Check for updates

OPEN ACCESS

EDITED BY Fanfan Zhou, The University of Sydney, Australia

REVIEWED BY Aristeidis Konstantinidis, University Hospital of Alexandroupolis, Greece Lijie Pan, Shandong University, China Lin Mu, Shanghai University of Traditional Chinese Medicine, China Xiaoli Shi, Chinese Academy of Sciences (CAS), China Noni Novisari Soeroso, University of North Sumatra, Indonesia

*CORRESPONDENCE Qinghua Peng ⊠ pengqinghua@hnucm.edu.cn

RECEIVED 07 April 2025 ACCEPTED 20 June 2025 PUBLISHED 08 July 2025

CITATION

Xun X, Liu X, Zhou P, Wu C and Peng Q (2025) Feasibility analysis of Sinomenine alleviating fibrosis of filtering bleb after glaucoma filtering surgery: a mini review. *Front. Med.* 12:1607500. doi: 10.3389/fmed.2025.1607500

COPYRIGHT

© 2025 Xun, Liu, Zhou, Wu and Peng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Feasibility analysis of Sinomenine alleviating fibrosis of filtering bleb after glaucoma filtering surgery: a mini review

Xin Xun¹, Xiyuan Liu¹, Pai Zhou¹, Chengliang Wu¹ and Qinghua Peng^{1,2,3*}

¹Hunan University of Chinese Medicine, Changsha, China, ²Key Laboratory of Traditional Chinese Medicine for Prevention and Treatment of Eye, Ear, Nose and Throat Diseases, Changsha, China, ³First Affiliated Hospital of Hunan University of Chinese Medicine, Changsha, China

Fibrosis of the filtering bleb remains the predominant cause of glaucoma filtering surgery failure, mediated by interconnected pathological processes including postoperative local inflammation, aberrant fibroblast proliferation, and deposition of the extracellular matrix (ECM). The antimetabolite drugs 5-fluorouracil (5-FU) and mitomycin C (MMC) are effective in preventing filtering bleb fibrosis, but their non-specific cytotoxic effects necessitate the development of targeted therapeutic alternatives. Fibrosis is a group of diseases with similar pathological mechanisms and molecular features. By analyzing evidence of Sinomenine's (SIN) anti-fibrotic effects across multiple organs, this study explores its potential use in glaucoma filtration surgery (GFS) to reduce scarring: (1) SIN inhibits trauma-induced NF-kB activation in Tenon's fibroblasts (TFs), reduces neutrophil and macrophage infiltration, and suppresses cytokine cascades. Besides, SIN targets the phosphatidylinositol-3-kinase (PI3K)/Akt pathway to attenuate macrophage M2 polarization and neutrophil recruitment, thereby interrupting fibrotic progression. (2) SIN suppresses transforming growth factor- β (TGF- β)/Smad3 signaling and inhibits the transdifferentiation of fibroblasts into α smooth muscle actin (α SMA) expressing myofibroblasts (MFs). SIN also blocks fibroblast proliferation and migration via PI3K/Akt/mTORC1 axis inhibition, restraining myofibroblast differentiation-the central pathological event in filtering bleb scarring. SIN shows antifibrotic efficacy, and feasibility studies on its application may offer novel insights into antifibrotic strategies.

KEYWORDS

Sinomenine, glaucoma, glaucoma filtering surgery, filtering bleb, fibrosis

1 Introduction

Glaucoma is a group of optic neuropathies characterized by progressive degeneration of retinal ganglion cells, leading to optic nerve damage and visual field loss. Primary open-angle glaucoma (POAG) is the most common type of glaucoma worldwide. Most cases of POAG may progress to total blindness without the patient experiencing any pain or discomfort. Glaucoma has become the leading cause of irreversible blindness worldwide (1–3). It is projected that by 2040, the number of glaucoma cases will reach 110 million worldwide, with up to 60% of the cases in Asia, posing a serious threat to global visual health and quality of life (4, 5). Multiple factors influence the risk of developing glaucoma, including age, race, family history, corneal thickness, systemic hypotension, cerebrospinal fluid pressure, intraocular pressure (IOP), and vascular disorders (6).



Pathological elevation of intraocular pressure (IOP) is an independent risk factor for glaucoma. Lowering IOP through various treatments is an established strategy for preventing vision loss in patients with glaucoma (7). When medication and laser therapies fail to adequately control IOP, surgical intervention becomes the primary therapeutic consideration. Glaucoma filtration surgery (GFS) is the first-line surgical option in current clinical practice. This surgical procedure facilitates aqueous humor drainage through the creation of a subconjunctival filtration channel to regulate IOP. The key determinant of this procedure is the establishment and maintenance of a patent and functioning filtering bleb. However, the procedure damages the ocular tissues, activating the wound repair cascade. Inflammatory response during the early healing phase may drive fibrosis. This process involves the proliferation and differentiation of fibroblasts and the excessive deposition of extracellular matrix (ECM). These changes cause postoperative fibrosis of the filtering bleb, which in turn damages the function of the filtering bleb, block the outflow of aqueous humor, and ultimately lead to failure of GFS (8, 9). Therefore, the prevention of excessive fibrosis of the surgical area has been a major research focus in GFS.

Antimetabolite agents such as 5-fluorouracil (5-FU) and mitomycin C (MMC) are standard therapeutic agents to prevent fibrosis of filtering blebs. Although these agents effectively prolong the survival duration of filtering blebs and enhance the longterm success rate of glaucoma filtration surgery (GFS), they carry significant risks of ocular complications. These include thin-walled cystic blebs predisposing to late leakage, heightened infection risks, chronic hypotony-associated pathologies, and corneal epithelial damage (10, 11). Investigating alternative therapeutic approaches could help improve clinical outcomes.

Sinomenine (SIN) is an alkaloid monomer derived from *Sinomenium acutum*, a plant of the Menispermaceae family. Its molecular formula is $C_{19}H_{23}NO_4$, and its molecular weight is 329.39 (12). SIN shows immunosuppressive, anti-inflammatory, apoptosis-inducing, antihypertensive, analgesic, and other

pharmacological effects, and it is potentially efficacious for antifibrotic applications. SIN exerts its antifibrotic effects primarily through inhibiting signaling pathways [including transforming growth factor- β (TGF- β)/Smad, phosphatidylinositol-3-kinase (PI3K)/Akt, and NF- κ B], suppressing inflammatory cytokine release, and downregulating fibroblast activation. Fibrosis in the filtering bleb shares a similar pathology with fibrosis of other tissues and organs, but there have been no reports of SIN inhibiting fibrosis in the filtering bleb. In this review, we analyze the mechanisms of fibrosis of the filtering bleb and the pharmacological effects of SIN in order to evaluate the feasibility of using SIN to inhibit fibrosis after GFS.

2 Mechanism of fibrosis of filtering bleb after GFS

2.1 Healing process

The healing process of the filtering bleb after GFS is in accordance with general wound healing, following the pattern of hemostasis, inflammation, cellular proliferation, and tissue remodeling (13, 14). After the procedure, platelets aggregate at damaged vessels and form fibrin clots, with platelets releasing inflammatory factors, growth factors, and activating the inflammatory response. During the inflammatory phase, cellular infiltration is increased. The cytokine secretion drives fibroblast activation, and fibroblasts sustainably proliferate and transdifferentiate into myofibroblasts (MFs) which express α -smooth muscle actin (α -SMA) (15). Large amounts of ECM are released by MFs and promote protein deposition. This causes migration and epithelialization of epithelial cells, neovascularization, and granulation tissue formation, leading to tissue remodeling and eventual scar formation (16).

