### Check for updates

### **OPEN ACCESS**

EDITED BY Masaki Saito, Tohoku University, Japan

REVIEWED BY Issei Shimada, Nagoya City University, Japan

\*CORRESPONDENCE Tatsuo Miyamoto, ⊠ t-miyamoto@yamaguchi-u.ac.jp

#### SPECIALTY SECTION

This article was submitted to Lipids, Membranes and Membranous Organelles, a section of the journal Frontiers in Molecular Biosciences

RECEIVED 07 February 2023 ACCEPTED 06 March 2023 PUBLISHED 16 March 2023

#### CITATION

Kimura S, Morita T, Hosoba K, Itoh H, Yamamoto T and Miyamoto T (2023), Cholesterol in the ciliary membrane as a therapeutic target against cancer. *Front. Mol. Biosci.* 10:1160415. doi: 10.3389/fmolb.2023.1160415

### COPYRIGHT

© 2023 Kimura, Morita, Hosoba, Itoh, Yamamoto and Miyamoto. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Cholesterol in the ciliary membrane as a therapeutic target against cancer

Sotai Kimura<sup>1</sup>, Tomoka Morita<sup>2</sup>, Kosuke Hosoba<sup>3,4</sup>, Hiroshi Itoh<sup>1</sup>, Takashi Yamamoto<sup>3,4</sup> and Tatsuo Miyamoto<sup>2</sup>\*

<sup>1</sup>Department of Molecular Pathology, Graduate School of Medicine, Yamaguchi University, Ube, Japan, <sup>2</sup>Department of Molecular and Cellular Physiology, Graduate School of Medicine, Yamaguchi University, Ube, Japan, <sup>3</sup>Program of Biomedical Science, Graduate School of Integrated Sciences for Life, Hiroshima University, Hiroshima, Japan, <sup>4</sup>Program of Mathematical and Life Science, Graduate School of Integrated Sciences for Life, Hiroshima University, Hiroshima, Japan

Primary cilium is a non-motile, antenna-like structure that develops in the quiescent G<sub>0</sub> phase-cell surface. It is composed of an array of axonemal microtubules polymerized from the centrosome/basal body. The plasma membrane surrounding the primary cilium, which is called the ciliary membrane, contains a variety of receptors and ion channels, through which the cell receives extracellular chemical and physical stimuli to initiate signal transduction. In general, primary cilia disappear when cells receive the proliferative signals to re-enter the cell cycle. Primary cilia thus cannot be identified in many malignant and proliferative tumors. In contrast, some cancers, including basal cell carcinoma, medulloblastoma, gastrointestinal stromal tumor, and other malignancies, retain their primary cilia. Importantly, it has been reported that the primary cilia-mediated oncogenic signals of Hedgehog, Wnt, and Aurora kinase A are involved in the tumorigenesis and tumor progression of basal cell carcinoma and some types of medulloblastoma. It has also been demonstrated that cholesterol is significantly more enriched in the ciliary membrane than in the rest of the plasma membrane to ensure Sonic hedgehog signaling. A series of epidemiological studies on statin drugs (cholesterol-lowering medication) demonstrated that they prevent recurrence in a wide range of cancers. Taken together, ciliary cholesterol could be a potential therapeutic target in primary ciliadependent progressive cancers.

### KEYWORDS

primary cilia, cholesterol, cancers, sonic hedgehog signal, Wnt signal

### **1** Introduction

Primary cilium is a single, non-motile, antenna-like structure on the quiescent  $G_0$  phasecell surface. The plasma membrane surrounding the primary cilium is composed of axonemal microtubules that originate from the centrosome-converted basal body, and is known as the ciliary membrane (Singla and Reiter, 2006). The ciliary membrane contains a variety of receptors and ion channels, thereby receiving the extracellular mechanical and chemical stimuli to transduce the signals involved in cell proliferation and differentiation (Fabbri et al., 2019).

