



# **Editorial: Molecular Tracing of Aquatic Viruses: Where Epidemiology Needs to Meet Genomics**

Jean-Christophe Avarre\*

ISEM, IRD, CNRS, Univ. Montpellier, EPHE, Montpellier, France

Keywords: aquaculture, virus, genomics, tracing, epidemiology

Editorial on the Research Topic

## Molecular Tracing of Aquatic Viruses: Where Epidemiology Needs to Meet Genomics

Viruses are highly abundant in aquatic environments (Bergh et al., 1989; Suttle, 2007) and can infect a wide range of organisms, from plankton cells to whales (Suttle, 2005). If aquatic viruses can cause mortalities among wild fish and shellfish populations, the rapid (and often uncontrolled) development of intensive aquaculture over the last decades has been a major driver of the emergence of many viral diseases (Walker and Winton, 2010). As a result, viral diseases now constitute a real threat for the sustainability of this ever-growing worldwide industry. In addition to their devastating economic and social impact resulting in losses of several \$US billion (Walker and Mohan, 2009), viruses may also have substantial environmental impacts on the surrounding ecosystems, either directly (Bunce and Norman, 2000; Dann et al., 2000) or indirectly (Le et al., 2005; Ali, 2006; Phan et al., 2011).

# **OPEN ACCESS**

## Edited by:

George S. Bullerjahn, Bowling Green State University, United States

## Reviewed by:

Steven Wilhelm, University of Tennessee, Knoxville, United States

## \*Correspondence:

Jean-Christophe Avarre jean-christophe.avarre@ird.fr

#### Specialty section:

This article was submitted to Aquatic Microbiology, a section of the journal Frontiers in Microbiology

Received: 02 June 2017 Accepted: 26 July 2017 Published: 08 August 2017

## Citation:

Avarre J-C (2017) Editorial: Molecular Tracing of Aquatic Viruses: Where Epidemiology Needs to Meet Genomics. Front. Microbiol. 8:1498. doi: 10.3389/fmicb.2017.01498 As stated by Walker and Winton, the extent of disease spread and impacts is greatly affected by the availability of suitable diagnostic tools for identification of the causative agent or by the ability to understand the genetic relation of the causative agent with other characterized pathogens (Walker and Winton, 2010). An example is the emergence of the white spot syndrome in 1992, which has spread to almost all shrimp-producing countries before its causative agent, the white spot syndrome virus, could be identified and fully characterized nearly 9 years later (Yang et al., 2001).

In this context, the objectives of this research topic were: (i) to illustrate how sequencing technologies and associated bioinformatics tools can be utilized to trace aquatic viruses, and (ii) to discuss the new opportunities they offer for understanding the emergence of viral diseases and controlling their spread.

The two first articles of this topic show how sequencing a single gene of a virus may help understand the ways it propagates between aquaculture settings. Sequence comparison of VP2 gene from infectious pancreatic necrosis virus isolates showed the existence of two clades that differed by a distinct signature motif in the hypervariable region, each motif being associated with a different level of infection (Mutoloki et al.). In the second paper, Abbadi et al. looked at the evolution of two viruses that cause high mortalities in rainbow trout farms, the infectious haematopoietic necrosis virus (IHNV) and the viral haemorrhagic septicaemia virus (VHSV). They analyzed the glycoprotein (G) gene sequences of many isolates collected over a period of nearly 30 years and concluded that the two viruses have distinct evolutionary rates. Integration of these molecular data with high-quality epidemiological information led to propose different patterns of virus spread among trout farms.

In a third article, Klafack et al. describe an improved method to genotype and trace the highly pathogenic Cyprinid herpesvirus 3. They developed a qPCR test that enables to discriminate between the Asian and European lineages, and found that this virus was able to switch between the Asian and European genotypes after many *in vitro* passages.

