Check for updates

OPEN ACCESS

EDITED BY Kaifang Zeng, Southwest University, China

REVIEWED BY Porfirio Gutierrez-Martinez, Instituto Tecnológico de Tepic, Mexico Elsherbiny A. Elsherbiny, Mansoura University, Eqvot

*CORRESPONDENCE Abir El-araby ⊠ abir.elaraby@usmba.ac.ma

SPECIALTY SECTION This article was submitted to Food Microbiology, a section of the journal Frontiers in Microbiology

RECEIVED 03 December 2022 ACCEPTED 21 December 2022 PUBLISHED 10 January 2023

CITATION

El-araby A, Azzouzi A, Ayam IM, Samouh KF and Errachidi F (2023) Survey on technical management of strawberries in Morocco and evaluation of their postharvest microbial load. *Front. Microbiol.* 13:1115340. doi: 10.3389/fmicb.2022.1115340

COPYRIGHT

© 2023 El-araby, Azzouzi, Ayam, Samouh and Errachidi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Survey on technical management of strawberries in Morocco and evaluation of their post-harvest microbial load

Abir El-araby ¹*, Amal Azzouzi¹, Iman Msegued Ayam¹, Khaoula Filali Samouh² and Faouzi Errachidi¹

¹Functional Ecology and Environment Engineering Laboratory, Faculty of Science and Technology, Sidi Mohamed Ben Abdellah University, Fez, Morocco, ²Department of Biology, Immunology and Biodiversity Laboratory, Faculty of Sciences Ain Chock, University Hassan II, Casablanca, Morocco

The climatic conditions of the growing regions influence the fruit's microbiological quality and their tolerance to post-harvest pathogens. The present work aims to identify the prevalence of bacterial and fungal strains of strawberries (Fragaria x ananassa) in the Gharb and Loukkos regions of Morocco. Thus, to establish a correlation between the microbial load and the climatic conditions of the two targeted regions. The bacteriological analyses were studied according to the International Organization for Standardization (ISO) methodologies. Regarding the mycological study, fungal species determination was performed using identification keys. Yeast species determination was done using genus analysis, assimilation, and fermentation tests. Emberger bioclimatic quotients (Q₂) were calculated for the Gharb and Loukkos regions and bioclimatic stages were determined. Salmonella spp. was not detected in the studied samples. However, Listeria monocytogenes and Escherichia coli were isolated from the Gharb samples. Sulfite-reducing clostridia spores were found in two Gharb samples versus one Loukkos sample. Coagulase-positive staphylococci were negative in all samples analyzed. Loukkos and Gharb regions were contaminated by Bacillus cereus with percentages of occurrence of 5.2 and 7.8%, respectively. The fungi found in strawberries from both regions were Aspergillus niger, Botrytis cinerea, Fusarium spp., Penicillium spp., Rhizopus spp., and Alternaria alternata with a significant predominance in the Loukkos samples. Indeed, a marked presence is noted for Candida sake and Rhodotorula glutinis in strawberries from Loukkos. Gharb is located on the semi-arid stage, while Loukkos is located on the sub-humid stage. Climatic conditions have a strong influence on plant microbial load, which explains the prevalence of bacteria in strawberries from Gharb and the prevalence of fungi in strawberries from Loukkos.

KEYWORDS

climatic conditions, microbiological study, Morocco, post-harvest, pre-harvest, strawberries

1. Introduction

Fruits are living organisms with a shelf life largely influenced by ambient temperature, relative humidity, precipitation, atmosphere composition during and after harvest, and type and degree of infection by microorganisms (Singh and Sharma, 2018). Strawberry fruits are considered to be a source of vitamins, minerals, and natural substances necessary for the proper functioning of the human body. These fruits are considered beneficial to health due to their antioxidant and antibacterial properties (Steinka and Kukułowicz, 2018). In general, red fruits have limited storage potential due to their extreme tenderness, vulnerability to mechanical damage, high respiration and transpiration rates, and susceptibility to fungal spoilage (Matar et al., 2020). These soft fruits are highly perishable, which leads to a rapid deterioration of their appearance and texture once they reach the market, generating significant economic losses for the strawberry industry worldwide.

Therefore, it is essential to apply adequate practices before and after harvesting strawberry fruits. The most widely used method to maintain strawberry quality is rapid post-harvest cooling also continuous low-temperature $(0-4^{\circ}C)$ storage (Tahir et al., 2018). High CO₂ atmospheres are one of the best methods to control post-harvest strawberries spoilage (Nakata and Izumi, 2020). Other postharvest techniques have also been applied to extend the shelf life of these susceptible fruits, such as gamma rays (de Jesus Filho et al., 2018), chemical application (Shu et al., 2020), and edible coatings (Mohammadi et al., 2021). Post-harvest treatments of strawberry fruits are of great importance to improve the quality and shelf life of these fruits considered extremely perishable, while minimizing losses and maximizing benefits.

Rots development caused by certain fungal pathogens is one of the major causes of postharvest losses in strawberries. Fungal strains commonly affecting strawberries in postharvest are Botrytis cinerea, Rhizopus stolonifer, Penicillium spp., and Aspergillus spp. (Feliziani and Romanazzi, 2016; Saleh and Al-Thani, 2019; Weber and Hahn, 2019; Trinetta et al., 2020). Studies have reported that microflora isolated from strawberries included bacteria, such as Escherichia coli, Bacillus cereus, Staphylococcus aureus, and Staphylococcus epidermidis (Yoon et al., 2010; Delbeke et al., 2015; Oliveira et al., 2019). Data from the literature confirm the probable presence of microorganisms such as Salmonella spp., Listeria monocytogenes, and B. cereus on strawberry fruits surface (Dziedzinska et al., 2018; Ortiz-Solà et al., 2020). In fact, the climatic conditions of the locality of origin have a strong influence on the microbial load of fruits and vegetables. Pre-harvest weather factors, including rainfall, humidity, and temperature, affect the microbiological quality of the fruit and its tolerance to infection and the development of post-harvest pathogens (Machado-Moreira et al., 2019; Bui et al., 2021).

The present work aims to identify the prevalence of bacterial and fungal strains of strawberry fruits (Fragaria × Ananassa) in the Gharb (Rabat-Sale-Kenitra) and Loukkos (Tangier-Tetouan-Al Hoceima) regions. Thus, to establish a correlation between microbial load and climatic conditions of the two targeted regions.

