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Amyloid- β and heart failure in Alzheimer's disease: the new vistas

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Alzheimer's disease (AD) is the most common cause of dementia and represents 75% of all dementia types. AD neuropathology is due to the progressive deposition of extracellular amyloid-beta (A β) peptide and intracellular hyperphosphorylated tau protein. The accumulated Aß forms amyloid plaques, while the hyperphosphorylated tau protein forms neurofibrillary tangles (NFTs). Both amyloid plaques and NFTs are hallmarks of AD neuropathology. The fundamental mechanism involved in the pathogenesis of AD is still elusive, although A β is the more conceivable theory. A β induced neurodegeneration and associated neuroinflammation, oxidative stress, endoplasmic reticulum stress (ER), and mitochondrial dysfunction contribute to the development of cognitive impairment and dementia. Of note, A β is not only originated from the brain but also produced peripherally and, via the bloodbrain barrier (BBB), can accumulate in the brain and result in the development of AD. It has been shown that cardiometabolic conditions such as obesity, type 2 diabetes (T2D), and heart failure (HF) are regarded as possible risk factors for the development of AD and other types of dementia, such as vascular dementia. HFinduced chronic cerebral hypoperfusion, oxidative stress, and inflammation can induce the development and progression of AD. Interestingly, AD is regarded as a systemic disease that causes systemic inflammation and oxidative stress, which in turn affects peripheral organs, including the heart. A β through deranged BBB can be transported into the systemic circulation from the brain and accumulated in the heart, leading to the development of HF. These findings suggest a close relationship between AD and HF. However, the exact mechanism of AD-induced HF is not fully elucidated. Therefore, this review aims to discuss the link between AD and the risk of HF regarding the potential role of $A\beta$ in the pathogenesis of HF.

KEYWORDS

Alzheimer's disease, cardiovascular diseases, heart failure, pathogenesis, amyloid peptide

1 Introduction

Alzheimer's disease (AD) is the most common cause of dementia and represents 75% of all dementia types (1, 2). AD neuropathology is due to the progressive accumulation of extracellular amyloid-beta (A β) peptide and intracellular deposition of hyperphosphorylated tau protein (3). The accumulated A β forms amyloid plaque, while the hyperphosphorylated tau protein forms neurofibrillary tangles (NFTs) (4, 5). Both A β and NFTs trigger progressive neurodegeneration directly or through the initiation of oxidative stress and inflammation (6, 7).

A_β-induced neurodegeneration associated and neuroinflammation, oxidative stress, endoplasmic reticulum stress (ER), and mitochondrial dysfunction contribute to the development of cognitive impairment and dementia (8). Under physiological conditions, a low percentage of amyloid precursor protein (APP) is processed via β and γ secretases in the amyloidogenic pathway to produce oligomer A β , which is eliminated by many pathways into the systemic circulation, where it is metabolized by the liver and excreted by the kidney (9). In addition, A β is eliminated by cellular autophagy and degraded by enzymes such as neprilysin (NEP) and insulindegrading enzyme (IDE). Therefore, defective autophagy and downregulation of NEP and IDE are linked with the progression of AD neuropathology (10, 11). Most of the APP processing in young healthy subjects is by α -secretase in the non-amyloidogenic pathway to produce the neuroprotective soluble APP alpha (sAPP α) (9). In the aging process, hypoxia, and ischemia, APP processing is shifted toward the amyloidogenic pathway (12). AD is considered an agingrelated disorder due to the augmentation of oxidative stress, low-grade inflammatory reactions, and abnormal immune responses that affect APP processing and shift it toward the amyloidogenic pathway (12). Interestingly, the functional capacity of neuronal autophagy and the expression of NEP and IDE are extremely reduced by aging, leading to the impairment of A β elimination (10). Furthermore, an ischemic stroke increases the risk of AD and other types of dementia (13). In addition, genetic and environmental risk factors trigger the development of AD neuropathology through the induction of oxidative stress and inflammation (14).

AD occurs in two forms: early-onset familial and late-onset sporadic; genetic mutations in presenilin 1 (PS1), presenilin 2 (*PS2*), and *APP* genes cause early-onset familial AD, and a combination of lifestyle, environment, and genetic factors causes the late-onset sporadic form of the disease (15). However, accelerated disease progression is noticed in patients with familial AD. The early-onset familial AD represents 10% of all AD cases, although late-onset sporadic AD is the most common type and contributes to 90% of AD cases (16, 17). The main cause of early-onset familial AD is the overproduction of neurotoxic A β ; nevertheless, the chief cause of late-onset sporadic AD is impairment of A β (18, 19).

Thus, the pathophysiology of AD is multifarious and related to the activation of different signaling pathways (20) (Figure 1).

Research indicates that cardiometabolic conditions such as obesity, type 2 diabetes (T2D), and heart failure (HF) may serve as potential risk factors for the onset of Alzheimer's disease (AD) and



amyloid plaques, which induces the dissociation of tau protein and formation of hyperphosphorylated tau protein and formation of neurofibirlilary tangles (NFTs). $A\beta_{1,2}$ -induced activation of both astrocytes and microglia provokes the development of neuroinflammation. In addition, $A\beta_{1,2}$ leads to the impairment of neurotransmitter release and the development of synaptic failure. Furthermore, $A\beta_{1,2}$, by inducing oxidative stress, results in the development of mitochondrial dysfunction, which causes neuronal apoptosis.

other dementia types such as vascular dementia (21, 22). Of note, $A\beta$ is not only originated from the brain but also produced peripherally and, via the blood-brain barrier (BBB), can accumulate in the brain and result in the development of AD (23). HF-induced chronic cerebral hypoperfusion, oxidative stress, and inflammation can induce the development and progression of AD (12, 24). Remarkably, AD is considered a systemic disease causing systemic inflammation and oxidative stress, thereby affecting peripheral organs, including the heart (22). The development of HF due to the deposition of $A\beta$ is called cardiac amyloidosis (13, 25). Aβ through deranged BBB can be transported into the systemic circulation from the brain and accumulate in the heart, leading to the development of HF (25). These findings suggest a close relationship between AD and HF (Figure 2). However, the exact mechanism of AD-induced HF is not fully elucidated. Therefore, this review aims to discuss the link between AD and the risk of HF regarding the potential role of $A\beta$ in the pathogenesis of HF.

2 The pathophysiology of HF

HF is defined as the failure of the heart to support blood flow and the circulatory system during physical activity and rest (26). The universal definition of heart failure (HF) is a clinical syndrome characterized by symptoms and/or signs that are caused by a functional and structural abnormality of the heart, accompanied by elevated levels of natriuretic peptide and/or evidence of systemic and pulmonary congestion (27). The revised stages of HF define Stage A as at risk of HF, Stage B as pre-HF, Stage C as symptomatic HF, and Stage D as advanced HF (27). Generally, HF is classified as HF with reduced ejection fraction (HFrEF) and HF with preserved ejection fraction (HFpEF) (28).

However, the revised classification of HF proposed a new classification including HFrEF (symptomatic HF with LVEF \leq 40%), HF with mildly reduced ejection fraction (HFmrEF) with LVEF \geq 50%, and HF with improved ejection fraction (HFimpEF): symptomatic HF with a baseline LVEF \leq 40%, a \geq 10 point increase from baseline LVEF, and a second measurement of LVEF >40% (24). HFrEF, which typically develops subsequent to ischemic heart disease and is related to left ventricle hypertrophy, is most common in men and associated with pressure overload (29). However, HFpEF is typically developed in patients with type 2 diabetes (T2D), hypertension, obesity, and chronic kidney insufficiency (30). The pathophysiology of HFpEF differs from that of HFrEF as it is mostly attributed to coronary endothelial dysfunction and systemic inflammation (31). HFpEF is characterized by diastolic dysfunction and mild systolic dysfunction leading to increased left ventricle filling pressure (32). HFrEF, which is also called systolic HF, is mainly due to cardiomyocyte injury and defect in myocardial contractility (33).

The international prevalence of HF is in elevation due to the aging process and the obtainability of effective treatments. The prevalence of HF affects 1–2% of the general population and is expected to increase by approximately 64% from 2012 to 2030 with an equivalent increase in healthcare services by 127% (26, 34). HF is regarded as a pandemic disease affecting >64 million people globally (35). HF frequency increases with the progression of aging and reaches 10% over the age of 80 years (36). In 2022, HF will affect 64 million subjects globally with an incidence of 2% (37). The potential risk factors involved in the development and progression of HF are hypertension, ischemic heart disease, obesity, T2D, and smoking (38).

