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Can a diet rich in Brassicaceae help control *Helicobacter pylori* infection? A systematic review

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Introduction: *Helicobacter pylori* (*Hp*) infection is highly prevalent globally and poses a significant public health challenge due to its link with chronic gastritis, peptic ulcers, and gastric malignancies. *Hp*'s persistence within the gastric environment, particularly in case of infection with virulent strains, triggers chronic inflammatory responses and mucosal damage. Antibiotic therapy is the primary approach for *Hp* eradication, but antibiotic resistance and adverse effects hinder treatment efficacy. Emerging evidence suggests that Brassicaceae-derived metabolites could serve as adjunctive therapy for *Hp* infection, offering potential antimicrobial and anti-inflammatory benefits.

Methods: A systematic literature review was conducted following PRISMA guidelines to assess the impact of Brassicaceae-rich diets on *Hp* infection control. Searches were performed in MEDLINE PubMed, Web of Science, and the Cochrane Library until 18 October 2023, without language or date restrictions. Eligible studies meeting PICOS criteria were included, encompassing populations infected with *Hp* or *Hp*-infected human cell cultures, interventions involving Brassicaceae consumption or its bioactive molecules, and outcomes related to *Hp* infection control, antibiotic therapy interactions, reduction of antibiotic side effects, and inflammation mitigation. Animal studies, cell line experiments, reviews unrelated to the research objectives, and studies on *Hp*-related gastric cancer were excluded.

Results: Available evidence indicates that Brassicaceae consumption exhibits the potential to reduce *Hp* colonization but achieving complete eradication of the pathogen remains challenging. Conflicting results regarding the efficacy of broccoli in *Hp* treatment emerge, with certain investigations suggesting limited effectiveness. Other studies point to a potential for heightened eradication rates when combined with standard triple therapy. Furthermore, promising outcomes are observed with broccoli extract supplements, indicating their role in mitigating *Hp*-induced gastric mucosal damage. In fact, it is noteworthy that sulforaphane and its derivatives manifest notable reductions in pro-inflammatory markers, indicative of their anti-inflammatory properties. Adverse events associated with antibiotic therapy seem unaffected by sulforaphane derivatives or probiotics. However, individual responses to these treatments vary, underscoring the unpredictability of their efficacy in ameliorating antibiotic therapy-related side effects.

Conclusion: Our systematic review highlights the potential of Brassicaceaerich diets as adjunctive therapy for *Hp* infection, offering synergistic interactions with antibiotics and possibly mitigating antibiotic side effects and inflammation. Further research, particularly well-designed randomized trials, is warranted to elucidate the therapeutic efficacy and optimal utilization of Brassicaceaederived metabolites in managing human *Hp*-related diseases.

KEYWORDS

Helicobacter pylori infections/diagnosis, Helicobacter pylori, Brassicaceae plants, glucosinolates, thiocyanate

1 Introduction

Helicobacter pylori (*Hp*) is a Gram-negative microaerophilic bacterium (1), that colonizes one-half of the world's population (2). *Hp* is the main risk factor for gastric cancer (3). Disseminating through oro-oral and/or oro-fecal routes (4), generally within families during early childhood (2), before the age of 10 years (5). However, high prevalence and early age of infection do not always translate in high incidence of gastric cancer (6). Risk and protective factors may modulate gastric cancer risk in *Hp* infected patients and consumption of Brassicaceae has been proposed as a possible protective factor (7).

Over the past century, there has been a remarkable but not uniform global reduction in the prevalence of Hp infection. Recent studies indicate a significant decrease in the global prevalence of Hpinfection from 58% in the 1980–1990 period to 43% in 2011–2022 (5). However, this overall decline masks substantial regional and demographic variability. The most notable reductions have been observed in high-income countries, reflecting sanitation, healthcare access, spread of Hp eradication treatment, and other public health initiatives. In contrast, the prevalence remains high in many developing countries, particularly in parts of Africa, South America, and Asia, where the prevalence of infection often exceeds 70%. These areas experience slower declines due to persistent socioeconomic challenges, limited access to clean water, and crowded living conditions, which facilitate the transmission of Hp (8).

Moreover, in high-income countries, younger populations exhibit significantly lower prevalence rates compared to older cohorts, reflecting the impact of improved living conditions and healthcare advancements over recent decades (8). Despite the fact that approximately 80% of colonized individuals display no symptoms, persistent Hp infection almost always leads to the development of chronic gastritis (9). The severity of the chronic inflammatory process associated with Hp infection is influenced by various factors, including the colonizing strain's virulence, host genetics, immune response, and dietary habits (10). The disease spectrum associated with Hp chronic infection encompasses duodenal ulceration, often associated with antral gastritis, gastric ulceration in cases of corpus gastritis, gastric malignancy (10), and MALT (Mucosa Associated Lymphoid Tissue) lymphoma (11–13). Hp's survival and proliferation within the acidic gastric milieu is facilitated by its secretion of urease, which enzymatically hydrolyzes host urea into ammonia and nitric oxide (14) and facilitates flagellar motility through the mucus layer by changing the viscoelasticity properties of gastric mucins (15). Hp's adhesion and colonization of the gastric mucosa are mediated by adhesins (BabA, blood group antigen-binding adhesion (16), SabA, sialic acid-binding adherence (16), and LabA, lacdiNAc-binding adhesin (17)) and virulence factors like VacA (Vacuolating-toxin A, apoptotic cell death inductor or necrosis inductor through mitochondrial membrane perturbation. VacA suppresses T and B cell proliferation, as well as various immune cell populations, thereby downregulating immune responses to *Hp* infection and promoting host tolerance to the organism (18)) and CagA (Cytotoxin-associated gene A involved in adhesion mechanisms and pro-inflammatory cytokine release (13)), predominantly expressed by more pathogenic *Hp* strains (13). Host receptors (TLR and NOD-like receptors) bind to pathogen components, eliciting pro-inflammatory gene activation (13). This immune response activates the NF- κ B (Nuclear factor kappa-light-chain-enhancer of activated B cells) pro-inflammatory signaling pathway (19) and the NRF2 (Nuclear factor erythroid 2-related factor 2) anti-inflammatory signaling pathway (20), leading to cytokine release including TNF- α , IL-1 β , IL-8, and IFN- γ (21).

Following Hp colonization, the gastric mucosa undergoes cellular infiltration, primarily consisting of neutrophils and mononuclear cells. These infiltrating cells actively engage in phagocytosis of the pathogen and, by expressing inducible nitric oxide synthase (iNOS), secrete nitric oxide and reactive oxygen species (ROS) (22). In fact, in Hp-infected gastric mucosa, neutrophils are believed to be the main source of ROS (23). After the recognition of pathogenic bacteria, neutrophils immediately phagocytize and kill them through ROS (24). However, in the Hp-infected gastric mucosa, immune cells cannot eradicate the bacteria. Therefore, a persistent inflammatory status of gastric mucosa is established where the phagocytes produce inflammatory cytokines and ROS in response to bacteria. This excessive production of ROS is believed to be a major cause of gastric mucosal damage (24), and contributes, together with direct mucosal damage from virulent Hp strains, to the multi-step pathogenesis of chronic lesions of the stomach (22, 25). Furthermore, Hp infection elevates blood levels of Pepsinogen I (PGI), Pepsinogen II (PGII), and Gastrin-17 (G-17), serving as biological markers to assess gastric mucosal inflammation (26) and functionality (27).

An effective treatment regimen for pathogen eradication and control is critical to mitigate the clinical manifestations of Hpinfection (28). International guidelines advocate assessing local clarithromycin resistance prevalence to select first-line therapy selection. In regions where the prevalence of clarithromycinresistant Hp strains is high (>15%) or unknown, a 14-day quadruple therapy incorporating a proton pump inhibitor (PPI), amoxicillin or bismuth, clarithromycin, and metronidazole is recommended for eradication (29). PPIs are believed to synergize with antimicrobials by attenuating gastric acidity (30), inhibiting Hp urease activity (31), and exhibiting intrinsic antibacterial properties (30). It is also known that PPIs can significantly impact the gut microbiota reducing microbial diversity and increasing the risk of overgrowth of harmful bacteria (32).