2.2 Injury-induced activation

The Tenon's capsule, also known as the fascial sheath of the eyeball, is a dense connective tissue that wraps around the outer sclera and is populated by fibroblasts (17). Fibroblasts are able to transdifferentiate into MFs, which play a central role in ECM synthesis and secretion. They facilitate wound healing, and are also involved in numerous fibrotic diseases. Fibrosis of the filtering bleb is mainly induced by the proliferation, migration and contraction of Tenon's fibroblasts (TFs). Hyperproliferation and differentiation of TFs are important for postoperative fibrosis in the filtering area. Surgical procedures inevitably cause some degree of damage to the cornea, conjunctiva, Tenon's capsule, and sclera. During the healing process of the filtering area, excessive fibrosis develops. Studies have shown (18) that patients with glaucoma exhibit significant fibrotic changes in TFs, as evidenced by the transdifferentiation of fibroblasts to myofibroblasts, and associated changes such as mitochondrial fission, ECM remodeling, proliferation, inflammation, and apoptosis. These changes may be related to their pathogenesis and/or the damage caused by local treatment.

2.3 Inflammatory response

Inflammation is one of the crucial factors in the formation of scarring and occurs in the early stages of wound healing. Excessive and prolonged inflammation impairs wound healing and promotes scar formation (19). The acute inflammatory response is characterized by increased exudation, thickening of the filtering bleb, dense collagenous tissue, and hyperproliferation of fibroblasts, and excessive angiogenesis. As a result of the activation of the endogenous coagulation cascade reaction, large amounts of cytokines and growth factors are released, causing the wound healing phase to prolong the inflammatory phase. These factors include tumor necrosis factor- α (TNF- α), interleukin (IL), transforming growth factor-β (TGF-β), plateletderived growth factor (PDGF), and vascular endothelial growth factor (VEGF), which are able to recruit and activate fibroblasts and vascular endothelial cells in turn (20-22). TNF- α , as an inflammatory mediator, promotes inflammatory cell recruitment in subconjunctival tissues and exerts a destructive effect on trabecular cells by up-regulating the expression of pro-inflammatory factors, such as IL-1 and IL-6. Fibroblast proliferation exhibits a higher rate and longer duration after TNF-α intervention (22). IL-6 plays a crucial role in the development of scar formation after GFS by accelerating the fibrotic process by promoting the proliferation of CD4+ T cells, inhibiting autophagy, enhancing endoplasmic reticulum stress, and promoting the transformation of fibroblasts in the early stages of inflammation (23, 24). TGF- β in atrial fluid has been shown to promote fibroblast migration, proliferation, and differentiation, as well as increase the expression of type I collagen and fibronectin, leading to scarring of postoperative filtration channels (25, 26). VEGF directly modulates a spectrum of pro-fibrotic genetic pathways by orchestrating myofibroblast differentiation through upregulation of collagen synthesis and α -smooth muscle actin (α -SMA) expression, thereby mediating fibrotic progression (27). PDGF induces vascular repair, promotes proliferation and migration of macrophages and fibroblasts to the wound site, and stimulates fibroblasts to transdifferentiate into myofibroblasts, which enhances the ECM and angiogenesis and promotes scarring (28, 29).

2.4 Fibroblast transdifferentiation

TFs are activated by a combination of cytokines and growth factors, start to proliferate and migrate, and undergo sustained transdifferentiation into MFs. MFs synthesize large amounts of collagen-rich ECM. The disordered collagen fibers induce abnormal cell movement around the wound, leading to excessive blood vessel proliferation, scar tissue formation, and tissue tightening during scar formation. During the fibrotic remodeling phase, TFs and MFs gradually undergo apoptosis, and the ECM forms a dense scar through the dehydration of collagen cross-links induced by selective degradation (30), which blocks the functional filtering bleb and disrupts aqueous humor drainage.

In summary, fibrosis is a common pathological outcome of trauma and inflammatory responses, and excessive fibrosis leads to scar formation (Figure 1).



proliferation, migration, and contractile activity of TFs. Surgical trauma triggers vascular disruption, releasing platelets and bload components that initiate coagulation cascades. Necrotic tissue debris, coagulative processes, and microbial infiltration collectively induce inflammatory activation. Inflammatory cells infiltrate the wound site to phagocytize cellular debris and pathogens while secreting growth factors and cytokines, including TGF- β , VEGF, PDGF, and IL-6. During the proliferative phase, neovascularization occurs while TFs are activated and transdifferentiate into MFs expressing α -SMA. Concurrently, ECM synthesis establishes a granulation tissue scaffold, facilitating wound contraction and repair. Ultimately, ECM remodeling occurs, culminating in the maturation of granulation tissue into dense fibrotic scar tissue).

3 Current status of fibrosis of filtering bleb medication after GFS

To mitigate the fibrosis of the filtering bleb after GFS, glucocorticoids (31), and antimetabolites are commonly used in clinical practice. Corticosteroids are able to modulate fibroblast recruitment and inhibit their activity by alleviating the inflammatory response and reducing the release of inflammatory mediators. This supports their role in regulating the fibrotic process. However, glucocorticoids may increase the risk of postoperative infection (32). The antimetabolites mitomycin C (MMC) and 5-Fluorouracil (5-FU) are now commonly used clinical antifibrotic drugs. MMC, a broad-spectrum antitumor antibiotic isolated from Streptomyces, is a DNA cross-linking alkylating agent. MMC inhibits cellular DNA synthesis and replication, reduces cell proliferation, induces apoptosis in target TFs, and partially mitigates the fibrotic process (33). Compared with 5-FU, MMC is more effective and long-lasting in inhibiting the proliferation of fibroblasts, and it is considered the gold standard for mitigating postoperative fibrosis (34, 35). However, both MMC and 5-FU are associated with side effects such as thin-walled cystic blebs, late bleb leakage, bleb-related infections, chronic hypotony, hypotony maculopathy, and corneal epithelial toxicity (36-38).

Emerging experimental studies have revealed that immunosuppressive agents such as rapamycin augment cellular autophagic activity and, thus inhibit tissue fibrosis (39, 40). In *in vitro* experiments, bevacizumab has been shown to inhibit fibroblast proliferation by reducing new blood vessel formation and collagen deposition. It also suppress scar formation progression in animal models (41). Another study (42) demonstrated that the combination of MMC and bevacizumab enhances the success rate of GFS compared to MMC alone. Postoperative follow-up further revealed improved maintenance of filtering bleb morphology in patients. Intravitreal injection of ranibizumab has been shown to reduce angiogenesis and maintain postoperative filtering bleb morphology (43). Although anti-vascular endothelial growth factor agents exhibit anti-fibrotic effects, their use in this context remains controversial and requires further investigation (44). Rosiglitazone blocks the p38 signaling pathway. This inhibition suppresses TGF-\u00c31-induced proliferation and differentiation of TFs and prolongs functional bleb survival (45). A novel protein, the S58 aptamer, targeting TGF-β receptor II, suppresses fibroblast transdifferentiation into myofibroblasts mediated by TGF- β 2 (46). Comparative studies with mitomycin C (MMC) demonstrated a significant reduction in myofibroblast numbers in the S58 aptamer-treated group (47). However, the free S58 aptamer is susceptible to nuclease degradation and requires nanocarrier-based delivery systems (e.g., exosomes) (48). IP-10 peptide is a smallmolecule cytokine that inhibits fibroblast migration, angiogenesis, and collagen deposition by binding to the CXCR3 receptor. It blocks VEGF-induced angiogenesis and promotes the regression of neovascularization (49). Studies have shown that IP-10p-treated filtering blebs exhibit reduced collagen deposition, decreased cell density, and inhibited scar formation (50). The vascular density of filtering blebs in the IP-10p group was also lower than that in the MMC-treated group. Additionally, the IP-10p combined with MMC group demonstrated reduced conjunctival damage compared to MMC alone. Animal experimental studies on Rho kinase (51) and matrix metalloproteinases (52) have shown

antifibrotic efficacy. All these agents exhibit certain anti-fibrotic potential in filtering blebs and represent viable alternatives. However, their efficacy and safety in human glaucoma patients still require further clinical testing.

