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*CORRESPONDENCE Gang Liu ⊠ liugang8966@163.com

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Metagenomic analysis of gut microbiota and antibiotic-resistant genes in *Anser erythropus* wintering at Shengjin and Caizi Lakes in China

Gang Liu*, Na Xu and Jiahui Feng

School of Life Sciences, Anhui Medical University, Hefei, China

Migratory birds are the primary source and reservoir of antibiotic-resistant genes (ARGs) related to their gut microbes. In this study, we performed metagenomics analysis to study the gut microbial communities and ARGs of Anser erythropus wintering at Shengjin (SJ) and Caizi (CZ) Lakes. The results showed that bacteria, fungi, viruses, and archaea were the dominant gut microbes. Principal component analysis (PCA) indicated that the microbiota compositions significantly differed between the two populations. Diet may be the most crucial driver of the gut microbial communities for A. erythropus. This species fed exclusively on Poaceae spp. at Shengjin Lake and primarily on Carex spp. at Caizi Lake. Tetracycline, macrolide, fluoroquinolone, phenicol, and peptide antibiotics were the dominant resistant types. ARGs had a significantly higher abundance of operational taxonomic units (OTUs) in the Shengjin Lake samples than in Caizi Lake samples. PCA indicated that most Shengjin Lake samples significantly differed in gut microbiota composition from those obtained at Caizi Lake. This difference in gut microbiota composition between the two lakes' samples is attributed to more extensive aquaculture operations and poultry farms surrounding Shengjin Lake than Caizi Lake. ARGs-microbes associations indicated that 24 bacterial species, commonly used as indicators of antibiotic resistance in surveillance efforts, were abundant in wintering A. erythropus. The results revealed the composition and structural characteristics of the gut microbiota and ARGs of A. erythropus, pointing to their high sensitivities to diet habits at both lakes. This study also provides primary data for risk prevention and control of potential harmful pathogens that could endanger public health and therefore are of major significance to epidemiological and public health.

KEYWORDS

Anser erythropus, antibiotic-resistant genes (ARGs), gut microbiota, metagenomics, Shengjin and Caizi Lakes

Introduction

Globally, antibiotic pollution is a cause of growing concern and has become a serious threat to public health (Zhao D. D. et al., 2020). With the development of the modern pharmaceutical industry, more and more antibiotics, such as chloramphenicol, quinolones, tetracyclines, vancomycin, and sulfonamides, are being widely used in clinical treatment and in livestock and poultry breeding industries (Mu et al., 2015). As an emergent environmental pollutant, almost all antibiotics are not completely absorbed in nature, and their use causes the selection and spread of antibiotic-resistant genes (ARGs) (Salerno et al., 2022). Thus, ARGs have become a critical concern for public health and have attracted much attention in recent decades (Huang et al., 2017). Agricultural activities, human and animal disease treatments, and the breeding industry are considered potential reservoirs for ARGs (Zhao et al., 2010; Zhu et al., 2013). ARGs can spread or circulate among all ecosystems, such as humans and animals, thus threatening environmental and public health (Huang et al., 2017; Wang et al., 2020; Zhao H. et al., 2020).

Since wild animals are the major source and reservoir of ARGs, which are related to gut microbes, they are used as bioindicators of ARGs in the environment (Li et al., 2016; Cevidanes et al., 2020). Compared to other wild vertebrates, birds, in particular waterbirds, can fly long distances and develop unique diets, lifestyles, and complex physiological traits (Li et al., 2016; Cevidanes et al., 2020). Studies indicated that migratory waterbirds are an important group for the transmission of ARGs in ecosystems, as ARGs may quickly colonize in their gut as a result of the ingestion of polluted food or water (Franklin et al., 2020; Jarma et al., 2021). Migratory waterbirds have a relatively high population density, may encounter ARGs or acquire these genes through contact with each other, and may spread ARGs across large distances (Bauer and Hoye, 2014; Marcelino et al., 2019). Studies indicated that habitats and changes in the avian host diet significantly affect ARGs in the gut of avian species. These avian species, with different ecological patterns, are also likely to play different roles in ARG dispersion (Huang et al., 2017). However, knowledge of the role played by ARGs in the gut of migratory waterbirds is deficient. Only a small number of studies describe how ARGs can change in the same migratory waterbird wintering in different habitats.

As an important wetland indicator species and long-distance migratory waterbird, *A. erythropus* migrate from nesting and breeding areas (Fennoscandian Lapland to northeastern Siberia) in summer to wintering areas (Japan, China, and South Korea). In China, *A. erythropus* mainly stay at the Shengjin (SJ) and Caizi (CZ). Lakes in the Anhui Province in winter. *Carex* and subterranean tubers are their main food in the mudflats of Caizi Lake, whereas *Poaceae* spp. is their main diet in the grasslands of Shengjin Lake (Zhao et al., 2013). The present study hypothesizes that the different food sources that wintering *A. erythropus* consume at the two lakes change the composition and structure of ARGs in their gut. In this study, we performed metagenomics analysis to characterize the gut microbial communities of *A. erythropus* wintering at the two lakes, assessed whether or not their gut ARGs significantly differed between the two lakes, tested if the same migratory waterbird species at the two different wintering habitats played a different role in ARGs, and elucidated the relationship between ARGs and microbes in wintering *A. erythropus*.

Materials and methods

Ethical standards

No birds were harmed or killed in this study, and this research was approved by Anhui Medical University (No. 81220270).