Furthermore, various natural products and food components exhibit anti-*Hp* activity. Key agents include polyphenols, flavonoids, essential oils, and compounds from medicinal plants. These substances work through mechanisms such as inhibiting *Hp* growth, reducing adhesion to gastric cells, and modulating inflammation (33).

TABLE 1 Search strategy.

Search engine	Search strategy
MEDLINE/PubMed	From 1000/01/01 to 18/10/2023:
	(Helicobacter pylori OR helicobacter pylori infection) AND (quercetin OR sulforaphane OR glucosinolates OR isothiocyanate) AND (Brassicaceae
	OR Brassica OR cruciferous vegetables OR broccoli OR cabbage OR cauliflower OR Brussels sprouts OR mustard plants OR mustard oil OR
	sauerkraut OR coleslaw OR collards OR bok choy OR turnip greens OR vegetables OR rocket OR wasabi)
Web of science	From 1000/01/01 to 18/10/2023:
	Query: TS = ((helicobacter pylori OR helicobacter pylori infection) AND (quercetin OR sulforaphane OR glucosinolates OR isothiocyanate) AND
	(Brassicaceae OR Brassica OR cruciferous vegetables OR broccoli OR cabbage OR cauliflower OR Brussels sprouts OR mustard plants OR mustard
	oil OR sauerkraut OR coleslaw OR collards OR bok choy OR turnip greens OR vegetables OR rocket OR wasabi)).
Cochrane library	From 1000/01/01 to 18/10/2023:
	(helicobacter pylori OR helicobacter pylori infection) AND (quercetin OR sulforaphane OR glucosinolates OR isothiocyanate) AND (Brassicaceae
	OR Brassica OR cruciferous vegetables OR broccoli OR cabbage OR cauliflower OR Brussels sprouts OR mustard plants OR mustard oil OR
	sauerkraut OR coleslaw OR collards OR bok choy OR turnip greens OR vegetables OR rocket OR wasabi).

A recent narrative review on *Hericium erinaceus*, commonly known as the Chinese mushroom, underscores the potential of specific mushroom fractions, particularly those derived from ethyl acetate or HEP25/75, which exhibit promising anti-*Hp* activities. These fractions demonstrate noteworthy minimum inhibitory concentration (MIC) values against the bacterium (34).

Previous investigations (35-37) underscore the potential of compounds in Brassicaceae vegetables (e.g., Cabbage, Broccoli, Cauliflower, Brussels sprouts, Kale, Collard greens, Turnip, Radish, Mustard greens, Horseradish, Rutabaga (Swede), Watercress, Arugula (Rocket), Bok choy (Pak choi), Chinese cabbage, Rapeseed (Canola), Wasabi (38)) to mitigate oxidative stress in vitro and exert antiinflammatory effects on Hp-infected gastric mucosa in animal models and humans. Cruciferous vegetables, particularly broccoli sprouts, contain glucosinolates, such as glucoraphanin, and quercetin, that are hydrolyzed by myrosinase within the vegetables themselves and by intestinal flora into active molecules (39), isothiocyanates such as sulforaphane, a potent inducer of phase 2 detoxification enzymes (40), implicated in pro-inflammatory cytokine downregulation. Sulforaphane further mitigates inflammation by reducing NF-KB expression and activating the NRF2 pathway (35), exhibiting bacteriostatic and bactericidal effects against Hp in vitro and efficacy against extensively antibiotic-resistant Hp clinical isolates (41). Sulforaphane has also been studied for its ability to reduce the risk of colorectal cancer (42) and for enhancing the sensitivity of cancer cells to chemotherapy, potentially overcoming chemoresistance (43).

Quercetin, a flavonoid abundant in Brassicaceae (44), exhibits anti-inflammatory and antioxidant properties in rodents via NRF2 upregulation and NF- κ B downregulation (45). However, confirmation of quercetin's anti-inflammatory and *Hp* infection control efficacy, as demonstrated in animal models (46, 47), in human subjects remains lacking. In fact, despite numerous investigations elucidating the impacts of Brassicaceae derivatives on animals or *in vitro* regarding *Hp* infection, scant evidence substantiates their efficacy in human cases of *Hp* infection. This systematic review explores the potential of a diet rich in Brassicaceae vegetables in influencing the clinical outcomes of *Hp* infection in humans. Particularly it investigates possible synergistic interactions between Brassicaceae-derived metabolites and antibiotic therapy for eradication, and the role of such metabolites in alleviating the adverse effects of antibiotics and mitigating *Hp*-induced inflammation.

2 Materials and methods

2.1 Research strategy

We conducted a systematic literature review, adhering to the PRISMA statement (48), to investigate the potential impact of a diet rich in Brassicaceae on the control of Hp infection. The protocol was registered and is available on PROSPERO (CRD42024517622)¹. Three electronic databases (MEDLINE PubMed, Web of Science, and Cochrane Library) were searched without language or date restrictions, from inception to 18/10/2023 for relevant original articles.

We started the search on MEDLINE and then identified an appropriate syntax for the other databases. Our search strategy included terms related to Hp infection, Brassicaceae vegetables, and their bioactive molecules.

The entire search strategy is reported in Table 1.

2.2 Eligibility criteria

Included studies met the following eligibility criteria described as PICOS (48): (1) P (Population): individuals infected with Hp regardless of age, gender, health status, and any other variable influenced by the infection and/or human cells *in vitro* infected with Hp; (2) I (Intervention): intake of brassicaceae vegetables and their active bioactive molecules; (3) C (Control): standard or non-necessary; (4) O (Outcome): interaction of treatments with eradication antibiotic therapy and/or reduction of adverse effects from eradication antibiotic therapy by treatments, inhibition and/or control of Hp infection, reduction of inflammation; (5) S (Studies): primary studies.

We excluded: (1) studies conducted exclusively on animals; (2) studies conducted on *Hp* cell lines maintained in culture broths; (3) narrative and systematic reviews, case series; (4) studies evaluating the

¹ http://www.crd.york.ac.uk/PROSPERO

relationship between *Hp* infection and gastric cancer; (5) studies for which the full text was not available; (6) studies concerning data irrelevant to this analysis; (7) duplicate studies.

2.3 Study selection

The first author (SP) imported the literature into the Rayyan online platform (49). After manually removing duplicates, two reviewers (SP and MR) independently screened the articles, first by title and abstract, then by full-text, to determine eligibility for final inclusion. Discrepancies during screening were resolved by consensus or referred to a third reviewer (AG). In cases where multiple publications were associated with

the same studies, a key paper for each study was selected, and then the other associated publications were used for supplementary information during the data extraction process. Sources deemed unsuitable for inclusion at this stage were systematically documented, accompanied by an outline of the reasons for their exclusion (Figure 1). The next phase involved data extraction and cumulative assessment by the reviewers.

2.4 Data extraction

Systematic retrieval of relevant information from each article was performed via a standardized data extraction form (Microsoft Excel 2019, Microsoft Corp). The extracted data included the



name of the first author, year of publication, geographic region under study, study setting, key characteristics of the study population, study outcomes, and respective quantitative assessment methods. In addition, the data were stratified according to the outcomes investigated the nature of the interventions and the data extraction process was performed by two independent reviewers (SP and MR). Cases of disparity or divergence in data extraction were resolved through extensive deliberation and consensus building.

2.5 Data synthesis

In the context of this systematic review, a narrative approach was used to succinctly summarize the data results. The results were described in separate paragraphs based on the type of intervention, to facilitate a comparison between studies. Limited number of studies and data availability across some of the included studies precluded meta-analytic synthesis.

2.6 Methodological quality assessment (risk of bias)

For quality assessment, the CASP Randomized Controlled Trial Checklist (50) was employed, which considers 11 aspects grouped into three sections. Specifically, "Section A" provides screening questions about the validity of the basic study design, "Section B" assesses whether the study was methodologically sound, "Section C" provides for the investigation of the results, "Section D" provides for the investigation of local usefulness of results. Responses to the questions were categorized as "Yes," "No," or "Cannot tell" with a score of one assigned to affirmative responses.

Two independent reviewers (SP and MR) conducted the assessment process. In cases where disparities arose in assigning scores to individual included studies, these were resolved through thorough reflection and discussion between the two reviewers. Additionally, in cases where consensus could not be reached, the expertise of a third reviewer (AG) was sought to facilitate resolution.

