



# Contribution of Mitochondrial DNA Variation to Chronic Disease in East Asian Populations

Dayan Sun<sup>1,2</sup>, Yang Wei<sup>1,2</sup>, Hong-Xiang Zheng<sup>3</sup>, Li Jin<sup>1,2\*</sup> and Jiucun Wang<sup>1,2\*</sup>

<sup>1</sup> State Key Laboratory of Genetic Engineering and Collaborative Innovation Center for Genetics and Development, School of Life Sciences, Fudan University, Shanghai, China, <sup>2</sup> Human Phenome Institute, Fudan University, Shanghai, China, <sup>3</sup> Ministry of Education Key Laboratory of Contemporary Anthropology, Department of Anthropology and Human Genetics, School of Life Sciences, Fudan University, Shanghai, China

## OPEN ACCESS

### Edited by:

Eleonora Napoli,  
University of California, Davis,  
United States

### Reviewed by:

Aurora Gomez-Duran,  
University of Cambridge,  
United Kingdom  
Prasun K. Datta,  
Temple University, United States

### \*Correspondence:

Li Jin  
lijin@fudan.edu.cn  
Jiucun Wang  
jcwang@fudan.edu.cn

### Specialty section:

This article was submitted to  
Cellular Biochemistry,  
a section of the journal  
*Frontiers in Molecular Biosciences*

Received: 18 July 2019

Accepted: 29 October 2019

Published: 15 November 2019

### Citation:

Sun D, Wei Y, Zheng H-X, Jin L and Wang J (2019) Contribution of Mitochondrial DNA Variation to Chronic Disease in East Asian Populations. *Front. Mol. Biosci.* 6:128.  
doi: 10.3389/fmbo.2019.00128

Mitochondria are the main producers of energy in eukaryotic cells. Mitochondrial dysfunction is associated with specific mitochondrial DNA (mtDNA) variations (haplogroups), and these variations can contribute to human disease. East Asian populations show enrichment of many mitochondrial haplogroups, including A, B, D, G, M7, M8, M9, N9, R9, and exhibit half of the known haplogroups of worldwide. In this review, we summarize the current research in the field of mtDNA variation and associated disease in East Asian populations and discuss the physiological and pathological relevance of mitochondrial biology. mtDNA haplogroups are associated with various metabolic disorders ascribed to altered oxidative phosphorylation. The same mitochondrial haplogroup can show either a negative or positive association with different diseases. Mitochondrial dynamics, mitophagy, and mitochondrial oxidative stress, ultimately influence susceptibility to various diseases. In addition, mitochondrial retrograde signaling pathways may have profound effects on nuclear-mitochondrial interactions, affecting cellular morphology, and function. Other complex networks including proteostasis, mitochondrial unfolded protein response and reactive oxygen species signaling may also play pivotal roles in metabolic performance.

**Keywords:** mitochondrial DNA, variation, haplogroup, chronic disease, East Asian

## INTRODUCTION

Mitochondria are cytoplasmic organelles of eukaryotic cells that provide more than 90% of the cell's adenosine triphosphate (ATP) through oxidative phosphorylation (OXPHOS) and the mitochondrial electron transport chain (ETC). OXPHOS enzymes include five complexes: complex I (NADH: ubiquinone oxidoreductase), complex II (FADH<sub>2</sub>: succinate dehydrogenase), complex III (coenzyme Q- Cytochrome C [Cyt-C] reductase), complex IV (Cyt-C oxidase), and complex V (F<sub>1</sub>F<sub>0</sub>-ATP synthase). These are embed in the inner mitochondrial membrane. There are about 1,500 proteins that maintain the normal structure and function of mitochondria. Thirteen of these proteins are mtDNA-encoded proteins. The remainder are encoded by nuclear genes and are synthesized in the cytoplasm and transported into mitochondria (Wallace, 2005). Electrons from NADH and FADH<sub>2</sub> are transferred to complexes I and II, respectively, and are donated to complex III via ubiquinone. Complex IV then receives electrons from complex III via Cyt-C to reduce molecular oxygen to water. Electrons transferred between these complexes generate a proton gradient across the inner mitochondrial membrane, which is then used by complex V to synthesize ATP from ADP (Wallace, 2013).

mtDNA is continuously synthesized throughout the cell cycle in early human embryonic development. mtDNA has an extremely high mutation rate, presumably due to the lack of histone protection and chronic exposure to mitochondrial reactive oxygen species (ROS). Pathogenic mtDNA mutations include rearrangement mutations (Holt et al., 1988), polypeptide gene missense mutations (Wallace et al., 1988a), and gene mutations (rRNA and tRNA) related to protein synthesis (Wallace et al., 1988b; Shoffner et al., 1990). These mismatches, designated single nucleotide polymorphisms (SNPs) by the Cambridge reference sequence for human mtDNA, determine mitochondrial haplogroups. A mitochondrial haplogroup is a combination of variants that are phylogenetically related (PhyloTree.org-mtDNA tree). A Sub-haplogroup is a sub-branch of haplogroup characterized by new variants, based on the major haplogroup branches. Numerous pathogenic variations in mtDNA or in nuclear DNA (nDNA) encoding mitochondrial proteins may lead to clinically and genetically heterogeneous disorders due to mitochondrial ETC dysfunction (Fang H. et al., 2015; van Rahden et al., 2015; Piekutowska-Abramczuk et al., 2018). Insufficient energy for cardiac and skeletal muscles, brain, liver, and kidney, may lead to metabolic disorders (Lu et al., 2010; Nishigaki et al., 2010; Wang et al., 2010; Liou et al., 2016). Tools for studies of mutant mtDNA have been established by fusing enucleated cells with mitochondria donors, which are called cybrids. Cybrids are useful for studying alterations of mitochondrial function at the cellular level without the influence of the nuclear background (King and Attardi, 1989; Wilkins et al., 2014).

As a multi-ethnic region, East Asia contains nearly half of the known mitochondrial haplogroups, most of which are associated with metabolic and degenerative diseases (Kong et al., 2006; Takasaki, 2008; van Oven and Kayser, 2009). No review has been published discussing the relationship between mtDNA variations and diseases in East Asia. It is important to correlate the East Asian mtDNA variations with different disease to determine their role in pathogenesis. Herein, we review the current research on mtDNA variations and haplogroups in various metabolic diseases and discuss the physiological and pathological relevance of mitochondrial biology in East Asia.

## MITOCHONDRIAL HAPLOGROUPS ASSOCIATED WITH DISEASE IN EAST ASIA

**Figure 1** and **Table 1** summarize previous studies of the East Asia mitochondrial haplogroups and their association with disease. Multi-ethnic East Asia populations (**Figure 1**) account for more than half of the known haplogroups worldwide. Haplogroups M and N are essentially equally represented. Haplogroup M includes sub-haplogroups D4, D5, M7, and G. Haplogroup N includes B4, B5, N9, A, and F. mtDNA variation is not uniform from south to north in East Asia. In the north, more haplogroup M is present, which includes the sub-haplogroups A, C, D4, D5, G, M8, M9, N9, and Z. In the south, more haplogroup N is found, specifically sub-haplogroups B4a, B5a, F, M7, and R9 (Xue et al., 2008). Most of these haplogroups are associated with metabolic and

degenerative diseases (Kong et al., 2006; Takasaki, 2008; van Oven and Kayser, 2009). **Table 1** shows mitochondrial haplogroups which are closely related to several diseases. Understanding the mechanisms of mitochondrial dysfunction is critical for clinical diagnosis and for development of therapies for patients with mitochondrial diseases.

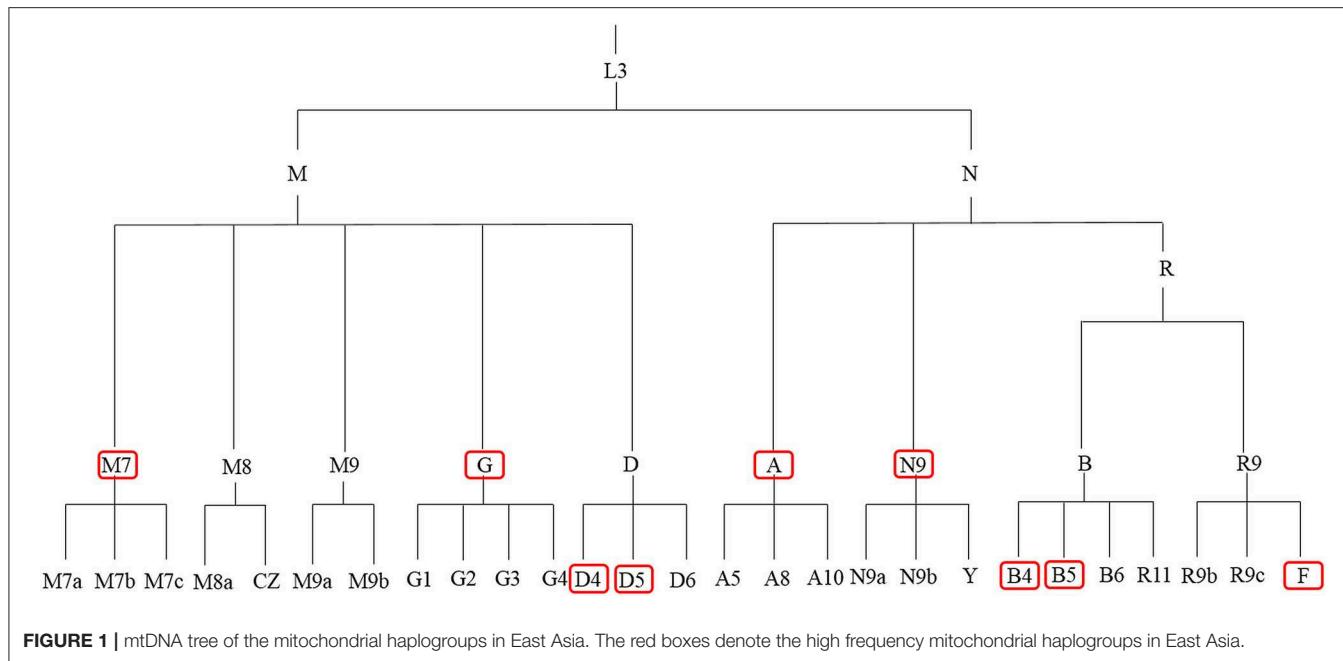
## Nervous System Diseases

Neurons need most energy produced from mitochondria to maintain neuron cellular function. In addition, normal mitophagy to eliminate misfolded and aggregated proteins is necessary for neuron cellular function. Mitophagy is a specific process that degrades damaged mitochondria to maintain cellular homeostasis based on the mechanism of autophagy (Youle and Narendra, 2011). Autophagy is dependent on enclosing ubiquitinated proteins in vesicles termed “autophagosomes,” and subsequently lysosomal fusion (Levine and Kroemer, 2008). mtDNA mutation can lead to mitophagy (Dombi et al., 2016) and mitochondrial dysfunction (Dolle et al., 2016; Pera et al., 2017; Ehrnhoefer et al., 2018; Lindqvist et al., 2018; Pereira et al., 2018; Puthumana and Regenold, 2019) have long been reported as pathogenic in psychiatric and neurodegenerative diseases.

### Alzheimer Disease and Parkinson's Disease

Mitochondrial abnormalities are associated with high levels of amyloid-beta protein in Alzheimer disease (AD) (Wang et al., 2007). Studies found increased tricarboxylic acid (TCA) cycle metabolism in AD patients due to an increased levels of ROS from deficiencies in OXPHOS (Bubber et al., 2005; Carvalho et al., 2009). Other papers suggest that mitochondrial dysfunction may start 3 months before extracellular deposition of amyloid-beta protein, and that progression accelerates with aging (Hauptmann et al., 2009). The mitophagy-related-protein-PINK1, which plays a pivotal role in Parkinson's disease (PD). PINK1 regulates mitochondrial stress through indirect interaction with mitochondrial proteases and the fission protein-Drp1 to further recruit Parkin by depolarized mitochondria (Chu, 2010). In addition, mitochondrial respiration complex I and IV deficiency may also contribute to the occurrence of AD and PD (Holper et al., 2019).

Comprehensive epidemiological analyses of mtDNA variations in Japanese patients with AD ( $n = 96$ ) or PD ( $n = 96$ ) showed that AD is uniquely associated with haplogroups G2a, B4c1, and N9b1, and PD with haplogroups M7a, M7b2, B4e, and B5b (Takasaki, 2008, 2009). In Han Chinese populations, haplogroup B5 is significantly associated with AD ( $n = 341$ ) in patients from Southwest China (Bi et al., 2015). Cells with the B5 haplogroup had higher levels of ROS, decreased mitochondrial mass, lower ATP generation, and lower respiration when compared with non-B5 haplogroup cells (Bi et al., 2015). A study of the distribution of mtDNA haplogroups of the Han population with sporadic PD ( $n = 279$ ) indicated that haplogroup B may confer a lower risk for PD, while haplogroup D may lead to a higher risk of PD in people younger than 50 years of age (Chen et al., 2015). Consistent with these findings, Liou et al. determined the association of mtDNA haplogroups with PD patients ( $n = 725$ ) in Taiwan. They also found that



mitochondrial haplogroup B5 confers resistance to PD. In cybrid cellular models, the B5 cybrid showed lower ROS generation and a lower rate of apoptosis compared with the B4 cybrid (Liou et al., 2016).

