Check for updates

OPEN ACCESS

EDITED BY Vivek P. Chavda, L M College of Pharmacy, India

REVIEWED BY Monica Cricca, University of Bologna, Italy Kiran Bala, All India Institute of Medical Sciences, India

*CORRESPONDENCE Guoqing Qian ⊠ qian@outlook.com Yijun Mo ⊠ moyijun@126.com

RECEIVED 13 December 2024 ACCEPTED 20 March 2025 PUBLISHED 09 April 2025

CITATION

Wang S, Liu Y, Hu L, Qian G and Mo Y (2025) Case Report: Pulmonary mucormycosis caused by *Rhizopus microsporus* in a patient with chronic obstructive pulmonary disease. *Front. Med.* 12:1544621. doi: 10.3389/fmed.2025.1544621

COPYRIGHT

© 2025 Wang, Liu, Hu, Qian and Mo. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Case Report: Pulmonary mucormycosis caused by *Rhizopus microsporus* in a patient with chronic obstructive pulmonary disease

Shanshan Wang¹, Yanqing Liu¹, Liqing Hu¹, Guoqing Qian^{2*} and Yijun Mo^{1*}

¹Department of Laboratory Medicine, The First Affiliated Hospital of Ningbo University, Ningbo, China, ²Department of Infectious Diseases, The First Affiliated Hospital of Ningbo University, Ningbo, China

Background: We report a rare case of pulmonary mucormycosis caused by *Rhizopus microsporus*, which is rare in patients with chronic obstructive pulmonary disease. *Rhizopus microsporus* had been reported as the most common etiological agent associated with human infections, except *Rhizopus oryzae* in some studies.

Case presentation: We described a case of 81-year-old man with pulmonary mucormycosis caused by *Rhizopus microsporus* but no other apparent risk factors including diabetes. The diagnosis mainly relied on sputum cultures and clinical manifestations. Despite antifungal therapy, his condition worsened, resulting in mortality.

Conclusion: In this case, the patient had no underlying diseases such as diabetes or solid tumors. Clinicians should be aware of routine pathogenic microbiological tests of pulmonary mucormycosis in patients with chronic obstructive pulmonary disease. Early and aggressive treatment can lead to improved prognosis.

KEYWORDS

Rhizopus microsporus, pulmonary mucormycosis, chronic obstructive pulmonary disease, posaconazole, infection

Introduction

Mucormycosis is defined as an opportunistic and life-threatening fungal infection that originates from the fungi order Mucorales. It is associated with high morbidity and mortality rates in patients with immune deficiencies. Mucormycosis is the third most common invasive fungal infection following candidiasis and aspergillosis (1). The infection site of involvement of mucormycosis correlates with specific predisposing factors rhino-orbital mucormycosis occurs in uncontrolled diabetes and the recipients of hematopoietic stem cell transplant and solid-organ transplant. Pulmonary mucormycosis (PM) is the second most common manifestation of mucormycosis after nasal (2). Moreover, pulmonary mucormycosis have a rapid fatal clinical progression that can be fatal if it is untreated (3). The mortality rate of infection was high (50–70%) in recent studies. The predominant fungus species causing pulmonary mucormycosis were *Rhizopus, Lichtheimia, Cunninghamella*, and *Mucor* (4). Here, we described a case of pulmonary mucormycosis in an elderly male patient diagnosed with

chronic obstructive pulmonary disease (COPD) without diabetes mellitus (DM). Furthermore, we also present the data available so far on mucormycosis in COPD. Our case report emphasized the significance of considering fungal infections in patients with COPD.

Case presentation

The patient was diagnosed with chronic obstructive pulmonary disease (COPD) at a local hospital 30 years ago, but he had no longterm follow-up or maintenance therapy. He remained clinically stable until 20 days before admission, when he developed paroxysmal dry cough without identifiable triggers. He was presented to the emergency department with a three-day history of recurrent hemoptysis. The patient was treated with hemostasis in a local hospital, but the symptoms had not improved significantly. CT angiography enhanced examination of the pulmonary arteries revealed lung consolidation in the right upper lobe in a local hospital.

On admission, laboratory investigations revealed an elevated level of C-reactive protein (10.5 mg/L), hemoglobin of 120 g/L, neutrophil counts of 22.6×10^{9} /L, and platelet counts of 146×10^{9} /L. The level of alanine aminotransferase (95 U/L) was elevated, but albumin level was 36 g/L. He was not suffering from DM, hepatitis virus infections (hepatitis A, B, C, D, and E), HIV, and syphilis. RT-PCR detection for COVID19 was negative. The physical examination revealed that his temperature was 37.3°C, with a pulse of 99 bpm, hypertensive with a blood pressure of 168/87 mmHg, and respiratory rate with oxygen saturation of 95% under supplemental oxygen therapy via nasal cannula at 2 L/min on room air.