Study areas

Both the Shengjin (30.25°-30.50°N, 116.92°-117.25°E) and Caizi Lakes (30.75°-30.97°N, 117.00°-117.15°E) are located in the southern part of Anhui Province, China, and are shallow lakes connected to the Yangtze River. Both lakes are major international wintering and stopover wetlands for migratory waterbirds in the East Asian-Australasian Flyway (Chen et al., 2011). Both lakes are surrounded by economically developed areas, with the highest population densities in China and with the pursuing of industrial and agricultural activities. However, the two lakes significantly differ from one another in terms of their environmental conditions. Shengjin Lake suffers from severe pollution, overexploitation, in particular due to cage aquaculture operations in recent decades, and pond and lowhead dams (Zhao et al., 2013; Li et al., 2015). In recent years, artificial aquaculture enclosures have been demolished at Shengjin Lake; however, many aquaculture and poultry farms still use large quantities of antibiotics for animal treatment. Pollution and overexploitation are less prevalent at Caizi Lake. Mainly paddy fields and a small number of cage aquaculture operations and poultry farms surround Caizi Lake. The water quality (total phosphorus, total nitrogen, permanganate value, chlorophyll-a, and transparency) of Caizi Lake is better than that of Shengjin Lake (Gu et al., 2014).

Sample collection and avian species determination

Twenty fecal samples were collected from paddy fields at the two lakes (10 samples each) in December 2021. A

large sample size of more than 100–150 geese was selected to collect fecal samples at foraging sites from the two lakes. Foraging behavior was observed using telescopes or binoculars before the fecal samples were collected. After the geese finished foraging and had defecated, we immediately collected the fresh fecal samples. To avoid individual repetition, human disturbance, and soil contamination, the distance between successive fecal samples was at least 5 m, and we collected only the center of each fecal mass. All samples were immediately transported to the laboratory and stored at -80° C (Dong et al., 2019; Xiang et al., 2019; Liu et al., 2020).

A DNA Stool Mini kit was used to extract the DNA from the avian fecal sample, and the *cox1* gene of the mitochondrial genome was used to confirm the avian species. The primers (BIRDF1: 5[']-TTC TCC AAC CAC AAA GAC ATT GGC AC-3['] and BIRDR1: 5[']-ACG TGG GAG ATA ATT CCA AAT CCT G-3[']) were used to identify the goose species *via* polymerase chain reaction (PCR) amplification (Liu et al., 2020). The successive cycling conditions of PCR amplification consisted of denaturation at 95°C for 5 min, 35 cycles of denaturation at 95°C for 30 s, annealing at 55°C for 45 s, extension at 72°C for 1.5 min, and a final extension at 72°C for 10 min. The PCR products were sequenced, and all resulting sequences were blasted in GenBank to ensure that all fecal samples in this study were taken from *A. erythropus* alone.

Illumina high-throughput sequencing and bioinformatics analysis

Metagenomic sequencing and analysis of the fecal samples were conducted using an Illumina HiSeq 2500 at OE Biotech Co., Ltd. (Shanghai, China). MEGAHIT (v1.1.2) software was used to perform metagenomic assembly after the recording of valid reads. Clean reads were aligned against the nonredundant gene set (95% identity) using Bowtie 2 (v2.2.9). The gene set representative sequence of the fecal samples in this study was annotated with the Kyoto Encyclopedia of Genes and Genomes (KEGG), Swiss-Prot, Non-Redundant Protein Sequence Database (NRDB), Clusters of Orthologous Groups of proteins (COGs), and Gene Ontology (GO) databases. KEGG, Carbohydrate-Active Enzymes (CAZy), Evolutionary Genealogy of Genes: Non-supervised Orthologous Groups (eggNOG) Database, and Comprehensive Antibiotic Resistance Database (CARD) of the unigenes were obtained. The linear discriminant analysis effect size (LEfSe) method was used to compare the two groups in order to determine the differentially abundant taxonomic features by using the non-parametric Kruskal-Wallis rank sum test.

Statistical analysis

Differences in the gut microbial communities and ARGs between the two geese populations were analyzed using the non-parametric Kruskal–Wallis rank sum test. All statistical analyses and associated plots, such as principal component analysis (PCA), pairwise Spearman's correlations, R scores, and *p*-values, were performed using GraphPad Prism v 7.0, SPSS 22 and Oebiotech tools available at https://cloud.oebiotech.cn/ task/.

