



Homologous Recombination in Protozoan Parasites and Recombinase Inhibitors

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Homologous recombination (HR) is a DNA double-strand break (DSB) repair pathway that utilizes a homologous template to fully repair the damaged DNA. HR is critical to maintain genome stability and to ensure genetic diversity during meiosis. A specialized class of enzymes known as recombinases facilitate the exchange of genetic information between sister chromatids or homologous chromosomes with the help of numerous protein accessory factors. The majority of the HR machinery is highly conserved among eukaryotes. In many protozoan parasites, HR is an essential DSB repair pathway that allows these organisms to adapt to environmental conditions and evade host immune systems through genetic recombination. Therefore, small molecule inhibitors, capable of disrupting HR in protozoan parasites, represent potential therapeutic options. A number of small molecule inhibitors were identified that disrupt the activities of the human recombinase RAD51. Recent studies have examined the effect of two of these molecules on the *Entamoeba* recombinases. Here, we discuss the current understandings of HR in the protozoan parasites *Trypanosoma*, *Leishmania*, *Plasmodium*, and *Entamoeba*, and we review the small molecule inhibitors known to disrupt human RAD51 activity.

Keywords: homologous recombination, DNA repair, double-strand break repair, protozoan parasites, recombination inhibitors, RAD51, DMC1

INTRODUCTION

DNA double-stranded breaks (DSBs) can occur due to exogenous or endogenous events. Exogenous sources of DSBs include reactive oxygen species generated from exposure to ionizing radiation and radiomimetic chemicals (Mehta and Haber, 2014), while endogenous sources of DSBs include erroneous DNA replication that can lead to the collapse of replication forks. Additionally, DSBs can be intentionally introduced into the genome through programmed events such as in meiosis, where the production of crossover products is essential. Whether they are produced intentionally or as a result of genotoxic events, unrepaired DSBs threaten the genome stability of an organism. Defects in DSB repair pathways have been associated with sterility, cancer, and chromosomal rearrangements (San Filippo et al., 2008; Moynahan and Jasin, 2010; Hunter, 2015).

Four major pathways function in DSB repair: non-homologous end joining (NHEJ), single-strand annealing (SSA), alternative end joining (ALT-EJ), and homologous recombination (HR) (**Figure 1**). NHEJ (also known as classical-NHEJ or canonical-NHEJ) is the major DSB repair pathway, and is mediated by many factors, which include Ku70, Ku80, XRCC4, and DNA ligase IV (Lieber, 2010). In this pathway, the ends of the DSB are protected to prevent nucleolytic end resection (Mimori and Hardin, 1986; Paillard and Strauss, 1991), followed by ligation to mend the DSB (**Figure 1A**). NHEJ is active during the entire cell cycle, and is the pathway responsible for V(D)J recombination in antibody variation (Soulas-Sprauel et al., 2007; Malu et al., 2012). NHEJ can result in full restoration of the DSB. However, if there is any enzymatic processing or degradation at the ends of the DSB, there will be a loss of genetic information once repaired (Betermier et al., 2014); thus, NHEJ is often associated with chromosomal alterations at the break site (Ghezraoui et al., 2014). Repair by SSA is initiated when a DSB occurs at a locus with extensive homology in the sequences flanking the break site (**Figure 1B**). Once homology is located (≥ 100 base pairs), the sequences are annealed by the DNA annealing factor RAD52 (Morales et al., 2015) and the non-homologous overlapping ends are endonucleolytically processed followed by DNA polymerase gap filling and ligation to restore the break (Bhargava et al., 2016). Similarly, ALT-EJ (also known as a microhomology mediated-EJ) DSB repair occurs through the recognition and annealing of short sequences called microhomology (≤ 10 base pairs) within the sequences flanking the DSB (Wang and Xu, 2017) (**Figure 1C**). The two main protein factors involved in mediating ALT-EJ are poly-ADP-ribose polymerase (PARP) and DNA polymerase theta (POLQ) (Bhargava et al., 2016). SSA and ALT-EJ are both active early in the S and G2 phases of the cell cycle. Both pathways result in the loss of DNA sequence between the annealed substrates (SSA results in large deletion events and ALT-EJ results in small deletions). As a result, both SSA and ALT-EJ are considered to be mutagenic DSB repair pathways (Bhargava et al., 2016). In contrast, HR relies on a homologous template to restore the damaged DNA in its entirety (**Figure 1D**). HR is active during the S and G2 phases of the cell cycle due to the presence of sister chromatids or homologous chromosomes (in meiosis) to act as the template. The HR pathway requires enzymatic resection of the ends of the DSB to produce 3' single-stranded DNA (ssDNA) overhangs. These overhangs serve as the nucleation site for a recombinase, the central enzyme of HR. There are two conserved recombinases among eukaryotes, Rad51 (radiation-sensitive 51) and Dmc1 (disrupted meiotic cDNA 1) (Moore, 1978; Bishop et al., 1992). Rad51 is the lead recombinase during mitosis, and Dmc1 serves as the major recombinase during meiosis, in which Rad51 plays a supportive role (Bishop, 2012). These different processes result in the requirement for different accessory proteins to modulate the functions of each recombinase. Nonetheless, the mechanisms of action for both recombinases are similar. The active ATP-bound form of the recombinase forms a right-handed presynaptic, nucleoprotein filament on the ssDNA overhang. The presynaptic filament then

conducts a homology search to identify a homologous sequence that will serve as a template for repair. Rad51 preferentially utilizes the sister chromatid as a template in order to prevent loss of heterozygosity, whereas Dmc1 prefers the homologous chromosome with the goal of generating crossover products. In either case, the presynaptic filament invades the duplex DNA and facilitates the base-pairing of the 3' ssDNA with its complementary sequence. As a result, the homologous strand of the duplex DNA is displaced to form a displacement loop (D-loop) structure. The 3' end of the invading strand primes DNA synthesis by a polymerase to replace DNA that was lost at the DSB site. As a result of strand invasion and DNA synthesis to repair the damaged DNA, two distinct DNA joint molecules can be formed. These DNA intermediates (the initial D-loop or Holliday junctions, which are associated with meiosis and form after D-loop extension, second end capture, ligation, and a second synthesis event) are resolved by one of two major pathways to ultimately yield non-crossover products or crossover products (Matos and West, 2014; Morrill, 2015).

The complex process of DSB repair by HR requires numerous accessory proteins to proceed with accuracy and efficiency (**Table 1**). RPA (replication protein A) is a heterotrimeric complex composed of three subunits (RPA1, RPA2, RPA3) that coat the ssDNA overhangs to protect them from nucleases and to prevent the formation of secondary structure (Binz et al., 2004). Although the DNA binding activity of RPA is necessary for HR, this function also represents a barrier to the recombinase loading. Various recombination mediators help to overcome this inhibition by displacing RPA and loading the recombinase onto the ssDNA, including *Saccharomyces cerevisiae* Rad52 (radiation-sensitive 52) and human BRCA2 (breast cancer susceptibility gene 2) (Sung, 1997; Jensen et al., 2010; Liu et al., 2010; Thorslund et al., 2010). Recently, DSS1 (deleted in split hand/split foot syndrome) was shown to aid BRCA2 in loading RAD51 onto RPA-coated ssDNA (Yang et al., 2002; Zhao et al., 2015). DSS1 functions as a DNA mimic that competes with authentic DNA to displace RPA, allowing BRCA2 to load RAD51 onto ssDNA (Zhao et al., 2014). Another factor, Hop2-Mnd1 (homologous-pairing protein 2—meiotic nuclear division protein 1), is a meiosis-specific heterodimeric protein complex that promotes recombinase-mediated D-loop formation by stabilizing the presynaptic filament and bringing the duplex DNA into close proximity with the presynaptic filament for more efficient homologous DNA pairing (Chi et al., 2007; Pezza et al., 2007). Other accessory proteins, such as RAD54 (radiation-sensitive 54), promote the search for homology by the recombinase and help to dissociate the recombinase from the DNA after strand exchange has occurred (Petukhova et al., 1998; Mason et al., 2015). The high degree of conservation of the HR pathway across divergent eukaryotic species emphasizes its indispensable nature. The clinical relevance of HR in non-communicable human disease, such as cancer, makes it a potential therapeutic target. Furthermore, since HR plays a role in virulence of eukaryotic pathogens (see below), the components of this DNA repair system may also serve as a target for the development of new drugs to fight infectious disease.