In recent years, active ingredients of traditional Chinese medicine have shown unique potential in inhibiting postoperative fibrosis in glaucoma due to their multi-target regulatory properties. Studies have found that homoharringtonine, an antimetabolic agent, acts as an anti-fibroblast proliferative agent by inhibiting DNA synthesis (53). It has been shown that Hansenulae mitigate fibrosis progression by inhibiting fibroblast proliferation through apoptosis and down-regulating filtration bleb fibrosis (54). Quercetin is a flavonoid compound. In vitro experiments revealed that it inhibits postoperative glaucoma fibrosis by inhibiting collagen synthesis and cell proliferation (55, 56). Although significant progress has been made in previous studies, herbal monomers still face challenges in antifibrosis research. Current research is still at the stage of in vitro and animal experiments, with a lack of dynamic simulation systems capable of replicating physiological aqueous microenvironments. Clarifying the mechanisms of herbal monomers in filtering bleb fibrosis and developing optimized delivery methods with precise dosages will establish a robust foundation for clinical trials, thereby enhancing the evidence base for clinical translation. Identification of herbal components with high efficacy, low toxicity, and high bioavailability is critical for postoperative antifibrotic studies in glaucoma.

SIN is a purified alkaloid from the traditional Chinese medicine *Sinomenium acutum* (57). SIN has been shown to reduce the formation of scar tissue in organs and tissues (58), including mitigating fibrosis in the lungs, liver, kidneys, and other tissues (59–63). Because of the similarity in the pathological mechanisms of fibrotic diseases, SIN is hypothesized to exert antifibrotic effects after GFS. Although the antifibrotic effect of SIN has not been widely applied in the field of ophthalmic diseases, its extensive research background, low toxicity, and high production yield suggest potential value (64). Investigating the pharmacological properties of SIN will help to understand its antifibrotic mechanisms after GFS and provide new therapies for postoperative fibrosis management. As a promising alternative or adjuvant agent, SIN warrants further exploration in preclinical models and clinical trials to validate its translational potential.

4 SIN prevents and suppresses fibrosis of filtering bleb after GFS

4.1 Anti-inflammatory

As the initiating phase of tissue repair and scar formation, reducing the inflammatory response helps inhibit postoperative filtering bleb fibrosis. Studies have shown that SIN reduces the secretion of inflammatory factors like IL-6, GM-CSF, IL-12 p40, IL-1 α , IL-1 β , TNF- α and other inflammatory factors in the serum of mice, and demonstrates significant anti-inflammatory activity (65). SINO-WCJ-33, a SIN derivative, significantly reverses elevated serum levels of IL-2, IL-6, and TNF- α in mice and plays an important role in modulating inflammatory responses (66). In ophthalmology research, SIN eye drops given to mice with

experimental dry eye showed that the SIN-treated group exhibited significantly reduced corneal expression of IL-1 β and TNF- α compared to controls, along with increased tear production (67).

Damaged cells and pathogens are cleared when the inflammatory response initiates, and subsequently neutrophils, macrophages, and lymphocytes are recruited to the injury site, producing clots, platelet-derived mediators, cytokines, and other factors. These mediators in turn induce fibroblast migration and transdifferentiate fibroblasts into MFs. These changes cause protein deposition and the formation of dense scar tissue (68). Previous research has demonstrated that therapeutic strategies targeting the inflammatory response after GFS reduce the progression of filtering bleb fibrosis. SIN exhibits significant anti-inflammatory effects, inhibiting the infiltration of inflammatory cells and the production of various cytokines, thereby reducing postoperative filtering bleb fibrosis (69, 70).

4.2 Inhibits fibroblast proliferation and promotes apoptosis

SIN significantly inhibits fibroblast proliferation and promotes apoptosis. By inhibiting TGF-β1/Smad3, PI3K/Akt, and NF-κB signaling pathways, SIN inhibits the migration and proliferation of fibroblasts and A549 cells. It prevents myofibroblast transdifferentiation and epithelial mesenchymal transition, resulting in reduced ECM protein expression (71). SIN has been shown to inhibit fibrosis progression by promoting collagen-I and collagen-III degradation through upregulation of ADAMTS-1 expression (72). After SIN treatment, fibroblasts exhibit reduced activity, decreased colony formation, and increased apoptosis rates. This suggests that SIN directly inhibits fibroblast proliferation while promoting apoptosis. These changes may be produced by upregulation of miR-23b-3p expression and downregulation of FGF9 expression in the miR-23b-3p/FGF9 axis (73). Similar to other fibrotic pathologies, involves excessive collagen and ECM deposition mediated by fibroblasts. Furthermore, another study reported the dose-dependent pro-apoptotic effect of SIN on human Tenon fibroblasts (74).

Western blot analysis showed that the TGF- β 1 signaling pathway in human fibroblasts was significantly inhibited after SIN treatment. Meanwhile, the expression levels of Cyclin D1, Bcl-2, and MMP2 were significantly reduced. This study demonstrated that SIN suppresses fibroblast growth and migration, an effect mediated through inhibition of the TGF- β 1 signaling pathway. These findings establish a mechanistic foundation for further exploration of SIN in anti-fibrotic therapy.

Fibroblasts are involved in fibrotic activities in the body and play an important role in promoting angiogenesis and granulation tissue formation. As wound healing proceeds to the cell proliferation stage, fibroblasts are recruited and activated to transdifferentiate into MFs, leading to increased collagen synthesis and deposition. This further drives fibrosis (75). In this stage, the key steps to mitigate fibrosis of the filtering bleb after GFS include: (1) inhibition of fibroblast migration and proliferation, and promotion of fibroblast apoptosis; and (2) suppression of MF transdifferentiation and induction of their apoptosis. A large body of evidence shows that SIN not only inhibits fibroblast migration, and proliferation but also promotes fibroblast apoptosis, reduces α -SMA expression, and enhances collagen degradation. SIN effectively inhibits the progression of fibrosis. TFs are one of the key effector cells in fibrosis of filtering bleb after GFS (76). SIN may suppress the fibrosis of the filtering bleb after GFS by modulating the activity of TFs and thereby maintaining functional filtering bleb to increase the success of the procedure.

4.3 Possible molecular mechanisms

The antifibrotic effect of SIN is mainly reflected in: (1) the anti-inflammatory effect of SIN attenuates the local inflammatory reaction after trauma or surgery, reduces inflammatory cell infiltration and cytokine release, and inhibits fibrosis at the injury site; and (2) SIN inhibits the activation, migration, and proliferation of fibroblasts, promotes their apoptosis, and suppresses their transdifferentiation into myofibroblasts. Based on its anti-inflammatory and antifibrotic pharmacological effects, evidence suggests that SIN helps maintain the function of the filtering bleb after GFS and prevent fibrosis. At present, studies on the molecular mechanism of SIN's antifibrotic effect on filtering bleb have mainly focused on the TGF- β /Smad, PI3K/AKT and NF- κ B signaling pathways.

