

OPEN ACCESS

EDITED BY Giuseppe Castaldo, University of Naples Federico II, Italy

REVIEWED BY Chonin Cheana. Macau Society for Health Economics, Macao SAR, China Yujun Xiong, Peking University, China Xiaopeng Ma, Hebei Medical University, China

*CORRESPONDENCE Shenglan Tian ≥ 2480596805@qq.com

†These authors share first authorship

RECEIVED 07 May 2025 ACCEPTED 13 August 2025 PUBLISHED 29 August 2025

Wang K, Zheng J, Yin S, Zhang H, Yan H, Jin Y, Qiu Y, Wang X, Zhu X, Yao L and Tian S (2025) Association between Helicobacter pylori infection and serum uric acid levels in a Chinese community population: a cross-sectional study stratified by renal function. Front. Med. 12:1615161.

doi: 10.3389/fmed.2025.1615161

© 2025 Wang, Zheng, Yin, Zhang, Yan, Jin, Qiu, Wang, Zhu, Yao and Tian. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms

Association between Helicobacter pylori infection and serum uric acid levels in a Chinese community population: a cross-sectional study stratified by renal function

Keding Wang^{1†}, Jiaojiao Zheng^{1†}, Shali Yin^{1†}, Hui Zhang^{1†}, Hongying Yan¹, Yuling Jin¹, Yuzhen Qiu¹, Xvru Wang², Xinyu Zhu², Lan Yao³ and Shenglan Tian^{1*}

¹Department of General Medicine, Tianyou Hospital Affiliated to Wuhan University of Science and Technology, Wuhan, China, ²School of Clinical Medicine, Wuhan University of Science and Technology, Wuhan, China, ³Hongshan Street Community Health Service Center Affiliated to Wuhan University of Science and Technology, Wuhan, China

Background: The relationship between Helicobacter pylori (H. pylori) infection and serum uric acid levels remains debated. This study investigates the association between H. pylori infection and serum uric acid levels in a Chinese community, exploring renal function as a potential modifier.

Methods: We conducted a cross-sectional study involving 8,439 adults who underwent health examinations at a hospital in Wuhan from January 2022 to January 2024. H. pylori infection was assessed via the 14C-urea breath test, and serum uric acid levels were measured by the uricase method. Multivariable linear regression models evaluated the associations, and interaction analysis identified potential effect modifiers. Subgroup analyses were stratified by estimated glomerular filtration rate (eGFR).

Results: The prevalence of H. pylori infection was 21.5% (1,816/8,439). Initial analysis showed higher serum uric acid levels in individuals with H. pylori infection compared to those without $(403.76 \pm 102.89 \text{ vs. } 395.87 \pm 102.13 \,\mu\text{mol/L},$ p = 0.004). However, after adjusting for age, sex, body mass index, lipid profiles, and hepatorenal function, this association was no longer significant in the overall cohort ($\beta = 1.92$, 95% CI: -2.38 to 6.23, p = 0.381). Interaction analysis revealed a significant modification by eGFR (p for interaction = 0.007). Stratified analysis showed an inverse association between H. pylori infection and serum uric acid in individuals with mild renal impairment (eGFR 60-80 mL/min/1.73m², n = 824; adjusted $\beta = -17.86$, 95% CI: -31.28 to -4.44, p = 0.009), while no such association was observed in those with normal renal function (eGFR ≥80 mL/ $min/1.73m^2$, n = 7,531; $\beta = 3.92$, 95% CI: -0.66 to 8.50, p = 0.094). Sensitivity analyses confirmed the robustness of these findings.

Conclusion: Renal function modulates the association between H. pylori infection and serum uric acid levels, with an inverse correlation observed in individuals with mild renal impairment. These findings suggest that renal function may influence the impact of H. pylori infection on uric acid metabolism.

KEYWORDS

Helicobacter pylori, serum uric acid, renal function, effect modification, Chinese population

Introduction

Helicobacter pylori, a Gram-negative bacterium that colonizes the human gastric mucosa, infects nearly half of the global population, presenting a significant public health concern (1–4). Its prevalence varies across regions, with developing countries exhibiting higher infection rates (5–7). In China, approximately 50% of the population is infected, though regional variations exist due to socioeconomic factors, lifestyle choices, and demographic characteristics (8–12). Since Warren and Marshall first isolated *H. pylori* in 1982, research has shown that its pathogenic effects extend beyond the gastrointestinal tract, contributing to conditions such as peptic ulcers, chronic gastritis, gastric cancer, and even neurodegenerative diseases (13–17).

Recent studies have highlighted that H. pylori influences host metabolic homeostasis through multiple mechanisms. For instance, VacA targets mitochondria, leading to Adenosine Triphosphate (ATP) depletion and cellular energy stress, which activates AMP-activated protein kinase (AMPK) and shifts cellular processes from anabolic to catabolic through mechanisms including autophagy induction (18-20). Chronic low-grade inflammation induced by *H. pylori* infection elevates pro-inflammatory cytokines, such as Tumor Necrosis Factor- α (TNF- α) and Interleukin-6 (IL-6), impairing insulin signaling and contributing to insulin resistance (21, 22). Additionally, H. pylori alters the gut microbiota, affecting short-chain fatty acid production and the secretion of metabolic hormones such as ghrelin and leptin, which in turn influence systemic energy metabolism (23-25). These extragastric effects provide a biological basis for linking *H. pylori* infection with metabolic disorders, including disturbances in uric acid metabolism.

Uric acid, the final product of purine metabolism, acts as an antioxidant at normal concentrations by scavenging reactive oxygen species and protecting against oxidative stress (26-28). However, elevated serum uric acid levels are an independent risk factor for metabolic and cardiovascular diseases, including diabetes, hypertension, and metabolic syndrome (29-31). Uric acid homeostasis is closely tied to renal function, with the kidneys playing a central role in its clearance. Approximately 70% of uric acid is cleared by the kidneys through transporters such as Urate Transporter 1 (URAT1) and Glucose Transporter 9 (GLUT9) (32, 33). As renal function declines, reduced renal clearance of uric acid triggers compensatory excretion via intestinal ATP-Binding Cassette Subfamily G Member 2 (ABCG2) transporters (33, 34). This interaction between the kidneys and gut—referred to as the "kidney-gut axis"—suggests that factors such as H. pylori infection may affect uric acid metabolism differently depending on renal function, offering a new perspective on the variability seen in past studies (34-36).

The relationship between *H. pylori* infection and serum uric acid levels remains contentious. Some studies suggest that *H. pylori* infection is a risk factor for gout, while others report no significant association (37, 38). This inconsistency may be attributed to the modifying role of renal function. Research has shown that patients with chronic kidney disease (CKD) have lower *H. pylori* infection rates (48.2% vs. 59.3%),

and the relationship between uric acid and clinical outcomes varies significantly across different stages of renal function (39, 40). In the early stages of renal decline, uric acid levels are positively correlated with cardiovascular risk, while this association weakens or disappears in later stages (40, 41). Furthermore, *H. pylori's* impact on renal health appears to be stage-dependent, suggesting that renal function may modulate the interaction between *H. pylori* infection and uric acid metabolism through altered gastrointestinal conditions, inflammatory responses, and the function of uric acid transporters (32, 34, 35, 39).

