



Distribution of the Most Prevalent Spa Types among Clinical Isolates of Methicillin-Resistant and -Susceptible *Staphylococcus aureus* around the World: A Review

Parisa Asadollahi¹, Narges Nodeh Farahani¹, Mehdi Mirzaei², Seyed Sajjad Khoramrooz³, Alex van Belkum⁴, Khairollah Asadollahi^{5,6}, Masoud Dadashi⁷ and Davood Darban-Sarokhalil^{1*}

OPEN ACCESS

Edited by:

John W. A. Rossen,
University Medical Center Groningen,
Netherlands

Reviewed by:

Livia Visai,
Università degli Studi di Pavia and
Fondazione Salvatore Maugeri, Italy
Balaji Veeraraghavan,
Christian Medical College & Hospital,
India

*Correspondence:

Davood Darban-Sarokhalil
davood_darban@yahoo.com;
darban.d@iums.ac.ir

Specialty section:

This article was submitted to
Infectious Diseases,
a section of the journal
Frontiers in Microbiology

Received: 19 September 2017

Accepted: 24 January 2018

Published: 12 February 2018

Citation:

Asadollahi P, Farahani NN, Mirzaei M,
Khoramrooz SS, van Belkum A,
Asadollahi K, Dadashi M and
Darban-Sarokhalil D (2018)

Distribution of the Most Prevalent Spa
Types among Clinical Isolates of
Methicillin-Resistant and -Susceptible
Staphylococcus aureus around the
World: A Review.
Front. Microbiol. 9:163.

doi: 10.3389/fmicb.2018.00163

¹ Department of Microbiology, Faculty of Medicine, Iran University of Medical Sciences, Tehran, Iran, ² Department of Microbiology, School of Medicine, Shahrood University of Medical Sciences, Shahrood, Iran, ³ Department of Microbiology, Faculty of Medicine, Cellular and Molecular Research Center, Yasuj University of Medical Sciences, Yasuj, Iran, ⁴ Data Analytics Unit, bioMérieux 3, La Balme Les Grottes, France, ⁵ Department of Social Medicine, Faculty of Medicine, Ilam University of Medical Sciences, Ilam, Iran, ⁶ Faculty of Medicine, Biotechnology and Medicinal Plants Researches Center, Ilam University of Medical Sciences, Ilam, Iran, ⁷ Department of Microbiology, Faculty of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Background: *Staphylococcus aureus*, a leading cause of community-acquired and nosocomial infections, remains a major health problem worldwide. Molecular typing methods, such as spa typing, are vital for the control and, when typing can be made more timely, prevention of *S. aureus* spread around healthcare settings. The current study aims to review the literature to report the most common clinical spa types around the world, which is important for epidemiological surveys and nosocomial infection control policies.

Methods: A search via PubMed, Google Scholar, Web of Science, Embase, the Cochrane library, and Scopus was conducted for original articles reporting the most prevalent spa types among *S. aureus* isolates. The search terms were “*Staphylococcus aureus*, spa typing.”

Results: The most prevalent spa types were t032, t008 and t002 in Europe; t037 and t002 in Asia; t008, t002, and t242 in America; t037, t084, and t064 in Africa; and t020 in Australia. In Europe, all the isolates related to spa type t032 were MRSA. In addition, spa type t037 in Africa and t037 and t437 in Australia also consisted exclusively of MRSA isolates. Given the fact that more than 95% of the papers we studied originated in the past decade there was no option to study the dynamics of regional clone emergence.

Conclusion: This review documents the presence of the most prevalent spa types in countries, continents and worldwide and shows big local differences in clonal distribution.

Keywords: *Staphylococcus aureus*, spa typing, MRSA, prevalent, SCCmec typing, MLST, clonal complex

INTRODUCTION

Staphylococcus aureus, a leading cause of community-acquired and nosocomial infections, remains a major health problem around the world causing a variety of different conditions including wound infections, osteomyelitis, food poisoning, endocarditis, as well as more life-threatening diseases, such as pneumonia and bacteremia (Goudarzi et al., 2016b). Since the introduction of penicillin into medical therapy in the early 1940s, resistance against beta-lactams started to develop among staphylococcal isolates. To overcome this problem, a narrow spectrum semi-synthetic penicillin (methicillin) was introduced. However, soon after its first use in 1961, the first methicillin-resistant *S. aureus* (MRSA) strain was identified (Turlej et al., 2011). Methicillin resistance is caused by the *mecA* gene product, a modified form of penicillin binding protein (PBP), called PBP2a or PBP2', which has a lower affinity for all beta-lactam antibiotics (Hanssen and Ericson Sollid, 2006). The *mecA* gene is located within the *mec* operon of the staphylococcal cassette chromosome *mec* (SCCmec). SCCmec typing, which classifies SCCmec elements on the basis of their structural differences, is applied in several epidemiological studies of MRSA strains (Turlej et al., 2011). Molecular characterization of *S. aureus* is vital for the rapid identification of prevalent strains and will contribute to the control and prevention of *S. aureus* spread around healthcare settings if results are provided in real time (Siegel et al., 2007; Bosch et al., 2015; O'Hara et al., 2016). Phage typing was originally used for the formal typing of *S. aureus* isolates, but it was gradually replaced by pulsed-field gel electrophoresis (PFGE), the most recent gold standard method for the typing of *S. aureus* isolates (Bannerman et al., 1995; Murchan et al., 2003; Bosch et al., 2015). However, due to its laborious character and difficulties in exchanging data between laboratories, and the requirement for inter-laboratory standardization, PFGE was replaced by multi-locus sequence typing (MLST) and staphylococcal protein A (*spa*) typing (Harmsen et al., 2003). MLST is a great tool for evolutionary investigations and differentiates isolates according to nucleotide variations in 7 housekeeping genes. *Spa* typing, which relies only on the assessment of the number of and sequence variation in repeats at the X region of the *spa* gene, exhibits excellent discriminatory power and has become a useful typing tool for the sake of its ease of performance, cheaper procedure, and standardized nomenclature (Frenay et al., 1996; Koreen et al., 2004; Strommenger et al., 2008; Bosch et al., 2015; Darban-Sarokhalil et al., 2016; O'Hara et al., 2016). The *spa* gene contains three distinct regions: Fc, X, and C (Verweij, 1940; Harmsen et al., 2003; Goudarzi et al., 2016b). The polymorphic X region, which encodes a part of the staphylococcal protein A (Spa), contains variations in the number of tandem repeats and the base sequence within each repeat. In other words, each new sequence motif, with a length of 24 bp, found in any *S. aureus* strain is assigned a unique repeat code and the repeat succession and the precise sequences of the individual repeats for a given strain determines its *spa* type (Mazi et al., 2015). The primary binding site for protein A is the Fc region of mammalian immunoglobulins, most notably IgGs, which renders the bacteria inaccessible to opsonins,

thus impairing phagocytosis by immune system attack (Graille et al., 2000).

According to the literature, the prevalence of *spa* types among *S. aureus* isolates varies in different areas around the world. According to the authors' knowledge, no comprehensive data, during the last decade, have been made available on the distribution of diverse *spa* types within different geographical areas, so the aim of the present study was to review the literature to report the most common clinical *spa* types which is important for discriminating *S. aureus* outbreak isolates and nosocomial infection control policies worldwide.

SeqNet.org has shown the 10 most frequent *spa* types on the seqNet during 2004–2008 which includes only the European countries plus Lebanon. These data seem to include MRSA from both human and veterinary sources which is different from the present review which includes only human clinical data and a larger geographic domain.

METHODS

Search Strategy and Selection Criteria

The PubMed, Google Scholar, Web of Science, Embase, Cochrane library, and Scopus databases were searched for original articles, reporting regionally prevalent *spa* types among *S. aureus* isolates. The search terms were “*Staphylococcus aureus*, *spa* typing.”

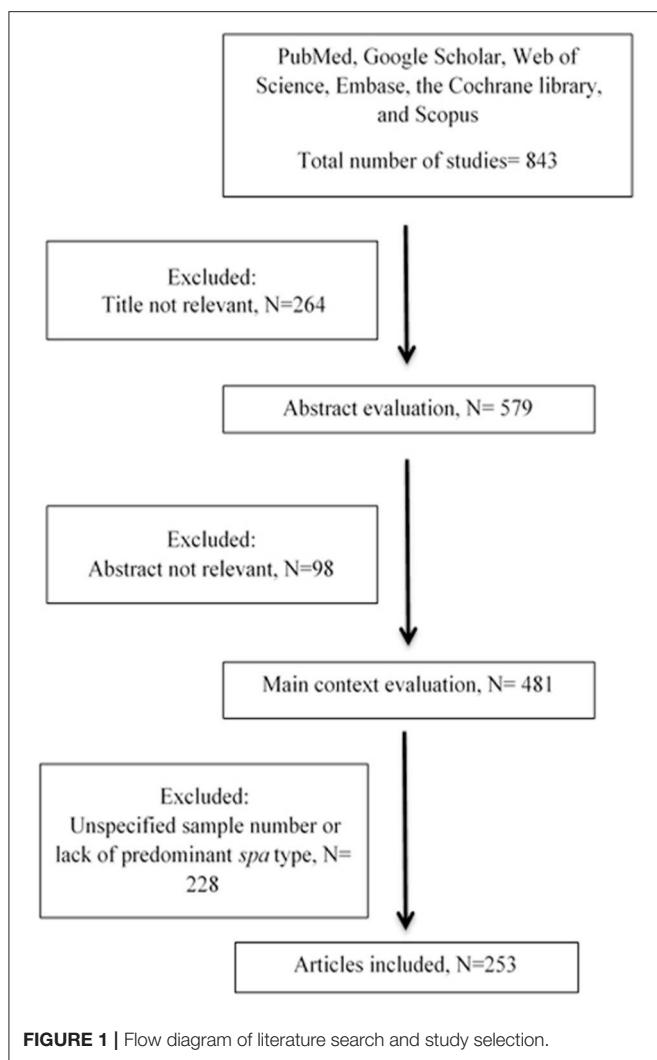
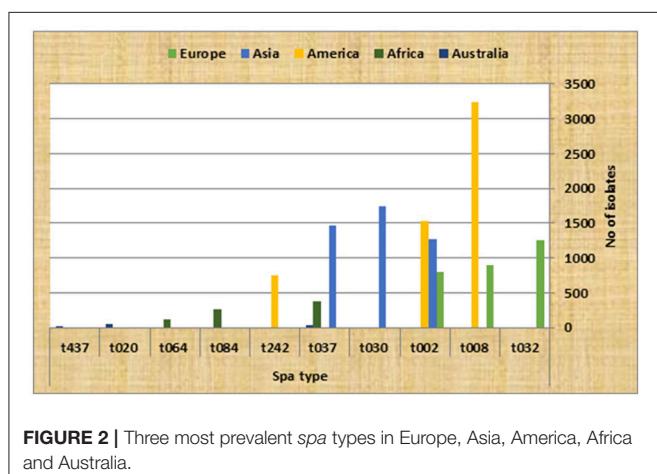
The articles were selected according to evaluations on titles, abstracts and the main text. The reasons for exclusion of certain articles were: non-human clinical isolates of *S. aureus*, old *spa*-typing methods (e.g., RFLP), non-English articles, and isolates from patients with certain specific diseases including HIV and other immune mediated afflictions. Following main text assessment, several articles were excluded for unspecified sample size or the lack of a predominant *spa* type in the study. Studies presented only in abstract form were also excluded. Papers occurring in more than a single database were cited once.

Data Extraction

The following data were extracted from each article: first author's name, year of publication, country, number of isolates, number of methicillin resistant and susceptible *S. aureus* isolates (MRSA and MSSA), the predominant *spa* type, SCCmec types of the predominant *spa* types, MLST and *spa* clonal complexes belonging to the most prevalent *spa* types.

RESULTS

During the initial database search, a total of 843 articles, from 5 continents (Europe, Asia, America, Africa and Australia), were collected among which 264 and 98 were excluded based on title and abstract evaluations, respectively (Figure 1). Out of the remaining articles, 253 fulfilled our inclusion criteria (Shopsin et al., 1999; Graille et al., 2000; Fey et al., 2003; Arakere et al., 2005; Denis et al., 2005; Ko et al., 2005; Aires-de-Sousa et al., 2006, 2008; Deplano et al., 2006; Durand et al., 2006; Ferry et al., 2006, 2010; Fossum and Bukholm, 2006; Jury et al., 2006; Kuhn et al., 2006; Mellmann et al., 2006, 2008; Montesinos et al., 2006; Ruppitsch et al., 2006, 2007; Sabat et al., 2006; Cai et al., 2007,

**FIGURE 1 |** Flow diagram of literature search and study selection.**FIGURE 2 |** Three most prevalent spa types in Europe, Asia, America, Africa and Australia.

2009; Conceicao et al., 2007; Cookson et al., 2007; Ellington et al., 2007a,b; Ghebremedhin et al., 2007; Hallin et al., 2007, 2008; Krasuski et al., 2007; Matussek et al., 2007; Otter et al., 2007;

Tristan et al., 2007; Van Loo et al., 2007; Vourli et al., 2007; Werbick et al., 2007; Witte et al., 2007; von Eiff et al., 2007; Bartels et al., 2008, 2013, 2014; Chaberny et al., 2008; Chmelnitsky et al., 2008; Gardella et al., 2008; Fenner et al., 2008a,b; Golding et al., 2008, 2011; Ho et al., 2008a,b, 2012, 2016, 2017; Jappe et al., 2008; Karynski et al., 2008; Larsen et al., 2008, 2009; Nulens et al., 2008; Pérez-Vázquez et al., 2008; Strommenger et al., 2008; Vainio et al., 2008, 2011; Zhang et al., 2008, 2009; Alp et al., 2009; Argudín et al., 2009, 2011; Atkinson et al., 2009; Bekkhoucha et al., 2009; Chen et al., 2009; Chen H.-J. et al., 2010; Chen L. et al., 2010; Chen et al., 2012, 2013, 2014; Croes et al., 2009; Khandavilli et al., 2009; Köck et al., 2009; Lamaro-Cardoso et al., 2009; Lindqvist et al., 2009, 2012, 2015; Liu et al., 2009, 2010; Melin et al., 2009; Peck et al., 2009; Rasschaert et al., 2009; Rijnders et al., 2009; Shet et al., 2009; Soliman et al., 2009; Sun et al., 2009, 2013; Vindel et al., 2009; Argudín et al., 2010; Borghi et al., 2010; Coombs et al., 2010, 2012; Geng et al., 2010a,b,c; Ghaznavi-Rad et al., 2010; Graveland et al., 2010; Grundmann et al., 2010, 2014; Holzkench et al., 2010; Ionescu et al., 2010; Laurent et al., 2010; Lee et al., 2010, 2013; Monaco et al., 2010; Moodley et al., 2010; Nadig et al., 2010, 2012; O'Sullivan et al., 2010; Petersson et al., 2010; Raulin et al., 2010; Ruimy et al., 2010; Shore et al., 2010, 2012, 2014; Valaperta et al., 2010; Wang et al., 2010, 2012, 2017; Alvarellos et al., 2011; Babouee et al., 2011; Blanco et al., 2011; Breurec et al., 2011a,b; Boakes et al., 2011; Cheng et al., 2011; Church et al., 2011; Conceição et al., 2011, 2012; García-Álvarez et al., 2011; Hesje et al., 2011; Jansen van Rensburg et al., 2011; Kechrid et al., 2011; Kim et al., 2011; Longtin et al., 2011; Miller et al., 2011; Skråmm et al., 2011; Pfingsten-Würzburg et al., 2011; Sanchini et al., 2011, 2014; Sangvik et al., 2011; Turlej et al., 2011; Uglottoli et al., 2011; Valentín-Domelier et al., 2011; Vandendriessche et al., 2011; Aamot et al., 2012, 2015; Adler et al., 2012; Berktold et al., 2012; Brennan et al., 2012; Cupane et al., 2012; Hafer et al., 2012; Hudson et al., 2012, 2013; Kriegeskorte et al., 2012; Lamand et al., 2012; Lim et al., 2012; Maeda et al., 2012; Marimón et al., 2012; Ngoa et al., 2012; Otokunefor et al., 2012; Ruffing et al., 2012; Sangal et al., 2012; Shambat et al., 2012; Sobral et al., 2012; Velasco et al., 2012; Blumenthal et al., 2013; Brauner et al., 2013; Camoëz et al., 2013; Chroboczek et al., 2013a,b; David et al., 2013; Fernandez et al., 2013; García-Garrote et al., 2013; Gómez-Sanz et al., 2013; He et al., 2013; Japoni-Nejad et al., 2013; Kwak et al., 2013; Li et al., 2013; Lozano et al., 2013; Machuca et al., 2013; Medina et al., 2013; Miko et al., 2013; Murphy et al., 2013; Price et al., 2013; Prosperi et al., 2013; Sabri et al., 2013; Schmid et al., 2013; Song et al., 2013; Tian et al., 2013; Uzunović-Kamberović et al., 2013; van der Donk et al., 2013a,b; Williamson et al., 2013; Xiao et al., 2013; Aiken et al., 2014; Casey et al., 2014; Egyir et al., 2014; Faires et al., 2014; Harastani and Tokajian, 2014; Harastani et al., 2014; Havaei et al., 2014; Holmes et al., 2014; Kachrimanidou et al., 2014; Limbago et al., 2014; Luxner et al., 2014; Mohammadi et al., 2014; Rodríguez et al., 2014; Shakeri and Ghaemi, 2014; Tavares et al., 2014; Udo et al., 2014, 2016; Uzunović et al., 2014; Wiśniewska et al., 2014; Al Laham et al., 2015; Ayepola et al., 2015; Bartoloni et al., 2015; Biber et al., 2015; de Oliveira et al., 2015; Cirković et al., 2015,?; Mirzaei et al., 2015; O'Malley et al., 2015; Perovic et al., 2015; Rajan et al., 2015; Seidl et al., 2015; Shittu et al., 2015; Yu et al., 2015; Darban-Sarokhalil

et al., 2016; Dündar et al., 2016; Garcia et al., 2016; Goudarzi et al., 2016a, 2017a,b; Jotić et al., 2016; O’Hara et al., 2016; Omuse et al., 2016; Parhizgari et al., 2016; Ahmed et al., 2017; Amissah et al., 2017; Bayat et al., 2017; Blomfeldt et al., 2017; Chmielarczyk et al., 2017; Gostev et al., 2017; Khemiri et al., 2017; Kong et al., 2017; Múnera et al., 2017; Pomorska-Wesołowska et al., 2017). In total, 127 articles were included from Europe, 70 from Asia, 33 from North and South America, 18 from Africa and 5 from Australia. More than 95% of the articles included in this study were published since 2007 and onwards. The frequent *spa* types on the different continents are shown in **Figures 2, 3 and Table 1**. The 3 most prevalent *spa* types were reported by 14, 33, and 22 out of the 127 studies in Europe, 13, 18, and 18 respective studies out of 70 in Asia, 13, 16, 2 out of 33 in America, and 3, 3, 4 out of 18 studies in Africa. Finally, in Australia, the 3 most prevalent *spa* types were reported by 1 article each out the total of 5 studies. In total, t202, t037, t437, t172, and t011 were the only *spa* types reported in Australia.