The pathophysiology of HF is related to the augmentation of preload (venous return), afterload (peripheral vascular resistance), and decreasing of cardiac contractility (26). Cardiac injury due to exaggerated preload and afterload and associated reduction of cardiac



output initiates a series of neurohormonal activation such as activation of the renin–angiotensin–aldosterone system (RAAS) and sympathoadrenal system (39). These alterations affect the cardiomyocytes to compensate for the failure status. Furthermore, increasing pressure overload provokes the development of cardiomyocyte hypertrophy, which causes cardiomyocyte ischemia and promotes the release of hypoxia-inducible factor 1 (HIF-1) (40). Importantly, HIF-1 promotes the expression of angiogenic growth factors and provokes angiogenesis (41). In the adaptive phase of HF, angiogenesis induces cardiac hypertrophy to preserve cardiac function (39–41). However, lack of angiogenesis prevents adaptive cardiac hypertrophy, leading to cardiac dysfunction (42) (Figure 3).

3 HF and risk of AD

The association between HF and cognitive impairment was initially proposed by Emerson et al. in 1981 as cardiogenic dementia (43). Previous epidemiological studies illustrated that up to 50% of HF patients developed some degree of cognitive decline, and 10% of them experienced severe cognitive decline (44–46). However, only 26% of managed HF patients developed vascular cognitive decline (47). Prominently, the biomarkers of cardiac dysfunction and myocardial injury, such as cardiac troponin and pro-N-terminal B-type natriuretic peptide (NT-proBNP), are correlated with lower cognitive function in old patients with HF (48–50). These findings underscore the close relationship between brain and cardiac functions.

Many studies have shown that HF may increase the risk of developing AD because it lowers blood flow to the brain and raises neurohormonal activity, which leads to neuronal energy crises and neurovascular unit dysfunction (44-46, 51, 52). Long-term low blood flow to the brain from HF starts the amyloidogenic pathway, which makes neurotoxic A (2) from APP (52). An experiment showed that long-term cerebral hypoperfusion raises the levels of β -secretase (BACE1) and sAPP (2) while lowering the levels of α -secretase and sAPP α in the hippocampus of rats (52). Babusikova et al. (53) discovered that experimental global brain ischemia starts the amyloidogenic pathway of APP processing, which leads to the buildup of neurotoxic A (2). Chronic cerebral hypoperfusion leads to oxidative stress, neuroinflammation, tau protein hyperphosphorylation, Aß buildup, synaptic dysfunction, and neurodegeneration (54). All of these factors can increase the risk of developing Alzheimer's disease, vascular dementia, or mixed dementia. Particularly, $A\beta$ is the key mediator for the development of cognitive impairment in agingassociated vascular pathologies (55). HF makes cognitive impairment worse because it causes chronic cerebral hypoperfusion, which damages the BBB and causes glial activation that cannot be controlled (56). Interestingly, dysregulation of the heart-brain axis seems a basic contributor to the development of cognitive impairment and dementia in HF (56).

Furthermore, exaggeration of the inflammatory and oxidative responses, injury to cerebral vasculatures, destruction of the BBB, and abnormal glial activation in HF contribute to the development and progression of AD (57–61). Research has demonstrated a link between the reduction of cardiac contractility and function, the dysregulation of the inflammatory milieu in the brain, and exaggerated sympathoexcitation (57). The HF mouse model consistently increases the expression of TLR4 and pro-inflammatory cytokines in the hippocampus and cortex (58). In addition, HF-induced oxidative stress is associated with the development of cognitive impairment in



FIGURE 3

Pathophysiology of HF: Pressure overload provokes cardiomyocyte hypertrophy, which causes cardiomyocyte ischemia and promotes the release of hypoxia-inducible factor 1 (HIF-1). HIF-1 stimulates the expression of angiogenesis and angiogenic growth factors. In the adaptive phase of HF, angiogenesis induces cardiac hypertrophy to preserve cardiac function. Failure of angiogenesis prevents adaptive cardiac hypertrophy, resulting in cardiac dysfunction.

a rat model (59). Oxidative stress is a key factor in the development of Alzheimer's disease, vascular dementia, and other neurodegenerative diseases. It does this by causing the formation of amyloid plaques and NFTs, which are characteristic of Alzheimer's disease (60). Furthermore, HF-induced cerebral vasculature injury is associated with cerebral vascular endothelial dysfunction and the development of cognitive impairment (60). In addition, the destruction of the BBB due to chronic cerebral hypoperfusion and brain ischemia in HF triggers AD neuropathology (61). When the BBB is damaged, neurotoxic substances such as thrombin and fibrinogen can easily enter the brain from the bloodstream. This can lead to neuroinflammation and Alzheimer's disease (62). Of interest, abnormal glial activation in HF promotes the release of pro-inflammatory cytokines the and development of neuroinflammation and AD (63, 64).

These verdicts indicated that the development of HF is influenced by abnormal inflammatory response, oxidative response, injury to cerebral vasculatures, destruction of BBB, and abnormal glial activation (Figure 4).

4 AD and risk of HF

AD has been regarded as a brain-specific disease though; for decades, researchers have observed a connection between various cardiovascular abnormalities and AD, such as HF, coronary artery disease, atrial fibrillation, and vasculopathy (65). A considerable volume of work has pointed to this head-to-heart connection, focusing mainly on connotations between chronic cerebral hypoperfusion and neuronal degradation. However, new evidence of a possible systemic or metastatic profile of AD calls for further analysis of this association. Aß aggregations are now known to be present in the hearts of individuals with idiopathic dilated cardiomyopathy as well as in the hearts of patients with AD (65-26). These findings suggest a potential systemic profile of proteinopathies and a new hypothesis for the link between peripheral and central symptoms of HF and AD. It has been shown that there is a close relationship between AD and HF. It has been shown that HF, atrial fibrillation, and hypertension through the induction of chronic cerebral hypoperfusion trigger the development of dementia and AD (65). In addition, AD increases the risk of HF and other cardiovascular disorders (65). AB, tau protein, and mutations of the PSN gene lead to arterial stiffness and diastolic dysfunction with subsequent development of HF and other cardiovascular disorders (Figure 5).

Many epidemiological studies illustrated that AD patients are associated with compromised cardiac function and early diastolic dysfunction due to the deposition of A β in the myocardium (66, 67). The population-based Rotterdam Study revealed that higher levels of A β 40 were associated with worse cardiac function and a higher risk of new-onset HF in the general population (68). Interestingly, A β 40 and A β 42 are accumulated in the cardiomyocytes and lead to HF in AD patients compared to healthy controls (67) suggesting that AD neuropathology affects the myocardium and is implicated in the development of HF. A retrospective study illustrated that myocardial





function was reduced in AD patients compared to healthy controls due to progressive accumulation of A β 40 in HF (67). In addition, cardiac index and function are low in patients with an aging brain (66). Numerous clinical studies highlighted that aortic stiffness, impairment of cardiac diastolic dysfunction, and reduction of left ventricular ejection fraction are common in AD patients compared to healthy controls (69, 70).

An echocardiographic study revealed that AD patients exhibit diastolic dysfunction, higher atrial conduction times, and increased arterial stiffness when compared to matched healthy controls (25). A case–control study found that people with AD had lower ejection fractions and cerebral blood flow velocities, as well as higher resistance and pulsatility indices in the basilar artery, the left terminal internal carotid artery, and the right terminal internal carotid artery. The aortic and carotid arteries also had more plaques than healthy controls (26). It has been shown that the prevalence of diastolic dysfunction and QRS abnormality in AD patients was 70 and 28%, respectively (65). AD and HFpEF are common age-related disorders that can coexist (65). In addition, AD patients have more left ventricular hypertrophy and different valvular heart disease compared to healthy controls (65).

Therefore, AD may be considered a systemic disease and not limited to the brain. The relationship between AD and various physical and systemic manifestations suggests that AD is a complex disease that impacts both the central nervous system and the peripheral nervous system. Remarkably, a common feature of many systemic processes linked to AD is involvement in energy metabolism due to genetics, mitochondria, and vascular mechanisms (71). For example, mitochondrial dysfunction in AD is not isolated to neurons but occurs systemically (72). Thus, abnormal systemic changes might not only develop secondary to brain dysfunction but might also affect AD progression, suggesting that the interactions between the brain and the periphery have a crucial role in the development and progression of AD (22).

Furthermore, HF is regarded as a proteinopathy disorder due to the accumulation of A β in the myocardium with subsequent membrane injury and dysregulation of intracellular Ca⁺² (73).

In addition, high circulating Aβ40 levels in AD patients are correlated with the development of ischemic heart disease, a risk factor for the development and progression of HF (74). The accumulated A β in the myocardium of AD is much lower than in the brain (74). Of note, exaggerated of AB level in AD not only accumulated in the brain but also in other organ such as the heart, skin, lungs, intestine, and kidney (75). Progressive accumulation of misfolded proteins such as Aβ, wild-type transthyretin (TTR), and PSN-1 results in the development of cardiomyopathy and HF in AD patients (76). Mutations in PSN-1 and PSN-2 genes are associated with the development of dilated cardiomyopathy (77). However, deletion of the PSN-1 gene is linked with the development of severe HF in mice (78) suggesting that the PSN-1 gene is essential for the development of the heart. Mutation of the PSN-1 gene increases the cleavage of APP and reduces $A\beta$ clearance resulting in the formation of amyloid aggregates and NFTs in both the brain and heart (79). Therefore, PSN genes are regarded as a potential link in the development of neurodegenerative diseases and cardiovascular disorders (Figure 6).