Given the conflicting evidence, the lack of consideration for potential effect modifiers, and the scarcity of studies in Chinese populations, this study aims to evaluate the relationship between *H. pylori* infection and serum uric acid levels in a large cohort from China, with a specific focus on renal function as a modifying factor. We hypothesize that the association between *H. pylori* infection and serum uric acid levels will vary depending on renal function status, which may help explain the inconsistencies observed in previous studies and offer a theoretical framework for more personalized infection management and metabolic monitoring strategies.

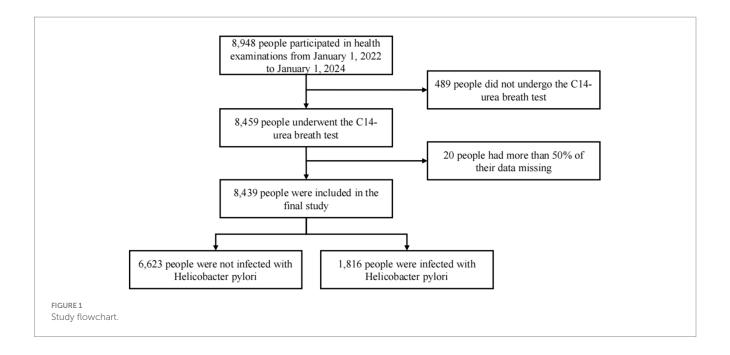
Methods

Study design and population

This retrospective cross-sectional study utilized health examination data from Tianyou Hospital, affiliated with Wuhan University of Science and Technology, and its community health service centers. Data were collected between January 2022 and January 2024. The study protocol was approved by the Ethics Committee of Tianyou Hospital (approval number: No. LL2024100901), and written informed consent was obtained from all participants. Initially, 8,948 adults (≥18 years) undergoing health examinations were included, predominantly asymptomatic individuals with a small proportion presenting mild, non-specific gastrointestinal symptoms, such as dyspepsia or epigastric discomfort. Inclusion criteria were: (1) completion of the 14C-urea breath test (14C-UBT); (2) availability of complete demographic and blood biochemistry data; (3) no use of proton pump inhibitors, bismuth preparations, antibiotics, or other anti-H. pylori medications within 4 weeks prior to testing; (4) voluntary participation with consent for data use. Exclusion criteria included: (1) missing key test data (14C-UBT, demographic data, or blood tests); (2) current or past use of uric acid-lowering medications (e.g., allopurinol, febuxostat, benzbromarone); (3) history of gastric cancer or gastrectomy; (4) missing data rate >50%. After rigorous screening, 8,439 participants were included in the final analysis (Figure 1).

Helicobacter pylori infection detection

Helicobacter pylori infection status was assessed using the 14C-UBT. Participants fasted for at least 8 h before the test. The



procedure involved ingesting a capsule containing 1.0 μ Ci of 14C-labeled urea, followed by breath sample collection 15 min later using a GP1000 breath analyzer (Furui Kang, China). A diagnostic threshold of 100 dpm/mmol CO₂ was set, with values \geq 100 dpm/mmol CO₂ considered positive for *H. pylori* and values <100 dpm/mmol CO₂ considered negative. This method has demonstrated high diagnostic accuracy, with sensitivity and specificity exceeding 95% (42).

Serum uric acid measurement

Venous blood samples were collected after a minimum 8-h fasting period. Serum uric acid concentrations were measured using the uricase-peroxidase method on an ADVIA 2400 automatic biochemistry analyzer (Siemens, Germany). The enzymatic method converts uric acid to allantoin and hydrogen peroxide, which then reacts with chromogenic substrates for colorimetric quantification. The detection range was 0–20 mg/dL (0–1,190 μ mol/L), with intraassay and inter-assay coefficients of variation (CV) of <2 and <3%, respectively. Hyperuricemia was defined as serum uric acid >420 μ mol/L in males and >360 μ mol/L in females (43).

Covariate collection and measurement

Covariates were selected based on existing literature and included age, sex, body mass index (BMI), mean arterial pressure (MAP), smoking status, alcohol consumption, occupational activity, hypertension, diabetes, and various laboratory parameters. Age and sex were obtained via structured questionnaires. BMI was measured using a standardized electronic height-weight scale (MSG005-H, Meilun Medical, China), with BMI calculated as weight (kg)/height² (m²). Blood pressure was measured according to American Heart Association guidelines, with the average of three measurements taken after a 5–10 min rest. MAP was calculated using the formula: MAP = diastolic pressure + (systolic pressure - diastolic pressure)/3.

Smoking and alcohol consumption were classified into three categories: (1) never exposed; (2) former exposure (≥ 1 year cessation); (3) current exposure (cessation <1 year). Occupational intensity was assessed using metabolic equivalents (METs) based on the Compendium of Physical Activities (44). Participants reported their occupational type and the activity composition of a typical workday. Corresponding METs values were assigned to each activity, and participants were classified into three categories: light (<3.0 METs, primarily sedentary work, such as office workers); moderate (3.0-6.0 METs, involving frequent standing or moderate physical activity, such as sales staff and nurses); and heavy (>6.0 METs, involving continuous heavy physical labor, such as construction workers and porters). Hypertension was defined as: (1) prior physician diagnosis; (2) current antihypertensive medication; (3) systolic ≥140 mmHg or diastolic \geq 90 mmHg (based on \geq 2 measurements). Diabetes was defined as: (1) glucose-lowering medications; (2) fasting glucose ≥7.0 mmol/L (based on ≥2 measurements); or fasting glucose between 6.1-6.9 mmol/L (considered as diabetes for binary analysis). Blood samples were analyzed for lipid profile, liver function markers (alanine aminotransferase, aspartate aminotransferase, bilirubin), fasting glucose, and serum creatinine. Renal function was evaluated using the estimated glomerular filtration rate (eGFR), calculated via the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) equation. A complete blood count was measured using standard methods. All tests were performed within 4 h of blood collection, and laboratory procedures adhered to National Center for Clinical Laboratory quality control standards.

Statistical analysis

Continuous variables were expressed as means ± standard deviation, and categorical variables as frequencies (percentages). Baseline characteristics were compared between *H. pylori*-positive and negative groups using one-way Analysis of Variance (ANOVA) for continuous variables and chi-square tests for categorical variables.

Linear regression assumptions were tested, and multicollinearity was assessed using variance inflation factors (VIF), excluding variables with VIF > 5. The association between *H. pylori* infection and serum uric acid was analyzed using multivariable linear regression in three hierarchical models: Model 1 (unadjusted), Model 2 (adjusted for age and sex), and Model 3 (fully adjusted for all covariates). Interaction analysis was conducted to identify effect modifiers by including interaction terms between H. pylori infection and potential modifiers, with likelihood ratio tests used to assess statistical significance (p < 0.05). For continuous effect modifiers showing significant interactions, grid search algorithms were applied to determine optimal stratification thresholds based on model fit criteria. Sensitivity analyses included logistic regression with hyperuricemia as a binary outcome and linear regression using log-transformed uric acid values to account for non-normality. All analyses were performed using R version 4.5.1, with two-sided *p*-values <0.05 considered statistically significant.