### Psychiatric Disorders

Mitochondrial abnormalities may be involved in the pathophysiology of psychiatric disorders, such as schizophrenia, bipolar disorder, and attention deficit hyperactivity disorder (ADHD). Studies showed decreased protein and transcript levels of mitochondrial complex I and IV, decreased mitochondrial fusion levels, increased fission levels, and impaired OXPHOS in patients with schizophrenia or with bipolar disorder (Bubber et al., 2004; Hjelm et al., 2015; Flippo and Strack, 2017; Haghightafard et al., 2017; Rollins et al., 2018; Holper et al., 2019).

A recent study of 11 families with schizophrenia demonstrated that mtDNA A15395G and A8536G were deleterious (Bi et al., 2016). Functional characterization further confirmed the potential pathogenicity of the two variants which includes lower mitomass, mtDNA copy number, respiration, ATP, and higher ROS (Bi et al., 2016). The T3644C mutation was found in Japanese patients with bipolar disorder ( $n = 199$ ) but not in healthy controls. This mutation converts a well-conserved valine, to alanine in the complex I ND1 subunit, and may impair assembly of complex I. The m.3644T>C (MT-ND1) variant alters mitochondrial function by decreasing mitochondrial membrane potential (MMP) and complex I activity in 3644C cybrids compared with 3644T cybrids (Munakata et al., 2004). An epidemiologic study of Korean ADHD children ( $n = 150$ ) revealed that haplogroup B4 increases the occurrence of ADHD, and haplogroup B5 and D4b are significantly associated with ADHD boys and girls, respectively. These results suggest that

mtDNA plays an important role in the genetic etiology of ADHD in Korean children (Hwang et al., 2018). Cybrids of the SH-SY5Y neuroblastoma cell line showed decreased complex V activity and MMP, but elevated oxidative stress (Verma et al., 2016).

### Optic Neuropathy

Mitochondrial dysfunction can also cause optic neuropathy. The relationship between several mtDNA variants (G11778A, G3460A, T14484C, G11696A, G13708A, G10680A, and T12338C) and Leber's hereditary optic neuropathy (LHON) have been reported (Yoneda et al., 1989; Hotta et al., 1995; Brown et al., 2000; Ji et al., 2008). The typical LHON-related G11778A mutation in different families belongs to the Chinese haplogroups B5b, G2a, C4a1, M7b102, and M8a; the Thai urban population haplogroups M and B; and European haplogroups J, K, and H, respectively. Several groups constructed cybrids of LHON probands carrying the G3460A, G11778A, and T14484C LHON primary mutations to confirm that mitochondrial dysfunction is caused by these mtDNA variants. They detected complex I-dependent defects in respiration, decreased ATP synthesis, increased ROS production, disrupted glutamate transport, increased mitochondrial-dependent apoptotic death, delayed complex I assembly kinetics, and instability of complexes III and IV (Ghelli et al., 2003; Baracca et al., 2005; Floreani et al., 2005; Pello et al., 2008). The mtDNA G13051A led to variable neurology and activated mitophagy in LHON patients (Dombi et al., 2016). Mitophagy activation can also repair LHON-associated mitochondrial dysfunction and improve cell survival (Sharma et al., 2019).

These results point to promising targets for predicting the probabilities and initial diagnosis of nervous system diseases, although the etiology of these diseases remains unclear. Targeting

**TABLE 1 |** Diseases associated with mitochondrial haplogroups in East Asia.

Haplogroups	Associated diseases	References	
A	Hearing loss	Wang et al., 2006	
	LHON	Ji et al., 2014	
	Gastric cancer	Bi et al., 2011	
	Oncocytic tumors	Lyu et al., 2019	
	Periodontitis	Wang et al., 2015	
	Cerebral infarction	Nishigaki et al., 2007a	
	Coronary atherosclerosis	Sawabe et al., 2011	
	COPD	Zheng et al., 2012	
	Hearing loss	Ying et al., 2015	
	Osteoarthritis	Koo et al., 2019	
B	Acute mountain sickness	Li et al., 2011	
	B4	ADHD	Hwang et al., 2018
	Deafness	Lu et al., 2010	
	B5	Alzheimer's disease	Bi et al., 2015
	ADHD	Hwang et al., 2018	
	Hypertension	Liu et al., 2009	
	Leigh syndrome	Hao et al., 2013	
	Deafness	Lu et al., 2010	
	OSCC	Lai et al., 2012	
	C4a1	Hearing loss	Yuan et al., 2007
C	LHON	Zhou et al., 2010	
	D	Parkinson's Disease	Chen et al., 2015
	Endometrial cancer	Xu et al., 2006	
	End-stage renal disease	Zhang et al., 2017	
	Lung cancer	Zheng et al., 2012	
	Seasonal cold periodontitis	Wang et al., 2015	
	D4	Deafness	Liao et al., 2007
	LHON	Qian et al., 2005	
	Diabetes mellitus	Liou et al., 2012	
	Longevity	Cai et al., 2009	
E	Nonalcoholic fatty liver disease	Lu et al., 2012	
	OSCC	Lai et al., 2012	
	Acute myeloid leukemia	Kim et al., 2018	
	D5	LHON	Qu et al., 2006
	Diabetes mellitus	Zhong et al., 2014	
	Breast cancer	Fang et al., 2010	
	Chronic hepatitis B virus infection	Li et al., 2018	
	Oncocytic tumors	Lyu et al., 2019	
	OSCC	Lai et al., 2012	
	Longevity	Alexe et al., 2007	
F	Metabolic Syndrome	Tanaka et al., 2007	
	E2b1	Diabetes mellitus	Loo et al., 2014
	ADHD	Hwang et al., 2018	
	Diabetes mellitus	Niu et al., 2015	
	Lung cancer	Fang Y. et al., 2015	
	Longevity	Feng et al., 2011	
	F1	Hearing loss	Yuan et al., 2007
	LHON	Qian et al., 2005	

(Continued)

**TABLE 1 |** Continued

Haplogroups	Associated diseases	References		
F2	Gastric cancer	Bi et al., 2011		
	Nasopharyngeal Carcinoma	Hu et al., 2014		
	Deafness	Lu et al., 2010		
	LHON	Liu et al., 2011		
	Gastric cancer	Bi et al., 2011		
	Diabetes mellitus	Liao et al., 2008		
	Lung cancer	Zheng et al., 2012		
	Osteoarthritis	Fang et al., 2014		
	G1	Metabolic Syndrome	Tanaka et al., 2007	
	H	Diabetes mellitus	Jiang et al., 2017	
M	M7	Breast cancer	Fang et al., 2010	
	M8	Hepatocellular carcinoma	Zhang et al., 2010	
	M1	LHON	Qu et al., 2010	
	M9	Diabetes mellitus	Liao et al., 2008	
	N	N9a	Hearing loss	Kato et al., 2012
		SCJD	Zhang et al., 2015	
		Diabetes mellitus	Fuku et al., 2007	
	R	N9b1	Metabolic Syndrome	Tanaka et al., 2007
		Alzheimer's disease	Takasaki, 2009	
		Hearing loss	Yuan et al., 2007	
W	R11	Azoospermia	Feng et al., 2013	
	Y	Diabetes mellitus	Liao et al., 2008	
	R9a	Hearing loss	Young et al., 2005	
	W3a	SCJD	Zhang et al., 2015	
		LHON	Yu et al., 2010	
		Leigh syndrome	Hao et al., 2013	
	Y2	Deafness	Ding et al., 2009	
		OSCC	Lai et al., 2012	
	Z3	LHON	Qu et al., 2010	

mtDNA, mitochondrial DNA; ATP, adenosine triphosphate; OXPHOS, oxidative phosphorylation; ETC, electron transport chain; cyt-C, Cytochrome C; ROS, reactive oxygen species; SNP, single nucleotide polymorphism; nDNA, nuclear DNA; AD, Alzheimer disease; TCA, tricarboxylic acid; PD, Parkinson's disease; ADHD, attention deficit hyperactivity disorder; LHON, Leber's hereditary optic neuropathy; DM, diabetes mellitus; DKD, diabetic kidney disease; HCC, hepatocellular carcinoma; EC, esophageal cancer; HCM, hypertrophic cardiomyopathy; LVNC, left ventricular non-compaction; UPR, unfolded protein response; COPD, chronic obstructive pulmonary disease; sCJD, sporadic Creutzfeldt-Jakob disease; OSCC, oral squamous cell carcinoma.

mitochondrial function and oxidative stress, with antioxidants, coenzyme Q10, and vitamin C may be effective strategies to ameliorate the progress of nervous system diseases.

## Endocrine System Diseases

Steroid hormone biosynthesis is carried out in mitochondria. The ATP produced by the mitochondria also provides energy for hormone generation and trafficking. mtDNA variants may result in endocrine organ defects due to impaired OXPHOS (Chow et al., 2017).

## Diabetes Mellitus

Diabetes mellitus (DM) is a major global health problem. It is a challenge to understand the physiological and pathological conditions that lead to the development of this disease (Zimmet et al., 2001; Roglic and Unwin, 2010). Mitochondria are essential for providing energy to maintain insulin metabolism; mtDNA mutations of OXPHOS complexes can lead to pancreatic islet dysfunction. Increased mitochondrial fission may impair endothelial function via increased ROS in DM (Shenouda et al., 2011). Decreased OXPHOS and fatty acid oxidation in insulin-sensitive tissues has been reported (Kwak and Park, 2016). Mitochondrial uncoupling may protect the mitochondrial matrix against lipid-induced mitochondrial damage (Schrauwen and Hesselink, 2004).

The association between mtDNA haplogroups and the risk of DM is controversial. Some studies found diabetes susceptibility genes located in mitochondria-encoded genomes, such as G3316A and C3310T in a Japanese family (Nakano et al., 1998; Hattori et al., 2005), and G3316A, C3310T, A3243G T3394C, G4491A, T16189C, and T16519C in a Chinese population ( $n = 826$ ) (Liao et al., 2008; Li M. Z. et al., 2008; Wang et al., 2013; Zhong et al., 2014). The N9a, M8a, B4 and D4 haplogroups appeared to be related to DM in East Asia ( $n = 1289$ ) (Fuku et al., 2007; Loo et al., 2014; Li et al., 2015). A cybrid with a C3310T mutation showed that mitochondrial complex I activity, ATP generation, oxygen consumption were significantly decreased (Chen et al., 2006). Fuku et al. found that mitochondrial haplogroup N9a was a significant protective factor for DM in a Korean study ( $n = 732$ ) (Fuku et al., 2007). In contrast, Niu et al. found that N9a was a risk factor for diabetic nephropathy ( $n = 235$ ) (Niu et al., 2015). Subsequently, Fang et al. confirmed that the N9a haplogroup increased the risk of DM in the Chinese population by altering mitochondrial function and intracellular mitochondrial signals. The N9a haplogroup cybrids exhibited lower respiratory chain complex activity, ATP, MMP and oxygen consumption; however, they contained more ROS and fragmented mitochondria than non-N9a haplogroup cybrids. Insulin-stimulated glucose uptake was partially inhibited through enhanced stimulation of ERK1/2 phosphorylation and subsequent TLR4 activation in N9a haplogroup cybrids (Fang et al., 2018). Taken together, these studies show that mitochondrial haplogroups profoundly affect the occurrence and development of diabetes. Some antioxidants, including vitamin C and vitamin E, can ameliorate the oxidative stress associated with diabetes (Victor et al., 2011).

## Diabetic Kidney Disease

Diabetic kidney disease (DKD) is the most common cause of end-stage kidney disease worldwide. Mitochondrial dysfunction plays a role in the pathophysiology of diabetes (Susztak et al., 2006; Fakhruddin et al., 2017; Forbes and Thorburn, 2018). Overproduction of ROS, activation of apoptosis, and defective mitophagy have been shown to contribute to the progression of the disease (Wei and Szeto, 2019). A study of mitochondria-targeted metabolic tubular injury in diabetic kidney disease, which included healthy controls ( $n = 65$ ), diabetes patients without kidney disease ( $n = 48$ ), and DKD patients ( $n = 60$ )

was carried out in China. The accumulation of damaged mtDNA, fragmented mitochondria, activated apoptosis, loss of MMP, and perturbations in glycolysis and TCA cycle were detected in tubules and PBMCs from the patients. These results indicate that mitochondrial damage may be the hallmark of DKD patients (Jiang et al., 2019).

Salidroside, an active component from the traditional Chinese medicine Rhodiola rosea L, stimulated the Sirt1/PGC-1alpha axis and ameliorated diabetic nephropathy through enhanced mitochondrial DNA copy number and ETC protein expression in mice (Xue et al., 2019). SIRT3 overexpression also inhibited kidney tumor cells and improved mitochondrial biogenesis rather than exhibiting the Warburg effect (Liu et al., 2018). Overexpression of MnSOD can abrogate mitochondrial dysfunction and effectively prevent the development of diabetic retinopathy (Madsen-Bouterse et al., 2010). Pyruvate kinase M2 activation may protect against the progression of diabetic glomerular pathology and mitochondrial dysfunction by increasing glucose metabolic flux, and inhibiting the production of toxic glucose metabolites (Qi et al., 2017).

## Obesity

Obesity is one of the most important pathogenic factors of DM. Impaired mitochondrial lipid oxidation, increased inflammation, and increased mitochondrial OXPHOS in the liver have been observed in obese subjects (Khasawneh et al., 2009; Rogge, 2009; Buchner et al., 2011). Guo et al. found that the C8684T transition of haplogroup M8a, and the C3497T and T1119C transitions of haplogroup B4c caused increased susceptibility to DM.