A β aggregation in AD is associated with progressive deposition of A β in the heart and other peripheral tissues due to the disturbance of APP processing in the peripheral tissues (75). In addition, BBB injury in AD promotes the permeability of A β from the brain into the systemic circulation and deposition in peripheral tissues, including the myocardium (80). It has been shown that plasma A β level is correlated with brain A β load in AD patients (81). A systematic review and meta-analysis found that high plasma A β levels in elderly subjects



predict the development of AD (82). Therefore, AD may increase the risk of HF by augmenting the plasma A β level, which is involved in the pathogenesis of cardiovascular injury and HF.

5 Peripheral Aβ and HF

It has been established that $A\beta$ is also produced from peripheral tissues and implicated in the pathogenesis of AD and HF (83, 84). Citron et al. (85) found that peripheral $A\beta$ is highly produced from skin fibroblasts of AD patients by decades before the development of symptomatic AD. Therefore, the overproduction of peripheral $A\beta$ is involved in the pathogenesis of AD before the development of central neural abnormalities. Mounting evidence from preclinical studies illustrated that AD neuropathology can be extended systematically, causing peripheral oxidative stress and inflammation (22). Supporting this claim, left ventricular hypertrophy and aortic stenosis are developing in transgenic mice with AD model (86).

Furthermore, APP processing can occur in different cells, such as platelets, leukocytes, skeletal muscles, and cardiomyocytes, resulting in the peripheral accumulation of A β (87). In addition, overexpression of tau protein is associated with the development of HF and cardiac hypertrophy (22). However, the CSF level of A β is higher than the plasma level due to the rapid clearance of peripheral A β by erythrocytes (88). Unlike A β 42, which is predominantly expressed in the brain, A β 40 is more prominent peripherally (89). Importantly, plasma A β level fluctuates due to the contribution of both brain and peripheral tissues to the production of plasma A β level (89). The difference in the distribution of A β isoforms is related to the type of distributed APP in the brain and peripheral tissues. Research has shown that the brain expresses APP₆₉₅, while leukocytes and platelets predominantly express APP_{751/770} (90). In addition, the difference in the expression of secretase enzyme (89) may explain the differences in the A β isoforms in the brain and periphery. The peripheral production of A β is augmented in HF due to the upregulation of β - and γ -secretase in the myocardium (91, 92). However, the expression of the cardioprotective α -secretase is reduced in coronary dysfunction and HF (93).

The coronary vasculature is crucial for normal heart function, especially the maturation of the coronary arterial endothelium. It has been shown that endothelial inactivation of α -secretase, a key regulator of Notch signaling, leads to defects in coronary arterial differentiation, as evidenced by dysregulated genes related to Notch signaling and arterial identity (93). Further analysis revealed that α -secretase knockout mice have enlarged dysfunctional hearts with abnormal myocardial compaction and increased the expression of venous and immature endothelium markers (93). These findings indicated a potential role for endothelial α -secretase in the cardioprotective homeostatic pathway.

It has been stated that the plasma A β 42 level is augmented in patients with ischemic heart disease. A cohort study found that the plasma A β 42 level was high in patients with non-ST elevation myocardial infarction (NSTEMI) compared to patients with STEMI (94). Therefore, the A β 42 plasma level is value in the risk stratification of patients with NSTEMI. Importantly, plasma sAPP is increased prior to the elevation of cardiac enzymes following experimental myocardial injury (95). Therefore, endothelial APP770 is regarded as an early biomarker of acute coronary syndrome (95).

The mechanism of $A\beta$ -induced myocardial injury is not fully elucidated, although it is related to the induction of coronary thrombosis and induction of platelet activation via the activation of the PKC signaling pathway and the production of thromboxane A2

(96, 97). In addition, Aβ40 triggers the release of MMP-9 from activated monocytes, which provokes the rupture of coronary atherosclerotic plaques (98). Consistently, high plasma A β is implicated in the development and progression of acute coronary syndrome by inducing thrombosis via factor XII activation, platelet activation, and monocyte activation (74, 97). Moreover, Aß reduces the function of cardiomyocytes by inducing oxidative stress and mitochondrial dysfunction (99). Furthermore, AB is released from activated platelets in response to inflammation and hypoxia (100). Platelet activation is augmented in HF due to an increment in inflammation and oxidative stress (101). Plasma AB40 level is correlated with worsening of cardiac contractility, and a high plasma A β 40 level predicts the development of HF (68). Plasma A β 40 level is correlated with cardiac enzymes during myocardial injury (102). Cardiac enzymes are elevated in patients with severe HF (103); therefore, a positive correlation between plasma Aβ40 level and cardiac enzymes indicates a severe form of HF. Interestingly, plasma Aβ40 is derived from peripheral tissues, while plasma Aβ42 is derived from the brain. However, the plasma $A\beta 42/A\beta 42$ ratio is not a reliable biomarker of HF or AD (104).

Of note, > 60% of A β is cleared from the brain into the systemic circulation (105). In addition, peripheral clearance of A β is highly reduced in aging due to the impairment of renal function, and increasing efflux of A β across the injured BBB contributes to augmenting plasma A β levels (106). Therefore, both brain and peripheral tissues contribute to the elevated plasma A β level, which either accumulate in the peripheral organs, such as the heart or seeds back into the brain across the injured BBB (22).

Importantly, P-glycoprotein and low-density lipoprotein receptorrelated protein 1 (LPR-1) mediate efflux of AB from the brain to the peripheral circulation (107). However, receptors for advanced glycation end product (RAGE) mediate the efflux of Aβ from the peripheral circulation to the brain (108). Moreover, the deregulation of P-glycoprotein in the myocardium reduces the clearance of Aβ40, resulting in cardiac dysfunction and the development of HF (109). Thus, impairment of the expression of P-glycoprotein and LPR-1 and overexpression of RAGE are involved in the pathogenesis of AD (107, 108, 110). It has been reported that LPR-1 plays a critical role in regulating inflammatory signaling in the myocardium and has a cardioprotective effect. Findings from preclinical studies demonstrated that LPR-1 agonists such as α1-antitrypsin reduce infarct size in acute myocardial infraction (111). Indeed, cardiac LPR-1 mediates the efflux of A β from the myocardium during ischemic-reperfusion injury (112). Many experimental studies highlighted that cardiac ischemic-reperfusion injury increases the expression of myocardium APP with subsequent overproduction of A β 40, which induces the disturbance of intracellular Ca²⁺. In addition, a reduction in the expression of LPR-1 reduces the elimination of Aβ40 from the myocardium, leading to progressive myocardial injury by the initiation of inflammatory signaling (113, 114). Furthermore, RAGE is upregulated in ischemic heart disease and involved in the pathogenesis of HF. RAGE enhances the accumulation of Aβ40 in the myocardium, causing myocardial dysfunction and the development of HF (115). Upregulated RAGE is implicated in the pathogenesis of HF and other cardiovascular disorders (116).

It has been illustrated that the peripheral clearance of $A\beta$ is by the liver, kidney, skin, and intestine, although the liver is involved in >60% of $A\beta$ clearance (117). Similarly, the kidney eliminates plasma $A\beta$ in a concentration-dependent manner (118). Furthermore, many blood

enzymes, such as angiotensin-converting enzyme (ACE), endothelinconverting enzyme 1 (ECE-1), IDE, and NEP, are intricate in the metabolism and clearance of peripheral A β (119). Moreover, the soluble NEP level which is involved in the metabolism of A β is increased and correlated with cardiac dysfunction (120). Nevertheless, NEP is linked to the metabolism of neuronal A β 42 rather than peripheral A β 40 (121). Therefore, NEP inhibitors have cardioprotective effects by increasing circulating natriuretic peptides (120).

Of note, the generated $A\beta$ from APP in the plasma is not free in the body fluids but binds different binding proteins such as albumin, antithrombin III, and lipoproteins before transport and catabolism by peripheral organs (122). In addition, peripheral cells such as erythrocytes, monocytes, macrophages, neutrophils, and lymphocytes promote the clearance of peripheral A β (123, 124). It has been shown that impairment of peripheral $A\beta$ clearance is implicated in the pathogenesis of HF (125). Increasing plasma A\beta level is correlated with the severity of cardiac stress in HF patients with normal cognitive function. As well, targeting peripheral Aß by the monoclonal antibody aducanumab may reduce the severity of myocardial dysfunction in patients with HF (125). In HF, most of the peripheral clearance mechanisms of AB are impaired. For example, a reduction in serum albumin, which binds 89% of plasma A_β, is correlated with severity and mortality in patients with HF (126). Moreover, hepato-renal dysfunction is common in patients with HF (127).