Results

Characteristics of the study population

A total of 8,439 participants were included in the study, of whom 6,623 (78.48%) were H. pylori-negative and 1,816 (21.52%) were H. pylori-positive. The H. pylori-positive group had a higher proportion of males (80.84% vs. 77.65%, p = 0.004), current smokers (37.06% vs. 32.09%, *p* < 0.001), current drinkers (35.35% vs. 32.90%, p = 0.036), and individuals engaged in heavy physical labor (45.48% vs. 40.45%, p = 0.001). Biochemically, the *H. pylori*-positive group exhibited significantly higher mean arterial pressure (MAP) $(94.02 \pm 13.69 \text{ vs. } 92.87 \pm 12.64 \text{ mmHg}, p = 0.001)$, white blood cell count $(6.60 \pm 1.68 \text{ vs. } 6.28 \pm 1.58 \times 10^9/\text{L}, p < 0.001)$, and platelet count $(246.55 \pm 62.27 \text{ vs. } 241.92 \pm 57.80 \times 10^9/\text{L}, p = 0.003).$ Conversely, high-density lipoprotein cholesterol (HDL-C) levels were significantly lower in the *H. pylori*-positive group $(1.33 \pm 0.33 \text{ vs.})$ $1.37 \pm 0.34 \text{ mmol/L}$, p < 0.001). Serum uric acid levels were also higher in the *H. pylori*-positive group compared to the negative group $(403.76 \pm 102.89 \text{ vs. } 395.87 \pm 102.13 \, \mu\text{mol/L}, p = 0.004) \, \text{(Table 1)}.$

Association between Helicobacter pylori infection and serum uric acid

In the unadjusted model (Model 1), *H. pylori* infection was positively associated with serum uric acid levels (β = 7.89, 95% CI: 2.58 to 13.20, p = 0.004). However, after adjusting for confounders, this association weakened. In the age- and sex-adjusted model (Model 2), the association became marginally significant (β = 4.39, 95% CI: -0.27 to 9.05, p = 0.065). In the fully adjusted model (Model 3), which accounted for metabolic parameters, the association was no longer significant (β = 1.92, 95% CI: -2.38 to 6.23, p = 0.381) (Table 2).

Identification of effect modifiers

Interaction analysis revealed that estimated glomerular filtration rate (eGFR) significantly modified the relationship

TABLE 1 Characteristics of study participants at baseline.

	7.				
Variables	<i>H. pylori</i> Negative (n = 6,623)	H. pylori Positive (n = 1816)	p-value		
Sex, %					
Female	1,480 (22.35%)	348 (19.16%)	0.004		
Male	5,143 (77.65%)	1,468 (80.84%)			
Drinking, %					
Never	3,100 (46.81%)	789 (43.45%)	0.036		
Former	1,344 (20.29%)	385 (21.20%)			
Current	2,179 (32.90%)	642 (35.35%)			
Smoking, %					
Never	3,137 (47.37%)	758 (41.74%)	<0.001		
Former	1,361 (20.55%)	385 (21.20%)			
Current	2,125 (32.09%)	673 (37.06%)			
Occupational intensity	, %				
Light	756 (11.41%)	195 (10.74%)	0.001		
Moderate	3,188 (48.14%)	795 (43.78%)			
Heavy	2,679 (40.45%)	826 (45.48%)			
Hypertension, %					
No	4,757 (71.83%)	1,262 (69.49%)	0.055		
Yes	1866 (28.17%)	554 (30.51%)			
Diabetes, %	1				
No	5,706 (86.15%)	1,549 (85.30%)	0.372		
Yes	917 (13.85%)	267 (14.70%)			
Age (years)	43.85 ± 12.42	44.14 ± 12.06	0.372		
BMI (kg/m²)	24.67 ± 3.53	24.81 ± 3.57	0.139		
MAP (mmHg)	92.87 ± 12.64	94.02 ± 13.69	0.001		
HDL-C (mmol/L)	1.37 ± 0.34	1.33 ± 0.33	< 0.001		
LDL-C (mmol/L)	2.95 ± 0.77	3.01 ± 0.76	0.01		
TC (mmol/L)	4.82 ± 0.93	4.85 ± 0.91	0.226		
TG (mmol/L)	1.86 ± 1.88	1.90 ± 1.72	0.415		
ALT (U/L)	29.39 ± 22.72	29.57 ± 22.72	0.76		
AST (U/L)	23.49 ± 12.40	23.82 ± 13.58	0.335		
TBil (μmol/L)	16.44 ± 6.23	16.13 ± 6.00	0.057		
DBil (μmol/L)	6.11 ± 2.23	6.03 ± 2.08	0.156		
eGFR (mL/ min/1.73 m²)	101.69 ± 18.92	101.46 ± 19.56	0.642		
UA (μmol/L)	395.87 ± 102.13	403.76 ± 102.89	0.004		
FBG (mmol/L)	5.35 ± 1.36	5.41 ± 1.48	0.067		
RBC (10 ¹² /L)	4.89 ± 0.47	4.90 ± 0.48	0.292		
WBC (10 ⁹ /L)	6.28 ± 1.58	6.60 ± 1.68	<0.001		
PLT (10 ⁹ /L)	241.92 ± 57.80	246.55 ± 62.27	0.003		
H. twlari. Helicobacter twlari. RMI. RoduMass Index: MAP. Mean Arterial Pressure: HDIC					

H. pylori, Helicobacter pylori; BMI, BodyMass Index; MAP, Mean Arterial Pressure; HDL-C, High-Density Lipoprotein Cholesterol; LDL-C, Low-Density Lipoprotein Cholesterol; TC, Total Cholesterol; TG, Triglycerides; ALT, Alanine Aminotransferase; AST, Aspartate Aminotransferase; TBil, Total Bilirubin; DBil, Direct Bilirubin; eGRF, estimated Glomerular Filtration Rate; UA, Serum Uric Acid; FBG, Fasting Blood Glucose; RBC, Red Blood Cell count; WBC, White Blood Cell count; PLT, Platelet count.

between H. pylori infection and serum uric acid (P for interaction = 0.007). Alanine aminotransferase (ALT) also showed a significant interaction (p=0.035). However, traditional effect modifiers such as sex, age, and body mass index (BMI) did not exhibit significant interactions (Supplementary Table 1). A systematic breakpoint analysis identified that the association between H. pylori infection and serum uric acid was most significant at eGFR = $85 \text{ mL/min}/1.73\text{m}^2$ (p=0.002). Based on clinical relevance and statistical considerations, a cut-off of $80 \text{ mL/min}/1.73\text{m}^2$ was selected for the primary analysis (p=0.006) (Supplementary Table 2).