A cardiopulmonary examination noted that thickening breathing sounds in both lungs. The remainder of the physical examination was unremarkable. His oral mucosa was positive for thrush. He was discharged with the diagnosis of lung abscess, COPD with acute lower respiratory tract infection and accompanied by hypertension.

Having presented with severe symptoms of lung infection, the patient was admitted to the infectious disease department, and was empirically treated with intravenous meropenem (1 g/quaque die 8 h). In addition, bronchoscopy was recommended to identify the causative pathogens in order to determine the etiology. However, the patient explicitly declined bronchoscopy. Sputum cultures were collected before empiric treatment. To mitigate sampling limitations, we strictly followed guidelines for expectorated sputum collection, including pre-procedural oral hygiene, deep cough technique, and immediate transportation to the microbiology laboratory.

Two days later, despite being hemodynamically stable, the patient continued to be feverish with hemoptysis. Computed tomography (CT) scan of the chest indicated that lungs appeared inflammation, including a thick-walled cavity in the upper lobe of the right lung, an air sac cavity in the middle lobe of the right lung, and arteriosclerosis (Figure 1A).

After 2 days of incubation, the sputum fungal culture grew white and woolly colonies with a short nap inoculated on sabouraud dextrose agar (SDA) under 28°C (Figure 2A). Fluorescent microscope examination showed shows sporangiospores in pairs terminating in sporangium with subspherical columella containing brownish sporangiospores with primitive rhizoids (Figure 2B). Three consecutive cultures of the sputum grew Rhizopus microsporus, which was identified by matrix-assisted laser desorption/ionisation time-offlight mass (MALDI-TOF). For further identification, we performed the amplification of the internal transcribed spacer (ITS) gene by PCR with primer ITS1 and primer ITS4. The amplified PCR products were purified and directly sequenced through targeted DNA method. The DNA sequences were searched in GenBank by using the BLAST server. The ITS sequence demonstrated 99% identity to R. microsporus (GenBank accession number: No. MT 620751). Serological assay for Aspergillus, galactomannan, β -D-glucan, and quantitative PCR in serum sample was normal. Based on morphological characteristics (Figure 2) and sequences findings in immunocompromised states, a diagnosis of pulmonary mucormycosis (PM) was confirmed, Rhizopus microsporus being the most likely etiology. At the same time, antifungal susceptibility testing was performed according to the CLSI M38-A2 method (5). The results showed that the isolate was sensitive



FIGURE 1

(A) CT scan image of upper lobes of the lungs demonstrating bilateral pulmonary infection, with cavitation (about 32 × 40 mm) in the upper lobe of the right lung, and an air sac cavity in the middle lobe of the right lung. (B) After 10 days of antifungal treatment, chest CT showed a decrease in lung inflammation and a diminution of cavitation (about 29 × 21 mm) in the upper lobe of the right lung.

02



FIGURE 2

(A) Sabouraud dextrose agar (SDA) plate (60 mm) after 2 days incubation at 28°C, demonstrating white and woolly colonies with a short nap.
(B) Fluorescent staining examination shows sporangiospores in pairs terminating in sporangium with subspherical columella containing brownish sporangiospores with primitive rhizoids (×400).

to posaconazole and amphotericin B. Meanwhile, the targeted nextgeneration sequencing (tNGS) of sputum was further detected. The tNGS suggests influenza A virus H3N2 (sequence number: 1674). Unexpectedly, the sequencing results did not cue that *Rhizopus microsporus*.

With the diagnosis of PM caused by Rhizopus microsporus, he was administered a high dosage of liposomal amphotericin B (10 mg/kg/d IV) with monitoring of renal functions and electrolytes. In addition, the patient was also treated with oseltamivir to fight the virus. However, after 5 days of antifungal therapy, the treatment was discontinued due to persistent severe hypokalemia, which was difficult to correct. Itraconazole (0.3 g/quaque die 1 day) was then initiated on day 5, accompanied by symptomatic supportive treatment for hypertension. After 10 days of antifungal therapy, repeated thoracic CT scan (Figure 1B) revealed the cavities in the lungs were significantly reduced in size, while the pleural effusion was slightly increased. But the WBC, CRP, and PCT levels remained elevated, and multiple plaques were found in the arteries of the left lower extremity. Furthermore, he remained afebrile with breathlessness. His sputum cultures showed Acinetobacter baumannii infection. The patient was added to intravenous meropenem (1 g/quaque die 12 h) for antiinfection treatment. Unfortunately, his symptoms of the infection did not improve and the disease progressed rapidly after 22 days of admission. He and his family decided to give up further treatment with progressive worsening of shortness of breath. The patient's blood oxygen saturation was low at discharge, maintained at about 75-85% under oxygen inhalation. The patient died soon after discharge during the subsequent follow-up.

