



Green Approach: "A Forwarding Step for Curing Leishmaniasis—A Neglected Tropical Disease"

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The present review focuses on a dreaded vector-mediated leishmaniasis, with the existing therapeutic approaches including a variety of drugs along with their limitations, the treatment with natural compounds, and different types of metal/metal oxide nanoparticles (NPs). As evidenced, various metallic NPs, comprising silver, silver oxide, gold, zinc oxide, titanium, lead oxide, *etc.*, played a curative role to treat leishmaniasis, are also presented. Keeping in view the advance success of vaccines against the prevalent dreaded diseases in the past and the present scenario, efforts are also being made to develop vaccines based on these NP formulations.

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INTRODUCTION

Neglected tropical diseases (NTDs) are contagious diseases that cause substantial illness in more than one billion people globally (Maheshwari and Bandyopadhyay, 2020). Various parasite-mediated diseases comprising giardiasis, Chagas disease, Babesiosis, toxoplasmosis, leishmaniasis, etc., befall in animals and further spread to human population (Oryan, 2015; Hotez et al., 2020). Leishmaniasis is one of the NTDs considered as imperative parasitic diseases, commonly caused by an etiologic agent Leishmania, a genus of trypanosomes. Leishmaniasis is located in the ninth place of the global burden of disease among individual infectious diseases. More than 22 species of infectious Leishmania have been reported (Maheshwari and Bandyopadhyay, 2020). Leishmania are transmitted to mammals through the bite of infected female sandflies belonging to Lutzomyia and Phlebotomus (Oryan and Akbari, 2016). It is endemic in 98 nations of the world, where more than 350 million people are at risk and more than 12 million cases of infection have been reported (Verma and Dey, 2004; Mcgwire and Satoskar, 2014). Based on the species and intensity of infection to the host, it has been classified into cutaneous leishmaniasis (CL), mucocutaneous leishmaniasis (MCL), and visceral leishmaniasis (VL) forms. Considering these classes, CL caused by L. aethiopica is most commonly found in human population and reported to infect 6,00,000 to one million people annually all around the world. It causes severe symptoms like ulcers, serious disabilities, and life-long marks (Surur et al., 2020). Previously, various chemical drugs including liposomal amphotericin B, amphotericin B, pentamidine, pentavalent antimonials, miltefosine, and paromomycin have been practiced against Leishmania. Among these drugs, pentavalent antimonials (sodium stibogluconate and meglumine) are existing chemical drugs, and they are a major therapeutic source to treat Leishmania infection. However, current treatment practices are associated with certain side effects like high toxicity, high cost, and most importantly, development of drug resistance. Hence, there is an instantaneous necessity to innovate new, harmless, and efficient prevention therapies to overcome these limitations (Mitropoulos et al., 2010). Currently, various approaches are involved to control the elevated level of infection, including nanoformulations and targeted drug

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delivery using nanocarriers as well as with the aid of particular bioactive compound obtained from plants (Javed et al., 2020; Santana et al., 2020). Inventions in nanoscience greatly contribute to overcome the problems allied with the treatment of infectious diseases. Owing to the small size of NPs (1-100 nm), the ability to penetrate easily into the cells, extensive circulation within the body, and the efficient targeted drug delivery system, they can be reflected as a better medication to treat endemic leishmaniasis (Ebrahimi et al., 2017). Further, plant-based nanoparticles have been reported as a successful approach for the preclusion of microbial infections as well as in the treatment of leishmaniasis (Alti et al., 2020). Owing to the green and eco-friendly nature, cost effectiveness, less hazardous nature, and involvement of phytoconstituents for capping and stabilization, plant-based metal oxide nanoparticles (NPs) (silver, zinc, nickel, iron, etc.) have been in use to cure leishmanial infection (Ismail et al., 2019; Alti et al., 2020). The present review focuses on the use of plantbased natural products, the phytosynthesized NPs, to cure Leishmania.

LIFE CYCLE

The life cycle of *Leishmania* parasite begins with the bite of infected female phlebotomine sandflies and is completed in two different morphological forms, that is, promastigote and amastigote. Flagellated metacyclic promastigote, formed in the infected sandflies firstly enter into the macrophages via phagocytosis and formed phagosome. Resultant phagosome

into of (membrane enter the stages maturation formed a new structure: the transformation) and parasitophorous vacuole. In this organelle, promastigotes metamorphose into amastigote, followed by its multiplication until explosion of the cells of the macrophage system $(4-6^{\circ} days)$ spreads infection further. The parasite either initiates infection superficial cells or visceral cell depending on its tropism characteristics. The life cycle of Leishmania into the host is completed when another uninfected sandfly sucks blood as a source of meal. The sensitivity of infection is based on the sandfly species, ecology, epidemiology, and pathogenicity (Bañuls et al., 2007). Further, detailed description of its infection period and transmission is clearly shown in Figure 1.