The Spa server has identified 17625 different *spa* types until the 17th of December, 2017¹. **Table 2** illustrates the distribution of diverse *spa* types among various *SCCmec* types in different continents. In Europe, 52 studies performed *SCCmec* typing on 3208 *spa* types and *SCCmec* types IV (1830 isolates) and II (800 isolates) were most associated and *SCCmec* type V (126 isolates) was least associated with the most common *spa* types. In Asia, *SCCmec* typing was performed on 4179 *spa* types by 41 studies and the most common *spa* types were classified into *SCCmec* types III (2725 isolates) and II (677 isolates), whilst the least number of *spa* types were categorized into *SCCmec* type V (104 isolates). A total of 12 studies in America performed *SCCmec* typing on 531 *spa* types, showing that *SCCmec* types IV (238 isolates) and II (167 isolates) were most associated with the frequent *spa* types. In Africa, 5 studies assessed the *SCCmec* types of 615 *spa* types and the common *spa* types were classified into *SCCmec* types IV (217 isolates) and V (185 isolates) whilst the least number of *spa* types were categorized into *SCCmec* type I (37 isolates). Finally, in Australia *SCCmec* typing was performed on 107 *spa* types by 5 studies and the most common *spa* types were classified into *SCCmec* types IV (49 isolates) and III (40 isolates), whilst the least number of *spa* types were categorized into *SCCmec* type I and II.

The total number of MRSA and MSSA isolates of the 3 most common *spa* types among different continents are shown in **Table 3**. In Europe, all the isolates related to *spa* type t032 were MRSA. In addition, *spa* type t037 in Africa and t037 and t437 in Australia were MRSA as well.

Spa clonal complex (S-CC) and MLST clonal complex (M-CC), plus the sequence types (STs) of the most common *spa* types among different continents are illustrated in **Table 4**. The number of studies that reported *spa* clonal complex for the common *spa* types were 30 in Europe, 12 in Asia, 10 in America and 9 in Africa. Common *spa* types categorized into distinct MLST clonal complexes were 43 in Europe, 19 in Asia, 14 in America, and 9 in

Africa. Forty eight studies in Europe, 29 in Asia, 18 in America, and 11 studies in Africa assessed the sequence types of the most common *spa* types. In Australia, no studies reported the *spa* or MLST complexes, nor any sequence types for the common *spa* types assessed.

The association of the most prevalent *spa* types with different countries among different continents is shown by Table S1 in Supplementary Material. The data exhibit that The Netherlands has reported the most diverse range of *spa* types (34 types), followed by China (22 *spa* types), Germany (16 types), UK (15 types), Spain (11 types), Sweden and USA (10 *spa* types each), Italy and Iran (8 *spa* types each), France and Portugal (7 *spa* types each) and Switzerland (6 *spa* types).

Dissemination of different *spa* types among different countries is illustrated by Table S2 in Supplementary Material. The *spa* types t008 and t002 were the most frequently repeated *spa* types among the others, each repeated in 16 countries among different continent. The next most frequently repeated *spa* types were respectively t037 (12 countries), t044 (11 countries), t084 (8 countries), t012 and 127 (7 countries each), t041 (6 countries), and t019, t011, t034, t355, t189, t304 (5 countries each). Almost 50% of the *spa* types (43 out of the 87) were only reported by 1 country.

DISCUSSION

Staphylococcus aureus is capable of adapting to a variety of conditions and successful clones can be epidemic and even pandemic as can be concluded by their spreading from one continent to another (Parhizgari et al., 2016). The current review reports the prevalence of *spa* types among clinical isolates, both as carriage and infectious isolates, across the world. Our analysis showed that t032 was the most prevalent *spa* type in Europe, predominantly centered in the UK and Germany (**Figure 3**), and among the 5 most predominant *spa* types in Austria. No other countries in Europe have reported t032 among its most frequent *spa* types. Moreover, t008 was the second most prevalent and, nonetheless, the most frequently identified *spa* type in the various European countries, distributed among 11 out of the 22 of them investigating local *spa* types while also being the predominant type in France and Italy (Table S2 in Supplementary Material). Germany and UK principally provided a larger sample size compared to other European countries, looking over 10081 and 2644 isolates, respectively. Despite the fact that a larger sample size could be a proof to the validity of acquired data, it might also be that the disparity of the sample size among different countries has caused deviance in the report of the most prevalent *spa* type in Europe by the present study. Sweden appeared to be the only European country to have t002 as its most predominant *spa* type, even though t002 was disseminated in 9 out of the 22 European countries included in this analysis. A comprehensive molecular-epidemiological analysis, investigating the geographical distribution of invasive *S. aureus* isolates in Europe (Grundmann et al., 2010), revealed that the 3 most common *spa* types in Europe were t032, t008, and t002, respectively; which was in agreement with the results

¹Ridom Gmb, H., info@ridom.de, Rothgaenger, J., Harmsen, D. Molecular diagnostic differentiation and typing of bacteria - software solutions. (8601); published online Epub(SCHEME=ISO8601) 2003-11-02 (<http://www.ridom.de/>).

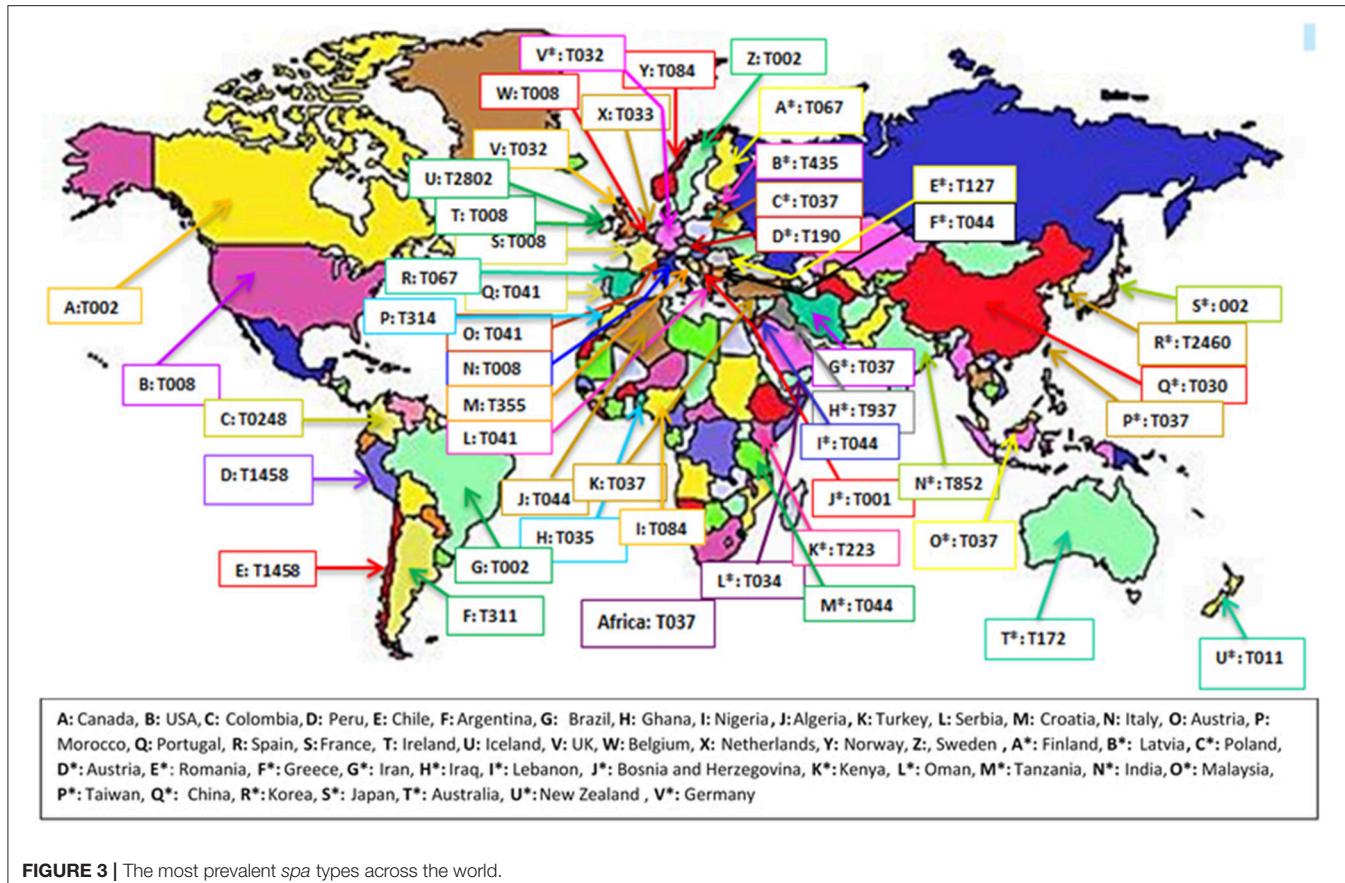


TABLE 1 | Frequency of the common spa types among different continents.

Continent	No. of isolates				The most predominant spa types (No. of isolates)
	MRSA	MSSA	Uncertified	Total	
Europe	13,988	9,767	4,565	28,320	t032 (1,250), t008 (964), t002 (794), t044 (609), t003 (596), t067 (532), t018 (458), t004 (385)
Asia	6,903	1,383	329	8,615	t030 (2,009), t037 (1,591), t002 (1,277), t437 (351), t1081 (118), t004 (116), t001 (99), t2460 (65)
America	4,828	1,126	2,187	8,141	t008 (2,100), t002 (1,569), t242 (752), t012 (285), t084 (147), t003 (99), t311 (79), t0149 (74)
Africa	1,223	577	326	2,126	t037 (394), t084 (267), t064 (123), t1257 (120), t045 (79), t012 (68), t1443 (66), t314 (37)
Australia	148	44	0	192	t202 (50), t037 (32), t437 (19), t172 (8), t011 (6)

of the current meta-analysis. In Asia, t030 was the predominant *spa* type mainly located in China (Figure 3), while also reported by Iran as the fifth most common *spa* type. Moreover, t037, as the second most common *spa* type in Asia, was reported by more Asian countries compared to other *spa* types (Korea, China, Taiwan, Iran and Malaysia out of 10 Asian countries under this survey). Similarly in Africa, t037 was the most prevalent and t084 and t064 the most frequently repeated *spa* types, reported by 3 African countries each. Even though t008 was the most prevalent *spa* type in America, it was only reported by the USA and Canada. Then again, t002, as the second most common *spa* type was distributed among the USA, Canada and Brazil. Again, for these 3 continents the distinct sample size variation within the conforming countries might account for the different reports

of the prevalent *spa* types among the associated continents. In Australia no precise information was revealed about the distribution of *spa* types.

The *spa* typing method, although being one of the valid schemes for the epidemiological surveillance of *S. aureus*, only considers a very limited portion of the whole genome and, therefore, could not possibly reflect the mutational events occurring in other parts of this organism's genome. Since certain *spa* types are still restricted to particular geographic locations, it might be considered that the polymorphic X region and, hence, the type of protein A have possible associations with the organism's adaptations to diverse conditions such as different host populations, the weather and geographical diversity.

TABLE 2 | Distribution of diverse spa types among various SCCmec types in different continents.

Continent	Spa types associated with SCCmec types (No.)				
	Type I	Type II	Type III	Type IV	Type V
Europe	t041 (189), t744 (8), t2023 (11), t002 (6), t022 (7), ?* (71)	t018 (369), t003 (33), t002 (113), t004 (196), ?* (89)	t037 (58), ?* (102)	t032 (375), t008 (328), t067 (314), t019 (66), t2802 (37), t044 (164), t002 (49), t051 (10), t038 (7), t744 (8), t304 (31), t005 (32), t515 (18), t148 (14), t024 (42), t022 (12), t127 (61), t189 (4), t030 (11), ?* (247)	t011 (60), t034 (31), t108 (11), t657 (23), t019 (1)
Asia	t127 (1), t2460 (1), t701 (2), t002 (8), t030 (3), t001 (99)	t002 (637), t2460 (36), t030 (4)	t037 (635), t071 (11), t030 (415), t002 (258), ?* (1,406)	t852 (44), t190 (7), t127 (6), t002 (23), t324 (13), t008 (31), t437 (94), t796 (3), t318 (12), t991 (12), ?* (203)	t701 (1), t002 (10), t030 (11), t081 (40), t437 (18), t657 (21), ?* (3)
America	t149 (25), t149 (18)	t002 (44), t008 (3), ?* (120)	t459 (29), t037 (7), ?* (47)	t084 (135), t002 (3), t008 (38), t045 (3), t019 (23), t024 (25), t216 (6), ?* (5)	-
Africa	t045 (37)	t311 (2), t012 (68)	t037 (106)	t044 (17), t311 (18), t186 (15), t064 (68), t1443 (66), t2196 (33)	t037 (1), t311 (1), ?* (183)
Australia	-	-	t172 (8), t037 (32)	t437 (7), t202 (42)	t437 (12), t011 (6)

*Unknown spa type.

TABLE 3 | Total number of MRSA and MSSA isolates of the most common spa types among different continents.

Continent	The most common spa types (No.): No. of MRSA and/or MSSA isolates		
Europe	t032 (1,250): 1250 MRSA	t008 (899): 510 MRSA, 229 MSSA, 90 uncertified	t002 (794): 450 MRSA, 100 MSSA, 244 uncertified
Asia	t030 (1,748): 1686 MRSA, 51 MSSA, 11uncertified	t037 (1,467): 1415 MRSA, 51 MSSA, 1 uncertified	T002 (1,285): 8064 MRSA, 9 MSSA, 340 uncertified
America	t008 (2,100): 2151 MRSA, 56 MSSA, 107 uncertified	t002 (1,525): 855 MRSA, 80 MSSA, 857 uncertified	t242 (752): 478 MRSA, 274 uncertified
Africa	t037 (381): 381 MRSA	t084 (267): 217MSSA, 50 uncertified	t064 (123): 256 MRSA, 11 uncertified
Australia	t202 (50): 50 uncertified	t037 (32): 32 MRSA	t437 (19): 19 MRSA

MRSA, methicillin resistant *Staphylococcus aureus*; MSSA, methicillin sensitive *Staphylococcus aureus*; Uncertified, not mentioned in the studies whether the isolates were MRSA or MSSA.

TABLE 4 | Spa and MLST clonal complexes plus sequence types of the most common spa types among different continents.

Continent	Prevalent spa types (No. of isolates)	Spa clonal complex/ S-CC (No. of spa types)	MLST clonal complex/M-CC (No. of spa types)	Sequence type/ST (No. of spa types)
Europe	t032 (1,250) t008 (899) t002 (794)	- S-CC008 (22) S-CC002 (58)	M-CC22 (97) M-CC8 (57) M-CC5 (162)	ST22 (173) ST8 (295), ST247 (51) ST5 (186)
Asia	t030 (1,748) t037 (1,467) t002 (1,285)	S-CC030 (121) S-CC001 (111) S-CC002 (431), SCC001/002 (8)	M-CC59 (11), M-CC8 (159) M-CC8 (198), M-CC5 (8), M-CC 188 (16) M-CC5 (145), M-CC8 (157)	ST239 (1,422), ST22 (99) ST 239 (1,124) ST5 (459)
America	t008 (2,100) t002 (1,525) t242 (752)	S-CC008 (97) S-CC002 (53)	M-CC 85 (85), M-CC5 (5) M-CC5 (30), M-CC8 (5)	ST8 (524), ST247 (100) ST5 (701) -
Africa	t037 (381) t084 (267) t064 (123)	- S-CC84 (75) S-CC64 (68)	M-CC239 (30) M-CC15 (75) M-CC8 (10), M-CC30 (68)	ST 239 (173) ST 15 (60) ST8 (68)
Australia	t202 (50) t037 (32) t437 (19)	- - -	- - -	- - -

As a vital virulence factor which enables the escape of *S. aureus* from innate and adaptive immune responses, the Spa protein may be an important target for adaptive evolution by means

of host specialization and other environmental factors (Santos-Júnior et al., 2016). The plasticity of the *spa* gene, as a result of intragenic recombination, non-synonymous mutations as well

as duplications events, can indeed influence the pathogenicity of *S. aureus*¹. It has been shown that the mosaic *spa* gene is composed of different segments, each with a distinct evolutionary histories which could provide *S. aureus* with increased fitness to colonize the host surfaces or bind the immunoglobulin subunits. This diversity of Spa domains has contributed to the epidemic phenotype of *S. aureus* strains implying that they represent selected adaptations to their environment (Santos-Júnior et al., 2016).