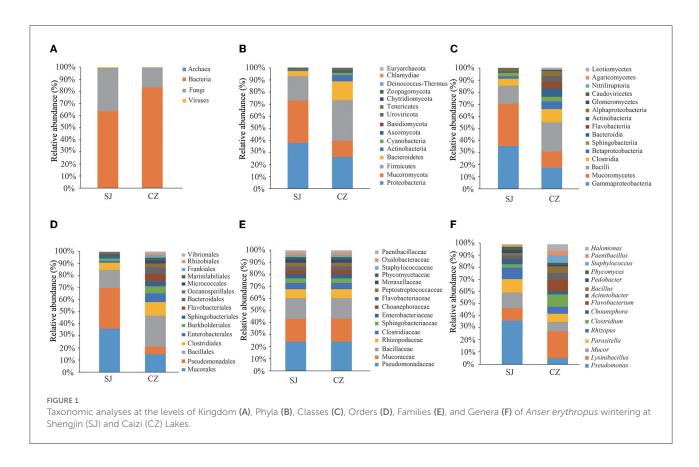
Results

Composition of microbial communities

The Illumina MiSeq 2500 sequencing run produced 1,186,628,964 raw reads. After the removal of low-quality reads, 1,182,900,206 clean reads corresponding to 7,935,970 operational taxonomic units (OTUs) were retained. Of the OTUs, 39.48% were found in both populations. Geese from Shengjin Lake had 10.02% unique OTUs, while geese from Caizi Lake had 11.02% unique OTUs. A Venn analysis indicated that 2,787 unique OTUs were identified in the 20 samples (Supplementary Figure 1). Four kingdoms, 164 phyla, 299 classes, 537 orders, 1,096 families, and 3,558 genera were identified in the gut microbes of A. erythropus. Bacteria, fungi, viruses, and archaea were the dominant gut microbes, accounting for 71.35, 28.06, 0.38, and 0.21% of the OTUs, respectively (Figure 1A). The abundance of bacteria was higher than those of archaea, eukaryotes, and viruses. Proteobacteria (32.69%), Mucoromycota (26.06%), Firmicutes (25.34%), Bacteroidetes (8.62%), and Actinobacteria (2.59%) were the dominant microbial phyla (Figure 1B). Gammaproteobacteria, Mucoromycetes, and Bacilli were the dominant microbial classes, accounting for 26.93, 2.545, and 1.75%, respectively (Figure 1C). Mucorales (1.273%) and Pseudomonadales (1.05%) were the dominant microbial orders (Figure 1D). Pseudomonadaceae, Mucoraceae, Bacillaceae, Rhizopodaceae, and Clostridiaceae were the dominant microbial families, accounting for 20.36, 15.73%, 14.32, 6.22, and 4.20%, respectively (Figure 1E). Pseudomonas (18.95%), Lysinibacillus (9.50%), Mucor (7.87%), and Parasitella (6.76%) were the dominant microbial genera (Figure 1F).

Diversity and differences in taxa of microbes

The results of PCA indicated that the microbiota compositions significantly differed between the two populations, and all the individuals were well-matched with their lakes (Figure 2A). The top 12 genera of microbial abundance were



found in all the samples between the two lakes (Figure 2B). Acinetobacter, Escherichia, Frankia, Halomonas, Klebsiella, and Staphylococcus had a significantly higher abundance of OTUs in the Shengjin Lake samples than in the Caizi Lake samples, while Choanephora, Phycomyces, Rhizopus, Mucor, and Parasitella were more abundant in the Caizi Lake samples than in the Shengjin Lake samples.

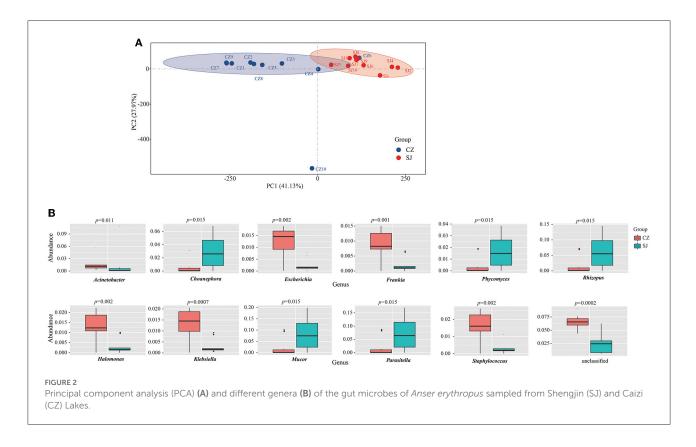
Functions of the gut microbes

The gut microbial functional pathways were annotated based on the eggNOG, CAZy, CARD, and KEGG databases. Most of the gene functions remained unknown (30.74%) according to the eggNOG annotation results. The eggNOG annotation results showed that replication, recombination, and repair (10.32%); energy production and conversion (7.86%); and amino acid transport and metabolism (6.99%) were the main functions (Figure 3A). The CAZy annotation results showed that the glycoside hydrolases (36.33%), glycosyl transferases (34.38%), and carbohydrate esterases (13.97%) were the primary functions (Figure 3B). Based on the CARD annotation results, antibiotic efflux was the most abundant function (72.21%) (Figure 3C). The KEGG results showed that carbohydrate metabolism (11.96%), energy metabolism (11.05%), and global

and overview maps (10.16%) were most abundant in the KEGG functions (Figure 3D). Between the gut microbial communities at the two lakes, there were no differences in eggNOG_description (p = 0.06), CAZy (p = 0.38), and CARD (p = 0.24). However, the KEGG results indicated a significant difference between the two communities (p = 0.02).

Abundance, diversity, network analysis, and differences in taxa of ARGs

In total, 2,218 ARGs were detected in the gut of *A. erythropus*, categorized into 34 resistant types. Tetracycline (14.55%), macrolide (13.72%), fluoroquinolone (13.41%), phenicol (12.34%), and peptide (10.14%) antibiotics were the dominant resistant types (Figure 4A). Of all the ARGs, 44.84% were found in both populations. Geese from Shengjin Lake had 10.32% unique ARGs, while those from Caizi Lake had none (Figure 4B). The ARG results indicated that the relative abundance of OTUs significantly differed between the samples from the two lakes (p = 0.0001) (Figure 4C). All samples were further analyzed using the PCA method according to the ARG abundance. All the Shengjin Lake samples, except for SJ5, were grouped together and significantly differed from the Caizi Lake samples, except for CZ6 (Figure 4D). The results of the top 100



ARGs' network analysis can be split into two major modules that consisted of 23 and 77 total nodes in each (Figure 5). *RpoB2* was the hub of Module I, whereas *MexK*, *MexN*, *MuxB*, and *MexW* were the hubs of Module II. The top 12 ARGs were found in all the samples from the two lakes and had a significantly higher abundance of OTUs in the Shengjin Lake samples than in the Caizi Lake samples (Figure 6).