2. Materials and methods

2.1. Strawberry sampling

In this microbiological study, we focused mainly on the two areas closest to our city (Fez) to avoid strawberry samples quality deterioration. Strawberry fruit (Fragaria × Ananassa) samples were collected from two suppliers during the year 2020. The samples (n = 50) were collected from two sites (one located in the Gharb area and the other located in the Loukkos area) and stored at 4°C at the Laboratory of Functional Ecology and Environment Engineering on the same days (Figure 1).

2.2. Bacteriological study of strawberry samples from the Loukkos and Gharb regions

2.2.1. Samples preparation

Twenty-five grams (25 g) of each strawberry fruit sample were suspended in 225 ml of sterile buffered peptone water (BPW) solution and homogenized with a grinder (Stomacher[®] 400 circulator) for 2 min. Dilution series were prepared from stock solution. A volume of 0.1 ml of each dilution was transferred to Petri dishes surface containing culture media suitable for each target microorganism. However, for the detection of *L. monocytogenes*, 25 g of each sample were homogenized in 225 ml of PALCAM broth and incubated at 30°C for 24 h.





2.2.2. Bacteriological analysis

Microbiological analyses were studied according to the International Organization for Standardization (ISO) methodologies. The bacteriological food safety and quality parameters studied were: *E. coli* (ISO, 2001), *Salmonella* spp. (ISO, 2002), *L. monocytogenes* (ISO, 1996; ISO, 1998), *B. cereus* (ISO, 2004), Coagulase-positive staphylococci (ISO, 1999), and Sulfitereducing clostridia spores (IPQ, 1986).

Presumptive *L. monocytogenes* colonies on PALCAM agar were streaked on sheep blood agar to assess hemolysis, carbohydrate zymogram (rhamnose, xylose, and mannitol), and the Christie-Atkins-Munch-Petersen (CAMP) test (Ortiz-Solà et al., 2020). For suspected *Salmonella* spp. colonies, biochemical confirmation was performed with Triple Sugar Iron (TSI) agar, urea agar, and L-lysine decarboxylase medium (Uyttendaele, 2014). For the determination of *E. coli* sample was placed in Petri dish center, subsequently, 20 ml of violet red bile agar (BD Difco, heated to 45° C) was added. After solidification, agar was overlaid with 5 ml of violet red bile agar (double strength) supplemented with 4-Methylumbelliferyl- β -D-glucuronide. The inverted plates were incubated at 35° C for 24 h, and colonies fluoresced under UV light as *E. coli* bacteria (Cárdenas et al., 2013).

For detection of Sulfite-reducing clostridia spores, 10 ml of each sample in buffered peptone water (BPW) solution was heatshocked at 80-85°C for 10 min before culture to inactivate vegetative bacteria and enhance sporulation phenomenon. Samples were then transferred to 10 ml of differential reinforced clostridium medium (DRCM) and incubated under anaerobic conditions in a jar at 37°C for 5 days. This method is based on ferric Sulfite reduction to iron sulfide and samples are considered positive when color changes (black dots) (Oliveira et al., 2019). Bacilli showing typical B. cereus group growth, i.e., rough, dry colonies with a pink-purple background surrounding an egg yolk precipitation, were counted, isolated, and selected for hemolytic activity, tested on sheep blood agar plates after 1 day of incubation at 30°C. Convex black colonies of staphylococci, with or without halo on Baird-Parker medium for 24-48h at 37°C, were isolated and tested for catalase reaction and coagulase activity using rabbit plasma (Evêncio-Luz et al., 2012).

2.3. Mycological study of strawberry samples from the Loukkos and Gharb regions

2.3.1. Molds study

Analysis of mycoflora associated with strawberry fruits was performed using the modified Buvard method (Benkirane, 1995). Strawberries with lesions were disinfected with 1% sodium hypochlorite. Then, strawberries pieces were placed in Petri dishes containing Potato Sucrose Agar (PSA) and incubated for 7 days in the dark at 24°C. Optical microscopic examination of the different cultures allowed for the determination of fungal species using the keys identification of Gilman (1957), Tarr (1962), Ellis (1971), Chidambaram et al. (1974), Domsch et al. (1980), and Champion (1997). Contamination percentage by different fungal species is calculated according to Ponchet (1966) method which defines the isolation frequency of the different fungi from 100 lesions present on the studied fruits according to the following formula (Eq. 1):

$$PC(\%) = \frac{NFI}{NTF} \times 100 \tag{1}$$

- PC is the percentage of contamination
- NFI is the number of lesions infected with a given fungal species
- NTF is the total number of lesions.

2.3.2. Yeasts study

Yeasts were isolated on YPG agar medium (Y: 1% yeast extract, P: 2% peptone, and G: 2% glucose). Three times a week for 5 weeks in June and July 2020, plastic trays each containing 500 g of strawberries were harvested during the harvest period. Each tray was placed in a sterile plastic bag at the farm and sampled at the laboratory 3 h later. Whole strawberries with calyxes (100 g) were mixed with 100 ml sterile chilled KH_2PO_4 (0.1 M) for 45 s at half speed, then 45 s at full speed. Four successive dilutions of 10 times were performed aseptically with KH_2PO_4 (0.1 M). An aliquot of 100 µl was put on Petri plates containing YPG agar supplemented with two antibiotics chloramphenicol and kanamycin at 5µg/ml and incubated at 30°C for 2 days.

Yeast colonies were counted on all Petri dishes and at all dilution levels when possible, after the longest possible incubation before colonies coalesced. Counts were expressed as the number of yeast per g of strawberries. Yeast purification was performed by at least five successive streaking on YPG agar (pH 4) incubated at 30°C. The identification was done on Wikierham medium for the assimilation (Auxanogram) and fermentation (Zymogram) tests. The identification key is the one proposed by Kreger-van Rij (1984).

2.4. Bioclimatic study of the two regions Loukkos and Gharb

A climatic study (Emberger bioclimatic quotient) was carried out to know the climate effect on strawberry fruits contamination in the two targeted regions (Gharb and Loukkos), the Emberger bioclimatic quotient (Q_2) was calculated according to the following formulas (Table 1). Climatograms were developed to identify the bioclimatic stage of the two stations studied (Loukkos and Gharb).

2.5. Statistical analyses

The statistical analyses were developed with SPSS statistical software. All data were analyzed for significant differences by applying analysis of variance test (ANOVA), with a probability of $p \leq 0.05$. Results of Coagulase-negative staphylococci and presumptive *B. cereus* strains were expressed as a percentage of positive samples.