The following two articles discuss the ability of metagenomics to uncover the environmental factors that influence virus spread in aquatic environments and to identify novel pathogenic viruses. Most of our knowledge on the epidemiology of viral diseases in aquaculture so far is derived from studies carried out on infected aquatic organisms. Metagenomics offers the possibility to study the epidemiology of viral diseases outside their host species by the direct analysis of environmental samples, which in turn enables to follow the environmental factors that influence the composition of viral communities (a recent example is provided by Hwang et al. (2017). Understanding the factors (both natural and anthropogenic) that influence the epidemiology of viral diseases may ultimately lead to the design of rational disease control strategies, especially in the aquaculture context (Munang'andu). Likewise, metagenomics has greatly accelerated the pace of virus discovery in the recent years, and also appears valuable to design proactive diagnostic tools able to identify novel viruses before they cause disease outbreaks (Munang'andu et al.).

In the particular context of viral aquatic diseases, analysis of outbreaks requires typing methods that offer a high level of strain discrimination. As outlined in the sixth article, whole genome sequencing (WGS) represents the "ultimate" typing methodology in terms of discriminatory power (Bayliss et al.). WGS not only enables to resolve micro-evolutionary distances, but it also has the power to discover new and rare variations, including polymorphisms that arise during an outbreak or that evolve *in vivo* during an infection. It thus offers the possibility to track the emergence and spread of a given variant, while it may also provide predictive information concerning key phenotypic traits (Feil, 2015).

Finally, in the last article of this topic, Naville and Volff raise an interesting question that is often overlooked in the field of viral (re)emergence: the potential role of fish endogenous retroviruses.

# REFERENCES

- Ali, A. M. S. (2006). Rice to shrimp: land use land cover changes and soil degradation in Southwestern Bangladesh. Land Use Policy 23, 421–435. doi: 10.1016/j.landusepol.2005.02.001
- Bergh, O., Borsheim, K. Y., Bratbak, G., and Heldal, M. (1989). High abundance of viruses found in aquatic environments. *Nature* 340, 467–468. doi: 10.1038/340467a0
- Bunce, A., and Norman, F. I. (2000). Changes in the diet of the Australasian gannet (Morus serrator) in response to the 1998 mortality of pilchards (Sardinops sagax). Mar. Freshw. Res. 51, 349–353. doi: 10.1071/MF99133
- Coffee, L. L., Casey, J. W., and Bowser, P. R. (2013). Pathology of tumors in fish associated with retroviruses. *Vet. Pathol.* 50, 390–403. doi: 10.1177/0300985813480529
- Dann, P., Norman, F. I., Cullen, J. M., Neira, F. J., and Chiaradia, A. (2000). Mortality and breeding failure of little penguins, Eudyptula minor, in Victoria, 1995-96, following a widespread mortality of pilchard, Sardinops sagax. *Mar. Freshw. Res.* 51, 355–362. doi: 10.1071/MF99114
- Eaton, W. D., Folkins, B., and Kent, M. L. (1994). Biochemical and histologic evidence of plasmacytoid leukemia and salmon leukemia virus (SLV) in wild-caught chinook salmon Oncorhynchus tshawytscha from british Columbia expressing plasmacytoid leukemia. *Dis. Aquat. Org.* 19, 147–151. doi: 10.3354/dao019147
- Feil, E. J. (2015). Toward a synthesis of genotypic typing and phenotypic inference in the genomics era. *Future Microbiol.* 10, 1897–1899. doi: 10.2217/fmb.15.113

While several exogenous retroviruses have been clearly identified as etiological agents of some fish diseases (Lepa and Siwicki, 2011; Coffee et al., 2013), little is known about the association between endogenous retroviruses and diseases in fish, with the exceptions of the zebrafish endogenous retrovirus (Frazer et al., 2012) and the salmon leukemia virus that may have an endogenous origin (Eaton et al., 1994). Naville and Volff speculate that scrutiny of more fish genomes will certainly uncover new types of retroviruses, some of which with a potential for retained infectivity or reemergence, as was recently discovered in the human genome (Wildschutte et al., 2016).