These verdicts indicated that overproduction and reduced clearance of peripheral A β contribute to the development and progression of HF. Together, exaggerated A β production in Alzheimer's disease (AD) and its transport across the damaged bloodbrain barrier (BBB) augment circulating A β , which is implicated in the pathogenesis of heart failure (HF).

6 Conclusion and future perspectives

HF and cardiometabolic disorders are implicated in the development and progression of AD, vascular dementia, and other neurodegenerative diseases. In addition, AD increases the pathogenesis of HF and other cardiovascular diseases, signifying a close relationship between AD and HF.

HF-induced AD is mediated by inducing chronic cerebral hypoperfusion, oxidative stress, and inflammation that promote AD neuropathology. However, AD-induced HF is mediated by the overproduction and impairment of Aß elimination. Impairment of cardiac diastolic dysfunction and reduction of left ventricular ejection fraction are common in AD patients. Interestingly, Aβ40 and Aβ42 are accumulated in the cardiomyocytes and lead to HF in AD, suggesting that AD neuropathology affects the myocardium and is implicated in the development of HF. In addition, myocardial function was reduced in AD patients due to progressive accumulation of A β 40. Furthermore, high circulating Aβ40 levels in AD patients is correlated with the development of ischemic heart disease, a risk factor for the development and progression of HF. Moreover, Aß is also produced from peripheral tissues and implicated in the pathogenesis of AD and HF. Mounting evidence illustrates that AD neuropathology can be extended systematically, causing peripheral oxidative stress and inflammation. Furthermore, APP processing can occur in different cells, resulting in the peripheral accumulation of Aβ.

The peripheral production of $A\beta$ is augmented in HF due to the upregulation of β and γ -secretase in the myocardium. However, the expression of the cardioprotective α -secretase is reduced in coronary dysfunction and HF. Moreover, plasma A β 42 levels are increased in patients with ischemic heart disease. A β reduces the function of cardiomyocytes by inducing oxidative stress and mitochondrial dysfunction. Plasma A β 40 level is correlated with the worsening of cardiac contractility, and a high plasma A β 40 level predicts the development of HF.

Furthermore, peripheral clearance of $A\beta$ is highly reduced in aging due to the impairment of renal function together with increasing efflux of $A\beta$ across the injured BBB contributes to augmenting plasma $A\beta$ level. Therefore, both brain and peripheral tissues contribute in elevating plasma $A\beta$ level which either accumulated in the peripheral organs such as heart or seeds back into the brain across the injured BBB.

Indeed, cardiac LPR-1 mediates the efflux of A β from the myocardium during ischemic-reperfusion injury. The expression of LPR-1 reduces the elimination of A β 40 from the myocardium, leading to progressive myocardial injury by the initiation of inflammatory signaling. However, RAGE is upregulated in ischemic heart disease and is involved in the pathogenesis of HF. RAGE enhances the accumulation of A β 40 in the myocardium, causing myocardial dysfunction and the development of HF.

Interestingly, medications used in the management of HF may affect cognitive function and AD development. Sodium-glucose co-transporter-2 (SGLT2) inhibitors, originally developed for diabetes management, are increasingly studied for their cognitive benefits and in the management of HF. The multifaceted effects and the relatively favorable side effect profile of SGLT2 inhibitors render them a promising therapeutic candidate for HF patients with cognitive impairments (128-130). Interestingly, the use of SGLT2 inhibitors in elderly patients with T2D and HFpEF improves cognitive function (131). Consistently, a prospective study revealed that empagliflozin improves cognitive and physical impairment in frail older adults with T2D and HFpEF (132). A nationwide study from Sewed found that SGLT2 inhibitors are associated with lower neurocognitive disorders in patients with HF (133). Moreover, the use of SGLT2 inhibitors has a neuroprotective effect in T2D patients with HF, reducing the incidence or progression of cognitive impairment and dementia (134). The neuroprotective effects of SGLT2 inhibitors are mediated by reducing oxidative stress and neuroinflammation, decreasing amyloid burdens, enhancing neuronal plasticity, and improving cerebral glucose utilization (131, 134). Thus, SGLT2 inhibitors, by targeting $A\beta$ and associated neuroinflammation and oxidative stress, can mitigate both HF and AD and reduce the incidence of HF in elderly patients with AD

Taken together, these findings point out that overproduction and reduced clearance of peripheral A β contributes to the development and progression of HF. In addition, the exaggeration of A β production in AD and transport across the damaged BBB augments circulating the A β level, which is implicated in the pathogenesis of HF. Therefore,

References

1. Al-Kuraishy HM, Jabir MS, Albuhadily AK, Al-Gareeb AI, Rafeeq MF. The link between Alzheimer disease and metabolic syndrome: a mutual relationship and long rigorous investigation. *Ageing Res Rev.* (2023) 91:102084. doi: 10.1016/j.arr.2023.102084

AD is regarded as a potential risk factor for the development of HF, although the fundamental mechanisms need to be verified by future studies.

Accordingly, targeting peripheral A β by inhibiting its production or increasing its elimination may reduce the risk of HF development in AD patients. In addition, the restoration of BBB integrity by novel agents may reduce the transport of A β from the brain into the systemic circulation. It is interesting that RAGE inhibitors and LPR-1 activators might help stop the movement of A β from the brain into the bloodstream and help control HF. Therefore, we recommend further preclinical and clinical studies in this area.

Author contributions

HA-K: Conceptualization, Writing – original draft, Writing – review & editing. GS: Conceptualization, Writing – original draft, Writing – review & editing. HM: Software, Writing – review & editing. SM: Writing – review & editing. AA-G: Writing – original draft, Writing – review & editing. AKA: Writing – original draft, Writing – review & editing. RD: Writing – review & editing. AAA: Writing – review & editing. MA-A: Writing – review & editing.

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Conflict of interest

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2. Dhahri M, Alghrably M, Mohammed HA, Badshah SL, Noreen N, Mouffouk F, et al. Natural polysaccharides as preventive and therapeutic horizon for neurodegenerative diseases. *Pharmaceutics*. (2021) 14:1. doi: 10.3390/pharmaceutics14010001 3. Association, A. 2012 Alzheimer's disease facts and figures. *Alzheimers Dement*. (2012) 8:131-68. doi: 10.1016/j.jalz.2012.02.001

4. Ashrafian H, Zadeh EH, Khan RH. Review on Alzheimer's disease: inhibition of amyloid beta and tau tangle formation. *Int J Biol Macromol.* (2021) 167:382–94. doi: 10.1016/j.ijbiomac.2020.11.192

5. Zhang H, Wei W, Zhao M, Ma L, Jiang X, Pei H, et al. Interaction between A β and tau in the pathogenesis of Alzheimer's disease. *Int J Biol Sci.* (2021) 17:2181–92. doi: 10.7150/ijbs.57078

6. Perluigi M, Di Domenico F, Butterfield DA. Oxidative damage in neurodegeneration: roles in the pathogenesis and progression of Alzheimer disease. *Physiol Rev.* (2024) 104:103–97. doi: 10.1152/physrev.00030.2022

 Abubakar MB, Sanusi KO, Ugusman A, Mohamed W, Kamal H, Ibrahim NH, et al. Alzheimer's disease: an update and insights into pathophysiology. *Front Aging Neurosci.* (2022) 14:742408. doi: 10.3389/fnagi.2022.742408

8. al-kuraishy H, Jabir M, Albuhadily A, al-Gareeb A, Jawad S, Swelum A, et al. Role of ketogenic diet in neurodegenerative diseases focusing on Alzheimer diseases: the guardian angle. *Ageing Res Rev.* (2024) 95:102233. doi: 10.1016/j.arr.2024.102233

9. al-kuraishy H, Jabir M, al-Gareeb A, Albuhadily A, Albukhaty S, Sulaiman G, et al. Evaluation and targeting of amyloid precursor protein (APP)/amyloid beta (A β) axis in amyloidogenic and non-amyloidogenic pathways: a time outside the tunnel. *Ageing Res Rev.* (2023) 92:102119. doi: 10.1016/j.arr.2023.102119

10. al-Kuraishy HM, Jabir MS, Sulaiman GM, Mohammed HA, al-Gareeb AI, Albuhadily AK, et al. The role of statins in amyotrophic lateral sclerosis: protective or not? *Front Neurosci.* (2024) 18:1422912. doi: 10.3389/fnins.2024.1422912

11. Ali NH, al-kuraishy HM, al-Gareeb AI, Alnaaim SA, Alexiou A, Papadakis M, et al. The probable role of tissue plasminogen activator/neuroserpin axis in Alzheimer's disease: a new perspective. *Acta Neurol Belg.* (2024) 124:377–88. doi: 10.1007/ s13760-023-02403-x