EXISTING THERAPEUTIC APPROACHES AND LIMITATIONS

Leishmaniasis is one of the most important NTDs coupled with various adverse as well as life-threating factors, including substantial morbidity, early death, and long-term infirmity. Treatment comprises control of disease spreading and use of existing parameters, while the currently used therapies including chemical drugs necessitate long-duration therapy and low efficiency with numerous toxic effects. Although no relevant therapies have been developed to prevent the infection which is extensively spread among human population, only few prevention methods are available (Gharirvand Eskandari et al., 2020). Among them, some kind of clinically approved drugs are



found to treat this endemic disease, including meglumine antimoniate (glucatime), sodium stibogluconate (pentostan), amphotericin B, and miltefosine. However, the excessive use of these chemotherapeutic sources is associated with antagonistic effects (Ghobakhloo et al., 2016). This has led to the search of some natural methods to treat leishmaniasis.

Chemical-Based Drugs for the Treatment of Leishmaniasis

Since last several years, various kinds of pharmaceutical drugs including amphotericin B, pentamidine, miltefosine, and paromomycin were involved in the treatment of leishmaniasis. None of the clinically approved drugs could be deliberated as the ultimate source of treatment due to their time-taking process and high toxicity combined with severe adversative effects. In addition, the most often used medicines do not eradicate the parasites entirely from all infected entities (De Menezes et al., 2015). Further, applications of some of these medicines with their limitations are described below:

- Pentavalent antimonials can be administrated by the intravenous, intramuscular, and intralymphatic routes with the optimum dosage of 20 mg/kg/day (28–30 days) and exhibited 35–95% potentiality. Continuous and excessive use of this drug causes toxicity like nephrotoxicity, hepatotoxicity, severe cardiotoxicity, and pancreatitis (De Menezes et al., 2015).
- Oral administration of miltefosine not only showed inhibitory effects on the growth of *Leishmania* but also affected adversely and created severe infecting symptoms comprising nephrotoxicity, teratogenicity, vomiting and diarrhea, and hepatotoxicity (Sundar et al., 2011).
- Paromomycin, also being used as a therapeutic agent to treat leishmaniasis, reported to show some toxic effects during its treatment phase, like severe nephrotoxicity, hepatotoxicity, and ototoxicity (Jhingran et al., 2009).
- Pentamidine with the prime dosage of 3 mg/kg/day can potentially involve in the retardation of *Leishmania* growth with some severe antagonistic effects such as hypotension, elevated rate of hyperglycemia, tachycardia, pancreatic damage, and electrocardiographic changes (De Menezes et al., 2015).

The existing chemotherapies have a list of short comings comprising high cost, higher toxicity, and acquired resistance toward parasitic strain, and other side effects during their prevention mechanism insisted scientists and medical practitioners to evolve a new therapeutic system to treat NTDs. During the last decades, green therapies involving plant extracts, bioactive compounds, and secondary metabolites derived from particular plant species and different kinds of NPs synthesized using plant extract become promising as well as safer prevention therapies.

Natural Methods

From ancient times, plant-based traditional methods are being used in the therapeutics against various infectious ailments.

Currently, plant extract and particular bioactive compound extracted from plants are either directly used as a therapeutic source or as derived herbal drugs for the treatment of leishmaniasis as well as other microbial infection (Oryan, 2015).

Involvement of Plant Extracts and Plant-Derived Secondary Metabolites

The consumption of herbal drugs derived from plants is being used from centuries as a prevention source for NTDs as well as other diseases including bacterial (and their vectors), helminth (and their vectors), fungal, ectoparasitic, protozoan (and their vectors), and viral infections (and their vectors). The use of medicinal plants becomes more advantageous over other chemotherapies due to their nontoxic, environment-friendly, and cost-effective properties. Further natural compounds obtained from plants are considered as a reliable therapeutic source to treat leishmaniasis (Cheuka et al., 2017).