3. Results and discussion

3.1. Bacteriological study of strawberry samples from the Loukkos and Gharb regions

Strawberry samples microbiological quality was determined by the detection of *Salmonella* spp., *L. monocytogenes*, *E. coli*, and Sulfite-reducing clostridia spores and by enumeration of *B. cereus* and Coagulase-positive staphylococci according to the International Organization for Standardization (ISO). The occurrence of *Salmonella* spp., *L. monocytogenes*, *E. coli*, and Sulfite-reducing clostridia spores is presented in Table 2.

TABLE 1 Emberger bioclimatic quotient (Q₂) formulas.

Formulas	References	Climatological parameters
Q ₂ =2000*P/ (M+m+546.4)*(M-m) Q ₂ =3.43*P/(M-m)	Mokhtari et al. (2013) Stewart (1968)	P: Annual precipitation in mm/m ² /year M: Maximum temperature of the warmest month in °C m: Minimum temperature of the coldest month in °C
Q ₂ =2000*P/(M ² -m ²)	Emberger (1942)	P: Annual precipitation in mm/m ² /year M: Maximum temperature of the warmest month in °K m: Minimum temperature of the coldest month in °K

Salmonella spp. was not detected in all samples from the two studied regions (n = 50). However, *L. monocytogenes* and *E. coli* were isolated from only two fruit samples of the Gharb region. Sulfite-reducing clostridia spores were found in two samples tested from the Gharb locality against one sample from the Loukkos area. Samples from the Gharb region have a relatively high prevalence of these microorganisms with 4 positive samples in total. Bacterial contamination level was below 100 CFU/g in both targeted areas (Gharb and Loukkos).

Table 3 shows the prevalence results of *B. cereus* and Coagulase-negative staphylococci in strawberry samples from the two targeted regions (Loukkos and Gharb).

Coagulase-positive staphylococci were negative in all samples from the two studied regions (n = 50). Therefore, the prevalence of Coagulase-negative staphylococci was reported. Coagulasenegative staphylococci were detected in 1.6 and 2.4% of strawberry samples from Loukkos and Gharb, respectively. According to Table 3, the two targeted areas samples were contaminated by *B. cereus* with occurrence percentages of 5.2 and 7.8% in Loukkos and Gharb, respectively. Samples from the Gharb region have a relatively high prevalence of *B. cereus* and Coagulase-negative staphylococci compared to those from the Loukkos region.

In this study, all samples from the Loukkos region tested negative for *Salmonella* spp., *L. monocytogenes*, and *E. coli*. However, two strawberry samples from the Gharb region were contaminated with *L. monocytogenes* and *E. coli*. Sulfitereducing clostridia spores were detected in both targeted areas, with a relatively high prevalence for the Gharb region. Coagulase-negative staphylococci and *B. cereus* were isolated from samples collected in the Gharb and Loukkos localities, samples from the Gharb region being the fruits with the highest occurrence percentage of these microorganisms. The overall results of this bacteriological analysis indicated that the bacteriological quality of the strawberry samples tested was satisfactory.

A study done by Ortiz-Solà et al. (2020) in Spain, reported that all tested samples (fresh and frozen strawberries) were negative for *Salmonella* spp., *L. monocytogenes*, and *E. coli*. These results are in accordance with our bacteriological analyses of strawberry samples from the Loukkos region. A survey conducted by Delbeke et al. (2015) in Belgium, reported that no *Salmonella* spp. or Shigatoxin-producing *E. coli* were isolated from the analyzed strawberry samples. In the same report, generic *E. coli* was present in only two of 72 strawberry samples at levels of 1.0 and 3.0 log CFU/g. Similarly, Macori et al. (2018) evaluated freshly harvested berries (Raspberries, blueberries, blackberries, and

TABLE 2 Occurrence of Salmonella spp., Listeria monocytogenes, Escherichia coli, and Sulfite-reducing clostridia spores in strawberry samples.

Samples source	N	Salmonella spp.	L. monocytogenes	E. coli	S. R. clostridia
Loukkos	25	ND	ND	ND	1
Gharb	25	ND	1	1	2

N: Number of analyzed samples. ND, not detected.

Samples source	N	Percentage of contaminated samples		
		Coagulase- negative staphylococci	Presumptive Bacillus cereus	
Loukkos	25	1.6%	5.2%	
Gharb	25	2.4%	7.8%	

TABLE 3 Prevalence (%) of presumptive *Bacillus cereus* and coagulase-negative staphylococci in strawberry fruit samples.

N: Number of analyzed samples.

TABLE 4 Prevalence of mold strains contaminating strawberry fruits in the two targeted localities Loukkos and Gharb.

Molds species	Loukkos region	Gharb region
Alternaria alternata	2	1
Aspergillus nidulans	0	1
Aspergillus niger	6	2
Botrytis cinerea	5	2
Colletotrichum acutatum	1	0
Fusarium oxysporum	0	1
Fusarium spp.	3	1
Mucor spp.	2	0
Penicillium notatum	1	0
Penicillium spp.	3	1
Rhizopus spp.	3	2

currants) and did not find any *Salmonella* spp. and Shigatoxinproducing *E. coli* in the tested samples.

Results were found by Dziedzinska et al. (2018) in Czech Republic, where detection of *L. monocytogenes* was reported as sporadic with only one positive sample among all field strawberries (0.6%) and one positive sample of purchased strawberries (1.4%). In another study, Delbeke et al. (2015) reported that *L. monocytogenes* was found in 3.8% of strawberry samples. Sulfite-reducing clostridia species are ubiquitously distributed in the environment and can be easily isolated from vegetation (Palese et al., 2009). This explains the presence of these microorganisms in two strawberry samples from the Gharb locality and one sample from the loukkos area. Oliveira et al. (2019) reported the presence of sulfite-reducing clostridia spores in berry fruits. However, blackberries had the highest prevalence of these microorganisms with 35 positive samples in total.

A study performed by Graça et al. (2017) in southern Portugal, reported the absence of coagulase-positive staphylococci in fresh-cut strawberries. Similar results were found by Oliveira et al. (2019) in berries. However, *Staphylococcus* spp. was detected in 10.1, 11.5, 44.9, and 14.4% of strawberry, blueberry, raspberry, and blackberry samples, respectively. *B. cereus* is ubiquitous and can be found in a wide range of foodstuffs, soil, raw materials, raw fruits and vegetables, raw herbs, dry foods, and processed foods (Park et al., 2018; Tango et al., 2018). In our study, *B. cereus* was

detected in samples from the two targeted localities (Loukkos and Gharb). Kim et al. (2014) reported that *B. cereus* was found in 98 strawberry samples (31.1%) out of 315 total samples analyzed in South Korea.