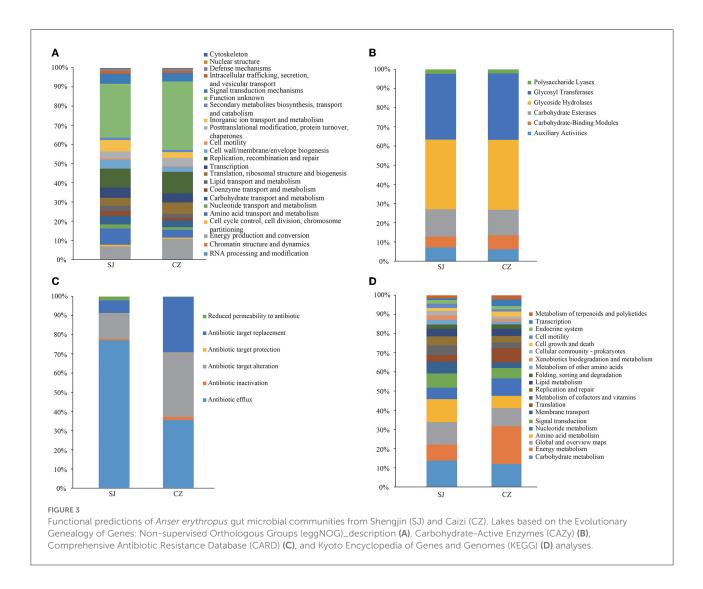
ARGs-microbe association analysis

Correlation analysis confirmed associations between ARGs and bacteria. One hundred and four bacterial species represented most of the ARGs. Twenty-four bacterial species, including *Arcobacter cloacae*, *Vibrio harveyi*, *Vibrio cholerae*, *Stenotrophomonas maltophilia*, *Serratia marcescens*, *Serratia fonticola*, and *Riemerella anatipestifer*, commonly used as bioindicators of antibiotic resistance in surveillance efforts, were abundant in wintering *A. erythropus*. The frequencies of *Efra*, *Rpob2*, *Tet*(38), and *Bmr* were most correlated with the abundance of microbes (Figure 7).

Discussion

In our study, Proteobacteria, Mucoromycota, and Firmicutes were the most abundant gut microbiota in A.

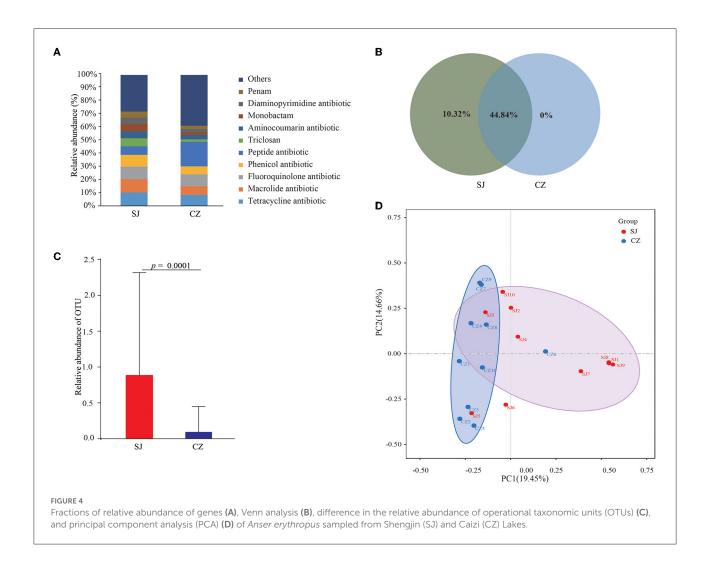
erythropus. During the wintering period, geese require a great amount of energy to cope with the cold, and Proteobacteria may play an important role in energy accumulation (Dong et al., 2019). Mucoromycota are common fungi in the highly versatile metabolic gut system, and these fungi may act as powerful microbial cell factories for biorefinery applications (Dzurendova et al., 2022). Firmicutes could help the geese to absorb nutrients and intake energy from the metabolism of polysaccharides, fatty acids, carbohydrates, and sugars (Flint et al., 2008; Tap et al., 2009). The gut microbial composition and characteristics of birds are different from those of other vertebrates due to complex and unique avian physiological traits, diets, and migration strategies (Song et al., 2020; Li et al., 2021). Most studies have highlighted that the avian gut microbiome is influenced by diet, age, genetics, captivity, and changes in their habitats (Hammons et al., 2010; Waite and Taylor, 2014; Barbosa et al., 2016; Wang et al., 2016; Díaz-Sánchez et al., 2019). The diet was the primary driver of the gut microbial community structure for the host, as the food source of wintering A. erythropus was different at the two lakes. The species fed exclusively on Poaceae spp. at Shengjin Lake and primarily on Carex spp. at Caizi Lake (Wang et al., 2012; Zhao et al., 2012). This feeding behavior suggested that the different food intakes of wintering A. erythropus at the two lakes changed their gut microbial community compositions. It was reported that migratory birds show a strong-site fidelity for wintering and breeding areas and often return to the same site each year



in winter (Møller and Szép, 2011). This phenomenon may be related to the fact that *A. erythropus* wintering at Shengjin Lake migrated from a different breeding location, compared to those at Caizi Lake (Liu et al., 2020). However, no detailed information regarding the migration routes of *A. erythropus* wintering at the two lakes is available. The gut microbial communities and functions of wintering *A. erythropus* differed between the two lakes, as supported by the 16S rRNA gene sequencing results (Liu et al., 2020). However, the dominant microbial phyla were different from the 16S rRNA gene sequencing results, in that Proteobacteria was the most abundant phyla in the metagenomic results, while Firmicutes was the highest in the 16S rRNA gene sequencing results (Liu et al., 2020).