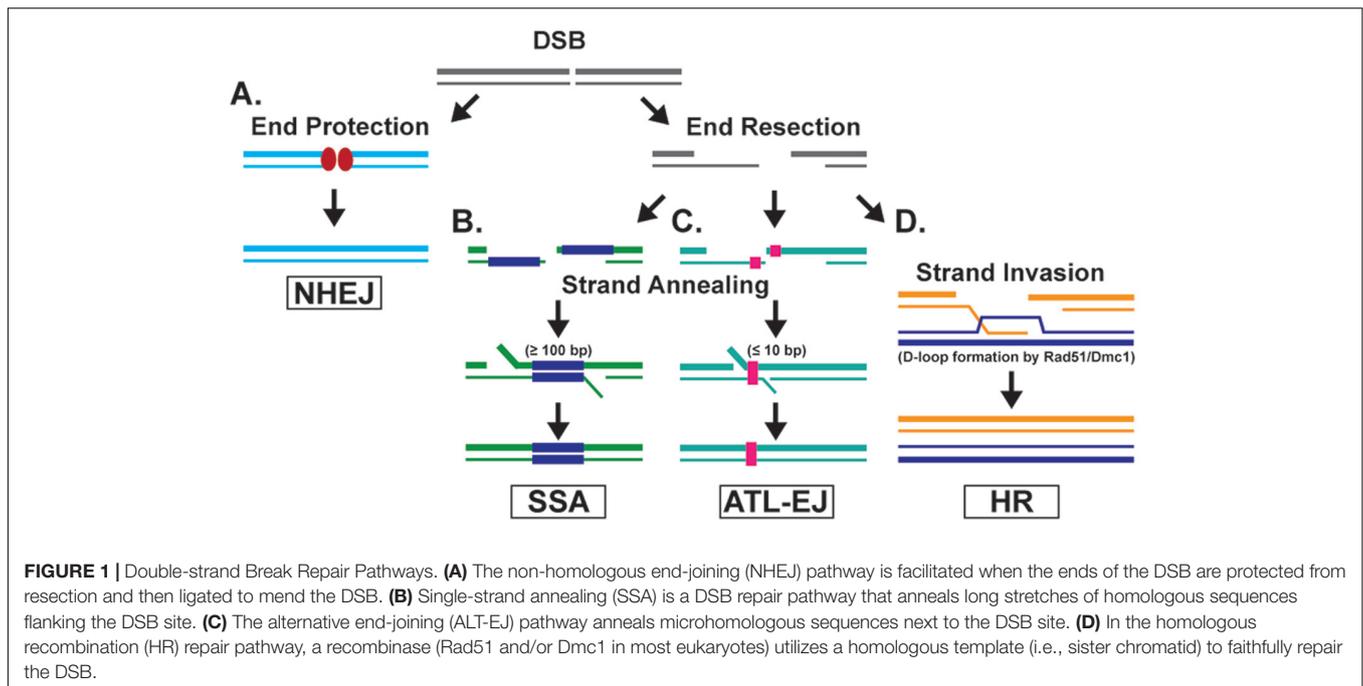


TABLE 1 | HR DSB repair proteins, species, functions, and UniProtKB identifiers of human, yeast, and putative protozoan parasite homologs.

Function	<i>Homo sapiens</i>	<i>Saccharomyces cerevisiae</i>	<i>Entamoeba histolytica</i>	<i>Trypanosoma brucei</i>	<i>Plasmodium falciparum</i>	<i>Leishmania major</i>
DSB End Resection	MRE11	Mre11	C4LVX7	Q8T8P1-?	A0A1C3KMQ6	E9ADG7
	RAD50	Rad50	N9V1K1	Q384J8	C6KSQ6	Q4Q8L7
	NSB1	Xrs2	?	?	?	?
Single-strand DNA Binding	RPA1-RPA2-RPA3	Rfa1-Rfa2-Rfa3	?-?-?	?-?-?	A0A1C3KN47/ A0A1C3KQ72	?-?-?
	RAD51	Rad51	Q86C17*	Q384K0*	Q8IIS8	O61127
Recombinase (Meiosis-specific)	DMC1	Dmc1	C4LTR6*	Q38E34*	A0A1C3KPB3	O61128
Accessory Proteins	RAD54	Rad54	C4LVM6*	Q385M5	Q8IAN4	Q4QH75
	RAD54B	Rdh54	N9TAM9	?	?	?
	RAD55	Rad55	?	?	?	?
	RAD57	Rad57	?	?	?	?
	-	RAD59	?	?	?	?
	HOP2	Hop2	?	?	?	?
	MND1	Mnd1	M3TLC0	?	C6S3J7	Q4QAN2
Single-Strand Annealing	RAD52	Rad52	C4M197	?	?	?
Mediator	BRCA2	Rad52	N9TLS7	Q4GZF5	?	Q4QD38

*Indicates characterized enzymes parasites.

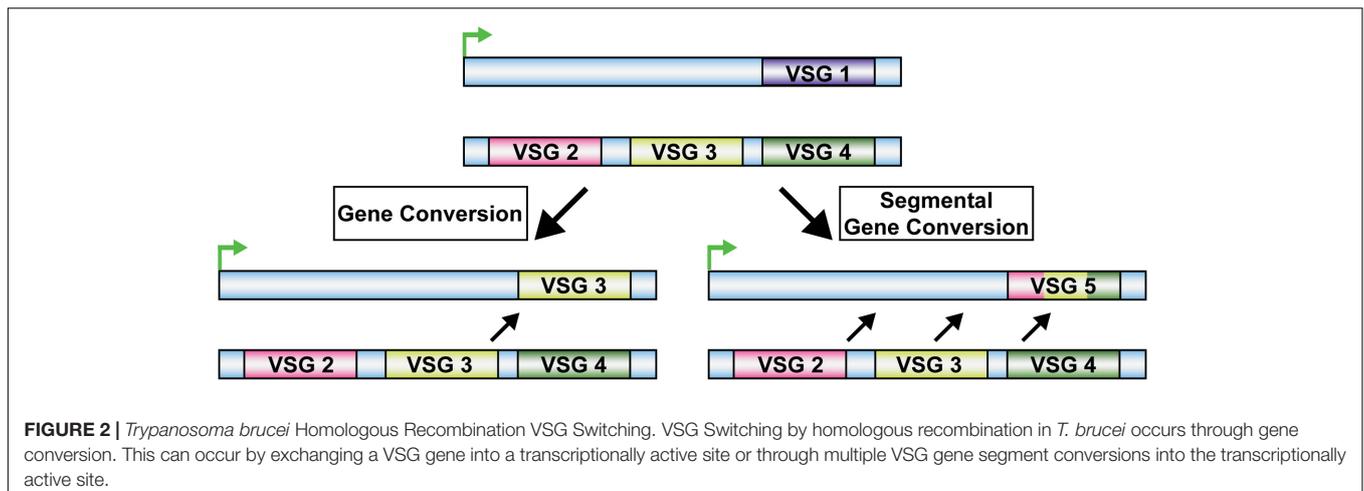
?Indicates no known homolog.

HOMOLOGOUS RECOMBINATION IN PROTOZOAN PARASITES

Homologous recombination is essential in many protozoan parasites. These parasites utilize HR to adapt to diverse environmental conditions, evade host immune systems, and respond to DSBs (Deitsch et al., 1997; Bhattacharyya et al., 2004). Many of the core eukaryotic enzymes involved in HR

are conserved among protozoan parasites (Table 1); however, the mechanisms of HR in many of these pathogens are largely unknown. Over the past few years, our understanding of HR in these organisms has increased from studies involving *Trypanosoma*, *Leishmania*, *Plasmodium*, and *Entamoeba*.

Homologous recombination plays a significant role in the survival of *T. brucei*, the protozoan parasite responsible for African trypanosomiasis (sleeping sickness). *T. brucei* generates



antigenic variation through DNA recombination to evade the host immune system (McCulloch and Barry, 1999). In *T. brucei*, antigenic variation consists of repeatedly changing the expression of the Variant Surface Glycoprotein (VSG) genes that encode an outer protective coat of the parasite (Cross, 1975). *T. brucei* expresses more than 1000 VSG genes and pseudogenes one at a time (Borst, 2002; Marcello and Barry, 2007; McCulloch et al., 2015). One of the ways in which *T. brucei* can switch the expression of the VSG gene is through gene conversion of entire VSG genes or partial VSG pseudogenes into a transcriptionally active site using HR (Figure 2) (Borst, 2002; Marcello and Barry, 2007; Vink et al., 2012). It was shown that Rad51 was a major factor in this process, as null mutations of Rad51 in *T. brucei* (*TbRad51*) led to a reduction in the VSG switching frequency (McCulloch and Barry, 1999). Moreover, mutations in genes involved in other DSB repair pathways (i.e., NHEJ) induced no change in VSG switching frequency (Conway et al., 2002a), indicating that VSG switching is likely dependent on *TbRad51*. However, the frequency of VSG switching was reduced rather than eliminated in *TbRad51* mutants, suggesting that RAD51-mediated HR is not the only pathway for VSG switching (McCulloch and Barry, 1999; Conway et al., 2002a,b). Indeed, ALT-EJ also exists as an alternative pathway to HR in this pathogen (Conway et al., 2002b; Barnes and McCulloch, 2007; Glover et al., 2008, 2011). Furthermore, *TbRad51* was reported to be important in response to DNA damage. When DSBs were induced in *T. brucei*, *TbRad51* was shown to localize to sub-nuclear foci (Glover and Horn, 2012), and HR served as the predominant DSB repair pathway (Barnes and McCulloch, 2007; Glover et al., 2008).