12. Yue X, Zhou Y, Qiao M, Zhao X, Huang X, Zhao T, et al. Intermittent hypoxia treatment alleviates memory impairment in the 6-month-old APPswe/PS1dE9 mice and reduces amyloid beta accumulation and inflammation in the brain. *Alzheimers Res Ther.* (2021) 13:1–16. doi: 10.1186/s13195-021-00935-z

13. Elman-Shina K, Efrati S. Ischemia as a common trigger for Alzheimer's disease. *Front Aging Neurosci.* (2022) 14:1012779. doi: 10.3389/fnagi.2022.1012779

14. Zhang X-X, Tian Y, Wang ZT, Ma YH, Tan L, Yu JT. The epidemiology of Alzheimer's disease modifiable risk factors and prevention. *J Prev Alzheimers Dis.* (2021) 8:313–21. doi: 10.14283/jpad.2021.15

15. Alexiou A, Kamal MA, Ashraf GM. The Alzheimer's disease challenge. Front Neurosci. (2019) 13:768 at. doi: 10.3389/fnins.2019.00768

16. Chatterjee S, Mudher A. Alzheimer's disease and type 2 diabetes: a critical assessment of the shared pathological traits. *Front Neurosci.* (2018) 12:383. doi: 10.3389/ fnins.2018.00383

17. Spina S, la Joie R, Petersen C, Nolan AL, Cuevas D, Cosme C, et al. Comorbid neuropathological diagnoses in early versus late-onset Alzheimer's disease. *Brain.* (2021) 144:2186–98. doi: 10.1093/brain/awab099

18. Gauvrit T, Benderradji H, Buée L, Blum D, Vieau D. Early-life environment influence on late-onset Alzheimer's disease. *Front Cell Dev Biol.* (2022) 10:834661. doi: 10.3389/fcell.2022.834661

19. du Y, Zhang S, Fang Y, Qiu Q, Zhao L, Wei W, et al. Radiomic features of the hippocampus for diagnosing early-onset and late-onset Alzheimer's disease. *Front Aging Neurosci.* (2022) 13:789099. doi: 10.3389/fnagi.2021.789099

20. Das TK, Ganesh BP, Fatima-Shad K. Common signaling pathways involved in Alzheimer's disease and stroke: two faces of the same coin. *J Alzheimers Dis Rep.* (2023) 7:381–98. doi: 10.3233/ADR-220108

21. Ryder REJ, Laubner K, Benes M, Haluzik M, Munro L, Frydenberg H, et al. Endoscopic duodenal-jejunal bypass liner treatment for type 2 diabetes and obesity: glycemic and cardiovascular disease risk factor improvements in 1,022 patients treated worldwide. *Diabetes Care*. (2023) 46:e89–91. doi: 10.2337/dc22-1952

22. Wang J, Gu BJ, Masters CL, Wang Y-J. A systemic view of Alzheimer disease—insights from amyloid- β metabolism beyond the brain. *Nat Rev Neurol.* (2017) 13:612–23. doi: 10.1038/nrneurol.2017.111

23. Cheng Y, Tian D-Y, Wang Y-J. Peripheral clearance of brain-derived A β in Alzheimer's disease: pathophysiology and therapeutic perspectives. *Transl Neurodegener*. (2020) 9:1–11. doi: 10.1186/s40035-020-00195-1

24. Tini G, Cannatà A, Canepa M, Masci PG, Pardini M, Giacca M, et al. Is heart failure with preserved ejection fraction a 'dementia' the heart? *Heart Fail Rev.* (2022) 27:587–94. doi: 10.1007/s10741-021-10114-9

25. Rubin J, Maurer MS. Cardiac amyloidosis: overlooked, underappreciated, and treatable. Annu Rev Med. (2020) 71:203–19. doi: 10.1146/annurev-med-052918-020140

26. Schwinger RHG. Pathophysiology of heart failure. *Cardiovasc Diagn Ther*. (2021) 11:263–76. doi: 10.21037/cdt-20-302

27. Bozkurt B, Coats AJS, Tsutsui H, Abdelhamid CM, Adamopoulos S, Albert N, et al. Universal definition and classification of heart failure: a report of the heart failure society of America, heart failure association of the European society of cardiology, Japanese heart failure society and writing committee of the universal definition of heart failure. *J Card Fail.* (2021) 27:387–413. doi: 10.1002/ejhf.2115

28. Lam CSP, Solomon SD. Classification of heart failure according to ejection fraction: JACC review topic of the week. *J Am Coll Cardiol*. (2021) 77:3217–25. doi: 10.1016/j.jacc.2021.04.070

29. Swaraj S, Kozor R, Arnott C, Di Bartolo BA, Figtree AG. Heart failure with reduced ejection fraction—does sex matter? Curr. *Heart Fail Rep.* (2021) 18:345–52. doi: 10.1007/s11897-021-00533-y

30. Abudureyimu M, Luo X, Wang X, Sowers JR, Wang W, Ge J, et al. Heart failure with preserved ejection fraction (HFpEF) in type 2 diabetes mellitus: from pathophysiology to therapeutics. *J Mol Cell Biol.* (2022) 14:mjac028. doi: 10.1093/jmcb/mjac028

31. Youn J-C, Ahn Y, Jung HO. Pathophysiology of heart failure with preserved ejection fraction. *Heart Fail Clin.* (2021) 17:327–35. doi: 10.1016/j.hfc.2021.02.001

32. Heinzel FR, Hegemann N, Hohendanner F, Primessnig U, Grune J, Blaschke F, et al. Left ventricular dysfunction in heart failure with preserved ejection fraction—molecular mechanisms and impact on right ventricular function. *Cardiovasc Diagn Ther.* (2020) 10:1541–60. doi: 10.21037/cdt-20-477

33. Simmonds SJ, Cuijpers I, Heymans S, Jones EAV. Cellular and molecular differences between HFpEF and HFrEF: a step ahead in an improved pathological understanding. *Cells.* (2020) 9:242. doi: 10.3390/cells9010242

34. Virani SS, Alonso A, Aparicio HJ, Benjamin EJ, Bittencourt MS, Callaway CW, et al. On behalf of the American Heart Association Council on epidemiology and prevention statistics committee and stroke statistics subcommittee heart disease and stroke statistics-2021 update: a report from the American Heart Association. *Circulation*. (2021) 143:e254–743. doi: 10.1161/CIR.0000000000000050

35. GBD 2017 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: A systematic analysis for the global burden of disease study 2017. *Lancet.* (2018) 392:1789–858. doi: 10.1016/S0140-6736(18)32279-7

36. Mosterd A, Hoes AW. Clinical epidemiology of heart failure. *Heart*. (2007) 93:1137-46. doi: 10.1136/hrt.2003.025270

37. Tsao CW, Aday AW, Almarzooq ZI, Alonso A, Beaton AZ, Bittencourt MS, et al. Heart disease and stroke statistics—2022 update: a report from the American Heart Association. *Circulation*. (2022) 145:e153–639. doi: 10.1161/CIR.000000000001052

38. Bui AL, Horwich TB, Fonarow GC. Epidemiology and risk profile of heart failure. *Nat Rev Cardiol.* (2011) 8:30–41. doi: 10.1038/nrcardio.2010.165

39. Dandel M. Load dependency of ventricular pump function: impact on the noninvasive evaluation of the severity and the prognostic relevance of myocardial dysfunction. *Rev Cardiovasc Med.* (2024) 25:272. doi: 10.31083/j.rcm2508272

40. Sant'Ana PG, Tomasi LC, Murata GM, Vileigas DF, Mota GAF, et al. Hypoxiainducible factor 1-alpha and glucose metabolism during cardiac remodeling progression from hypertrophy to heart failure. *Int J Mol Sci.* (2023) 24:6201. doi: 10.3390/ ijms24076201

41. Syukri A, Budu B, Hatta M, Amir M, Rohman MS, Mappangara I, et al. Doxorubicin induced immune abnormalities and inflammatory responses via HMGB1, HIF1- α and VEGF pathway in progressive of cardiovascular damage. *Ann Med Surg.* (2022) 76:501. doi: 10.1016/j.amsu.2022.103501

42. Carter K, Shah E, Waite J, Rana D, Zhao Z-Q. Pathophysiology of angiotensin IImediated hypertension, cardiac hypertrophy, and failure: a perspective from macrophages. *Cells*. (2024) 13:2001. doi: 10.3390/cells13232001

43. Emerson TR, Milne JR, Gardner AJ. Cardiogenic dementia—a myth? *Lancet*. (1981) 318:743-4. doi: 10.1016/S0140-6736(81)91063-1