Discussion

Here we present a case of pulmonary mucormycosis with COPD. Mucormycosis is a rare, rapidly progressing, and life-threatening fungal disease caused by fungi of the order Mucorales. It has commonly been seen in immunocompromised patients (1).

Risk factors for mucormycosis include solid organ, stem cell transplantation, hematologic malignancies, uncontrolled DM, corticosteroids use, neutropenia, trauma, and burns. In recent years, the incidence of PM, has been increasing. Predisposing factors play an important role in mucormycosis infection. Conditions involving immunodeficiency are most prevalent risk factor, among which diabetes mellitus emerged as the most significant. Other risk factors, such as stem cell or solid organ transplant, hematological malignancy, and autoimmune diseases, cannot be ignored. Several studies have suggested an increased relative abundance of Rhizopus microsporus in patients with DM. Evidence showed that exposure to spores from Rhizopus microsporus might cause several types of respiratory symptoms in wood trimmers (6). In this case report, we emphasized the significance of considering fungal infections in patients with common diseases, including COPD. The risk factors for mucormycosis in our patient included coughing and expectorating repeatedly for more than 30 years. However, the interesting aspects of our case were the absence of DM and malignancy cause for immunosuppression. But his prognosis was still fatal. As evidenced by previous studies, Rhizopus microsporus is an opportunistic pathogen that could cause serious and lifethreatening infections.

Diagnosis of pulmonary mucormycosis is challenging owing to its rarity. The symptoms of pulmonary mucormycosis are non-specific, such as fever, cough, dyspnea, and chest pain. Prolonged fever is mostly seen in many patients with this infection, although it might be symptomless (7). Both bronchoscopic and percutaneous lung biopsy are effective tools to help diagnose PM. But bronchoscopy was declined by the patient in this case. Previous studies have identified associations between sputum microbiology and various clinical factors in patients with COPD exacerbations (8). The microbiological reliability of sputum specimens critically depends on proper collection techniques, particularly deep-cough expectoration under supervision. As evidenced by multicentre studies, supervised deep coughing significantly increases the yield of lower respiratory tract pathogens compared to spontaneously expectorated samples (9). These results will be of value to clinicians, allowing them to select which patients with COPD exacerbation need to have sputum cultures from deepcough expectoration. Isolation of *Rhizopus microsporus* from three sputum cultures further authenticated this diagnosis.

PM is hard to diagnose as patient presentation is similar to patients with Aspergillus infections. In aspergillosis, the hyphae are thinner, more organised, septate and show acute angle branching. Demonstration of broad, non-septate, ribbon-like hyphae with rightangle branching is obligatory to diagnose mucormycosis (10). Mucormycosis ECMM MSG Global Guideline Writing Group pointed out that histology remains the gold standard for the diagnosis of infection, while culture of specimens is strongly recommended for genus and species identification, and for antifungal susceptibility testing (11). So the correct identification of fungi is essential in selecting the appropriate treatment. The gold standard diagnostic technique is culture and histopathological examination is usually for complementary. The patient had hemoptysis for 3 days while he was admitted at the local hospital, which was diagnosed pulmonary aspergillosis. Hemoptysis is possible due to erosion of the pulmonary artery into the tracheobronchial tree (7). We speculated that misdiagnosis was a result of the lack of morphological discrimination between Rhizopus microsporus, Rhizopus arrhizus, and Aspergillus, and local laboratory conditions were limited. Rhizopus arrhizus and R. microsporus are microscopically distinguished by unbranched sporangiophores with elliptical-cylindrical spores (5-8 µm) versus branched sporangiophores bearing striated subglobose spores (3–5 μ m), complemented by thermotolerance divergence (\leq 45°C vs. $30-35^{\circ}C$) (12). In the present case, given that he could not tolerate urgent surgery, definitive diagnosis was through sputum culture. Three sputum cultures during the hospitalization suggested Rhizopus microsporus. In recent years, NGS methods have been used to try to improve the detection and identification of pathogens and have become a topic of concern as routine pathogen identification tool (13). The identification of influenza A virus H3N2 in sputum by tNGS was achieved in the early stage. Long-term use of corticosteroids in COPD patients may lead to immunosuppression, increasing the risk of fungal infections. Inhalation of fungal spores in patients with compromised lung defenses may result in PM (14). The influenza virus can damage the respiratory epithelium, facilitating fungal invasion (15). But tNGS did not indicate Rhizopus microsporus infections. The reason for the failure may be that Trichoucales belongs to the pathogen with thicker cell wall. With low extraction rate of nucleic acid, tNGS of the sample resulted in low sensitivity and detection rate (16). Therefore, when PM was suspected clinically, it should not rely on mNGS or tNGS results simply. Isolation of fungi by culture is vital for definitive identification in the early clinical diagnosis. Clinicians should keep an eye on patients who are at high risk of acquiring this fatal disease and make early intervention strategies to reduce terrible outcomes. To achieve a more rapid diagnosis of mucormycosis, more modern laboratory tools are needed, besides improved awareness by clinicians.