Stratified analysis by renal function

Stratified analysis revealed heterogeneity in the effect of H. pylori infection on serum uric acid across different renal function groups. In the mild renal impairment group (eGFR 60–80 mL/min/1.73m², n=824), H. pylori infection was inversely associated with serum uric acid levels ($\beta=-17.86$, 95% CI: -31.28 to -4.44, p=0.009). In contrast, in the normal renal function group (eGFR ≥ 80 mL/min/1.73m², n=7.531), the association was positive but not statistically significant ($\beta=3.92$, 95% CI: -0.66 to 8.50, p=0.094). In the severe renal impairment group (eGFR < 60 mL/min/1.73m², n=84), no significant association was observed, likely due to the small sample size ($\beta=25.9$, 95% CI: -18.36 to 70.17, p=0.256) (Table 3).

Sensitivity analyses

Several sensitivity analyses were conducted to verify the robustness of the findings. First, logistic regression with hyperuricemia as a binary outcome revealed that in the mild renal impairment group (eGFR 60–80 mL/min/1.73m², n = 824), H. pylori infection was associated with a 33% reduced risk of hyperuricemia (odds ratio [OR] = 0.673, 95% CI: 0.461 to 0.983, p = 0.040) (Table 4). Second, analysis using log-transformed serum uric acid values showed that in the same group, *H. pylori*-positive individuals had 4.43% lower serum uric acid levels compared to H. pylori-negative individuals (95% CI: -7.92% to -1.15%, p = 0.009) (Table 5). These results from different statistical approaches further support the main findings. Collinearity diagnostics indicated severe multicollinearity for total cholesterol (VIF = 21.3) and direct bilirubin (VIF = 8.9) in the initial model. After excluding these variables, all VIF values were below 3, confirming no multicollinearity issues in the final model. Re-analysis with the adjusted variable yielded unchanged set (Supplementary Table 3). Linear regression assumption tests further supported the validity of model assumptions (Supplementary Figures 1–4).

Collinearity diagnostics indicated severe multicollinearity for total cholesterol (VIF = 21.3) and direct bilirubin (VIF = 8.9) in the initial model. After excluding these variables, all VIF values were below 3, confirming no multicollinearity issues in the final model. Re-analysis with the adjusted variable set yielded unchanged results (Supplementary Table 3). Linear regression assumption tests further supported the validity of model assumptions (Supplementary Figures 1-4).

Discussion

In this cross-sectional study of 8,439 Chinese adults, we found that the association between *H. pylori* infection and serum uric acid levels varied significantly by renal function. Although unadjusted analyses suggested a positive correlation, the association lost statistical significance after adjustment for demographic, metabolic, and hepatorenal covariates indicating that the observed relationship in the general population may reflect shared risk factors rather than a direct effect. Notably, stratified analyses identified a significant inverse association between *H. pylori* infection and serum uric acid levels among individuals with mild renal impairment (eGFR 60–80 mL/min/1.73 m²), which remained robust after multivariable adjustment. These findings suggest that renal function modulates the systemic metabolic effects of *H. pylori* infection. However, due to the cross-sectional design, causal inference and temporal directionality cannot be established.

The relationship between H. pylori and uric acid metabolism has been inconsistently reported in previous studies. Data from the U. S. National Health and Nutrition Examination Survey (NHANES) demonstrated a positive association, with H. pylori positive individuals exhibiting elevated uric acid to HDL cholesterol ratios (OR = 1.15; 95% CI: 1.02 to 1.30; p = 0.020) (45). In contrast, a large Chinese health screening cohort (n = 76,749) reported significantly lower serum uric acid concentrations among infected individuals (p < 0.001) (46). These conflicting findings remain unresolved. Our results provide a potential explanation by identifying renal function as an effect modifier. Variability in eGFR distribution across cohorts may have contributed to the discrepancies. Importantly, neither of the prior studies stratified participants by renal function, which may have masked subgroup specific associations. Our finding that H. pylori infection is inversely associated with serum uric acid only in those with mildly reduced eGFR highlights the critical role of organ-specific functional status in modulating systemic consequences of localized infections. This insight underscores the importance of renal

TABLE 2 Association between Helicobacter pylori infection and serum uric acid levels using hierarchical linear regression models.

Variable	n (%)	Model 1		Model 2		Model 3	
		β (95% CI)	p-value	β (95% CI)	p-value	β (95% CI)	<i>p</i> -value
H. Pylori							
Negative	6,623 (78.48)	Reference	0.004	Reference	0.065	Reference	0.381
Positive	1816 (21.52)	7.89 (2.58, 13.20)		4.39 (-0.27, 9.05)		1.92 (-2.38, 6.23)	

Model 1: Unadjusted; Model 2: Adjusted for age and sex; Model 3: Adjusted for all covariates; β: regression coefficient representing the difference in serum uric acid between H. pylori positive and negative groups; CI: Confidence Interval; Total cholesterol and direct bilirubin were excluded due to multicollinearity (VIF >5).

TABLE 3 Association between Helicobacter pylori infection and serum uric acid stratified by renal function.

Renal function category	N	β (95% CI) ^a	p-value	
Severe impairment (eGFR <60 mL/min/1.73m²)	84	25.90 (-18.36, 70.17)	0.256	
Mild impairment (eGFR 60-80 mL/min/1.73m²)	824	-17.86 (-31.28, -4.44)	0.009	
Normal function (eGFR ≥80 mL/min/1.73m²)	7,531	3.92 (-0.66, 8.50)	0.094	

CI, confidence interval; eGFR, estimated glomerular filtration rate.

TABLE 4 Association between Helicobacter pylori infection and hyperuricemia risk stratified by renal function: logistic regression analysis.

Renal function category	Total (n)	Hyperuricemia ^a n (%)	OR (95% CI) ^b	<i>p</i> -value
Severe impairment eGFR <60 mL/min/1.73m ²	84	32 (38.1)	6.78 (0.63, 72.76)	0.114
Mild impairment eGFR 60-80 mL/min/1.73m ²	824	350 (42.5)	0.67 (0.46, 0.98)	0.040
Normal function eGFR ≥80 mL/min/1.73m²	7,531	3,125 (41.5)	1.03 (0.91, 1.17)	0.595

CI, confidence interval; eGFR, estimated glomerular filtration rate; OR, odds ratio.

TABLE 5 Sensitivity analysis: association between Helicobacter pylori infection and log-transformed serum uric acid levels stratified by renal function.

Renal function category	Total (<i>n</i>)	β ^a	Percent change (%) (95% Cl) ^b	<i>p</i> -value
Severe impairment eGFR<60 mL/min/1.73m ²	84	0.063	6.45 (-4.88, 17.37)	0.275
Mild impairment eGFR 60-80 mL/min/1.73m ²	824	-0.045	-4.43 (-7.92, -1.15)	0.009
Normal function eGFR ≥80 mL/min/1.73m ²	7,531	0.008	0.83 (-0.35, 2.01)	0.168

CI, confidence interval; eGFR, estimated glomerular filtration rate.

stratification in metabolic epidemiology, particularly in studies of chronic infectious exposures.