44. Pressler SJ. Cognitive functioning and chronic heart failure: a review of the literature (2002-July 2007). *J Cardiovasc Nurs.* (2008) 23:239–49. doi: 10.1097/01. JCN.0000305096.09710.ec

45. Vogels RLC, Oosterman JM, van Harten B, Scheltens P, van der Flier WM, Schroeder-Tanka JM, et al. Profile of cognitive impairment in chronic heart failure. *J Am Geriatr Soc.* (2007) 55:1764–70. doi: 10.1111/j.1532-5415.2007.01395.x

46. Huijts M, van Oostenbrugge R, Duits A, Burkard T, Muzzarelli S, Maeder MT, et al. Cognitive impairment in heart failure: results from the trial of intensified versus standard medical therapy in elderly patients with congestive heart failure (TIME-CHF) randomized trial. *Eur J Heart Fail.* (2013) 15:699–707. doi: 10.1093/eurjhf/hft020

47. Zuccalà G, onder G, Pedone C, Carosella L, Pahor M, Bernabei R, et al. Hypotension and cognitive impairment. *Neurology*. (2001) 57:1986–92. doi: 10.1212/WNL.57.11.1986

48. Mirza SS, de Bruijn RFAG, Koudstaal PJ, van den Meiracker AH, Franco OH, Hofman A, et al. The N-terminal pro B-type natriuretic peptide, and risk of dementia and cognitive decline: a 10-year follow-up study in the general population. *J Neurol Neurosurg Psychiatry*. (2016) 87:356–62. doi: 10.1136/jnnp-2014-309968

49. Sabayan B, van Buchem MA, de Craen AJM, Sigurdsson S, Zhang Q, Harris TB, et al. N-terminal pro-brain natriuretic peptide and abnormal brain aging: the AGES-Reykjavik study. *Neurology*. (2015) 85:813–20. doi: 10.1212/WNL.000000000001885

50. van Peet PG, de Craen AJM, Gussekloo J, de Ruijter W. Plasma NT-proBNP as predictor of change in functional status, cardiovascular morbidity and mortality in the oldest old: the Leiden 85-plus study. *Age (Dordr)*. (2014) 36:9660. doi: 10.1007/s11357-014-9660-1

51. Cermakova P, Eriksdotter M, Lund LH, Winblad B, Religa P, Religa D. Heart failure and Alzheimer's disease. J Intern Med. (2015) 277:406–25. doi: 10.1111/joim.12287

52. Cai Z, Liu Z, Xiao M, Wang C, Tian F. Chronic cerebral Hypoperfusion promotes amyloid-Beta pathogenesis via activating β/γ -secretases. *Neurochem Res.* (2017) 42:3446–55. doi: 10.1007/s11064-017-2391-9

53. Babusikova E, Dobrota D, Turner AJ, Nalivaeva NN. Effect of global brain ischemia on amyloid precursor protein metabolism and expression of amyloid-degrading enzymes in rat cortex: role in pathogenesis of Alzheimer's disease. *Biochemistry (Mosc)*. (2021) 86:680–92. doi: 10.1134/S0006297921060067

54. Zhao Y, Gong C-X. From chronic cerebral hypoperfusion to Alzheimer-like brain pathology and neurodegeneration. *Cell Mol Neurobiol.* (2015) 35:101–10. doi: 10.1007/s10571-014-0127-9

55. Khan F, Qiu H. Amyloid-β: a potential mediator of aging-related vascular pathologies. *Vasc Pharmacol.* (2023) 152:107213. doi: 10.1016/j.vph.2023.107213

56. Xu C, Tao X, Ma X, Zhao R, Cao Z. Cognitive dysfunction after heart disease: a manifestation of the heart-brain Axis. *Oxidative Med Cell Longev*. (2021) 2021:4899688. doi: 10.1155/2021/4899688

57. Toledo C, Andrade DC, Díaz HS, Inestrosa NC, Del Rio R. Neurocognitive disorders in heart failure: novel pathophysiological mechanisms underpinning memory loss and learning impairment. *Mol Neurobiol.* (2019) 56:8035–51. doi: 10.1007/s12035-019-01655-0

58. Hong X, Bu L, Wang Y, Xu J, Wu J, Huang Y, et al. Increases in the risk of cognitive impairment and alterations of cerebral β -amyloid metabolism in mouse model of heart failure. *PLoS One.* (2013) 8:e63829. doi: 10.1371/journal.pone.0063829

59. Fu X, Zhang J, Guo L, Xu Y, Sun L, Wang S, et al. Protective role of luteolin against cognitive dysfunction induced by chronic cerebral hypoperfusion in rats. *Pharmacol Biochem Behav.* (2014) 126:122–30. doi: 10.1016/j.pbb.2014.09.005

60. Ovsenik A, Podbregar M, Fabjan A. Cerebral blood flow impairment and cognitive decline in heart failure. *Brain Behav.* (2021) 11:e02176. doi: 10.1002/brb3.2176

61. Yang M, Sun D, Wang Y, Yan M, Zheng J, Ren J. Cognitive impairment in heart failure: landscape, challenges, and future directions. *Front Cardiovasc Med.* (2021) 8:831734. doi: 10.3389/fcvm.2021.831734

62. McLarnon JG. A leaky blood-brain barrier to fibrinogen contributes to oxidative damage in Alzheimer's disease. *Antioxidants (Basel, Switzerland)*. (2021) 11:102. doi: 10.3390/antiox11010102

63. Li Y-L. Stellate ganglia and cardiac sympathetic Overactivation in heart failure. Int J Mol Sci. (2022) 23:311. doi: 10.3390/ijms232113311

64. Uddin MS, Lim LW. Glial cells in Alzheimer's disease: from neuropathological changes to therapeutic implications. *Ageing Res Rev.* (2022) 78:101622. doi: 10.1016/j. arr.2022.101622

65. Schaich CL, Maurer MS, Nadkarni NK. Amyloidosis of the brain and heart: two sides of the same coin? JACC. *Heart Failure*. (2019) 7:129–131 at. doi: 10.1016/j. jchf.2018.12.014

66. Jefferson AL, Himali JJ, Beiser AS, Au R, Massaro JM, Seshadri S, et al. Cardiac index is associated with brain aging: the Framingham heart study. *Circulation*. (2010) 122:690–7. doi: 10.1161/CIRCULATIONAHA.109.905091

67. Troncone L, Luciani M, Coggins M, Wilker EH, Ho CY, Codispoti KE, et al. Aβ amyloid pathology affects the hearts of patients with Alzheimer's disease: mind the heart. J Am Coll Cardiol. (2016) 68:2395–407. doi: 10.1016/j.jacc.2016.08.073

68. Zhu F, Wolters FJ, Yaqub A, MJG L, Ghanbari M, Boersma E, et al. Plasma amyloid-β in relation to cardiac function and risk of heart failure in general population. *JACC Heart Fail*. (2023) 11:93–102. doi: 10.1016/j.jchf.2022.09.006

69. Çalık AN, Özcan KS, Yüksel G, Güngör B, Aruğarslan E, Varlibas F, et al. Altered diastolic function and aortic stiffness in Alzheimer's disease. *Clin Interv Aging*. (2014) 9:1115–21. doi: 10.2147/CIA.S63337

70. Jin W-S, Bu XL, Wang YR, Li L, Li WW, Liu YH, et al. Reduced cardiovascular functions in patients with Alzheimer's disease. *J Alzheimers Dis.* (2017) 58:919–25. doi: 10.3233/JAD-170088

71. Mosconi L, de Leon M, Murray J, E L, Lu J, Javier E, et al. Reduced mitochondria cytochrome oxidase activity in adult children of mothers with Alzheimer's disease. *J Alzheimers Dis.* (2011) 27:483–90. doi: 10.3233/JAD-2011-110866

72. Morris JK, Honea RA, Vidoni ED, Swerdlow RH, Burns JM. Is Alzheimer's disease a systemic disease? *Biochim Biophys Acta.* (2014) 1842:1340–9. doi: 10.1016/j. bbadis.2014.04.012

73. Willis MS, Patterson C. Proteotoxicity and cardiac dysfunction—Alzheimer's disease of the heart? *N Engl J Med.* (2013) 368:455–64. doi: 10.1056/NEJMra1106180

74. Stamatelopoulos K, Sibbing D, Rallidis LS, Georgiopoulos G, Stakos D, Braun S, et al. Amyloid-beta (1-40) and the risk of death from cardiovascular causes in patients with coronary heart disease. *J Am Coll Cardiol.* (2015) 65:904–16. doi: 10.1016/j. jacc.2014.12.035

75. Kuo Y-M, Kokjohn TA, Watson MD, Woods AS, Cotter RJ, Sue LI, et al. Elevated A β 42 in skeletal muscle of Alzheimer disease patients suggests peripheral alterations of A β PP metabolism. *Am J Pathol.* (2000) 156:797–805. doi: 10.1016/S0002-9440(10)64947-4