4.3.1 TGF-β/Smad

TGF-B/Smad is a major pathway leading to scar formation, closely associated with ECM synthesis and fibroblast transdifferentiation (77, 78). Glaucoma patients exhibit elevated TGF-B concentrations in the aqueous humor and trabecular meshwork, suggesting that targeting TGF- β signaling represents a key therapeutic strategy for preventing postoperative fibrosis (79, 80). As a multifunctional dimeric polypeptide growth factor, TGF- β has been shown to promote proliferation, migration, and myofibroblast transdifferentiation of various target cells, and enhances the production of fibrosis-related proteins (81). The Smad protein family acts as TGF-B downstream intracellular effectors (82), with Smad2, Smad3, Smad4, and Smad7 being essential signaling components. Upon activation, TGF-β binds to the type II receptor (TBRII), which phosphorylates and activates the type I receptor (TBRI) kinase. This initiates the Smad-dependent signaling pathway, leading to phosphorylation and activation of downstream effectors such as Smad2 and Smad3, which mediate various biological effects (83-85). It was found that SIN-treated human fibroblasts not only exhibited cell cycle arrest but also underwent apoptosis. As the concentration of SIN increased, the number of apoptotic cells rose, accompanied by inhibition of TGF-B1 signaling. This was further evidenced by a decrease in the expression of Cyclin D1, a key regulator of the G1-to-S phase transition, and Bcl-2, an anti-apoptotic protein. In addition, the expression of MMP-2, which is associated with ECM remodeling and fibroblast migration, was also reduced. These findings indicate that SIN inhibits fibroblast proliferation and migration by regulating the TGF- β 1/Smad signaling pathway (74). On the one hand, SIN reduces TGF-^β release, inhibits its binding to cell surface receptors, and decreases ECM synthesis. On the other hand, it inhibits Smad protein phosphorylation, reduces their activation, and disrupts TGF- β signaling (86, 87).

4.3.2 PI3K/AKT

The PI3K/AKT pathway consists of phosphatidylinositol-3kinase (PI3K), protein kinase B (PKB), and its downstream molecules (88). PKB, also known as AKT, is a serine/threonine protein kinase and the principal downstream effector of the PI3K signaling pathway. Upon exogenous stimulation, PI3K is activated, leading to phosphorylation of AKT in cells and tissues. This activation induces a variety of biological effects, including the regulation of cell metabolism, growth, proliferation, and apoptosis. It also modulates oxidative stress, inflammatory responses, and energy homeostasis through multiple downstream targets (89). SIN attenuates renal fibrosis by modulating the PI3K-AKT pathway and affecting autophagy levels through BMSC-exo carrying miR-204-5p (90). Upon inhibition of PI3K activity, Akt phosphorylation and activation are reduced, which disrupts downstream signaling pathways, decreases cell proliferation and survival, and suppresses tissue fibrosis (91, 92).

4.3.3 NF-κB

NF-*κ*B is a family of transcription factors widely involved in regulation of cellular immunity, inflammation, proliferation, and apoptosis, primarily modulating inflammatory responses and cell survival (93, 94). As one of the classical inflammatory signaling pathways, NF-*κ*B induces the transcription of various proinflammatory cytokines, promotes ECM synthesis, and contributes to fibrosis (95). SIN inhibits the nuclear translocation of NF-*κ*B p65 subunit andthe DNA-binding activity of NF-*κ*B in synoviocytes, which might be one of anti-inflammatory mechanisms (96). SIN increases A2A receptor and suppresses NF-*κ*B pathway activation via the α 7 nicotinic acetylcholine receptor in adjuvant-induced arthritis rats (97, 98). The inflammatory response is a key contributor to fibrosis. Suppressing the expression of inflammatory cytokines helps slow fibrotic progression, and SIN's antifibrotic effect is likely mediated through inhibition of NF-*κ*B signaling.

5 Discussion and conclusion

GFS remains the gold-standard intervention for refractory glaucoma, but postoperative fibrosis of the filtering bleb is a leading cause of surgical failure. The pathogenesis of filtering bleb fibrosis is multifactorial, involving inflammatory, fibroblast proliferation and transdifferentiation, ECM deposition, aberrant angiogenesis, and pro-fibrotic cytokine signaling. This process is originally part of postoperative injury healing, but excessive fibrosis may lead to filtering bleb dysfunction. Although current therapies show some efficacy, they are frequently associated with considerable toxicity and adverse effects. Furthermore, several emerging therapies remain at the experimental stage, and the development of safe and effective agents to prevent or attenuate filtering bleb fibrosis after GFS remains a significant challenge.

Fibrosis of the filtering bleb after GFS shares a similar pathological mechanism with other fibrotic diseases. Although

no studies have reported the effect of SIN on filtering bleb fibrosis, analysis of fibrosis mechanisms and SIN's pharmacological properties suggests its therapeutic potential. SIN exerts antiinflammatory effects by reducing inflammatory reactions, decreasing inflammatory cell infiltration and cytokine release, thereby inhibiting fibrosis at the injury site. SIN inhibits the activation, migration, and proliferation of fibroblasts while promoting their apoptosis. It suppresses the transdifferentiation of fibroblasts into myofibroblasts. It decreases the expression of α -SMA, reduces protein deposition, and promotes collagen degradation. SIN's antifibrotic effects on filtering bleb fibrosis after GFS may involve the TGF- β /Smad, PI3K/Akt, and NF- κ B signaling pathways.

The strong histamine-releasing effects of SIN may induce skin edema, pruritus, and gastrointestinal reactions (99), suggesting the need to be vigilant for patients with asthma and a history of allergy during clinical application. The study showed (100) that the distribution pattern of SIN in organs after multiple administrations was similar to that of a single administration, and no drug residues were detected in any tissue after 1 week of drug withdrawal, suggesting no accumulation in the body. It is worth noting that SIN concentrations are highest in the liver, and lower in the heart. Histological observation suggests relatively obvious morphological changes in liver tissue. However, no significant abnormalities were detected in liver function tests (ALT, AST), renal function (BUN), or urinary sediment in rats following 6 weeks of continuous administration and 1 week of withdrawal. This indicates that high-dose SIN did not compromise hepatic or renal enzymatic profiles, despite causing subtle histological alterations in the liver. Prolonged oral administration of SIN may induce hepatotoxicity, nephrotoxicity, and cardiotoxicity. Regular monitoring is therefore recommended for patients with severe hepatic or cardiovascular diseases. Additionally, caution is advised when using SIN in patients undergoing systemic immunosuppressive therapy for autoimmune diseases, as well as in pregnant or breastfeeding women (101, 102).

The safety profile of Sinomenine in ophthalmic applications is currently under investigation, with topical ocular administration considered effective for mitigating the aforementioned risks. After treatment with 0.05 and 0.1% Sinomenine (SIN) eye drops administered four times daily, slit lamp examination and corneal staining revealed no significant ocular irritation or corneal damage in mouse dry eye models (67). In rabbit uveitis models, administration of 0.5% SIN solution or gel showed no significant ocular irritation. Histopathological examination further confirmed that neither corneal epithelial detachment nor stromal damage occurred after treatment with 0.5% SIN solution or gel (103). In another study (104), the use of 1% SIN eye drops (four times daily for 7 days) significantly inhibited inflammation, reduced neovascularization, and promoted epithelial repair, effectively treating acute phase damage to the cornea in alkali burns. No corneal opacity, iris inflammation, or persistent conjunctival hyperemia was observed during the trial. The corneal epithelium and stromal structures remained intact with no pathological damage. However, the study lasted only 7 days, so long-term observation is still needed to explore its effect on subsequent scar formation.

By analyzing the mechanism of fibrosis in filtering bleb formation, current therapies, and SIN's pharmacological effects, SIN shows potential to maintain filtering bleb function after GFS. This analysis offers novel perspectives for studying anti-fibrotic strategies targeting the filtering bleb. Further studies are required to evaluate SIN's efficacy and safety, with the aim of providing a better therapeutic option for glaucoma surgery patients.