3.2. Mycological study of strawberry samples from the Loukkos and Gharb regions

3.2.1. Molds study

Optical microscopic examination of the different cultures allowed us to determine the fungal species using the identification keys and consultation of the fungal collections of the National School of Agriculture (ENA-Meknes). Table 4 groups the results obtained in the two localities targeted (Loukkos and Gharb) by this microbiological study.

From these preliminary results, we notice that Aspergillus niger, B. cinerea, Fusarium spp., Penicillium spp., Rhizopus spp., and Alternaria alternata species are dominant in both study areas with a priority of fungal contamination in strawberry samples from Loukkos locality. Indeed, a marked presence is noted for A. niger on strawberry samples, with six positive samples from the Loukkos region and three positive samples from the Gharb region. B. cinerea was also found on five samples from Loukkos and two samples from Gharb. Three samples from the Loukkos locality were contaminated with Fusarium spp. while only one strawberry sample from the Gharb locality was positive. The same results were reported for Penicillium spp. Moreover, the presence of Rhizopus spp. was revealed on three samples from the Loukkos region against two samples from the Gharb region. Two strawberry samples from Loukkos and one sample from Gharb were positive for the Alternaria alternate strain. However, Colletotrichum acutatum, Mucor spp., and Penicillium notatum were isolated only from the Loukkos samples. While Aspergillus nidulans and Fusarium oxysporum species were found in the samples from the Gharb region. In general, strawberry samples from the Loukkos area have a relatively high prevalence of these microorganisms with 26 positive samples in total against 11 positive samples in the Gharb area.

According to the molds study, the most common fungi found in strawberry samples from both regions (Loukkos and Gharb) were *A. niger*, *B. cinerea*, *Fusarium* spp., *Penicillium* spp., *Rhizopus* spp., and *A. alternata* with a significant predominance in samples from the Loukkos region. In a study done by Saleh and Al-Thani (2019), samples of cucumber, tomato, strawberry, and orange were tested to identify spoilage agents. Various fungal types were isolated from strawberry samples, including 5 *Aspergillus* spp., 8 *Rhizopus* spp., 3 *Penicillium* spp., and 5 *Botrytis* spp. In the same report, results indicate that strawberry samples contain the greatest fungi variety of all the fruits. *B. cinerea* causes gray mold, the most important disease of strawberries harvested worldwide (Petrasch et al., 2019; Weber and Hahn, 2019). *A. niger* is by far the most common Aspergillus species responsible for the post-harvest rot of fresh fruit, including apples, pears, peaches, citrus fruits, grapes, and strawberries (Pitt and Hocking, 2009).

Strawberries are fruits closer to the ground and therefore are more susceptible to soft rot mainly Rhizopus spp. and Mucor spp. (Feliziani and Romanazzi, 2016). In a study conducted by Agyare et al. (2020) in the United Kingdom, Mucor piriformis and Rhizopus spp. were isolated from decaying strawberry fruit. Tournas and Katsoudas (2005) revealed the presence of Penicillium spp. in all types of berries tested (strawberries, blackberries, and raspberries). Penicillium expansum and some other species of the genus Penicillium can cause significant post-harvest losses in strawberries (Feliziani and Romanazzi, 2016). A study carried out by Mouden et al. (2013) marked the presence of B. cinerea on strawberry samples, with a percentage of contamination ranging from 48.4 to 58.1%. In the same report, an attack by A. alternata was revealed on the samples analyzed with a percentage of contamination ranging from 19.8 to 23.4% depending on the varieties studied. A study done by El-araby et al. (2022) reported that the fungal strain A. niger was isolated from spoiled strawberries.

3.2.2. Yeasts study

The analysis of the genus (nitrate assimilation, mycelium form, and sporulation) and the assimilation (Auxanogram) and fermentation (Zymogram) tests of the yeasts (Tables 5, 6) allowed us to establish Table 7.

In our study, the prevalence of yeast strains contaminating strawberry fruits from the Loukkos and Gharb localities was negligible, with a priority on fungal contamination for strawberries from the loukkos. Indeed, a marked presence is noted for *Candida sake* in strawberry samples, with four positive samples from the Loukkos region and one positive sample from the Gharb region. While *Rhodotorula glutinis* was found in only three samples from the Loukkos.

A study by Tournas and Katsoudas (2005) reported the presence of yeasts in only about 5% of raspberries and 3% of strawberry samples. The lower incidence of yeasts may be partly explained by the fact that these organisms cannot penetrate the epidermis of the fruit and infect the internal tissues. In addition, yeasts are probably more sensitive to chlorine and are therefore inactivated or washed out during the disinfection step. Isolation and identification of *Rhodotorula* spp. and *Cryptococcus laurentii* from strawberries could be considered in light of bio-control of spoilage molds (*Rhizopus* spp. and *B. cinerea*; Campaniello et al., 2008). Similar results were reported by Zhang et al. (2007) for *Cryptococcus laurentii* against *Rhizopus* spp. in strawberry fruit.

A study performed by Zhang et al. (2007) reported that *R. glutinis* was isolated from the surfaces of strawberries harvested in unsprayed orchards. Another report mentioned the presence of *R. glutinis* and *Debaryomyces melissophilus* in packaged strawberries of the Belgian variety Elsanta (Ragaert et al., 2006). The most frequently isolated yeasts on strawberries from conventional and organic growers belonged to the genus Candida, Cryptococcus, and Rhodotorula (Jensen et al., 2013). *Geotrichum*

candidum was isolated from diseased strawberries by Ma et al. (2018).

3.3. Bioclimatic study of the two regions Loukkos and Gharb

3.3.1. Average temperature and precipitation

In Gharb, summers are hot, arid, and generally clear and winters are cool and partly cloudy. Over the course of the year, the temperature generally varies between 8°C and 32°C and is rarely below 4°C or above 37°C. The very hot season in Gharb lasts for 3 months, from June 20th to September 20th, with an average daily maximum temperature above 29°C. The hottest month of the year in Gharb is August, with an average maximum temperature of 31°C and a minimum temperature of 20°C. The cool season in Gharb region lasts 3.5 months, from November 24th to March 7th, with an average daily maximum temperature below 20°C. The coldest month of the year in Gharb is January, with an average minimum temperature of 8°C and a maximum of 18°C.