Ageratum conyzoides, Bidens pilosa, and Eugenia uniflora showed efficient leishmanicidal effects. Bidens pilosa (root) has been reported for its antileishmanial properties (against L. amazonensis, promastigote) with the least IC₅₀ value (1.5 µg/ ml) as compared to other plant species (Table 1). Essential oils from Eugenia uniflorae potentially inhibit the growth of both the parasitic forms, that is, promastigote and amastigote, of L. amazonensis, and Ageratum conyzoides has been reported to treat infection caused by L. donovani (amastigote form) (Silveira et al., 2021). E-caryophyllene, the main component of Melampodium divaricatum and Casearia sylvestris essential oil, has been reported for its promising antileishmanial response against L. amazonensis (IC50 values of 10.7, 10.7, and 14.0 µg/ ml) (Moreira et al., 2019). Moreover, 1,8-cineole, α-pinene, and p-cymene active constituents of Protium altsonii and P. hebetatum (Burseraceae) exhibited dose-dependent amastigote inhibition with IC₅₀ of 48.4, 37, and 46 µg/ml, respectively (Santana et al., 2020). Butanol fraction of K. odoratissima with 154.1 µg/ml IC₅₀ value showed antileishmanial properties against L. major promastigote and amastigote (Mirzaei et al., 2020).

Role of Plant-Based Nanoparticles in the Treatment of Leishmaniasis

Methods in controlling infectious diseases have modernized translational sciences to develop a better controlling method for infectious diseases. The field of nanomedicine has shown enormous potential in developing highly sensitive diagnostic tools with excellent drug delivery properties. Recently, nanoparticle-conjugated drugs have increasingly been studied as an alternative, cost-effective therapy with increased effectiveness. However, toxicity is a major barrier that needs to be encountered. Several reports have shown the effective antimicrobial activities of various metal/metal oxide nanoparticles as well as against the *Leishmania* causing organism through their wide surface area and unique properties.

Nanoparticles synthesized using crude as well as various solvent-fractionated extracts of medically important plants are considered as efficient agents for the delivery of specific phytoconstituents into the cells. Keeping in view the effective antimicrobial activities of silver metal, silver/silver oxide NPs

S. no	Plant used	Plant part used for extract preparation	Bioactive compound involved	Mode of study and optimum dosages	Organism tested	Structural formula	Mechanism of action	References
1	Baccharis uncinella (groundsel)	Leaves	Ursolic acid	<i>In vivo</i> 1.0 mg/kg or 2.0 mg/kg (body weight)	L. infantum		Treatment with ursolic acid causes a remarkable reduction in liver as well as splenic parasitism	Jesus et al. (2017)
2	Allium sativum (garlic)	Bulb	Allicin	In vitro and in vivo 50 μM for in vitro studies	L. major	°∥ s∽s∽s∽	-	Metwally et al. (201)
3	Eremurus persicus (desert candles)	Root extract	Aloesaponol III 8-methyl ether	<i>In vitr</i> o IC ₅₀ 73 μg/ml	L. infantum	OH OH	After treatment with isolated compound, mitochondrial potential and few structural alterations in the promastigote form of tested organism were observed	Rossi et al. (2017)
1	<i>Olea europaea</i> (wild olive, Indian olive, and brown olive)	Air-dried, pulverized leaves	Oleuropein	<i>In vitro</i> and <i>in vivo</i> 128.4 μΜ (69.4 μg/ ml), for <i>in vitro</i> studies	L. donovani	$= \left(\begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\$	Oleuropein responsible to raise ROS production, upregulation of host antioxidant enzymes, and downregulation of other enzymes of the parasite. Furthermore, <i>in vivo</i> model delayed-type	Kyriazis et al. (2016 Sharma et al. (2019
5	Zingiber zerumbet (awapuhi and bitter ginger)	Fresh rhizome	Zerumbone	<i>In vitro</i> 10 μM	L. donovani		hypersensitivity and elevation of IgG2a/IgG1 ratio (leishmania-specific) were observed Zerumbone extracted from Zingiber zerumbet causes apoptosis in promastigotes by affecting ROS production coupled with reduction of intracellular amastigotes in infected macrophages	Mukherjee et al. (201