Trypanosoma cruzi is a protozoan parasite that causes Chagas disease, or American trypanosomiasis. *T. cruzi* Rad51 (*TcRad51*) has a significant role in the response to and repair of DNA damage caused by ionizing radiation, as shown by *in vivo* analyses (Regis-da-Silva et al., 2006). Interestingly, the genomes of *T. brucei* and *T. cruzi* contain Dmc1 genes with 65% and 70% identity to human DMC1, respectively (Proudfoot and McCulloch, 2006; Kelso et al., 2017). However, heterozygous and homozygous knockout *DMC1* mutants (*DMC1*^{+/-} and

dmc1^{-/-}, respectively) in *T. brucei* were not sensitive to DNA damage, failed to impact VSG switching frequency, and had unaltered recombination efficiency, suggesting that *TbDmc1* played no role in these processes (Proudfoot and McCulloch, 2006). These results were in direct contrast to the studies of *T. brucei rad51*^{-/-} mutants (McCulloch and Barry, 1999; Conway et al., 2002b). Currently, there is no evidence to suggest that *TbRad51* compensates for the loss of *TbDmc1*, or vice versa. Future work will be necessary to reveal the biochemical underpinnings of Rad51 and Dmc1 recombinases from *T. brucei* and *T. cruzi*.

Leishmaniasis is a diverse disease characterized by three main forms: fever/anemia/weight loss, skin lesions, or mucous membrane destruction. Leishmaniasis is caused by the protozoan parasite *Leishmania*. *Leishmania* utilize HR to induce gene rearrangement events in response to oxidative stress, while HR-mediated DNA amplification events serve as a mechanism for drug resistance (Beverley, 1991; Ouellette and Papadopoulou, 1993; Nathan and Shiloh, 2000; Ubeda et al., 2008, 2014; Montenegro et al., 2015). Although HR is exploited *in vitro* to create null mutants of *L. major* for study (Cruz et al., 1991), the mechanisms of HR in *L. major* are largely unknown. There are a few studies demonstrating the existence of functional Rad51 in *Leishmania*. Namely, *in vivo* studies of *L. major* showed that the typically low levels of Rad51 (*LmRad51*) expression increased in response to the DNA damaging agent phleomycin (McKean et al., 2001). Similarly, *L. infantum* Rad51 (*LiRAD51*) was highly expressed in response to phleomycin exposure, while the expression levels of the *L. infantum* ortholog of BRCA2 (*LiBRCA2*) were unchanged, similar to what has been reported in human cells (Lu et al., 2005). *LiRAD51* and *LiBRCA2* both localize to punctate foci in the nucleus; however, *LiRAD51* fails to localize to the nucleus in *LiBRCA2*-deficient cells (Genois et al., 2012). Biochemically, *LmRad51* ATP hydrolysis activity was stimulated by the presence of both ssDNA and dsDNA (McKean et al., 2001). Another biochemical study showed that *LiRAD51* binds DNA—with an apparent higher affinity for ssDNA over dsDNA (Genois et al., 2012), and *LiRAD51* can facilitate homologous DNA pairing (Genois et al., 2012).

Furthermore, *LiBRCA2* was reported to load *LiRAD51* onto RPA coated ssDNA (Genois et al., 2012), similar to its human homolog (Jensen et al., 2010; Liu et al., 2010; Thorslund et al., 2010).

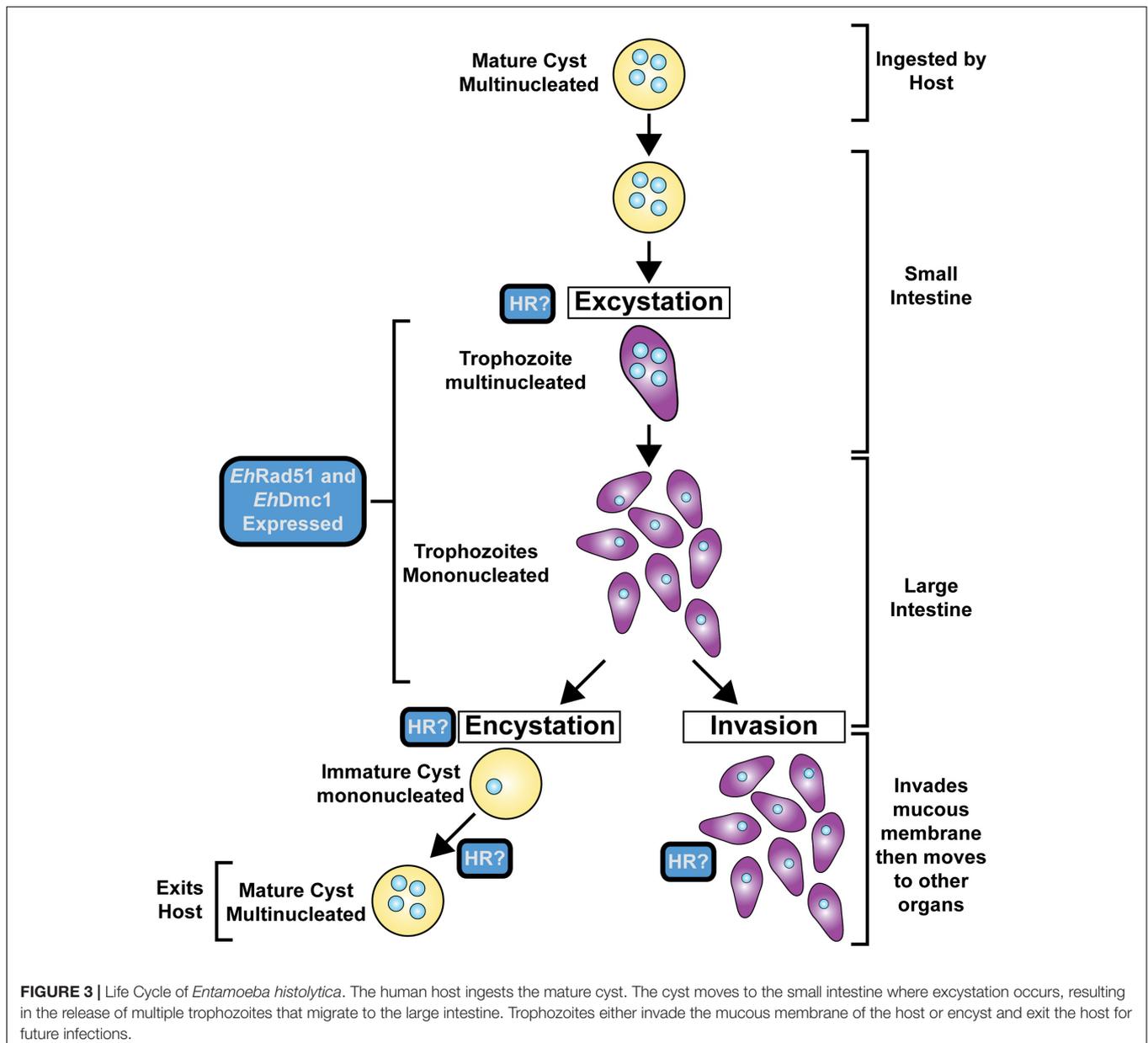
P. falciparum causes malaria in humans. The genome of *P. falciparum* contains many of the HR proteins (**Table 1**) (Gardner et al., 2002; Kirkman et al., 2014). *In vivo* studies using an exogenous HR reporter substrate with an inducible DSB site found that HR was the preferred DSB repair pathway when a homologous template was available, and during the more common haploid state, ALT-EJ was preferred (Kirkman et al., 2014). Upon exposure to the DNA damaging agent, methyl methanesulfonate (MMS), *P. falciparum* Rad51 (*PfRad51*) expression increased (Bhattacharyya and Kumar, 2003). Biochemical analysis of *PfRad51* showed that it hydrolyzed ATP and facilitated efficient DNA strand exchange (Bhattacharyya et al., 2005). *PfRad51* required ATP binding but not hydrolysis to catalyze DNA strand exchange *in vitro* (Bhattacharyya et al., 2005), similar to human RAD51. In support of this notion, mutational analysis of the ATP binding motif of *PfRad51* showed that *PfRad51K143R* significantly impacted the *in vivo* function of *PfRad51* (Roy et al., 2014). Furthermore, *P. berghei* (responsible for malaria in rodents) and *S. cerevisiae* expressing the *PfRad51-K143R* variant were found to be hypersensitive to MMS treatment (Roy et al., 2014).