The observed inverse association in individuals with mild renal impairment warrants mechanistic consideration. We hypothesize that this finding reflects an interaction between H. pylori induced metabolic changes and compensatory extra-renal uric acid elimination, which becomes physiologically relevant as renal clearance begins to decline. Chronic *H. pylori* colonization induces low-grade systemic inflammation, characterized by increased levels of IL-1β, IL-6, and TNF- α (47, 48). These cytokines upregulate xanthine oxidoreductase (XOR) activity in gastric and intestinal epithelia, enhancing both uric acid production and reactive oxygen species (49-51). Concurrently, inflammatory signaling upregulates intestinal urate transporters particularly ATP-binding cassette subfamily G member 2 (ABCG2) on enterocyte apical membranes, promoting urate excretion into the gut lumen (47, 52). In addition, H. pylori urease activity raises local ammonia concentrations and alters pH, potentially enhancing the expression and function of these transporters (53, 54). In individuals with normal renal function (eGFR >90 mL/min/1.73 m²), such intestinal adaptations are likely of limited systemic consequence, as renal clearance responsible for 70% of urate elimination remains efficient (55). However, when eGFR declines into the 60-80 mL/min/1.73 m² range, modest reductions in renal excretory capacity may render these intestinal compensatory mechanisms biologically significant. In this setting, H. pylori driven inflammatory and transporter mediated effects may enhance gut-renal axis activity, leading to a net reduction in serum uric acid levels (34, 52). This hypothesis is supported by the specificity of our finding to this eGFR category, where impairment is sufficient to trigger compensation but not yet severe enough to overwhelm it. Despite statistical significance, the absolute magnitude of urate reduction observed was modest and unlikely to influence clinical decisionmaking thresholds for pharmacological intervention, which typically begin at 360-420 µmol/L. Nevertheless, the biological relevance remains compelling. Our findings reveal a previously unrecognized interaction between gastrointestinal infection and uric acid homeostasis, particularly under conditions of early renal dysfunction. The renal function-dependent gradient observed in our stratified

a, β represent the difference in serum uric acid levels between H. pylori-positive and H. pylori-negative participants. Models were adjusted for age, sex, body mass index, mean arterial pressure, smoking, drinking, occupational intensity, hypertension, diabetes, high-density lipoprotein cholesterol, low-density lipoprotein cholesterol, triglycerides, alanine aminotransferase, aspartate aminotransferase, total bilirubin, estimated glomerular filtration rate, fasting blood glucose, red blood cell count, white blood cell count and platelet count. Total cholesterol and direct bilirubin were excluded due to multicollinearity (VIF >5).

a, Hyperuricemia defined as serum uric acid >420 µmol/L for males and >360 µmol/L for females, according to Chinese guidelines for diagnosis and treatment of hyperuricemia and gout. b, Odds ratios adjusted for age, sex, body mass index, mean arterial pressure, smoking, drinking, occupational intensity, hypertension, diabetes, high-density lipoprotein cholesterol, low-density lipoprotein cholesterol, triglycerides, alanine aminotransferase, aspartate aminotransferase, total bilirubin, estimated glomerular filtration rate, fasting blood glucose, red blood cell count, white blood cell count and platelet count. Total cholesterol and direct bilirubin were excluded due to multicollinearity (VIF >5).

a, β represent the difference in log-transformed serum uric acid levels between H. pylori-positive and H. pylori-negative groups, adjusted for age, sex, body mass index, mean arterial pressure, smoking, drinking, occupational intensity, hypertension, diabetes, high-density lipoprotein cholesterol, low-density lipoprotein cholesterol, triglycerides, alanine aminotransferase, aspartate aminotransferase, total bilirubin, estimated glomerular filtration rate, fasting blood glucose, red blood cell count, white blood cell count and platelet count. Total cholesterol and direct bilirubin were excluded due to multicollinearity (VIF >5).

b, Percent change calculated as $(e^{\beta}-1)\times 100\%,$ representing the relative difference in serum uric acid levels.

analyses further supports this interpretation. Among participants with normal renal function (eGFR \geq 80 mL/min/1.73 m²), no significant association was detected. In contrast, a clear inverse relationship emerged in the mildly impaired group. Among those with severe renal dysfunction (eGFR <60 mL/min/1.73 m²), no association was found (β = 25.90; p = 0.256), and the wide confidence interval (-18.36 to 70.17) suggests substantial imprecision. Several explanations may account for this null finding. The subgroup sample size was limited (n = 84), with a relatively low prevalence of H. pylori infection, reducing statistical power. Furthermore, advanced CKD is associated with elevated systemic urea, altered gastric acid profiles, and frequent antibiotic exposure factors that may suppress H. pylori colonization. At this stage of renal decline, uric acid homeostasis is often severely dysregulated due to impaired excretion, transporter dysfunction, and polypharmacy (56-58). Thus, the compensatory mechanisms operative in early CKD may be either saturated or disrupted in advanced disease. This gradient, from null association in normal function, to significant inverse association in mild impairment, to absence of effect in severe CKD supports our central hypothesis: H. pylori influences uric acid metabolism through renal intestinal compensatory pathways that are active within a specific window of declining renal function.

Our findings carry several important implications. First, they suggest that the metabolic impact of *H. pylori* infection is modulated by renal function, underscoring the need for individualized consideration in both *H. pylori* eradication strategies and hyperuricemia management. In individuals with mild renal impairment, infection status may influence serum urate levels and should be integrated into metabolic risk assessments. Furthermore, future investigations into *H. pylori* associated metabolic effects should incorporate renal function stratification to avoid obscured associations and more accurately capture biological heterogeneity.

This study possesses several notable strengths. The large, community-based cohort enhances external validity, while standardized assessments of *H. pylori* status and renal function support measurement reliability. Importantly, this work introduces renal function as a novel and clinically meaningful effect modifier within the framework of host microbe metabolism interactions. Adjustment for a wide range of demographic, metabolic, and hepatorenal covariates further strengthens internal validity and mitigates confounding.

Nonetheless, several limitations should be considered when interpreting the findings. First, we did not assess strain level characteristics or bacterial load. H. pylori is genetically heterogeneous, with virulence factors such as CagA, VacA, and urease activity influencing host inflammatory responses and potentially affecting systemic metabolism (59–61). Without molecular characterization, we cannot determine whether specific bacterial phenotypes contributed to the observed associations. Second, the cross-sectional design precludes conclusions regarding causality or temporal direction. The 14C-UBT reflects current infection status but provides no insight into infection duration or cumulative exposure. Given the typically chronic nature of *H. pylori* colonization, infection duration may play a pivotal role in host metabolic adaptation (60). Prospective studies incorporating seroconversion data and serial uric acid measurements are needed to elucidate temporal dynamics. Third, although H. pylori is anatomically confined to the gastric mucosa, its systemic metabolic effects may be mediated through several established pathways, including low-grade systemic inflammation, altered gastric acidity affecting intestinal transporter expression, and modulation of the gut-kidney axis (34, 52, 62). While the effect size observed in this study was modest and unlikely to affect clinical thresholds for urate lowering therapy, the biological relevance remains noteworthy particularly in individuals with early renal dysfunction, where cumulative metabolic burden may evolve over time. Fourth, residual confounding cannot be excluded. Data on dietary purine intake, alcohol consumption, and use of uric acid modulating medications were not available. Endoscopic or histological markers of gastric pathology were also lacking, precluding correlation between mucosal inflammatory severity and systemic metabolic outcomes. Additional unmeasured variables such as prior H. pylori eradication, bacterial density, and host microbiome interactions may have influenced the findings. Finally, the subgroup with severe renal impairment (eGFR <60 mL/min/1.73 m²) was relatively small (n = 84) and exhibited a low prevalence of H. pylori infection (20.2%), limiting statistical power and resulting in wide confidence intervals. The reduced infection rate in this subgroup may itself have biological significance, as advanced chronic kidney disease is associated with elevated urea, altered gastric pH, and frequent antibiotic use factors that may inhibit H. pylori colonization. Moreover, the findings may not be generalizable to non-Chinese populations, in whom genetic variation in urate transporters and microbial susceptibility may differ.