Author contributions

XX: Writing – review & editing, Writing – original draft. XL: Writing – review & editing, Writing – original draft. PZ: Writing – original draft, Writing – review & editing. CW: Writing – review & editing, Writing – original draft. QP: Conceptualization, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was supported by the National Natural Science Foundation of China (82274588), Hunan Provincial Health High-Level Talent Scientific Research Project (grant number: R2023099), Talent Support Program of the State Administration of Traditional Chinese Medicine-Qihuang Scholars (Letter of the State Administration of Traditional Chinese Medicine [2022] No. 6), "Academician Liu Liang's Workstation" Guidance Project (21YS002), Hunan Province Health and Healthcare High-level Talents Major Scientific Research Special Grant (R2023099), Chinese Medicine Prevention and Treatment of Eye, Ear, Nose and Throat Diseases Hunan Province Key Laboratory Open Fund Project (2018YZD02) and Construction Project of Hunan Provincial Key Laboratory of Traditional Chinese Medicine for Prevention and Treatment of five senses Diseases (2017TP1018).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Gen AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Kyari F, Entekume G, Rabiu M, Spry P, Wormald R, Nolan W, et al. A populationbased survey of the prevalence and types of glaucoma in Nigeria: results from the Nigeria National Blindness and visual impairment survey. *BMC Ophthalmol.* (2015) 15:176. doi: 10.1186/s12886-015-0160-6

2. Jung KI, Woo JE, Park CK. Effects of aqueous suppressants and prostaglandin analogues on early wound healing after glaucoma implant surgery. *Sci Rep.* (2019) 9:5251. doi: 10.1038/s41598-019-41790-1

3. Parikh KS, Josyula A, Omiadze R, Ahn JY, Ha Y, Ensign LM, et al. Nanostructured glaucoma drainage implant safely and significantly reduces intraocular pressure in rabbits via post-operative outflow modulation. *Sci Rep.* (2020) 10:12911. doi: 10.1038/s41598-020-69687-4

4. Tham Y-C, Li X, Wong TY, Quigley HA, Aung T, Cheng C-Y. Global prevalence of glaucoma and projections of glaucoma burden through 2040. *Ophthalmology*. (2014) 121:2081–90. doi: 10.1016/j.ophtha.2014.05.013

5. Zhang N, Wang J, Li Y, Jiang B. Prevalence of primary open angle glaucoma in the last 20 years: a meta-analysis and systematic review. *Sci Rep.* (2021) 11:13762. doi: 10.1038/s41598-021-92971-w

6. Križaj D. "What is glaucoma?" In: Kolb H, Fernandez E, Jones B, Nelson R, editors. *Webvision: The Organization of the Retina and Visual System*. Salt Lake City, UT: University of Utah Health Sciences Center (2019). p. 1586. Available online at: http://www.ncbi.nlm.nih.gov/books/NBK543075/ (Accessed June 27, 2025).

7. Lusthaus J, Goldberg I. Current management of glaucoma. Med J Aust. (2019) 210:180-7. doi: 10.5694/mja2.50020

8. Henderson NC, Rieder F, Wynn TA. Fibrosis: from mechanisms to medicines. *Nature*. (2020) 587:555–66. doi: 10.1038/s41586-020-2938-9

9. O'Regan A, O'Brien CJ, Eivers SB. The lysophosphatidic acid axis in fibrosis: Implications for glaucoma. *Wound Repair Regen*. (2021) 29:613–26. doi: 10.1111/wrr.12929

10. Peng Y-J, Pan C-Y, Hsieh Y-T, Wu S, Chu H-N, Hsu W-C. The application of tissue engineering in reversing mitomycin C-induced ischemic conjunctiva. *J Biomed Mater Res A*. (2012) 100A:1126–35. doi: 10.1002/jbm.a.34041

11. Joshi AB, Parrish RK, Feuer WF. 2002 Survey of the American Glaucoma Society: practice preferences for glaucoma surgery and antifibrotic use. *J Glaucoma*. (2005) 14:172–4. doi: 10.1097/01.ijg.0000151684.12033.4d

12. Hou W, Huang L, Huang H, Liu S, Dai W, Tang J, et al. Bioactivities and mechanisms of action of sinomenine and its derivatives: a comprehensive review. *Molecules.* (2024) 29:540. doi: 10.3390/molecules29020540

13. Shu DY, Lovicu FJ. Myofibroblast transdifferentiation: the dark force in ocular wound healing and fibrosis. *Prog Retin Eye Res.* (2017) 60:44–65. doi: 10.1016/j.preteyeres.2017.08.001

14. Sacchi M, Tomaselli D, Ruggeri ML, Aiello FB, Sabella P, Dore S, et al. Fighting bleb fibrosis after glaucoma surgery: updated focus on key players and novel targets for therapy. *Int J Mol Sci.* (2025) 26:2327. doi: 10.3390/ijms26052327

15. Zhao X, Kwan JYY, Yip K, Liu PP, Liu F-F. Targeting metabolic dysregulation for fibrosis therapy. *Nat Rev Drug Discov.* (2020) 19:57–75. doi: 10.1038/s41573-019-0040-5

16. Zada M, Pattamatta U, White A. Modulation of fibroblasts in conjunctival wound healing. *Ophthalmology*. (2018) 125:179–92. doi: 10.1016/j.ophtha.2017.08.028

17. Kakizaki H, Takahashi Y, Nakano T, Asamoto K, Ikeda H, Ichinose A, et al. Anatomy of Tenons capsule. *Clin Exper Ophthalmol.* (2012) 40:611-6. doi: 10.1111/j.1442-9071.2011.02745.x

18. Roodnat AW, Callaghan B, Doyle C, Vallabh NA, Atkinson SD, Willoughby CE. Genome-wide RNA sequencing of ocular fibroblasts from glaucomatous and normal eyes: Implications for glaucoma management. *PLoS ONE.* (2024) 19:e0307227. doi: 10.1371/journal.pone.0307227

19. Qian L-W, Fourcaudot AB, Yamane K, You T, Chan RK, Leung KP. Exacerbated and prolonged inflammation impairs wound healing and increases scarring: excess inflammation deteriorates wound outcomes. *Wound Rep and Reg.* (2016) 24:26–34. doi:10.1111/wrr.12381

20. Cunliffe IA, Richardson PS, Rees RC, Rennie IG. Effect of TNF, IL-1, and IL-6 on the proliferation of human Tenon's capsule fibroblasts in tissue culture. *Br J Ophthalmol.* (1995) 79:590–5. doi: 10.1136/bjo.79.6.590

21. He Y-H, Zhang H-N, Zhang G-P, Hou N, Xiao Q, Huang Y, et al. A physiological concentration of glucocorticoid inhibits the pro-inflammatory cytokineinduced proliferation of adult rat cardiac fibroblasts: roles of extracellular signalregulated kinase 1/2 and nuclear factor-κB: GC and cardiac fibroblast proliferation. *Clin Exp Pharmacol Physiol.* (2011) 38:739–46. doi: 10.1111/j.1440-1681.2011.05581.x

22. Gater R, Ipek T, Sadiq S, Nguyen D, Jones L, El Haj A, et al. Investigation of conjunctival fibrosis response using a 3D glaucoma Tenon's capsule + conjunctival model. *Invest Ophthalmol Vis Sci.* (2019) 60:605. doi: 10.1167/iovs.18-25335

23. Liu L, Zhou W, Fan Y, Zhang L, Liu S, Song S, et al. Effect of interleukin 6 on scleral fibroblast proliferation, differentiation, and apoptosis involved in myopic scleral remodeling. *Ophthalmic Res.* (2022) 65:529–39. doi: 10.1159/0005 24502

24. Montero P, Milara J, Roger I, Cortijo J. Role of JAK/STAT in interstitial lung diseases; molecular and cellular mechanisms. *Int J Mol Sci.* (2021) 22:6211. doi: 10.3390/ijms22126211

25. Wilson SE. Topical losartan: practical guidance for clinical trials in the prevention and treatment of corneal scarring fibrosis and other eye diseases and disorders. *J Ocul Pharmacol Ther.* (2023) 39:191–206. doi: 10.1089/jop.2022.0174

26. Zhang S, Yao Y, Zheng Y, Wu P, Zhu Y. Effect of biological amniotic membrane soaked in pirfenidone on scar formation after glaucoma filtration surgery in rabbit eyes. *Int Eye Sci.* (2024) 24:189–95. Available online at: https://pesquisa.bvsalud.org/portal/resource/pt/wpr-1005379

