



OPEN ACCESS

EDITED AND REVIEWED BY
Axl Cloeckaert,
Institut National de recherche pour
l'agriculture, l'alimentation et l'environnement
(INRAE), France

*CORRESPONDENCE

Jinlin Li
✉ jinlin.li@imbim.uu.se

RECEIVED 29 May 2025
ACCEPTED 30 May 2025
PUBLISHED 17 June 2025

CITATION

Yadavalli T, Awasthi S and Li J (2025) Editorial:
Herpesvirus: transmission, pathogenesis,
host-pathogen interaction, and treatment.
Front. Microbiol. 16:1637344.
doi: 10.3389/fmicb.2025.1637344

COPYRIGHT

© 2025 Yadavalli, Awasthi and Li. This is an
open-access article distributed under the
terms of the [Creative Commons Attribution
License \(CC BY\)](#). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that the
original publication in this journal is cited, in
accordance with accepted academic practice.
No use, distribution or reproduction is
permitted which does not comply with these
terms.

Editorial: Herpesvirus: transmission, pathogenesis, host-pathogen interaction, and treatment

Tejabhram Yadavalli¹, Sita Awasthi² and Jinlin Li^{3*}

¹Department of Ophthalmology and Visual Sciences, University of Illinois Chicago, Chicago, IL, United States, ²Infectious Disease Division, Department of Medicine, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, United States, ³Department of Medical Biochemistry and Microbiology, Zoonosis Science Center, Uppsala University, Uppsala, Sweden

KEYWORDS

EBV, HSV-1, BHV 1, HSV-2, CMV, HHV-7, pulmonary infection

Editorial on the Research Topic

Herpesvirus: transmission, pathogenesis, host-pathogen interaction, and treatment

Herpesviruses are a large group of viruses including the members that can infect humans, bovines, birds, and a wide range of other hosts. These viruses have large double-stranded DNA genomes and express a large of envelope, tegument, and capsid proteins. At least five of the nine known human herpesviruses are particularly widespread, including Epstein-Barr virus (EBV), Herpes simplex virus 1 and 2 (HSV-1 and HSV-2), Varicella zoster virus (VZV), and Human cytomegalovirus (HCMV) (Grinde, 2013). In addition to human herpesviruses, one of the most important viruses linked to cattle diseases is Bovine herpesvirus 1 (BHV1) (Nandi et al., 2009). This Research Topic explores the transmission, pathogenesis, host-pathogen interactions, and treatment of herpesviruses.

Human herpesvirus reactivation remains an extremely interesting and complex topic of research. The EBV trans-activator, ZEBRA (also known as Zta, Z, EB1, encoded by EBV gene BZLF1) was discovered 40 years ago (Countryman and Miller, 1985), and known to play an important role in regulating EBV reactivation. In this Research Topic, Wang et al. comprehensively summarized the key factors that regulate the reactivation of EBV at the different stages of the lytic cycle including the immediate-early (IE) genes expression, the DNA replication, and progeny virus production. They discussed the regulation of the viral immediate-early proteins ZEBRA and Rta at the transcriptional levels by various activators, repressors, and epigenetic modifications. Additionally, they pointed out the essential roles of the host partners and modifications in remodeling ZEBRA and Rta activities. The authors summarized the regulation of the EBV lytic cycle after the initiation by different EBV-encoded proteins, viral miRNAs, and host factors, and discussed the emerging tools and technologies, such as structural prediction tools (AlphaFold3), literature data mining tools [Natural Language Processing (NLP)] and genome editing tools (CRISPRx, derived from CRISPR systems) for advancing our understanding of mechanisms of latency and reactivation.