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S. no	Plant used	Plant part used for extract preparation	Bioactive compound involved	Mode of study and optimum dosages	Organism tested	Structural formula	Mechanism of action	References
6	<i>Morinda lucida</i> (brimstone tree)	-	Molucidin	In vitro IC ₅₀ 4.24 μM for <i>Leishmania hertigi</i> and anti-010 activity with MIC of 4.167 μM	<i>L. hertigi</i> and field strain-010	CH2	Molucidin: Normal cells have single set of nucleus and kinetoplast, that is, 1N/1K, but molucidin stimulates two different sets of kinetoplast and nucleus in the cells of parasite. After division of both the sets, this compound obstructs the cytokinesis and causes cell cycle arrest which leads to death of parasites	Amoa-Bosompem et al. (2016);Sharma et al. (2019)
7	Artemisia annua (sweet annie, annual mugwort, sweet sagewort, or annual wormwood)	-	Artemisinin	In vivo and in vitro 100 µg/ml for in vivo studies	L. major	H ₃ C	-	Ghaffarifar et al. (2015); Sharma et al. (2019)
8	Hypericum Carinatum (St John's wort)	Flowering aerial parts	Cariphenone A (1), isouliginosin B (2), and uliginosin B (3)	In vitro IC_{50} values of 10.5, 17.5, and 11.3 μ M for compound 1, 2, and 3, respectively.	L. amazonensis		Inhibition of parasites mediated by oxidative stress (ROS production) and alteration in mitochondrial potential-like hyperpolarization condition	Dagnino et al. (2018)
9	<i>Euphorbia peplus</i> (radium weed)	Peplus aerial parts	Simiarenol	<i>In vitro</i> IC ₅₀ values of 20.24, 34.87, and 32.05 μg/ml	L. donovani	HO N H	- (Continu	Moawad et al. (2016) ed on following page)

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Protium altsonii (PaEO) and P. hebetatum (PhEO) Artemisia aucheri Clerodendrum myricoides (blue-flowered tinder wood) and Salvadora persica (arak, jhak, pīlu, Salvadora indica, toothbrush tree, mustard tree) Croton blanchetianus Baill	- Whole plant extract Aqueous extract of stem	Essential oil	In vitro PaEO IC ₅₀ were 14.8 µg/ml and 7.8 µg/ml and PhEO IC ₅₀ were 0.46 µg/ml and 30.5 µg/ml In Vitro and in vivo IC ₅₀ 90 µg/ml In vitro MIC = 625 µg/ml	L. amazonensis L. major L. major	-	Mitochondrial membrane potential associated with NO production could be an effective mechanism of leishmaniasis -	Santana et al. (2020) KarimiPourSaryazdi et al. (2020) Maina et al.(2020)
Clerodendrum myricoides (blue-flowered tinder wood) and Salvadora persica (arak, jhak, pīlu, Salvadora indica, toothbrush tree, mustard tree)	Aqueous extract of stem	-	90 µg/ml In vitro MIC =		-	-	et al. (2020)
(blue-flowered tinder wood) and <i>Salvadora</i> <i>persica</i> (arak, jhak, pilu, <i>Salvadora indica</i> , toothbrush tree, mustard tree)	stem	-		L. major	-	-	Maina et al.(2020)
,	Ethanolic extract	-					
			In vitro IC ₅₀ values of 208.6 and 8.8 µg/ml for <i>Leishmania</i> <i>infantum</i> and IC50 values of 73.6 and 3.1 µg/ml for <i>Leishmania</i> <i>amazonensis</i> promastigotes and	L. amazonensis and L. infantum	-	Ethanolic extract of <i>Croton</i> <i>blanchetianus</i> targets a significant depolarization of mitochondrial membrane potential and leads to mitochondrial dysfunction	Pereira et al. (2020)
<i>Prunus armeniaca</i> (Armenian plum)	Leaf extract	1, 2-benzenedicarboxylic acid and diisooctyl ester	In vitro anti- promastigote activity with $ C_{50} 11.48 \pm$ 0.82 µg/ml and anti- amastigotes activity with IC50 21.03 ± 0.98 µg/ml	L. tropica		-	Shaheen et al. (2020)
Urtica dioica (common nettle, stinging nettle)	Aqueous extract	-	In vivo and in vitro 3,500 and 6,000 µg/ ml for promastigotes and amastigotes, respectively.	L. major	-	It proficiently killed the amastigote form of <i>L. major</i> ; additionally, remarkable reduction of parasite load, skin lesion size, and IL-4, and significant increase of NO and IFN- γ were observed	Badirzadeh et al. (2020)
<i>Piper marginatum</i> (cake bush, anesi wiwiri,	Leaves (ethanolic extract)	3,4- Methylenedioxypropiophenone	In vivo	L. amazonensis		-	Macêdo et al. (2020)
L n Fb	Armenian plum) <i>Irtica dioica</i> (common ettle, stinging nettle) Piper marginatum (cake	Armenian plum) I <i>rtica dioica</i> (common Aqueous extract ettle, stinging nettle)	Armenian plum) diisooctyl ester <i>Irtica dioica</i> (common Aqueous extract - ettle, stinging nettle) <i>Piper marginatum</i> (cake Leaves (ethanolic 3,4- ush, anesi wiwiri, extract) Methylenedioxypropiophenone	values of 73.6 and 3.1 µg/ml for Leishmania amazonensis promastigotes and amastigotes activity with ICso 11.48 ± 0.82 µg/ml and anti- amastigotes activity with ICS0 21.03 ± 0.98 µg/mlIn vito and in vitro 3,500 and 6,000 µg/ ml for promastigotes, respectively.Viper marginatum (cake ush, anesi wiwiri,Leaves (ethanolic extract)3,4- MethylenedioxypropiophenoneIn vivo	In vivo and in vitro L. major Armenian plum) Aqueous extract - In vivo and in vitro 3,500 and 5,000 µg/ml In vivo and in vitro 1,2-benzenedicarboxylic acid and diisooctyl ester In vivo and in vitro 0.82 µg/ml and anti-amastigotes activity with IC50 21.03 ± O.98 µg/ml - In vivo and in vitro 3,500 and 6,000 µg/ml for promastigotes, respectively. Viper marginatum (cake Leaves (ethanolic ush, anesi wiwiri, extract) 3,4- In vivo L. major Artact 3,4- In vivo L. armazonensis	$ \frac{1}{2} 1$	values of 73.6 and 3.1 µg/ml for Lishmania amazonensis promastigotes and amastigotes and disocctyl estervalues of 73.6 and 3.1 µg/ml for Lishmania amazonensis promastigotes and amastigotes and to X2 µg/ml and anti- amastigotes activity with ICGs 21.0 3 ± $0.38 µg/ml$ L. tropica tropicaIn vivo and in vitro atter, stinging nettle)In vivo and in vitro 3,500 and 6,000 µg/ ml for promastigotes and amastigotes, and amastigotes, respectively.L. major It proficiently killed the amastigotes and amastigotes and amastigotes, and amastigotes, and amastigotes, respectivelyIt proficiently killed the amastigotes and amastigotes, and amastigotes, and amastigotes, respectivelyIt major additionally, remarkable observedtiper marginatum (cake Leaves (ethanolic ush, anesi wiwiri, extract)3,4- MethylenedioxypropiophenoneIn vivo additionally amazonensisL. major additionally, remarkable amazonensis-tiper marginatum (cake ush, anesi wiwiri, extract)3,4- MethylenedioxypropiophenoneIn vivo amazonensisL. μ_{max} amazonensis-tiper marginatum (cake ush, anesi wiwiri, extract3,4- MethylenedioxypropiophenoneIn vivo amazon