## Asthenozoospermia

Asthenozoospermia is a multi-factor disorder that affects approximately half of males with infertility, and nearly 15% of cases result from genetic abnormalities (Moore and Reijo-Pera, 2000; Ferlin et al., 2007). Mitochondrial ATP, appropriate MMP levels, respiration activity, and low ROS levels are necessary to sustain normal sperm motility (Kasai et al., 2002; Marchetti et al., 2002; Ferramosca et al., 2012). The first study of human mtDNA haplogroups associated with asthenozoospermia was performed in Spain. The authors showed that haplogroups H and T conferred susceptibility to non-asthenozoospermic and asthenozoospermic, respectively. More importantly, complex IV activity was significantly decreased in haplogroup T cybrids (Ruiz-Pesini et al., 2000). In the Han population, men with haplogroup R exhibited decreased frequency of asthenozoospermia ( $n = 312$ ) (Feng et al., 2013). Recently, Wang's group reported that haplogroup M8a played a critical role in the penetrance of variant C8684T in the pathogenesis of non-obstructive azoospermia due to increased mtDNA damage and impaired normal spermatogenesis (Ji et al., 2018). These studies indicated that mitochondria play a pivotal role of human fertility.

## Cancer

Cancer is a chronic non-communicable disease affecting most of the people worldwide. Cancer cells attack the immune

system of the human body. Mitochondria play a critical role in providing energy for T cells, B cells, and macrophages, albeit the metabolism of the various cells are diverse. For example, M1 macrophages utilize a modified TCA cycle to drive inflammation and are characterized by increased lactate production. However, M2 macrophages require both mitochondrial TCA metabolism and glycolysis; they use acetyl-CoA to drive forward flux through the ETC, but also require glycolysis, in part to support hexosamine biosynthesis (Vats et al., 2006; Haschemi et al., 2012; Huang et al., 2014; Jin et al., 2014). Each cell type possesses its own unique immune status, it can be pro-inflammatory or anti-inflammatory, depending on the different metabolic pathways. Additionally, new mitochondrial DNA synthesis enables NLRP3 inflammasome activation (Zhong et al., 2018). Therefore, mtDNA mutations profoundly affect the human immune system. Here, we focus on some common diseases associated with cancer.

The “Warburg effect” is a feature of most cancer cells, they prefer aerobic glycolysis from glucose to lactate in the presence of oxygen, rather than complete oxidation of glucose and mitochondrial respiration (Warburg, 1956; Wallace, 2005). Mitochondrial DNA mutation would affect mitochondrial biogenesis and turnover, mitophagy, fission and fusion dynamics, oxidative stress, and cell death (Smith et al., 2012). The balance of mitochondrial metabolism plays a crucial role in the pathogenesis and progression of human malignancies, and mtDNA mutations play bidirectional roles in tumorigenesis; some are protective factors, while others are risk factors (Xu et al., 2006; Li et al., 2007; Ma L. et al., 2018).

### Endometrial Cancer

A PGC1 $\alpha$ -dependent pathway increases mitochondrial biogenesis, mitochondrial fission, mitophagy, proteolysis, and antioxidant response in endometrial cancer (Cormio et al., 2017), and haplogroup D shows significant correlation with the incidence of endometrial cancer in the Yunnan province in China (Xu et al., 2006). Treatment of 1, 1-bis (3'-indolyl)-1-(p-substituted phenyl) methane has been reported to decrease mitochondrial membrane potential and induce apoptosis in endometrial cancer cell lines (Hong et al., 2008).

### Breast Cancer

Mitochondrial dysfunction and abnormal intracellular mitochondrial signaling has been detected in breast cancer cells (Santidrian et al., 2013; Pelicano et al., 2014). Haplogroup M may be a risk factor for breast cancer. Mutations in the D-loop region are more likely to be detected in benign breast tumors ( $n = 104$ ) (Fang et al., 2010). The frequency of haplogroup D5 is significantly increased in patients with breast cancer. It was found that mitochondrial respiration, ATP content, and MMP levels were decreased in D5 haplogroup cybrids compared to those with non-D5 haplogroups. The D5 cybrids were also more susceptible to tumorigenesis through activation of the AKT pathway, mediated by ROS generation (Ma L. et al., 2018). A potential therapeutic strategy for breast cancer may depend on improving the NAD+/NADH balance through treatment with

NAD $^{+}$  precursors, which can inhibit metastasis and prevent progression of breast cancer (Santidrian et al., 2013).

### Cervical Cancer

mtDNA variations located in the D-loop, coding region, and tRNA and rRNA genes are potential biomarkers in cervical carcinogenesis (Kabekkodu et al., 2014). Two groups investigated mtDNA mutations in cervical cancers of Chinese women. Zhai et al. reported that an mtDNA C150T polymorphism in HPV-positive cervical cancer patients was significantly increased compared to HPV-negative controls (Zhai et al., 2011). Li's group found that mitochondrial haplogroup D4b1 enhanced the risk of cervical cancer initiation in Chinese women ( $n = 150$ ) (Li et al., 2016).

There are many therapeutic drugs targeted to mitochondrial dysfunction available today: benzimidazolethiol induces apoptosis by regulating the PI3K/Akt signaling pathway, interferon alpha activates both the intrinsic mitochondrial pathway and endoplasmic reticulum stress-induced pathway, mefloquine impairs mitochondrial function and inhibits mTOR pathway, nicotinamide induces mitochondrial-mediated apoptosis through oxidative stress, tocotrienol inhibits proliferation and inducing apoptosis, betulinic acid induces apoptosis by regulating PI3K/Akt signaling, and pterostilbene targets m-TOR/PI3K/Akt signaling pathway via disruption of MMP (Hoti et al., 2003; Shi et al., 2016; Feng et al., 2017; Li et al., 2017; Xu T. et al., 2017; Xu W. et al., 2017; Hong Bin et al., 2018; Tian et al., 2018).

### Prostate Cancer

Prostate cancer is associated with dysregulation of OXPHOS. A multiethnic cohort epidemiological study of 4,086 prostate cancer cases and 3,698 controls from African, Asian, American, European, Latino, and Native Hawaiian patients was performed in order to examine the association of mtDNA and prostate cancer. This study revealed that haplogroup N contributed to overall prostate cancer, however, the mtDNA-encoded OXPHOS genes were not associated with prostate cancer risk in this cohort (Giorgi et al., 2016). A study of a Korean population also revealed no association of the mtDNA-encoded OXPHOS genes with prostate cancer ( $n = 139$ ) (Kim et al., 2008). However, several studies of prostate cancer demonstrated that mitochondrial dysfunction and altered intermediary metabolism, especially high ROS levels, occurred in prostate cancer cells (Dakubo et al., 2006; Mizumachi et al., 2008; Altieri, 2010; Chaudhary et al., 2017). ROS production accelerated mtDNA mutations in prostate cancer and further stimulated malignant transformation of prostate through increased ETC activity (Dakubo et al., 2006).

### Lung Cancer

Cell migration and invasion, which occurs through the induction of AKT and AMPK pathways in lung cancer cells, has been associated with mitochondrial dysfunction (Han et al., 2018). Two case-control cohort studies found an association between mtDNA variation and lung cancer risk in a Han Chinese population from southwestern China ( $n = 422$ ). Zheng et al. revealed that haplogroups D and F were protective factors for

lung cancer, while haplogroups G and M7 increased susceptibility (Zheng et al., 2012). However, Fang's group demonstrated that haplogroups F and G predisposed people to lung cancer ( $n = 237$ ). Although the results varied, both studies suggest that haplogroup G is a risk factor for lung cancer due to excess ROS generated by the impaired mitochondrial respiration chain (Fang Y. et al., 2015). In lung cancer, altered rates of mitochondrial fission and fusion were seen, which can influence metabolic function, proliferation, and cell survival. Therefore, altering mitochondrial dynamics may be a therapeutic strategy, for example, inhibiting mitochondrial fission can prevent cell cycle progression in lung cancer (Rehman et al., 2012; Lennon and Salgia, 2014).

### Hepatocellular Carcinoma

Mutations in the mitochondrial D-loop region have been reported in hepatocellular carcinoma (HCC), which may partly contribute to cancer development (Zhang et al., 2010). Mitochondrial pyruvate carrier (MPC1/2) protein expression was significantly downregulated in HCC, and may serve as a biomarker for the identification of patients with this disease (Ma X. et al., 2018). Mitochondrial fission significantly promoted the reprogramming of focal-adhesion dynamics and lamellipodia formation in HCC cells, mainly by activating  $\text{Ca}^{2+}/\text{CaMKII/ERK/FAK}$  pathway (Sun et al., 2018). In addition, Drp1-mediated mitochondrial fission promoted cell proliferation through crosstalk between the p53 and NF-kappaB pathways in HCC (Zhan et al., 2016). Therefore, treatment with mitochondrial division inhibitor-1 may decrease proliferation in HCC cells.

### Esophageal Cancer

Esophageal cancer (EC) has a very high mortality rate in China. Casticin treatment plays a pivotal role in inhibiting proliferation and inducing apoptosis of EC cells through activation of JNK signaling pathway, and hesperetin induces apoptosis via increased intracellular reactive oxygen species (Wu et al., 2016; Qiao et al., 2019). A study of mitochondrial haplogroups and esophageal cancer ( $n = 30$ ) in the Taihang Mountain and Chaoshan areas of China has shown that haplogroups D4a and D5 in Taihang Mountain, and haplogroups D and D5 in Chaoshan areas, were related to higher susceptibility to esophageal cancer (Li et al., 2007). Overall, haplogroup D, specifically sub-haplogroups D4a and D5a, can serve as potential biomarkers for esophageal cancer, at least in these two areas. Other papers showed that haplogroup D4a was associated with an increased risk of thyroid cancer ( $n = 100$ ) in China (Fang et al., 2010). As previously mentioned, haplogroup D5 was also a risk factor of breast cancer. It appears that haplogroup D is a risk factor for a diverse group of diseases because it impairs mitochondrial OXPHOS.

## Cardiovascular and Cerebrovascular Diseases

Cardiovascular and cerebrovascular diseases are major health problems worldwide, but Asian countries have higher mortality rates for stroke than Western countries, although these rates have recently decreased in Japan and urban areas in China. South

Asian, but not East Asian countries have a higher mortality rate for coronary heart disease than Western countries (Zhang et al., 2007; Ueshima et al., 2008). There are about 290 million patients, or one in five adults, with cardiovascular or cerebrovascular diseases in China (Wallace, 2000).

### Hypertension

Hypertension is one of the most common risk factors of cardiovascular disease, affecting ~168.1 million in China (Wang et al., 2019). It can be caused by both hereditary and genetic factors. Mutations in the mitochondrial genome are associated with essential hypertension; several mtDNA mutations associated with hypertension are found in haplogroup D4j (A4295G) (Li Z. et al., 2008), haplogroup G2a1 (A4435G) (Lu et al., 2011), and haplogroup B5b1 (T16189C) (Zhu et al., 2016). Mitochondrial dysfunction associated with increased ROS production may be involved in the pathogenesis of hypertension (Dikalov and Ungvari, 2013; Lahera et al., 2017). Lymphocyte cell lines with a tRNA(Met) C4467A mutation showed oxidative stress and mitochondrial biogenesis dysfunction, including lower ATP generation, MMP activity, and oxygen consumption, and increased ROS levels (Liu et al., 2017).

### Myocardial and Cerebral Infarction

Myocardial and cerebral infarction are multifactorial disorders affected by both genetic and environmental conditions, such as coronary atherosclerosis (Sawabe et al., 2011), carotid artery stenosis (Iizuka et al., 2009), hypertrophic cardiomyopathy (Wei et al., 2009), and left ventricular non-compaction (Tang et al., 2010). Mitochondria-derived ROS plays a role in myocardial and cerebral infarction due to the fact that mitochondria in vascular endothelial cells are the major source of superoxide (Guzik et al., 2006). Therefore, it is important to determine the variants of mtDNA associated with myocardial and cerebral infarction. In the Japanese population, mtDNA C5178A transversion causes leucine to methionine substitution in ND2, resulting in anti-atherosclerotic effects in diabetic subjects and a lower prevalence of myocardial infarction (Takagi et al., 2004). The study found that mitochondrial haplogroup A contributes to atherothrombotic cerebral infarction ( $n = 1,181$ ), but only in females. Haplogroup N9b protects against myocardial infarction; however, haplogroup G1 is a risk factor for this disease in Japanese males (Nishigaki et al., 2007a,b). Mitochondrial haplogroups A and M7a increase the risk for coronary atherosclerosis in a Japanese population ( $n = 1,536$ ). Surprisingly, a haplogroup associated with extreme longevity, D4a, conferred a risk of myocardial infarction (Alexe et al., 2007; Cai et al., 2009; Sawabe et al., 2011). Inhibition of mitochondrial permeability transition improved functional recovery and reduced mortality following acute myocardial infarction in mice (Gomez et al., 2007). Therefore, targeting mitochondrial calcium transport and inhibiting mitochondrial fission may be effective strategies for myocardial infarction (Cooper and Eguchi, 2018; Frangogiannis, 2018).