PM has a rapid fatal clinical progression. Its prognosis is dependent on early diagnosis and aggressive treatment. The infection has a high mortality rate (50–70%). In recent studies, surgical resection was recommended if hemoptysis was massive (10). An unprecedented outbreak of intestinal zygomycosis due to *Rhizopus microsporus* occurred in China, and similar cases have been reported in USA and elsewhere in Europe (17). Supplementary Table S1 shows clinical characteristics and outcomes of Rhizopus microsporus infections in patients in recently reported clinical cases (n = 10), including year, country, age (y), infection type and outcome. Current guidelines recommend a combined medical and local surgical debridement to management. As antifungal agents may have poor penetration at the site of the infection and the disease is rapidly progressive and associated with a bad prognosis (11). In the medical treatment of mucormycosis, the timely initiation of a liposomal formulation of amphotericin B monotherapy is the first line treatment for mucormycosis (18). Posaconazole and isavuconazole are acceptable salvage and long-term treatment options when patients are at a high risk of amphotericin-induced nephrotoxicity (19). In this case, although the patient was immediately started on a lipid formulation of amphotericin B delivered at a higher dose through aerosolized therapy with antifungal coverage before being switched to posaconazole, our patient had an unfavorable prognosis eventually. We speculated that it might be related to the rapid course of underlying disease and angioinvasive infection by Rhizopus microsporus.

Conclusion

In conclusion, this case illustrates the rapid progression of PM associated with deadly outcomes. Timely prevention, early diagnosis and symptomatic treatment are effective ways to prevent mucormycosis from spreading and to ameliorate patient outcomes despite the low survival rates in disseminated cases. Laboratory personnel and doctors should cooperate with each other and summarize clinical outcomes to improve the diagnosis and efficiency from the initial pity.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary material, further inquiries can be directed to the corresponding authors.

Ethics statement

The studies involving humans were approved by the First Affiliated Hospital of Ningbo University (2024202RS). The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article. Written informed consent was obtained from the participant/patient(s) for the publication of this case report.

Author contributions

SW: Conceptualization, Data curation, Writing - original draft, Writing - review & editing. YL: Formal analysis,

Investigation, Writing – original draft. LH: Data curation, Software, Supervision, Writing – review & editing. GQ: Methodology, Project administration, Resources, Writing – review & editing. YM: Conceptualization, Data curation, Formal analysis, Resources, Validation, Visualization, Writing – review & editing.

Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Kurosawa M, Yonezumi M, Hashino S, Tanaka J, Nishio M, Kaneda M, et al. Epidemiology and treatment outcome of invasive fungal infections in patients with hematological malignancies. *Int J Hematol.* (2012) 96:748–57. doi: 10.1007/s12185-012-1210-y

2. Iqbal N, Irfan M, Jabeen K, Kazmi MM, Tariq MU. Chronic pulmonary mucormycosis: an emerging fungal infection in diabetes mellitus. *J Thorac Dis.* (2017) 9:E121–5. doi: 10.21037/jtd.2017.02.31

3. Zhang M, Luo Q, Li X, Mu X, Wu W. Pulmonary artery pseudoaneurysm caused by pulmonary mucormycosis. *Am J Respir Crit Care Med.* (2024) 209:879–80. doi: 10.1164/rccm.202307-1125IM

4. Petrikkos G, Skiada A, Lortholary O, Roilides E, Walsh TJ, Kontoyiannis DP. Epidemiology and clinical manifestations of mucormycosis. *Clin Infect Dis.* (2012) 54:S23–34. doi: 10.1093/cid/cir866

5. Rex JH, Alexander BD, Andes D, Arthington-Skaggs B, Brown SD, Chaturveli V, et al. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi, approved standard M38-A2. 2nd ed. Wayne, PA: National Committee for Clinical Laboratory Standards (2008).