27. Jiang C, Yin X, Chang J, Liu G, Lu P. Topical administration of bevacizumab to facilitate the functional filtering bleb after trabeculectomy in the rabbit. *J Ocul Pharmacol Therap.* (2023) 39:716–24. doi: 10.1089/jop.2023.0046

28. Siedlecki J, Asani B, Wertheimer C, Hillenmayer A, Ohlmann A, Priglinger C, et al. Combined VEGF/PDGF inhibition using axitinib induces α SMA expression and a pro-fibrotic phenotype in human pericytes. *Graefes Arch Clin Exp Ophthalmol.* (2018) 256:1141–9. doi: 10.1007/s00417-018-3987-8

29. Uutela M, Wirzenius M, Paavonen K, Rajantie I, He Y, Karpanen T, et al. PDGF-D induces macrophage recruitment, increased interstitial pressure, and blood vessel maturation during angiogenesis. *Blood.* (2004) 104:3198–204. doi: 10.1182/blood-2004-04-1485

30. Weiskirchen R, Weiskirchen S, Tacke F. Organ and tissue fibrosis: molecular signals, cellular mechanisms and translational implications. *Mol Aspects Med.* (2019) 65:2–15. doi: 10.1016/j.mam.2018.06.003

31. Bergen TV, Velde SV de, Vandewalle E, Moons L, Stalmans I. Improving patient outcomes following glaucoma surgery: state of the art and future perspectives. *Clin Ophthalmol.* (2014) 8:857–67. doi: 10.2147/OPTH.S48745

32. Koval MS, Moster MR, Freidl KB, Waisbourd M, Jain SG, Ichhpujani P, et al. Intracameral triamcinolone acetonide in glaucoma surgery: a prospective randomized controlled trial. *Am J Ophthalmol.* (2014) 158:395–401.e2. doi: 10.1016/j.ajo.2014.04.027

33. Swogger J, Conner IP, Rosano M, Kemmerer M, Happ-Smith C, Wells A, et al. Injected vs. sponge-applied mitomycin c (MMC) during modified trabeculectomy in New Zealand white rabbit model. *Trans Vis Sci Tech.* (2020) 9:23. doi: 10.1167/tvst.9.11.23

34. Shao CG, Sinha NR, Mohan RR, Webel AD. Novel therapies for the prevention of fibrosis in glaucoma filtration surgery. *Biomedicines.* (2023) 11:657. doi: 10.3390/biomedicines11030657

35. Bell K, De Padua Soares Bezerra B, Mofokeng M, Montesano G, Nongpiur ME, Marti MV, et al. Learning from the past: mitomycin C use in trabeculectomy and its application in bleb-forming minimally invasive glaucoma surgery. *Surv Ophthalmol.* (2021) 66:109–23. doi: 10.1016/j.survophthal.2020.05.005

36. Palanca-Capistrano AM, Hall J, Cantor LB, Morgan L, Hoop J, WuDunn D. Long-term outcomes of intraoperative 5-fluorouracil vs. intraoperative mitomycin C in primary trabeculectomy surgery. *Ophthalmology.* (2009) 116:185–90. doi: 10.1016/j.ophtha.2008.08.009

37. Wolters JEJ, Van Mechelen RJS, Al Majidi R, Pinchuk L, Webers CAB, Beckers HJM, et al. History, presence, and future of mitomycin C in glaucoma filtration surgery. *Curr Opin Ophthalmol.* (2021) 32:148–59. doi: 10.1097/ICU.0000000000 00729

38. Anand N, Arora S, Clowes M. Mitomycin C augmented glaucoma surgery: evolution of filtering bleb avascularity, transconjunctival oozing, and leaks. Br J Ophthalmol. (2006) 90:175–80. doi: 10.1136/bjo.2005.077800

39. Fakhraie G, Lopes JF, Spaeth GL, Almodin J, Ichhpujani P. Moster MR. Effects of postoperative cyclosporine ophthalmic emulsion 005% (Restasis) following glaucoma surgery. *Clin Exp Ophthalmol.* (2009) 37:842–8. doi: 10.1111/j.1442-9071.2009.02134.x

40. Song Y, Wang F, Luo H, Hu H, Pang Y, Xu K, et al. Rapamycin protects glucocorticoid-induced glaucoma model mice against trabecular meshwork fibrosis by suppressing mTORC1/2 signaling. *Eur J Pharmacol.* (2025) 990:177269. doi: 10.1016/j.ejphar.2025.177269

41. Lin X, Wen J, Liu R, Gao W, Qu B, Yu M. Nintedanib inhibits TGF- β -induced myofibroblast transdifferentiation in human Tenon's fibroblasts. *Mol Vis.* (2018) 24:789–800.

42. José P, Teixeira FJ, Barão R, Sousa DC, Marques RE, Barata ADDO, et al. Trabeculectomy with mitomycin C alone or coupled with intracamerular bevacizumab? A 2-year comparative study. *Br J Ophthalmol.* (2022) 106:1399–405. doi: 10.1136/bjophthalmol-2021-319039

43. Zhang Y, Zhu S, Xu X, Zuo L. *In vitro* study of combined application of bevacizumab and 5-fluorouracil or bevacizumab and mitomycin C to inhibit scar formation in glaucoma filtration surgery. *J Ophthalmol.* (2019) 2019:7419571. doi: 10.1155/2019/7419571

44. Hwang YH, Jung S-A, Lyu J, Kim YY, Lee JH. Transforming growth factor-β1induced human subconjunctival fibrosis is mediated by microRNA 143/145 expression. *Invest Ophthalmol Vis Sci.* (2019) 60:2064. doi: 10.1167/iovs.19-26797

45. Zhang F, Liu K, Cao M, Qu J, Zhou D, Pan Z, et al. Rosiglitazone treatment prevents postoperative fibrosis in a rabbit model of glaucoma filtration surgery. *Invest Ophthalmol Vis Sci.* (2019) 60:2743–52. doi: 10.1167/iovs.18-26526

46. Yu A, Shi H, Liu H, Bao Z, Dai M, Lin D, et al. Mucoadhesive dexamethasoneglycol chitosan nanoparticles for ophthalmic drug delivery. *Int J Pharm.* (2020) 575:118943. doi: 10.1016/j.ijpharm.2019.118943

47. Tatischeff I, Alfsen A. A new biological strategy for drug delivery: eucaryotic cell-derived nanovesicles. *J Biomater Nanobiotechnol.* (2011) 02:494. doi: 10.4236/jbnb.2011.225060

48. van den Boorn JG, Schlee M, Coch C, Hartmann G. SiRNA delivery with exosome nanoparticles. *Nat Biotechnol.* (2011) 29:325-6. doi: 10.1038/nbt.1830

49. Yates-Binder CC, Rodgers M, Jaynes J, Wells A, Bodnar RJ, Turner T. An IP-10 (CXCL10)-derived peptide inhibits angiogenesis. *PLoS ONE.* (2012) 7:e40812. doi: 10.1371/journal.pone.0040812

50. Swogger J, Conner IP, Happ-Smith C, Kemmerer MC, Julian DR, Davis R, et al. Novel combination therapy reduces subconjunctival fibrosis after glaucoma filtration surgery in the rabbit model. *Clin Exp Ophthalmol.* (2021) 49:60–9. doi: 10.1111/ceo.13884

51. Fujimoto T, Inoue-Mochita M, Inoue T. A ROCK inhibitor suppresses the transforming growth factor-beta-2-induced endothelial-mesenchymal transition in Schlemm's canal endothelial cells. *Sci Rep.* (2023) 13:9655. doi: 10.1038/s41598-023-36808-8

52. Kishimoto T, Ishida W, Nakajima I, Taguchi O, Sugioka K, Kusaka S, et al. Promotion of conjunctival fibroblast-mediated collagen gel contraction by mast cells through up-regulation of matrix metalloproteinase release and activation. *Exp Eye Res.* (2022) 218:108980. doi: 10.1016/j.exer.2022.108980