Considering the fact that the primary binding site for protein A is the Fc region of mammalian immunoglobulins, and most notably IgGs (Graillie et al., 2000), one possible justification for such an association might be the likely difference in the incidence rates of immunoglobulin subclasses among different geographical populations and, hence, the different binding strength of protein A types to these immunoglobulins. This might consequently cause a difference in the extent of opsonization and phagocytosis and, hence, the survival rates of particular *S. aureus* *spa* types within different populations (Sasso et al., 1991)².

Overall, t008 (2692 MRSA, 258 MSSA, 222 uncertified) and t002 (9364 MRSA, 189 MSSA and 1441 uncertified) were the most widely distributed *spa* types worldwide, disseminated each through 16 out of the 34 countries assessing *spa* types, followed by t037 (1971 MRSA, 51 MSSA, 62 uncertified) and t044 (590MRSA, 0 MSSA, 77 uncertified) respectively occurring in 12 and 11 countries worldwide. Almost half of the *spa* types (43 out of the 87) were yet localized and limited to 1 country each (Table S2 in Supplementary Material). Migrations from one country/continent to another provides a reasonable justification as to why some *spa* types are common between certain countries/continents. In Europe, all the isolates related to *spa* type t032 were MRSA isolates. In addition, *spa* type t037 in Africa and t037and t437 in Australia consisted only of MRSA isolates; however, as shown in Table 3, the majority of predominant *spa* types consist of both MRSA and MSSA isolates (Adler et al., 2012; Jiménez et al., 2013; Aiken et al., 2014). Here again, a notable number of studies have not deduced whether the predominant *spa* types are MRSA or MSSA and there is therefore some missing points in the data regarding the association of prevalent *spa* types and methicillin resistance among different continents. Furthermore, results are dependent on the original sample collection to be *spa* typed. Most studies have, in the first place, *spa* typed methicillin resistant *S. aureus* isolates because of their epidemiological importance among clinical settings (Ruppitsch et al., 2006; Zhang et al., 2008; Miller et al., 2011) and therefore no specific conclusion is to be invoked as to whether MRSA/MSSA isolates belong to specific *spa* types or vice versa.

In Europe, SCCmec types IV and II were most associated with the common *spa* types. In Asia, the most common *spa* types were classified into SCCmec types III and II. In America, SCCmec types IV and II were most associated with frequent *spa* types. In Africa, the common *spa* types were classified into SCCmec types IV and V and finally, in Australia the most common *spa* types were classified into SCCmec types IV and III. The *spa* and SCCmec typing methods focus on two distinct locations within the genome of *S. aureus*. The last SCCmec type reported in 2015 in Germany was the SCCmec type XII (Wu et al., 2015), whereas the studies assessed in this review, have only ascertained limited SCCmec types (I, II, II, IV, and V). Moreover, a significant number of *spa* types have not been associated to any specific SCCmec type and the number of studies which have assessed SCCmec typing for the prevalent *spa* types are limited. For the above mentioned reasons, the association between certain *spa* and SCCmec types found in this review might be of questionable reliability.

Data relating to the *spa* and MLST clonal complexes, and sequence types of the most common *spa* types revealed that the *spa* clonal complexes (S-CC) 001 and 002 were common among Europe and Asia and had the highest association with prevalent *spa* types in this continents. Similarly, S-CC012 contained some frequent *spa* types reported by Europe, America and Africa while S-CC84 was only common among America and Africa. This means that some related *spa* types exist among different continents. On the other hand, MLST clonal complex (M-CC) 5 was associated with prevalent *spa* types in Europe, Asia, America and Africa. Meanwhile, some of the most frequently encountered *spa* types were associated to M-CC 8 which were common among Asia, America and Africa. It seems that there is a virtually sustained association between the *spa* and sequence types irrelevant of the continent. For example, t032 has almost always been associated with ST22 across all continents; the same is true for t008 which has been associated with either ST8 or ST247, among all the studies being assessed in this review. As some of the most prevalent *spa* types reported by many different studies, t002, t030, and t037 have been constantly associated with ST5; ST239 and ST22; and ST239, respectively. In Australia, no studies reported the *spa*, MLST complexes or sequence types for the common *spa* types assessed. *S. aureus*, as an organism with a relatively stable genome, tends to present as clones which are relatively stable and generally diversify by the accumulation of single nucleotide substitutions without frequent inter-strain recombination (Grundmann et al., 2010; Shittu et al., 2011; van der Donk et al., 2013b). It is also noteworthy to mention that *S. aureus* clones might vary among different clinical settings within the same country or even among different wards of the same hospital (Shittu et al., 2011; van der Donk et al., 2013b; Seidl et al., 2015). Since a majority of studies under this review did not specifically discern the exact location of sampling, the data presented in this review presents a general information about the prevalent *spa* types and the associated clonal complexes in each country/continent; so, it would have been valuable to provide information on the exact sampling time and location within each country among different continents.

²G. healthcare. (<http://www.gelifesciences.com>), pp. DF-1.6 %ââÍÓ 1333 1330 obj <</Linearized 1331/L 3256349/O 3251336/E 3272957/N 3256173/T 3254047/H [3256516 3251548]>> endobj 3251354 3256340 obj <</DecodeParms<</Columns 3256344/Predictor 3256312>>/Filter/FlateDecode/ID/<3256387AC3256329D3256370FC3256363F3256344D3256 345CCEFB3256374F3256335F3256811><3256819D3256377F3256347 F3829348BEF3256342C3256343C3256931C3256359C3256346F>]/Index[325 1333 3256336]/Info 3251332 3256340 R/Length 3256100/Prev 3254048/Root 3251334 3256340 R/Size 3251369/Type/XRef/W[3256341 3256342 3256341]>>stream hþbbd "bâ".

CONCLUSION

This review shows the spread of the most prevalent *spa* types in countries, continents and worldwide. Such data can be used for epidemiological purposes, such as defining the geographical spread of the predominant *spa* types of *S. aureus*, the interpretation of relative frequencies, comparing the worldwide diverse evolutionary trajectories of *S. aureus* lineages, and the understanding of molecular epidemiological dynamics of *S. aureus* transmission.

AUTHOR CONTRIBUTIONS

DD-S designed the first concept, helped in the literature review, data extraction, and preparation of the manuscript.

REFERENCES

- Aamot, H. V., Blomfeldt, A., and Eskesen, A. N. (2012). Genotyping of 353 *Staphylococcus aureus* bloodstream isolates collected between 2004 and 2009 at a Norwegian university hospital and potential associations with clinical parameters. *J. Clin. Microbiol.* 50, 3111–3114. doi: 10.1128/JCM.01352-12
- Amot, H., Stavem, K., and Skråmm, I. (2015). No change in the distribution of types and antibiotic resistance in clinical *Staphylococcus aureus* isolates from orthopaedic patients during a period of 12 years. *Eur. J. Clin. Microbiol. Infect. Dis.* 34, 1833–1837. doi: 10.1007/s10096-015-2420-z
- Adler, A., Chmelnitsky, I., Shitrit, P., Sprecher, H., Navon-Venezia, S., Embon, A., et al. (2012). Molecular epidemiology of methicillin-resistant *Staphylococcus aureus* in Israel: dissemination of global clones and unique features. *J. Clin. Microbiol.* 50, 134–137. doi: 10.1128/JCM.05446-11
- Ahmed, M. O., Baptiste, K. E., Daw, M. A., Elramalli, A. K., Abouzeeda, Y. M., and Petersen, A. (2017). Spa typing and identification of pvl genes of meticillin-resistant *Staphylococcus aureus* isolated from a Libyan hospital in Tripoli. *J. Global Antimicrob. Resist.* 10, 179–181. doi: 10.1016/j.jgar.2017.06.004
- Aiken, A. M., Mutuku, I. M., Sabat, A. J., Akkerboom, V., Mwangi, J., Scott, J. A. G., et al. (2014). Carriage of *Staphylococcus aureus* in Thika Level 5 Hospital, Kenya: a cross-sectional study. *Antimicrob. Resist. Infect. Control* 3:22. doi: 10.1186/2047-2994-3-22
- Aires-de-Sousa, M., Conceicao, T. D., and De Lencastre, H. (2006). Unusually high prevalence of nosocomial Panton-Valentine leukocidin-positive *Staphylococcus aureus* isolates in Cape Verde Islands. *J. Clin. Microbiol.* 44, 3790–3793. doi: 10.1128/JCM.01192-06
- Aires-de-Sousa, M., Correia, B., and de Lencastre, H. (2008). Changing patterns in frequency of recovery of five methicillin-resistant *Staphylococcus aureus* clones in Portuguese hospitals: surveillance over a 16-year period. *J. Clin. Microbiol.* 46, 2912–2917. doi: 10.1128/JCM.00692-08
- Al Laham, N., Mediavilla, J. R., Chen, L., Abdelateef, N., Elamreen, F. A., Ginocchio, C. C., et al. (2015). MRSA clonal complex 22 strains harboring toxic shock syndrome toxin (TSST-1) are endemic in the primary hospital in Gaza, Palestine. *PLoS ONE* 10:e0120008. doi: 10.1371/journal.pone.0120008
- Alp, E., Klaassen, C. H., Doganay, M., Altoparlak, U., Aydin, K., Engin, A., et al. (2009). MRSA genotypes in Turkey: persistence over 10 years of a single clone of ST239. *J. Infect.* 58, 433–438. doi: 10.1016/j.jinf.2009.04.006
- Alvarellos, C. P., Carames, L. C., Castro, S. P., Garcia, P. A., Pi-on, J. T., and Fernandez, M. A. (2011). Usefulness of the restriction-modification test plus staphylococcal cassette chromosome mec types and Panton-Valentine leukocidin encoding phages to identify *Staphylococcus aureus* methicillin-resistant clones. *Scand. J. Infect. Dis.* 43, 943–946. doi: 10.3109/00365548.2011.589078
- Amissah, N. A., van Dam, L., Ablordey, A., Ampomah, O.-W., Prah, I., Tetteh, C. S., et al. (2017). Epidemiology of *Staphylococcus aureus* in a burn unit of a tertiary care center in Ghana. *PLoS ONE* 12:e0181072. doi: 10.1371/journal.pone.0181072
- PA prepared the manuscript, interpreted the data, and helped in the literature review and data extraction. NF helped in the literature review and data extraction. MM, SSK, and MD participated in the manuscript preparation. AvB made critical revision and helped in the preparation of the manuscript. KA helped in the analysis, interpretation of data, and preparation of the manuscript. All authors read and confirmed the content of the paper.
- SUPPLEMENTARY MATERIAL**
- The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmicb.2018.00163/full#supplementary-material>
- Arakere, G., Nadig, S., Swedberg, G., Macaden, R., Amarnath, S. K., and Ragunath, D. (2005). Genotyping of methicillin-resistant *Staphylococcus aureus* strains from two hospitals in Bangalore, South India. *J. Clin. Microbiol.* 43, 3198–3202. doi: 10.1128/JCM.43.7.3198-3202.2005
- Argudín, M. A., Mendoza, M. C., Vazquez, F., Guerra, B., and Rodicio, M. R. (2011). Molecular typing of *Staphylococcus aureus* bloodstream isolates from geriatric patients attending a long-term care Spanish hospital. *J. Med. Microbiol.* 60, 172–179. doi: 10.1099/jmm.0.021758-0
- Argudín, M., Fetsch, A., Tenhagen, A.-B., Hammerl, J., Hertwig, S., Schroeter, A., et al. (2010). High heterogeneity within methicillin-resistant *Staphylococcus aureus* ST398 isolates, defined by Cfr9I macrorestriction-pulsed-field gel electrophoresis profiles and spa and SCCmec types. *Appl. Environ. Microbiol.* 76, 652–658. doi: 10.1128/AEM.01721-09
- Argudín, M., Mendoza, M., Méndez, F., Martín, M., Guerra, B., and Rodicio, M. (2009). Clonal complexes and diversity of exotoxin gene profiles in methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* isolates from patients in a Spanish hospital. *J. Clin. Microbiol.* 47, 2097–2105. doi: 10.1128/JCM.01486-08
- Atkinson, S., Paul, J., Sloan, E., Curtis, S., and Miller, R. (2009). The emergence of methicillin-resistant *Staphylococcus aureus* among injecting drug users. *J. Infect.* 58, 339–345. doi: 10.1016/j.jinf.2009.03.004
- Ayepola, O. O., Olasupo, N. A., Egwari, L. O., Becker, K., and Schaumburg, F. (2015). Molecular characterization and antimicrobial susceptibility of *Staphylococcus aureus* isolates from clinical infection and asymptomatic carriers in Southwest Nigeria. *PLoS ONE* 10:e0137531. doi: 10.1371/journal.pone.0137531
- Babouee, B., Frei, R., Schultheiss, E., Widmer, A., and Goldenberger, D. (2011). Comparison of the DiversiLab repetitive element PCR system with spa typing and pulsed-field gel electrophoresis for clonal characterization of methicillin-resistant *Staphylococcus aureus*. *J. Clin. Microbiol.* 49, 1549–1555. doi: 10.1128/JCM.02254-10
- Bannerman, T. L., Hancock, G. A., Tenover, F. C., and Miller, J. M. (1995). Pulsed-field gel electrophoresis as a replacement for bacteriophage typing of *Staphylococcus aureus*. *J. Clin. Microbiol.* 33, 551–555.
- Bartels, M. D., Boye, K., Oliveira, D. C., Worning, P., Goering, R., and Westh, H. (2013). Associations between dru types and SCCmec cassettes. *PLoS ONE* 8:e61860. doi: 10.1371/journal.pone.0061860
- Bartels, M. D., Nanuashvili, A., Boye, K., Rohde, S. M., Jashiashvili, N., Faria, N., et al. (2008). Methicillin-resistant *Staphylococcus aureus* in hospitals in Tbilisi, the Republic of Georgia, are variants of the Brazilian clone. *Eur. J. Clin. Microbiol. Infect. Dis.* 27:757. doi: 10.1007/s10096-008-0500-z
- Bartels, M. D., Petersen, A., Worning, P., Nielsen, J. B., Larner-Svensson, H., Johansen, H. K., et al. (2014). Comparing whole-genome sequencing with Sanger sequencing for spa typing of methicillin-resistant *Staphylococcus aureus*. *J. Clin. Microbiol.* 52, 4305–4308. doi: 10.1128/JCM.01979-14
- Bartoloni, A., Riccobono, E., Magnelli, D., Villagran, A. L., Di Maggio, T., Mantella, A., et al. (2015). Methicillin-resistant *Staphylococcus aureus* in