S. no	Plant used	Plant part used for extract preparation	Bioactive compound involved	Mode of study and optimum dosages	Organism tested	Structural formula	Mechanism of action	References
21	Kelussia odoratissima (kelus celery and wild celery)	Dried leaves (butanol fraction)	-	In vitro half (IC_{50}) 264.1 and 154.1 µg/ml for promastigotes and amastigotes, respectively.	L. major		-	Mirzaei et al. (2020)
22	Tabernaemontana coronaria (milkwood)	Dried powder of stem bark	Voacamine	In vivo IC ₅₀ value was found to be 14.702 \pm 0.101 mM	L. donovani		Voacamine supresses the relaxation potential of LdTop1B (<i>L. donovani</i> toposoisomerase IB) and makes the clevable complex steady	Chowdhury et al. (2017)
23	Picramnia gracilis (bitterbush)	Powder of dried leaves	5,3'-hydroxy-7,4'- dimethoxyflavanone	In vitro and in vivo EC ₅₀ 17.0 + 2.8 mg/ ml, 53.7 µM for <i>in vitro</i> and 2 mg/kg/day for <i>in</i> vivo studies	L. braziliensis	H,CO CH, OCH, OCH, OCH, OCH, OCH, OCH, O		Robledo et al. (2015
24	<i>Lindera aggregate</i> (spice bush)	Leaves/bark	Boldine	<i>In vitr</i> o 600 μg/ml	L. amazonensis		-	Salama et al. (2017)
25	Hypericum andinum	Dried and powered materials of aerial parts	Uliginosin B	<i>In vitr</i> o (IC ₅₀) of 36.1 g/ml	L. amazonensis		-	Dagnino et al. (2015
26	Amphilophium crucigerum (monkey's comb)	Aerial parts	Ipolamiide	<i>In vitro</i> IC ₅₀ = 100 μM	L. amazonensis	HO HO HO HO HO HO HO HO HO HO HO HO HO H	-	Vendruscolo et al. (2018)

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TABLE 1 | (Continued) Role of plant-based natural products in the treatment of leishmaniasis.