To conclude, this research topic brings evidence that genomics has now become a routine tool for tracing viral pathogens. However, in spite of the promises brought by new sequencing technologies, it is critical to keep in mind that a molecular typing dataset, including whole genome sequencing, will rarely be fully informative if it is not associated with rich epidemiological (meta)data. Besides, both metagenomics and WGS are facing technological challenges when applied to aquatic viruses, which the third and fourth generations of sequencing will probably only partly overcome (Nkili-Meyong et al., 2016).

# **AUTHOR CONTRIBUTIONS**

The author confirms being the sole contributor of this work and approved it for publication.

# FUNDING

Part of the research presented in this topic has benefited from the ERA-NET EMIDA funded project MOLTRAQ (molecular tracing of viral pathogens in aquaculture).

- Frazer, J. K., Batchelor, L. A., Bradley, D. F., Brown, K. H., Dobrinski, K. P., Lee, C., et al. (2012). Genomic amplification of an endogenous retrovirus in zebrafish T-cell malignancies. *Adv. Hematol.* 2012, 12. doi: 10.1155/2012/6 27920
- Hwang, J., Park, S. Y., Park, M., Lee, S., and Lee, T.-K. (2017). Seasonal dynamics and metagenomic characterization of marine viruses in Goseong Bay, Korea. *PLoS ONE* 12:e0169841. doi: 10.1371/journal.pone.0169841
- Le, T. X., Munekage, Y., and Kato, S. (2005). Antibiotic resistance in bacteria from shrimp farming in mangrove areas. *Sci. Tot. Environ.* 349, 95–105. doi: 10.1016/j.scitotenv.2005.01.006
- Lepa, A., and Siwicki, A. K. (2011). Retroviruses of wild and cultured fish. Pol. J. Vet. Sci. 14, 703–709. doi: 10.2478/v10181-011-0106-8
- Nkili-Meyong, A. A., Bigarré, L., Labouba, I., Vallaeys, T., Avarre, J. C., and Berthet, N. (2016). Contribution of next-generation sequencing to aquatic and fish virology. *Intervirology* 59, 285–300. doi: 10.1159/000477808
- Phan, T. P. H., Managaki, S., Nakada, N., Takada, H., Shimizu, A., Anh, D. H., et al. (2011). Antibiotic contamination and occurrence of antibiotic-resistant bacteria in aquatic environments of northern Vietnam. *Sci. Tot. Environ.* 409, 2894–2901. doi: 10.1016/j.scitotenv.2011.04.030
- Suttle, C. A. (2005). Viruses in the sea. Nature 437, 356–361. doi: 10.1038/nature04160
- Suttle, C. A. (2007). Marine viruses: major players in the global ecosystem. Nat. Rev. Microbiol. 5, 801–812. doi: 10.1038/nrmicro1750
- Walker, P. J., and Mohan, C. V. (2009). Viral disease emergence in shrimp aquaculture: origins, impact and the effectiveness of health management

strategies. *Rev. Aquacult.* 1, 125–154. doi: 10.1111/j.1753-5131.2009.0 1007.x

- Walker, P. J., and Winton, J. R. (2010). Emerging viral diseases of fish and shrimp. Vet. Res. 41:51. doi: 10.1051/vetres/2010022
- Wildschutte, J. H., Williams, Z. H., Montesion, M., Subramanian, R. P., Kidd, J. M., and Coffin, J. M. (2016). Discovery of unfixed endogenous retrovirus insertions in diverse human populations. *Proc. Natl. Acad. Sci. U.S.A.* 113, E2326–E2334. doi: 10.1073/pnas.1602336113
- Yang, F., He, J., Lin, X., Li, Q., Pan, D., Zhang, X., et al. (2001). Complete genome sequence of the shrimp white spot bacilliform virus. J. Virol. 75, 11811–11820. doi: 10.1128/JVI.75.23.11811-11820.2001

**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Avarre. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.