In general, primary cilia are assembled in the quiescent  $G_0$  phase and disassembled in the cell cycle after receiving proliferative stimuli (Fabbri et al., 2019). It has been



### FIGURE 1

Schematic longitudinal section and cholesterol imaging of a primary cilium. (Left) The mother central body (MC) and daughter central bodies (DC) of the basal body (centrosome) are located at the base of the primary cilium, and microtubular axonemes develop from the mother central body docked to the cell membrane. The ciliary membrane covering the axonemes is rich in cholesterol and sphingolipids. At the base of the primary cilia, the plasma membrane is concave, forming a serial pocket. At the transition zone (TZ), the cytoplasm and ciliary membrane of primary cilia are biochemically compartmentalized from other cytoplasmic and cell membranes. (Right) Imaging of ciliary cholesterol in quiescent G<sub>0</sub>-phase hTERT-RPE1 cells stained with Filipin III. They were also immunostained with anti-acetylated-tubulin and anti-pericentrin antibodies. The merged image indicates the enriched ciliary cholesterol. Scale bar, 5 µm.

reported that primary cilia are not formed in many types of malignant tumors to dampen cilia-related signal transduction (Seeley et al., 2009; Hassounah et al., 2013; Nobutani et al., 2014; Liu et al., 2019). However, certain types of medulloblastoma, gastrointestinal stromal tumor, and basal cell carcinoma retain the primary cilia, which transduce signals such as Hedgehog and Wnt to contribute to the malignancy (Hassounah et al., 2012; Castiella et al., 2013; Park et al., 2021; Marino, 2022; Youn et al., 2022). Thus, whether primary cilia promote or suppress the progression of carcinoma is an important but difficult question in the field of oncology.

For primary cilia to receive extracellular signals effectively, not only the protein, but also the lipid composition of the ciliary membrane is compartmentalized from the rest of the plasma membrane (Nishimura et al., 2021; Miyamoto et al., 2022). Cholesterol, which is a major lipid for controlling cell membrane fluidity and is used as a precursor of steroid hormones and bile acids, is enriched in the ciliary membrane to transmit cilia-related oncogenic sonic hedgehog (SHH) signal (Miyamoto et al., 2022; Figure 1). Notably, epidemiological and experimental studies have suggested that cholesterol-lowering drug statins, which strongly inhibit the de novo cholesterol synthesis-limiting enzyme, hydroxymethylglutaryl-CoA (HMG-CoA) reductase, significantly prevent the recurrence in several cancers (Beckwitt et al., 2018; Riscal et al., 2019; Yang et al., 2020). These findings suggest that downregulation of ciliary cholesterol acts as tumor suppression. In this minireview, we summarize the function of primary cilia in the context of cancer formation and progression, and the potential of ciliary cholesterol as a therapeutic target.

# 2 Primary cilia-related signal transduction in the context of tumorigenesis

Primary cilia mediate cancer-related signaling such as the Sonic hedgehog, Wingless/INT (Wnt), NOTCH, platelet-derived growth factor (PDGF), mammalian target of rapamycin (mTOR), and Hippo pathways (Basten and Giles, 2013; Liu et al., 2018; Fabbri et al., 2019). Here, we focus on two major cilia-related cancer signal pathways.

### 2.1 SHH signaling pathway

SHH signaling is involved in the dorsal-ventral axis and in midline formation, digit patterning, and cerebellum development in embryogenesis (Harfe et al., 2004; Ribes and Briscoe, 2009; Wang et al., 2022). When the SHH ligand binds to the PTCH1 receptor, which is the 12-transmembrane patched 1 protein localized on the ciliary membrane in the absence of SHH ligand, it translocates to the outside of the cilia. Following the ciliary exit of PTCH1, the seventransmembrane protein smoothened (SMO) enters the ciliary membrane to induce the release of GLI transcriptional protein known as glioma-associated oncogene from the Suppressor of fused (SUFU) protein in the ciliary cytoplasm, thereby shuttling GLI from the primary cilium compartment to the nucleus to control the transcription of GLI-target genes (Figure 2; Basten and Giles, 2013; Byrne et al., 2016; Qi et al., 2018; Liu et al., 2018; Fabbri et al., 2019; Niewiadomski et al., 2019). Germline mutations of the PTCH1 gene causes autosomal-dominant Gorlin syndrome with basal cell