3 Results

3.1 Study selection

A total of 133 potential records were identified from the searched electronic databases. After removing duplicates (31), 102 articles were reviewed by title and abstract. The full text of 23 studies was assessed for possible inclusion. Eleven studies were excluded for the following reasons: full text unavailable (4), different study population (4), incorrect study design (1), and irrelevant study (2). Twelve studies (51–62), which met all inclusion criteria, were considered in the review (Figure 1).

3.2 Study characteristics

Among the included studies, 4 were conducted in Japan (52, 53, 55, 56), 2 in the United States (54, 57), 2 in Korea (51, 58), 2 in Iran

The overall quality of the papers was rated as moderate, based on assigned scores of 7/11 (54, 57, 59, 60, 62), 7.5/11 (52, 53, 55, 56), and 8/11 (51, 58, 61) (Table 2).

The study population comprised individuals with confirmed Hp infection (51–59, 61, 62), alongside xenografted human cell populations infected with Hp (60) (Table 2).

The primary criteria for subject exclusion were consistent across studies, including recent Hp eradication therapy (57–59, 61), prior gastric surgery (51, 58, 59), recent use of anti-inflammatory drugs (58), antibiotics (51, 54, 57, 59), PPIs (51, 57, 59), histamine H2-receptor antagonists and bismuth-based formulations (57, 59).

3.3 Findings by intervention

3.3.1 Interaction of treatments with eradication antibiotic therapy and/or changes in adverse effects from eradication antibiotic therapy by treatments

In an Iranian RCT (59) in total, 362 patients diagnosed with type 2 diabetes were initially assessed for potential inclusion in the study. Stool samples were collected from the 191 eligible patients to detect Hp infection using the HpSAg test. Of the 108 patients identified with Hp infection, 86 were randomly assigned to treatment: group A (n = 33) received standard triple therapy (STT) with omeprazole 20 mg, clarithromycin 500 mg, and amoxicillin 1.000 mg, administered twice daily for 14 days; group B (n = 28) received 6 g/ day broccoli sprout powder (BSP) for 28 days; group C (n = 25) received standard triple therapy (STT) for 14 days plus 6 g/day broccoli sprout powder (BSP) for 28 days. After accounting for losses at follow-up or treatment discontinuation, 77 of the 86 randomized patients completed the study (STT = 28; BSP = 25; STT + BSP = 24). The eradication rates of Hp after the treatment were 85.3, 36.0%, and 83.3 (assessed by UBT) and 89.3, 56.0, and 91.7% (assessed by HpSAg test) respectively in the STT, BSP, and STT + BSP groups. The levels of HpSAg were significantly lower in STT and STT + BSP groups compared to the BSP group, but there were no significant differences in serum PGI, PGII, and PGI/PGII ratio between the three groups. In a Korean trial (51) patients diagnosed with chronic gastritis or individuals with peptic ulcers and Hp infection were recruited for the study. Diagnosis of Hp infection was confirmed through either rapid urease testing or UBT. Additionally, patients underwent clarithromycin resistance testing and excluded from the study if resistant. The subjects (n = 183) were randomly allocated into three treatment groups. Group A (n = 61) received triple therapy consisting of 40 mg of pantoprazole, 1 g of amoxicillin, and 500 mg of clarithromycin administered twice daily for 7 days. Group B (n = 61) received the same triple therapy regimen for 7 days along with probiotics (Saccharomyces boulardii capsules, 3×10^{10} colony-forming units/g). Group C (n = 61) received triple therapy for 7 days in addition to sulforaphane supplementation (capsules containing 250 mg of standardized broccoli sprouts, yielding 1.000 µg of sulforaphane) 3 times daily for 4 weeks. The eradication rates -as determined by intention-to-treat (ITT) and per-protocol (PP) analyses- were 85.2 and 89.6% in Group A, 81.9 and 89.2% in Group B, and 86.8 and 96.3% in Group C. No significant difference in eradication rates emerged by treatment group, (ITT analysis: A vs. B,

TABLE 2 Studies characteristics.

Author, year	Country	Study design	Population	Intervention	Outcome
 A. Yanaka et al, 2009 (56), A. Yanaka et al, 2011 (55), A. Yanaka et al, 2017 (52) A. Yanak et al, 2018 (53) 	Japan	RCT	50 <i>Hp</i> positive volunteers, whose endoscopy showed no abnormalities other than gastritis.	BS group: 25 <i>Hp</i> positive received 70g/day of SGS-rich 3-days-old BS (~420 μmol of glucoraphanin) for 8 weeks; AS group: 25 <i>Hp</i> positive received 70gr of alfalfa sprouts lacking glucoraphanin daily for 8 weeks.	Inhibition a/o control of <i>Hp</i> infection; reduction of inflammation.
Y. W. Chang et al, 2015 (58)	Korea	DB-RCT	100 subjects with functional dyspepsia underwent upper gastrointestinal endoscopies, UBT, assessment of MDA and GSH levels. 67 subjects <i>Hp</i> positive, 33 subjects <i>Hp</i> negative. 11 subjects lost at follow up. 89 Included.	Group A: 33 <i>Hp</i> positive take BSES (250 mg standardized broccoli sprout equal to 1,000 mg of sulforaphane). Group B: 28 <i>Hp</i> positive take placebo. Group C: 28 <i>Hp</i> negative take BSES.	Inhibition a/o control of <i>Hp</i> infection; reduction of inflammation.
K. Guo et al, 2022 (61)	China	DB-RCT	533 volunteers with no atrophic gastritis screened by 14C-UBT, of these 138 retested by 13C-UBT, 110 of the 138 retested were included as <i>Hp</i> positive.	55 patients randomly allocated to receive 4.5g of AESB (aqueous extract of seed of broccoli) a day, for 2 months before bedtime; 55 patients randomly allocated to receive 4.5g of placebo a day, for 2 months before bedtime.	Inhibition a/o control of <i>Hp</i> infection; reduction of inflammation.
P. Mirmiran et al, 2017 (62)	Iran	RCT	362 patients with DM2 assessed for potential inclusion; 191 eligible patients undergo HpSAg test to detect <i>Hp</i> infection. Of 108 patients <i>Hp</i> positive, 86 were randomly assigned into one of the 3 groups.	Group A (STT $n=33$) received omeprazole 20mg, clarithromycin 500mg, and amoxicillin 1.000mg, twice daily for 14 days; Group B (BSP $n = 28$) received 6 g/day broccoli sprout powder for 28 days; Group C (STT + BSP $n = 25$) received standard triple therapy for 14 days plus 6 g/day broccoli sprout powder for 28 days.	Inhibition a/o control of <i>Hp</i> infection; reduction of inflammation.
M. V. Galan et al, 2004 (54)	Michigan	RCT	Patients with evidence of active <i>Hp</i> infection established by gastric biopsy or stool antigen testing.	9 patients random assigned to receive 7, 14, or 28 g of broccoli sprouts on an empty stomach twice daily for 7 days.	Inhibition a/o control of <i>Hp</i> infection a/o changes in adverse effects from eradication antibiotion therapy by treatments.
A. R. Opekun et al, 2005 (57)	Texas	СТ	12 volunteers <i>Hp</i> positive at UBT.	5 volunteers <i>Hp</i> positive received 135 gr of fresh, finely minced juvenile broccoli tips (135 μmol of glucoraphanin) added to ½ cup commercial plain yogurt; 7 volunteers <i>Hp</i> positive received 120ml of Tibetan yogurt whey twice daily for 3.5 days.	Inhibition a/o control of <i>Hp</i> infection.
X. Haristoy et al, 2003 (60)	France	RCT	22 xenografts, derived from mature human gastric epithelium and implanted in nude mice, uniformly infected with <i>Hp</i> -26695 type.	Intervention group received a solution of 0.5 ml of sterile water with 0.5% acetonitrile containing 7.5 μ mol of sulforaphane (1.33 mg) once daily for 5 consecutive days. Control group received an identical solution without sulforaphane once daily for 5 days.	Inhibition a/o control of <i>Hp</i> infection.
Z. Bahadoran et al, 2014 (59)	Iran	DB-RCT	362 patients with DM2 assessed for potential inclusion; 191 eligible patients undergo HpSAg test to detect <i>Hp</i> infection. Of 108 patients <i>Hp</i> positive, 86 were randomly assigned into one of the 3 groups.	Group A ($n = 33$) received standard triple therapy with omeprazole 20mg, clarithromycin 500mg, and amoxicillin 1.000mg, administered twice daily for 14 days; group B ($n = 28$) received 6 g/day broccoli sprout powder for 28 days; group C ($n = 25$) received standard triple therapy for 14 days plus 6 g/day broccoli sprout powder for 28 days.	Interaction of treatments with eradication antibiotic therapy.
Y. W. Chang et al, 2020 (51)	Korea	RCT	Patients with <i>Hp</i> positive chronic gastritis or peptic ulcer disease, no clarithromycin resistance.	Group A (61 patients): pantoprazole 40mg, amoxicillin 1g, clarithromycin 500mg twice daily/7 days. Groups B (61 patients) & C (61 patients): same PPI-based triple therapy for 7 days. Additionally, a capsule of probiotics or sulforaphane provided three times daily for 4 weeks to patients in groups B and C, respectively.	Interaction of treatments with eradication antibiotic therapy a/c changes in adverse effects from eradication antibiotic therapy by treatments.