- hospitalized patients from the Bolivian Chaco. *Int. J. Infect. Dis.* 30, 156–160. doi: 10.1016/j.ijid.2014.12.006
- Bayat, B., Zade, M. H., Mansouri, S., Kalantar, E., Kabir, K., Zahmatkesh, E., et al. (2017). High frequency of methicillin-resistant *Staphylococcus aureus* (MRSA) with SCC mec type III and spa type t030 in Karaj's teaching hospitals, Iran. *Acta Microbiol. Immunol. Hung.* 64, 1–11. doi: 10.1556/030.64.2017.020
- Bekkhoucha, S., Cady, A., Gautier, P., Itim, F., and Donnio, P.-Y. (2009). A portrait of *Staphylococcus aureus* from the other side of the Mediterranean Sea: molecular characteristics of isolates from Western Algeria. *Eur. J. Clin. Microbiol. Infect. Dis.* 28:553. doi: 10.1007/s10096-008-0660-x
- Berkhold, M., Grif, K., Mäser, M., Witte, W., Würzner, R., and Orth-Höller, D. (2012). Genetic characterization of Panton–Valentine leukocidin-producing methicillin-resistant *Staphylococcus aureus* in Western Austria. *Wien. Klin. Wochenschr.* 124, 709–715. doi: 10.1007/s00508-012-0244-8
- Biber, A., Parizade, M., Taran, D., Jaber, H., Berla, E., Rubin, C., et al. (2015). Molecular epidemiology of community-onset methicillin-resistant *Staphylococcus aureus* infections in Israel. *Eur. J. Clin. Microbiol. Infect. Dis.* 34, 1603–1613. doi: 10.1007/s10096-015-2395-9
- Blanco, R., Tristan, A., Ezpeleta, G., Larsen, A. R., Bes, M., Etienne, J., et al. (2011). Molecular epidemiology of Panton–Valentine leukocidin-positive *Staphylococcus aureus* in Spain: emergence of the USA300 clone in an autochthonous population. *J. Clin. Microbiol.* 49, 433–436. doi: 10.1128/JCM.02201-10
- Blomfeldt, A., Larssen, K., Moghen, A., Gabrielsen, C., Elström, P., Aamot, H., et al. (2017). Emerging multidrug-resistant Bengal Bay clone ST772-MRSA-V in Norway: molecular epidemiology 2004–2014. *Eur. J. Clin. Microbiol. Infect. Dis.* 36, 1911–192. doi: 10.1007/s10096-017-3014-8
- Blumenthal, S., Deplano, A., Jourdain, S., De Mendonça, R., Hallin, M., Nonhoff, C., et al. (2013). Dynamic pattern and genotypic diversity of *Staphylococcus aureus* nasopharyngeal carriage in healthy pre-school children. *J. Antimicrob. Chemother.* 68, 1517–1523. doi: 10.1093/jac/dkt080
- Boakes, E., Kearns, A., Ganner, M., Perry, C., Warner, M., Hill, R., et al. (2011). Molecular diversity within clonal complex 22 methicillin-resistant *Staphylococcus aureus* encoding Panton–Valentine leukocidin in England and Wales. *Clin. Microbiol. Infect.* 17, 140–145. doi: 10.1111/j.1469-0691.2010.03199.x
- Borghi, E., Cainarca, M., Sciota, R., Biassoni, C., and Morace, G. (2010). Molecular picture of community-and healthcare-associated methicillin-resistant *Staphylococcus aureus* circulating in a teaching hospital in Milan. *Scand. J. Infect. Dis.* 42, 873–878. doi: 10.3109/00365548.2010.508465
- Bosch, T., Pluister, G. N., Van Luit, M., Landman, F., van Santen-Verheuel, M., Schot, C., et al. (2015). Multiple-locus variable number tandem repeat analysis is superior to spa typing and sufficient to characterize MRSA for surveillance purposes. *Future Microbiol.* 10, 1155–1162. doi: 10.2217/fmb.15.35
- Brauner, J., Hallin, M., Deplano, A., De Mendonça, R., Nonhoff, C., De Ryck, R., et al. (2013). Community-acquired methicillin-resistant *Staphylococcus aureus* clones circulating in Belgium from 2005 to 2009: changing epidemiology. *Eur. J. Clin. Microbiol. Infect. Dis.* 32, 613–620. doi: 10.1007/s10096-012-1784-6
- Brennan, G. I., Shore, A. C., Corcoran, S., Tecklenborg, S., Coleman, D. C., and O'Connell, B. (2012). Emergence of hospital-and community-associated panton–valentine leukocidin-positive methicillin-resistant *Staphylococcus aureus* genotype ST772-MRSA-V in Ireland and detailed investigation of an ST772-MRSA-V cluster in a neonatal intensive care unit. *J. Clin. Microbiol.* 50, 841–847. doi: 10.1128/JCM.06354-11
- Breurec, S., Fall, C., Pouillot, R., Boisier, P., Brisse, S., Diene-Sarr, F., et al. (2011a). Epidemiology of methicillin-susceptible *Staphylococcus aureus* lineages in five major African towns: high prevalence of Panton–Valentine leukocidin genes. *Clin. Microbiol. Infect.* 17, 633–639. doi: 10.1111/j.1469-0691.2010.03320.x
- Breurec, S., Zriouil, S., Fall, C., Boisier, P., Brisse, S., Djibo, S., et al. (2011b). Epidemiology of methicillin-resistant *Staphylococcus aureus* lineages in five major African towns: emergence and spread of atypical clones. *Clin. Microbiol. Infect.* 17, 160–165. doi: 10.1111/j.1469-0691.2010.03219.x
- Cai, L., Kong, F., Wang, Q., Wang, H., Xiao, M., Sintchenko, V., et al. (2009). A new multiplex PCR-based reverse line-blot hybridization (mPCR/RLB) assay for rapid staphylococcal cassette chromosome mec (SCCmec) typing. *J. Med. Microbiol.* 58, 1045–1057. doi: 10.1099/jmm.0.007955-0
- Cai, Y., Kong, F., Wang, Q., Tong, Z., Sintchenko, V., Zeng, X., et al. (2007). Comparison of single-and multilocus sequence typing and toxin gene profiling for characterization of methicillin-resistant *Staphylococcus aureus*. *J. Clin. Microbiol.* 45, 3302–3308. doi: 10.1128/JCM.01082-07
- Camozz, M., Sierra, J. M., Pujol, M., Hornero, A., Martin, R., and Domínguez, M. A. (2013). Prevalence and molecular characterization of methicillin-resistant *Staphylococcus aureus* ST398 resistant to tetracycline at a Spanish hospital over 12 years. *PLoS ONE* 8:e72828. doi: 10.1371/journal.pone.0072828
- Casey, J. A., Shopsin, B., Cosgrove, S. E., Nachman, K. E., Curriero, F. C., Rose, H. R., et al. (2014). High-density livestock production and molecularly characterized MRSA infections in Pennsylvania. *Environ. Health Perspect.* 122:464. doi: 10.1289/ehp.1307370
- Chaberny, I. F., Bindseil, A., Sohr, D., and Gastmeier, P. (2008). A point-prevalence study for MRSA in a German university hospital to identify patients at risk and to evaluate an established admission screening procedure. *Infection* 36, 526–532. doi: 10.1007/s1510-008-7436-1
- Chen, F.-J., Hiramatsu, K., Huang, I.-W., Wang, C.-H., and Lauderdale, T.-L. Y. (2009). Panton–Valentine leukocidin (PVL)-positive methicillin-susceptible and resistant *Staphylococcus aureus* in Taiwan: identification of oxacillin-susceptible mecA-positive methicillin-resistant *S. aureus*. *Diagn. Microbiol. Infect. Dis.* 65, 351–357. doi: 10.1016/j.diagmicrobio.2009.07.024
- Chen, H.-J., Hung, W.-C., Tseng, S.-P., Tsai, J.-C., Hsueh, P.-R., and Teng, L.-J. (2010). Fusidic acid resistance determinants in *Staphylococcus aureus* clinical isolates. *Antimicrob. Agents Chemother.* 54, 4985–4991. doi: 10.1128/AAC.00523-10
- Chen, J. H.-K., Cheng, V. C.-C., Chan, J. F.-W., She, K. K.-K., Yan, M.-K., Yau, M. C.-Y., et al. (2013). The use of high-resolution melting analysis for rapid spa typing on methicillin-resistant *Staphylococcus aureus* clinical isolates. *J. Microbiol. Methods* 92, 99–102. doi: 10.1016/j.mimet.2012.11.006
- Chen, L., Mediavilla, J. R., Smyth, D. S., Chavda, K. D., Ionescu, R., Roberts, R. B., et al. (2010). Identification of a novel transposon (Tn6072) and a truncated staphylococcal cassette chromosome chromosome mec element in methicillin-resistant *Staphylococcus aureus* ST239. *Antimicrob. Agents Chemother.* 54, 3347–3354. doi: 10.1128/AAC.00001-10
- Chen, X., Yang, H.-H., Huangfu, Y.-C., Wang, W.-K., Liu, Y., Ni, Y.-X., et al. (2012). Molecular epidemiologic analysis of *Staphylococcus aureus* isolated from four burn centers. *Burns* 38, 738–742. doi: 10.1016/j.burns.2011.12.023
- Chen, Y., Liu, Z., Duo, L., Xiong, J., Gong, Y., Yang, J., et al. (2014). Characterization of *Staphylococcus aureus* from distinct geographic locations in China: an increasing prevalence of spa-t030 and SCCmec type III. *PLoS ONE* 9:e96255. doi: 10.1371/journal.pone.0096255
- Cheng, V., Chan, J., Lau, E., Yam, W., Ho, S., Yau, M., et al. (2011). Studying the transmission dynamics of methicillin-resistant *Staphylococcus aureus* in Hong Kong using spa typing. *J. Hosp. Infect.* 79, 206–210. doi: 10.1016/j.jhin.2011.03.024
- Chmelnitsky, I., Navon-Venezia, S., Leavit, A., Somekh, E., Regev-Yochai, G., Chowers, M., et al. (2008). SCCmec and spa types of methicillin-resistant *Staphylococcus aureus* strains in Israel. Detection of SCCmec type V. *Eur. J. Clin. Microbiol. Infect. Dis.* 27, 385–390. doi: 10.1007/s10096-007-0426-x
- Chmielarczyk, A., Pomorska-Wesołowska, M., Szczępta, A., Romaniszyn, D., Pobiega, M., and Wójkowska-Mach, J. (2017). Molecular analysis of methicillin-resistant *Staphylococcus aureus* strains isolated from different types of infections from patients hospitalized in 12 regional, non-teaching hospitals in southern Poland. *J. Hosp. Infect.* 95, 259–267. doi: 10.1016/j.jhin.2016.10.024
- Chroboczek, T., Boisset, S., Rasigade, J.-P., Tristan, A., Bes, M., Meugnier, H., et al. (2013b). Clonal complex 398 methicillin susceptible *Staphylococcus aureus*: a frequent unspecialized human pathogen with specific phenotypic and genotypic characteristics. *PLoS ONE* 8:e68462. doi: 10.1371/journal.pone.0068462
- Chroboczek, T., Boisset, S., Rasigade, J. P., Meugnier, H., Akpaka, P. E., Nicholson, A., et al. (2013a). Major West Indies MRSA clones in human beings: do they travel with their hosts? *J. Travel Med.* 20, 283–288. doi: 10.1111/jtm.12047
- Church, D. L., Chow, B. L., Lloyd, T., and Gregson, D. B. (2011). Comparison of automated repetitive-sequence-based polymerase chain reaction and spa typing versus pulsed-field gel electrophoresis for molecular typing of methicillin-resistant *Staphylococcus aureus*. *Diagn. Microbiol. Infect. Dis.* 69, 30–37. doi: 10.1016/j.diagmicrobio.2010.09.010
- Cirković, I., Knežević, M., Božić, D. D., Rašić, D., Larsen, A. R., and Đukić, S. (2015). Methicillin-resistant *Staphylococcus aureus* biofilm formation on

- dacryocystorhinostomy silicone tubes depends on the genetic lineage. *Graefes Arch. Clin. Exp. Ophthalmol.* 253, 77–82. doi: 10.1007/s00417-014-2786-0
- Conceição, T., Aires de Sousa, M., Miragaia, M., Paulino, E., Barroso, R., Brito, M. J., et al. (2012). *Staphylococcus aureus* reservoirs and transmission routes in a Portuguese Neonatal Intensive Care Unit: a 30-month surveillance study. *Microb. Drug Resist.* 18, 116–124. doi: 10.1089/mdr.2011.0182
- Conceicao, T., Aires-de-Sousa, M., Füzi, M., Toth, A., Paszti, J., Ungvári, E., et al. (2007). Replacement of methicillin-resistant *Staphylococcus aureus* clones in Hungary over time: a 10-year surveillance study. *Clin. Microbiol. Infect.* 13, 971–979. doi: 10.1111/j.1469-0691.2007.01794.x
- Conceição, T., Aires-de-Sousa, M., Pona, N., Brito, M., Barradas, C., Coelho, R., et al. (2011). High prevalence of ST121 in community-associated methicillin-susceptible *Staphylococcus aureus* lineages responsible for skin and soft tissue infections in Portuguese children. *Eur. J. Clin. Microbiol. Infect. Dis.* 30, 293–297. doi: 10.1007/s10096-010-1087-8
- Cookson, B. D., Robinson, D. A., Monk, A. B., Murchan, S., Deplano, A., De Ryck, R., et al. (2007). Evaluation of molecular typing methods in characterizing a European collection of epidemic methicillin-resistant *Staphylococcus aureus* strains: the HARMONY collection. *J. Clin. Microbiol.* 45, 1830–1837. doi: 10.1128/JCM.02402-06
- Coombs, G. W., Goering, R. V., Chua, K. Y., Monecke, S., Howden, B. P., Stinear, T. P., et al. (2012). The molecular epidemiology of the highly virulent ST93 Australian community *Staphylococcus aureus* strain. *PLoS ONE* 7:e43037. doi: 10.1371/journal.pone.0043037
- Coombes, G. W., Monecke, S., Ehricht, R., Slickers, P., Pearson, J. C., Tan, H.-L., et al. (2010). Differentiation of clonal complex 59 community-associated methicillin-resistant *Staphylococcus aureus* in Western Australia. *Antimicrob. Agents Chemother.* 54, 1914–1921. doi: 10.1128/AAC.01287-09
- Croes, S., Deurenberg, R. H., Boumans, M.-L. L., Beisser, P. S., Neef, C., and Stobberingh, E. E. (2009). *Staphylococcus aureus* biofilm formation at the physiologic glucose concentration depends on the *S. aureus* lineage. *BMC Microbiol.* 9:229. doi: 10.1186/1471-2180-9-229
- Cupane, L., Pugacova, N., Berzina, D., Cauce, V., Gardovska, D., and Miklaševics, E. (2012). Patients with Panton-Valentine leukocidin positive *Staphylococcus aureus* infections run an increased risk of longer hospitalisation. *Int. J. Mol. Epidemiol. Genet.* 3:48.
- Darban-Sarokhalil, D., Khoramrooz, S. S., Marashfard, M., Hosseini, S. A. A. M., Parhizgari, N., Yazdanpanah, M., et al. (2016). Molecular characterization of *Staphylococcus aureus* isolates from southwest of Iran using spa and SCCmec typing methods. *Microb. Pathog.* 98, 88–92. doi: 10.1016/j.micpath.2016.07.003
- David, M. Z., Taylor, A., Lynfield, R., Boxrud, D. J., Short, G., Zychowski, D., et al. (2013). Comparing pulsed-field gel electrophoresis with multilocus sequence typing, spa typing, staphylococcal cassette chromosome *mec* (SCCmec) typing, and PCR for Panton-Valentine leukocidin, *arcA*, and *opp3* in methicillin-resistant *Staphylococcus aureus* isolates at a US medical center. *J. Clin. Microbiol.* 51, 814–819. doi: 10.1128/JCM.02429-12
- de Oliveira, L. M., van der Heijden, I. M., Golding, G. R., Abdala, E., Freire, M. P., Rossi, F., et al. (2015). *Staphylococcus aureus* isolates colonizing and infecting cirrhotic and liver-transplantation patients: comparison of molecular typing and virulence factors. *BMC Microbiol.* 15:264. doi: 10.1186/s12866-015-0598-y
- Denis, O., Deplano, A., De Beenhouwer, H., Hallin, M., Huysmans, G., Garrino, M. G., et al. (2005). Polyclonal emergence and importation of community-acquired methicillin-resistant *Staphylococcus aureus* strains harbouring Panton-Valentine leukocidin genes in Belgium. *J. Antimicrob. Chemother.* 56, 1103–1106. doi: 10.1093/jac/dki379
- Deplano, A., De Mendonça, R., De Ryck, R., and Struelens, M. (2006). External quality assessment of molecular typing of *Staphylococcus aureus* isolates by a network of laboratories. *J. Clin. Microbiol.* 44, 3236–3244. doi: 10.1128/JCM.00789-06
- Dündar, D., Willke, A., Sayan, M., Koc, M. M., Akan, O. A., Sumerkan, B., et al. (2016). Epidemiological and molecular characteristics of methicillin-resistant *Staphylococcus aureus* in Turkey: a multicentre study. *J. Global Antimicrob. Resist.* 6, 44–49. doi: 10.1016/j.jgar.2016.02.006
- Durand, G., Bes, M., Meugnier, H., Enright, M. C., Forey, F., Liassine, N., et al. (2006). Detection of new methicillin-resistant *Staphylococcus aureus* clones containing the toxic shock syndrome toxin 1 gene responsible for hospital- and community-acquired infections in France. *J. Clin. Microbiol.* 44, 847–853. doi: 10.1128/JCM.44.3.847-853.2006
- Egyir, B., Guardabassi, L., Esson, J., Nielsen, S. S., Newman, M. J., Addo, K. K., et al. (2014). Insights into nasal carriage of *Staphylococcus aureus* in an urban and a rural community in Ghana. *PLoS ONE* 9:e96119. doi: 10.1371/journal.pone.0096119
- Ellington, M. J., Hope, R., Ganner, M., East, C., Brick, G., et al. (2007a). Is Panton-Valentine leucocidin associated with the pathogenesis of *Staphylococcus aureus* bacteraemia in the UK? *J. Antimicrob. Chemother.* 60, 402–405. doi: 10.1093/jac/dkm206
- Ellington, M. J., Yearwood, L., Ganner, M., East, C., and Kearns, A. M. (2007b). Distribution of the ACME-arcA gene among methicillin-resistant *Staphylococcus aureus* from England and Wales. *J. Antimicrob. Chemother.* 61, 73–77. doi: 10.1093/jac/dkm422
- Faires, M. C., Pearl, D. L., Ciccotelli, W. A., Berke, O., Reid-Smith, R. J., and Weese, J. S. (2014). The use of the temporal scan statistic to detect methicillin-resistant *Staphylococcus aureus* clusters in a community hospital. *BMC Infect. Dis.* 14:375. doi: 10.1186/1471-2334-14-375
- Fenner, L., Widmer, A. F., Dangel, M., and Frei, R. (2008a). Distribution of spa types among methicillin-resistant *Staphylococcus aureus* isolates during a 6 year period at a low-prevalence university hospital. *J. Med. Microbiol.* 57, 612–616. doi: 10.1099/jmm.0.47757-0
- Fenner, L., Widmer, A., and Frei, R. (2008b). Molecular epidemiology of invasive methicillin-susceptible *Staphylococcus aureus* strains circulating at a Swiss University Hospital. *Eur. J. Clin. Microbiol. Infect. Dis.* 27, 623–626. doi: 10.1007/s10096-008-0463-0
- Fernandez, S., de Vedia, L., Furst, M. L., Gardella, N., Di Gregorio, S., Ganaha, M., et al. (2013). Methicillin-resistant *Staphylococcus aureus* ST30-SCCmec IV clone as the major cause of community-acquired invasive infections in Argentina. *Infect. Genet. Evol.* 14, 401–405. doi: 10.1016/j.meegid.2012.12.018
- Ferry, T., Bes, M., Dauwalder, O., Meugnier, H., Lina, G., Forey, F., et al. (2006). Toxin gene content of the Lyon methicillin-resistant *Staphylococcus aureus* clone compared with that of other pandemic clones. *J. Clin. Microbiol.* 44, 2642–2644. doi: 10.1128/JCM.00430-06
- Ferry, T., Uckay, I., Vaudaux, P., Francois, P., Schrenzel, J., Harbarth, S., et al. (2010). Risk factors for treatment failure in orthopedic device-related methicillin-resistant *Staphylococcus aureus* infection. *Eur. J. Clin. Microbiol. Infect. Dis.* 29, 171–180. doi: 10.1007/s10096-009-0837-y
- Fey, P., Said-Salim, B., Rupp, M., Hinrichs, S., Boxrud, D., Davis, C., et al. (2003). Comparative molecular analysis of community-or hospital-acquired methicillin-resistant *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* 47, 196–203. doi: 10.1128/AAC.47.1.196-203.2003
- Fossum, A., and Bukholm, G. (2006). Increased incidence of methicillin-resistant *Staphylococcus aureus* ST80, novel ST125 and SCCmecIV in the south-eastern part of Norway during a 12-year period. *Clin. Microbiol. Infect.* 12, 627–633. doi: 10.1111/j.1469-0691.2006.01467.x
- Frenay, H., Bunschoten, A., Schouls, L., Van Leeuwen, W., Vandebroucke-Grauls, C., Verhoef, J., et al. (1996). Molecular typing of methicillin-resistant *Staphylococcus aureus* on the basis of protein A gene polymorphism. *Eur. J. Clin. Microbiol. Infect. Dis.* 15, 60–64. doi: 10.1007/BF01586186
- Garcia, C., Acuña-Villaorduña, A., Dulanto, A., Vandendriessche, S., Hallin, M., Jacobs, J., et al. (2016). Dynamics of nasal carriage of methicillin-resistant *Staphylococcus aureus* among healthcare workers in a tertiary-care hospital in Peru. *Eur. J. Clin. Microbiol. Infect. Dis.* 35, 89–93. doi: 10.1007/s10096-015-2512-9
- García-Álvarez, L., Holden, M. T., Lindsay, H., Webb, C. R., Brown, D. F., Curran, M. D., et al. (2011). Meticillin-resistant *Staphylococcus aureus* with a novel *mecA* homologue in human and bovine populations in the UK and Denmark: a descriptive study. *Lancet Infect. Dis.* 11, 595–603. doi: 10.1016/S1473-3099(11)70126-8
- García-Garrote, F., Cercenado, E., Marín, M., Bal, M., Trincado, P., Corredoira, J., et al. (2013). Methicillin-resistant *Staphylococcus aureus* carrying the *mecC* gene: emergence in Spain and report of a fatal case of bacteraemia. *J. Antimicrob. Chemother.* 69, 45–50. doi: 10.1093/jac/dkt327
- Gardella, N., von Specht, M., Cuirolo, A., Rosato, A., Gutkind, G., and Mollerach, M. (2008). Community-associated methicillin-resistant *Staphylococcus aureus*, eastern Argentina. *Diagn. Microbiol. Infect. Dis.* 62, 343–347. doi: 10.1016/j.diagmicrobio.2008.07.008
- Geng, W., Yang, Y., Wang, C., Deng, L., Zheng, Y., and Shen, X. (2010a). Skin and soft tissue infections caused by community-associated methicillin-resistant