### Hypertrophic Cardiomyopathy

Hypertrophic cardiomyopathy (HCM) is a common genetic disorder, affecting 1 in 500 individuals worldwide (Maron et al.,

2006). Mitochondrial haplogroup M10 may be a risk factor for HCM, specifically, three mutations - G7967A in the COX II of complex IV, and T12477C and G13135A in the ND5 of complex I. Mitochondrial complex I activity was markedly decreased in the HCM individuals, resulting in disrupted mitochondrial respiratory function (Wei et al., 2009). The mitochondrial ND5 T12338C variant which belongs to haplogroup F2a was associated with hypertrophic cardiomyopathy in a Chinese pedigree (Liu et al., 2012). A characteristic T2336C homoplasmic mutation in the mitochondrial 16S rRNA gene of HCM has also been found in a Chinese family. Reduced ATP and MMP levels, and increased ROS generation in the mutant 2336C cybrids may lead to deterioration of mitochondrial function (Li et al., 2018).

### Left Ventricular Non-compaction

Left ventricular non-compaction (LVNC) is a genetically heterogeneous pathological disorder that is affected by both the nuclear and mitochondrial genomes, leading to congenital heart disease (Digilio et al., 1999; Xing et al., 2006). Tang et al. found that mtDNA A3397G and T3398C of the complex I ND1 subunit may disrupt mitochondrial function to initiate LVNC (Tang et al., 2010).

### Stroke

Stroke is a complex multifactorial disorder caused by both genetics and environment in the Chinese population (Della-Morte et al., 2012). One study shows that mtDNA C5178A belongs to haplogroup D4b and is a protective factor for IS in the Chinese Han population ( $n = 200$ ) (Yang et al., 2014). Other studies reported that m.5178C, but not m.5178A, is a risk factor for several diseases including myocardial infarction ( $n = 517$ ), cerebrovascular diseases ( $n = 127$ ), and diabetes ( $n = 270$ ) (Wang et al., 2001; Ohkubo et al., 2002; Takagi et al., 2004). One explanation may be that the production of ROS in complex I disrupts the structure and respiratory chain function, and further damages the cardiovascular system. Rapamycin treatment attenuated mitochondrial dysfunction following cerebral ischemia, possibly through enhancement of mitophagy (Li et al., 2014).

## REFERENCES

- Alexe, G., Fuku, N., Bilal, E., Ueno, H., Nishigaki, Y., Fujita, Y., et al. (2007). Enrichment of longevity phenotype in mtDNA haplogroups D4b2b, D4a, and D5 in the Japanese population. *Hum. Genet.* 121, 347–356. doi: 10.1007/s00439-007-0330-6
- Altieri, D. C. (2010). Mitochondrial Hsp90 chaperones as novel molecular targets in prostate cancer. *Fut. Oncol.* 6, 487–489. doi: 10.2217/fon.10.26
- Baracca, A., Solaini, G., Sgarbi, G., Lenaz, G., Baruzzi, A., Schapira, A. H., et al. (2005). Severe impairment of complex I-driven adenosine triphosphate synthesis in leber hereditary optic neuropathy cybrids. *Arch. Neurol.* 62, 730–736. doi: 10.1001/archneur.62.5.730
- Bi, R., Li, W. L., Chen, M. Q., Zhu, Z., and Yao, Y. G. (2011). Rapid identification of mtDNA somatic mutations in gastric cancer tissues based on the mtDNA phylogeny. *Mutat. Res.* 709–710, 15–20. doi: 10.1016/j.mrfmmm.2011.02.016
- Bi, R., Tang, J., Zhang, W., Li, X., Chen, S. Y., Yu, D., et al. (2016). Mitochondrial genome variations and functional characterization in Han Chinese families with schizophrenia. *Schizophr. Res.* 171, 200–206. doi: 10.1016/j.schres.2016.01.011
- Bi, R., Zhang, W., Yu, D., Li, X., Wang, H. Z., Hu, Q. X., et al. (2015). Mitochondrial DNA haplogroup B5 confers genetic susceptibility to Alzheimer's disease in Han Chinese. *Neurobiol. Aging* 36, 1604.e1607–1616. doi: 10.1016/j.neurobiolaging.2014.10.009
- Brown, M. D., Trounce, I. A., Jun, A. S., Allen, J. C., and Wallace, D. C. (2000). Functional analysis of lymphoblast and cybrid mitochondria containing the 3460, 11778, or 14484 Leber's hereditary optic neuropathy mitochondrial DNA mutation. *J. Biol. Chem.* 275, 39831–39836. doi: 10.1074/jbc.M006476200
- Bubber, P., Haroutunian, V., Fisch, G., Blass, J. P., and Gibson, G. E. (2005). Mitochondrial abnormalities in Alzheimer brain: mechanistic implications. *Ann. Neurol.* 57, 695–703. doi: 10.1002/ana.20474
- Bubber, P., Tang, J., Haroutunian, V., Xu, H., Davis, K. L., Blass, J. P., et al. (2004). Mitochondrial enzymes in schizophrenia. *J. Mol. Neurosci.* 24, 315–321. doi: 10.1385/JMN:24:2:315

## CONCLUSIONS

In summary, the studies described in this review shed light on the pathogenesis of diseases associated with mitochondrial dysfunction. They provide biological plausibility for the observed epidemiological surveys, although a number of limitations must also be considered; notably, the specific mechanisms of disease require further investigation. An intriguing finding is that the same mitochondrial haplogroup can have negative or positive association with different diseases. These associations remain unclear, although haplogroups defined by using common SNPs in each ethnic group may be a potential explanation. The distribution of mtDNA allele frequency varies considerably with the populations studied, which implies that haplogroup analysis is insufficient across populations. The results suggest that there are mechanisms other than mitochondrial pathways that affect susceptibility to these diseases. Mitochondrial retrograde signaling pathways may have profound effects on nuclear-mitochondrial interactions in cellular morphology and functionality. A possible explanation is that the nucleus attempts to make more mitochondria compensate for the energy deficiency. Other complex networks including proteostasis, mitochondrial unfolded protein response (UPR) and ROS signaling may also play pivotal roles in the organism's metabolism and deserve future investigation.

## AUTHOR CONTRIBUTIONS

DS and H-XZ designed this review. DS searched literature and wrote the initial manuscript. YW made the table. JW and LJ supervised and provided critical comments on the manuscript. DS, YW, H-XZ, LJ, and JW read, amended, and discussed the article.

## FUNDING

This study was supported by the grants from the National Science Foundation of China (31871436) and Shanghai Municipal Science and Technology Major Project (2017SHZDZX01).

- Buchner, D. A., Yazbek, S. N., Solinas, P., Burrage, L. C., Morgan, M. G., Hoppel, C. L., et al. (2011). Increased mitochondrial oxidative phosphorylation in the liver is associated with obesity and insulin resistance. *Obesity* 19, 917–924. doi: 10.1038/oby.2010.214
- Cai, X. Y., Wang, X. F., Li, S. L., Qian, J., Qian, D. G., Chen, F., et al. (2009). Association of mitochondrial DNA haplogroups with exceptional longevity in a Chinese population. *PLoS ONE* 4:e6423. doi: 10.1371/journal.pone.0006423
- Carvalho, C., Correia, S. C., Santos, R. X., Cardoso, S., Moreira, P. I., Clark, T. A., et al. (2009). Role of mitochondrial-mediated signaling pathways in Alzheimer disease and hypoxia. *J. Bioenerg. Biomembr.* 41, 433–440. doi: 10.1007/s10863-009-9247-1
- Chaudhary, A. K., O’Malley, J., Kumar, S., Inigo, J. R., Kumar, R., Yadav, N., et al. (2017). Mitochondrial dysfunction and prostate cancer racial disparities among American men. *Front. Biosci.* 9, 154–164. doi: 10.2741/s479
- Chen, J., Hattori, Y., Nakajima, K., Eizawa, T., Ehara, T., Koyama, M., et al. (2006). Mitochondrial complex I activity is significantly decreased in a patient with maternally inherited type 2 diabetes mellitus and hypertrophic cardiomyopathy associated with mitochondrial DNA C3310T mutation: a cybrid study. *Diabetes Res. Clin. Pract.* 74, 148–153. doi: 10.1016/j.diabres.2006.03.024
- Chen, Y. F., Chen, W. J., Lin, X. Z., Zhang, Q. J., Cai, J. P., Liou, C. W., et al. (2015). Mitochondrial DNA haplogroups and the risk of sporadic parkinson’s disease in han Chinese. *Chin. Med. J.* 128, 1748–1754. doi: 10.4103/0366-6999.159348
- Chow, J., Rahman, J., Achermann, J. C., Dattani, M. T., and Rahman, S. (2017). Mitochondrial disease and endocrine dysfunction. *Nat. Rev. Endocrinol.* 13, 92–104. doi: 10.1038/nrendo.2016.151
- Chu, C. T. (2010). A pivotal role for PINK1 and autophagy in mitochondrial quality control: implications for Parkinson disease. *Hum. Mol. Genet.* 19, R28–R37. doi: 10.1093/hmg/ddq143
- Cooper, H. A., and Eguchi, S. (2018). Inhibition of mitochondrial fission as a novel therapeutic strategy to reduce mortality upon myocardial infarction. *Clin. Sci.* 132, 2163–2167. doi: 10.1042/CS20180671
- Cormio, A., Musicco, C., Gasparre, G., Cormio, G., Pesce, V., Sardanelli, A. M., et al. (2017). Increase in proteins involved in mitochondrial fission, mitophagy, proteolysis and antioxidant response in type I endometrial cancer as an adaptive response to respiratory complex I deficiency. *Biochem. Biophys. Res. Commun.* 491, 85–90. doi: 10.1016/j.bbrc.2017.07.047
- Dakubo, G. D., Parr, R. L., Costello, L. C., Franklin, R. B., and Thayer, R. E. (2006). Altered metabolism and mitochondrial genome in prostate cancer. *J. Clin. Pathol.* 59, 10–16. doi: 10.1136/jcp.2005.027664
- Della-Morte, D., Guadagni, F., Palmirota, R., Testa, G., Caso, V., Paciaroni, M., et al. (2012). Genetics of ischemic stroke, stroke-related risk factors, stroke precursors and treatments. *Pharmacogenomics* 13, 595–613. doi: 10.2217/pgs.12.14
- Digilio, M. C., Marino, B., Bevilacqua, M., Musolino, A. M., Giannotti, A., and Dallapiccola, B. (1999). Genetic heterogeneity of isolated noncompaction of the left ventricular myocardium. *Am. J. Med. Genet.* 85, 90–91. doi: 10.1002/(SICI)1096-8628(19990702)85:1<90::AID-AJMG19>3.0.CO;2-U
- Dikalov, S. I., and Ungvari, Z. (2013). Role of mitochondrial oxidative stress in hypertension. *Am. J. Physiol. Heart Circ. Physiol.* 305, H1417–H1427. doi: 10.1152/ajpheart.00089.2013
- Ding, Y., Li, Y., You, J., Yang, L., Chen, B., Lu, J., et al. (2009). Mitochondrial tRNA(Glu) A14693G variant may modulate the phenotypic manifestation of deafness-associated 12S rRNA A1555G mutation in a Han Chinese family. *J. Genet. Genomics* 36, 241–250. doi: 10.1016/S1673-8527(08)60111-3
- Dolle, C., Flones, I., Nido, G. S., Miletic, H., Osuagwu, N., Kristoffersen, S., et al. (2016). Defective mitochondrial DNA homeostasis in the substantia nigra in Parkinson disease. *Nat. Commun.* 7:13548. doi: 10.1038/ncomms13548
- Dombi, E., Diot, A., Morten, K., Carver, J., Lodge, T., Fratter, C., et al. (2016). The m.13051G>A mitochondrial DNA mutation results in variable neurology and activated mitophagy. *Neurology* 86, 1921–1923. doi: 10.1212/WNL.0000000000002688
- Ehrnhofer, D. E., Southwell, A. L., Sivasubramanian, M., Qiu, X., Villanueva, E. B., Xie, Y., et al. (2018). HACE1 is essential for astrocyte mitochondrial function and influences Huntington disease phenotypes *in vivo*. *Hum. Mol. Genet.* 27, 239–253. doi: 10.1093/hmg/ddx394
- Fakhruddin, S., Alanazi, W., and Jackson, K. E. (2017). Diabetes-induced reactive oxygen species: mechanism of their generation and role in renal injury. *J. Diabetes Res.* 2017:8379327. doi: 10.1155/2017/8379327
- Fang, H., Hu, N., Zhao, Q., Wang, B., Zhou, H., Fu, Q., et al. (2018). mtDNA haplogroup N9a increases the risk of type 2 diabetes by altering mitochondrial function and intracellular mitochondrial signals. *Diabetes* 67, 1441–1453. doi: 10.2337/db17-0974
- Fang, H., Liu, X., Shen, L., Li, F., Liu, Y., Chi, H., et al. (2014). Role of mtDNA haplogroups in the prevalence of knee osteoarthritis in a southern Chinese population. *Int. J. Mol. Sci.* 15, 2646–2659. doi: 10.3390/ijms15022646
- Fang, H., Shen, L., Chen, T., He, J., Ding, Z., Wei, J., et al. (2010). Cancer type-specific modulation of mitochondrial haplogroups in breast, colorectal and thyroid cancer. *BMC Cancer* 10:421. doi: 10.1186/1471-2407-10-421
- Fang, H., Shi, H., Li, X., Sun, D., Li, F., Li, B., et al. (2015). Exercise intolerance and developmental delay associated with a novel mitochondrial ND5 mutation. *Sci. Rep.* 5:10480. doi: 10.1038/srep10480
- Fang, Y., Yang, H. Y., Shi, Y. H., Cui, J. H., Li, L. Y., Xu, Y. C., et al. (2015). Mitochondrial DNA haplogroups and somatic mutations are associated with lung cancer in patients from Southwest China. *Genet. Mol. Res.* 14, 5031–5043. doi: 10.4238/2015.May.12.6
- Feng, G. F., Zhang, J., Feng, L. M., Shen, N. X., Li, L. J., and Zhu, Y. M. (2013). Mitochondrial DNA haplogroup associated with sperm motility in the Han population. *Asian J. Androl.* 15, 630–633. doi: 10.1038/aja.2013.83
- Feng, J., Zhang, J., Liu, M., Wan, G., Qi, K., Zheng, C., et al. (2011). Association of mtDNA haplogroup F with healthy longevity in the female Chuang population, China. *Exp. Gerontol.* 46, 987–993. doi: 10.1016/j.exger.2011.09.001
- Feng, Y., Wang, Y., Jiang, C., Fang, Z., Zhang, Z., Lin, X., et al. (2017). Nicotinamide induces mitochondrial-mediated apoptosis through oxidative stress in human cervical cancer HeLa cells. *Life Sci.* 181, 62–69. doi: 10.1016/j.lfs.2017.06.003
- Perlin, A., Raicu, F., Gatta, V., Zuccarello, D., Palka, G., and Foresta, C. (2007). Male infertility: role of genetic background. *Reprod. Biomed. Online* 14, 734–745. doi: 10.1016/S1472-6483(10)60677-3
- Ferramosca, A., Provenzano, S. P., Coppola, L., and Zara, V. (2012). Mitochondrial respiratory efficiency is positively correlated with human sperm motility. *Urology* 79, 809–814. doi: 10.1016/j.urology.2011.12.042
- Flippo, K. H., and Strack, S. (2017). An emerging role for mitochondrial dynamics in schizophrenia. *Schizophr. Res.* 187, 26–32. doi: 10.1016/j.schres.2017.05.003
- Floreani, M., Napoli, E., Martinuzzi, A., Pantano, G., De Riva, V., Trevisan, R., et al. (2005). Antioxidant defences in cybrids harboring mtDNA mutations associated with Leber’s hereditary optic neuropathy. *FEBS J.* 272, 1124–1135. doi: 10.1111/j.1742-4658.2004.04542.x
- Forbes, J. M., and Thorburn, D. R. (2018). Mitochondrial dysfunction in diabetic kidney disease. *Nat. Rev. Nephrol.* 14, 291–312. doi: 10.1038/nrneph.2018.9
- Frangogiannis, N. G. (2018). Targeting mitochondrial calcium transport in myocardial infarction. *Hellenic J. Cardiol.* 59, 223–225. doi: 10.1016/j.hjc.2018.06.010
- Fuku, N., Park, K. S., Yamada, Y., Nishigaki, Y., Cho, Y. M., Matsuo, H., et al. (2007). Mitochondrial haplogroup N9a confers resistance against type 2 diabetes in Asians. *Am. J. Hum. Genet.* 80, 407–415. doi: 10.1086/512202
- Ghelli, A., Zanna, C., Porcelli, A. M., Schapira, A. H., Martinuzzi, A., Carelli, V., et al. (2003). Leber’s hereditary optic neuropathy (LHON) pathogenic mutations induce mitochondrial-dependent apoptotic death in transmtochondrial cells incubated with galactose medium. *J. Biol. Chem.* 278, 4145–4150. doi: 10.1074/jbc.M2120285200
- Giorgi, E. E., Li, Y., Caberto, C. P., Beckman, K. B., Lum-Jones, A., Haiman, C. A., et al. (2016). No association between the mitochondrial genome and prostate cancer risk: the multiethnic cohort. *Cancer Epidemiol. Biomarkers Prev.* 25, 1001–1003. doi: 10.1158/1055-9965.EPI-16-0111
- Gomez, L., Thibault, H., Gharib, A., Dumont, J. M., Vuagniaux, G., Scalfaro, P., et al. (2007). Inhibition of mitochondrial permeability transition improves functional recovery and reduces mortality following acute myocardial infarction in mice. *Am. J. Physiol. Heart Circ. Physiol.* 293, H1654–H1661. doi: 10.1152/ajpheart.01378.2006
- Guzik, T. J., Sadowski, J., Guzik, B., Jopek, A., Kapelak, B., Przybylowski, P., et al. (2006). Coronary artery superoxide production and nox isoform expression in human coronary artery disease. *Arterioscler. Thromb. Vasc. Biol.* 26, 333–339. doi: 10.1161/01.ATV.0000196651.64776.51