p = 0.744; A vs. C, p = 1.000; PP analysis: A vs. B, p = 0.313; A vs. C, p = 0.273). There was also no significant variation in the overall frequency of adverse events (taste disorders, diarrhea, headache, epigastric pain, nausea, and urticaria) among the three groups, although group B showed a lower frequency of gastrointestinal disorders than group A (comparison test A vs. B, p = 0.339). The study, therefore, does not demonstrate statistically significant effects resulting from supplementary therapy with probiotics or sulforaphane. In line with those findings, in a RCT conducted in Michigan (54) at baseline, 4 patients reported symptoms (abdominal discomfort such as pain, nausea, and bloating); one patient declared symptom improvement, one no change, and one worsening of symptoms following broccoli sprout therapy.

3.3.2 Inhibition and/or control of *Hp* infection and reduction of inflammation

A Japanese RCT, reported in 4 different publications (52, 53, 55, 56), was conducted to assess the efficacy of a treatment protocol on a cohort of 50 individuals infected by Hp, presenting with gastritis as the sole endoscopic abnormality. Participants were divided into two groups: the treatment group (Broccoli Sprouts BS group = 25) which involved the daily consumption of 70 g (~420 µmol) of glucoraphanin-rich 3-day-old germinated broccoli sprouts for 8 weeks, and the control group (Alfalfa Sprouts AS group = 25) which ingested an equivalent quantity of alfalfa sprouts lacking glucoraphanin, glucosinolates, or isothiocyanates for the same duration. Blood and stool samples of participants were collected at baseline (0 weeks), mid-intervention (4 weeks), post-intervention (8 weeks), and follow-up (16 weeks, corresponding to 8 weeks after the intervention period), and used to evaluate, respectively, PGI, PGII and the PGI/PGII ratio, and the presence of Hp stool antigen using HpSAg-ELISA. During the intervention, there were significant reductions of PGII, compared with baseline, only in the BS group, but these returned to baseline values 2 months after the intervention. There was also a significant increase in PGI/PGII ratio in the BS group. HpSAg levels measured in the BS group were significantly lower during the intervention than at baseline but returned to baseline 2 months after the intervention. The placebo group receiving AS showed no significant change in HpSAg. Of the 25 subjects in the BS treatment group, 8 had HpSAg values below the cutoff (0.100) at the end of the 8-week treatment period. In 6 of these subjects, the HpSAg values became positive again at 8 weeks after cessation of BS consumption, and the values for the remaining two subjects became positive again 6 months after intervention, indicating that BS treatment significantly reduced Hp colonization during the treatment but did not result in complete eradication (52, 53, 55, 56).

A Chinese RCT (61) enrolled a total of 533 volunteers screened by 14C-UBT for *Hp* infection. Among these, 138 were retested by 13C-UBT for *Hp* infection to exclude false positives. Subsequently, 55 individuals were randomly allocated to the AESB (aqueous extract of seed of broccoli containing 16.31% of glucosinolate, 14.10% glucoraphanin) group, while another 55 were assigned to the placebo group. Participants ingested either 4.5 g of AESB or an equivalent amount of placebo, visually indistinguishable, daily for 2 months before bedtime. Blood samples were obtained following an overnight fasting period to evaluate plasma levels of PG I and PG II and the PGI/ PGII ratio, plasma concentrations of inflammatory cytokines, including IFN-γ, TNF-α, CRP, IL-17A, IL-1β, IL-8, IL-18, and G-17. Normal reference ranges for PG and G-17 were defined as PG I > 70 ng/mL, PGR > 3, and G-17 levels between 1 and 7 pmol/L. The 13C-UBT was performed also at 60 days. No significant differences were observed in any of the inflammatory cytokines between the two groups at baseline and at 60 days follow-up; IL-8 and IL-18 at 60 days were reduced in the AESB groups compared to baseline, but only IL-8 significantly. IFN- γ decreased in both the placebo and AESB group at 60 days, but the reduction was significant only in AESB group. In the placebo group, PGR increased significantly at 60 days compared to baseline, but no significant differences were found in PG I, PG II and G-17 before and after treatment. In contrast, PGR, PG I and G-17 significantly decreased in AESB group at 60 days compared to baseline, but no significant difference was found in PG II after AESB treatment. The eradication rate in the AESB group was greater than in the placebo group at 60 days, but not significantly (11.11% vs. 3.70%, p = 0.270) (61). These promising preliminary results suggest that AESB treatment may contribute to prevent Hp-induced gastric cancer by reducing inflammation levels (mainly IL-8 and IFN- γ).

A RCT (62), additionally conducted in Iran in the framework of the previously mentioned RCT (59) with which shared the *Hp* eradication rate evaluation, examined the possible effect of high SFN broccoli sprout powder versus STT on serum levels of NO metabolites (NOx) as an indicator of systemic NO synthesis, in patients with *Hp* infection. There was a significant decrease in serum NOx levels in BSP (25.2 vs. 51.6 µmol/L) and STT + BSP (31.9 vs. 44.3, µmol/L) groups after intervention. The median, interquartile range, change percent of serum NOx levels were -3.7 (-29.4, 20.3), -42.7 (-57.1, -16.1), and -27.1 (-55.7, -0.92), in the STT, BSP, and STT + BSP groups, respectively. Data suggest that the use of high-SFN broccoli sprouts may attenuate undesirable overproduction of NO in *Hp*-infected patients.

In a Korean DB-RCT (58), 100 participants with functional dyspepsia and non-atrophic erythematous gastritis, without clarithromycin resistance, were randomly assigned in a double-blind manner. Participants underwent upper gastrointestinal endoscopies, UBT, and evaluations of malondialdehyde (MDA) derived from lipid peroxidation, and reduced glutathione (GSH) before and after the intervention. Follow-up assessments were performed within 1 week of taking the broccoli sprout extract supplements (BSES). 67 subjects out of 100 tested subjects were Hp positive and were randomly assigned to take BSES capsules (34 subjects) containing 250 mg (1,000 mg of sulforaphane) standardized broccoli sprout extract or placebo (33 subjects) for 4 weeks. 33 Hp negative subjects received BSES. Following losses to follow-up, 89 subjects were included, grouped as 33 BSES Hp positive (Group A), 28 placebo Hp positive (Group B) and 28 BSES Hp negative (Group C). In the Hp positive BSES group, no statistically significant differences were found in Δ UBT values before and after treatment. Only in one subject did the Δ UBT value reduce to less than 50% of the baseline level. Overall, at baseline, no statistically significant differences were detected in MDA and reduced GSH concentrations in the mucosa between the Hp positive and Hp negative group, but a significant decrease in MDA concentrations in the mucosa was observed in BSES Hp positive subjects compared to before treatment. Furthermore, a significant decrease in MDA concentrations was noted in all subjects treated with BSES, regardless of Hp infection status. At baseline, GSH concentrations in the 61 Hp positive subjects were lower, but not significantly, than those in the 28 Hp negative subjects.