S. no	Plant used	Plant part used for extract preparation	Bioactive compound involved	Mode of study and optimum dosages	Organism tested	Structural formula	Mechanism of action	References
27	<i>Valeriana jatamansi</i> (Indian Valerian or Tagar)	Rhizomes	Valepotriates	In vitro IC ₅₀ = 2.96 μ M	L. major	-	-	Glaser et al. (2015)
28	<i>Nymphoides indica</i> (banana plant, robust marshwort, and water snowflake)	Leaves	3-O-methylquercetin-7-O- β-glucoside	<i>In vitro</i> IC ₅₀ 32 μM	L. infantum		-	Amin et al. (2016)
29	<i>Vitex grandifolia</i> (black plum, chocolate and berry tree)	Air-dried leaves	Bartioside	<i>In vitro</i> IC ₅₀ 27.51 μΜ	L. donovani	но о он но он он	-	Bello et al. (2018)
30	<i>Scrophularia syriaca</i> (figworts)	Aerial parts	6-0-a-Irhamnopyranosylcatalpol	<i>In vitro</i> EC ₅₀ 100 µM	L. major	-	-	Alkhaldi et al. (2020)

Section 2: Natural drugs for treating leishmaniasis

S. no	Compound (s)	Company/ originator	Country	Year	Mode of studies	Probable mechanism involved	Patent and IPC	Ref.
31	Ethyl 3-(2- chloroacetamido) benzoate, dihydroquercetin, and bisabolol	Auclair et al., AC Bioscience SA	Switzerland	2019	<i>In vitro</i> and <i>in vivo</i> studies	Inhibition of some important parasitic enzymes with tryparedoxin peroxidase and tubulin	WO2019043212 and A61K A61P	Hajaji et al. (2018)
32	Diterpenoid membranolides	Baker et al., the University of South Florida	United States of America	2016	In vitro	Impedes lipid synthesis	US2016003O388 and A61K	Baker et al. (2016)
33	Withaferin-A and miltefosine	Maurya et al., the University of Hyderabad	India	2017	<i>In vitro</i> and <i>in vivo</i> studies	Inhibits pteridine reductase-1 enzyme, and phosphatidylcholine synthesis, and cytochrome c oxidase	WO2017046778and A61K A61P	Maurya and Chandrasekaran (2017)

synthesized using a variety of medicinally important plant species, including Mentha arvensis L., Ficus benghalensis, Cuminum cyminum, Moringa oleifera, Silvbum marianum, and Sechium edule, at a dosage of 10, 300, 0.5, 246, and 51.88 µg/ml tested against L. tropica, L. donovani, L. tropica, L. major, L. tropica, and Leishmania donovani, respectively (Baranwal et al., 2018; El-Khadragy et al., 2018; Hameed et al., 2019; Ismail et al., 2019; Bagirova et al., 2020; Javed et al., 2020). Gold and silver bimetallic NPs synthesized by using medically important plants have also been reported to be used for the prevention of this disease (Alti et al., 2020). However, Cannabis sativa-based Au-NPs accomplished virtuous antileishmanial activity against amastigote forms (IC₅₀: 171.00 ± 2.28 μg/ml) (Hameed et al., 2020). 7, 8dihydroxyflavone, a type of flavonoid found abundantly in plants used to produce gold nanoparticles, has also been reported to preclude leishmaniasis (Prasanna et al., 2020).

Zno-NPs were also reported to show dose-dependent cytotoxicity against L. tropica (IC50: 8.30 µg/ml) (Iqbal et al., 2019). Rod-shaped zinc oxide NPs produced by using Lilium ledebourii tuber extract potentially inhibited the growth of L. *major* with the IC₅₀ value of 0.001 mg ml⁻¹ (Khatami et al., 2020). Saleh (2019) also concluded that green TiO₂ nanoparticles have shown effective roles to counter noxiousness of Leishmania tropica in male rats. Hematite (Fe₂O₃) NPs fabricated with the Rhus punjabensis extract played an efficient role in the treatment of leishmaniasis (Naz et al., 2019). Khalil et al. (2020) prepared lead oxide NPs (PbO-NPs) by green route using aqueous leaf extracts of Sageretia thea. The experimental data showed that PbO-NPs were significantly active in arresting the growth of promastigote and amastigote forms of Leishmania tropica, with 14.7 µg/ml and 11.95 µg/ml IC₅₀ values, respectively.