53. Shi F, Shi H, Li Z. Low dosage of homoharringtonine for prevention of cicatrization after glaucoma filtering surgery. *Zhonghua Yan Ke Za Zhi*. (1995) 31:345–6. Available online at: https://europepmc.org/article/med/8706581

54. Li D, Sun M, Li Q, Yang F, Lin S, Wu K. Effect of tetrandrine on bax, bcl-2 and TGF-β2 mRNA expressions in cultured human Tenon's capsule fibroblasts. *Nan Fang Yi Ke Da Xue Xue Bao.* (2012) 32:97–100. Available online at: https://europepmc.org/ article/med/22366014

55. Liu S, Chen L. Inhibitive effects of quercetin on rabbit Tenon capsule fibroblasts proliferation. *Yan Ke Xue Bao.* (2005) 21:175–8. Available online at: https://europepmc.org/article/med/17162857

56. Qi B, Zhao Y, Wei X, Xiao W, Zheng H, Chen Z, et al. A further investigation concerning correlation between anti-fibrotic effect of liposomal quercetin and inflammatory cytokines in pulmonary fibrosis. *Eur J Pharmacol.* (2010) 642:134–9. doi: 10.1016/j.ejphar.2010.05.019

57. Zhao X-X, Peng C, Zhang H, Qin L-P. *Sinomenium acutum* : a review of chemistry, pharmacology, pharmacokinetics, and clinical use. *Pharm Biol.* (2012) 50:1053-61. doi: 10.3109/13880209.2012.656847

58. Jabbar AA j, Ahmed KA-A, Abdulla MA, Abdullah FO, Salehen NA, Mothana RA, et al. Sinomenine accelerate wound healing in rats by augmentation of antioxidant, anti-inflammatory, immunuhistochemical pathways. *Heliyon.* (2024) 10:e23581. doi: 10.1016/j.heliyon.2023.e23581

59. Liu Y, Chen H, Wu Y, Ai F, Li W, Peng H, et al. Sinomenine attenuates bleomycin-induced pulmonary fibrosis, inflammation, and oxidative stress by inhibiting TLR4/NLRP3/TGF β signaling. *Inhal Toxicol.* (2024) 36:217–27. doi: 10.1080/08958378.2024.2335193

60. Nie Z, Wu J, Xie J, Yin W. Sinomenine ameliorates bleomycin-induced pulmonary fibrosis by inhibiting the differentiation of fibroblast into myofibroblast. *Heliyon.* (2024) 10:e33314. doi: 10.1016/j.heliyon.2024.e33314

61. Cheng Z, Liangkun W. Improvement effects and mechanism of sinomenine on non-alcoholic steatohepatitis in mice. *zggx.* (2024) 35:1701-7. doi: 10.6039/j.issn.1001-0408.2024.14.05

62. Li J-J, Li L, Li S, Tang X-Y, Sun H-F, Liu J-X. Sinomenine hydrochloride protects IgA nephropathy through regulating cell growth and apoptosis of T and B lymphocytes. *Drug Des Devel Ther.* (2024) 18:1247–62. doi: 10.2147/DDDT.S449119

63. Qin T, Yin S, Yang J, Zhang Q, Liu Y, Huang F, et al. Sinomenine attenuates renal fibrosis through Nrf2-mediated inhibition of oxidative stress and TGF β signaling. *Toxicol Appl Pharmacol.* (2016) 304:1–8. doi: 10.1016/j.taap.2016.05.009

64. Li D, Zhong Z, Ko C-N, Tian T, Yang C. From mundane to classic: sinomenine as a multi-therapeutic agent. *Br J Pharmacol.* (2025) 182:2159–80. doi: 10.1111/bph.16267

65. Liu W, Zhang Y, Zhu W, Ma C, Ruan J, Long H, et al. Sinomenine inhibits the progression of rheumatoid arthritis by regulating the secretion of

inflammatory cytokines and monocyte/macrophage subsets. Front Immunol. (2018) 9:2228. doi: 10.3389/fimmu.2018.02228

66. Zheng X, Li W, Xu H, Liu J, Ren L, Yang Y, et al. Sinomenine ester derivative inhibits glioblastoma by inducing mitochondria-dependent apoptosis and autophagy by PI3K/AKT/mTOR and AMPK/mTOR pathway. *Acta Pharmaceutica Sinica B.* (2021) 11:3465–80. doi: 10.1016/j.apsb.2021.05.027

67. Li H, Wei F, Li S, Yan L, Lu P. The effect of sinomenine eye drops on experimental dry eye in mice. *Cutan Ocul Toxicol.* (2020) 39:389–95. doi: 10.1080/15569527.2020.1840580

68. Zhang W-J, Chen S-J, Zhou S-C, Wu S-Z, Wang H. Inflammasomes and fibrosis. Front Immunol. (2021) 12:643149. doi: 10.3389/fimmu.2021.643149

69. Yin N, Xiong Y, Tao W, Chen J. Wang Z. Sinomenine alleviates lipopolysaccharide-induced inflammatory responses in RAW2647 macrophages. *Immunopharmacol Immunotoxicol.* (2020) 42:147– 55. doi: 10.1080/08923973.2020.1732407

70. Wang Y, Fang Y, Huang W, Zhou X, Wang M, Zhong B, et al. Effect of sinomenine on cytokine expression of macrophages and synoviocytes in adjuvant arthritis rats. *J Ethnopharmacol.* (2005) 98:37–43. doi: 10.1016/j.jep.2004.12.022

71. Yao F, Xu M, Dong L, Shen X, Shen Y, Jiang Y, et al. Sinomenine attenuates pulmonary fibrosis by downregulating TGF- β 1/Smad3, PI3K/Akt and NF- κ B signaling pathways. *BMC Pulm Med.* (2024) 24:229. doi: 10.1186/s12890-024-03050-5

72. Liu L, Qian H, Meng Q, Zhang X, Wei Y, He J. [Sinomenine ameliorates bleomycin A5-induced pulmonary fibrosis by blocking the miR-21/ADAMTS-1 signaling pathway in rats]. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi.* (2023) 39:721–8. Available online at: https://europepmc.org/article/med/37515339

73. Li J, Fu T, Zhang GF, et al. Sinomenine induces apoptosis of human rheumatoid arthritis fibroblast-like synoviocytes via miR-23b-3p/FGF9 signaling pathway. *Chin J Pathophysiol.* (2020) 36:1653–60.

74. Li HB. Effects of cymbidium alkaloids on the biological function of proliferative scar fibroblasts [master's thesis]. Jinzhou Medical University, Jinzhou (2019).

75. Xie N, Tan Z, Banerjee S, Cui H, Ge J, Liu R-M, et al. Glycolytic reprogramming in myofibroblast differentiation and lung fibrosis. *Am J Respir Crit Care Med.* (2015) 192:1462–74. doi: 10.1164/rccm.201504-0780OC

76. Stahnke T, Löbler M, Kastner C, Stachs O, Wree A, Sternberg K, et al. Different fibroblast subpopulations of the eye: a therapeutic target to prevent postoperative fibrosis in glaucoma therapy. *Exp Eye Res.* (2012) 100:88–97. doi: 10.1016/j.exer.2012.04.015

77. Kramer EL, Clancy JP. TGF β as a therapeutic target in cystic fibrosis. *Expert Opin Ther Targets.* (2018) 22:177–89. doi: 10.1080/14728222.2018.1406922

78. Rajabi S, Saberi S, Najafipour H, Askaripour M, Rajizadeh MA, Shahraki S, et al. Interaction of estradiol and renin-angiotensin system with microRNAs-21 and–29 in renal fibrosis: focus on TGF- β /smad signaling pathway. *Mol Biol Rep.* (2024) 51:137. doi: 10.1007/s11033-023-09127-4