3.3.3 Inhibition and/or control of Hp infection

In a trial conducted in Texas (57) the study population was comprised of volunteers with Hp infection. After a baseline UBT, volunteers received 135 grams of broccoli tips (equivalent to 135 µmol of glucoraphanin) added to ½ cup commercial plain yogurt, at breakfast, lunch and supper for servings (3.3 days). A final UBT was done 4 h after the last dose. This study also evaluated the effect of freshly made Tibetan yogurt whey in treating human Hp infection; volunteers consumed 120 mL of whey twice daily for 3.5 days. There was no significant difference in UBT results before and after a high dose of broccoli (mean UBT results before treatment = 15.8/mil, mean UBT results after treatment = 19.4/mil). Furthermore, there was no significant decrease in the UBT of participants who took Tibetan yogurt whey (UBT before treatment = 35.5/mil ±12.8, UBT after treatment = 40.7/mil±12.2).

In a French study (60), 22 xenografts, derived from mature human gastric epithelium and implanted in nude mice, were uniformly infected with Hp-26695 type. 2 weeks post-infection, mucus samples were collected from the xenografts and cultured on blood agar plates to evaluate colonization levels. Additionally, three biopsies were obtained to detect Hp and assess gastritis severity. The xenografted animals were subsequently randomized into two groups of 11 each. In the intervention group, a solution comprising 0.5 mL of sterile water with 0.5% acetonitrile containing 7.5 µmol of sulforaphane (approximately 1.33 mg) was administered via catheter once daily for 5 consecutive days. Conversely, the control group received an identical solution devoid of sulforaphane via the same route and schedule for 5 days. 1 month following the cessation of treatment, eradication of Hp was noted in 8 out of 11 grafts subjected to sulforaphane treatment. Conversely, the control group exhibited no variation in bacterial concentrations within the mucosa. These findings illustrate the potential for human gastric xenografts to effectively eradicate Hp following short-term administration of sulforaphane at a dose of 1.33 mg/day per xenograft.

In another RCT conducted in Michigan (54) eligible patients were identified based on evidence of active Hp infection confirmed through gastric biopsy or stool antigen testing. Through random assignment, 9 patients were allocated to receive doses of broccoli sprouts totalling 7, 14, or 28 grams administered on an empty stomach twice daily for 7 days. Stool antigen testing was conducted immediately post-treatment completion and on day 35 following the final sprout dose. Patients who remained Hp antigen-negative at day 35 underwent 14C-UBT. The PPI therapy was suspended for 2 weeks before testing. Following each treatment, patients completed a questionnaire regarding symptoms and the palatability of broccoli sprouts. The HpSAg test, conducted immediately following the completion of therapy (day 8), revealed that 7 out of 9 patients (78%) tested Hp negative. By day 35, 6 out of 9 patients (67%) remained negative, comprising 2 with negative UBT results, 2 with positive UBT results, and 2 with indeterminate UBT results. Immunohistochemical staining of gastric biopsies confirmed the absence of *Hp* in a patient with an indeterminate test result. Eradication of *Hp* was observed in one patient from each of the three dosage groups (14, 28, and 56 grams/day). Those who did not respond to the study regimen were offered standard antibiotic-based anti-Hp therapy. Results seem to suggest that consumption of oral broccoli sprouts was temporally associated with eradication of Hp infection in three of nine patients.

4 Discussion

The primary objective of this study was to gather empirical evidence regarding the consumption of Brassicaceae and the effects of its constituent sulforaphane, focusing particularly on its potential benefits in addressing human Hp infection. Our findings delve into the plausible synergistic interactions between metabolites derived from Brassicaceae, particularly sulforaphane, and antibiotic therapy in addressing Hp infection, as well as the role of these metabolites in mitigating Hp-induced inflammation and modifying the adverse effects associated with eradication antibiotic therapy.

4.1 Summary of findings

Our evidence shows that consumption of Brassicaceae appears to reduce Hp colonization (52, 53, 55, 56). Results on Hp eradication are controversial and highlight the variability in outcomes. A RCT conducted in Texas (57) suggests that broccoli itself, administered at least as part of a meal, is unlikely to be useful for the treatment of Hp infection. This finding aligns with existing literature (63) and is likely due to the lower sulforaphane content in broccoli compared to broccoli sprouts. An RCT conducted in Michigan (54), reported that broccoli sprout treatment led to Hp eradication in some patients, but without a clear dose-response relationship. Three RCTs have shown (51, 59) a higher, though not statistically significant, eradication rates, respectively combining Standard Triple Therapy with sulforaphane-compared to Standard Triple Therapy alone- or administering aqueous broccoli seed extract- instead of placebo (61). Beyond eradication rates, the potential protective effects of sulforaphane against Hp-induced gastric mucosal damage are noteworthy. A Korean RCT (58), found that consumption of broccoli extract supplements did not affect the density of Hp infection, but significantly prevented lipid peroxidation in the gastric mucosa and protected against Hp-induced gastritis. Indeed, the evidence (62) from our review revealed significant reductions in NOx levels in all intervention groups. Elevated NOx levels play a critical role in the development of gastric mucosal inflammation, gastritis, and gastric cancer (64, 65). The most pronounced reduction was observed in the BSP group that had received 6 g/day of broccoli sprout powder for 28 days alone or combined with standard triple therapy (62).

The literature robustly demonstrates that the production of ROS and iNOS, along with the resultant increase in NO levels due to Hp exposure, can lead to genetic alterations (66, 67). Notably, mutations in p53, induced by elevated NO levels associated with Hp infection, occur progressively as the gastric mucosa transitions from gastritis, through intestinal metaplasia and dysplasia, to GC (67, 68). Furthermore supporting this, a Japanese study (52, 53, 55, 56) found that treatment with broccoli sprouts led to a reduction in PGII levels, whose high levels are associated with gastric mucosal inflammation following Hp infection (69-71), along with a significant increase in the PGI/PGII ratio. Those findings highlight the potential protective role of sulforaphane against Hp-induced oxidative damage to the gastric mucosa, mitigating the inflammation that underlies carcinogenesis. In contrast, an RCT conducted in Iran (59) found no significant changes in serum levels of PGI, PGII, and the PGI/PGII ratio before and after treatment between the groups studied, potentially due to the low amount of broccoli sprout powder administered and the limited duration of treatment. Further affirmation of the potential anti-inflammatory efficacy of sulforaphane is evidenced in the RCT undertaken in China (61), which observed a notable decrease in pro-inflammatory cytokines associated with gastric mucosal injury, peptic ulceration, and gastric malignancy (21, 72, 73) after 2 months of daily AESB dosage. Our systematic review also indicates that the administration of sulforaphane derivatives or probiotics did not significantly alter the incidence of adverse events typically associated with antibiotic therapy, such as taste disorders, diarrhea, headache, epigastric pain, nausea, and urticaria (51). Moreover, the RCT conducted in Michigan (54), highlighted the unpredictable nature of sulforaphane derivatives' efficacy in ameliorating antibiotic therapy-related side effects, with not concord outcomes in symptom improvement among patients.

As delineated in preceding investigations (41), metabolic derivatives of Brassicaceae exhibit a notable bacteriostatic effect on Hp strains conserved in Brucella broth. Our data indicate that the administration of high-dose sulforaphane appears to be efficacious in eradicating Hp in short-term human gastric xenografts (60).

Despite the known anticarcinogenic effects of glucosinolates (74), it cannot be excluded that high-dose extracts of Brassicaceae may have undesirable effects, such as hepatocellular damage (75), irritation and increased apoptosis in gastric cells, especially at high concentrations (76). Furthermore, thiocyanates in large quantities formed by hydrolysis of glucosinolates by myrosinase can induce goiter (77). This highlights the continued need for studies to elucidate the various potential effects of these molecules in relation to their concentrations.

