and appropriate surgical intervention. The key to this successful treatment relies on the appropriate application of molecular detection of NST for pathogen identification, precise antiinfective treatment, and collaborative efforts of surgical and multidisciplinary teams. NST is of great importance for the detection of unknown pathogens, and we expect further application of advanced sequencing technologies in acute and serious infectious diseases in the future.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found at: https://db.cngb.org/cnsa/, CNP0004703.

Ethics statement

Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Author contributions

MZ: Conceptualization, Data curation, Formal analysis, Methodology, Writing – original draft, Writing – review & editing. XL: Investigation, Methodology, Resources, Writing – original draft. JX: Project administration, Software, Writing – original draft. JC: Project administration, Supervision, Writing – original draft. SL: Conceptualization, Validation, Visualization, Writing – original draft, Writing – review & editing. WZ: Conceptualization, Funding acquisition, Resources, Supervision, Validation, Writing – review & editing, Writing – original draft.

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References

 Stevens DL, Longo DL, Bryant AE. Necrotizing soft-tissue infections. N Engl J Med. (2017) 377:2253–65. doi: 10.1056/NEJMra1600673

2. Faraklas I, Yang D, Eggerstedt M, Zhai Y, Liebel P, Graves G, et al. A multi-center review of care patterns and outcomes in necrotizing soft tissue infections. *Surg Infect.* (2016) 17:773–8. doi: 10.1089/sur.2015.238

 Quereshy FA, Baskin J, Barbu AM, Zechel MA. Report of a case of cervicothoracic necrotizing fasciitis along with a current review of reported cases. J Oral Maxillofac Surg. (2009) 67:419–23. doi: 10.1016/j.joms.2008.07.017

4. Miller JM, Binnicker MJ, Campbell S, Carroll KC, Chapin KC, Gonzalez MD, et al. Guide to utilization of the microbiology Laboratory for Diagnosis of infectious diseases: 2024 update by the Infectious Diseases Society of America (IDSA) and the American Society for Microbiology (ASM). *Clin Infect Dis.* (2024) 104. doi: 10.1093/cid/ciae104

5. Bonne SL, Kadri SS. Evaluation and Management of Necrotizing Soft Tissue Infections. *Infect Dis Clin N Am.* (2017) 31:497–511. doi: 10.1016/j.idc.2017.05.011

6. Diao Z, Han D, Zhang R, Li J. Metagenomics next-generation sequencing tests take the stage in the diagnosis of lower respiratory tract infections. J Adv Res. (2022) 38:201–12. doi: 10.1016/j.jare.2021.09.012

7. Jin X, Li J, Shao M, Lv X, Ji N, Zhu Y, et al. Improving suspected pulmonary infection diagnosis by Bronchoalveolar lavage fluid metagenomic next-generation sequencing: a multicenter retrospective study. *Microbiol Spectr.* (2022) 10:e0247321. doi: 10.1128/spectrum.02473-21

8. Gaston DC, Miller HB, Fissel JA, Jacobs E, Gough E, Wu J, et al. Evaluation of metagenomic and targeted next-generation sequencing workflows for detection of respiratory pathogens from Bronchoalveolar lavage fluid specimens. *J Clin Microbiol.* (2022) 60:e0052622. doi: 10.1128/jcm.00526-22

9. Zhao M, Zhang Y, Chen L, Yan X, Xu T, Fu M, et al. Nanopore sequencing of infectious fluid is a promising supplement for gold-standard culture in real-world clinical scenario. *Front Cell Infect Microbiol.* (2024) 14:1330788. doi: 10.3389/ fcimb.2024.1330788

10. Hsiao CT, Chang CP, Huang TY, Chen YC, Fann WC. Prospective validation of the laboratory risk Indicator for necrotizing fasciitis (LRINEC) score for necrotizing fasciitis of the extremities. *PLoS One.* (2020) 15:e0227748. doi: 10.1371/journal.pone.0227748

11. Jabbour G, El-Menyar A, Peralta R, Shaikh N, Abdelrahman H, Mudali IN, et al. Pattern and predictors of mortality in necrotizing fasciitis patients in a single tertiary hospital. *World J Emerg Surg.* (2016) 11:40. doi: 10.1186/s13017-016-0097-y

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Conflict of interest

SL was employed by the company Dian Diagnostics Group Co., Ltd. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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12. Misiakos EP, Bagias G, Patapis P, Sotiropoulos D, Kanavidis P, Machairas A. Current concepts in the management of necrotizing fasciitis. *Front Surg.* (2014) 1:36. doi: 10.3389/fsurg.2014.00036

13. Moskowitz E, Schroeppel T. Necrotizing fasciitis following abdominal gunshot wound. *Trauma Surg Acute Care Open.* (2018) 3:e000163. doi: 10.1136/tsaco-2018-000163

14. Diab J, Bannan A, Pollitt T. Necrotising fasciitis. *BMJ*. (2020) 369:m1428. doi: 10.1136/bmj.m1428

15. Henry SM, Davis KA, Morrison JJ, Scalea TM. Can necrotizing soft tissue infection be reliably diagnosed in the emergency department? *Trauma Surg Acute Care Open*. (2018) 3:e000157. doi: 10.1136/tsaco-2017-000157

16. Al-Wageeh S, Ahmed F, Alyhari Q, Mohammed F. Synchronous cervical necrotizing fasciitis and pharyngocutaneous fistula: a case report. *Int J Surg Case Rep.* (2022) 93:106988. doi: 10.1016/j.ijscr.2022.106988