The research article from Ripa et al. investigated the interaction between HSV-1 and autophagy in glial cells and found that HSV-1 inhibited the formation of autophagosomes. Knocking out the ATG5 gene in the HOG and U-87 MG cell lines using the CRISPR/Cas9

led to the suppression of HSV-1 transcription and replication. No significant differences in viral production were observed when knocking out the MAP1LC3B gene. Based on these results, Ripa et al. proposed that HSV-1 hijacks non-canonical functions of certain components of the autophagic machinery, such as ATG-5, to facilitate its replication by inhibiting the complete autophagy in glial cells. Shi et al. established an HSV-2 infection model using HaCaT cells and evaluated the potential role of ACV in treating HSV-2 infection by enhancing the host antiviral immune response through regulation of the TLR9 signaling pathway.

Another interesting topic related to herpesviruses and pulmonary infections was investigated by Liu et al. By employing metagenomic next-generation sequencing (mNGS) of 100 respiratory samples from patients with pulmonary infection, they detected a total of 43 bacterial species, 12 fungal species, and 5 viral species. EBV, *Candida albicans*, and *Haemophilus parainfluenzae* are the most frequently detected viruses, fungi, and bacteria, respectively. It is worth noting that herpesviruses were the only DNA viruses detected. The average hospitalization duration was significantly longer for patients who tested positive for herpesvirus compared to those who tested negative. The patients who tested positive for viral infection were more likely to have co-infections with other pathogens, with *Pneumocystis jirovecii* and *Aspergillus fumigatus* being the most commonly identified.

Significant disparities in microbial community composition were observed between the virus-positive and virus-negative groups. Through the analysis of the correlation between herpesvirus and high abundance species, a distinct positive correlation was observed between *Haemophilus parainfluenzae* and three herpesviruses: EBV, CMV, and HHV-7. This study pointed out the important association of active herpesvirus and pulmonary infections.

In addition to the human herpesviruses, in this Research Topic, Yan et al. studied the potential role of *Serratia marcescens* on BHV1 infection. *Serratia marcescens* is a Gram-negative bacterium frequently found in a wide range of environments and commonly co-infected with BHV1. Yan et al. assessed the effects of recombinant serralyisin-like protease D (rSPD) on BHV1 infection. Serralyisin-like protease D is an extracellular enzyme secreted by *Serratia marcescens*. They found that rSPD significantly promoted BHV1 production in Madin-Darby bovine kidney (MDBK) cells. Furthermore, the transcriptomic analysis showed that rSPD curbs innate immune responses, evidenced by the downregulation of innate immunity-associated genes, such as *ISG15*, *OAS2*, *IFIT1*, *IFIT2*, *IFIT3*, *MX1*, *RSAD2*, *MX2*, *SAA3*, *DDX58*, *IFI44*, and *IRF1*. In addition, rSPD was found to upregulate the genes associated with

inflammatory response, including *IL-6*, *IL-8*, *CCL2*, *CX3CL1*, *CCL3*, and *CXCL3* which may increase cell damage. Based on these results, Yan et al. propose that rSPD may enhance BHV-1 replication by suppressing the expression of antiviral genes and promoting viral spread through upregulated inflammatory responses.

In summary, the articles in this Research Topic will enhance our understanding of herpesviruses and associated diseases.

Author contributions

TY: Writing – review & editing. SA: Writing – review & editing. JL: Writing – review & editing, Writing – original draft.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. TY was supported by NEI (R01EY034508) and JL was supported by Åke Wiberg Stiftelse grant (M20–0130).

Acknowledgments

We thank all the contributors of this Research Topic and appreciate the efforts of the reviewers.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Countryman, J., and Miller, G. (1985). Activation of expression of latent epstein-barr herpesvirus after gene transfer with a small cloned subfragment of heterogeneous viral DNA. *Proc. Natl. Acad. Sci. USA* 82, 4085–4089. doi: 10.1073/pnas.82.12.4085
- Grinde, B. (2013). Herpesviruses: latency and reactivation - viral strategies and host response. *J. Oral Microbiol.* 5:22766. doi: 10.3402/jom.v5i0.22766
- Nandi, S., Kumar, M., Manohar, M., and Chauhan, R. S. (2009). Bovine herpes virus infections in cattle. *Anim. Health Res. Rev.* 10, 85–98. doi: 10.1017/S1466252309990028