79. Pasquale LR, Dorman-Pease ME, Lutty GA, Quigley HA, Jampel HD. Immunolocalization of TGF-beta 1, TGF-beta 2, and TGF-beta 3 in the anterior segment of the human eye. *Invest Ophthalmol Vis Sci.* (1993) 34:23–30. Available online at: https://iovs.arvojournals.org/article.aspx?articleid=2179146

80. Tovar-Vidales T, Clark AF, Wordinger RJ. Transforming growth factor-beta2 utilizes the canonical Smad-signaling pathway to regulate tissue transglutaminase expression in human trabecular meshwork cells. *Exp Eye Res.* (2011) 93:442–51. doi: 10.1016/j.exer.2011.06.011

 Khalil H, Kanisicak O, Prasad V, Correll RN, Fu X, Schips T, et al. Fibroblastspecific TGF-β-Smad2/3 signaling underlies cardiac fibrosis. J Clin Invest. (2017) 127:3770–83. doi: 10.1172/JCI94753

82. Luo K. Signaling Cross Talk between TGF-β/Smad and other signaling pathways. Cold Spring Harb Perspect Biol. (2017) 9:a022137. doi: 10.1101/cshperspect.a022137

83. Xu F, Liu C, Zhou D, Zhang L. TGF-β/SMAD pathway and its regulation in hepatic fibrosis. J Histochem Cytochem. (2016) 64:157-67. doi: 10.1369/0022155415627681

84. Liang R, Šumová B, Cordazzo C, Mallano T, Zhang Y, Wohlfahrt T, et al. The transcription factor GLI2 as a downstream mediator of transforming growth factor- β -induced fibroblast activation in SSc. Ann Rheum Dis. (2017) 76:756–64. doi: 10.1136/annrheumdis-2016-209698

85. Tomcik M, Palumbo-Zerr K, Zerr P, Sumova B, Avouac J, Dees C, et al. Tribbles homologue 3 stimulates canonical TGF- β signalling to regulate fibroblast activation and tissue fibrosis. Ann Rheum Dis. (2016) 75:609–16. doi: 10.1136/annrheumdis-2014-206234

86. Aashaq S, Batool A, Mir SA, Beigh MA, Andrabi KI, Shah ZA. TGF- β signaling: a recap of SMAD-independent and SMAD-dependent pathways. *J Cell Physiol.* (2022) 237:59–85. doi: 10.1002/jcp.30529

87. He H, Cao L, Wang Z, Wang Z, Miao J, Li X-M, et al. Sinomenine relieves airway remodeling by inhibiting epithelial-mesenchymal transition through downregulating TGF- β 1 and Smad3 expression *in vitro* and *in vivo*. Front Immunol. (2021) 12:736479. doi: 10.3389/fimmu.2021.736479

88. Wang J, Hu K, Cai X, Yang B, He Q, Wang J, et al. Targeting PI3K/AKT signaling for treatment of idiopathic pulmonary fibrosis. *Acta Pharmaceutica Sinica B.* (2022) 12:18–32. doi: 10.1016/j.apsb.2021.07.023

89. Chen K, Li Y, Zhang X, Ullah R, Tong J, Shen Y. The role of the PI3K/AKT signalling pathway in the corneal epithelium: recent updates. *Cell Death Dis.* (2022) 13:1–13. doi: 10.1038/s41419-022-04963-x

90. Gu H, Li J, Ni Y. Sinomenine improves renal fibrosis by regulating mesenchymal stem cell-derived exosomes and affecting autophagy levels. *Environ Toxicol.* (2023) 38:2524–37. doi: 10.1002/tox.23890

91. He Z, Deng Y, Li W, Chen Y, Xing S, Zhao X, et al. Overexpression of PTEN suppresses lipopolysaccharide-induced lung fibroblast proliferation, differentiation and collagen secretion through inhibition of the PI3-K-Akt-GSK3beta pathway. *Cell Biosci.* (2014) 4:2. doi: 10.1186/2045-3701-4-2

92. Song B, Li X-F, Yao Y, Xu Q-Q, Meng X-M, Huang C, et al. BMP9 inhibits the proliferation and migration of fibroblast-like synoviocytes in rheumatoid arthritis via the PI3K/AKT signaling pathway. *Int Immunopharmacol.* (2019) 74:105685. doi: 10.1016/j.intimp.2019.105685

93. Zhang Q, Lenardo MJ, Baltimore D. 30 years of NF-KB: a blossoming of relevance to human pathobiology. *Cell.* (2017) 168:37–57. doi: 10.1016/j.cell.2016.12.012

94. Ilchovska D. (Daisy), Barrow DM. An overview of the NF-kB mechanism of pathophysiology in rheumatoid arthritis, investigation of the NF-kB ligand RANKL and related nutritional interventions. *Autoimmun Rev.* (2021) 20:102741. doi: 10.1016/j.autrev.2020.102741

95. Luedde T, Schwabe RF. NF-κB in the liver—linking injury, fibrosis and hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol.* (2011) 8:108-18. doi: 10.1038/nrgastro.2010.213

96. Huang X, Hao F, Wang Y, Fang Y. Inhibition of sinomenine on nuclear factor- κ B of synoviocytes in collagen-induced arthritis rats. *Acta Acad. Med. Mil. Tertiae J. Army Med. Univ.* (2007) 13:1269–72. Available online at: https://europepmc.org/article/cba/639726

97. Yi L, Ke J, Liu J, Lai H, Lv Y, Peng C, et al. Sinomenine increases adenosine A2A receptor and inhibits NF-κB to inhibit arthritis in adjuvant-induced-arthritis rats and fibroblast-like synoviocytes through α7nAChR. J Leukoc Biol. (2021) 110:1113–20. doi: 10.1002/JLB.3MA0121-024RRRR

98. Ru-Bing Yao R-BY, Zhi-Ming Zhao Z-MZ, Ling-Jie Zhao L-JZ, Hui Cai HC. Sinomenine inhibits the inflammatory responses of human fibroblast-like synoviocytes via the TLR4/MyD88/NF-kB signaling pathway in rheumatoid arthrits. *Pharmazie*. (2017) 72:355–360. doi: 10.1691/ph.2017.6946

99. Duan D, Wang Y, Pan D, Jin X, Yan Y, Song P, et al. Rheumatoid arthritis drug sinomenine induces apoptosis of cervical tumor cells by targeting thioredoxin reductase *in vitro* and *in vivo*. *Bioorg Chem.* (2022) 122:105711. doi: 10.1016/j.bioorg.2022.105711

100. Ye M, Liu L, Zeng Y, Zhang L, Tan Y, Deng S, et al. Studies on the relationship between sinomenine distribution and its organic toxicology. *Chin Pharmacol Bullet*. (2001) 12: 65–9. Available online at: https://pesquisa.bvsalud.org/portal/resource/pt/ wpr-412076

101. Tang J, Raza A, Chen J, Xu H, A. Systematic review on the sinomenine derivatives. *MRMC*. (2018) 18:906–17. doi: 10.2174/1389557517666171123212 557

102. Chen X, Huang R, Huang Q, Chu Y, Yan J. Systemic review and meta-analysis of the clinical efficacy and adverse effects of Zhengqing Fengtongning combined with methotrexate in rheumatoid arthritis. *Evid Based Complement Alter Med.* (2015) 2015;910376. doi: 10.1155/2015/910376

103. Song J, Bi H, Xie X, Guo J, Wang X, Liu D. Preparation and evaluation of sinomenine hydrochloride *in situ* gel for uveitis treatment. *Int Immunopharmacol.* (2013) 17:99–107. doi: 10.1016/j.intimp.2013.05.020

104. Konuk SG, Özdemir S, Kiliç R, Türkyilmaz B, Bektur G, Güneş A. Exploring the therapeutic potential: sinomenine and melatonin in alkali-induced corneal burns. *Int Ophthalmol.* (2024) 44:422. doi: 10.1007/s10792-024-033 27-y