- Staphylococcus aureus* among children in China. *Acta Paediatr.* 99, 575–580. doi: 10.1111/j.1651-2227.2009.01645.x
- Geng, W., Yang, Y., Wu, D., Huang, G., Wang, C., Deng, L., et al. (2010b). Molecular characteristics of community-acquired, methicillin-resistant *Staphylococcus aureus* isolated from Chinese children. *FEMS Immunol. Med. Microbiol.* 58, 356–362. doi: 10.1111/j.1574-695X.2009.00648.x
- Geng, W., Yang, Y., Wu, D., Zhang, W., Wang, C., Shang, Y., et al. (2010c). Community-acquired, methicillin-resistant *Staphylococcus aureus* isolated from children with community-onset pneumonia in China. *Pediatr. Pulmonol.* 45, 387–394. doi: 10.1002/ppul.21202
- Ghaznavi-Rad, E., Shamsudin, M. N., Sekawi, Z., Khoon, L. Y., Aziz, M. N., Hamat, R. A., et al. (2010). Predominance and emergence of clones of hospital-acquired methicillin-resistant *Staphylococcus aureus* in Malaysia. *J. Clin. Microbiol.* 48, 867–872. doi: 10.1128/JCM.01112-09
- Ghebremedhin, B., König, W., Witte, W., Hardy, K., Hawkey, P., and König, B. (2007). Subtyping of ST22-MRSA-IV (Barnim epidemic MRSA strain) at a university clinic in Germany from 2002 to 2005. *J. Med. Microbiol.* 56, 365–375. doi: 10.1099/jmm.0.46883-0
- Golding, G. R., Campbell, J. L., Spreitzer, D. J., Veyhl, J., Surynicz, K., Simor, A., et al. (2008). A preliminary guideline for the assignment of methicillin-resistant *Staphylococcus aureus* to a Canadian pulsed-field gel electrophoresis epidemic type using spa typing. *Can. J. Infect. Dis. Med. Microbiol.* 19, 273–281. doi: 10.1155/2008/754249
- Golding, G. R., Levett, P. N., McDonald, R. R., Irvine, J., Quinn, B., Nsungu, M., et al. (2011). High rates of *Staphylococcus aureus* USA400 infection, Northern Canada. *Emerging Infect. Dis.* 17:722. doi: 10.3201/eid1704.100482
- Gómez-Sanz, E., Torres, C., Lozano, C., and Zarazaga, M. (2013). High diversity of *Staphylococcus aureus* and *Staphylococcus pseudintermedius* lineages and toxicogenic traits in healthy pet-owning household members. Underestimating normal household contact? *Comp. Immunol. Microbiol. Infect. Dis.* 36, 83–94. doi: 10.1016/j.cimid.2012.10.001
- Gostev, V., Kruglov, A., Kalinogorskaya, O., Dmitrenko, O., Khokhlova, O., Yamamoto, T., et al. (2017). Molecular epidemiology and antibiotic resistance of methicillin-resistant *Staphylococcus aureus* circulating in the Russian Federation. *Infect. Genet. Evol.* 53, 189–194. doi: 10.1016/j.meegid.2017.06.006
- Goudarzi, M., Bahramian, M., Tabrizi, M. S., Udo, E. E., Figueiredo, A. M. S., Fazeli, M., et al. (2017a). Genetic diversity of methicillin resistant *Staphylococcus aureus* strains isolated from burn patients in Iran: ST239-SCCmec III/t037 emerges as the major clone. *Microb. Pathog.* 105, 1–7. doi: 10.1016/j.micpath.2017.02.004
- Goudarzi, M., Fazeli, M., Goudarzi, H., Azad, M., and Seyedjavadi, S. S. (2016b). Spa typing of *Staphylococcus aureus* strains isolated from clinical specimens of patients with nosocomial infections in Tehran, Iran. *Jundishapur J. Microbiol.* 9:e35685. doi: 10.5812/jjm.35685
- Goudarzi, M., Goudarzi, H., Figueiredo, A. M. S., Udo, E. E., Fazeli, M., Asadzadeh, M., et al. (2016a). Molecular characterization of methicillin resistant *Staphylococcus aureus* strains isolated from intensive care units in Iran: ST22-SCCmec IV/t790 emerges as the major clone. *PLoS ONE* 11:e0155529. doi: 10.1371/journal.pone.0155529
- Goudarzi, M., Seyedjavadi, S. S., Nasiri, M. J., Goudarzi, H., Nia, R. S., and Dabiri, H. (2017b). Molecular characteristics of methicillin-resistant *Staphylococcus aureus* (MRSA) strains isolated from patients with bacteremia based on MLST, SCCmec, spa, and agr locus types analysis. *Microb. Pathog.* 104, 328–335. doi: 10.1016/j.micpath.2017.01.055
- Graille, M., Stura, E. A., Corper, A. L., Sutton, B. J., Taussig, M. J., Charbonnier, J.-B., et al. (2000). Crystal structure of a *Staphylococcus aureus* protein A domain complexed with the Fab fragment of a human IgM antibody: structural basis for recognition of B-cell receptors and superantigen activity. *Proc. Natl. Acad. Sci. U.S.A.* 97, 5399–5404. doi: 10.1073/pnas.97.10.5399
- Graveland, H., Wagenaar, J. A., Heesterbeek, H., Mevius, D., Van Duijkeren, E., and Heederik, D. (2010). Methicillin resistant *Staphylococcus aureus* ST398 in veal calf farming: human MRSA carriage related with animal antimicrobial usage and farm hygiene. *PLoS ONE* 5:e10990. doi: 10.1371/journal.pone.0010990
- Grundmann, H., Aanensen, D. M., Van Den Wijngaard, C. C., Spratt, B. G., Harmsen, D., Friedrich, A. W., et al. (2010). Geographic distribution of *Staphylococcus aureus* causing invasive infections in Europe: a molecular-epidemiological analysis. *PLoS Med.* 7:e1000215. doi: 10.1371/journal.pmed.1000215
- Grundmann, H., Schouls, L. M., Aanensen, D. M., Pluister, G. N., Tami, A., Chlebowicz, M., et al. (2014). The dynamic changes of dominant clones of *Staphylococcus aureus* causing bloodstream infections in the European region: results of a second structured survey. *Euro Surveill.* 19:20987.
- Hafer, C., Lin, Y., Kornblum, J., Lowy, F. D., and Uhlemann, A.-C. (2012). Contribution of selected gene mutations to resistance in clinical isolates of vancomycin-intermediate *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* 56, 5845–5851. doi: 10.1128/AAC.01139-12
- Hallin, M., Denis, O., Deplano, A., De Mendonça, R., De Ryck, R., Rottiers, S., et al. (2007). Genetic relatedness between methicillin-susceptible and methicillin-resistant *Staphylococcus aureus*: results of a national survey. *J. Antimicrob. Chemother.* 59, 465–472. doi: 10.1093/jac/dkl535
- Hallin, M., Denis, O., Deplano, A., De Ryck, R., Crèvecoeur, S., Rottiers, S., et al. (2008). Evolutionary relationships between sporadic and epidemic strains of healthcare-associated methicillin-resistant *Staphylococcus aureus*. *Clin. Microbiol. Infect.* 14, 659–669. doi: 10.1111/j.1469-0961.2008.02015.x
- Hanssen, A. M., and Ericson Söllid, J. U. (2006). SCCmec in staphylococci: genes on the move. *Pathog. Dis.* 46, 8–20. doi: 10.1111/j.1574-695X.2005.00009.x
- Harastani, H. H., and Tokajian, S. T. (2014). Community-associated methicillin-resistant *Staphylococcus aureus* clonal complex 80 type IV (CC80-MRSA-IV) isolated from the Middle East: a heterogeneous expanding clonal lineage. *PLoS ONE* 9:e103715. doi: 10.1371/journal.pone.0103715
- Harastani, H. H., Araj, G. F., and Tokajian, S. T. (2014). Molecular characteristics of *Staphylococcus aureus* isolated from a major hospital in Lebanon. *Int. J. Infect. Dis.* 19, 33–38. doi: 10.1016/j.ijid.2013.10.007
- Harmsen, D., Claus, H., Witte, W., Rothgänger, J., Claus, H., Turnwald, D., et al. (2003). Typing of methicillin-resistant *Staphylococcus aureus* in a university hospital setting by using novel software for spa repeat determination and database management. *J. Clin. Microbiol.* 41, 5442–5448. doi: 10.1128/JCM.41.12.5442-5448.2003
- Havaei, S. A., Ghanbari, F., Rastegari, A. A., Azimian, A., Khademi, F., Hosseini, N., et al. (2014). Molecular typing of hospital-acquired *Staphylococcus aureus* isolated from Isfahan, Iran. 2014, 1–6. *Int. Schol. Res. Not.* doi: 10.1155/2014/185272
- He, W., Chen, H., Zhao, C., Zhang, F., Li, H., Wang, Q., et al. (2013). Population structure and characterisation of *Staphylococcus aureus* from bacteraemia at multiple hospitals in China: association between antimicrobial resistance, toxin genes and genotypes. *Int. J. Antimicrob. Agents* 42, 211–219. doi: 10.1016/j.ijantimicag.2013.04.031
- Hesje, C. K., Sanfilippo, C. M., Haas, W., and Morris, T. W. (2011). Molecular epidemiology of methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* isolated from the eye. *Curr. Eye Res.* 36, 94–102. doi: 10.3109/02713683.2010.534229
- Ho, C.-M., Ho, M.-W., Lee, C.-Y., Tien, N., and Lu, J.-J. (2012). Clonal spreading of methicillin-resistant SCCmec *Staphylococcus aureus* with specific spa and dru types in central Taiwan. *Eur. J. Clin. Microbiol. Infect. Dis.* 31, 499–504. doi: 10.1007/s10096-011-1338-3
- Ho, C.-M., Lin, C.-Y., Ho, M.-W., Lin, H.-C., Peng, C.-T., and Lu, J.-J. (2016). Concomitant genotyping revealed diverse spreading between methicillin-resistant *Staphylococcus aureus* and methicillin-susceptible *Staphylococcus aureus* in central Taiwan. *J. Microbiol. Immunol. Infect.* 49, 363–370. doi: 10.1016/j.jmii.2014.07.010
- Ho, P.-L., Chuang, S.-K., Choi, Y.-F., Lee, R. A., Lit, A. C., Nig, T.-L., et al. (2008a). Community-associated methicillin-resistant and methicillin-sensitive *Staphylococcus aureus*: skin and soft tissue infections in Hong Kong. *Diagn. Microbiol. Infect. Dis.* 61, 245–250. doi: 10.1016/j.diamond.2007.12.015
- Ho, P.-L., Lai, E. L., Chow, K.-H., Chow, L. S., Yuen, K.-Y., and Yung, R. W. (2008b). Molecular epidemiology of methicillin-resistant *Staphylococcus aureus* in residential care homes for the elderly in Hong Kong. *Diagn. Microbiol. Infect. Dis.* 61, 135–142. doi: 10.1016/j.diamond.2007.12.017
- Ho, W.-Y., Choo, Q.-C., and Chew, C.-H. (2017). Predominance of Three Closely Related Methicillin-Resistant *Staphylococcus aureus* Clones Carrying a Unique ccrC-Positive SCC mec type III and the Emergence of spa t304 and t690 SCC mec type IV pvl+ MRSA Isolates in Kinta Valley, Malaysia. *Microbial. Drug Resist.* 23, 215–223. doi: 10.1089/mdr.2015.0250
- Holmes, A., McAllister, G., McAdam, P., Choi, S. H., Girvan, K., Robb, A., et al. (2014). Genome-wide single nucleotide polymorphism-based assay for high-resolution epidemiological analysis of the methicillin-resistant *Staphylococcus*