In addition to the studies on *PfRad51*, homologs of other HR proteins were shown to have similar functions and/or responses to DNA DSBs in *Plasmodium* as reported for their human counterparts. For example, Rad54 is a Rad51 stimulating protein, and *PfRad54* similarly stimulated the homologous DNA pairing activity of *PfRad51* (Gopalakrishnan and Kumar, 2013). BLM (Bloom syndrome protein) is a DNA helicase involved in DSB end-resection, and *PfBlm* possessed helicase activity (Rahman et al., 2016). The protein complex Mre11 (meiotic recombination 11), Rad50 (radiation-sensitive 50), and NBS1 (involved in Nijmegen breakage syndrome) recognizes and end-resects DNA DSBs, and *PfMre11* demonstrated nuclease activity and interacted with *PfRad50* (Badugu et al., 2015). Lastly, diploid zygotes of *Plasmodium* undergo meiosis in the insect vector to produce haploid cells, a state in which Dmc1 becomes relevant. One study in *P. berghei* null for *dmc1* demonstrated that *PbDmc1* was essential for proper oocyst development (Mlambo et al., 2012). *PbDmc1* null cells were also shown to be highly sensitive to a DNA damaging agent (Mlambo et al., 2012).

Entamoeba histolytica is the parasite responsible for amoebic dysentery and amoebic liver abscess. The life cycle of *E. histolytica* consists of a tetra-nucleated cyst that is transmitted through water or food sources by fecal contamination (**Figure 3**). When ingested by the host, the mature, tetra-nucleated cyst can undergo excystation in the small intestine resulting in the release of eight trophozoites that can colonize the large intestine. Trophozoites multiply by binary fission and produce multinucleated cysts through the process of encystation (Dobell, 1928; Cleveland and Sanders, 1930; Ratcliffe and Geiman, 1934; Koushik et al., 2014). There is ample support for HR in *E. histolytica*. For example, the genome of *E. histolytica* contains the majority of the HR core genes (**Table 1**) (Bhattacharya

et al., 2000). Also, genome duplication events, unscheduled gene amplification events, and genetic rearrangements are reported in *E. histolytica* and thought to be mediated by HR (Orozco et al., 1988; Baez-Camargo et al., 1996; Bhattacharya et al., 2000; Mukherjee et al., 2008). The first study on HR in *E. histolytica* monitored the response of *E. histolytica* to UV-C radiation, which is known to cause DNA damage (Lopez-Casamichana et al., 2008). Many hallmarks of HR were observed upon DNA damage induction in *E. histolytica*: the histone H2AX was phosphorylated (indicating DNA damage), cell survivability was not affected (indicating DSB repair occurred), the core HR genes were differentially expressed in response to irradiation, *EhRad51* expression (mRNA and protein) peaked rapidly in response to DNA damage, and *EhRad51* formed nuclear foci in response to DNA damage (Lopez-Casamichana et al., 2008; Charcas-Lopez Mdel et al., 2014). During growth stresses (heat shock, oxygen stress, serum starvation) and in response to DNA damage (UV irradiation), inverted repeat sequences located either on a plasmid or in the *E. histolytica* genome underwent recombination (Singh et al., 2013). This represents the first direct evidence for HR in *E. histolytica*. Biochemically, Lopez-Casamichana et al. (2008) showed that partially purified *EhRad51* bound DNA and facilitated homologous DNA pairing (Lopez-Casamichana et al., 2008). The findings were recently extended by a report that demonstrated *EhRad51* was not only capable of binding DNA and forming D-loops, but that *EhRad51* hydrolyzed ATP, formed presynaptic filaments, and catalyzed DNA strand exchange over thousands of base pairs (Kelso et al., 2016). Although *E. histolytica* reproduce asexually through binary fission, the occurrence of meiosis in *E. histolytica* was first proposed to occur during encystation based on increased expression of meiosis-specific genes (Ehrenkauf et al., 2013). Additional support for this notion came from monitoring the expression of meiosis-specific genes in the reptilian amoebozoan parasite, *E. invadens* during encystation (Singh et al., 2013). More recently, biochemical analysis of *EhDmc1* demonstrated the enzyme hydrolyzed ATP in the presence of DNA, bound DNA, formed nucleoprotein filaments, and catalyzed homologous DNA pairing and DNA strand exchange (Kelso et al., 2015). Taken together, the evidence suggests that mitotic and meiotic HR occur and are likely important for encystation in *Entamoeba* (**Figure 3**).

Currently, the roles of other DSB repair pathways in protozoan parasites are largely uncharacterized. As mentioned above, ALT-EJ has been reported for some parasites, but the mechanisms remain elusive (Conway et al., 2002b; Barnes and McCulloch, 2007; Glover et al., 2008, 2011; Kirkman et al., 2014). As for SSA, although many genomes of protozoan parasites contain the *RAD52* gene, *T. brucei* is the only pathogen to demonstrate a potential for SSA (Glover and Horn, 2009). Lastly, the genomes of some parasites, like *T. brucei* and *E. histolytica*, contain factors necessary for NHEJ (e.g., Ku); however, many of the key components of this pathway are missing such as DNA ligase IV and XRCC4 homologs (Burton et al., 2007). In fact, *T. brucei* Ku was shown to have a role in telomere length maintenance, but DSB repair occurred independent of Ku (Conway et al., 2002a).

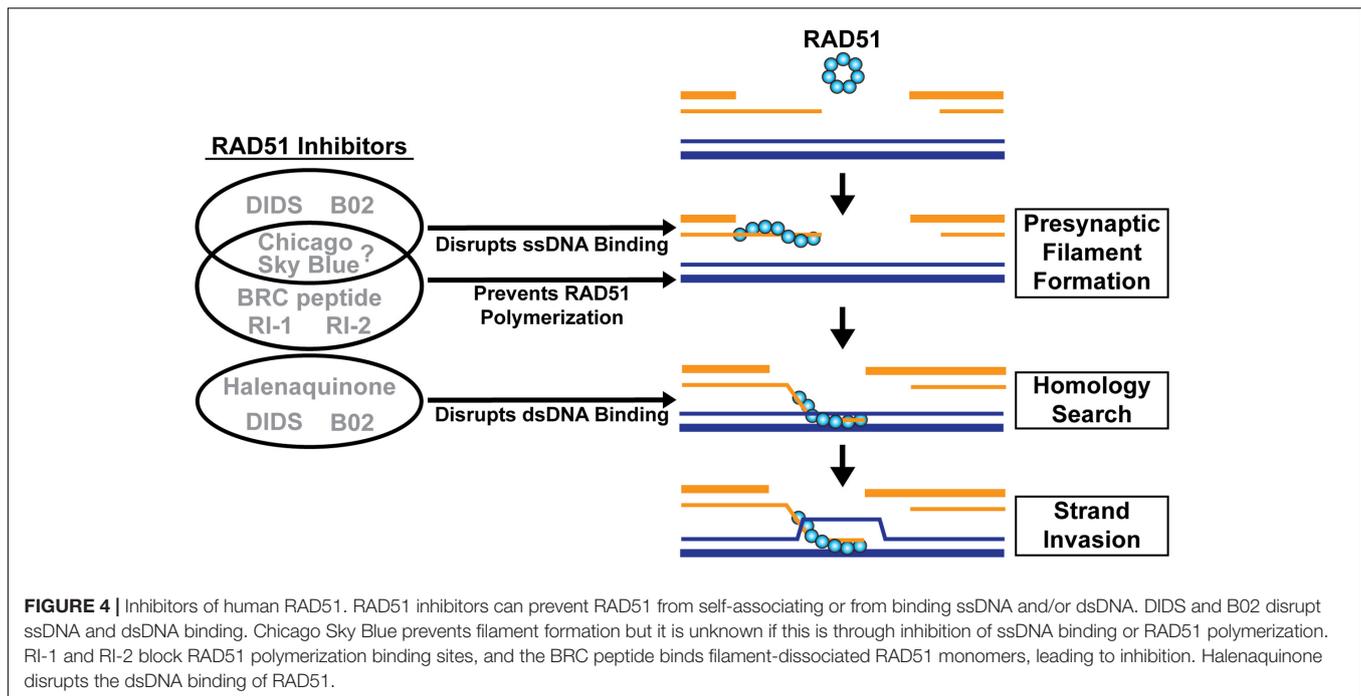


TARGETING HOMOLOGOUS RECOMBINATION WITH SMALL MOLECULE INHIBITORS

Studies of small molecule inhibitors targeting the proteins involved in HR have increased in recent years (Bryant et al., 2005; Ishida et al., 2009; Huang et al., 2011, 2012; Takaku et al., 2011; Budke et al., 2012, 2013; Huang and Mazin, 2014; Normand et al., 2014). Many current anticancer therapies, such as irradiation and chemotherapy, are designed to induce DNA damage and provoke an apoptotic response. By targeting major HR enzymes like RAD51, RAD54, and BRCA2, the susceptibility of cancer cells could increase when combined with other DNA-damaging therapies. An analogous argument could be made for

the targeting of enzymes involved in HR in protozoan parasites, which rely on HR for diverse environmental adaptations, host immune evasion, and drug resistances (Deitsch et al., 1997; Bhattacharyya et al., 2004).