FIGURE 2

Ciliary cholesterol-mediated signal transduction. (Left) In the presence of Shh ligands, Ptch exits from ciliary membrane, while Smo enters and is then activated. Ciliary Smo allows Sufu to release Gli. Activated Gli is translocated into the nucleus, thereby controlling the transcription of Shh responsive genes. Ciliary cholesterol directly binds to Smo to ensure the activation. (Right) In the presence of Wnt1 class ligands, Frizzled-LRP5/6 activates DVL that destabilizes the destruction complex composed of APC and GSK3, thereby allowing  $\beta$ -catenin cytosolic accumulation and nuclear translocation to regulate the Wnt responsive gene expression.

carcinoma and medulloblastoma predisposition. In Gorlin syndrome with dominant negative mutations of *PTCH1*, constitutive activation of SMO triggers the misregulation of SHH-mediated gene expression to contribute to the cancer disposition (Athar et al., 2014).

### 2.2 Wnt signaling pathway

Wnt signaling controls many biological events, including embryogenesis, stem cell maintenance, and tumorigenesis (Wang et al., 2012). In the canonical Wnt signaling pathway, the Wnt1 class ligands (Wnt2, Wnt3, Wnt3a, and Wnt8) bind the membrane protein complex of Frizzled (FZ) family proteins and lipoprotein receptor-related protein (LRP) 5/6 to form the FZ, LRP5/6, and cytoplasmic Dishevelled (DVL) protein complex and trigger the downstream cascade. Importantly, FZ and LRP5/6 localize to the ciliary membrane, implying that primary cilia might sense the Wnt signals. In the absence of canonical Wnt ligands, β-catenin is phosphorylated and ubiquitinated by the destruction complex (DC), which consists of adenomatous polyposis coli (APC), the scaffolding protein Axin, glycogen synthase kinase (GSK)-3β, and other regulators to be degraded (Haseeb et al., 2019; He et al., 2015; Phesse et al., 2016; Chae and Bothwell, 2018; Aros et al., 2021). The canonical Wnt-activated DVL inhibits the DC to stabilize β-catenin, thereby allowing  $\beta$ -catenin nuclear translocation to induce target gene expression for cell proliferation and cell differentiation

# (Figure 2; He et al., 2015; Phesse et al., 2016; Chae and Bothwell, 2018; Aros et al., 2021).

In the non-classical Wnt signaling pathway, the Wnt5a type ligands (Wnt4, Wnt5a, Wnt5b, Wnt6, Wnt7a, and Wnt11) trigger Wnt/planar cell polarity (PCP) or the Wnt/Ca<sup>2+</sup> pathways for cell polarization and cell migration. Both pathways are initiated by binding of the Wnt5a-type ligands to FZ/retinoic acid-related orphan receptor (ROR)/tyrosine-protein kinase RYK. The Wnt/ Ca<sup>2+</sup> pathway is mediated by phospholipase C (PLC) and protein kinase C (PKC) to increase the intracellular Ca2+ concentration, thereby activating the calcium/calmodulin-dependent protein kinase II (CAMKII) and nuclear factor of activated T-cells (NFAT) pathways to regulate target gene transcription. In the Wnt/PCP pathway, the binding of a Wnt ligand to FZ, ROR, and RYK receptors involves DVL-mediated small GTPase RHOA and Rac1 for the control of actin dynamics and Jun N terminal kinase (JNK) activation in the context of cell polarization and cell migration (Haseeb et al., 2019; Chae and Bothwell, 2018). Notably, the interaction of Wnt5a and FZ stimulates casein kinase 1 (CK1) to phosphorylate DVL2 at the centrosome, thereby forming DVL2 and a mitotic kinase polo-like kinase 1 (PLK1) complex. The DVL2-PLK1 complex activates human enhancer of filamentation 1 (HEF1) and Aurora Kinase A (AurA) to disassemble primary cilia (Pugacheva et al., 2007). HEF1/AurA phosphorylates histone deacetylase (HDAC) 6 to deacetylate not only histone but also tubulin, thereby inhibiting axonemal tubulin polymerization. Interestingly, the HEF1/AurA-HDAC6-mediated primary cilia

disassembly axis is also activated by platelet-derived growth factor signaling. Together, primary cilia mediate the Wnt signals, while their formation is controlled by the Wnt signals (Pugacheva et al., 2007; Liu et al., 2018; Lee, 2020).