- aureus* hospital clone EMRSA-15. *Clin. Microbiol. Infect.* 20, O124–O131. doi: 10.1111/1469-0691.12328
- Holzknecht, B. J., Hardardottir, H., Haraldsson, G., Westh, H., Valsdottir, F., Boye, K., et al. (2010). Changing epidemiology of methicillin-resistant *Staphylococcus aureus* in Iceland from 2000 to 2008: a challenge to current guidelines. *J. Clin. Microbiol.* 48, 4221–4227. doi: 10.1128/JCM.01382-10
- Hudson, L. O., Murphy, C. R., Spratt, B. G., Enright, M. C., Elkins, K., Nguyen, C., et al. (2013). Diversity of methicillin-resistant *Staphylococcus aureus* (MRSA) strains isolated from inpatients of 30 hospitals in Orange County, California. *PLoS ONE* 8:e62117. doi: 10.1371/journal.pone.0062117
- Hudson, L. O., Murphy, C. R., Spratt, B. G., Enright, M. C., Terpstra, L., Gombosov, A., et al. (2012). Differences in methicillin-resistant *Staphylococcus aureus* strains isolated from pediatric and adult patients from hospitals in a large county in California. *J. Clin. Microbiol.* 50, 573–579. doi: 10.1128/JCM.05336-11
- Ionescu, R., Mediavilla, J. R., Chen, L., Grigorescu, D. O., Idomir, M., Kreiswirth, B. N., et al. (2010). Molecular characterization and antibiotic susceptibility of *Staphylococcus aureus* from a multidisciplinary hospital in Romania. *Microbial. Drug Resist.* 16, 263–272. doi: 10.1089/mdr.2010.0059
- Jansen van Rensburg, M., Eliya Madikane, V., Whitelaw, A., Chachage, M., Haffejee, S., and Gay Elisha, B. (2011). The dominant methicillin-resistant *Staphylococcus aureus* clone from hospitals in Cape Town has an unusual genotype: ST612. *Clin. Microbiol. Infect.* 17, 785–792. doi: 10.1111/j.1469-0691.2010.03373.x
- Japoni-Nejad, A., Rezazadeh, M., Kazemian, H., Fardmousavi, N., van Belkum, A., and Ghaznavi-Rad, E. (2013). Molecular characterization of the first community-acquired methicillin-resistant *Staphylococcus aureus* strains from Central Iran. *Int. J. Infect. Dis.* 17, e949–e954. doi: 10.1016/j.ijid.2013.03.023
- Jappe, U., Heuck, D., Strommenger, B., Wendt, C., Werner, G., Altmann, D., et al. (2008). *Staphylococcus aureus* in dermatology outpatients with special emphasis on community-associated methicillin-resistant strains. *J. Investig. Dermatol.* 128, 2655–2664. doi: 10.1038/jid.2008.133
- Jiménez, J. N., Ocampo, A. M., Vanegas, J. M., Rodriguez, E. A., Mediavilla, J. R., Chen, L., et al. (2013). A comparison of methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* reveals no clinical and epidemiological but molecular differences. *Int. J. Med. Microbiol.* 303, 76–83. doi: 10.1016/j.ijmm.2012.12.003
- Jotić, A., Božić, D. D., Milovanović, J., Pavlović, B., Ješić, S., Pelešić, M., et al. (2016). Biofilm formation on tympanostomy tubes depends on methicillin-resistant *Staphylococcus aureus* genetic lineage. *Eur. Arch. Oto Rhinol Laryngol.* 273, 615–620. doi: 10.1007/s00405-015-3607-8
- Jury, F., Al-Mahrouse, M., Apostolou, M., Sandiford, S., Fox, A., Ollier, W., et al. (2006). Rapid cost-effective subtyping of methicillin-resistant *Staphylococcus aureus* by denaturing HPLC. *J. Med. Microbiol.* 55, 1053–1060. doi: 10.1099/jmm.0.46409-0
- Kachrimanidou, M., Tsorlini, E., Katsifa, E., VLachou, S., Kyriakidou, S., Xanthopoulou, K., et al. (2014). Prevalence and molecular epidemiology of methicillin-resistant *Staphylococcus aureus* in a tertiary Greek hospital. *Hippokratia* 18:24.
- Karynski, M., Sabat, A. J., Empel, J., and Hryniewicz, W. (2008). Molecular surveillance of methicillin-resistant *Staphylococcus aureus* by multiple-locus variable number tandem repeat fingerprinting (formerly multiple-locus variable number tandem repeat analysis) and spa typing in a hierachic approach. *Diagn. Microbiol. Infect. Dis.* 62, 255–262. doi: 10.1016/j.diagmicrobio.2008.06.019
- Kechedid, A., Pérez-Vázquez, M., Smaoui, H., Hariga, D., Rodríguez-Baños, M., Vindel, A., et al. (2011). Molecular analysis of community-acquired methicillin-susceptible and resistant *Staphylococcus aureus* isolates recovered from bacteraemic and osteomyelitis infections in children from Tunisia. *Clin. Microbiol. Infect.* 17, 1020–1026. doi: 10.1111/j.1469-0691.2010.03367.x
- Khandavilli, S., Wilson, P., Cookson, B., Cepeda, J., Bellangan, G., and Brown, J. (2009). Utility of spa typing for investigating the local epidemiology of MRSA on a UK intensive care ward. *J. Hosp. Infect.* 71, 29–35. doi: 10.1016/j.jhin.2008.09.011
- Khemiri, M., Alhusain, A. A., Abbassi, M. S., El Ghaieb, H., Costa, S. S., Belas, A., et al. (2017). Clonal spread of methicillin-resistant *Staphylococcus aureus*-t6065-CC5-SCCmecV-agrII in a Libyan hospital. *J. Glob. Antimicrobial Resist.* 10, 101–105. doi: 10.1016/j.jgar.2017.04.014
- Kim, T., Yi, J., Hong, K. H., Park, J.-S., and Kim, E.-C. (2011). Distribution of virulence genes in spa types of methicillin-resistant *Staphylococcus aureus* isolated from patients in intensive care units. *Korean J. Lab. Med.* 31, 30–36. doi: 10.3343/kjlm.2011.31.1.30
- Ko, K. S., Lee, J.-Y., Suh, J. Y., Oh, W. S., Peck, K. R., Lee, N. Y., et al. (2005). Distribution of major genotypes among methicillin-resistant *Staphylococcus aureus* clones in Asian countries. *J. Clin. Microbiol.* 43, 421–426. doi: 10.1128/JCM.43.1.421-426.2005
- Köck, R., Brakensiek, L., Mellmann, A., Kipp, F., Henderikx, M., Harmsen, D., et al. (2009). Cross-border comparison of the admission prevalence and clonal structure of methicillin-resistant *Staphylococcus aureus*. *J. Hosp. Infect.* 71, 320–326. doi: 10.1016/j.jhin.2008.12.001
- Kong, H., Yu, F., Zhang, W., Li, X., and Wang, H. (2017). Molecular epidemiology and antibiotic resistance profiles of methicillin-resistant *Staphylococcus aureus* strains in a tertiary hospital in China. *Front. Microbiol.* 8:838. doi: 10.3389/fmicb.2017.00838
- Koreen, L., Ramaswamy, S. V., Graviss, E. A., Naidich, S., Musser, J. M., and Kreiswirth, B. N. (2004). spa typing method for discriminating among *Staphylococcus aureus* isolates: implications for use of a single marker to detect genetic micro-and macrovariation. *J. Clin. Microbiol.* 42, 792–799. doi: 10.1128/JCM.42.2.792-799.2004
- Krasuski, A., Michnowska-Swincow, E. and Jarzemowski, T., (2007). Cytidine deamination assay to differentiate *Staphylococcus aureus* from other staphylococci. *Lett. Appl. Microbiol.* 45, 497–503. doi: 10.1111/j.1472-765X.2007.02218.x
- Kriegeskorte, A., Ballhausen, B., Idelevich, E. A., Köck, R., Friedrich, A. W., Karch, H., et al. (2012). Human MRSA isolates with novel genetic homolog. Germany. *Emerging Infect. Dis.* 18:1016. doi: 10.3201/eid1806.110910
- Kuhn, G., Francioli, P., and Blanc, D. (2006). Evidence for clonal evolution among highly polymorphic genes in methicillin-resistant *Staphylococcus aureus*. *J. Bacteriol.* 188, 169–178. doi: 10.1128/JB.188.1.169-178.2006
- Kwak, Y. G., Truong-Bolduc, Q. C., Bin Kim, H., Song, K.-H., Kim, E. S., and Hooper, D. C. (2013). Association of norB overexpression and fluoroquinolone resistance in clinical isolates of *Staphylococcus aureus* from Korea. *J. Antimicrobial Chemother.* 68, 2766–2772. doi: 10.1093/jac/dkt286
- Lamand, V., Dauwalder, O., Tristan, A., Casalegno, J., Meugnier, H., Bes, M., et al. (2012). Epidemiological data of staphylococcal scalded skin syndrome in France from 1997 to 2007 and microbiological characteristics of *Staphylococcus aureus* associated strains. *Clin. Microbiol. Infect.* 18, 514–521. doi: 10.1111/j.1469-0691.12053
- Lamaro-Cardoso, J., de Lencastre, H., Kipnis, A., Pimenta, F. C., Oliveira, L. S., Oliveira, R. M., et al. (2009). Molecular epidemiology and risk factors for nasal carriage of *Staphylococcus aureus* and methicillin-resistant *S. aureus* in infants attending day care centers in Brazil. *J. Clin. Microbiol.* 47, 3991–3997. doi: 10.1128/JCM.01322-09
- Larsen, A. R., Goering, R., Stegger, M., Lindsay, J. A., Gould, K. A., Hinds, J., et al. (2009). Two distinct clones of methicillin-resistant *Staphylococcus aureus* (MRSA) with the same USA300 pulsed-field gel electrophoresis profile: a potential pitfall for identification of USA300 community-associated MRSA. *J. Clin. Microbiol.* 47, 3765–3768. doi: 10.1128/JCM.00934-09
- Larsen, A., Böcher, S., Stegger, M., Goering, R., Pallesen, L., and Skov, R. (2008). Epidemiology of European community-associated methicillin-resistant *Staphylococcus aureus* clonal complex 80 type IV strains isolated in Denmark from 1993 to 2004. *J. Clin. Microbiol.* 46, 62–68. doi: 10.1128/JCM.01381-07
- Laurent, C., Bogaerts, P., Schoevaerdts, D., Denis, O., Deplano, A., Swine, C., et al. (2010). Evaluation of the Xpert MRSA assay for rapid detection of methicillin-resistant *Staphylococcus aureus* from nares swabs of geriatric hospitalized patients and failure to detect a specific SCCmec type IV variant. *Eur. J. Clin. Microbiol. Infect. Dis.* 29, 995–1002. doi: 10.1007/s10096-010-0958-3
- Lee, H., Lim, H., Bae, I. K., Yong, D., Jeong, S. H., Lee, K., et al. (2013). Coexistence of mupirocin and antiseptic resistance in methicillin-resistant *Staphylococcus aureus* isolates from Korea. *Diagn. Microbiol. Infect. Dis.* 75, 308–312. doi: 10.1016/j.diagmicrobio.2012.11.025
- Lee, S. S., Kim, Y. J., Chung, D. R., Jung, K.-S., and Kim, J.-S. (2010). Invasive infection caused by a community-associated methicillin-resistant

- Staphylococcus aureus* strain not carrying Panton-Valentine leukocidin in South Korea. *J. Clin. Microbiol.* 48, 311–313. doi: 10.1128/JCM.00297-09
- Li, J., Wang, L., Ip, M., Sun, M., Sun, J., Huang, G., et al. (2013). Molecular and clinical characteristics of clonal complex 59 methicillin-resistant *Staphylococcus aureus* infections in Mainland China. *PLoS ONE* 8:e70602. doi: 10.1371/journal.pone.0070602
- Lim, K.-T., Hanifah, Y. A., Yusof, M. Y. M., and Thong, K.-L. (2012). Characterisation of the virulence factors and genetic types of methicillin susceptible *Staphylococcus aureus* from patients and healthy individuals. *Indian J. Microbiol.* 52, 593–600. doi: 10.1007/s12088-012-0286-7
- Limbago, B. M., Kallen, A. J., Zhu, W., Eggers, P., McDougal, L. K., and Albrecht, V. S. (2014). Report of the 13th vancomycin-resistant *Staphylococcus aureus* isolate from the United States. *J. Clin. Microbiol.* 52, 998–1002. doi: 10.1128/JCM.02187-13
- Lindqvist, M., Isaksson, B., Grub, C., Jonassen, T., and Häggman, A. (2012). Detection and characterisation of SCCmec remnants in multiresistant methicillin-susceptible *Staphylococcus aureus* causing a clonal outbreak in a Swedish county. *Eur. J. Clin. Microbiol. Infect. Dis.* 31, 141–147. doi: 10.1007/s10096-011-1286-y
- Lindqvist, M., Isaksson, B., Samuelsson, A., Nilsson, L. E., and Häggman, A. (2009). A clonal outbreak of methicillin-susceptible *Staphylococcus aureus* with concomitant resistance to erythromycin, clindamycin and tobramycin in a Swedish county. *Scand. J. Infect. Dis.* 41, 324–333. doi: 10.1080/00365540902801202
- Lindqvist, M., Isaksson, B., Swanberg, J., Skov, R., Larsen, A., Larsen, J., et al. (2015). Long-term persistence of a multi-resistant methicillin-susceptible *Staphylococcus aureus* (MR-MSSA) clone at a university hospital in southeast Sweden, without further transmission within the region. *Eur. J. Clin. Microbiol. Infect. Dis.* 34, 1415–1422. doi: 10.1007/s10096-015-2366-1
- Liu, Q.-Z., Wu, Q., Zhang, Y.-B., Liu, M.-N., Hu, F.-P., Xu, X.-G., et al. (2010). Prevalence of clinical methicillin-resistant *Staphylococcus aureus* (MRSA) with high-level mupirocin resistance in Shanghai and Wenzhou, China. *Int. J. Antimicrob. Agents* 35, 114–118. doi: 10.1016/j.ijantimicag.2009.09.018
- Liu, Y., Wang, H., Du, N., Shen, E., Chen, H., Niu, J., et al. (2009). Molecular evidence for spread of two major methicillin-resistant *Staphylococcus aureus* clones with a unique geographic distribution in Chinese hospitals. *Antimicrobial Agents Chemother.* 53, 512–518. doi: 10.1128/AAC.00804-08
- Longtin, J., Seah, C., Siebert, K., McGeer, A., Simor, A., Longtin, Y., et al. (2011). Distribution of antiseptic resistance genes qacA, qacB, and smr in methicillin-resistant *Staphylococcus aureus* isolated in Toronto, Canada, from 2005 to 2009. *Antimicrobial Agents Chemother.* 55, 2999–3001. doi: 10.1128/AAC.01707-10
- Lozano, C., Porres-Osante, N., Crettaz, J., Rojo-Bezares, B., Benito, D., Olarte, I., et al. (2013). Changes in genetic lineages, resistance, and virulence in clinical methicillin-resistant *Staphylococcus aureus* in a Spanish hospital. *J. Infect. Chemother.* 19, 233–242. doi: 10.1007/s10156-012-0486-4
- Luxner, J., Zarfel, G., Johler, S., Feierl, G., Leitner, E., Hoenigl, M., et al. (2014). Genetic characterization of *Staphylococcus aureus* isolates causing bloodstream infections in Austria. *Diagn. Microbiol. Infect. Dis.* 78, 153–156. doi: 10.1016/j.diagmicrobio.2013.10.010
- Machuca, M. A., Sosa, L. M., and González, C. I. (2013). Molecular typing and virulence characteristic of methicillin-resistant *Staphylococcus aureus* isolates from pediatric patients in Bucaramanga, Colombia. *PLoS ONE* 8:e73434. doi: 10.1371/journal.pone.0073434
- Maeda, T., Saga, T., Miyazaki, T., Kouyama, Y., Harada, S., Iwata, M., et al. (2012). Genotyping of skin and soft tissue infection (SSTI)-associated methicillin-resistant *Staphylococcus aureus* (MRSA) strains among outpatients in a teaching hospital in Japan: application of a phage-open reading frame typing (POT) kit. *J. Infect. Chemother.* 18, 906–914. doi: 10.1007/s10156-012-0506-4
- Marimón, J. M., Villar, M., García-Arenzana, J. M., Caba de la, L., and Pérez-Trallero, E. (2012). Molecular characterization of *Staphylococcus aureus* carrying the panton-valentine leucocidin genes in northern Spain. *J. Infect.* 64, 47–53. doi: 10.1016/j.jinf.2011.10.010
- Matussek, A., Taipalensuu, J., Einemo, M., Tiefenthal, M., and Löfgren, S. (2007). Transmission of *Staphylococcus aureus* from maternity unit staff members to newborns disclosed through spa typing. *Am. J. Infect. Control* 35, 122–125. doi: 10.1016/j.ajic.2006.08.009
- Mazi, W., Sangal, V., Sandstrom, G., Saeed, A., and Yu, J. (2015). Evaluation of spa-typing of methicillin-resistant *Staphylococcus aureus* using high-resolution melting analysis. *Int. J. Infect. Dis.* 38, 125–128. doi: 10.1016/j.ijid.2015.05.002
- Medina, G., Egea, A., Otth, C., Otth, L., Fernández, H., Bocco, J., et al. (2013). Molecular epidemiology of hospital-onset methicillin-resistant *Staphylococcus aureus* infections in Southern Chile. *Eur. J. Clin. Microbiol. Infect. Dis.* 32, 1533–1540. doi: 10.1007/s10096-013-1907-8
- Melin, S., Melin, S., Häggman, S., Melin, S., Häggman, S., Olsson-Liljequist, B., et al. (2009). Epidemiological typing of methicillin-resistant *Staphylococcus aureus* (MRSA): spa typing versus pulsed-field gel electrophoresis. *Scand. J. Infect. Dis.* 41, 433–439. doi: 10.1080/00365540902962749
- Mellmann, A., Friedrich, A. W., Rosenkötter, N., Rothgänger, J., Karch, H., Reintjes, R., et al. (2006). Automated DNA sequence-based early warning system for the detection of methicillin-resistant *Staphylococcus aureus* outbreaks. *PLoS Med.* 3:e33. doi: 10.1371/journal.pmed.0030033
- Mellmann, A., Weniger, T., Berssenbrügge, C., Keckevoet, U., Friedrich, A. W., Harmsen, D., et al. (2008). Characterization of clonal relatedness among the natural population of *Staphylococcus aureus* strains by using spa sequence typing and the BURP (based upon repeat patterns) algorithm. *J. Clin. Microbiol.* 46, 2805–2808. doi: 10.1128/JCM.00071-08
- Miko, B. A., Hafer, C. A., Lee, C. J., Sullivan, S. B., Hackel, M. A., Johnson, B. M., et al. (2013). Molecular characterization of methicillin-susceptible *Staphylococcus aureus* clinical isolates in the United States, 2004 to 2010. *J. Clin. Microbiol.* 51, 874–879. doi: 10.1128/JCM.00923-12
- Miller, C. E., Batra, R., Cooper, B. S., Patel, A. K., Klein, J., Otter, J. A., et al. (2011). An association between bacterial genotype combined with a high-vancomycin minimum inhibitory concentration and risk of endocarditis in methicillin-resistant *Staphylococcus aureus* bloodstream infection. *Clin. Infect. Dis.* 54, 591–600. doi: 10.1093/cid/cir858
- Mirzaii, M., Emameini, M., Jabalameli, F., Halimi, S., and Taherikalani, M. (2015). Molecular investigation of *Staphylococcus aureus* isolated from the patients, personnel, air and environment of an ICU in a hospital in Tehran. *J. Infect. Public Health* 8, 202–206. doi: 10.1016/j.jiph.2014.09.002
- Mohammadi, S., Sekawi, Z., Monjezi, A., Maleki, M.-H., Soroush, S., Sadeghfard, N., et al. (2014). Emergence of SCCmec type III with variable antimicrobial resistance profiles and spa types among methicillin-resistant *Staphylococcus aureus* isolated from healthcare-and community-acquired infections in the west of Iran. *Int. J. Infect. Dis.* 25, 152–158. doi: 10.1016/j.ijid.2014.02.018
- Monaco, M., Sanchini, A., Grundmann, H., and Pantosti, A. (2010). Vancomycin-heteroresistant phenotype in invasive methicillin-resistant *Staphylococcus aureus* isolates belonging to spa type 041. *Eur. J. Clin. Microbiol. Infect. Dis.* 29, 771–777. doi: 10.1007/s10096-010-0922-2
- Montesinos, I., Delgado, T., Riverol, D., Salido, E., Miguel, M., Jimenez, A., et al. (2006). Changes in the epidemiology of methicillin-resistant *Staphylococcus aureus* associated with the emergence of EMRSA-16 at a university hospital. *J. Hosp. Infect.* 64, 257–263. doi: 10.1016/j.jhin.2006.07.004
- Moodley, A., Oosthuysen, W., Duse, A., Marais, E., and Group, S. A. M. S. (2010). Molecular characterization of clinical methicillin-resistant *Staphylococcus aureus* isolates in South Africa. *J. Clin. Microbiol.* 48, 4608–4611. doi: 10.1128/JCM.01704-10
- Múnera, J. M. V., Ríos, A. M. O., Urrego, D. M., and Quiceno, J. N. J. (2017). In vitro susceptibility of methicillin-resistant *Staphylococcus aureus* isolates from skin and soft tissue infections to vancomycin, daptomycin, linezolid, and tedizolid. *Braz. J. Infect. Dis.* 21, 493–499. doi: 10.1016/j.bjid.2017.03.010
- Murchan, S., Kaufmann, M. E., Deplano, A., de Ryck, R., Struelens, M., Zinn, C. E., et al. (2003). Harmonization of pulsed-field gel electrophoresis protocols for epidemiological typing of strains of methicillin-resistant *Staphylococcus aureus*: a single approach developed by consensus in 10 European laboratories and its application for tracing the spread of related strains. *J. Clin. Microbiol.* 41, 1574–1585. doi: 10.1128/JCM.41.4.1574-1585.2003
- Murphy, C. R., Hudson, L. O., Spratt, B. G., Elkins, K., Terpstra, L., Gombossev, A., et al. (2013). Predictors of hospitals with endemic community-associated methicillin-resistant *Staphylococcus aureus*. *Infect. Control Hosp. Epidemiol.* 34, 581–587. doi: 10.1086/670631
- Nadig, S., Raju, S. R., and Arakere, G. (2010). Epidemic methicillin-resistant *Staphylococcus aureus* (EMRSA-15) variants detected in healthy and diseased individuals in India. *J. Med. Microbiol.* 59, 815–821. doi: 10.1099/jmm.0.017632-0