Antibiotic-resistant genes have been considered an emerging environmental contaminant in aquaculture, wastewater, soil, sewage plants, and livestock feces, threatening worldwide public health (Durso et al., 2012; Cevidanes et al., 2020). Previous studies showed that ARGs have increased in the environment

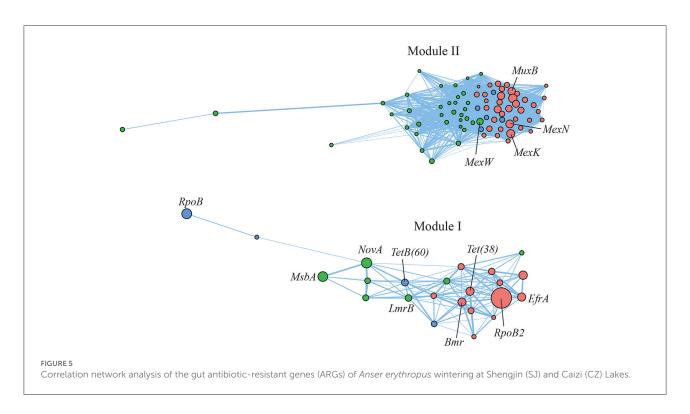
due to animal and human disease treatments and agricultural activities. The spread and aggregation of ARGs into resistant pathogens have challenged life-saving antibiotic therapies in recent decades (Zhu et al., 2013; Cevidanes et al., 2020; Wang et al., 2020). ARGs can be transferred among species in the wild through several mobile genetic elements, such as integrons, plasmids, and transposons (Bellanger et al., 2014; Blair et al., 2015; Chen et al., 2015; Mu et al., 2015; Kipkorir et al., 2020). Wild birds are under the selective pressure of antibiotics, in that not only do these antibiotics kill beneficial gut bacteria, but they also alter the composition, structure, and function of the gut microbiome, even causing some bacteria to completely disappear from the gut (Wang et al., 2017). Migratory birds can acquire and transmit ARGs from contaminated environments along migration flyways (Foti et al., 2011). ARGs in the gut of migratory birds are considered an emerging public health problem since their gut microbes can carry and spread ARGs to humans, and these ARGs render antibiotic drugs obsolete

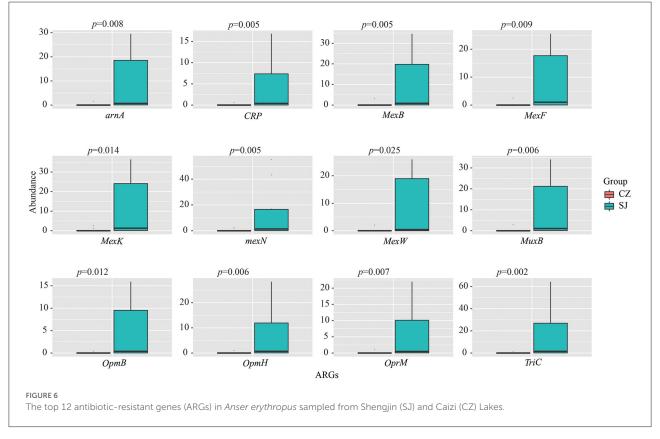


(Yuan et al., 2021). As a long-distance migratory waterbird, A. erythropus is found in various ecosystems; however, ARGs in this species have received little attention. This study is the first to explore ARGs in A. erythropus wintering at the two Lakes. The abundance of ARGs in wintering A. erythropus was found to be higher at Shengjin Lake than at Caizi Lake. This result was consistent with the abundance of their gut microbial communities and may be due to the heterogeneous environmental factors of the two lakes (Liu et al., 2020). It was reported that microbes can harbor ARGs, selected by a variety of antibiotics, acquire antibiotic resistance, frequently switch hosts within the same microbial community or among different microbial communities, and spread from the environment to migratory waterbirds (Sommer et al., 2009; Bonnedahl and Jarhult, 2014; Liu et al., 2016). ARGs are not only resistant to antibiotics but are also critical to the environment. A recent study demonstrated that the external living environment affects the gut ARG diversities and the origin of ARGs in the guts of waterbirds in the environment (Jarma et al., 2021). With

the rapid rates of economic, population, and consumption growth, ARGs have been increasingly used for agriculture and aquaculture around Shengjin and Caizi Lakes (Li et al., 2015). Therefore, the ARGs in the gut of *A. erythropus* appear to originate from the habitats at the two lakes. A higher ARG diversity was observed in the geese wintering at Shengjin Lake than at Caizi Lake, probably because aquaculture operations and poultry farms affected Shengjin Lake more than Caizi Lake. The CRP and cpxR types were highly abundant in the gut of *A. erythropus*. The CRP type is a regulator that modulates bacterial antibiotic resistance, and its mutants can increase the resistance ability of bacterial strains (Nishino et al., 2008). CpxR is identified as a regulator of the cell envelope stress response and acts as an activator to enhance the expression of the efflux pump and improve drug resistance (Tian et al., 2016).

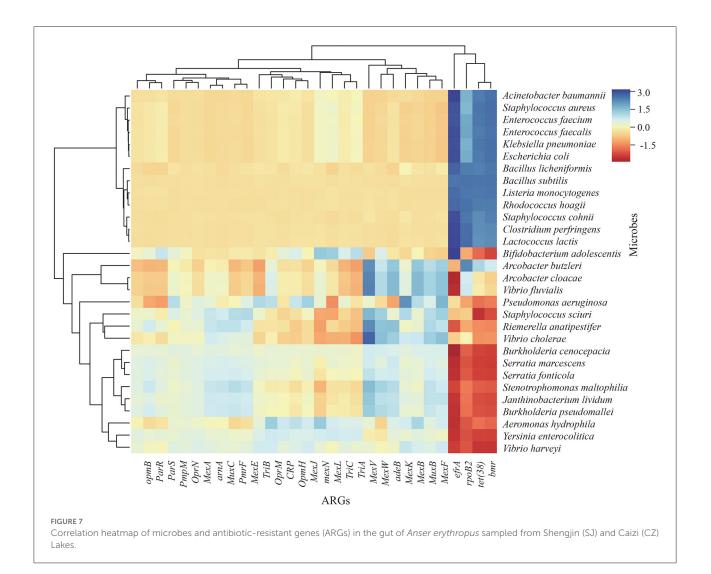
The gut microbiota and ARGs in *A. erythropus* wintering at the two lakes were correlated with the physicochemical properties of the two lakes. It was previously reported that the concentrations of chemical ions in the surface





and groundwater of the study area were higher than those of atmospheric precipitation, while total phosphorus, total

nitrogen, chlorophyll-a, and transparency values of the surface water pointed to eutrophication (Cui et al., 2021; Gao et al.,



2022; Zhao et al., 2022). The trophic state index and bacterial eutrophic index values showed that the two lakes were slightly eutrophic (Gao et al., 2022; Zhao et al., 2022). These previously reported results indicate that the physicochemical properties of these two lakes affect the structure and function of the gut microbiota and ARGs in *A. erythropus*. These inferences remain to be validated in the future.