- Haghightafard, A., Andalib, S., Amini Faskhodi, M., Sadeghi, S., Ghaderi, A. H., Moradkhani, S., et al. (2017). Gene expression study of mitochondrial complex I in schizophrenia and paranoid personality disorder. *World J. Biol. Psychiatry* 19(Sup3):S133–S146. doi: 10.1080/15622975.2017.1282171
- Han, S. Y., Jeong, Y. J., Choi, Y., Hwang, S. K., Bae, Y. S., and Chang, Y. C. (2018). Mitochondrial dysfunction induces the invasive phenotype, and cell migration and invasion, through the induction of AKT and AMPK pathways in lung cancer cells. *Int. J. Mol. Med.* 42, 1644–1652. doi: 10.3892/ijmm.2018.3733
- Hao, X. D., Yang, Y. L., Tang, N. L., Kong, Q. P., Wu, S. F., and Zhang, Y. P. (2013). Mitochondrial DNA haplogroup Y is associated to Leigh syndrome in Chinese population. *Gene* 512, 460–463. doi: 10.1016/j.gene.2012.10.054
- Haschemi, A., Kosma, P., Gille, L., Evans, C. R., Burant, C. F., Starkl, P., et al. (2012). The sedoheptulose kinase CARKL directs macrophage polarization through control of glucose metabolism. *Cell Metab.* 15, 813–826. doi: 10.1016/j.cmet.2012.04.023
- Hattori, Y., Takeoka, M., Nakajima, K., Ehara, T., and Koyama, M. (2005). A heteroplasmic mitochondrial DNA 3310 mutation in the ND1 gene in a patient with type 2 diabetes, hypertrophic cardiomyopathy, and mental retardation. *Exp. Clin. Endocrinol. Diabetes* 113, 318–323. doi: 10.1055/s-2005-865646
- Hauptmann, S., Scherping, I., Drose, S., Brandt, U., Schulz, K. L., Jendrach, M., et al. (2009). Mitochondrial dysfunction: an early event in Alzheimer pathology accumulates with age in AD transgenic mice. *Neurobiol. Aging* 30, 1574–1586. doi: 10.1016/j.neurobiolaging.2007.12.005
- Hjelm, B. E., Rollins, B., Mamdani, F., Lauterborn, J. C., Kirov, G., Lynch, G., et al. (2015). Evidence of mitochondrial dysfunction within the complex genetic etiology of schizophrenia. *Mol. Neuropsychiatr.* 1, 201–219. doi: 10.1159/000441252
- Holper, L., Ben-Shachar, D., and Mann, J. J. (2019). Multivariate meta-analyses of mitochondrial complex I and IV in major depressive disorder, bipolar disorder, schizophrenia, Alzheimer disease, and Parkinson disease. *Neuropsychopharmacology* 44, 837–849. doi: 10.1038/s41386-018-0090-0
- Holt, I. J., Harding, A. E., and Morgan-Hughes, J. A. (1988). Deletions of muscle mitochondrial DNA in patients with mitochondrial myopathies. *Nature* 331, 717–719. doi: 10.1038/331717a0
- Hong Bin, W., Da, L. H., Xue, Y., and Jing, B. (2018). Pterostilbene (3',5'-dimethoxy-resveratrol) exerts potent antitumor effects in HeLa human cervical cancer cells via disruption of mitochondrial membrane potential, apoptosis induction and targeting m-TOR/PI3K/Akt signalling pathway. *J. Buon.* 23, 1384–1389.
- Hong, J., Samudio, I., Chinthalapalli, S., and Safe, S. (2008). 1,1-bis(3'-indolyl)-1-(p-substituted phenyl)methanes decrease mitochondrial membrane potential and induce apoptosis in endometrial and other cancer cell lines. *Mol. Carcinog.* 47, 492–507. doi: 10.1002/mc.20407
- Hoti, N., Ma, J., Tabassum, S., Wang, Y., and Wu, M. (2003). Triphenyl tin benzimidazolethiol, a novel antitumor agent, induces mitochondrial-mediated apoptosis in human cervical cancer cells via suppression of HPV-18 encoded E6. *J. Biochem.* 134, 521–528. doi: 10.1093/jb/mvg169
- Hotta, Y., Fujiki, K., Hayakawa, M., Nakajima, A., Kanai, A., Mashima, Y., et al. (1995). Clinical features of Japanese Leber's hereditary optic neuropathy with 11778 mutation of mitochondrial DNA. *Jpn. J. Ophthalmol.* 39, 96–108.
- Hu, S. P., Du, J. P., Li, D. R., and Yao, Y. G. (2014). Mitochondrial DNA haplogroup confers genetic susceptibility to nasopharyngeal carcinoma in Chaoshanese from Guangdong, China. *PLoS ONE* 9:e87795. doi: 10.1371/journal.pone.0087795
- Huang, S. C., Everts, B., Ivanova, Y., O'Sullivan, D., Nascimento, M., Smith, A. M., et al. (2014). Cell-intrinsic lysosomal lipolysis is essential for alternative activation of macrophages. *Nat. Immunol.* 15, 846–855. doi: 10.1038/ni.2956
- Hwang, I. W., Kwon, B. N., Kim, H. J., Han, S. H., Lee, N. R., Lim, M. H., et al. (2018). Assessment of associations between mitochondrial DNA haplogroups and attention deficit and hyperactivity disorder in Korean children. *Mitochondrion* 47, 174–178. doi: 10.1016/j.mito.2018.11.003
- Iizuka, T., Goto, Y., Miyakawa, S., Sato, M., Wang, Z., Suzuki, K., et al. (2009). Progressive carotid artery stenosis with a novel tRNA phenylalanine mitochondrial DNA mutation. *J. Neurol. Sci.* 278, 35–40. doi: 10.1016/j.jns.2008.11.016
- Ji, J., Xu, M., Wang, R., Wang, Y., Qin, Y., Li, L., et al. (2018). Human mitochondrial DNA haplogroup M8a influences the penetrance of m.8684C>T in Han Chinese men with non-obstructive azoospermia. *Reprod. Biomed. Online* 37, 480–488. doi: 10.1016/j.rbmo.2018.08.004
- Ji, Y., Liang, M., Zhang, J., Zhang, M., Zhu, J., Meng, X., et al. (2014). Mitochondrial haplotypes may modulate the phenotypic manifestation of the LHON-associated ND1 G3460A mutation in Chinese families. *J. Hum. Genet.* 59, 134–140. doi: 10.1038/jhg.2013.134
- Ji, Y., Zhang, A. M., Jia, X., Zhang, Y. P., Xiao, X., Li, S., et al. (2008). Mitochondrial DNA haplogroups M7b1'2 and M8a affect clinical expression of leber hereditary optic neuropathy in Chinese families with the m.11778G>a mutation. *Am. J. Hum. Genet.* 83, 760–768. doi: 10.1016/j.ajhg.2008.11.002
- Jiang, H., Shao, X., Jia, S., Qu, L., Weng, C., Shen, X., et al. (2019). The mitochondria-targeted metabolic tubular injury in diabetic kidney disease. *Cell Physiol. Biochem.* 52, 156–171. doi: 10.33594/000000011
- Jiang, W., Li, R., Zhang, Y., Wang, P., Wu, T., Lin, J., et al. (2017). Mitochondrial DNA mutations associated with type 2 diabetes mellitus in Chinese Uyghur population. *Sci. Rep.* 7:16989. doi: 10.1038/s41598-017-17086-7
- Jin, Z., Wei, W., Yang, M., Du, Y., and Wan, Y. (2014). Mitochondrial complex I activity suppresses inflammation and enhances bone resorption by shifting macrophage-osteoclast polarization. *Cell Metab.* 20, 483–498. doi: 10.1016/j.cmet.2014.07.011
- Kabekkodu, S. P., Bhat, S., Mascarenhas, R., Mallya, S., Bhat, M., Pandey, D., et al. (2014). Mitochondrial DNA variation analysis in cervical cancer. *Mitochondrion* 16, 73–82. doi: 10.1016/j.mito.2013.07.001
- Kasai, T., Ogawa, K., Mizuno, K., Nagai, S., Uchida, Y., Ohta, S., et al. (2002). Relationship between sperm mitochondrial membrane potential, sperm motility, and fertility potential. *Asian J. Androl.* 4, 97–103.
- Kato, T., Fuku, N., Noguchi, Y., Murakami, H., Miyachi, M., Kimura, Y., et al. (2012). Mitochondrial DNA haplogroup associated with hereditary hearing loss in a Japanese population. *Acta. Otolaryngol.* 132, 1178–1182. doi: 10.3109/00016489.2012.693624
- Khasawneh, J., Schulz, M. D., Walch, A., Rozman, J., Hrabe de Angelis, M., Klingenspor, M., et al. (2009). Inflammation and mitochondrial fatty acid beta-oxidation link obesity to early tumor promotion. *Proc. Natl. Acad. Sci. U.S.A.* 106, 3354–3359. doi: 10.1073/pnas.0802864106
- Kim, H. R., Kang, M. G., Lee, Y. E., Na, B. R., Noh, M. S., Yang, S. H., et al. (2018). Spectrum of mitochondrial genome instability and implication of mitochondrial haplogroups in Korean patients with acute myeloid leukemia. *Blood Res.* 53, 240–249. doi: 10.5045/br.2018.53.3.240
- Kim, W., Yoo, T. K., Shin, D. J., Rho, H. W., Jin, H. J., Kim, E. T., et al. (2008). Mitochondrial DNA haplogroup analysis reveals no association between the common genetic lineages and prostate cancer in the Korean population. *PLoS ONE* 3:e2211. doi: 10.1371/journal.pone.0002211
- King, M. P., and Attardi, G. (1989). Human cells lacking mtDNA: repopulation with exogenous mitochondria by complementation. *Science* 246, 500–503. doi: 10.1126/science.2814477
- Kong, Q. P., Bandelt, H. J., Sun, C., Yao, Y. G., Salas, A., Achilli, A., et al. (2006). Updating the East Asian mtDNA phylogeny: a prerequisite for the identification of pathogenic mutations. *Hum. Mol. Genet.* 15, 2076–2086. doi: 10.1093/hmg/ddl130
- Koo, B. S., Song, Y., Lee, S., Sung, Y. K., Shin, K. J., Cho, N. H., et al. (2019). Association of Asian mitochondrial DNA haplogroup B with new development of knee osteoarthritis in Koreans. *Int. J. Rheum. Dis.* 22, 411–416. doi: 10.1111/1756-185X.13453
- Kwak, S. H., and Park, K. S. (2016). Role of mitochondrial DNA variation in the pathogenesis of diabetes mellitus. *Front. Biosci.* 21, 1151–1167. doi: 10.2741/4447
- Lahera, V., de Las Heras, N., Lopez-Farre, A., Manucha, W., and Ferder, L. (2017). Role of mitochondrial dysfunction in hypertension and obesity. *Curr. Hypertens. Rep.* 19:11. doi: 10.1007/s11906-017-0710-9
- Lai, C. H., Huang, S. F., Chen, I. H., Liao, C. T., Wang, H. M., and Hsieh, L. L. (2012). The mitochondrial DNA Northeast Asia CZD haplogroup is associated with good disease-free survival among male oral squamous cell carcinoma patients. *PLoS ONE* 7:e49684. doi: 10.1371/journal.pone.0049684
- Lennon, F. E., and Salgia, R. (2014). Mitochondrial dynamics: biology and therapy in lung cancer. *Exp. Opin. Investig. Drugs* 23, 675–692. doi: 10.1517/13543784.2014.899350
- Levine, B., and Kroemer, G. (2008). Autophagy in the pathogenesis of disease. *Cell* 132, 27–42. doi: 10.1016/j.cell.2007.12.018