17. Fu G, Yang L, Wu G. Difficult intubation in a patient with acute epiglottitis and abscess complicated with cervical necrotizing fasciitis: a case report. *Medicine* (*Baltimore*). (2024) 103:e38658. doi: 10.1097/md.00000000038658

18. Cho W, Jang EA, Kim KN. Reconstruction of cervical necrotizing fasciitis defect with the modified keystone flap technique: two case reports. *World J Clin Cases*. (2024) 12:1305–12. doi: 10.12998/wjcc.v12.i7.1305

19. Sidam S, Bhagat A, Chavan A, Sahoo AK. Cervical necrotizing fasciitis: a surgical outcome analysis. *Cureus*. (2023) 15:e44678. doi: 10.7759/cureus.44678

20. Lauček P, Šiška D, Lučenič M, Juhos P, Janík M. Complicated case of patient with cervical necrotising fasciitis and descending necrotizing mediastinitis case report. *Rozhl Chir.* (2020) 99:189–93. doi: 10.33699/pis.2020.99.4.189-193

21. Xia J, Xia L, Zhou H, Lin X, Xu F. Empyema caused by *Streptococcus constellatus*: a case report and literature review. *BMC Infect Dis.* (2021) 21:1267. doi: 10.1186/s12879-021-06955-2

22. Claridge JE 3rd, Attorri S, Musher DM, Hebert J, Dunbar S. Streptococcus intermedius, Streptococcus constellatus, and Streptococcus anginosus ("Streptococcus milleri group") are of different clinical importance and are not equally associated with abscess. Clin Infect Dis. (2001) 32:1511-5. doi: 10.1086/320163

23. Tett A, Pasolli E, Masetti G, Ercolini D, Segata N. Prevotella diversity, niches and interactions with the human host. *Nat Rev Microbiol.* (2021) 19:585–99. doi: 10.1038/s41579-021-00559-y

24. Hisamoto T, Hirabayashi M, Nakatani M, Akiyama Y, Takehana A, Jikuya S, et al. Subcutaneous abscess in the shoulder caused by *Prevotella bivia* infection. *Anaerobe.* (2022) 76:102609. doi: 10.1016/j.anaerobe.2022.102609

25. Chang N, McKee J, Marmolejo V. Necrotizing fasciitis due to *Streptococcus constellatus* in a patient with uncontrolled diabetes and bilateral diabetic foot ulceration. *Wounds*. (2023) 35:E74–e77. doi: 10.25270/wnds/22044

26. Ling J, Hirase T. Necrotizing fasciitis due to *Prevotella denticola* infection in an intravenous drug user. *Cureus.* (2022) 14:e20901. doi: 10.7759/cureus.20901

27. Zhao-Fleming HH, Wilkinson JE, Larumbe E, Dissanaike S, Rumbaugh K. Obligate anaerobes are abundant in human necrotizing soft tissue infection samples - a metagenomics analysis. *APMIS*. (2019) 127:577–87. doi: 10.1111/apm.12969

28. Baldan R, Cliff PR, Burns S, Medina A, Smith GC, Batra R, et al. Development and evaluation of a nanopore 16S rRNA gene sequencing service for same day targeted treatment of bacterial respiratory infection in the intensive care unit. *J Inf Secur.* (2021) 83:167–74. doi: 10.1016/j.jinf.2021.06.014

29. Abbasi Z, Inam H, Das S, Neel S, Fatimi SH. Fungal cervical abscess complicated by necrotizing fasciitis leading to descending necrotizing Mediastinitis: a case report. *Cureus*. (2019) 11:e5369. doi: 10.7759/cureus.5369

30. Chaurasiya PS, Gurung S, Karki S, Timilsina B, Shah R, Neupane S. *Pseudomonas aeruginosa* as a culprit of cervical necrotizing fasciitis: a case report. *Int J Surg Case Rep.* (2022) 99:107713. doi: 10.1016/j.ijscr.2022.107713

31. Ghuneim LJ, Raghuvanshi R, Neugebauer KA, Guzior DV, Christian MH, Schena B, et al. Complex and unexpected outcomes of antibiotic therapy against a polymicrobial infection. *ISME J.* (2022) 16:2065–75. doi: 10.1038/ s41396-022-01252-5

32. Liu Q, Liu X, Hu B, Xu H, Sun R, Li P, et al. Diagnostic performance and clinical impact of blood metagenomic next-generation sequencing in ICU patients suspected monomicrobial and polymicrobial bloodstream infections. *Front Cell Infect Microbiol.* (2023) 13:1192931. doi: 10.3389/fcimb.2023.1192931

33. Stebner A, Ensser A, Geißdörfer W, Bozhkov Y, Lang R. Molecular diagnosis of polymicrobial brain abscesses with 16S-rDNA-based next-generation sequencing. *Clin Microbiol Infect*. (2021) 27:76–82. doi: 10.1016/j.cmi.2020.03.028

34. Kanellakis NI, Wrightson JM, Gerry S, Ilott N, Corcoran JP, Bedawi EO, et al. The bacteriology of pleural infection (TORPIDS): an exploratory metagenomics analysis through next generation sequencing. *Lancet Microbe*. (2022) 3:e294–302. doi: 10.1016/s2666-5247(21)00327-x