Wild animals serve as important carriers of ARGs and play a vital role in their spread worldwide. Migratory birds serve as an important reservoir of resistant bacteria and ARGs and can carry them over long distances, even across continents. Thus, migratory birds may operate as a transmission route and are partly responsible for the global dissemination of ARGs. This study is the first step in describing the characterization of the presence of ARGs in the gut of *A. erythropus* wintering at the two lakes. The presence of elevated levels of ARGs in *A. erythropus* feces poses a significant risk of acquisition by human pathogens. *A. erythropus* has an enormous potential to spread ARGs to places where humans are most vulnerable to them. These results underscore the need to limit human exposure to bird feces and to proactively manage the associated risks. Although the origin of ARGs in A. erythropus is still to be clarified, as wild birds are not directly exposed to antibiotics, contact with sewage, soil, or animal manure may affect humans. According to this study, these geese carry ARGs during their migration and become a major source of ARGs in the local environment. Overall, this study suggests that different food resources and habitat heterogeneity affect the microbiota community's composition and structure in A. erythropus and their exposition and acquisition of ARGs. However, there are some limitations in this study that need to be addressed in future studies. First, its relatively small sample size should be increased so as to represent a more spatiotemporal sample. Second, soil, water, plant, and air samples are required to correlate A. erythropus and their gut microbial communities as well as ARG compositions with environmental factors.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: https://www.ncbi. nlm.nih.gov/genbank/, PRJNA894380.

Ethics statement

The animal study was reviewed and approved by this research was approved by Anhui Medical University (No. 81220270).

Author contributions

GL designed the experiments, analyzed the data, and wrote the manuscript. JF and NX performed the experiments. All authors contributed to the article and approved the submitted version.

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References

Barbosa, A., Balagué, V., Valera, F., Martínez, A., Benzal, J., Motas, M., et al. (2016). Age-related differences in the gastrointestinal microbiota of chinstrap penguins (*Pygoscelis antarctica*). *PLoS ONE* 11, 0153215. doi: 10.1371/journal.pone.0153215

Bauer, S., and Hoye, B. J. (2014). Migratory animals couple biodiversity and ecosystem functioning worldwide. *Science* 344, 1242552. doi: 10.1126/science.1242552

Bellanger, X., Guilloteau, H., Bonot, S., and Merlin, C. (2014). Demonstrating plasmidbased horizontal gene transfer in complex environmental matrices: a practical approach for a critical review. *Sci. Total Environ.* 493, 872–882. doi: 10.1016/j.scitotenv.2014.06.070

Blair, J. M. A., Webber, M. A., Baylay, A. J., Ogbolu, D. O., and Piddock, L. J. V. (2015). Molecular mechanisms of antibiotic resistance. *Nat. Rev. Microbiol.* 13, 42–51. doi: 10.1038/nrmicro3380

Bonnedahl, J., and Jarhult, J. D. (2014). Antibiotic resistance in wild birds. *Upsala J. Med. Sci.* 119, 113–116. doi: 10.3109/03009734.2014.905663

Cevidanes, A., Esperón, F., Cataldo, D. S., Neves, E., Sallaberry-Pincheira, N., and Millán, J. (2020). Antimicrobial resistance genes in Andean foxes inhabiting anthropized landscapes in central Chile. *Sci. Total Environ.* 724, 138247. doi: 10.1016/j.scitotenv.2020.138247

Chen, B., Hao, L. J., Guo, X. Y., Wang, N., and Ye, B. P. (2015). Prevalence of antibiotic resistance genes of wastewater and surface water in livestock farms of Jiangsu Province, China. *Environ. Sci. Pollut. Res.* 22, 13950–13959. doi: 10.1007/s11356-015-4636-y

Chen, J. Y., Zhou, L. Z., Zhou, B., Xu, R. X., Zhu, W. Z., and Xu, W. B. (2011). Seasonal dynamics of wintering waterbirds in two shallow lakes along Yangtze River in Anhui Province. *Zool. Res.* 32, 540–548. doi: 10.3724/SP.J.1141.2011.05540

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmicb. 2022.1081468/full#supplementary-material

Cui, Y. H., Wang, J., Liu, Y. C., Hao, S., and Gao, X. (2021). Hydrochemical characteristics and ion origin analysis of surface groundwater at the Shengjin Lake and Yangtze River interface. *Huan Jing Ke Xue* 42, 3223–3231. doi: 10.13227/j.hjkx.202012016

Díaz-Sánchez, S., Perrotta, A. R., Rockafellow, I., Alm, E. J., Okimoto, R., Hawken, R., et al. (2019). Using fecal microbiota as biomarkers for predictions of performance in the selective breeding process of pedigree broiler breeders. *PLoS ONE* 14, e0216080. doi: 10.1371/journal.pone. 0216080

Dong, Y., Xiang, X., Zhao, G., Song, Y., and Zhou, L. (2019). Variations in gut bacterial communities of hooded crane (*Grus monacha*) over spatial-temporal scales. *Peer J.* 7, e7045. doi: 10.7717/peerj.7045

Durso, L. M., Miller, D. N., and Wienhold, B. J. (2012). Distribution and quantification of antibiotic resistant genes and bacteria across agricultural and non- agricultural metagenomes. *PLoS ONE* 7, e48325 doi: 10.1371/journal.pone. 0048325