In Loukkos, summers are hot, humid, and generally clear and winters are long, cool, wet, windy and partly cloudy. During the year, the temperature generally varies between 9°C and 29°C and it is rarely below 5°C or above 36°C. The very hot season in Loukkos region lasts 3 months, from June 22nd to September 22nd, with an average daily maximum temperature of over 27°C. The hottest month of the year in Loukkos is August, with an average maximum temperature of 29°C and a minimum of 20°C. The cool season in Loukkos lasts 3.8 months, from November 25 to March 17, with an average daily maximum temperature of <19°C. The coldest month of the year in Loukkos is January, with an average minimum temperature of 9°C and a maximum of 17°C.

In the Gharb region, the rainy period of the year lasts 8 months, from September 20th to May 21st, with rainfall of at least 13 mm over a sliding period of 31 days. The rainiest month in Gharb is November, with an average rainfall of 65 mm. The dry period of the year lasts 4 months, from May 21st to September 20th. The least rainy month in Gharb is July, with an average rainfall of 1 mm (Figure 2).

In the Loukkos region, the rainy period of the year lasts 8.4 months, from September 14th to May 27th, with rainfall of at least 13 mm over a sliding period of 31 days. The rainiest month in Loukkos is November, with an average rainfall of 94 mm. The dry period of the year lasts 3.6 months, from May 27th to September 14th. The least rainy month in Loukkos is July, with an average rainfall of 1 mm.

3.3.2. Emberger bioclimatic quotient

The Emberger bioclimatic quotient (Q_2) was calculated according to the three formulas mentioned above (Table 1). Table 8 represents the Q_2 values calculated in the two regions Gharb and Loukkos.

The climatic parameters allowed us to locate the two target regions Gharb and Loukkos on the Emberger diagram (Figure 3). According to the latter, the Gharb region (Rabat-Sale-Kenitra) is

Assimilation tests	Yeast strains				
	C. sake	Cr. albidus	G. candidum	P. guilliermondii	R. glutinis
Glucose	+	+	+	+	+
Inulin	_	_	_	+	_
Sucrose	+	+	_	+	_
Raffinose	_	+	_	+	_
Melibiose	_	_	_	+	_
Galactose	+	_	_	+	_
Lactose	_	_	_	-	_
Trehalose	+	+	-	+	_
Maltose	_	+	-	+	_
Melezitose	_	+	_	+	_
Methyl α-D-glucoside	_	_	-	+	_
Soluble starch	_	+	-	_	_
Cellobiose	_	+	-	+	-
Salicin	_	+	-	+	_
L-sorbose	_	_	_	_	_
L-rhamnose	_	_	_	_	_
D-xylose	_	+	+	+	-
L-arabinose	_	+	-	+	-
D-arabinose	_	+	-	+	_
D-ribose	_	_	_	+	_
Ethanol	_	+	+	+	_
Glycerol	_	_	+	+	+
Erythritol	_	_	_	_	_
Ribitol	_	_	_	+	+
Galactitol	_	_	_	_	_
D-mannitol	_	+	_	+	+
D-glucitol	_	+	_	+	_
Myo-inositol	_	+	_	_	_
DL-lactate	_	+	-	+	+
Succinate	+	+	-	+	-
Citrate	_	+	_	_	_
D-gluconate	_	+	-	-	-
D-glucosamine	_	_	-	+	-
Nitrate	_	+	-	-	_
Vitamin-free	-	_	_	_	-

TABLE 5 Auxanogram of yeast strains contaminating strawberry fruits from the two localities Loukkos and Gharb.

located on the semi-arid stage. While, the Loukkos (Tangier-Tetouan-Al Hoceima) is located on the sub-humid stage. In Gharb, the climate is clearly Mediterranean in the inland cities and Mediterranean with an oceanic influence in the coastal cities. In Loukkos, the climate is Mediterranean in the coasts and surroundings, rather continental and with periods of heavy snowfall in the inland cities of the region. Bioclimatic indices have a strong influence on plant growth as well as its contamination, which explains the differential effect on the prevalence of bacteria in strawberry samples from Gharb (Tables 2, 3) and on the prevalence of fungi in strawberry samples from Loukkos (Tables 4, 5).

Strawberries are perishable fruits that, during handling, can be attacked by certain microorganisms, especially bacteria. Also,

Fermentation tests	Yeast strains				
	C. sake	Cr. albidus	G. candidum	P. guilliermondii	R. glutinis
Glucose	+-	-	+	+	-
Galactose	+	_	_	_	-
Sucrose	_	_	_	_	-
Maltose	_	_	_	_	_
Lactose	_	_	_	_	_
Raffinose	_	_	_	_	-
Trehalose	_	_	_	_	_

TABLE 6 Zymogram of yeast strains contaminating strawberry fruits from the two localities Loukkos and Gharb.

TABLE 7 Prevalence of yeast strains contaminating strawberry fruits from the two localities Loukkos and Gharb.

Yeasts species	Loukkos region	Gharb region
Candida sake	4	1
Cryptococcus albidus	1	0
Geotrichum candidum	0	1
Pichia guilliermondii	1	1
Rhodotorula glutinis	3	0

TABLE 8 Emberger bioclimatic quotient $(\ensuremath{\mathsf{Q}}_2)$ of the two regions Gharb and Loukkos.

	Gharb	Loukkos
Q ₂ Mokhtari et al. (2013)	102.91	171.70
Q2 Stewart (1968)	103.15	171.50
Q ₂ Emberger (1942)	102.89	171.69
Means	102.98	171.63



the main causes of strawberry diseases during storage and shelf life are the development of rots caused by a series of fungi. Climatic conditions were suspected to have a differential effect on the prevalence of bacteria in the Gharb strawberry samples and on the prevalence of fungi in the Loukkos strawberry samples. A climatological study was necessary to explain this regional distribution. Analysis of the association between microbial indicators and climatology demonstrates the plausibility of the idea that climatology can affect foodborne pathogens pre-harvest. Climatic conditions are an important factor to consider regarding microbial contamination of fresh products (Machado-Moreira et al., 2019). It is known that changes in climatic conditions determine the development of strawberry plants, affecting



flowering, fruiting, and fruit quality, among other characteristics (Khammayom et al., 2022). However, it is pertinent to mention that the influence of climatic conditions on plant contamination may be specific to each microorganism/plant/climatic conditions combination, meaning that no clear trend can be established (Ward et al., 2015). Climatic conditions during the flowering and ripening of strawberry fruit influence fruit quality and microbial contamination as well as affect shelf life more than the irrigation method used (Parikka et al., 2009).