4.2 Limitations and implications for future research

It is imperative to acknowledge certain constraints inherent in this review. The heterogeneity of outcomes across studies may stem from suboptimal adherence to therapy, variations in the administration of PPIs not uniformly investigated, disparate dosages and timings of sulforaphane and its derivatives. Subsequent investigations should aim to identify an "optimal" dosage capable of mitigating infection, inflammation, and antibiotic-related side effects, since our data shows that too low a dosage could be responsible for a lack of effectiveness of the treatment. Concomitant research endeavors should explore alternative interventions, such as probiotics or other plant derivatives like green tea, garlic or liquorice, in demographically congruent populations. Given escalating antibiotic resistance, comprehensive analyses should encompass resistance profiles to employed antibiotics. Furthermore, evaluating the efficacy of sulforaphane and its derivatives in conjunction with quadruple therapy, including probing resistance to metronidazole, warrants attention.

It is imperative for forthcoming studies to incorporate representative sample cohorts to enhance generalizability. To this end, longitudinal methodologies facilitating the longitudinal observation of phenomena and multicentric approaches, fostering diversity in research contexts and participant demographics, are essential.

Despite strict adherence to PRISMA guidelines, potential selection biases could not be entirely mitigated. Regrettably, a quantitative synthesis was unattainable due to disparate outcome measures and assessment tools across included studies. Standardization of treatment protocols in future investigations would facilitate evaluative comparability and enhance interpretive coherence. Our results suggest that although Brassicaceae consumption may reduce Hp colonization, complete eradication of the infection is not consistently achieved in all patients or in all doses administered. However, Brassicaceae derivatives such as sulforaphane appear to have beneficial effects in controlling Hp infection. Further studies are needed to fully understand the therapeutic potential of these treatments, with prolonged administration of sulforaphane at different doses, possibly after phase I RCTs to identify the optimal dose, in representative samples using cross-sectional studies and/or multicentre approaches. These efforts would contribute to a deeper understanding of the therapeutic potential of sulforaphane in the management of Hp infection and maintenance of gastric health, potentially playing a prominent role in the prevention of gastric malignancies.

5 Conclusion

In conclusion, our review show that consumption of Brassicaceae and its component sulforaphane may have an impact on human Hpinfection through its bacteriostatic role and its inflammation-reducing effect. We have observed promising, but contrasting results regarding the synergistic interactions between Brassicaceae metabolites and antibiotic therapy in eradicating Hp, as well as their role in alleviating adverse effects associated with the eradication antibiotic therapy for Hp-induced conditions. Instead, several pieces of evidence point to potential protective effects against Hp-induced gastric mucosal damage and inflammation, which could be useful in counteracting the progression toward gastric cancer.

Further investigation are recommended to better elucidate a potential protective effect of Brassicaceae or plant extracts on *Hp*-associated gastric cancer risk and their addition to eradication treatment such as Standard Triple Therapy, or Quadruple Therapy, also considering antibiotic resistance patterns, as well as the influence of PPI intake.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author.

Author contributions

SP: Data curation, Formal analysis, Investigation, Methodology, Resources, Validation, Visualization, Writing – original draft, Writing – review & editing. FS: Conceptualization, Funding acquisition, Project administration, Resources, Supervision, Validation, Writing – original draft, Writing – review & editing. MR: Data curation, Investigation, Methodology, Validation, Writing – review & editing. CL: Supervision, Writing – review & editing. AV: Supervision, Writing – review & editing. AG: Conceptualization, Data curation, Formal analysis, Methodology, Supervision, Validation, Writing – original draft, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Rautelin H, Kosunen TU. Helicobacter pylori and associated gastroduodenal diseases. *APMIS.* (1991) 99:677–95. doi: 10.1111/j.1699-0463.1991.tb01246.x

2. Salama NR, Hartung ML, Müller A. Life in the human stomach: persistence strategies of the bacterial pathogen *Helicobacter Pylori*. *Nat Rev Microbiol*. (2013) 11:385–99. doi: 10.1038/nrmicro3016

3. Wroblewski LE, Peek RM Jr, Wilson KT. Helicobacter pylori and gastric cancer: factors that modulate disease risk. *Clin Microbiol Rev.* (2010) 23:713–39. doi: 10.1128/CMR.00011-10

4. Stefano K, Marco M, Federica G, Laura B, Barbara B, Gioacchino L, et al. *Helicobacter Pylori*, transmission routes and recurrence of infection: state of the art. *Acta Biomed*. (2018) 89:72–6. doi: 10.23750/abm.v89i8-S.7947

5. Malaty HM, El-Kasabany A, Graham DY, Miller CC, Reddy SG, Srinivasan SR, et al. Age at Acquisition of *Helicobacter Pylori* Infection: a follow-up study from infancy to adulthood. *Lancet*. (2002) 359:931–5. doi: 10.1016/S0140-6736(02)08025-X

6. Jung YS, Tran MTX, Song H, Park B, Moon CM. Association between age at *Helicobacter Pylori* eradication and the risk of gastric Cancer stratified by family history of gastric Cancer: a Nationwide population-based study. *Cancers*. (2023) 15:1604. doi: 10.3390/cancers15051604

7. Wu Q-J, Yang Y, Wang J, Han L-H, Xiang Y-B. Cruciferous vegetable consumption and gastric Cancer risk: a meta-analysis of epidemiological studies. *Cancer Sci.* (2013) 104:1067–73. doi: 10.1111/cas.12195

8. Li Y, Choi H, Leung K, Jiang F, Graham DY, Leung WK. Global prevalence of *Helicobacter Pylori* infection between 1980 and 2022: a systematic review and Metaanalysis. *Lancet Gastroenterol Hepatol.* (2023) 8:553–64. doi: 10.1016/ S2468-1253(23)00070-5

9. Malfertheiner P, Camargo MC, El-Omar E, Liou J-M, Peek R, Schulz C, et al. *Helicobacter Pylori* infection. *Nat Rev Dis Primers*. (2023) 9:1–24. doi: 10.1038/s41572-023-00431-8

10. Kusters JG, van Vliet AHM, Kuipers EJ. Pathogenesis of *Helicobacter Pylori* infection. *Clin Microbiol Rev.* (2006) 19:449–90. doi: 10.1128/CMR.00054-05

11. Conteduca V, Sansonno D, Lauletta G, Russi S, Ingravallo G, Dammacco FH. Pylori infection and gastric Cancer: state of the art (review). *Int J Oncol.* (2013) 42:5–18. doi: 10.3892/ijo.2012.1701

12. Du M-Q, Isaccson PG. Gastric MALT lymphoma: from Aetiology to treatment. Lancet Oncol. (2002) 3:97–104. doi: 10.1016/s1470-2045(02)00651-4

13. Robinson K, Atherton JC. The Spectrum of Helicobacter-mediated diseases. Annu Rev Pathol. (2021) 16:123–44. doi: 10.1146/annurev-pathol-032520-024949

14. Montecucco C, Rappuoli R. Living dangerously: how *Helicobacter Pylori* survives in the human stomach. *Nat Rev Mol Cell Biol.* (2001) 2:457–66. doi: 10.1038/35073084

15. Celli JP, Turner BS, Afdhal NH, Keates S, Ghiran I, Kelly CP, et al. *Helicobacter Pylori* moves through mucus by reducing mucin viscoelasticity. *Proc Natl Acad Sci USA*. (2009) 106:14321–6. doi: 10.1073/pnas.0903438106

16. Doohan D, Rezkitha YAA, Waskito LA, Yamaoka Y, Miftahussurur M. *Helicobacter Pylori* BabA–SabA key roles in the adherence phase: the synergic mechanism for successful colonization and disease development. *Toxins*. (2021) 13:485. doi: 10.3390/toxins13070485

17. Rossez Y, Gosset P, Boneca IG, Magalhães A, Ecobichon C, Reis CA, et al. The LacdiNAc-specific Adhesin LabA mediates adhesion of *Helicobacter Pylori* to human gastric mucosa. *J Infect Dis.* (2014) 210:1286–95. doi: 10.1093/infdis/jiu239

18. Foegeding NJ, Caston RR, McClain MS, Ohi MD, Cover TL. An overview of *Helicobacter Pylori* VacA toxin biology. *Toxins*. (2016) 8:173. doi: 10.3390/toxins8060173

19. Lawrence T. The nuclear factor NF-kappaB pathway in inflammation. *Cold Spring Harb Perspect Biol.* (2009) 1:a001651. doi: 10.1101/cshperspect.a001651