- Li, D., Sun, Y., Zhuang, Q., Song, Y., Wu, B., Jia, Z., et al. (2018). Mitochondrial dysfunction caused by m.2336T>C mutation with hypertrophic cardiomyopathy in cybrid cell lines. *Mitochondrion* 46, 313–320. doi: 10.1016/j.mito.2018.08.005
- Li, F. X., Ji, F. Y., Zheng, S. Z., Yao, W., Xiao, Z. L., and Qian, G. S. (2011). MtDNA haplogroups M7 and B in southwestern Han Chinese at risk for acute mountain sickness. *Mitochondrion* 11, 553–558. doi: 10.1016/j.mito.2011.02.003
- Li, H., Jiao, S., Li, X., Banu, H., Hamal, S., and Wang, X. (2017). Therapeutic effects of antibiotic drug mefloquine against cervical cancer through impairing mitochondrial function and inhibiting mTOR pathway. *Can. J. Physiol. Pharmacol.* 95, 43–50. doi: 10.1139/cjpp-2016-0124
- Li, M. Z., Yu, D. M., Yu, P., Liu, D. M., Wang, K., and Tang, X. Z. (2008). Mitochondrial gene mutations and type 2 diabetes in Chinese families. *Chin. Med. J.* 121, 682–686. doi: 10.1097/00029330-200804020-00004
- Li, Q., Zhang, T., Wang, J., Zhang, Z., Zhai, Y., Yang, G. Y., et al. (2014). Rapamycin attenuates mitochondrial dysfunction via activation of mitophagy in experimental ischemic stroke. *Biochem. Biophys. Res. Commun.* 444, 182–188. doi: 10.1016/j.bbrc.2014.01.032
- Li, W., Wen, C., Li, W., Wang, H., Guan, X., Zhang, W., et al. (2015). The tRNA(Gly) T10003C mutation in mitochondrial haplogroup M11b in a Chinese family with diabetes decreases the steady-state level of tRNA(Gly), increases aberrant reactive oxygen species production, and reduces mitochondrial membrane potential. *Mol. Cell. Biochem.* 408, 171–179. doi: 10.1007/s11010-015-2493-0
- Li, X. Y., Su, M., Huang, H. H., Li, H., Tian, D. P., and Gao, Y. X. (2007). mtDNA evidence: genetic background associated with related populations at high risk for esophageal cancer between Chaoshan and Taihang Mountain areas in China. *Genomics* 90, 474–481. doi: 10.1016/j.ygeno.2007.06.006
- Li, Y., Li, X., Wang, Z., Feng, Z., Li, L., and Ke, X. (2016). Subhaplogroup D4b1 enhances the risk of cervical cancer initiation: a case-control study in southern China. *J. Obstet. Gynaecol. Res.* 42, 325–330. doi: 10.1111/jog.12879
- Li, Z., Liu, Y., Yang, L., Wang, S., and Guan, M. X. (2008). Maternally inherited hypertension is associated with the mitochondrial tRNA(Ile) A4295G mutation in a Chinese family. *Biochem. Biophys. Res. Commun.* 367, 906–911. doi: 10.1016/j.bbrc.2007.12.150
- Liao, W. Q., Pang, Y., Yu, C. A., Wen, J. Y., Zhang, Y. G., and Li, X. H. (2008). Novel mutations of mitochondrial DNA associated with type 2 diabetes in Chinese Han population. *Tohoku J. Exp. Med.* 215, 377–384. doi: 10.1620/tjem.215.377
- Liao, Z., Zhao, J., Zhu, Y., Yang, L., Yang, A., Sun, D., et al. (2007). The ND4 G1169A mutation may influence the phenotypic manifestation of the deafness-associated 12S rRNA A1555G mutation in a four-generation Chinese family. *Biochem. Biophys. Res. Commun.* 362, 670–676. doi: 10.1016/j.bbrc.2007.08.034
- Lindqvist, D., Wolkowitz, O. M., Picard, M., Ohlsson, L., Bersani, F. S., Fernstrom, J., et al. (2018). Circulating cell-free mitochondrial DNA, but not leukocyte mitochondrial DNA copy number, is elevated in major depressive disorder. *Neuropsychopharmacology* 43, 1557–1564. doi: 10.1038/s41386-017-0001-9
- Liou, C. W., Chen, J. B., Tiao, M. M., Weng, S. W., Huang, T. L., Chuang, J. H., et al. (2012). Mitochondrial DNA coding and control region variants as genetic risk factors for type 2 diabetes. *Diabetes* 61, 2642–2651. doi: 10.2337/db11-1369
- Liou, C. W., Chuang, J. H., Chen, J. B., Tiao, M. M., Wang, P. W., Huang, S. T., et al. (2016). Mitochondrial DNA variants as genetic risk factors for Parkinson disease. *Eur. J. Neurol.* 23, 1289–1300. doi: 10.1111/ene.13020
- Liu, H., Li, S., Liu, X., Chen, Y., and Deng, H. (2018). SIRT3 overexpression inhibits growth of kidney tumor cells and enhances mitochondrial biogenesis. *J. Proteome Res.* 17, 3143–3152. doi: 10.1021/acs.jproteome.8b00260
- Liu, X. L., Zhou, X., Zhou, J., Zhao, F., Zhang, J., Li, C., et al. (2011). Leber's hereditary optic neuropathy is associated with the T12338C mutation in mitochondrial ND5 gene in six Han Chinese families. *Ophthalmology* 118, 978–985. doi: 10.1016/j.ophtha.2010.09.003
- Liu, Y., Gao, L., Li, Y., Li, Z., Xu, H., Wang, L., et al. (2009). Voltage-dependent anion channel (VDAC) is involved in apoptosis of cell lines carrying the mitochondrial DNA mutation. *BMC Med. Genet.* 10:114. doi: 10.1186/1471-2350-10-114
- Liu, Y., Li, Y., Zhu, C., Tian, L., Guan, M., and Chen, Y. (2017). Mitochondrial biogenesis dysfunction and metabolic dysfunction from a novel mitochondrial tRNA(Met) 4467 C>A mutation in a Han Chinese family with maternally inherited hypertension. *Sci. Rep.* 7:3034. doi: 10.1038/s41598-017-03303-w
- Liu, Z., Song, Y., Gu, S., He, X., Zhu, X., Shen, Y., et al. (2012). Mitochondrial ND5 12338T>C variant is associated with maternally inherited hypertrophic cardiomyopathy in a Chinese pedigree. *Gene* 506, 339–343. doi: 10.1016/j.gene.2012.06.071
- Loo, J. H., Trejaut, J. A., Yen, J. C., Chen, Z. S., Ng, W. M., Huang, C. Y., et al. (2014). Mitochondrial DNA association study of type 2 diabetes with or without ischemic stroke in Taiwan. *BMC Res.* 7:223. doi: 10.1186/1756-0500-7-223
- Lu, J., Qian, Y., Li, Z., Yang, A., Zhu, Y., Li, R., et al. (2010). Mitochondrial haplotypes may modulate the phenotypic manifestation of the deafness-associated 12S rRNA 1555A>G mutation. *Mitochondrion* 10, 69–81. doi: 10.1016/j.mito.2009.09.007
- Lu, M. Y., Huang, J. F., Liao, Y. C., Bai, R. K., Trieu, R. B., Chuang, W. L., et al. (2012). Mitochondrial polymorphism 12361A>G is associated with nonalcoholic fatty liver disease. *Transl. Res.* 159, 58–59. doi: 10.1016/j.trsl.2011.10.011
- Lu, Z., Chen, H., Meng, Y., Wang, Y., Xue, L., Zhi, S., et al. (2011). The tRNAMet 4435A>G mutation in the mitochondrial haplogroup G2a1 is responsible for maternally inherited hypertension in a Chinese pedigree. *Eur. J. Hum. Genet.* 19, 1181–1186. doi: 10.1038/ejhg.2011.111
- Lyu, L., Wang, Q., Song, S., Li, L., Zhou, H., Li, M., et al. (2019). Oncocytic tumors are marked by enhanced mitochondrial content and mtDNA mutations of complex I in Chinese patients. *Mitochondrion* 45, 1–6. doi: 10.1016/j.mito.2018.01.008
- Ma, L., Fu, Q., Xu, B., Zhou, H., Gao, J., Shao, X., et al. (2018). Breast cancer-associated mitochondrial DNA haplogroup promotes neoplastic growth via ROS-mediated AKT activation. *Int. J. Cancer* 142, 1786–1796. doi: 10.1002/ijc.31207
- Ma, X., Cui, Y., Zhou, H., and Li, Q. (2018). Function of mitochondrial pyruvate carriers in hepatocellular carcinoma patients. *Oncol. Lett.* 15, 9110–9116. doi: 10.3892/ol.2018.8466
- Madsen-Bouterse, S. A., Zhong, Q., Mohammad, G., Ho, Y. S., and Kowluru, R. A. (2010). Oxidative damage of mitochondrial DNA in diabetes and its protection by manganese superoxide dismutase. *Free Radic. Res.* 44, 313–321. doi: 10.3109/10715760903494168
- Marchetti, C., Obert, G., Deffosez, A., Formstecher, P., and Marchetti, P. (2002). Study of mitochondrial membrane potential, reactive oxygen species, DNA fragmentation and cell viability by flow cytometry in human sperm. *Hum. Reprod.* 17, 1257–1265. doi: 10.1093/humrep/17.5.1257
- Maron, B. J., Towbin, J. A., Thiene, G., Antzelevitch, C., Corrado, D., Arnett, D., et al. (2006). Contemporary definitions and classification of the cardiomyopathies: an American Heart Association Scientific Statement from the Council on Clinical Cardiology, Heart Failure and Transplantation Committee; Quality of Care and Outcomes Research and Functional Genomics and Translational Biology Interdisciplinary Working Groups; and Council on Epidemiology and Prevention. *Circulation* 113, 1807–1816. doi: 10.1161/CIRCULATIONAHA.106.174287
- Mizumachi, T., Muskhelishvili, L., Naito, A., Furusawa, J., Fan, C. Y., Siegel, E. R., et al. (2008). Increased distributional variance of mitochondrial DNA content associated with prostate cancer cells as compared with normal prostate cells. *Prostate* 68, 408–417. doi: 10.1002/pros.20697
- Moore, F. L., and Reijo-Pera, R. A. (2000). Male sperm motility dictated by mother's mtDNA. *Am. J. Hum. Genet.* 67, 543–548. doi: 10.1086/303061
- Munakata, K., Tanaka, M., Mori, K., Washizuka, S., Yoneda, M., Tajima, O., et al. (2004). Mitochondrial DNA 3644T>C mutation associated with bipolar disorder. *Genomics* 84, 1041–1050. doi: 10.1016/j.ygeno.2004.08.015
- Nakano, S., Fukuda, M., Hotta, F., Ito, T., Ishii, T., Kitazawa, M., et al. (1998). Mitochondrial DNA point mutation at nucleotide pair 3316 in a Japanese family with heterogeneous phenotypes of diabetes. *Endocr. J.* 45, 625–630. doi: 10.1507/endocrj.45.625
- Nishigaki, Y., Fuku, N., and Tanaka, M. (2010). Mitochondrial haplogroups associated with lifestyle-related diseases and longevity in the Japanese population. *Geriatr. Gerontol. Int.* 10 (Suppl 1), S221–235. doi: 10.1111/j.1447-0594.2010.00599.x
- Nishigaki, Y., Yamada, Y., Fuku, N., Matsuo, H., Segawa, T., Watanabe, S., et al. (2007a). Mitochondrial haplogroup A is a genetic risk factor for atherosclerotic cerebral infarction in Japanese females. *Mitochondrion* 7, 72–79. doi: 10.1016/j.mito.2006.11.002