Climatic conditions, including precipitation, humidity, temperature, wind, and orchard management activities, have previously been shown to be correlated with the risk of postharvest fungal disease development during apple storage (Dutot et al., 2013; Bui et al., 2021). Studies on plant pathogens have shown that high humidity and moderate temperatures promote fungal growth (Griffin, 1996; Rowan et al., 1999). *B. cinerea* infections are correlated positively with relative humidity and negatively with temperature (Feliziani and Romanazzi, 2016). Weather factors, particularly humidity and temperature, had a direct influence on fungal abundance and richness in the air and on vegetation leaf surfaces, showing large differences among habitats (Talley et al., 2002).

According to the bacteriological and mycological study, there is a difference in the contamination level of strawberry fruits according to the locality of origin. The Gharb region shows a prevalence of bacterial contamination and the Loukkos region reveals a prevalence of fungal contamination. The climatological analyses carried out confirm this hypothesis. Evidently, post-harvest management determines food quality and safety, market competitiveness, and producer profits. Temperature and humidity management is the most important factor to control in order to minimize spoilage of strawberry fruit and maximize its post-harvest life. This management must take into consideration the climatic conditions of the cultivated areas.

4. Conclusion

Pre-harvest weather conditions affect strawberries' quality and susceptibility to post-harvest pathogen attacks. The objective of this survey was to study the microbiological quality and the prevalence of major foodborne pathogens on strawberry fruits in the Gharb and Loukkos regions of Morocco. Thus, to establish a correlation between the postharvest microbial load and the pre-harvest bioclimatic conditions. The Gharb region has a prevalence of fungal contamination, while the Loukkos region has a prevalence of bacterial contamination. The climatological study confirms this hypothesis. Post-harvest storage and conditioning of strawberries must take into consideration the climatology of the growing regions to improve food safety practices for a good strawberry agricultural system.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work, and approved it for publication.

Funding

This work was supported by the PRIMA project (Enhancing Mediterranean Fresh Produce Shelf-Life using Sustainable Preservative Technologies and Communicating Knowledge on Dynamic Shelf-Life using Food Cloud Services and Predictive Modeling BIOFRESHCLOUD).

Acknowledgments

The authors are indebted to "Sidi Mohamed Ben Abdellah" University of Fez, Morocco. AE-a thanks all those who directly or indirectly contributed to this research.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

Agyare, S., Magan, N., and Xu, X. (2020). Conditions for infection of strawberry fruit by M. piriformis and *Rhizopus* spp. *Eur. J. Plant Pathol.* 157, 65–75. doi: 10.1007/s10658-020-01981-3

Benkirane, R., (1995). Contribution à l'étude des Maladies du riz au Maroc. Cas de la Pyriculariose due à Pyricularia Oryzae. Thèse de 3ème Cycle. Université Ibn Tofail. Faculté des Sciences, Kénitra, Maroc, 145.

Bui, T. A. T., Stridh, H., and Molin, M. (2021). Influence of weather conditions on the quality of 'Ingrid Marie' apples and their susceptibility to grey mould infection. *J. Agric. Food Res.* 3:100104. doi: 10.1016/j.jafr.2021.100104

Campaniello, D., Bevilacqua, A., Sinigaglia, M., and Corbo, M. R. (2008). Chitosan: antimicrobial activity and potential applications for preserving minimally processed strawberries. *Food Microbiol.* 25, 992–1000. doi: 10.1016/j.fm.2008.06.009

Cárdenas, C., Molina, K., Heredia, N., and García, S. (2013). Evaluation of microbial contamination of tomatoes and peppers at retail Markets in Monterrey, Mexico. *J. Food Prot.* 76, 1475–1479. doi: 10.4315/0362-028X.JFP-12-561

Champion, R., (1997). Identifier les champignons transmis par les semences. INRA, Paris, 398.

Chidambaram, P., Mathur, S., and Neergaard, P. (1974). "Identification of seedborne Drechslera species," in Handbook on Seed Health Testing. (Norway: International Seed Testing Association), 165–168.

de Jesus Filho, M., Scolforo, C. Z., Saraiva, S. H., Pinheiro, C. J. G., Silva, P. I., and Della Lucia, S. M. (2018). Physicochemical, microbiological and sensory acceptance alterations of strawberries caused by gamma radiation and storage time. *Sci. Hortic.* 238, 187–194. doi: 10.1016/j.scienta.2018.04.053

Delbeke, S., Ceuppens, S., Hessel, C. T., Castro, I., Jacxsens, L., De Zutter, L., et al. (2015). Microbial safety and sanitary quality of strawberry primary production in Belgium: risk factors for *Salmonella* and Shiga toxin-producing *Escherichia coli* contamination. *Appl. Environ. Microbiol.* 81, 2562–2570. doi: 10.1128/AEM.03930-14

Domsch, K. H., Gams, W., and Anderson, T. H., (1980). Compendium of Soil Fungi, Volume 1. Academic Press, London, 859.

Dutot, M., Nelson, L. M., and Tyson, R. C. (2013). Predicting the spread of postharvest disease in stored fruit, with application to apples. *Postharvest Biol. Technol.* 85, 45–56. doi: 10.1016/j.postharvbio.2013.04.003

Dziedzinska, R., Vasickova, P., Hrdy, J., Slany, M., Babak, V., and Moravkova, M. (2018). Foodborne bacterial, viral, and protozoan pathogens in field and market strawberries and environment of strawberry farms. *J. Food Sci.* 83, 3069–3075. doi: 10.1111/1750-3841.14401

El-araby, A., El Ghadraoui, L., and Errachidi, F. (2022). Usage of biological chitosan against the contamination of post-harvest treatment of strawberries by *Aspergillus niger. Front. Sustain. Food Syst.* 6:881434. doi: 10.3389/fsufs.2022.881434

Ellis, M.B., (1971). *Dematiaceous Hyphomycetes*. Commonwealth Mycological Institute Kew, Surrey, England, 608.

Emberger, L. (1942). A project to classify climates from a phytogeographical point of view. *Bull. Soc. d'histoire Nat. Toulouse* 77, 97–124.