76. Parry TL, Melehani JH, Ranek MJ, Willis MS. Functional amyloid signaling via the inflammasome, necrosome, and signalosome: new therapeutic targets in heart failure. *Front. Cardiovasc. Med.* (2015) 2:25. doi: 10.3389/fcvm.2015.00025

77. Li D, Parks SB, Kushner JD, Nauman D, Burgess D, Ludwigsen S, et al. Mutations of presenilin genes in dilated cardiomyopathy and heart failure. *Am J Hum Genet*. (2006) 79:1030–9. doi: 10.1086/509900

78. Song X, Yuan QN, Tang Y, Cao M, Shen YF, Zeng ZY, et al. Conditionally targeted deletion of PSEN1 leads to diastolic heart dysfunction. *J Cell Physiol*. (2018) 233:1548–57. doi: 10.1002/jcp.26057

79. Yang Y, Bagyinszky E, An SSA. Presenilin-1 (PSEN1) mutations: clinical phenotypes beyond Alzheimer's disease. *Int J Mol Sci.* (2023) 24:8417. doi: 10.3390/ ijms24098417

80. Huang Z, Wong LW, Su Y, Huang X, Wang N, Chen H, et al. Blood-brain barrier integrity in the pathogenesis of Alzheimer's disease. *Front Neuroendocrinol.* (2020) 59:100857. doi: 10.1016/j.yfrne.2020.100857

81. Cheng L, Li W, Chen Y, Lin Y, Wang B, Guo Q, et al. Plasma Aβ as a biomarker for predicting Aβ-PET status in Alzheimer's disease: a systematic review with meta-analysis. *J Neurol Neurosurg Psychiatry*. (2022) 93:513–20. doi: 10.1136/jnnp-2021-327864

82. Song F, Poljak A, Valenzuela M, Mayeux R, Smythe GA, Sachdev PS. Meta-analysis of plasma amyloid- β levels in Alzheimer's disease. J Alzheimers Dis. (2011) 26:365–75. doi: 10.3233/JAD-2011-101977

83. Uddin MS, Kabir MT, Tewari D, Mamun AA, Mathew B, Aleya L, et al. Revisiting the role of brain and peripheral A β in the pathogenesis of Alzheimer's disease. *J Neurol Sci.* (2020) 416:116974. doi: 10.1016/j.jns.2020.116974

84. Daniele G, DiLucia S, Masci P-G, Del Monte F. Heart and brain: complex relationships for left ventricular dysfunction. *Curr Cardiol Rep.* (2020) 22:1–11. doi: 10.1007/s11886-020-01318-w

85. Citron M, Vigo-Pelfrey C, Teplow DB, Miller C, Schenk D, Johnston J, et al. Excessive production of amyloid beta-protein by peripheral cells of symptomatic and presymptomatic patients carrying the Swedish familial Alzheimer disease mutation. *Proc Natl Acad Sci.* (1994) 91:11993–7. doi: 10.1073/pnas.91.25.11993

86. Grigorova Y, Camandola S, Petrashevskaya N, McPherson R, Rumian NL, Hagood M, et al. P3-465: increase in aortic stiffness and development of cardiac hypertrophy is associated with cognitive impairment in 2XTG-ad mice. *Alzheimers Dement.* (2018) 14:P1296–8. doi: 10.1016/j.jalz.2018.06.1829

87. Abyadeh M, Gupta V, Paulo JA, Mahmoudabad AG, Shadfar S, Mirshahvaladi S, et al. Amyloid-beta and tau protein beyond Alzheimer's disease. *Neural Regen Res.* (2024) 19:1262–76. doi: 10.4103/1673-5374.386406

88. Veerabhadrappa B, Delaby C, Hirtz C, Vialaret J, Alcolea D, Lleó A, et al. Detection of amyloid beta peptides in body fluids for the diagnosis of alzheimer's disease: where do we stand? *Crit Rev Clin Lab Sci.* (2020) 57:99–113. doi: 10.1080/10408363.2019.1678011

89. Roher AE, Esh CL, Kokjohn TA, Castaño EM, van Vickle GD, Kalback WM, et al. Amyloid beta peptides in human plasma and tissues and their significance for Alzheimer's disease. *Alzheimers Dement*. (2009) 5:18–29. doi: 10.1016/j.jalz.2008.10.004

90. Bernini F, Malferrari D, Pignataro M, Bortolotti CA, di Rocco G, Lancellotti L, et al. Pre-amyloid oligomers budding: a metastatic mechanism of proteotoxicity. *Sci Rep.* (2016) 6:35865. doi: 10.1038/srep35865

91. Sen S, Hallee L, Lam CK. The potential of gamma secretase as a therapeutic target for cardiac diseases. J Pers Med. (2021) 11:1294. doi: 10.3390/jpm11121294

92. Bampatsias D, Mavroeidis I, Tual-Chalot S, Vlachogiannis NI, Bonini F, Sachse M, et al. Beta-Secretase-1 antisense rna is associated with vascular ageing and atherosclerotic cardiovascular disease. *Thromb Haemost*. (2022) 122:1932–42. doi: 10.1055/a-1914-2094

93. Farber G, Parks MM, Lustgarten Guahmich N, Zhang Y, Monette S, Blanchard SC, et al. ADAM10 controls the differentiation of the coronary arterial endothelium. *Angiogenesis.* (2019) 22:237–50. doi: 10.1007/s10456-018-9653-2

94. Del L, Lerma C, González-Pacheco H, Chávez-Lázaro A, Massó F, Rodriguez E. Correlation of plasmatic amyloid Beta peptides (Aβ-40, Aβ-42) with myocardial injury and inflammatory biomarkers in acute coronary syndrome. J Clin Med. (2024) 13:1117. doi: 10.3390/jcm13041117

95. Kitazume S, Yoshihisa A, Yamaki T, Oikawa M, Tachida Y, Ogawa K, et al. Soluble amyloid precursor protein 770 is a novel biomarker candidate for acute coronary syndrome. *J Biol Chem.* (2013) 7:657–63. doi: 10.1002/prca.201200135

96. Gowert NS, Donner L, Chatterjee M, Eisele YS, Towhid ST, Münzer P, et al. Blood platelets in the progression of Alzheimer's disease. *PLoS One*. (2014) 9:e90523. doi: 10.1371/journal.pone.0090523

97. Zamolodchikov D, Renné T, Strickland S. The Alzheimer's disease peptide β -amyloid promotes thrombin generation through activation of coagulation factor XII. J Thromb Haemost. (2016) 14:995–1007. doi: 10.1111/jth.13209

98. Li T, Li X, Feng Y, Dong G, Wang Y, Yang J. The role of matrix Metalloproteinase-9 in atherosclerotic plaque instability. *Mediat Inflamm.* (2020) 2020:3872367. doi: 10.1155/2020/3872367

99. Jang S, Chapa-Dubocq XR, Parodi-Rullán RM, Fossati S, Javadov S. Beta-amyloid instigates dysfunction of mitochondria in cardiac cells. *Cells.* (2022) 11:373. doi: 10.3390/cells11030373

100. Wolska N, Celikag M, Failla AV, Tarafdar A, Renné T, Torti M, et al. Human platelets release amyloid peptides β and β in response to haemostatic, immune, and hypoxic stimuli. *Res Pract Thromb Haemost.* 7:100154. doi: 10.1016/j.rpth.2023.100154

101. Sun Y, Jiang X, Lv Y, Liang X, Zhao B, Bian W, et al. Circular RNA expression profiles in plasma from patients with heart failure related to platelet activity. *Biomol Ther.* (2020) 10:187. doi: 10.3390/biom10020187

102. Stamatelopoulos K, Pol CJ, Ayers C, Georgiopoulos G, Gatsiou A, Brilakis ES, et al. Amyloid-beta (1-40) peptide and subclinical cardiovascular disease. J Am Coll Cardiol. (2018) 72:1060–1. doi: 10.1016/j.jacc.2018.06.027

103. Hamada M, Shigematsu Y, Ohtani T, Ikeda S. Elevated cardiac enzymes in hypertrophic cardiomyopathy patients with heart failure–a 20-year prospective follow-up study–. *Circ J.* (2015) 80:218–26. doi: 10.1253/circj.CJ-15-0872

104. Guo Y, Shen XN, Wang HF, Chen SD, Zhang YR, Chen SF, et al. The dynamics of plasma biomarkers across the Alzheimer's continuum. *Alzheimers Res Ther.* (2023) 15:31. doi: 10.1186/s13195-023-01174-0

105. Kanekiyo T, Bu G. The low-density lipoprotein receptor-related protein 1 and amyloid- β clearance in Alzheimer's disease. *Front Aging Neurosci.* (2014) 6:93. doi: 10.3389/fnagi.2014.00093