RAD51 serves as a reasonable drug target for protozoan parasites because it is important for the repair of damaged DNA by HR. However, for RAD51 to be an effective target for small molecule inhibitors in these pathogens, it is essential to develop inhibitors that are species-specific, since RAD51 is also ubiquitously expressed in the human host. Currently, there is limited structural information about the recombinases, which hinders the potential for structure-based drug design. Due to the availability of purified recombinases, high throughput screens could provide a powerful platform for identifying potential



compounds that demonstrate efficient inhibition of parasitic recombinases but not human recombinases. Additionally, there are many well-characterized biochemical assays that could act as secondary analyses to aid in the understanding of mechanisms of inhibition. On a related note, recent studies have utilized small molecules, previously demonstrated to inhibit human RAD51, to target the activities of the *E. histolytica* recombinases (Kelso et al., 2015, 2016). The small molecule 4,4'-diisothiocyanostilbene-2,2'-disulfonic acid (DIDS) was found to directly interact with human RAD51 and interfere with its ability to bind ssDNA and dsDNA (Figure 4) (Ishida et al., 2009). By interfering with the ability of RAD51 to bind DNA, DIDS decreases the presynaptic filament formation of RAD51, which is essential for DNA strand exchange, D-loop formation, and ATP hydrolysis (Ishida et al., 2009). This small molecule was also shown to disrupt the recombinase activities of both *EhDmc1* and *EhRad51* *in vitro* (Kelso et al., 2015, 2016). Notably, encystation was significantly hindered in *E. invadens* when DIDS was present (Kelso et al., 2016). Despite its efficacy in these pathogens, DIDS exhibits high toxicity for human cells, complicating its use as a potential anti-pathogen treatment (Ishida et al., 2009). Interestingly, in the same study by Kelso et al. (2016), another human RAD51 small molecule inhibitor, B02, was tested. B02 was reported to disrupt RAD51 binding to ssDNA during presynaptic filament formation. In addition, B02 disrupts the subsequent binding of dsDNA to the RAD51-ssDNA complex (Figure 4) (Huang et al., 2011). As a result, B02 inhibits human RAD51-mediated DNA strand exchange and D-loop formation activities (Huang et al., 2011). *In vivo* studies revealed that B02 also prevents RAD51 foci formation at DSBs and effectively sensitizes human cells to DNA damage by

the interstrand crosslinking agents cisplatin and mitomycin-C and to PARP1 inhibitors, which target the PARP1 system responsible for repair of ssDNA breaks (Huang et al., 2011, 2012; Huang and Mazin, 2014). When tested with *EhDmc1* and *EhRad51*, B02 failed to disrupt their DNA strand exchange activity at concentrations that inhibited human RAD51 (Kelso et al., 2016). This was the first study to demonstrate species selectivity by small molecule inhibitors of human and protozoan recombinases. Importantly, these findings provide evidence that inhibitors could potentially be tailored to target specific recombinases.

There are additional small molecule inhibitors reported to disrupt human RAD51 activities that have not been characterized for their efficacy against protozoan recombinases. Halenaquinone disrupts the ability of RAD51 to bind dsDNA regardless of the presence of ssDNA (Takaku et al., 2011). This suggests that the compound binds to the dsDNA binding site, severely compromising the ability of RAD51 to catalyze homologous DNA pairing (Figure 4). Thus, *in vivo* studies demonstrated that halenaquinone suppressed the formation and retention of RAD51 foci at DSB sites (Takaku et al., 2011). Although halenaquinone interacts with RAD51 in a manner similar to DIDS, it does not exhibit the same level of toxicity for human cells (Takaku et al., 2011).

RI-1 is a compound that was shown to inhibit human RAD51 presynaptic filament formation by irreversibly binding to cysteine-319 at the oligomerization interface (Figure 4) (Budke et al., 2012). By inhibiting monomer-monomer interaction, RI-1 prevents polymerization of the RAD51 filament onto ssDNA and consequently inhibits the assembly of RAD51 foci *in vivo* (Budke et al., 2012). Additionally, RI-1 was shown to sensitize cancer cells to mitomycin-C, a compound that induces DNA

damage via cross-linking. RI-1 reacts with thiol groups (hence conjugation to cysteine-319) and therefore has the potential for off-target effects. Thus, RI-2, a derivative of RI-1, was developed to inhibit RAD51 via the same mechanism but in a reversible capacity in order to reduce the likelihood of serious off-target binding effects (**Figure 4**) (Budke et al., 2013). RI-2 was also shown to have a longer half-life than RI-1 and is a functional inhibitor of RAD51 in human cells. However, due to the nature of its reversible binding to the cysteine-319 of RAD51, RI-2 requires higher concentrations to achieve the same efficacy as the irreversibly bound RI-1 (Budke et al., 2013).

Chicago Sky Blue (CSB) is a potent inhibitor of human RAD51 activity, with sufficient inhibition observed in the 400 nM range (Normand et al., 2014). CSB inhibits D-loop formation and DNA strand exchange activity of RAD51 by preventing filament assembly of RAD51 onto ssDNA, effectively inhibiting its HR activities (Normand et al., 2014). However, it is unclear if the suppression of RAD51-ssDNA binding occurs by interfering with the ability of RAD51 to bind ssDNA (as seen with DIDS) or by disruption of protein polymerization onto ssDNA via binding at the protein-protein interface (as seen with RI-1 and RI-2) (**Figure 4**). Importantly, CSB exhibits the lowest IC₅₀ (400 nM) of the RAD51 inhibitory compounds, making it a potent RAD51 inhibitor with significant clinical potential (Normand et al., 2014).

Lastly, a short peptide of the BRC4 repeat from human BRCA2 was shown to dissociate the RAD51 filament by binding to the filament-dissociated monomers of RAD51 thereby rendering RAD51 inactive through sequestration (**Figure 4**) (Nomme et al., 2008).

Given that RAD51 is highly conserved among eukaryotes and is ubiquitously expressed, other HR proteins might serve as alternative therapeutic targets. Currently, HR in protozoan parasites is an emerging field, and therefore, the enzymes involved are not well understood. As this field develops, new HR targets that are essential and specific to parasites may emerge. Consequently, identifying if these exist in protozoan parasites would be valuable for potential pathogenic targets.

CONCLUSION AND PERSPECTIVES

Homologous recombination is an essential DNA repair process. Dysfunction of this pathway can challenge the viability and fecundity of an organism. The majority of the HR repair machinery is highly conserved among eukaryotes, including RAD51. RAD51 is responsible for DNA strand exchange and homology search via the formation of a nucleoprotein filament on ssDNA overhangs after end resection of a DSB. As a result of its central role in HR, there is merit for considering RAD51 as potential therapeutic targets in human pathogens. Presently, there is no evidence to suggest that protozoa can use the repair machinery of their host; thus, targeting protozoan RAD51 remains feasible. Interestingly, one study demonstrated the ability of a mammalian accessory protein, murine Hop2-Mnd1, to stimulate the recombinase activities of *EhRad51*, so it is reasonable to consider the possibility. Further studies will be

necessary to examine this hypothesis. Many of the RAD51 inhibitors reviewed here (DIDS, Halenaquinone, B02, CBS, BRC peptide) impede the interaction between RAD51 and DNA. As this function is essential for RAD51 filament formation and localization to DSB sites, it is an effective target to achieve inhibition of RAD51 activity in HR. Another class of RAD51 inhibitors (including RI-1 and RI-2) disrupt presynaptic filament formation by physically preventing RAD51 polymerization onto ssDNA. Again, inhibition of RAD51 at this early HR stage abolishes repair of DSBs, and therefore can lead to cell death. The ability to disrupt RAD51 in parasites and not inhibit human RAD51 will be critical for the development of new species-specific small molecule inhibitors. Analysis of the targeted impact of some of these molecules on *EhRad51* versus human RAD51 suggests that there is some degree of variance between the homologs that allows molecule specificity (note the specificity of B02 for human RAD51). Since protozoan parasites rely on HR for host immune evasion and drug resistance, targeting the major enzymes involved in the HR pathway could lead to new therapeutic intervention strategies. For example, under stressful environmental conditions, *E. histolytica* can undergo encystation for survival, which involves multiple rounds of DNA replication that produces a polyploid cyst. It has been suggested that HR plays a substantial role in encystation in *E. histolytica*, which has been modeled in the related reptilian parasite, *E. invadens*. Thus, inhibition of *Entamoeba* HR via *EhRad51* small molecule inhibitors could limit *Entamoeba* pathogenicity by obstructing the encystation pathways. Future studies are necessary to determine the efficacy of these human recombinase inhibitors on the survivability of protozoan parasites. Additionally, small molecule inhibitor studies on human RAD51 could provide a platform for future compound design against the RAD51 recombinase from other organisms.