Dzurendova, S., Losada, C. B., Dupuy-Galet, B. X., Fjær, K., and Shapaval, V. (2022). Mucoromycota fungi as powerful cell factories for modern biorefinery. *Appl. Microbiol. Biotechnol.* 106, 101–115. doi: 10.1007/s00253-021-11720-1

Flint, H. J., Bayer, E. A., Rincon, M. T., Lamed, R., and White, B. A. (2008). Polysaccharide utilization by gut bacteria: potential for new insights from genomic analysis. *Nat. Rev. Microbiol.* 6, 121-131. doi: 10.1038/nrmicro 1817

Foti, M., Rinaldo, D., Guercio, A., Giacopello, C., Aleo, A., and De Leo, F. (2011). Pathogenic microorganisms carried by migratory birds passing through

the territory of the island of Ustica, Sicily (Italy). Avian Pathol. 40, 405–409. doi: 10.1080/03079457.2011.588940

Franklin, A. B., Ramey, A. M., Bentler, K. T., Barrett, N. L., Mccurdy, L. M., Ahlstrom, C. A., et al. (2020). Gulls as sources of environmental contamination by Colistin-resistant bacteria. *Sci. Rep.* 10:4408. doi: 10.1038/s41598-020-61318-2

Gao, N., Liang, Y., Li, J., Cui, K., and Lu, W. (2022). Bacterial community composition and indicators of water quality in Caizi Lake, a typical Yangtze-connected freshwater lake. *FEMS Microbiol. Lett.* 369, fnac084. doi: 10.1093/femsle/fnac084

Gu, Y., Li, F. S., Li, C. X., Huang, W. X., Yao, J. B., and Zhang, H. (2014). Comprehensive evaluation of Anqing Yangtze River wetland eutrophication of lakes. *J. Anqing Teach. Coll.* 20, 121–124. doi: 10.13757/j.cnki.cn34-1150/n.2014.04.029

Hammons, S., Oh, P. L., Martínez, I., Clark, K., Schlegel, V. L., Sitorius, E., et al. (2010). A small variation in diet influences the *Lactobacillus strain* composition in the crop of broiler chickens. *Syst. Appl. Microbiol.* 33, 275–281. doi: 10.1016/j.syapm.2010.04.003

Huang, L., Xu, Y. B., Xu, J. X., Ling, J. Y., Chen, J. L., and Zhou, J. L. (2017). Antibiotic resistance genes (ARGs) in duck and fish production ponds with integrated or non-integrated mode. *Chemosphere* 168, 1107–1114. doi: 10.1016/j.chemosphere.2016.10.096

Jarma, D., Sánchez, M. I., Green, A. J., Peralta-Sánchez, J. M., Hortas, F., and Sánchez-Melsió, A. (2021). Faecal microbiota and antibiotic resistance genes in migratory waterbirds with contrasting habitat use. *Sci. Total. Environ.* 783, 146872. doi: 10.1016/j.scitotenv.2021.146872

Kipkorir, C. K., Angienda, O. P., Onyango, M. D., and Onyango, O. P. (2020). Antibiotic resistance of *Escherichia coli* from humans and black rhinoceroses in Kenya. *Ecohealth* 17, 41–51. doi: 10.1007/s10393-019-01461-z

Li, C., Liu, Y., Gong, M., Zheng, C., Zhang, C., Li, H., et al. (2021). Diet-induced microbiome shifts of sympatric overwintering birds. *Appl. Microbiol. Biotechnol.* 105, 5993–6005. doi: 10.1007/s00253-021-11448-y

Li, C., Zhou, L., Xu, L., Zhao, N., and Beauchamp, G. (2015). Vigilance and activity time-budget adjustments of wintering hooded cranes, *Grus monacha*, in human-dominated foraging habitats. *PLoS ONE* 10, e0118928. doi: 10.1371/journal.pone.0118928

Li, T. T., Long, M., Ji, C., Shen, Z., Gatesoupe, F. J., Zhang, L., et al. (2016). Alterations of the gut microbiome of largemouth bronze gudgeon (*Coreius guichenoti*) suffering from furunculosis. *Sci. Rep.* 6, 30606. doi: 10.1038/srep30606

Liu, G., Gong, Z., and Li, Q. (2020). Variations in gut bacterial communities between lesser white-fronted geese wintering at Caizi and Shengjin lakes in China. *Microbiol. Open* 9, e1037. doi: 10.1002/mb03.1037

Liu, Y. Y., Wang, Y., Walsh, T. R., Yi, L. X., Zhang, R., Spencer, J., et al. (2016). Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect. Dis.* 16, 161–168. doi: 10.1016/S1473-3099(15)00424-7

Marcelino, V. R., Wille, M., Hurt, A. C., González-Acuña, D., Klaassen, M., Schlub, T. E., et al. (2019). Meta-transcriptomics reveals a diverse antibiotic resistance gene pool in avian microbiomes. *BMC Biol.* 17, 31. doi: 10.1186/s12915-019-0649-1

Møller, A. P., and Szép, T. (2011). The role of parasites in ecology and evolution of migration and migratory connectivity. J. Ornithol. 152, 141–150. doi: 10.1007/s10336-010-0621-x

Mu, Q. H., Li, J., Sun, Y. X., Mao, D. Q., Wang, Q., and Luo, Y. (2015). Occurrence of sulfonamide-, tetracycline-, plasmid-mediated quinolone- and macrolide-resistance genes in livestock feedlots in Northern China. *Environ. Sci. Pollut. Res.* 22, 6932–6940. doi: 10.1007/s11356-014-3905-5

Nishino, K., Senda, Y., and Yamaguchi, A. (2008). CRP regulator modulates multidrug resistance of *Escherichia coli* by repressing the mdtEFMulti drug efflux genes. *J. Antibiot.* 61, 120–127. doi: 10.1038/ja.2008.120