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20. He F, Ru X, Wen T. NRF2, a transcription factor for stress response and beyond. *Int J Mol Sci.* (2020) 21:4777. doi: 10.3390/ijms21134777

21. Camilo V, Sugiyama T, Touati E. Pathogenesis of *Helicobacter Pylori* infection. *Helicobacter*. (2017) 22:12405. doi: 10.1111/hel.12405

22. Butcher LD, den Hartog G, Ernst PB, Crowe SE. Oxidative stress resulting from *Helicobacter Pylori* infection contributes to gastric carcinogenesis. *Cell Mol Gastroenterol Hepatol.* (2017) 3:316–22. doi: 10.1016/j.jcmgh.2017.02.002

23. Naito Y, Yoshikawa T. Molecular and cellular mechanisms involved in *Helicobacter Pylori*-induced inflammation and oxidative stress. *Free Radic Biol Med.* (2002) 33:323–36. doi: 10.1016/s0891-5849(02)00868-7

24. Handa O, Naito Y, Yoshikawa T. Redox biology and gastric carcinogenesis: the role of *Helicobacter Pylori. Redox Rep.* (2011) 16:1–7. doi: 10.1179/17432921 1X12968219310756

25. Davies GR, Simmonds NJ, Stevens TR, Sheaff MT, Banatvala N, Laurenson IF, et al. *Helicobacter Pylori* stimulates antral mucosal reactive oxygen metabolite production in vivo. *Gut.* (1994) 35:179–85. doi: 10.1136/gut.35.2.179

26. Gong Y, Wei W, Jingwei L, Nannan D, Yuan Y. *Helicobacter Pylori* infection status correlates with serum parameter levels responding to multi-organ functions. *Dig Dis Sci.* (2015) 60:1748–54. doi: 10.1007/s10620-015-3522-2

27. Yuan L, Zhao J-B, Zhou Y-L, Qi Y-B, Guo Q-Y, Zhang H-H, et al. Type I and type II *Helicobacter Pylori* infection status and their impact on gastrin and pepsinogen level in a gastric Cancer prevalent area. *World J Gastroenterol.* (2020) 26:3673–85. doi: 10.3748/wjg.v26.i25.3673

28. Aumpan N, Mahachai V, Vilaichone R. Management of Helicobacter Pylori Infection. JGH Open. (2022) 7:3–15. doi: 10.1002/jgh3.12843

29. Sige-Sied. Trattamento dell'infezione da *helicobacter pylori* in italia. Rome: Italian Society of Digestive Endoscopy (2023).

30. Sidebotham RL, Worku ML, Karim QN, Dhir NK, Baron JH. How *Helicobacter Pylori* urease may affect external pH and influence growth and motility in the mucus environment: evidence from in-vitro studies. *Eur J Gastroenterol Hepatol.* (2003) 15:395–401. doi: 10.1097/00042737-200304000-00010

31. Park JB, Imamura L, Kobashi K. Kinetic studies of *Helicobacter Pylori* urease inhibition by a novel proton pump inhibitor, Rabeprazole. *Biol Pharm Bull.* (1996) 19:182–7. doi: 10.1248/bpb.19.182

32. Zhang J, Zhang C, Zhang Q, Yu L, Chen W, Xue Y, et al. Meta-analysis of the effects of proton pump inhibitors on the human gut microbiota. *BMC Microbiol.* (2023) 23:171. doi: 10.1186/s12866-023-02895-w

33. Takeuchi H, Trang VT, Morimoto N, Nishida Y, Matsumura Y, Sugiura T. Natural products and food components with anti-*Helicobacter Pylori* activities. *World J Gastroenterol.* (2014) 20:8971–8. doi: 10.3748/wjg.v20.i27.8971

34. Gravina AG, Pellegrino R, Auletta S, Palladino G, Brandimarte G, D'Onofrio R, et al. Hericium Erinaceus, a medicinal fungus with a centuries-old history: evidence in gastrointestinal diseases. *World J Gastroenterol.* (2023) 29:3048–65. doi: 10.3748/wjg. v29.i20.3048

35. Fahey JW, Zhang Y, Talalay P. Broccoli sprouts: an exceptionally rich source of inducers of enzymes that protect against chemical carcinogens. *Proc Natl Acad Sci USA*. (1997) 94:10367–72. doi: 10.1073/pnas.94.19.10367

36. Houghton CA. Sulforaphane: its "coming of age" as a clinically relevant nutraceutical in the prevention and treatment of chronic disease. Oxidative Med Cell Longev. (2019) 2019:2716870–27. doi: 10.1155/2019/2716870

37. Sturm C, Wagner AE. Brassica-derived plant bioactives as modulators of Chemopreventive and inflammatory signaling pathways. *Int J Mol Sci.* (1890) 18:18. doi: 10.3390/ijms18091890

38. Francis A, Lujan-Toro BE, Warwick SI, Macklin JA, Martin SL. Update on the Brassicaceae species checklist. *Biodivers Data J.* (2021) 9:e58773. doi: 10.3897/BDJ.9.e58773

39. Dinkova-Kostova AT, Fahey JW, Kostov RV, Kensler TW. KEAP1 and done? Targeting the NRF2 pathway with Sulforaphane. *Trends Food Sci Technol.* (2017) 69:257–69. doi: 10.1016/j.tifs.2017.02.002

40. Al-Ishaq RK, Overy AJ, Büsselberg D. Phytochemicals and gastrointestinal Cancer: cellular mechanisms and effects to change Cancer progression. *Biomol Ther.* (2020) 10:105. doi: 10.3390/biom10010105

41. Fahey JW, Haristoy X, Dolan PM, Kensler TW, Scholtus I, Stephenson KK, et al. Sulforaphane inhibits extracellular, intracellular, and antibiotic-resistant strains of Helicobacter pylori and prevents benzo[a]pyrene-induced stomach tumors. *Proc Natl Acad Sci USA*. (2002) 99:7610–5. doi: 10.1073/pnas.112203099

42. Coutinho LDL, Junior TCT, Rangel MC. Sulforaphane: an emergent anti-Cancer stem cell agent. *Front Oncol.* (2023) 13:1089115. doi: 10.3389/fonc.2023.1089115

43. Sailo BL, Liu L, Chauhan S, Girisa S, Hegde M, Liang L, et al. Harnessing Sulforaphane potential as a Chemosensitizing agent: a comprehensive review. *Cancers*. (2024) 16:244. doi: 10.3390/cancers16020244

44. Mlcek J, Jurikova T, Skrovankova S, Sochor J. Quercetin and its anti-allergic immune response. *Molecules*. (2016) 21:623. doi: 10.3390/molecules21050623

45. Carrasco-Pozo C, Castillo RL, Beltrán C, Miranda A, Fuentes J, Gotteland M. Molecular mechanisms of gastrointestinal protection by quercetin against indomethacininduced damage: role of NF-κB and Nrf2. J Nutr Biochem. (2016) 27:289–98. doi: 10.1016/j.jnutbio.2015.09.016

46. González-Segovia R, Quintanar JL, Salinas E, Ceballos-Salazar R, Aviles-Jiménez F, Torres-López J. Effect of the flavonoid quercetin on inflammation and lipid peroxidation induced by *Helicobacter Pylori* in gastric mucosa of Guinea pig. J Gastroenterol. (2008) 43:441–7. doi: 10.1007/s00535-008-2184-7

47. Wang Y-C. Medicinal plant activity on *Helicobacter Pylori* related diseases. *World J Gastroenterol.* (2014) 20:10368–82. doi: 10.3748/wjg.v20.i30.10368

48. EQUATOR Network (2021). Enhancing the quality and transparency of health research. Available at: https://www.equator-network.org/).

49. Ouzzani M, Hammady H, Fedorowicz Z, Elmagarmid A. Rayyan-a web and Mobile app for systematic reviews. *Syst Rev.* (2016) 5:210. doi: 10.1186/s13643-016-0384-4

50. CASP (2024). Checklists - critical appraisal skills programme. Available at: (https:// casp-uk.net/casp-tools-checklists/).