Plant-mediated (*Trigonella foenum-graecum*) iron oxide nanoparticles have been reported to exhibit significant inhibitory effects on *L. tropica* (Ain et al., 2019). Further, Abbasi et al. (2019) also stated the antileishmanial efficacy of Nio-NPs fabricated by using *Geranium wallichianum* against *L. tropica*.

Besides, the nanostructured drug delivery system was also reported in ameliorate NTDs including leishmaniasis. Furthermore, crude plant extracts and precise phytoconstituents obtained from plant which is involved in the prevention mechanism were also loaded in the nanostructured drug delivery system and used as a therapeutic source to cure leishmaniasis, and the mechanism is depicted below:

• Liposome NPs consisting of phospholipids are assisted as a transport system for the delivery of hydrophilic as well as lipophilic pharmaceutical drugs (Momeni et al., 2013). They provide improved pharmacokinetic assets along with target diligence which offers a foremost advantage (Kaye and Scott, 2011). Liposome can spear the macrophages through phagocytosis and offers direct delivery of the drugs at their targeted sites. Different drug formulations including AmB colloidal formulations, liposomal AmB, and AmB lipid network can overwhelm the toxic effects of conventional drugs (Moreno et al., 2015). Liposome-encapsulated *Curcuma longa* and *Combretum leprosum*

extracts were also reported for their antileishmanial properties (Aditya et al., 2012; Barros et al., 2013).

- Beta-lapachone extracted from Lapacho tree with the use of lecithin-chitosan NP encapsulation method has been reported in the treatment of leishmaniasis (Moreno et al., 2015).
- 8-hydroxyquinoline with the polymeric micelle encapsulation method has been used to treat *Leishmania* (Duarte et al., 2016). Berberin, an isoquinoline alkaloid extracted from medicinal plants, has been reported to possess various biologic properties, including antileishmanial properties. A previous study addressed the preparation of BER-loaded liposomes with the aim to prevent its rapid liver metabolism and improve the drug selective delivery to the infected organs in visceral leishmaniasis (VL) (Calvo et al., 2020).

As per the literature survey, plant-based nanoparticles contribute efficient roles in the treatment of leishmaniasis as compared to other existing practices. Phytosynthesized NPs revealed an identical effect on the inhibition of parasitic growth at a comparatively lesser concentration than the prescribed dose of Amp B to cure this disease. Additionally, bimetallic nanoparticles including Au-Ag, Zn-Ag, and Ti-Ag were synthesized using the green approach and proficiently used as a therapeutic source to treat leishmaniasis (Alti et al., 2020). NPs are preferred over other therapeutic sources to treat this dreaded disease because of their nontoxic, harmless, and efficient delivery system for vaccine. Currently, with the advancement of nanosciences, there is a new method of synthesizing vaccines using NPs as carriers of antigen preparation. Solid lipid nanoparticles can assist as an effective tool to produce leishmanial vaccine (Saljoughian et al., 2013). However, any kind of NP-based vaccine is not accessible, and it needs more consideration.

RESTORATIVE MECHANISM OF NANOFORMULATIONS AGAINST LEISHMANIASIS

Leishmania sp. are protozoal parasites which result in cutaneous and visceral leishmaniasis. Different clinical studies exhibit the development of self-curable to detrimental conditions, depending upon the immune responses triggered by the affected host (Noormehr et al., 2018). Chemotherapy with pentavalent antimonials (like sodium stibogluconate or meglumine antimoniate) and other antileishmanial drugs (amphotericin B, fluconazole, pentamidine, and miltefosine) are optimal for leishmanial therapy. However, due to adverse effects, high cost, difficult infusion routes, low cure, and increasing resistance are of significant concern in developing more efficient ways in leishmaniasis therapy. Moreover, the efficacy of the drug used in the treatment also varies for different leishmanial sp. (Noormehr et al., 2018; Alti et al., 2020; Calvo et al., 2020). In self-treatment, the innate immune cells (phagocytes) detect and engulf the causal agents, which induces Leishmania assassination by producing reactive oxygen species, nitric oxide, and tumor necrosis factors

(Olekhnovitch et al., 2014). After innate immune responses, respective activation and production of CD8⁺, NK, and IFN cells by TH1 immunity results in killing of *Leishmania* parasites (Noormehr et al., 2018). In susceptible conditions, the defense system fails to overcome infections, and follows incorrect TH2 immune responses along with antibody response, which is the key factor to generate new ways of parasite elimination. Metal nanoparticles inhibit proliferation and viability of infected cells, which is contingent with the NP strength and time of exposure (Rosas-Hernández et al., 2009; Fanti et al., 2018).