- Nadig, S., Velusamy, N., Lalitha, P., Kar, S., Sharma, S., and Arakere, G. (2012). *Staphylococcus aureus* eye infections in two Indian hospitals: emergence of ST772 as a major clone. *Clin. Ophthalmol.* 6:165. doi: 10.2147/OPTH.S23878
- Ngoa, U. A., Schaumburg, F., Adegnika, A. A., Kösters, K., Möller, T., Fernandes, J. F., et al. (2012). Epidemiology and population structure of *Staphylococcus aureus* in various population groups from a rural and semi urban area in Gabon, Central Africa. *Acta Trop.* 124, 42–47. doi: 10.1016/j.actatropica.2012.06.005
- Nulens, E., Stobberingh, E. E., van Dessel, H., Sebastian, S., van Tiel, F. H., Beisser, P. S., et al. (2008). Molecular characterization of *Staphylococcus aureus* bloodstream isolates collected in a Dutch University Hospital between 1999 and 2006. *J. Clin. Microbiol.* 46, 2438–2441. doi: 10.1128/JCM.00808-08
- O'Hara, F. P., Suaya, J. A., Ray, G. T., Baxter, R., Brown, M. L., Mera, R. M., et al. (2016). spa Typing and multilocus sequence typing show comparable performance in a macroepidemiologic study of *Staphylococcus aureus* in the United States. *Microbial. Drug Resist.* 22, 88–96. doi: 10.1089/mdr.2014.0238
- O'Malley, S., Emele, F., Nwaokorie, F., Idika, N., Umeizudike, A., Emeka-Nwabunnia, I., et al. (2015). Molecular typing of antibiotic-resistant *Staphylococcus aureus* in Nigeria. *J. Infect. Public Health* 8, 187–193. doi: 10.1016/j.jiph.2014.08.001
- Omuse, G., Zyl, K. N., Hoek, K., Abdulgader, S., Kariuki, S., Whitelaw, A., et al. (2016). Molecular characterization of *Staphylococcus aureus* isolates from various healthcare institutions in Nairobi, Kenya: a cross sectional study. *Ann. Clin. Microbiol. Antimicrob.* 15:51. doi: 10.1186/s12941-016-0171-z
- O'Sullivan, M. V., Kong, F., Sintchenko, V., and Gilbert, G. L. (2010). Rapid identification of methicillin-resistant *Staphylococcus aureus* transmission in hospitals by use of phage-derived open reading frame typing enhanced by multiplex PCR and reverse line blot assay. *J. Clin. Microbiol.* 48, 2741–2748. doi: 10.1128/JCM.02201-09
- Otokunefor, K., Sloan, T., Kearns, A. M., and James, R. (2012). Molecular characterization and Panton-Valentine leucocidin typing of community-acquired methicillin-sensitive *Staphylococcus aureus* clinical isolates. *J. Clin. Microbiol.* 50, 3069–3072. doi: 10.1128/JCM.00602-12
- Otter, J., Klein, J., Watts, T., Kearns, A., and French, G. (2007). Identification and control of an outbreak of ciprofloxacin-susceptible EMRSA-15 on a neonatal unit. *J. Hosp. Infect.* 67, 232–239. doi: 10.1016/j.jhin.2007.07.024
- Parhizgari, N., Khoramroozi, S. S., Hosseini, M., Asghar, S. A., Marashifard, M., Yazdanpanah, M., et al. (2016). High frequency of multidrug-resistant *Staphylococcus aureus* with SCCmec type III and Spa types t037 and t631 isolated from burn patients in southwest of Iran. *APMIS* 124, 221–228. doi: 10.1111/apm.12493
- Peck, K. R., Baek, J. Y., Song, H.-J., and Ko, K. S. (2009). Comparison of genotypes and enterotoxin genes between *Staphylococcus aureus* isolates from blood and nasal colonizers in a Korean hospital. *J. Korean Med. Sci.* 24, 585–591. doi: 10.3346/jkms.2009.24.4.585
- Pérez-Vázquez, M., Vindel, A., Marcos, C., Oteo, J., Cuevas, O., Trincado, P., et al. (2008). Spread of invasive Spanish *Staphylococcus aureus* spa-type t067 associated with a high prevalence of the aminoglycoside-modifying enzyme gene ant (4')-Ia and the efflux pump genes msra/msrb. *J. Antimicrobial. Chemother.* 63, 21–31. doi: 10.1093/jac/dkn430
- Perovic, O., Iyaloo, S., Kularatne, R., Lowman, W., Bosman, N., Wadula, J., et al. (2015). Prevalence and trends of *Staphylococcus aureus* bacteraemia in hospitalized patients in South Africa, 2010 to 2012: laboratory-based surveillance mapping of antimicrobial resistance and molecular epidemiology. *PLoS ONE* 10:e0145429. doi: 10.1371/journal.pone.0145429
- Pettersson, A., Olsson-Liljequist, B., Miörner, H., and Haeggman, S. (2010). Evaluating the usefulness of spa typing, in comparison with pulsed-field gel electrophoresis, for epidemiological typing of methicillin-resistant *Staphylococcus aureus* in a low-prevalence region in Sweden 2000–2004. *Clin. Microbiol. Infect.* 16, 456–462. doi: 10.1111/j.1469-0691.2009.02881.x
- Pfingsten-Würzburg, S., Pieper, D., Bautsch, W., and Probst-Kepper, M. (2011). Prevalence and molecular epidemiology of methicillin-resistant *Staphylococcus aureus* in nursing home residents in northern Germany. *J. Hosp. Infect.* 78, 108–112. doi: 10.1016/j.jhin.2011.02.011
- Pomorska-Wesołowska, M., Chmielarczyk, A., Chlebowicz, M., Ziolkowski, G., Szczypta, A., Natkaniec, J., et al. (2017). Virulence and Antibiotic Resistance of *Staphylococcus aureus* Isolated from Bloodstream Infections and Pneumonia in Southern Poland. *J. Global Antimicrobial Resist.* 11, 100–104. doi: 10.1016/j.jgar.2017.07.009
- Price, J. R., Golubchik, T., Cole, K., Wilson, D. J., Crook, D. W., Thwaites, G. E., et al. (2013). Whole-genome sequencing shows that patient-to-patient transmission rarely accounts for acquisition of *Staphylococcus aureus* in an intensive care unit. *Clin. Infect. Dis.* 58, 609–618. doi: 10.1093/cid/cit807
- Prosperi, M., Veras, N., Azarian, T., Rathore, M., Nolan, D., Rand, K., et al. (2013). Molecular epidemiology of community-associated methicillin-resistant *Staphylococcus aureus* in the genomic era: a cross-sectional study. *Sci. Reports* 3. doi: 10.1038/srep01902
- Rajan, V., Schoenfelder, S. M., Ziebuhr, W., and Gopal, S. (2015). Genotyping of community-associated methicillin resistant *Staphylococcus aureus* (CA-MRSA) in a tertiary care centre in Mysore, South India: ST2371-SCCmec IV emerges as the major clone. *Infect. Genet. Evol.* 34, 230–235. doi: 10.1016/j.meegid.2015.05.032
- Rasschaert, G., Vanderhaegen, W., Dewaele, I., N., Janež, Huijsdens, X., Butaye, P., et al. (2009). Comparison of fingerprinting methods for typing methicillin-resistant *Staphylococcus aureus* sequence type 398. *J. Clin. Microbiol.* 47, 3313–3322. doi: 10.1128/JCM.00910-09
- Raulin, O., Durand, G., Gillet, Y., Bes, M., Lina, G., Vandenesch, F., et al. (2010). Toxin profiling of *Staphylococcus aureus* strains involved in varicella superinfection. *J. Clin. Microbiol.* 48, 1696–1700. doi: 10.1128/JCM.02018-09
- Rijnders, M., Deurenberg, R., Boumans, M., Hoogkamp-Korstanje, J., Beisser, P., Stobberingh, E., et al. (2009). Population structure of *Staphylococcus aureus* strains isolated from intensive care unit patients in the Netherlands over an 11-year period (1996 to 2006). *J. Clin. Microbiol.* 47, 4090–4095. doi: 10.1128/JCM.00820-09
- Rodríguez, E. A., Correa, M. M., Ospina, S., Atehortúa, S. L., and Jiménez, J. N. (2014). Differences in epidemiological and molecular characteristics of nasal colonization with *Staphylococcus aureus* (MSSA-MRSA) in children from a university hospital and day care centers. *PLoS ONE* 9:e101417. doi: 10.1371/journal.pone.0101417
- Ruffing, U., Akulenko, R., Bischoff, M., Helms, V., Herrmann, M., and von Müller, L. (2012). Matched-cohort DNA microarray diversity analysis of methicillin sensitive and methicillin resistant *Staphylococcus aureus* isolates from hospital admission patients. *PLoS ONE* 7:e52487. doi: 10.1371/journal.pone.0052487
- Ruimy, R., Angebault, C., Djossou, F., Dupont, C., Epelboin, L., Jarraud, S., et al. (2010). Are host genetics the predominant determinant of persistent nasal *Staphylococcus aureus* carriage in humans? *J. Infect. Dis.* 202, 924–934. doi: 10.1086/655901
- Ruppitsch, W., Indra, A., Stöger, A., Mayer, B., Stadlbauer, S., Wewalka, G., et al. (2006). Classifying spa types in complexes improves interpretation of typing results for methicillin-resistant *Staphylococcus aureus*. *J. Clin. Microbiol.* 44, 2442–2448. doi: 10.1128/JCM.00113-06
- Ruppitsch, W., Stöger, A., Braun, O., Strommenger, B., Nübel, U., Wewalka, G., et al. (2007). Meticillin-resistant *Staphylococcus aureus*: occurrence of a new spa type in two acute care hospitals in Austria. *J. Hosp. Infect.* 67, 316–322. doi: 10.1016/j.jhin.2007.09.011
- Sabat, A., Malachowa, N., Miedzobrodzki, J., and Hryniewicz, W. (2006). Comparison of PCR-based methods for typing *Staphylococcus aureus* isolates. *J. Clin. Microbiol.* 44, 3804–3807. doi: 10.1128/JCM.00395-06
- Sabri, I., Adwan, K., Essawi, T. A., and Farraj, M. A. (2013). Molecular characterization of methicillin-resistant *Staphylococcus aureus* isolates in three different Arab world countries. *Eur. J. Microbiol. Immunol.* 3, 183–187. doi: 10.1556/EJM.3.2013.3.5
- Sanchini, A., Campanile, F., Monaco, M., Cafiso, V., Rasigade, P.-J., Pantosti, A., et al. (2011). DNA microarray-based characterisation of Panton-Valentine leukocidin-positive community-acquired methicillin-resistant *Staphylococcus aureus* from Italy. *Eur. J. Clin. Microbiol. Infect. Dis.* 30, 1399–1408. doi: 10.1007/s10096-011-1234-x
- Sanchini, A., Del Grosso, M., Villa, L., Ammendolia, M., Superti, F., Monaco, M., et al. (2014). Typing of Panton-Valentine leukocidin-encoding phages carried by methicillin-susceptible and methicillin-resistant *Staphylococcus aureus* from Italy. *Clin. Microbiol. Infect.* 20, 0840–0846. doi: 10.1111/1469-0691.12679
- Sangal, V., Girvan, E. K., Jadhav, S., Lawes, T., Robb, A., Vali, L., et al. (2012). Impacts of a long-term programme of active surveillance and chlorhexidine baths on the clinical and molecular epidemiology of methicillin-resistant *Staphylococcus aureus* (MRSA) in an Intensive Care Unit in Scotland. *Int. J. Antimicrob. Agents* 40, 323–331. doi: 10.1016/j.ijantimicag.2012.06.007