AUTHOR CONTRIBUTIONS

AK and MS conceived and designed the study. AK, SW, AL, and MS wrote and edited the manuscript. All authors approved the content of the manuscript for accuracy.

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REFERENCES

- Badugu, S. B., Nabi, S. A., Vaidyam, P., Laskar, S., Bhattacharyya, S., and Bhattacharyya, M. K. (2015). Identification of *Plasmodium falciparum* DNA repair protein Mre11 with an evolutionarily conserved nuclease function. *PLoS ONE* 10:e0125358. doi: 10.1371/journal.pone.0125358
- Baez-Camargo, M., Gharaibeh, R., Riveron, A. M., De La Cruz Hernandez, F., Luna, J. P., Gariglio, P., et al. (1996). Gene amplification in *Entamoeba histolytica*. *Invasion Metastasis* 16, 269–279.
- Barnes, R. L., and McCulloch, R. (2007). *Trypanosoma brucei* homologous recombination is dependent on substrate length and homology, though displays a differential dependence on mismatch repair as substrate length decreases. *Nucleic Acids Res.* 35, 3478–3493. doi: 10.1093/nar/gkm249
- Betermier, M., Bertrand, P., and Lopez, B. S. (2014). Is non-homologous end-joining really an inherently error-prone process? *PLOS Genet.* 10:e1004086. doi: 10.1371/journal.pgen.1004086
- Beverley, S. M. (1991). Gene amplification in *Leishmania*. *Annu. Rev. Microbiol.* 45, 417–444. doi: 10.1146/annurev.mi.45.100191.002221
- Bhargava, R., Onyango, D. O., and Stark, J. M. (2016). Regulation of single-strand annealing and its role in genome maintenance. *Trends Genet.* 32, 566–575. doi: 10.1016/j.tig.2016.06.007
- Bhattacharya, A., Satish, S., Bagchi, A., and Bhattacharya, S. (2000). The genome of *Entamoeba histolytica*. *Int. J. Parasitol.* 30, 401–410. doi: 10.1016/S0020-7519(99)00189-7
- Bhattacharyya, M. K., Bhattacharyya Nee Deb, S., Jayabalasingham, B., and Kumar, N. (2005). Characterization of kinetics of DNA strand-exchange and ATP hydrolysis activities of recombinant PfRad51, a *Plasmodium falciparum* recombinase. *Mol. Biochem. Parasitol.* 139, 33–39. doi: 10.1016/j.molbiopara.2004.09.007
- Bhattacharyya, M. K., and Kumar, N. (2003). Identification and molecular characterisation of DNA damaging agent induced expression of *Plasmodium falciparum* recombination protein PfRad51. *Int. J. Parasitol.* 33, 1385–1392. doi: 10.1016/S0020-7519(03)00212-1
- Bhattacharyya, M. K., Norris, D. E., and Kumar, N. (2004). Molecular players of homologous recombination in protozoan parasites: implications for generating antigenic variation. *Infect. Genet. Evol.* 4, 91–98. doi: 10.1016/j.meegid.2004.01.008
- Binz, S. K., Sheehan, A. M., and Wold, M. S. (2004). Replication protein A phosphorylation and the cellular response to DNA damage. *DNA Repair* 3, 1015–1024. doi: 10.1016/j.dnarep.2004.03.028
- Bishop, D. K. (2012). Rad51, the lead in mitotic recombinational DNA repair, plays a supporting role in budding yeast meiosis. *Cell Cycle* 11, 4105–4106. doi: 10.4161/cc.22396
- Bishop, D. K., Park, D., Xu, L., and Kleckner, N. (1992). DMC1: a meiosis-specific yeast homolog of *E. coli* recA required for recombination, synaptonemal complex formation, and cell cycle progression. *Cell* 69, 439–456. doi: 10.1016/0092-8674(92)90446-J
- Borst, P. (2002). Antigenic variation and allelic exclusion. *Cell* 109, 5–8. doi: 10.1016/S0092-8674(02)00711-0
- Bryant, H. E., Schultz, N., Thomas, H. D., Parker, K. M., Flower, D., Lopez, E., et al. (2005). Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase. *Nature* 434, 913–917. doi: 10.1038/nature03443
- Budke, B., Kalin, J. H., Pawlowski, M., Zelivianskaia, A. S., Wu, M., Kozikowski, A. P., et al. (2013). An optimized RAD51 inhibitor that disrupts homologous recombination without requiring Michael acceptor reactivity. *J. Med. Chem.* 56, 254–263. doi: 10.1021/jm301565b
- Budke, B., Logan, H. L., Kalin, J. H., Zelivianskaia, A. S., Cameron Mcguire, W., Miller, L. L., et al. (2012). RI-1: a chemical inhibitor of RAD51 that disrupts homologous recombination in human cells. *Nucleic Acids Res.* 40, 7347–7357. doi: 10.1093/nar/gks353
- Burton, P., McBride, D. J., Wilkes, J. M., Barry, J. D., and McCulloch, R. (2007). Ku heterodimer-independent end joining in *Trypanosoma brucei* cell extracts relies upon sequence microhomology. *Eukaryot. Cell* 6, 1773–1781. doi: 10.1128/EC.00212-07
- Charcas-Lopez Mdel, S., Garcia-Morales, L., Pezet-Valdez, M., Lopez-Camarillo, C., Zamorano-Carrillo, A., and Marchat, L. A. (2014). Expression of EhRAD54, EhRAD51, and EhBLM proteins during DNA repair by homologous recombination in *Entamoeba histolytica*. *Parasite* 21:7. doi: 10.1051/parasite/2014006
- Chi, P., San Filippo, J., Sehorn, M. G., Petukhova, G. V., and Sung, P. (2007). Bipartite stimulatory action of the Hop2-Mnd1 complex on the Rad51 recombinase. *Genes Dev.* 21, 1747–1757. doi: 10.1101/gad.1563007
- Cleveland, L. R., and Sanders, E. P. (1930). The production of bacteria-free amoebic abscesses in the liver of cats and observations on the amoebae in various media with and without bacteria. *Science* 72, 149–151. doi: 10.1126/science.72.18.149
- Conway, C., Mcculloch, R., Ginger, M. L., Robinson, N. P., Browitt, A., and Barry, J. D. (2002a). Ku is important for telomere maintenance, but not for differential expression of telomeric VSG genes, in African trypanosomes. *J. Biol. Chem.* 277, 21269–21277.
- Conway, C., Proudfoot, C., Burton, P., Barry, J. D., and Mcculloch, R. (2002b). Two pathways of homologous recombination in *Trypanosoma brucei*. *Mol. Microbiol.* 45, 1687–1700.
- Cross, G. A. (1975). Identification, purification and properties of clone-specific glycoprotein antigens constituting the surface coat of *Trypanosoma brucei*. *Parasitology* 71, 393–417. doi: 10.1017/S003118200004717X
- Cruz, A., Coburn, C. M., and Beverley, S. M. (1991). Double targeted gene replacement for creating null mutants. *Proc. Natl. Acad. Sci. U.S.A.* 88, 7170–7174. doi: 10.1073/pnas.88.16.7170
- Deitsch, K. W., Moxon, E. R., and Wellems, T. E. (1997). Shared themes of antigenic variation and virulence in bacterial, protozoal, and fungal infections. *Microbiol. Mol. Biol. Rev.* 61, 281–293.
- Dobell, C. (1928). Researches on the intestinal protozoa of monkeys and man. *Parasitology* 20, 357–412. doi: 10.1017/S003118200001177X
- Ehrenkafer, G. M., Weedall, G. D., Williams, D., Lorenzi, H. A., Caler, E., Hall, N., et al. (2013). The genome and transcriptome of the enteric parasite *Entamoeba invadens*, a model for encystation. *Genome Biol.* 14:R77. doi: 10.1186/gb-2013-14-7-r77
- Gardner, M. J., Hall, N., Fung, E., White, O., Berriman, M., Hyman, R. W., et al. (2002). Genome sequence of the human malaria parasite *Plasmodium falciparum*. *Nature* 419, 498–511. doi: 10.1038/nature01097
- Genois, M. M., Mukherjee, A., Ubeda, J. M., Buisson, R., Paquet, E., Roy, G., et al. (2012). Interactions between BRCA2 and RAD51 for promoting homologous recombination in *Leishmania infantum*. *Nucleic Acids Res.* 40, 6570–6584. doi: 10.1093/nar/gks306
- Ghezraoui, H., Piganeau, M., Renouf, B., Renaud, J. B., Sallmyr, A., Ruis, B., et al. (2014). Chromosomal translocations in human cells are generated by canonical nonhomologous end-joining. *Mol. Cell.* 55, 829–842. doi: 10.1016/j.molcel.2014.08.002
- Glover, L., and Horn, D. (2009). Site-specific DNA double-strand breaks greatly increase stable transformation efficiency in *Trypanosoma brucei*. *Mol. Biochem. Parasitol.* 166, 194–197. doi: 10.1016/j.molbiopara.2009.03.010
- Glover, L., and Horn, D. (2012). Trypanosomal histone gammaH2A and the DNA damage response. *Mol. Biochem. Parasitol.* 183, 78–83. doi: 10.1016/j.molbiopara.2012.01.008
- Glover, L., Jun, J., and Horn, D. (2011). Microhomology-mediated deletion and gene conversion in African trypanosomes. *Nucleic Acids Res.* 39, 1372–1380. doi: 10.1093/nar/gkq981
- Glover, L., Mcculloch, R., and Horn, D. (2008). Sequence homology and microhomology dominate chromosomal double-strand break repair in African trypanosomes. *Nucleic Acids Res.* 36, 2608–2618. doi: 10.1093/nar/gkn104
- Gopalakrishnan, A. M., and Kumar, N. (2013). Opposing roles for two molecular forms of replication protein A in Rad51-Rad54-mediated DNA recombination in *Plasmodium falciparum*. *MBio* 4:e00252-13. doi: 10.1128/mBio.00252-13
- Huang, F., and Mazin, A. V. (2014). A small molecule inhibitor of human RAD51 potentiates breast cancer cell killing by therapeutic agents in mouse xenografts. *PLoS ONE* 9:e100993. doi: 10.1371/journal.pone.0100993
- Huang, F., Mazina, O. M., Zentner, I. J., Cocklin, S., and Mazin, A. V. (2012). Inhibition of homologous recombination in human cells by targeting RAD51 recombinase. *J. Med. Chem.* 55, 3011–3020. doi: 10.1021/jm201173g
- Huang, F., Motlekar, N. A., Burgwin, C. M., Napper, A. D., Diamond, S. L., and Mazin, A. V. (2011). Identification of specific inhibitors of human RAD51 recombinase using high-throughput screening. *ACS Chem. Biol.* 6, 628–635. doi: 10.1021/cb100428c