Evêncio-Luz, L., Lima-Filho, J. V., and Evêncio-Neto, J. (2012). Occurrence of *Salmonella* sp. and coagulase-positive staphylococci in raw eggs and Coalho cheese: comparative study between two cities of Brazil's northeast. *Braz. J. Microbiol.* 43, 1463–1466. doi: 10.1590/S1517-83822012000400030

Feliziani, E., and Romanazzi, G. (2016). Postharvest decay of strawberry fruit: etiology, epidemiology, and disease management. *J. Berry Res.* 6, 47–63. doi: 10.3233/JBR-150113

Gilman, C. J., (1957). A Manual of Soil Fungi, 2nd Edn. The Iowa State College Press, Ames, Iowa, USA, 183.

Graça, A., Esteves, E., Nunes, C., Abadias, M., and Quintas, C. (2017). Microbiological quality and safety of minimally processed fruits in the marketplace of southern Portugal. *Food Control* 73, 775–783. doi: 10.1016/j. foodcont.2016.09.046

Griffin, D. H. (1996). Fungal Physiology. New York: John Wiley & Sons.

IPQ (1986). Food microbiology. General rules for identifying reducing clostridium sulphite spores. *Port. Stand. NP.* 2262:1986.

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

ISO (1996). Microbiology of Food and Animal Feeding Stuffs - Horizontal Method for the Detection and Enumeration of *Listeria monocytogenes* - Part 1: Detection Method. Amendment 1: Modification of the Isolation Media and the Haemolysis Test, and Inclusion of Precision Data. Int. Stand. ISO 11290-1:1996/AMD 1:2004.

ISO (1998). Microbiology of Food and Animal Feeding Stuffs - Horizontal Method for the Detection and Enumeration of *Listeria monocytogenes* - Part 2: Enumeration Method. Amendment 1: Modification of the Enumeration medium. Int. Stand. ISO 11290-2: 1998/AMD 1:2004.

ISO (1999). Microbiology of Food and Animal Feeding Stuffs - Horizontal Method for the Enumeration of Coagulase-Positive Staphylococci (*Staphylococcus aureus* and Other Species) - Part 1: Technique Using Baird-Parker Agar Medium. Int. Stand. ISO 6888-1: 1999.

ISO (2001). Microbiology of Food and Animal Feeding Stuffs - Horizontal Method for the Enumeration of *beta-glucuronidasepositive Escherichia coli*-Part 2: Colony-count technique at 44 °C using 5-bromo-4-chloro-3-indolyl beta-D-glucuronide. Zagreb: Croatian Standards Institute. ISO 16649-2:2001.

ISO (2002). Microbiology of Food and Animal Feeding Stuffs - Horizontal Method for the Detection of *Salmonella* spp. Int. Stand. ISO Norm 6579:2002.

ISO (2004). Microbiology of Food and Animal Feeding Stuffs-Horizontal Method for the Enumeration of Presumtive Bacillus Cereus-Colony-Count Technique at 30° C. Int. Stand. ISO 7932:2004.

Jensen, B., Knudsen, I. M. B., Andersen, B., Nielsen, K. F., Thrane, U., Jensen, D. F., et al. (2013). Characterization of microbial communities and fungal metabolites on field grown strawberries from organic and conventional production. *Int. J. Food Microbiol.* 160, 313–322. doi: 10.1016/j.ijfoodmicro.2012.11.005

Khammayom, N., Maruyama, N., and Chaichana, C. (2022). The effect of climatic parameters on strawberry production in a small walk-in greenhouse. *Agric. Eng.* 4:1, 104–121. doi: 10.3390/agriengineering4010007

Kim, W. I., Jo, A. R., Kim, S. R., Ryu, S. H., Nam, K. W., Yoon, Y., et al. (2014). Microbial contamination levels of strawberries at domestic farms of South Korea. *Korean J. Soil Sc. Fertilizer* 47, 437–442. doi: 10.7745/KJSSE2014.47.6.437

Kreger-van Rij, N. J. W., (1984). Yeasts. Amsterdam: Elsevier.

Ma, W., Zhang, Y., Wang, C., Liu, S., and Liao, X. (2018). A new disease of strawberry, fruit rot, caused by *Geotrichum candidum* in China. *Plant Prot. Sci.* 54, 92–100. doi: 10.17221/76/2017-PPS

Machado-Moreira, B., Richards, K., Brennan, F., Abram, F., and Burgess, C. M. (2019). Microbial contamination of fresh produce: what, where, and how? *Compr. Rev. Food Sci. Food Saf.* 18, 1727–1750. doi: 10.1111/1541-4337.12487

Macori, G., Gilardi, G., Bellio, A., Bianchi, D., Gallina, S., Vitale, N., et al. (2018). Microbiological parameters in the primary production of berries: a pilot study. *Foods* 7:105. doi: 10.3390/foods7070105

Matar, C., Guillard, V., Gauche, K., Costa, S., Gontard, N., Guilbert, S., et al. (2020). Consumer behaviour in the prediction of postharvest losses reduction for fresh strawberries packed in modified atmosphere packaging. *Postharvest Biol. Technol.* 163:111119. doi: 10.1016/j.postharvbio.2020.111119

Mohammadi, L., Ramezanian, A., Tanaka, F., and Tanaka, F. (2021). Impact of Aloe vera gel coating enriched with basil (*Ocimum basilicum L.*) essential oil on postharvest quality of strawberry fruit. *J. Food Meas. Characterization* 15, 353–362. doi: 10.1007/s11694-020-00634-7

Mokhtari, N., Mrabet, R., Lebailly, P., and Bock, L. (2013). Spatialization of bioclimates, aridity and vegetation levels of Morocco. *Revue Marocaine Sci.* Agronomiques et Vétérinaires 2, 50–66.

Mouden, N., Benkirane, R., Touhami, A. O., and Douira, A. (2013). Mycoflore de quelques variétés du fraisier (*Fragaria ananassa* L.), cultivées dans la région du Gharb et le Loukkos (Maroc). *J. Appl. Biosci.* 61, 4490–4514. doi: 10.4314/jab.v61i0.85597

Nakata, Y., and Izumi, H. (2020). Microbiological and quality responses of strawberry fruit to high CO2 controlled atmosphere and modified atmosphere storage. *HortScience* 55, 386–391. doi: 10.21273/HORTSCI14771-19

Oliveira, M., Rodrigues, C. M., and Teixeira, P. (2019). Microbiological quality of raw berries and their products: a focus on foodborne pathogens. *Heliyon* 5:e02992. doi: 10.1016/j.heliyon.2019.e02992

Ortiz-Solà, J., Viñas, I., Colás-Medà, P., Anguera, M., and Abadias, M. (2020). Occurrence of selected viral and bacterial pathogens and microbiological quality of fresh and frozen strawberries sold in Spain. *Int. J. Food Microbiol.* 314:108392. doi: 10.1016/j.ijfoodmicro.2019.108392