- Nishigaki, Y., Yamada, Y., Fuku, N., Matsuo, H., Segawa, T., Watanabe, S., et al. (2007b). Mitochondrial haplogroup N9b is protective against myocardial infarction in Japanese males. *Hum. Genet.* 120, 827–836. doi: 10.1007/s00439-006-0269-z
- Niu, Q., Zhang, W., Wang, H., Guan, X., Lu, J., and Li, W. (2015). Effects of mitochondrial haplogroup N9a on type 2 diabetes mellitus and its associated complications. *Exp. Ther. Med.* 10, 1918–1924. doi: 10.3892/etm.2015.2751
- Ohkubo, R., Nakagawa, M., Ikeda, K., Kodama, T., Arimura, K., Akiba, S., et al. (2002). Cerebrovascular disorders and genetic polymorphisms: mitochondrial DNA5178C is predominant in cerebrovascular disorders. *J. Neurol. Sci.* 198, 31–35. doi: 10.1016/S0022-510X(02)00055-2
- Pelicano, H., Zhang, W., Liu, J., Hammoudi, N., Dai, J., Xu, R. H., et al. (2014). Mitochondrial dysfunction in some triple-negative breast cancer cell lines: role of mTOR pathway and therapeutic potential. *Breast Cancer Res.* 16:434. doi: 10.1186/s13058-014-0434-6
- Pello, R., Martin, M. A., Carelli, V., Nijtmans, L. G., Achilli, A., Pala, M., et al. (2008). Mitochondrial DNA background modulates the assembly kinetics of OXPHOS complexes in a cellular model of mitochondrial disease. *Hum. Mol. Genet.* 17, 4001–4011. doi: 10.1093/hmg/ddn303
- Pera, M., Larrea, D., Guardia-Laguarta, C., Montesinos, J., Velasco, K. R., Agrawal, R. R., et al. (2017). Increased localization of APP-C99 in mitochondria-associated ER membranes causes mitochondrial dysfunction in Alzheimer disease. *EMBO J.* 36, 3356–3371. doi: 10.1525/embj.201796797
- Pereira, C., Chavarria, V., Vian, J., Ashton, M. M., Berk, M., Marx, W., et al. (2018). Mitochondrial agents for bipolar disorder. *Int. J. Neuropsychopharmacol.* 21, 550–569. doi: 10.1093/ijnp/ppy018
- Piekutowska-Abramczuk, D., Assouline, Z., Matakovic, L., Feichtinger, R. G., Konarikova, E., Jurkiewicz, E., et al. (2018). NDUFB8 mutations cause mitochondrial complex I deficiency in individuals with Leigh-like encephalomyopathy. *Am. J. Hum. Genet.* 102, 460–467. doi: 10.1016/j.ajhg.2018.01.008
- Puthumana, J. S., and Regenold, W. T. (2019). Glucose-6-phosphate dehydrogenase activity in bipolar disorder and schizophrenia: relationship to mitochondrial impairment. *J. Psychiatr. Res.* 112, 99–103. doi: 10.1016/j.jpsychires.2019.03.004
- Qi, W., Keenan, H. A., Li, Q., Ishikado, A., Kannt, A., Sadowski, T., et al. (2017). Pyruvate kinase M2 activation may protect against the progression of diabetic glomerular pathology and mitochondrial dysfunction. *Nat. Med.* 23, 753–762. doi: 10.1038/nm.4328
- Qian, Y., Zhou, X., Hu, Y., Tong, Y., Li, R., Lu, F., et al. (2005). Clinical evaluation and mitochondrial DNA sequence analysis in three Chinese families with Leber's hereditary optic neuropathy. *Biochem. Biophys. Res. Commun.* 332, 614–621. doi: 10.1016/j.bbrc.2005.05.003
- Qiao, Z., Cheng, Y., Liu, S., Ma, Z., Li, S., and Zhang, W. (2019). Casticin inhibits esophageal cancer cell proliferation and promotes apoptosis by regulating mitochondrial apoptotic and JNK signaling pathways. *Naunyn Schmiedebergs. Arch. Pharmacol.* 392, 177–187. doi: 10.1007/s00210-018-1574-5
- Qu, J., Li, R., Zhou, X., Tong, Y., Lu, F., Qian, Y., et al. (2006). The novel A4435G mutation in the mitochondrial tRNAMet may modulate the phenotypic expression of the LHON-associated ND4 G11778A mutation. *Invest. Ophthalmol. Vis. Sci.* 47, 475–483. doi: 10.1167/iov.05-0665
- Qu, J., Wang, Y., Tong, Y., Zhou, X., Zhao, F., Yang, L., et al. (2010). Leber's hereditary optic neuropathy affects only female matrilineal relatives in two Chinese families. *Invest. Ophthalmol. Vis. Sci.* 51, 4906–4912. doi: 10.1167/iov.09-5027
- Rehman, J., Zhang, H. J., Toth, P. T., Zhang, Y., Marsboom, G., Hong, Z., et al. (2012). Inhibition of mitochondrial fission prevents cell cycle progression in lung cancer. *FASEB J.* 26, 2175–2186. doi: 10.1096/fj.11-196543
- Rogge, M. M. (2009). The role of impaired mitochondrial lipid oxidation in obesity. *Biol. Res. Nurs.* 10, 356–373. doi: 10.1177/1099800408329408
- Roglic, G., and Unwin, N. (2010). Mortality attributable to diabetes: estimates for the year 2010. *Diabetes Res. Clin. Pract.* 87, 15–19. doi: 10.1016/j.diabres.2009.10.006
- Rollins, B. L., Morgan, L., Hjelm, B. E., Sequeira, A., Schatzberg, A. F., Barchas, J. D., et al. (2018). Mitochondrial complex I deficiency in Schizophrenia and bipolar disorder and medication influence. *Mol. Neuropsychiatr.* 3, 157–169. doi: 10.1159/000484348
- Ruiz-Pesini, E., Lapena, A. C., Diez-Sanchez, C., Perez-Martos, A., Montoya, J., Alvarez, E., et al. (2000). Human mtDNA haplogroups associated with high or reduced spermatozoa motility. *Am. J. Hum. Genet.* 67, 682–696. doi: 10.1086/303040
- Santidrian, A. F., Matsuno-Yagi, A., Ritland, M., Seo, B. B., LeBoeuf, S. E., Gay, L. J., et al. (2013). Mitochondrial complex I activity and NAD+/NADH balance regulate breast cancer progression. *J. Clin. Invest.* 123, 1068–1081. doi: 10.1172/JCI64264
- Sawabe, M., Tanaka, M., Chida, K., Arai, T., Nishigaki, Y., Fuku, N., et al. (2011). Mitochondrial haplogroups A and M7a confer a genetic risk for coronary atherosclerosis in the Japanese elderly: an autopsy study of 1,536 patients. *J. Atheroscler. Thromb.* 18, 166–175. doi: 10.5551/jat.6742
- Schrauwen, P., and Hesselink, M. K. (2004). Oxidative capacity, lipotoxicity, and mitochondrial damage in type 2 diabetes. *Diabetes* 53, 1412–1417. doi: 10.2337/diabetes.53.6.1412
- Sharma, L. K., Tiwari, M., Rai, N. K., and Bai, Y. (2019). Mitophagy activation repairs Leber's hereditary optic neuropathy-associated mitochondrial dysfunction and improves cell survival. *Hum. Mol. Genet.* 28, 422–433. doi: 10.1093/hmg/ddy354
- Shenouda, S. M., Widlansky, M. E., Chen, K., Xu, G., Holbrook, M., Tabit, C. E., et al. (2011). Altered mitochondrial dynamics contributes to endothelial dysfunction in diabetes mellitus. *Circulation* 124, 444–453. doi: 10.1161/CIRCULATIONAHA.110.014506
- Shi, W. Y., Cao, C., and Liu, L. (2016). Interferon  $\alpha$  induces the apoptosis of cervical cancer HeLa cells by activating both the intrinsic mitochondrial pathway and endoplasmic reticulum stress-induced pathway. *Int. J. Mol. Sci.* 17:1832. doi: 10.3390/ijms17111832
- Shoffner, J. M., Lott, M. T., Lezza, A. M., Seibel, P., Ballinger, S. W., and Wallace, D. C. (1990). Myoclonic epilepsy and ragged-red fiber disease (MERRF) is associated with a mitochondrial DNA tRNA(Lys) mutation. *Cell* 61, 931–937. doi: 10.1016/0092-8674(90)90059-N
- Smith, R. A., Hartley, R. C., Cocheme, H. M., and Murphy, M. P. (2012). Mitochondrial pharmacology. *Trends Pharmacol. Sci.* 33, 341–352. doi: 10.1016/j.tips.2012.03.010
- Sun, X., Cao, H., Zhan, L., Yin, C., Wang, G., Liang, P., et al. (2018). Mitochondrial fission promotes cell migration by Ca(2+)/CaMKII/ERK/FAK pathway in hepatocellular carcinoma. *Liver Int.* 38, 1263–1272. doi: 10.1111/liv.13660
- Susztak, K., Raff, A. C., Schiffer, M., and Bottiger, E. P. (2006). Glucose-induced reactive oxygen species cause apoptosis of podocytes and podocyte depletion at the onset of diabetic nephropathy. *Diabetes* 55, 225–233. doi: 10.2337/diabetes.55.01.06.db05-0894
- Takagi, K., Yamada, Y., Gong, J. S., Sone, T., Yokota, M., and Tanaka, M. (2004). Association of 5178C->A (Leu237Met) polymorphism in the mitochondrial DNA with a low prevalence of myocardial infarction in Japanese individuals. *Atherosclerosis* 175, 281–286. doi: 10.1016/j.atherosclerosis.2004.03.008
- Takasaki, S. (2008). Mitochondrial SNPs associated with Japanese centenarians, Alzheimer's patients, and Parkinson's patients. *Comput. Biol. Chem.* 32, 332–337. doi: 10.1016/j.combiolchem.2008.03.014
- Takasaki, S. (2009). Mitochondrial haplogroups associated with Japanese Alzheimer's patients. *J. Bioenerg. Biomembr.* 41, 407–410. doi: 10.1007/s10863-009-9240-8
- Tanaka, M., Fuku, N., Nishigaki, Y., Matsuo, H., Segawa, T., Watanabe, S., et al. (2007). Women with mitochondrial haplogroup N9a are protected against metabolic syndrome. *Diabetes* 56, 518–521. doi: 10.2337/db06-1105
- Tang, S., Batra, A., Zhang, Y., Ebenroth, E. S., and Huang, T. (2010). Left ventricular noncompaction is associated with mutations in the mitochondrial genome. *Mitochondrion* 10, 350–357. doi: 10.1016/j.mito.2010.02.003
- Tian, S., Chen, H., and Tan, W. (2018). Targeting mitochondrial respiration as a therapeutic strategy for cervical cancer. *Biochem. Biophys. Res. Commun.* 499, 1019–1024. doi: 10.1016/j.bbrc.2018.04.042
- Ueshima, H., Sekikawa, A., Miura, K., Turin, T. C., Takashima, N., Kita, Y., et al. (2008). Cardiovascular disease and risk factors in Asia: a selected review. *Circulation* 118, 2702–2709. doi: 10.1161/CIRCULATIONAHA.108.790048
- van Oven, M., and Kayser, M. (2009). Updated comprehensive phylogenetic tree of global human mitochondrial DNA variation. *Hum. Mutat.* 30, E386–394. doi: 10.1002/humu.20921
- van Rahden, V. A., Fernandez-Vizarra, E., Alawi, M., Brand, K., Fellmann, F., Horn, D., et al. (2015). Mutations in NDUFB11, encoding a complex