- Hunter, N. (2015). Meiotic recombination: the essence of heredity. *Cold Spring Harb. Perspect. Biol.* 7:a016618. doi: 10.1101/cshperspect.a016618
- Ishida, T., Takizawa, Y., Kainuma, T., Inoue, J., Mikawa, T., Shibata, T., et al. (2009). DIDS, a chemical compound that inhibits RAD51-mediated homologous pairing and strand exchange. *Nucleic Acids Res.* 37, 3367–3376. doi: 10.1093/nar/gkp200
- Jensen, R. B., Carreira, A., and Kowalczykowski, S. C. (2010). Purified human BRCA2 stimulates RAD51-mediated recombination. *Nature* 467, 678–683. doi: 10.1038/nature09399
- Kelso, A. A., Goodson, S. D., Chavan, S., Say, A. F., Turchick, A., Sharma, D., et al. (2016). Characterization of the recombination activities of the *Entamoeba histolytica* Rad51 recombinase. *Mol. Biochem. Parasitol.* 210, 71–84. doi: 10.1016/j.molbiopara.2016.09.001
- Kelso, A. A., Goodson, S. D., Temesvari, L. A., and Sehorn, M. G. (2017). Data on Rad51 amino acid sequences from higher and lower eukaryotic model organisms and parasites. *Data Brief* 10, 364–368. doi: 10.1016/j.dib.2016.12.002
- Kelso, A. A., Say, A. F., Sharma, D., Ledford, L. L., Turchick, A., Saski, C. A., et al. (2015). *Entamoeba histolytica* Dmc1 catalyzes homologous DNA pairing and strand exchange that is stimulated by calcium and Hop2-Mnd1. *PLOS ONE* 10:e0139399. doi: 10.1371/journal.pone.0139399
- Kirkman, L. A., Lawrence, E. A., and Deitsch, K. W. (2014). Malaria parasites utilize both homologous recombination and alternative end joining pathways to maintain genome integrity. *Nucleic Acids Res.* 42, 370–379. doi: 10.1093/nar/gkt881
- Koushik, A. B., Welter, B. H., Rock, M. L., and Temesvari, L. A. (2014). A genomewide overexpression screen identifies genes involved in the phosphatidylinositol 3-kinase pathway in the human protozoan parasite *Entamoeba histolytica*. *Eukaryot. Cell* 13, 401–411. doi: 10.1128/EC.00329-13
- Lieber, M. R. (2010). The mechanism of double-strand DNA break repair by the nonhomologous DNA end-joining pathway. *Annu. Rev. Biochem.* 79, 181–211. doi: 10.1146/annurev.biochem.052308.093131
- Liu, J., Doty, T., Gibson, B., and Heyer, W. D. (2010). Human BRCA2 protein promotes RAD51 filament formation on RPA-covered single-stranded DNA. *Nat. Struct. Mol. Biol.* 17, 1260–1262. doi: 10.1038/nsmb.1904
- Lopez-Casamichana, M., Orozco, E., Marchat, L. A., and Lopez-Camarillo, C. (2008). Transcriptional profile of the homologous recombination machinery and characterization of the EhRAD51 recombinase in response to DNA damage in *Entamoeba histolytica*. *BMC Mol. Biol.* 9:35. doi: 10.1186/1471-2199-9-35
- Lu, H., Guo, X., Meng, X., Liu, J., Allen, C., Wray, J., et al. (2005). The BRCA2-interacting protein BCCIP functions in RAD51 and BRCA2 focus formation and homologous recombinational repair. *Mol. Cell. Biol.* 25, 1949–1957. doi: 10.1128/MCB.25.5.1949-1957.2005
- Malu, S., Malshetty, V., Francis, D., and Cortes, P. (2012). Role of non-homologous end joining in V(D)J recombination. *Immunol. Res.* 54, 233–246. doi: 10.1007/s12026-012-8329-z
- Marcello, L., and Barry, J. D. (2007). Analysis of the VSG gene silent archive in *Trypanosoma brucei* reveals that mosaic gene expression is prominent in antigenic variation and is favored by archive substructure. *Genome Res.* 17, 1344–1352. doi: 10.1101/gr.6421207
- Mason, J. M., Dusad, K., Wright, W. D., Grubb, J., Budke, B., Heyer, W. D., et al. (2015). RAD54 family translocases counter genotoxic effects of RAD51 in human tumor cells. *Nucleic Acids Res.* 43, 3180–3196. doi: 10.1093/nar/gkv175
- Matos, J., and West, S. C. (2014). Holliday junction resolution: regulation in space and time. *DNA Repair* 19, 176–181. doi: 10.1016/j.dnarep.2014.03.013
- McCulloch, R., and Barry, J. D. (1999). A role for RAD51 and homologous recombination in *Trypanosoma brucei* antigenic variation. *Genes Dev.* 13, 2875–2888. doi: 10.1101/gad.13.21.2875
- McCulloch, R., Morrison, L. J., and Hall, J. P. (2015). DNA recombination strategies during antigenic variation in the African trypanosome. *Microbiol. Spectr.* 3, 1–25. doi: 10.1128/microbiolspec.MDNA3-0016-2014
- McKean, P. G., Keen, J. K., Smith, D. F., and Benson, F. E. (2001). Identification and characterisation of a RAD51 gene from *Leishmania major*. *Mol. Biochem. Parasitol.* 115, 209–216. doi: 10.1016/S0166-6851(01)00288-2
- Mehta, A., and Haber, J. E. (2014). Sources of DNA double-strand breaks and models of recombinational DNA repair. *Cold Spring Harb. Perspect. Biol.* 6:a016428. doi: 10.1101/cshperspect.a016428
- Mimori, T., and Hardin, J. A. (1986). Mechanism of interaction between Ku protein and DNA. *J. Biol. Chem.* 261, 10375–10379.
- Mlambo, G., Coppens, I., and Kumar, N. (2012). Aberrant sporogonic development of Dmc1 (a meiotic recombinase) deficient *Plasmodium berghei* parasites. *PLOS ONE* 7:e52480. doi: 10.1371/journal.pone.0052480
- Monte-Neto, R., Laffitte, M. C., Leprohon, P., Reis, P., Frezard, F., and Ouellette, M. (2015). Intrachromosomal amplification, locus deletion and point mutation in the aquaglyceroporin AQP1 gene in antimony resistant *Leishmania (Viannia) guyanensis*. *PLOS Negl. Trop. Dis.* 9:e0003476. doi: 10.1371/journal.pntd.0003476
- Moore, C. W. (1978). Responses of radiation-sensitive mutants of *Saccharomyces cerevisiae* to lethal effects of bleomycin. *Mutat. Res.* 51, 165–180. doi: 10.1016/S0027-5107(78)80016-5
- Morales, M. E., White, T. B., Strevia, V. A., Defreece, C. B., Hedges, D. J., and Deiner, P. L. (2015). The contribution of alu elements to mutagenic DNA double-strand break repair. *PLOS Genet.* 11:e1005016. doi: 10.1371/journal.pgen.1005016
- Morrill, S. W. (2015). DNA-pairing and annealing processes in homologous recombination and homology-directed repair. *Cold Spring Harb. Perspect. Biol.* 7:a016444. doi: 10.1101/cshperspect.a016444
- Moynahan, M. E., and Jasin, M. (2010). Mitotic homologous recombination maintains genomic stability and suppresses tumorigenesis. *Nat. Rev. Mol. Cell Biol.* 11, 196–207. doi: 10.1038/nrm2851
- Mukherjee, C., Clark, C. G., and Lohia, A. (2008). Entamoeba shows reversible variation in ploidy under different growth conditions and between life cycle phases. *PLOS Negl. Trop. Dis.* 2:e281. doi: 10.1371/journal.pntd.0000281
- Nathan, C., and Shiloh, M. U. (2000). Reactive oxygen and nitrogen intermediates in the relationship between mammalian hosts and microbial pathogens. *Proc. Natl. Acad. Sci. U.S.A.* 97, 8841–8848. doi: 10.1073/pnas.97.16.8841
- Nomme, J., Takizawa, Y., Martinez, S. F., Renodon-Corniere, A., Fleury, F., Weigel, P., et al. (2008). Inhibition of filament formation of human Rad51 protein by a small peptide derived from the BRC-motif of the BRCA2 protein. *Genes Cells* 13, 471–481. doi: 10.1111/j.1365-2443.2008.01180.x
- Normand, A., Riviere, E., and Renodon-Corniere, A. (2014). Identification and characterization of human Rad51 inhibitors by screening of an existing drug library. *Biochem. Pharmacol.* 91, 293–300. doi: 10.1016/j.bcp.2014.07.033
- Orozco, E., Solis, F. J., Dominguez, J., Chavez, B., and Hernandez, F. (1988). *Entamoeba histolytica*: cell cycle and nuclear division. *Exp. Parasitol.* 67, 85–95. doi: 10.1016/0014-4894(88)90011-2
- Ouellette, M., and Papadopolou, B. (1993). Mechanisms of drug resistance in *Leishmania*. *Parasitol. Today* 9, 150–153. doi: 10.1016/0169-4758(93)90135-3
- Paillard, S., and Strauss, F. (1991). Analysis of the mechanism of interaction of simian Ku protein with DNA. *Nucleic Acids Res.* 19, 5619–5624. doi: 10.1093/nar/19.20.5619
- Petukhova, G., Stratton, S., and Sung, P. (1998). Catalysis of homologous DNA pairing by yeast Rad51 and Rad54 proteins. *Nature* 393, 91–94. doi: 10.1038/30037
- Pezza, R. J., Voloshin, O. N., Vanevski, F., and Camerini-Otero, R. D. (2007). Hop2/Mnd1 acts on two critical steps in Dmc1-promoted homologous pairing. *Genes Dev.* 21, 1758–1766. doi: 10.1101/gad.1562907
- Proudfoot, C., and McCulloch, R. (2006). *Trypanosoma brucei* DMC1 does not act in DNA recombination, repair or antigenic variation in bloodstream stage cells. *Mol. Biochem. Parasitol.* 145, 245–253. doi: 10.1016/j.molbiopara.2005.10.007
- Rahman, F., Tarique, M., and Tuteja, R. (2016). *Plasmodium falciparum* Bloom homologue, a nucleocytoplasmic protein, translocates in 3' to 5' direction and is essential for parasite growth. *Biochim. Biophys. Acta* 1864, 594–608. doi: 10.1016/j.bbapap.2016.02.016
- Ratcliffe, H. L., and Geiman, Q. M. (1934). Amebiasis in reptiles. *Science* 79, 324–325. doi: 10.1126/science.79.2049.324
- Regis-da-Silva, C. G., Freitas, J. M., Passos-Silva, D. G., Furtado, C., Augusto-Pinto, L., Pereira, M. T., et al. (2006). Characterization of the *Trypanosoma cruzi* Rad51 gene and its role in recombination events associated with the parasite resistance to ionizing radiation. *Mol. Biochem. Parasitol.* 149, 191–200. doi: 10.1016/j.molbiopara.2006.05.012
- Roy, N., Bhattacharyya, S., Chakrabarty, S., Laskar, S., Babu, S. M., and Bhattacharyya, M. K. (2014). Dominant negative mutant of Plasmodium Rad51 causes reduced parasite burden in host by abrogating DNA double-strand break repair. *Mol. Microbiol.* 94, 353–366. doi: 10.1111/mmi.12762