106. Gu BJ, Huang X, Ou A, Rembach A, Fowler C, Avula PK, et al. Innate phagocytosis by peripheral blood monocytes is altered in Alzheimer's disease. *Acta Neuropathol.* (2016) 132:377–89. doi: 10.1007/s00401-016-1596-3

107. Sousa JA, Bernardes C, Bernardo-Castro S, Lino M, Albino I, Ferreira L, et al. Reconsidering the role of blood-brain barrier in Alzheimer's disease: from delivery to target. *Front Aging Neurosci.* (2023) 15:1102809. doi: 10.3389/fnagi.2023.1102809

108. Zeng F, Liu Y, Huang W, Qing H, Kadowaki T, Kashiwazaki H, et al. Receptor for advanced glycation end products up-regulation in cerebral endothelial cells mediates cerebrovascular-related amyloid β accumulation after *Porphyromonas gingivalis* infection. *J Neurochem.* (2021) 158:724–36. doi: 10.1111/jnc.15096

109. Meissner K, Sperker B, Karsten C, zu Schwabedissen HM, Seeland U, Böhm M, et al. Expression and localization of P-glycoprotein in human heart: effects of cardiomyopathy. J Histochem Cytochem. (2002) 50:1351–6. doi: 10.1177/002215540205001008

110. Cercy SP. Pericytes and the neurovascular unit: the critical Nexus of Alzheimer disease pathogenesis? *Explor Res Hypothesis Med.* (2021):125–34. doi: 10.14218/ERHM.2020.00062

111. Potere N, Del Buono MG, Mauro AG, Abbate A, Toldo S. Low density lipoprotein receptor-related protein-1 in cardiac inflammation and infarct healing. *Front Cardiovasc Med.* (2019) 6:51. doi: 10.3389/fcvm.2019.00051

112. Gagno G, Ferro F, Fluca AL, Janjusevic M, Rossi M, Sinagra G, et al. From brain to heart: possible role of amyloid- β in ischemic heart disease and ischemia-reperfusion injury. *Int J Mol Sci.* (2020) 21:9655. doi: 10.3390/ijms21249655

113. Alasmari F, Alshammari MA, Alasmari AF, Alanazi WA, Alhazzani K. Neuroinflammatory cytokines induce amyloid beta neurotoxicity through modulating amyloid precursor protein levels/metabolism. *Biomed Res Int.* (2018) 2018:3087475. doi: 10.1155/2018/3087475

114. Itkin A, Dupres V, Dufrêne YF, Bechinger B, Ruysschaert JM, Raussens V. Calcium ions promote formation of amyloid β -peptide (1–40) oligomers causally implicated in neuronal toxicity of Alzheimer's disease. *PLoS One.* (2011) 6:e18250. doi: 10.1371/journal.pone.0018250

115. Ramasamy R, Schmidt AM. Receptor for advanced glycation end products (RAGE) and implications for the pathophysiology of heart failure. *Curr Heart Fail Rep.* (2012) 9:107–16. doi: 10.1007/s11897-012-0089-5

116. Gutierrez-Mariscal FM, Cardelo MP, de la Cruz S, Alcala-Diaz JF, Roncero-Ramos I, Guler I, et al. Reduction in circulating advanced glycation end products by mediterranean diet is associated with increased likelihood of type 2 diabetes remission in patients with coronary heart disease: from the cordioprev study. *Mol Nutr Food Res.* (2021) 65:e1901290. doi: 10.1002/mnfr.201901290 117. Ghiso J, Shayo M, Calero M, Ng D, Tomidokoro Y, Gandy S, et al. Systemic catabolism of Alzheimer's Aβ40 and Aβ42. *J Biol Chem*. (2004) 279:45897–908. doi: 10.1074/jbc.M407668200

118. Tian D-Y, Cheng Y, Zhuang ZQ, He CY, Pan QG, Tang MZ, et al. Physiological clearance of amyloid-beta by the kidney and its therapeutic potential for Alzheimer's disease. *Mol Psychiatry*. (2021) 26:6074–82. doi: 10.1038/s41380-021-01073-6

119. Xiang Y, Bu XL, Liu YH, Zhu C, Shen LL, Jiao SS, et al. Physiological amyloidbeta clearance in the periphery and its therapeutic potential for Alzheimer's disease. *Acta Neuropathol.* (2015) 130:487–99. doi: 10.1007/s00401-015-1477-1

120. Bayés-Genís A, Barallat J, Galán A, de Antonio M, Domingo M, Zamora E, et al. Soluble neprilysin is predictive of cardiovascular death and heart failure hospitalization in heart failure patients. *J Am Coll Cardiol.* (2015) 65:657–65. doi: 10.1016/j. jacc.2014.11.048

121. Marr RA, Hafez DM. Amyloid-beta and Alzheimer's disease: the role of neprilysin-2 in amyloid-beta clearance. *Front Aging Neurosci.* (2014) 6:187. doi: 10.3389/fnagi.2014.00187

122. Ishima Y, Mimono A, Tuan Giam Chuang V, Fukuda T, Kusumoto K, Okuhira K, et al. Albumin domain mutants with enhanced A β binding capacity identified by phage display analysis for application in various peripheral A β elimination approaches of Alzheimer's disease treatment. *IUBMB Life*. (2020) 72:641–51. doi: 10.1002/iub.2203

123. Zenaro E, Pietronigro E, Bianca VD, Piacentino G, Marongiu L, Budui S, et al. Neutrophils promote Alzheimer's disease–like pathology and cognitive decline via LFA-1 integrin. *Nat Med.* (2015) 21:880–6. doi: 10.1038/nm.3913

124. Munawara U, Catanzaro M, Xu W, Tan C, Hirokawa K, Bosco N, et al. Hyperactivation of monocytes and macrophages in MCI patients contributes to the progression of Alzheimer's disease. *Immun Ageing*. (2021) 18:29. doi: 10.1186/s12979-021-00236-x

125. Peikert A, Cunningham JW. Amyloid-β and the risk of heart failure: cause or only association? *Heart Failure*. (2023) 11:103–5. doi: 10.1016/j.jchf.2022.09.010

126. El Iskandarani M, El Kurdi B, Murtaza G, Paul TK, Refaat MM. Prognostic role of albumin level in heart failure: a systematic review and meta-analysis. *Medicine (Baltimore)*. (2021) 100:e24785. doi: 10.1097/MD.000000000024785

127. Okano T, Motoki H, Minamisawa M, Kimura K, Kanai M, Yoshie K, et al. Cardio-renal and cardio-hepatic interactions predict cardiovascular events in elderly patients with heart failure. *PLoS One.* (2020) 15:e0241003. doi: 10.1371/journal. pone.0241003

128. Santulli G, Varzideh F, Forzano I, Wilson S, Salemme L, de A, et al. Functional and clinical importance of SGLT2-inhibitors in frailty: from the kidney to the heart. *Hypertension*. (2023) 80:1800–9. doi: 10.1161/HYPERTENSIONAHA.123.20598

129. Tsampasian V, Baral R, Chattopadhyay R, Debski M, Joshi SS, Reinhold J, et al. The role of SGLT2 inhibitors in heart failure: a systematic review and Meta-analysis. *Cardiol Res Pract.* (2021) 2021:9927533. doi: 10.1155/2021/9927533

130. Mei J, Li Y, Niu L, Liang R, Tang M, Cai Q, et al. SGLT2 inhibitors: a novel therapy for cognitive impairment via multifaceted effects on the nervous system. *Transl Neurodegener*. (2024) 13:41. doi: 10.1186/s40035-024-00431-y

131. Magurno M, Cassano V, Maruca F, Pastura CA, Divino M, Fazio F, et al. Effects of SGLT2-inhibitors on comprehensive geriatric assessment, biomarkers of oxidative stress, and platelet activation in elderly diabetic patients with heart failure with preserved ejection fraction. *Int J Mol Sci.* (2024) 25:811. doi: 10.3390/ijms25168811

132. Mone P, Lombardi A, Gambardella J, Pansini A, Macina G, Morgante M, et al. Empagliflozin improves cognitive impairment in frail older adults with type 2 diabetes and heart failure with preserved ejection fraction. *Diabetes Care*. (2022) 45:1247–51. doi: 10.2337/dc21-2434

133. Rankin L, Svahn S, Kindstedt J, Gustafsson M. Differences in pharmacological treatment of heart failure among persons with or without major cognitive disorder: a cross-sectional study based on National Registries in Sweden. *Drugs Aging.* (2024) 41:907–13. doi: 10.1007/s40266-024-01153-6

134. Lardaro A, Quarta L, Pagnotta S, Sodero G, Mariani S, del Ben M, et al. Impact of sodium glucose cotransporter 2 inhibitors (SGLT2i) therapy on dementia and cognitive decline. *Biomedicines*. (2024) 12:1750. doi: 10.3390/biomedicines12081750