In conclusion, this study identifies renal function as a key modifier of the association between *H. pylori* infection and serum uric acid levels. A significant inverse relationship was observed exclusively in individuals with mild renal impairment, likely reflecting synergistic interactions between *H. pylori* induced intestinal adaptations and compensatory urate excretion pathways. These findings provide mechanistic insight into the gut-kidney axis and illustrate how localized infections may exert systemic metabolic effects in the context of subclinical organ dysfunction. Although not yet practice changing, these results contribute to a growing framework in which host microbe interactions are interpreted through the lens of organ specific physiology. Given the inherent limitations of cross-sectional observational design, these findings should be regarded as hypothesis generating and warrant validation through prospective, mechanistic studies with longitudinal follow up.

Conclusion

Our results suggest that serum uric acid levels in the Chinese population studied are influenced by *Helicobacter pylori* infection, with renal function appearing to modify this relationship. While an initial positive association was observed, this became non-significant after adjusting for confounders. Notably, an inverse association was found in individuals with mild renal impairment. These findings highlight the potential role of renal function in the relationship between *H. pylori* infection and uric acid metabolism. Further longitudinal studies with more detailed renal assessments and molecular characterization of *H. pylori* are needed to better understand the underlying mechanisms and temporal dynamics of this association.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary material, further inquiries can be directed to the corresponding author.

Ethics statement

The studies involving humans were approved by the Ethics Committee of the Tianyou Hospital of Wuhan University of Science and Technology. The studies were conducted in accordance with the local legislation and institutional requirements. The ethics committee/institutional review board waived the requirement of written informed consent for participation from the participants or the participants' legal guardians/next of kin because this study is a retrospective study.

Author contributions

KW: Writing – review & editing, Writing – original draft. JZ: Writing – review & editing. SY: Writing – review & editing. HZ: Writing – review & editing. HY: Writing – review & editing. YJ: Writing – review & editing. YQ: Writing – review & editing. XW: Writing – review & editing. XZ: Writing – review & editing. LY: Writing – review & editing. ST: Writing – review & editing.

Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

References

- 1. Wu X, Wu D, Cui G, Lee KH, Yang T, Zhang Z, et al. Association between biofilm formation and structure and antibiotic resistance in *H. pylori. Infect Drug Resist.* (2024) 17:2501–12. doi: 10.2147/IDR.S468126
- 2. Salih BA. *Helicobacter pylori* infection in developing countries: the burden for how long? *Saudi J Gastroenterol.* (2009) 15:201–7. doi: 10.4103/1319-3767.54743
- 3. Yi M, Chen S, Yi X, Zhang F, Zhou X, Zeng M, et al. *Helicobacter pylori* infection process: from the molecular world to clinical treatment. *Front Microbiol.* (2025) 16:1541140. doi: 10.3389/fmicb.2025.1541140
- 4. Pu S, Zhuang Z, Liu N, Luo Q, Zhang D. Research progress on the relationship between *helicobacter pylori* infection and iron deficiency anemia. *Front Microbiol.* (2025) 16:1552630. doi: 10.3389/fmicb.2025.1552630
- 5. Shaaban SI, Talat D, Khatab SA, Nossair MA, Ayoub MA, Ewida RM, et al. An investigative study on the zoonotic potential of *helicobacter pylori*. *BMC Vet Res.* (2023) 19:16. doi: 10.1186/s12917-023-03572-w
- 6. Aminde JA, Dedino GA, Ngwasiri CA, Ombaku KS, Mahop Makon CA, Aminde LN. Helicobacter pylori infection among patients presenting with dyspepsia at a primary care setting in Cameroon: seroprevalence, five-year trend and predictors. BMC Infect Dis. (2019) 19:30. doi: 10.1186/s12879-019-3677-0
- 7. Saeed AY, Rashad BH, Ali BN, Sulaivany AH, Ibrahim KS. *Helicobacter pylori* infection: prevalence, risk factors, and treatment efficacy in symptomatic patients in Zakho city, kurdistan region, Iraq. *Cureus*. (2024) 16:e73873. doi: 10.7759/cureus.73873
- 8. Chen R-X, Zhang D-Y, Zhang X, Chen S, Huang S, Chen C, et al. A survey on *helicobacter pylori* infection rate in Hainan province and analysis of related risk factors. *BMC Gastroenterol.* (2023) 23:338. doi: 10.1186/s12876-023-02973-3
- 9. Marshall B. Epidemiology of helicobacter in Chinese families: a foundation for cost-effective eradication strategies? *Gut.* (2024) 73:870–1. doi: 10.1136/gutjnl-2023-329786

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The authors declare that no Gen AI was used in the creation of this manuscript.

Any alternative text (alt text) provided alongside figures in this article has been generated by Frontiers with the support of artificial intelligence and reasonable efforts have been made to ensure accuracy, including review by the authors wherever possible. If you identify any issues, please contact us.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed.2025.1615161/full#supplementary-material

- 10. Yu X-C, Shao Q-Q, Ma J, Yu M, Zhang C, Lei L, et al. Family-based *helicobacter pylori* infection status and transmission pattern in Central China, and its clinical implications for related disease prevention. *World J Gastroenterol.* (2022) 28:3706–19. doi: 10.3748/wjg.v28.i28.3706
- 11. Lu Y, Xiao F, Wang Y, Wang Z, Liu D, Hong F. Prevalence of *helicobacter pylori* in non-cardia gastric cancer in China: a systematic review and meta-analysis. *Front Oncol.* (2022) 12:850389. doi: 10.3389/fonc.2022.850389
- 12. Li F, Si Y-T, Tang J-W, Umar Z, Xiong X-S, Wang J-T, et al. Rapid profiling of carcinogenic types of *helicobacter pylori* infection via deep learning analysis of label-free SERS spectra of human serum. *Comput Struct Biotechnol J.* (2024) 23:3379–90. doi: 10.1016/j.csbj.2024.09.008
- 13. Ahmed N. 23 years of the discovery of helicobacter pylori: is the debate over? Ann Clin Microbiol Antimicrob. (2005) 4:17. doi: 10.1186/1476-0711-4-17
- 14. Abbasi J. Barry marshall, MD: H
 pylori 35 years later. JAMA. (2017) 317:1400–2. doi: 10.1001/jama.2017.2629
- 15. Watari J, Chen N, Amenta PS, Fukui H, Oshima T, Tomita T, et al. *Helicobacter pylori* associated chronic gastritis, clinical syndromes, precancerous lesions, and pathogenesis of gastric cancer development. *World J Gastroenterol*. (2014) 20:5461–73. doi: 10.3748/wjg.v20.i18.5461
- 16. Noori M, Mahboobi R, Nabavi-Rad A, Jamshidizadeh S, Fakharian F, Yadegar A, et al. *Helicobacter pylori* infection contributes to the expression of alzheimer's disease-associated risk factors and neuroinflammation. *Heliyon*. (2023) 9:e19607. doi: 10.1016/j.heliyon.2023.e19607
- 17. Lamb A, Chen L-F. Role of the *helicobacter pylori*-induced inflammatory response in the development of gastric cancer. *J Cell Biochem.* (2013) 114:491–7. doi: 10.1002/jcb.24389