- San Filippo, J., Sung, P., and Klein, H. (2008). Mechanism of eukaryotic homologous recombination. *Annu. Rev. Biochem.* 77, 229–257. doi: 10.1146/annurev.biochem.77.061306.125255
- Singh, N., Bhattacharya, A., and Bhattacharya, S. (2013). Homologous recombination occurs in *Entamoeba* and is enhanced during growth stress and stage conversion. *PLOS ONE* 8:e74465. doi: 10.1371/journal.pone.0074465
- Soulas-Sprauel, P., Rivera-Munoz, P., Malivert, L., Le Guyader, G., Abramowski, V., Revy, P., et al. (2007). V(D)J and immunoglobulin class switch recombinations: a paradigm to study the regulation of DNA end-joining. *Oncogene* 26, 7780–7791. doi: 10.1038/sj.onc.1210875
- Sung, P. (1997). Function of yeast Rad52 protein as a mediator between replication protein A and the Rad51 recombinase. *J. Biol. Chem.* 272, 28194–28197. doi: 10.1074/jbc.272.45.28194
- Takaku, M., Kainuma, T., Ishida-Takaku, T., Ishigami, S., Suzuki, H., Tashiro, S., et al. (2011). Halenaquinone, a chemical compound that specifically inhibits the secondary DNA binding of RAD51. *Genes Cells* 16, 427–436. doi: 10.1111/j.1365-2443.2011.01494.x
- Thorslund, T., McIlwraith, M. J., Compton, S. A., Lekomtsev, S., Petronczki, M., Griffith, J. D., et al. (2010). The breast cancer tumor suppressor BRCA2 promotes the specific targeting of RAD51 to single-stranded DNA. *Nat. Struct. Mol. Biol.* 17, 1263–1265. doi: 10.1038/nsmb.1905
- Ubeda, J. M., Legare, D., Raymond, F., Ouameur, A. A., Boisvert, S., Rigault, P., et al. (2008). Modulation of gene expression in drug resistant *Leishmania* is associated with gene amplification, gene deletion and chromosome aneuploidy. *Genome Biol.* 9:R115. doi: 10.1186/gb-2008-9-7-r115
- Ubeda, J. M., Raymond, F., Mukherjee, A., Plourde, M., Gingras, H., Roy, G., et al. (2014). Genome-wide stochastic adaptive DNA amplification at direct and inverted DNA repeats in the parasite *Leishmania*. *PLOS Biol.* 12:e1001868. doi: 10.1371/journal.pbio.1001868
- Vink, C., Rudenko, G., and Seifert, H. S. (2012). Microbial antigenic variation mediated by homologous DNA recombination. *FEMS Microbiol. Rev.* 36, 917–948. doi: 10.1111/j.1574-6976.2011.00321.x
- Wang, H., and Xu, X. (2017). Microhomology-mediated end joining: new players join the team. *Cell Biosci.* 7:6. doi: 10.1186/s13578-017-0136-8
- Yang, H., Jeffrey, P. D., Miller, J., Kinnucan, E., Sun, Y., Thoma, N. H., et al. (2002). BRCA2 function in DNA binding and recombination from a BRCA2-DSS1-ssDNA structure. *Science* 297, 1837–1848. doi: 10.1126/science.297.5588.1837
- Zhao, W., Saro, D., Hammel, M., Kwon, Y., Xu, Y., Rambo, R. P., et al. (2014). Mechanistic insights into the role of Hop2-Mnd1 in meiotic homologous DNA pairing. *Nucleic Acids Res.* 42, 906–917. doi: 10.1093/nar/gkt924
- Zhao, W., Vaithiyalingam, S., San Filippo, J., Maranon, D. G., Jimenez-Sainz, J., Fontenay, G. V., et al. (2015). Promotion of BRCA2-dependent homologous recombination by DSS1 via RPA targeting and DNA mimicry. *Mol. Cell.* 59, 176–187. doi: 10.1016/j.molcel.2015.05.032

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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