Salerno, B., Furlan, M., Sabatino, R., Di Cesare, A., Leati, M., Volanti, M., et al. (2022). Antibiotic resistance genes load in an antibiotic free organic broiler farm. *Poult. Sci.* 101, 101675. doi: 10.1016/j.psj.2021.101675

Sommer, M. O. A., Dantas, G., and Church, G. M. (2009). Functional characterization of the antibiotic resistance reservoir in the human microflora. *Science* 325, 1128–1131. doi: 10.1126/science.1176950

Song, S. J., Sanders, J. G., Delsuc, F., Metcalf, J., Amato, K., Taylor, M. W., et al. (2020). Comparative analyses of vertebrate gut microbiomes reveal convergence between birds and bats. *MBio* 11, 14. doi: 10.1128/mBio.02901-19

Tap, J., Mondot, S., Levenez, F., Pelletier, E., Caron, C., Furet, J. P., et al. (2009). Towards the human intestinal microbiota phylogenetic core. *Environ. Microbiol.* 11, 2574–2584. doi: 10.1111/j.1462-2920.2009.01982.x

Tian, Z. X., Yi, X. X., Cho, A., O'Gara, F., and Wang, Y. P. (2016). CpxR activates MexAB-OprM efflux pump expression and enhances antibiotic resistance in both laboratory and clinical nalB-type isolates of *Pseudomonas aeruginosa*. *PLoS Pathog*. 12, e1005932. doi: 10.1371/journal.ppat.1005932

Waite, D. W., and Taylor, M. W. (2014). Characterizing the avian gut microbiota: membership, driving influences, and potential function. *Front. Microbiol.* 5, 223. doi: 10.3389/fmicb.2014.00223

Wang, J., Ma, Z. B., Zeng, Z. L., Yang, X. W., Huang, Y., and Liu, J. H. (2017). The role of wildlife (wild birds) in the global transmission of antimicrobial resistance genes. *Zool Res.* 38, 55–80. doi: 10.24272/j.issn.2095-8137.2017.024

Wang, W., Zheng, S., Sharshov, K., Cao, J., Sun, H., Yang, F., et al. (2016). Distinctive gut microbial community structure in both the wild and farmed Swan goose (*Anser cygnoides*). *J. Basic Microbiol.* 56, 1299–1307. doi: 10.1002/jobm.201600155

Wang, X., Fox, A. D., Cong, P., Barter, M., and Cao, L. (2012). Changes in the distribution and abundance of wintering lesser white-fronted geese *Anser erythropus* in eastern China. *Bird Conserv. Int.* 22, 128. doi: 10.1017/S095927091100030X

Wang, Z., Han, M. Z., Lia, E. H., Liu, X., Wei, H. M., Yang, C., et al. (2020). Distribution of antibiotic resistance genes in an agriculturally disturbed lake in China: their links with microbial communities, antibiotics, and water quality. *J. Haz. Mat.* 393, 122426. doi: 10.1016/j.jhazmat.2020.122426

Xiang, X., Zhang, F., Fu, R., Yan, S., and Zhou, L. (2019). Significant differences in bacterial and potentially pathogenic communities between sympatric hooded crane and greater white-fronted goose. *Front. Microbiol.* 5, 163. doi: 10.3389/fmicb.2019.00163

Yuan, Y., Liang, B., Jiang, B. W., Zhu, L. W., Wang, T. C., Li, Y. G., et al. (2021). Migratory wild birds carrying multidrug-resistant *Escherichia coli* as potential transmitters of antimicrobial resistance in China. *PLoS ONE* 16, e0261444. doi: 10.1371/journal.pone.0261444

Zhao, D. D., Liu, G. J., Wang, X. F., Daraz, U., and Sun, Q. Y. (2020). Abundance of human pathogen genes in the phyllosphere of four landscape plants. *J. Environ. Manage.* 255, 109933. doi: 10.1016/j.jenvman.2019.109933

Zhao, F. T., Zhou, L. Z., and Xu, W. B. (2013). Habitat utilization and resource partitioning of wintering Hooded Cranes and three goose species at Shengjin Lake. *Chinese Birds* 4, 281–290. doi: 10.5122/cbirds.2013.0032

Zhao, H., Sun, R., Yu, P., and Alvarez, P. J. J. (2020). High levels of antibiotic resistance genes and opportunistic pathogenic bacteria indicators in urban wild bird feces. *Environ. Pollut.* 266(Pt 2), 115200. doi: 10.1016/j.envpol.2020. 115200

Zhao, L., Dong, Y. H., and Wang, H. (2010). Residues of veterinary antibiotics in manures from feedlot livestock in eight provinces of China. *Sci. Total Environ.* 408, 1069–1075. doi: 10.1016/j.scitotenv.2009.11.014

Zhao, M. J., Cong, P. H., Barter, M., Fox, A. D., and Cao, L. (2012). The changing abundance and distribution of greater white-fronted geese *Anser albifrons* in the Yangtze River floodplain: impacts of recent hydrological changes. *Bird Conserv. Int.* 22, 135–143. doi: 10.1017/S0959270911000542

Zhao, W., Liu, Z., Guo, W., and Zhou, Z. (2022). Effect of aquatic vegetation restoration after removal of culture purse seine on phytoplankton community structure in Caizi Lakes. *Diversity* 14, 395. doi: 10.3390/d14050395

Zhu, Y. G., Johnson, T. A., Su, J. Q., Qiao, M., Guo, G. X., Stedtfeld, R. D., et al. (2013). Diverse and abundant antibiotic resistance genes in Chinese swine farms. *Proc. Natl. Acad. Sci. U. S. A.* 110, 3435–3440. doi: 10.1073/pnas.1222743110