Palese, A. M., Pasquale, V., Celano, G., Figliuolo, G., Masi, S., and Xiloyannis, C. (2009). Irrigation of olive groves in southern Italy with treated municipal wastewater: effects on microbiological quality of soil and fruits. *Agric. Ecosyst. Environ.* 129, 43–51. doi: 10.1016/j.agee.2008.07.003

Parikka, P., Kivijärvi, P., Prokkola, S., and Kemppainen, R. (2009). Microbiological quality of organic strawberry fruit. *Acta Hortic.* 842, 377–380. doi: 10.17660/ActaHortic.2009.842.72

Park, K. M., Jeong, M., Park, K. J., and Koo, M. (2018). Prevalence, enterotoxin genes, and antibiotic resistance of *Bacillus cereus* isolated from raw vegetables in Korea. *J. Food Prot.* 81, 1590–1597. doi: 10.4315/0362-028X.JFP-18-205

Petrasch, S., Knapp, S. J., van Kan, J. A. L., and Blanco-Ulate, B. (2019). Grey mould of strawberry, a devastating disease caused by the ubiquitous necrotrophic fungal pathogen *Botrytis cinerea*. *Mol. Plant Pathol*. 20, 877–892. doi: 10.1111/mpp.12794

Pitt, J. I., and Hocking, A. D., (2009). Fungi and Food Spoilage. Springer USA.

Ponchet, A. (1966). Etude des contaminations mycopéricarpiques du caryopse du blé. Crop Res. 7, 554-460.

Ragaert, P., Devlieghere, F., Loos, S., Dewulf, J., Van Langenhove, H., and Debevere, J. (2006). Metabolite production of yeasts on a strawberry-agar during storage at 7°C in air and low oxygen atmosphere. *Food Microbiol.* 23, 154–161. doi: 10.1016/j.fm.2005.02.002

Rowan, N. J., Johnstone, C. M., McLean, R. C., Anderson, J. G., and Clarke, J. A. (1999). Prediction of toxigenic fungal growth in buildings by using a novel modelling system. *Appl. Environ. Microbiol.* 65, 4814–4821. doi: 10.1128/AEM.65.11.4814-4821.1999

Saleh, I., and Al-Thani, R. (2019). Fungal food spoilage of supermarkets' displayed fruits. Vet. World 12, 1877-1883. doi: 10.14202/vetworld.2019.1877-1883

Shu, P., Min, D., Ai, W., Li, J., Zhou, J., Li, Z., et al. (2020). L-arginine treatment attenuates postharvest decay and maintains quality of strawberry fruit by promoting nitric oxide synthase pathway. *Postharvest Biol. Technol.* 168:111253. doi: 10.1016/j. postharvbio.2020.111253

Singh, D., and Sharma, R. R. (2018). "Postharvest diseases of fruits and vegetables and their management" in *Postharvest Disinfection of Fruits and Vegetables* (Elsevier), 1–52.

Steinka, I., and Kukułowicz, A. (2018). Identification and study of the behavior of S. *aureus* and S. *epidermidis* in fresh and frozen strawberries. *Technol. Żywności.* Żywienia 17, 27–35. doi: 10.17306/J.AFS.2018.0535

Stewart, P. (1968). Rainfall quotient and biospheric degradation. Bull. Soc. d'histoire Nat. d'Afrique Nord 59, 23-36.

Tahir, H. E., Xiaobo, Z., Jiyong, S., Mahunu, G. K., Zhai, X., and Mariod, A. A. (2018). Quality and postharvest-shelf life of cold-stored strawberry fruit as affected by gum Arabic (Acacia Senegal) edible coating. *J. Food Biochem.* 42:e12527. doi: 10.1111/jfbc.12527

Talley, S. M., Coley, P. D., and Kursar, T. A. (2002). The effects of weather on fungal abundance and richness among 25 communities in the intermountain west. *BMC Ecol.* 2:7. doi: 10.1186/1472-6785-2-7

Tango, C. N., Wei, S., Khan, I., Hussain, M. S., Kounkeu, P.-F. N., Park, J., et al. (2018). Microbiological quality and safety of fresh fruits and vegetables at retail levels in Korea. *J. Food Sci.* 83, 386–392. doi: 10.1111/1750-3841.13992

Tarr, S., (1962). *Diseases of Sorghum, Sudan Grass and Broom Corn*. CAB, the Commonwealth Mycological Institute. Kew, 380p.

Tournas, V. H., and Katsoudas, E. (2005). Mould and yeast flora in fresh berries, grapes and citrus fruits. *Int. J. Food Microbiol.* 105, 11–17. doi: 10.1016/j. ijfoodmicro.2005.05.002

Trinetta, V., McDaniel, A., Batziakas, K. G., Yucel, U. K., Nwadike, L., and Pliakoni, E. (2020). Antifungal packaging film to maintain quality and control postharvest diseases in strawberries. *Antibiotics* 9:Art. 9. doi: 10.3390/antibiotics9090618

Uyttendaele, M. (2014). Microbiological safety of strawberries and lettuce for domestic consumption in Egypt. *J. Food Process. Technol.* 5:308. doi: 10.4172/2157-7110.1000308

Ward, M., Dhingra, R., Remais, J. V., Chang, H. H., Johnston, L. M., Jaykus, L.-A., et al. (2015). Associations between weather and microbial load on fresh produce prior to harvest. *J. Food Prot.* 78, 849–854. doi: 10.4315/0362-028X. JFP-14-381

Weber, R. W. S., and Hahn, M. (2019). Grey mould disease of strawberry in northern Germany: causal agents, fungicide resistance and management strategies. *Appl. Microbiol. Biotechnol.* 103, 1589–1597. doi: 10.1007/s00253-018-09590-1

Yoon, Y., Kim, K., Nam, M., Shim, W. B., Ryu, J. G., Kim, D. H., et al. (2010). Microbiological assessment in strawberry production and recommendations to establish a good agricultural practice system. *Foodborne Pathog. Dis.* 7, 1511–1519. doi: 10.1089/fpd.2010.0611

Zhang, H., Wang, L., Dong, Y., Jiang, S., Cao, J., and Meng, R. (2007). Postharvest biological control of gray mold decay of strawberry with *Rhodotorula glutinis*. *Biol. Control* 40, 287–292. doi: 10.1016/j.biocontrol.2006.10.008