Several in vitro as well as in vivo findings suggest leishmanicidal effects of bio-Ag-NPs by direct (exclusive of inflammatory mediators) or indirect (immunomodulatory) mechanisms (Fanti et al., 2018; Calvo et al., 2020). In the direct method, metal-NPs kill the parasitic cells by causing vacuolation inside parasites and to the cellular membrane without generating damage immunomodulatory intermediaries, that is, reactive oxygen species (ROS), nitric oxide (NO), and apoptotic and necrotic factors (Fanti et al., 2018). In situations when Leishmania parasites override the oxidative burst inside phagocytic cells and reside in phagolysosomes, nanoformulations assist site-specific delivery and accumulation of drugs, which is responsible for parasite killing (Shoaib Sarwar et al., 2020). According to Fanti et al. (2018), Ag-NPs after diffusion through the cellular membranes get oxidized due to acidic conditions within the phagolysosomes, and eventually, the release of free Ag + ions causes parasite assassination.

On the other hand, the indirect method involves immunomodulatory response generation at infection sites. Other ways to provide leishmanicidal effects are through activating immune response mediators in which the cell viability and proliferation get declined as an effect of metallic nanoparticles. NPs basically induce different morphological abrasions such as distorted membranal integrity, cytotoxicity, mitochondrial destruction, cell cycle arrest (G1), increased/decreased ROS and NO generation, affected enzymatic activities, and release of apoptotic or necrotic factors (Park et al., 2010; Kruszewski et al., 2011; Zahir et al., 2015). As a result of mitochondrial disintegration, ATP generation gets influenced, which causes cytotoxic effects, and ultimately affects the infection growth (AshaRani et al., 2009). Moreover, NP exposure exhibits decreased parasitic load and reduction in an essential parasitic enzyme trypanothione reductase system (Fanti et al., 2018).

CONCLUSION AND FUTURE PROSPECTIVE

Chemotherapy due to lack of effective therapies till date has become the only choice in treating leishmaniasis, as these therapies exhibit higher toxicity levels, treatment cost, and resistance development against leishmanial parasites, and encourage other side effects. In addition, it is evident that the efficiency of drugs varies from species to species due to leishmanial antigen variants and different immunological responses against the drug. To overcome these challenges, biogenic nanomaterials being nontoxic, biocompatible, cost effective, and having high targeted drug-loading potentials have been indicated as beneficial alternatives to formulate nanovaccines. Targeted drug delivery barriers can be conquered by using nanoformulations for enhanced parasiticidal proficiencies. Also, various studies have demonstrated leishmanicidal activities of plant-derived natural compounds (such as berberine, 7, 8dihydroxyflavone, E-caryophyllene, essential oil constituents, a-terpineol, glycosides, tannins, and anthraquinone flavonoids), which can further integrate beneficial outcomes. Besides, most of the studies conducted on leishmanicidal activities revealed only the basic outcomes like assessment of the effect of test drugs (crude extract, isolated bioactive compounds, essential oil, and purified fraction) on the parasite growth. Few of them identify the proper formulation as well as the effect on the promastigote stage, found in the sandflies (vector). As widely conferred in the literature, plants possess a variety of bioactive compounds, and most of them have been reported for their pharmaceutical properties. Thus, the standardization may conclude the identification of particular compound responsible for leishmanicidal activities. Biosynthesized nanoparticles majorly eliminate the infection either by triggering the immunomodulatory response of the host or sometimes directly by resulting in vacuolization of parasitic cells, leading to parasite killing. Nanovaccines are a relatively new concept in treating Leishmania although no vaccine is yet available, but studies are ongoing to find efficient nanovaccines. Although nanotechnology has provided a hope toward improved and successful eradication of neglected tropical diseases, the accurate molecular mechanism responsible still needs thorough transparency to bring utmost benefits.

AUTHOR CONTRIBUTIONS

LS: writing original draft and editing; MD: writing, review, and editing; AS: conceptualization and validation; MS: supervision, conceptualization, and validation.

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The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmolb.2021.655584/ full#supplementary-material.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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