- 18. Luo J, Bai L, Tao J, Wen Y, Li M, Zhu Y, et al. Autophagy induced by *H. pylori* VacA regulated the survival mechanism of the SGC7901 human gastric cancer cell line. *Genes Genomics*. (2021) 43:1223–30. doi: 10.1007/s13258-021-01151-7
- 19. Seeger AY, Zaidi F, Alhayek S, Jones RM, Zohair H, Holland RL, et al. Host cell sensing and restoration of mitochondrial function and metabolism within *helicobacter pylori* VacA intoxicated cells. *MBio*. (2023) 14:e0211723. doi: 10.1128/mbio.02117-23
- 20. Holland RL, Bosi KD, Seeger AY, Blanke SR. Restoration of mitochondrial structure and function within *helicobacter pylori* VacA intoxicated cells. *Adv Microbiol.* (2023) 13:399–419. doi: 10.4236/aim.2023.138026
- 21. Yu Y-Y, Tong Y-L, Wu L-Y, Yu X-Y. *Helicobacter pylori* infection eradication for nonalcoholic fatty liver disease: a randomized controlled trial. *Sci Rep.* (2022) 12:19530. doi: 10.1038/s41598-022-23746-0
- 22. Yu B, Xiang L, Peppelenbosch MP, Fuhler GM. Overlapping cytokines in *H. pylori* infection and gastric cancer: a tandem meta-analysis. *Front Immunol.* (2023) 14:1125658. doi: 10.3389/fimmu.2023.1125658
- 23. Yanagi H, Tsuda A, Matsushima M, Takahashi S, Ozawa G, Koga Y, et al. Changes in the gut microbiota composition and the plasma ghrelin level in patients with *Helicobacter pylori*-infected patients with eradication therapy. *BMJ Open Gastroenterol.* (2017) 4:e000182. doi: 10.1136/bmjgast-2017-000182
- 24. Francois F, Roper J, Joseph N, Pei Z, Chhada A, Shak JR, et al. The effect of *H. pylori* eradication on meal-associated changes in plasma ghrelin and leptin. *BMC Gastroenterol.* (2011) 11:37. doi: 10.1186/1471-230X-11-37
- 25. Mohammadi SO, Yadegar A, Kargar M, Mirjalali H, Kafilzadeh F. The impact of helicobacter pylori infection on gut microbiota-endocrine system axis; modulation of metabolic hormone levels and energy homeostasis. J Diabetes Metab Disord. (2020) 19:1855–61. doi: 10.1007/s40200-020-00608-y
- 26. Sautin YY, Johnson RJ. Uric acid: the oxidant-antioxidant paradox. Nucleosides Nucleotides Nucleic Acids. (2008) 27:608–19. doi: 10.1080/15257770802138558
- 27. Ames BN, Cathcart R, Schwiers E, Hochstein P. Uric acid provides an antioxidant defense in humans against oxidant- and radical-caused aging and cancer: a hypothesis. *Proc Natl Acad Sci USA*. (1981) 78:6858–62. doi: 10.1073/pnas.78.11.6858
- 28. Mikami T, Sorimachi M. Uric acid contributes greatly to hepatic antioxidant capacity besides protein. *Physiol Res.* (2017) 66:1001–7. doi: 10.33549/physiolres.933555
- 29. Chen S, Wu N, Yu C, Xu Y, Xu C, Huang Y, et al. Association between baseline and changes in serum uric acid and incident metabolic syndrome: a nation-wide cohort study and updated meta-analysis. *Nutr Metab.* (2021) 18:59. doi: 10.1186/s12986-021-00584-x
- 30. Liu L, Zhang X, Qie R, Han M, Li Q, Zhang L, et al. Dose-response association between serum uric acid levels and incident hypertension: a systematic review and meta-analysis of 17 prospective cohort studies of 32 thousand participants. *Acta Cardiol.* (2021) 76:748–53. doi: 10.1080/00015385.2020.1779476
- 31. Lv Q, Meng X-F, He F-F, Chen S, Su H, Xiong J, et al. High serum uric acid and increased risk of type 2 diabetes: a systemic review and meta-analysis of prospective cohort studies. *PLoS One.* (2013) 8:e56864. doi: 10.1371/journal.pone.0056864
- 32. Bobulescu IA, Moe OW. Renal transport of uric acid: evolving concepts and uncertainties. Adv Chronic Kidney Dis. (2012) 19:358–71. doi: 10.1053/j.ackd.2012.07.009
- 33. Chung S, Kim G-H. Urate transporters in the kidney: what clinicians need to know. Electrolyte Blood Press. (2021) 19:1–9. doi: 10.5049/EBP.2021.19.1.1
- 34. Ohashi Y, Toyoda M, Saito N, Koizumi M, Kanai G, Komaba H, et al. Evaluation of ABCG2-mediated extra-renal urate excretion in hemodialysis patients. *Sci Rep.* (2023) 13:93. doi: 10.1038/s41598-022-26519-x
- 35. Xu Y, Bi W-D, Shi Y-X, Liang X-R, Wang H-Y, Lai X-L, et al. Derivation and elimination of uremic toxins from kidney-gut axis. *Front Physiol.* (2023) 14:1123182. doi: 10.3389/fphys.2023.1123182
- 36. Nigam SK, Bhatnagar V. The systems biology of uric acid transporters: the role of remote sensing and signaling. *Curr Opin Nephrol Hypertens*. (2018) 27:305–13. doi: 10.1097/MNH.0000000000000427
- 37. Wawro N, Amann U, Butt J, Meisinger C, Akmatov MK, Pessler F, et al. *Helicobacter pylori* seropositivity: prevalence, associations, and the impact on incident metabolic diseases/risk factors in the population-based KORA study. *Front Public Health*. (2019) 7:96. doi: 10.3389/fpubh.2019.00096
- 38. Chu W-Y, Allegaert K, TPC D, ADR HALBINO Study Group. Semi-mechanistic modeling of hypoxanthine, xanthine, and uric acid metabolism in asphyxiated neonates. *Clin Pharmacokinet*. (2022) 61:1545–58. doi: 10.1007/s40262-022-01164-9
- 39. Shin SP, Bang CS, Lee JJ, Baik GH. *Helicobacter pylori* infection in patients with chronic kidney disease: a systematic review and meta-analysis. *Gut Liver.* (2019) 13:628–41. doi: 10.5009/gnl18517
- 40. Kwon Y-E, Ahn S-Y, Ko G-J, Kwon Y-J, Kim J-E. Impact of uric acid levels on mortality and cardiovascular outcomes in relation to kidney function. *J Clin Med.* (2024) 14:20. doi: 10.3390/jcm14010020