6. Tsyrkunou AV, Ellison RT 3rd, Akalin A, Wiederhold N, Sutton DA, Lindner J, et al. Multifocal *Rhizopus microsporus* lung infection following brush clearing. *Med Mycol Case Rep.* (2014) 6:14–7. doi: 10.1016/j.mmcr.2014.08.001

7. Cornely OA, Alastruey-Izquierdo A, Arenz D, Chen SCA, Dannaoui E, Hochhegger B, et al. Global guideline for the diagnosis and management of mucormycosis: an initiative of the European Confederation of Medical Mycology in cooperation with the Mycoses Study Group Education and Research Consortium. *Lancet Infect Dis.* (2019) 19:e405–21. doi: 10.1016/S1473-3099(19)30312-3

 Tsimogianni AM, Papiris SA, Kanavaki S, Stathopoulos GT, Sotiropoulou C, Manali ED, et al. Predictors of positive sputum cultures in exacerbations of chronic obstructive pulmonary disease. *Respirology*. (2009) 14:1114–20. doi: 10.1111/j.1440-1843.2009.01615.x

9. Murphy CN, Fowler R, Balada-Llasat JM, Carroll A, Stone H, Akerele O, et al. Multicenter evaluation of the BioFire FilmArray pneumonia/pneumonia plus panel for detection and quantification of agents of lower respiratory tract infection. J Clin Microbiol. (2020) 58:e00128. doi: 10.1128/JCM.00128-20

Generative AI statement

The authors declare that no Gen AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed.2025.1544621/ full#supplementary-material

10. Ibrahim AS, Spellberg B, Walsh TJ, Kontoyiannis DP. Pathogenesis of mucormycosis. *Clin Infect Dis.* (2012) 54:S16–22. doi: 10.1093/cid/cir865

11. Cornely OA, Arikan-Akdagli S, Dannaoui E, Groll AH, Lagrou K, Chakrabarti A, et al. ESCMID and ECMM joint clinical guidelines for the diagnosis and management of mucormycosis 2013. *Clin Microbiol Infect.* (2014) 20:5–26. doi: 10.1111/1469-0691.12371

12. Walther G, Wagner L, Kurzai O. Updates on the taxonomy of Mucorales with an emphasis on clinically important taxa. *J Fungi*. (2019) 5:106. doi: 10.3390/jof5040106

13. Pongas GN, Lewis RE, Samonis G, Kontoyiannis DP. Voriconazole-associated zygomycosis: a significant consequence of evolving antifungal prophylaxis and immunosuppression practices? *Clin Microbiol Infect.* (2009) 15:93–7. doi: 10.1111/j.1469-0691.2009.02988.x

14. Spira A, Brecher S, Karlinsky J. Pulmonary mucormycosis in the setting of chronic obstructive pulmonary disease. *Respiration*. (2002) 69:560–3. doi: 10.1159/000066467

15. Ahmadikia K, Hashemi SJ, Khodavaisy S, Getso MI, Alijani N, Badali H, et al. The double-edged sword of systemic corticosteroid therapy in viral pneumonia: a case report and comparative review of influenza-associated mucormycosis versus COVID-19 associated mucormycosis. *Mycoses.* (2021) 64:798–808. doi: 10.1111/myc.13256

16. Millon L, Larosa F, Lepiller Q, Legrand F, Rocchi S, Daguindau E, et al. Quantitative polymerase chain reaction detection of circulating DNA in serum for early diagnosis of mucormycosis in immunocompromised patients. *Clin Infect Dis.* (2013) 56:e95–e101. doi: 10.1093/cid/cit094

17. Cheng VC, Chan JF, Ngan AH, Too KK, Leung SY, Tsoi HW, et al. Outbreak of intestinal infection due to *Rhizopus microsporus*. J Clin Microbiol. (2009) 47:2834–43. doi: 10.1128/JCM.00908-09

18. Park BJ, Pappas PG, Wannemuehler KA, Alexander BD, Anaissie EJ, Andes DR, et al. Invasive non-aspergillus mold infections in transplant recipients, United States, 2001–2006. *Emerg Infect Dis.* (2011) 17:1855–64. doi: 10.3201/eid1710.110087

19. Chitasombat MN, Kontoyiannis DP. The 'cephalosporin era' of triazole therapy: isavuconazole, a welcomed newcomer for the treatment of invasive fungal infections. *Expert Opin Pharmacother*. (2015) 16:1543–58. doi: 10.1517/14656566.2015.1057500