51. Chang YW, Park YM, Oh CH, Oh SJ, Cho J-H, Kim J-W, et al. Effects of probiotics or broccoli supplementation on *Helicobacter Pylori* eradication with standard clarithromycin-based triple therapy. *Korean J Intern Med.* (2020) 35:574–81. doi: 10.3904/kjim.2019.139

52. Yanaka A. Role of Sulforaphane in protection of gastrointestinal tract against H. Pylori and NSAID-induced oxidative stress. *Curr Pharm Des.* (2017) 23:4066–75. doi: 1 0.2174/1381612823666170207103943

53. Yanaka A. Contribution of NRF2 in gastrointestinal protection from oxidative injury. *Curr Pharm Des.* (2018) 24:2023–33. doi: 10.2174/138161282466618052210310 3

54. Galan MV, Kishan AA, Silverman AL. Oral broccoli sprouts for the treatment of *Helicobacter Pylori* infection: a preliminary report. *Dig Dis Sci.* (2004) 49:1088–90. doi: 10.1023/b:ddas.0000037792.04787.8a

55. Yanaka A. Sulforaphane enhances protection and repair of gastric mucosa against oxidative stress in vitro, and demonstrates anti-inflammatory effects on *Helicobacter Pylori*-infected gastric mucosae in mice and human subjects. *Curr Pharm Des.* (2011) 17:1532–40. doi: 10.2174/138161211796196945

56. Yanaka A, Fahey JW, Fukumoto A, Nakayama M, Inoue S, Zhang S, et al. Dietary Sulforaphane-rich broccoli sprouts reduce colonization and attenuate gastritis in *Helicobacter Pylori*-infected mice and humans. *Cancer Prev Res.* (2009) 2:353–60. doi: 10.1158/1940-6207.CAPR-08-0192

57. Opekun AR, Yeh CW, Opekun JL, Graham DY. In vivo tests of natural therapy, Tibetan yogurt or fresh broccoli, for *Helicobacter Pylori* infection. *Methods Find Exp Clin Pharmacol.* (2005) 27:327–9. doi: 10.1358/mf.2005.27.5.896760

58. Chang YW, Jang JY, Kim YH, Kim J-W, Shim J-J. The effects of broccoli sprout extract containing Sulforaphane on lipid peroxidation and *Helicobacter Pylori* infection in the gastric mucosa. *Gut Liver*. (2015) 9:486–93. doi: 10.5009/gnl14040

59. Bahadoran Z, Mirmiran P, Yeganeh MZ, Hosseinpanah F, Zojaji H, Azizi F. Complementary and alternative medicinal effects of broccoli sprouts powder on *Helicobacter Pylori* eradication rate in type 2 diabetic patients: a randomized clinical trial. *J Funct Foods*. (2014) 7:390–7. doi: 10.1016/j.jff.2014.01.020

60. Haristoy X, Angioi-Duprez K, Duprez A, Lozniewski A. Efficacy of Sulforaphane in eradicating *Helicobacter Pylori* in human gastric xenografts implanted in nude mice. *Antimicrob Agents Chemother*. (2003) 47:3982–4. doi: 10.1128/ AAC.47.12.3982-3984.2003

61. Guo K, Wang L, Mahe J, Li L, Jiao S, Wang H, et al. Effect of aqueous extract of seed of broccoli on inflammatory cytokines and *Helicobacter Pylori* infection: a randomized, double-blind, controlled trial in patients without atrophic gastritis. *Inflammopharmacology*. (2022) 30:1659–68. doi: 10.1007/s10787-022-01030-x

62. Mirmiran P, Bahadoran Z, Ghasemi A, Jeddi S, Azizi F. High-Sulforaphane broccoli sprout powder reduces serum nitric oxide metabolites in *Helicobacter Pylori* infected patients. *J Funct Foods*. (2017) 34:356–8. doi: 10.1016/j.jff.2017.05.025

63. Sato K, Kawakami N, Ohtsu T, Tsutsumi A, Miyazaki S, Masumoto T, et al. Broccoli consumption and chronic atrophic gastritis among Japanese males: an epidemiological investigation. *Acta Med Okayama*. (2004) 58:127–33. doi: 10.18926/ AMO/32109

64. Wilson K, Ramanujam K, Mobley H, Musselman R, James S, Meltzer S. *Helicobacter Pylori* stimulates inducible nitric oxide synthase expression and activity in a murine macrophage cell line. *Gastroenterology*. (1996) 111:1524–33. doi: 10.1016/S0016-5085(96)70014-8

65. Fu S, Ramanujam KS, Wong A, Fantry GT, Drachenberg CB, James SP, et al. Increased expression and cellular localization of inducible nitric oxide synthase and cyclooxygenase 2 in *Helicobacter Pylori* gastritis. *Gastroenterology*. (1999) 116:1319–29. doi: 10.1016/S0016-5085(99)70496-8

66. Nardone G. Review article: molecular basis of gastric carcinogenesis. *Aliment Pharmacol Ther.* (2003) 17:75–81. doi: 10.1046/j.1365-2036.17.s2.10.x

67. Shimizu T, Chiba T, Marusawa H. *Helicobacter Pylori*-mediated genetic instability and gastric carcinogenesis. *Curr Top Microbiol Immunol.* (2017) 400:305–23. doi: 10.1007/978-3-319-50520-6_13

68. Fenoglio-Preiser CM, Wang J, Stemmermann GN, Noffsinger A. TP53 and gastric carcinoma: a review. *Hum Mutat*. (2003) 21:258–70. doi: 10.1002/humu.10180

69. Kiyohira K, Yoshihara M, Ito M, Haruma K, Tanaka S, Chayama K. Serum pepsinogen concentration as a marker of *Helicobacter Pylori*nfection and the histologic grade of gastriits; evaluation of gastric mucosa by serum pepsinogen levels. *J Gastroenterol.* (2003) 38:332–8. doi: 10.1007/s005350300060

70. Haj- Sheykholeslami A, Rakhshani N, Amirzargar A, Rafiee R, Shahidi SM, Nikbin B, et al. Pepsinogen II, and gastrin 17 in relatives of gastric Cancer patients: comparative study with type and severity of gastritis. *Clin Gastroenterol Hepatol.* (2008) 6:174–9. doi: 10.1016/j.cgh.2007.11.016

71. He C, Sun L, Gong Y, Xu Q, Dong N, Yuan Y. Serum pepsinogen II: a neglected but useful biomarker to differentiate between diseased and Normal stomachs. *J Gastroenterol Hepatol.* (2011) 26:1039–46. doi: 10.1111/j.1440-1746.2011.06692.x

72. Mommersteeg MC, Simovic I, Yu B, van Nieuwenburg SAV, Bruno IMJ, Doukas M, et al. Autophagy mediates ER stress and inflammation in *Helicobacter Pylori*-related gastric Cancer. *Gut Microbes*. (2022) 14:2015238. doi: 10.1080/19490976.2021.2015238

73. Tang L, Tang B, Lei Y, Yang M, Wang S, Hu S, et al. *Helicobacter Pylori*-induced Heparanase promotes *H. pylori* colonization and gastritis. *Front Immunol.* (2021) 12:675747. doi: 10.3389/fimmu.2021.675747

74. Soundararajan P, Kim JS. Anti-carcinogenic Glucosinolates in cruciferous vegetables and their antagonistic effects on prevention of cancers. *Molecules*. (2018) 23:2983. doi: 10.3390/molecules23112983

75. Kupke F, Herz C, Hanschen FS, Platz S, Odongo GA, Helmig S, et al. Cytotoxic and genotoxic potential of food-borne nitriles in a liver in vitro model. *Sci Rep.* (2016) 6:37631. doi: 10.1038/srep37631

76. Santos PW, Machado ART, De Grandis R, Ribeiro DL, Tuttis K, Morselli M, et al. Effects of Sulforaphane on the oxidative response, apoptosis, and the transcriptional profile of human stomach mucosa cells *in vitro. Mutat Res Genet Toxicol Environ Mutagen.* (2020) 854-855:503201. doi: 10.1016/j.mrgentox.2020.503201

77. Felker P, Bunch R, Leung AM. Concentrations of thiocyanate and Goitrin in human plasma, their precursor concentrations in Brassica vegetables, and associated potential risk for hypothyroidism. *Nutr Rev.* (2016) 74:248–58. doi: 10.1093/ nutrit/nuv110