- 41. Meremo A, Paget G, Duarte R, Dickens C, Dix-Peek T, Bintabara D, et al. Demographic and clinical profile of black patients with chronic kidney disease attending a tertiary hospital in johannesburg, South Africa. *PLoS One.* (2022) 17:e0266155. doi: 10.1371/journal.pone.0266155
- 42. Jambi LK. Systematic review and meta-analysis on the sensitivity and specificity of 13C/14C-urea breath tests in the diagnosis of *helicobacter pylori* infection. *Diagnostics*. (2022) 12:2428. doi: 10.3390/diagnostics12102428
- 43. Piao W, Zhao L, Yang Y, Fang H, Ju L, Cai S, et al. The prevalence of hyperuricemia and its correlates among adults in China: results from CNHS 2015-2017. *Nutrients*. (2022) 14:4095. doi: 10.3390/nu14194095
- 44. Mendes MA, da Silva I, Ramires V, Reichert F, Martins R, Ferreira R, et al. Metabolic equivalent of task (METs) thresholds as an indicator of physical activity intensity. *PLoS One.* (2018) 13:e0200701. doi: 10.1371/journal.pone.0200701
- 45. Qin Z, Fang Y, Liu Y, Zhang L, Zhang R, Zhang S. Association between hp infection and serum uric acid to high-density lipoprotein cholesterol ratio in adults. *Front Med.* (2025) 12:1509269. doi: 10.3389/fmed.2025.1509269
- 46. Gong R, Ling Y, Wang S, Wang W, Meng L, Zeng J, et al. *Helicobacter pylori* infection as an independent risk factor for kidney stone formation in China: a cross-sectional study. *Infect Drug Resist.* (2025) 18:2881–8. doi: 10.2147/IDR.S523335
- 47. Bravo D, Hoare A, Soto C, Valenzuela MA, Quest AF. *Helicobacter pylori* in human health and disease: mechanisms for local gastric and systemic effects. *World J Gastroenterol.* (2018) 24:3071–89. doi: 10.3748/wjg.v24.i28.3071
- 48. Sugimoto M, Furuta T, Yamaoka Y. Influence of inflammatory cytokine polymorphisms on eradication rates of *helicobacter pylori*. *J Gastroenterol Hepatol*. (2009) 24:1725–32. doi: 10.1111/j.1440-1746.2009.06047.x
- 49. McGee DW, Bamberg T, Vitkus SJ, McGhee JR. A synergistic relationship between TNF-alpha, IL-1 beta, and TGF-beta 1 on IL-6 secretion by the IEC-6 intestinal epithelial cell line. *Immunology*. (1995) 86:6–11.
- 50. Abooali M, Lall GS, Coughlan K, Lall HS, Gibbs BF, Sumbayev VV. Crucial involvement of xanthine oxidase in the intracellular signalling networks associated with human myeloid cell function. *Sci Rep.* (2014) 4:6307. doi: 10.1038/srep06307
- 51. Bortolotti M, Polito L, Battelli MG, Bolognesi A. Xanthine oxidoreductase: one enzyme for multiple physiological tasks. $Redox\ Biol.\ (2021)\ 41:101882.$ doi: 10.1016/j.redox.2021.101882
- 52. Chen M, Lu X, Lu C, Shen N, Jiang Y, Chen M, et al. Soluble uric acid increases PDZK1 and ABCG2 expression in human intestinal cell lines via the TLR4-NLRP3 inflammasome and PI3K/akt signaling pathway. *Arthritis Res Ther.* (2018) 20:20. doi: 10.1186/s13075-018-1512-4
- 53. Miller EF, Maier RJ. Ammonium metabolism enzymes aid helicobacter pylori acid resistance. J Bacteriol. (2014) 196:3074–81. doi: 10.1128/JB.01423-13
- 54. Ortiz-Ramirez C, Mora SI, Trejo J, Pantoja O. PvAMT1;1, a highly selective ammonium transporter that functions as H+/NH4(+) symporter. *J Biol Chem.* (2011) 286:31113–22. doi: 10.1074/jbc.M111.261693
- 55. Xiao X, Ye H, Yi C, Lin J, Peng Y, Huang X, et al. Roles of peritoneal clearance and residual kidney removal in control of uric acid in patients on peritoneal dialysis. *BMC Nephrol.* (2020) 21:148. doi: 10.1186/s12882-020-01800-1
- $56.\,\mathrm{Anders}$ H-J, Li Q, Steiger S. Asymptomatic hyperuricaemia in chronic kidney disease: mechanisms and clinical implications. Clin Kidney J. (2023) 16:928–38. doi: $10.1093/\mathrm{ckj/sfad006}$
- 57. Nishizawa K, Yoda N, Morokado F, Komori H, Nakanishi T, Tamai I. Changes of drug pharmacokinetics mediated by downregulation of kidney organic cation transporters Matel and Oct2 in a rat model of hyperuricemia. *PLoS One.* (2019) 14:e0214862. doi: 10.1371/journal.pone.0214862
- 58. Naseralallah L, Khatib M, Al-Khulaifi A, Danjuma M. Prevalence and global trends of polypharmacy in patients with chronic kidney disease: a systematic review and meta-analysis. *Front Pharmacol.* (2023) 14:1122898. doi: 10.3389/fphar.2023.1122898
- 59. Jain P, Luo Z-Q, Blanke SR. *Helicobacter pylori* vacuolating cytotoxin a (VacA) engages the mitochondrial fission machinery to induce host cell death. *Proc Natl Acad Sci USA*. (2011) 108:16032–7. doi: 10.1073/pnas.1105175108
- 60. Yamaoka Y, Kita M, Kodama T, Sawai N, Kashima K, Imanishi J. Induction of various cytokines and development of severe mucosal inflammation by cagA gene positive *helicobacter pylori* strains. *Gut.* (1997) 41:442–51. doi: 10.1136/gut.41.4.442
- 61. Rutherford JC. The emerging role of urease as a general microbial virulence factor. $PLoS\ Pathog.\ (2014)\ 10:e1004062.$ doi: 10.1371/journal.ppat.1004062
- 62. Li L, Bao B, Chai X, Chen X, Su X, Feng S, et al. The anti-inflammatory effect of callicarpa nudiflora extract on H. pylori-infected GES-1 cells through the inhibition of ROS/NLRP3/caspase-1/IL-1 β signaling axis. Can J Infect Dis Med Microbiol. (2022) 2022:5469236. doi: 10.1155/2022/5469236