- I component of the mitochondrial respiratory chain, cause microphthalmia with linear skin defects syndrome. *Am. J. Hum. Genet.* 96, 640–650. doi: 10.1016/j.ajhg.2015.02.002
- Vats, D., Mukundan, L., Odegaard, J. I., Zhang, L., Smith, K. L., Morel, C. R., et al. (2006). Oxidative metabolism and PGC-1beta attenuate macrophage-mediated inflammation. *Cell Metab.* 4, 13–24. doi: 10.1016/j.cmet.2006.05.011
- Verma, P., Singh, A., Nthenge-Ngumbau, D. N., Rajamma, U., Sinha, S., Mukhopadhyay, K., et al. (2016). Attention deficit-hyperactivity disorder suffers from mitochondrial dysfunction. *BBA Clin.* 6, 153–158. doi: 10.1016/j.bbaci.2016.10.003
- Victor, V. M., Rocha, M., Herance, R., and Hernandez-Mijares, A. (2011). Oxidative stress and mitochondrial dysfunction in type 2 diabetes. *Curr. Pharm. Des.* 17, 3947–3958. doi: 10.2174/138161211798764915
- Wallace, D. C. (2000). Mitochondrial defects in cardiomyopathy and neuromuscular disease. *Am. Heart J.* 139(2 Pt 3), S70–S85. doi: 10.1067/mhj.2000.103934
- Wallace, D. C. (2005). Mitochondria and cancer: warburg addressed. *Cold Spring Harb. Symp. Quant. Biol.* 70, 363–374. doi: 10.1101/sqb.2005.70.035
- Wallace, D. C. (2013). A mitochondrial bioenergetic etiology of disease. *J. Clin. Invest.* 123, 1405–1412. doi: 10.1172/JCI61398
- Wallace, D. C., Singh, G., Lott, M. T., Hodge, J. A., Schurr, T. G., Lezza, A. M., et al. (1988a). Mitochondrial DNA mutation associated with Leber's hereditary optic neuropathy. *Science* 242, 1427–1430. doi: 10.1126/science.3201231
- Wallace, D. C., Zheng, X. X., Lott, M. T., Shoffner, J. M., Hodge, J. A., Kelley, R. I., et al. (1988b). Familial mitochondrial encephalomyopathy (MERRF): genetic, pathophysiological, and biochemical characterization of a mitochondrial DNA disease. *Cell* 55, 601–610. doi: 10.1016/0092-8674(88)90218-8
- Wang, C. Y., Kong, Q. P., Yao, Y. G., and Zhang, Y. P. (2006). mtDNA mutation C1494T, haplogroup A, and hearing loss in Chinese. *Biochem. Biophys. Res. Commun.* 348, 712–715. doi: 10.1016/j.bbrc.2006.07.119
- Wang, D., Taniyama, M., Suzuki, Y., Katagiri, T., and Ban, Y. (2001). Association of the mitochondrial DNA 5178A/C polymorphism with maternal inheritance and onset of type 2 diabetes in Japanese patients. *Exp. Clin. Endocrinol. Diabetes* 109, 361–364. doi: 10.1055/s-2001-17407
- Wang, S., Wu, S., Zheng, T., Yang, Z., Ma, X., Jia, W., et al. (2013). Mitochondrial DNA mutations in diabetes mellitus patients in Chinese Han population. *Gene* 531, 472–475. doi: 10.1016/j.gene.2013.09.019
- Wang, X., Guo, Y., and Luan, Q. (2015). Association of mitochondrial DNA displacement loop polymorphisms and aggressive periodontitis in a Chinese population: a pilot study. *Mitochondrial DNA* 26, 389–395. doi: 10.3109/19401736.2013.840589
- Wang, X., Su, B., Perry, G., Smith, M. A., and Zhu, X. (2007). Insights into amyloid-beta-induced mitochondrial dysfunction in Alzheimer disease. *Free Radic. Biol. Med.* 43, 1569–1573. doi: 10.1016/j.freeradbiomed.2007.09.007
- Wang, Z., Hao, G., Wang, X., Chen, Z., Zhang, L., Zhang, Z., et al. (2019). Clinical outcomes and economic impact of the 2017 ACC/AHA guidelines on hypertension in China. *J. Clin. Hypertens.* 21, 1212–1220. doi: 10.1111/jch.13609
- Wang, Z., Ying, Z., Bosy-Westphal, A., Zhang, J., Schautz, B., Later, W., et al. (2010). Specific metabolic rates of major organs and tissues across adulthood: evaluation by mechanistic model of resting energy expenditure. *Am. J. Clin. Nutr.* 92, 1369–1377. doi: 10.3945/ajcn.2010.29885
- Warburg, O. (1956). On the origin of cancer cells. *Science* 123, 309–314. doi: 10.1126/science.123.3191.309
- Wei, P. Z., and Szeto, C. C. (2019). Mitochondrial dysfunction in diabetic kidney disease. *Clin. Chim. Acta* 496, 108–116. doi: 10.1016/j.cca.2019.07.005
- Wei, Y. L., Yu, C. A., Yang, P., Li, A. L., Wen, J. Y., Zhao, S. M., et al. (2009). Novel mitochondrial DNA mutations associated with Chinese familial hypertrophic cardiomyopathy. *Clin. Exp. Pharmacol. Physiol.* 36, 933–939. doi: 10.1111/j.1440-1681.2009.05183.x
- Wilkins, H. M., Carl, S. M., and Swerdlow, R. H. (2014). Cytoplasmic hybrid (cybrid) cell lines as a practical model for mitochondriopathies. *Redox Biol.* 2, 619–631. doi: 10.1016/j.redox.2014.03.006
- Wu, D., Zhang, J., Wang, J., Li, J., Liao, F., and Dong, W. (2016). Hesperetin induces apoptosis of esophageal cancer cells via mitochondrial pathway mediated by the increased intracellular reactive oxygen species. *Tumour Biol.* 37, 3451–3459. doi: 10.1007/s13277-015-4176-6
- Xing, Y., Ichida, F., Matsuoka, T., Isobe, T., Ikemoto, Y., Higaki, T., et al. (2006). Genetic analysis in patients with left ventricular noncompaction and evidence for genetic heterogeneity. *Mol. Genet. Metab.* 88, 71–77. doi: 10.1016/j.ymgme.2005.11.009
- Xu, L., Hu, Y., Chen, B., Tang, W., Han, X., Yu, H., et al. (2006). Mitochondrial polymorphisms as risk factors for endometrial cancer in southwest China. *Int. J. Gynecol. Cancer* 16, 1661–1667. doi: 10.1111/j.1525-1438.2006.00641.x
- Xu, T., Pang, Q., Wang, Y., and Yan, X. (2017). Betulinic acid induces apoptosis by regulating PI3K/Akt signaling and mitochondrial pathways in human cervical cancer cells. *Int. J. Mol. Med.* 40, 1669–1678. doi: 10.3892/ijmm.2017.3163
- Xu, W., Mi, Y., He, P., He, S., and Niu, L. (2017).  $\gamma$ -tocotrienol inhibits proliferation and induces apoptosis via the mitochondrial pathway in human cervical cancer HeLa cells. *Molecules* 22:E1299. doi: 10.3390/molecules22081299
- Xue, F., Wang, Y., Xu, S., Zhang, F., Wen, B., Wu, X., et al. (2008). A spatial analysis of genetic structure of human populations in China reveals distinct difference between maternal and paternal lineages. *Eur. J. Hum. Genet.* 16, 705–717. doi: 10.1038/sj.ejhg.5201998
- Xue, H., Li, P., Luo, Y., Wu, C., Liu, Y., Qin, X., et al. (2019). Salidroside stimulates the Sirt1/PGC-1 $\alpha$  axis and ameliorates diabetic nephropathy in mice. *Phytomedicine* 54, 240–247. doi: 10.1016/j.phymed.2018.10.031
- Yang, D., Wang, Q., Shi, Y., Fan, Y., Zheng, H. X., Song, G., et al. (2014). Mitochondrial DNA haplogroup D4b is a protective factor for ischemic stroke in Chinese Han population. *Mol. Genet. Genomics* 289, 1241–1246. doi: 10.1007/s00438-014-0884-7
- Ying, Z., Zheng, J., Cai, Z., Liu, L., Dai, Y., Yao, J., et al. (2015). Mitochondrial haplogroup B increases the risk for hearing loss among the Eastern Asian pedigrees carrying 12S rRNA 1555A>G mutation. *Protein Cell* 6, 844–848. doi: 10.1007/s13238-015-0203-z
- Yoneda, M., Tsuji, S., Yamauchi, T., Inuzuka, T., Miyatake, T., Horai, S., et al. (1989). Mitochondrial DNA mutation in family with Leber's hereditary optic neuropathy. *Lancet* 1, 1076–1077. doi: 10.1016/S0140-6736(89)92470-7
- Youle, R. J., and Narendra, D. P. (2011). Mechanisms of mitophagy. *Nat. Rev. Mol. Cell Biol.* 12, 9–14. doi: 10.1038/nrm3028
- Young, W. Y., Zhao, L., Qian, Y., Wang, Q., Li, N., Greinwald, J. H. Jr., et al. (2005). Extremely low penetrance of hearing loss in four Chinese families with the mitochondrial 12S rRNA A1555G mutation. *Biochem. Biophys. Res. Commun.* 328, 1244–1251. doi: 10.1016/j.bbrc.2005.01.085
- Yu, D., Jia, X., Zhang, A. M., Li, S., Zou, Y., Zhang, Q., et al. (2010). Mitochondrial DNA sequence variation and haplogroup distribution in Chinese patients with LHON and m.1448AT>C. *PLoS ONE* 5:e13426. doi: 10.1371/journal.pone.0013426
- Yuan, H., Chen, J., Liu, X., Cheng, J., Wang, X., Yang, L., et al. (2007). Coexistence of mitochondrial 12S rRNA C1494T and CO1/tRNA(Ser(UCN)) G7444A mutations in two Han Chinese pedigrees with aminoglycoside-induced and non-syndromic hearing loss. *Biochem. Biophys. Res. Commun.* 362, 94–100. doi: 10.1016/j.bbrc.2007.07.161
- Zhai, K., Chang, L., Zhang, Q., Liu, B., and Wu, Y. (2011). Mitochondrial C150T polymorphism increases the risk of cervical cancer and HPV infection. *Mitochondrion* 11, 559–563. doi: 10.1016/j.mito.2011.02.005
- Zhan, L., Cao, H., Wang, G., Lyu, Y., Sun, X., An, J., et al. (2016). Drp1-mediated mitochondrial fission promotes cell proliferation through crosstalk of p53 and NF-kappaB pathways in hepatocellular carcinoma. *Oncotarget* 7, 65001–65011. doi: 10.18632/oncotarget.11339
- Zhang, J., Zhang, Z. X., Du, P. C., Zhou, W., Wu, S. D., Wang, Q. L., et al. (2015). Analyses of the mitochondrial mutations in the Chinese patients with sporadic Creutzfeldt-Jakob disease. *Eur. J. Hum. Genet.* 23, 86–91. doi: 10.1038/ejhg.2014.52
- Zhang, R., Zhang, F., Wang, C., Wang, S., Shiao, Y. H., and Guo, Z. (2010). Identification of sequence polymorphism in the D-Loop region of mitochondrial DNA as a risk factor for hepatocellular carcinoma with distinct etiology. *J. Exp. Clin. Cancer Res.* 29:130. doi: 10.1186/1756-9966-29-130
- Zhang, X. H., Guan, T. R., Mao, J. W., and Liu, L. S. (2007). Disparity and its time trends in stroke mortality between urban and rural populations in China 1987 to 2001 - Changing patterns and their implications for public health policy. *Stroke* 38, 3139–3144. doi: 10.1161/STROKEAHA.107.494336
- Zhang, Y., Zhao, Y., Wen, S., Yan, R., Yang, Q., and Chen, H. (2017). Associations of mitochondrial haplogroups and mitochondrial DNA copy numbers with

- end-stage renal disease in a Han population. *Mitochondrial DNA A DNA Mapp. Seq. Anal.* 28, 725–731. doi: 10.1080/24701394.2016.1177038
- Zheng, S., Qian, P., Li, F., Qian, G., Wang, C., Wu, G., et al. (2012). Association of mitochondrial DNA variations with lung cancer risk in a Han Chinese population from southwestern China. *PLoS ONE* 7:e31322. doi: 10.1371/journal.pone.0031322
- Zhong, L., Tang, J., Kong, Q. P., Sun, C., Zhou, W. P., Yang, M., et al. (2014). Reappraising the relationship between mitochondrial DNA variant m.16189T>C and type 2 diabetes mellitus in East Asian populations. *Curr. Mol. Med.* 14, 1273–1278. doi: 10.2174/1566524014666141202161326
- Zhong, Z., Liang, S., Sanchez-Lopez, E., He, F., Shalapour, S., Lin, X. J., et al. (2018). New mitochondrial DNA synthesis enables NLRP3 inflammasome activation. *Nature* 560, 198–203. doi: 10.1038/s41586-018-0372-z
- Zhou, X., Zhang, H., Zhao, F., Ji, Y., Tong, Y., Zhang, J., et al. (2010). Very high penetrance and occurrence of Leber's hereditary optic neuropathy in a large Han Chinese pedigree carrying the ND4 G11778A mutation. *Mol. Genet. Metab.* 100, 379–384. doi: 10.1016/j.ymgme.2010.04.013
- Zhu, J. F., Zhang, X., and Ling, L. (2016). Molecular characterization of a Han Chinese family with essential hypertension. *Genet. Mol. Res.* 15:gmr.15028084. doi: 10.4238/gmr.15028084
- Zimmet, P., Alberti, K. G., and Shaw, J. (2001). Global and societal implications of the diabetes epidemic. *Nature* 414, 782–787. doi: 10.1038/414782a

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Sun, Wei, Zheng, Jin and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.