- Sangvik, M., Olsen, R. S., Olsen, K., Simonsen, G. S., Furberg, A.-S., and Ericson Sollid, J. U. (2011). Age-and gender-associated *Staphylococcus aureus* spa types found among nasal carriers in a general population: the Tromsø Staph and Skin Study. *J. Clin. Microbiol.* 49, 4213–4218. doi: 10.1128/JCM.05290-11
- Santos-Júnior, C. D., Veríssimo, A., and Costa, J. (2016). The recombination dynamics of *Staphylococcus aureus* inferred from spa gene. *BMC Microbiol.* 16:143. doi: 10.1186/s12866-016-0757-9
- Sasso, E., Silverman, G., and Mannik, M. (1991). Human IgA and IgG F (ab') 2 that bind to staphylococcal protein A belong to the VHIII subgroup. *J. Immunol.* 147, 1877–1883.
- Schmid, D., Simons, E., Ruppitsch, W., Hrvniaková, L., Stoeger, A., Wechsler-Fördös, A., et al. (2013). Limited value of routine spa typing: a cross-sectional study of methicillin-resistant *Staphylococcus aureus* positive patients in an Austrian hospital. *Am. J. Infect. Control* 41, 617–624. doi: 10.1016/j.ajic.2012.09.013
- Seidl, K., Leimer, N., Marques, M. P., Furrer, A., Holzmann-Bürgel, A., Senn, G., et al. (2015). Clonality and antimicrobial susceptibility of methicillin-resistant *Staphylococcus aureus* at the University Hospital Zurich, Switzerland between 2012 and 2014. *Ann. Clin. Microbiol. Antimicrob.* 14:14. doi: 10.1186/s12941-015-0075-3
- Shakeri, F., and Ghaemi, E. A. (2014). New Spa types among MRSA and MSSA isolates in north of Iran. *Adv. Microbiol.* 4:899. doi: 10.4236/aim.2014.413100
- Shambat, S., Nadig, S., Prabhakara, S., Bes, M., Etienne, J., and Arakere, G. (2012). Clonal complexes and virulence factors of *Staphylococcus aureus* from several cities in India. *BMC Microbiol.* 12:64. doi: 10.1186/1471-2180-12-64
- Shet, A., Mathema, B., Mediavilla, J. R., Kishii, K., Mehandru, S., Jeanne-Pierre, P., et al. (2009). Colonization and Subsequent Skin and Soft Tissue Infection Due to Methicillin-Resistant *Staphylococcus aureus* in a Cohort of Otherwise Healthy Adults Infected with HIV Type 1. *J. Infect. Dis.* 200, 88–93. doi: 10.1086/599315
- Shittu, A. O., Okon, K., Adesida, S., Oyedara, O., Witte, W., Strommenger, B., et al. (2011). Antibiotic resistance and molecular epidemiology of *Staphylococcus aureus* in Nigeria. *BMC Microbiol.* 11:92. doi: 10.1186/1471-2180-11-92
- Shittu, A. O., Oyedara, O., Okon, K., Raji, A., Peters, G., von Müller, L., et al. (2015). An assessment on DNA microarray and sequence-based methods for the characterization of methicillin-susceptible *Staphylococcus aureus* from Nigeria. *Front. Microbiol.* 6:1160. doi: 10.3389/fmicb.2015.01160
- Shopisin, B., Gomez, M., Montgomery, S., Smith, D., Waddington, M., Dodge, D., et al. (1999). Evaluation of protein A gene polymorphic region DNA sequencing for typing of *Staphylococcus aureus* strains. *J. Clin. Microbiol.* 37, 3556–3563.
- Shore, A. C., Brennan, O. M., Deasy, E. C., Rossney, A. S., Kinnevey, P. M., Ehrlicht, R., et al. (2012). DNA Microarray Profiling of a Diverse Collection of Nosocomial Methicillin-Resistant *Staphylococcus aureus* Assigns the Majority to the Correct Sequence Type and Staphylococcal Chromosome Cassette mec (SCCmec) Type and Resulted in the Subsequent Identification and Characterization of Novel SCCmec-SCCM1Composite Islands. *Antimicrobial Agents Chemother.* 62, 1247–1212. doi: 10.1128/AAC.01247-12
- Shore, A. C., Rossney, A. S., Kinnevey, P. M., Brennan, O. M., Creamer, E., Sherlock, O., et al. (2010). Enhanced discrimination of highly clonal ST22-methicillin-resistant *Staphylococcus aureus* IV isolates achieved by combining spa, dru, and pulsed-field gel electrophoresis typing data. *J. Clin. Microbiol.* 48, 1839–1852. doi: 10.1128/JCM.02155-09
- Shore, A. C., Tecklenborg, S. C., Brennan, G. I., Ehrlicht, R., Monecke, S., and Coleman, D. C. (2014). Panton-Valentine leukocidin-positive *Staphylococcus aureus* in Ireland from 2002 to 2011: 21 clones, frequent importation of clones, temporal shifts of predominant methicillin-resistant *S. aureus* clones, and increasing multiresistance. *J. Clin. Microbiol.* 52, 859–870. doi: 10.1128/JCM.02799-13
- Siegel, J. D., Rhinehart, E., Jackson, M., and Chiarello, L. (2007). Management of multidrug-resistant organisms in health care settings, 2006. *Am. J. Infect. Control* 35, S165–S193. doi: 10.1016/j.ajic.2007.10.006
- Skråmm, I., Moen, A. E. F., and Bukholm, G. (2011). Nasal carriage of *Staphylococcus aureus*: frequency and molecular diversity in a randomly sampled Norwegian community population. *APMIS* 119, 522–528. doi: 10.1111/j.1600-0463.2011.02758.x
- Sobral, D., Schwarz, S., Bergonier, D., Brisabois, A., Feßler, A. T., Gilbert, F. B., et al. (2012). High throughput multiple locus variable number of tandem repeat analysis (MLVA) of *Staphylococcus aureus* from human, animal and food sources. *PLoS ONE* 7:e33967. doi: 10.1371/journal.pone.0033967
- Soliman, R. S., Phillips, G., Whitty, P., and Edwards, D. H. (2009). Distribution of methicillin-resistant *Staphylococcus aureus* spa types isolated from health-care workers and patients in a Scottish university teaching hospital. *J. Med. Microbiol.* 58, 1190–1195. doi: 10.1099/jmm.0.010132-0
- Song, Y., Du, X., Li, T., Zhu, Y., and Li, M. (2013). Phenotypic and molecular characterization of *Staphylococcus aureus* recovered from different clinical specimens of inpatients at a teaching hospital in Shanghai between 2005 and 2010. *J. Med. Microbiol.* 62, 274–282. doi: 10.1099/jmm.0.050971-0
- Strommenger, B., Braulke, C., Heuck, D., Schmidt, C., Pasemann, B., Nübel, U., et al. (2008). spa typing of *Staphylococcus aureus* as a frontline tool in epidemiological typing. *J. Clin. Microbiol.* 46, 574–581. doi: 10.1128/JCM.01599-07
- Sun, D. D., Ma, X. X., Hu, J., Tian, Y., Pang, L., Shang, H., et al. (2013). Epidemiological and molecular characterization of community and hospital acquired *Staphylococcus aureus* strains prevailing in Shenyang, Northeastern China. *Braz. J. Infect. Dis.* 17, 682–690. doi: 10.1016/j.bjid.2013.02.007
- Sun, W., Chen, H., Liu, Y., Zhao, C., Nichols, W. W., Chen, M., et al. (2009). Prevalence and characterization of heterogeneous vancomycin-intermediate *Staphylococcus aureus* isolates from 14 cities in China. *Antimicrobial Agents Chemother.* 53, 3642–3649. doi: 10.1128/AAC.00206-09
- Tavares, A., Faria, N., De Lencastre, H., and Miragaia, M. (2014). Population structure of methicillin-susceptible *Staphylococcus aureus* (MSSA) in Portugal over a 19-year period (1992–2011). *Eur. J. Clin. Microbiol. Infect. Dis.* 33, 423–432. doi: 10.1007/s10096-013-1972-z
- Tian, S. F., Chu, Y. Z., Nian, H., Li, F. S., Sun, J. M., Wang, Y. L., et al. (2013). Genotype diversity of methicillin-resistant *Staphylococcus aureus* in Shenyang, China. *Scand. J. Infect. Dis.* 45, 915–921. doi: 10.3109/00365548.2013.830330
- Tristan, A., Bes, M., Meugnier, H., Lina, G., Bozdogan, B., Courvalin, P., et al. (2007). Global distribution of Panton-Valentine leukocidin-positive methicillin-resistant *Staphylococcus aureus*, 2006. *Emerging Infect. Dis.* 13:594. doi: 10.3201/eid1304.061316
- Turlej, A., Hryniwicz, W., and Empel, J. (2011). Staphylococcal cassette chromosome mec (SCCmec) classification and typing methods: an overview. *Pol. J. Microbiol.* 60, 95–103.
- Udo, E. E., Al-Lawati, H.-B., Al-Muharmi, Z., and Thukral, S. (2014). Genotyping of methicillin-resistant *Staphylococcus aureus* in the Sultan Qaboos University Hospital, Oman reveals the dominance of Panton-Valentine leucocidin-negative ST6-IV/t304 clone. *New Microb. New Infect.* 2, 100–105. doi: 10.1002/nmi.247
- Udo, E. E., Boswihi, S. S., and Al-Sweih, N. (2016). High prevalence of toxic shock syndrome toxin-producing epidemic methicillin-resistant *Staphylococcus aureus* 15 (EMRSA-15) strains in Kuwait hospitals. *New Microb. New Infect.* 12, 24–30. doi: 10.1016/j.nmni.2016.03.008
- Ugolotti, E., Bandettini, R., Marchese, A., Gualco, L., Vanni, I., Borzi, L., et al. (2011). Molecular characterization of hospital-acquired methicillin-resistant *Staphylococcus aureus* strains in pediatric outbreaks using variable tandem repeat analysis with spa and ClfB typing. *Diagn. Microbiol. Infect. Dis.* 69, 213–217. doi: 10.1016/j.diagmicrobio.2010.08.015
- Uzunović, S., Bedenić, B., Budimir, A., Kamberović, F., Ibrahimagić, A., Delić-Bikić, S., et al. (2014). Emergency (clonal spread) of methicillin-resistant *Staphylococcus aureus* (MRSA), extended spectrum (ESBL)-and AmpC beta-lactamase-producing Gram-negative bacteria infections at Pediatric Department, Bosnia and Herzegovina. *Wien. Klin. Wochenschr.* 126, 747–756. doi: 10.1007/s00508-014-0597-2
- Uzunović-Kamberović, S., Rijnders, M. I., Stobberingh, E. E., Ibrahimagić, A., Kamberović, F., and Ille, T. (2013). Molecular characterisation of methicillin-susceptible and methicillin-resistant *Staphylococcus aureus* in inpatients and outpatients in Bosnia and Herzegovina. *Wiener Medizinische Wochenschrift* 163, 13–20. doi: 10.1007/s10354-012-0142-8
- Vainio, A., Kardén-Lilja, M., Ibrahem, S., Kerttula, A., Salmenlinna, S., Virolainen, A., et al. (2008). Clonality of epidemic methicillin-resistant *Staphylococcus aureus* strains in Finland as defined by several molecular methods. *Eur. J. Clin. Microbiol. Infect. Dis.* 27, 545–555. doi: 10.1007/s10096-008-0470-1
- Vainio, A., Koskela, S., Virolainen, A., Vuopio, J., and Salmenlinna, S. (2011). Adapting spa typing for national laboratory-based surveillance of

- methicillin-resistant *Staphylococcus aureus*. *Eur. J. Clin. Microbiol. Infect. Dis.* 30, 789–797. doi: 10.1007/s10096-011-1158-5
- Valaperta, R., Tejada, M. R., Frigerio, M., Moroni, A., Ciulla, E., Cioffi, S., et al. (2010). *Staphylococcus aureus* nosocomial infections: the role of a rapid and low-cost characterization for the establishment of a surveillance system. *New Microbiol.* 33, 223–232.
- Valentin-Domeier, A.-S., Girard, M., Bertrand, X., Violette, J., François, P., Van Der Mee-Marquet, P.-N. et al. (2011). Methicillin-susceptible ST398 *Staphylococcus aureus* responsible for bloodstream infections: an emerging human-adapted subclone? *PLoS ONE* 6:e28369. doi: 10.1371/journal.pone.0028369
- van der Donk, C., Rijnders, M., Donker, G., de Neeling, A., Nys, S., and Stobberingh, E. (2013a). Is living in a border region a risk for a high prevalence of resistance? *Eur. J. Clin. Microbiol. Infect. Dis.* 32, 989–995. doi: 10.1007/s10096-013-1835-7
- van der Donk, C., Schols, J., Schneiders, V., Grimm, H.-K., and Stobberingh, E. (2013b). Antibiotic resistance, population structure and spread of *Staphylococcus aureus* in nursing homes in the Euregion Meuse-Rhine. *Eur. J. Clin. Microbiol. Infect. Dis.* 32, 1483–1489. doi: 10.1007/s10096-013-1901-1
- Van Loo, I., Huijsdens, X., Tiemersma, E., De Neeling, A., van de Sande-Bruinsma, N., Beaujean, D., et al. (2007). Emergence of methicillin-resistant *Staphylococcus aureus* of animal origin in humans. *Emerging Infect. Dis.* 13:1834. doi: 10.3201/eid1312.070384
- Vandendriessche, S., Kadlec, K., Schwarz, S., and Denis, O. (2011). Methicillin-susceptible *Staphylococcus aureus* ST398-t571 harbouring the macrolide-lincosamide-streptogramin B resistance gene erm (T) in Belgian hospitals. *J. Antimicrobial Chemother.* 66, 2455–2459. doi: 10.1093/jac/dkr348
- Velasco, C., López-Cortés, L., Caballero, F., Lepe, J., de Cueto, M., Molina, J., et al. (2012). Clinical and molecular epidemiology of methicillin-resistant *Staphylococcus aureus* causing bacteraemia in Southern Spain. *J. Hospital Infect.* 81, 257–263. doi: 10.1016/j.jhin.2012.05.007
- Verweij, W. (1940). A type-specific antigenic protein derived from the *Staphylococcus*. *J. Exp. Med.* 71, 635–644. doi: 10.1084/jem.71.5.635
- Vindel, A., Cuevas, O., Cercenado, E., Marcos, C., Bautista, V., Castellares, C., et al. (2009). Methicillin-resistant *Staphylococcus aureus* in Spain: molecular epidemiology and utility of different typing methods. *J. Clin. Microbiol.* 47, 1620–1627. doi: 10.1128/JCM.01579-08
- von Eiff, C., Taylor, K. L., Mellmann, A., Fattom, A. I., Friedrich, A. W., Peters, G., et al. (2007). Distribution of capsular and surface polysaccharide serotypes of *Staphylococcus aureus*. *Diagn. Microbiol. Infect. Dis.* 58, 297–302. doi: 10.1016/j.diagmicrobio.2007.01.016
- Vourli, S., Vagiakou, H., Ganteris, G., Orfanidou, M., Polemis, M., Vatopoulos, A., et al. (2007). High rates of community-acquired, Panton-Valentine leukocidin (PVL)-positive methicillin-resistant *S. aureus* (MRSA) infections in adult outpatients in Greece. *Age* 14:19089 doi: 10.2807/ese.14.02.19089-en
- Wang, J.-L., Wang, J.-T., Chen, S.-Y., Chen, Y.-C., and Chang, S.-C. (2010). Distribution of staphylococcal cassette chromosome mec types and correlation with comorbidity and infection type in patients with MRSA bacteremia. *PLoS ONE* 5:e9489. doi: 10.1371/journal.pone.0009489
- Wang, M., Zheng, Y., Mediavilla, J. R., Chen, L., Kreiswirth, B. N., Song, Y., et al. (2017). Hospital Dissemination of tst-1-Positive Clonal Complex 5 (CC5) Methicillin-Resistant *Staphylococcus aureus*. *Front. Cell. Infect. Microbiol.* 7:101. doi: 10.3389/fcimb.2017.00101
- Wang, W.-Y., Chiueh, T.-S., Sun, J.-R., Tsao, S.-M., and Lu, J.-J. (2012). Molecular typing and phenotype characterization of methicillin-resistant *Staphylococcus aureus* isolates from blood in Taiwan. *PLoS ONE* 7:e30394. doi: 10.1371/journal.pone.0030394
- Werbick, C., Becker, K., Mellmann, A., Juuti, K. M., Von Eiff, C., Peters, G., et al. (2007). Staphylococcal chromosomal cassette mec type I, spa type, and expression of PIs are determinants of reduced cellular invasiveness of methicillin-resistant *Staphylococcus aureus* isolates. *J. Infect. Dis.* 195, 1678–1685. doi: 10.1086/517517
- Williamson, D. A., Bakker, S., Coombs, G. W., Tan, H. L., Monecke, S., and Heffernan, H. (2013). Emergence and molecular characterization of clonal complex 398 (CC398) methicillin-resistant *Staphylococcus aureus* (MRSA) in New Zealand. *J. Antimicrobial Chemother.* 69, 1428–1430. doi: 10.1093/jac/dkt499
- Wiśniewska, K., Piórkowska, A., Kasprzyk, J., Bronk, M., and Świeć, K. (2014). Clonal distribution of bone sialoprotein-binding protein gene among *Staphylococcus aureus* isolates associated with bloodstream infections. *Folia Microbiol. (Praha)* 59, 465–471. doi: 10.1007/s12223-014-0321-7
- Witte, W., Strommenger, B., Cuny, C., Heuck, D., and Nuebel, U. (2007). Methicillin-resistant *Staphylococcus aureus* containing the Panton-Valentine leucocidin gene in Germany in 2005 and 2006. *J. Antimicrobial Chemother.* 60, 1258–1263. doi: 10.1093/jac/dkm384
- Wu, Z., Li, F., Liu, D., Xue, H., and Zhao, X. (2015). Novel type XII staphylococcal cassette chromosome mec harboring a new cassette chromosome recombinase, CcrC2. *Antimicrobial Agents Chemother.* 59, 7597–7601. doi: 10.1128/AAC.01692-15
- Xiao, M., Wang, H., Zhao, Y., Mao, L.-L., Brown, M., Yu, Y. S., et al. (2013). National surveillance of methicillin-resistant *Staphylococcus aureus* in China highlights a still-evolving epidemiology with 15 novel emerging multilocus sequence types. *J. Clin. Microbiol.* 51, 3638–3644. doi: 10.1128/JCM.01375-13
- Yu, F., Liu, Y., Lu, C., Jinnan, L., Qi, X., Ding, Y., et al. (2015). Dissemination of fusidic acid resistance among *Staphylococcus aureus* clinical isolates. *BMC Microbiol.* 15:210. doi: 10.1186/s12866-015-0552-z
- Zhang, K., McClure, J.-A., Elsayed, S., Tan, J., and Conly, J. M. (2008). Coexistence of Panton-Valentine Leukocidin—Positive and—Negative Community-Associated Methicillin-Resistant *Staphylococcus aureus* USA400 Sibling Strains in a Large Canadian Health-Care Region. *J. Infect. Dis.* 197, 195–204. doi: 10.1086/523763
- Zhang, W., Shen, X., Zhang, H., Wang, C., Deng, Q., Liu, L., et al. (2009). Molecular epidemiological analysis of methicillin-resistant *Staphylococcus aureus* isolates from Chinese pediatric patients. *Eur. J. Clin. Microbiol. Infect. Dis.* 28, 861–864. doi: 10.1007/s10096-009-0706-8

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Asadollahi, Farahani, Mirzaii, Khoramrooz, van Belkum, Asadollahi, Dadashi and Darban-Sarokhalil. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.