### 3 Ciliogenesis in the cancer tissues

Consistent with the basic relationship between cell cycle and primary cilia formation, it has been reported that primary cilia formation is disrupted in many cancer tissues and cell lines. Heterochronic activation of mitotic kinases at centrosomes during the G<sub>0</sub>/G<sub>1</sub> phases occurs in many cancers. AurA, Nemolike kinase 2 (NEK2), and PLK1 phosphorylate HDAC6, a mother centriole capping protein, KIF24, and a microtubuledepolymerizing enzyme, KIF2A, respectively, to activate the primary cilia resorption program in many cancers (Kinzel et al., 2010; Izawa et al., 2015; Kim et al., 2015; Miyamoto et al., 2015). In small intestinal and colorectal cancer, the frequency of primary cilia was very low (median, 0.49%) in the cells of small bowel adenocarcinoma and 0.22% (median) in the cells of colorectal adenocarcinoma. Overall survival of the patients with a higher frequency of primary cilia (≥0.187%) was significantly longer (Dvorak et al., 2016). However, several papers demonstrated that primary cilia exist even in cancer tissues. In 25% (25/100) of pancreatic adenocarcinoma cases, most primary cilia were found in the lumen of well-differentiated adenocarcinomas. Primary cilia formation was also reported in 100% (8/8) of adenocarcinoma of the small intestine cases, 100% (32/32) of adenocarcinoma of the colorectum cases, and 72% (69/96) of clear cell renal cell carcinoma of the kidney cases (Emoto et al., 2014; Dvorak et al., 2016; Rozsypalova et al., 2019). In thyroid cancer, primary cilia are found in papillary carcinoma and follicular carcinoma; however, primary cilia are negative in Hürthle cell carcinoma, oncocytic subtype of papillary carcinoma, and papillary carcinoma associated with Hashimoto thyroiditis, which are often associated with abnormal mitochondrial function (Lee et al., 2021). These findings suggest that primary cilia in heterogenous cancer cell populations contribute to the oncogenic characteristics.

Recently, epithelial-mesenchymal transition transcriptional factors (SNAIL, SLUG, and ZEB1) induce primary cilia formation in mammary tumor-initiating cells of claudin-low breast cancers. These cilia receive FGFR1 signals to decrease the amount of GLIS2 protein at the cilia base for maintaining cancer stemness (Guen et al., 2017; Wilson et al., 2021). Medulloblastoma, a well-known cilia-related cancer, is an embryonal malignant tumor that predominantly affects children and is now histopathologically classified into five major types: Wnt-activated; SHH-activated and TP53-mutated; SHH-activated and TP53 wild type; non-Wnt/non-SHH type; and histologically defined type, which contains classic desmoplastic/nodular medulloblastoma, medulloblastoma, medulloblastoma with extensive nodularity, and large cell/ anaplastic medulloblastoma (WHO Classification of Tumours Editorial Board, 2021). The first two have primary cilia and show activation of the Wnt and SHH pathways, respectively (Marino, 2022; Youn et al., 2022). In medulloblastoma-modeled mice driven by the constitutively active Smo transgene (SmoM2), depletion of Kif3a removes primary cilia to block medulloblastoma formation (Barakat et al., 2013). In other cilia-related tumors, skin basal cell carcinoma and rhabdomyosarcoma often have primary cilia to mediate the constitutive activation of SHH signaling as a major driver of pathogenesis (Fu et al., 2014; Kuonen et al., 2019). The SMO antagonists, vismodegib and sonidegib, have been approved by the U.S. FOOD & DRUG administration (FDA) for treating skin basal cell carcinoma (Meiss and Zeiser, 2014; Jain et al., 2017). Together, primary cilia in several cancers transmit the oncogenic signals involved in tumorigenesis and progression.

# 4 Ciliary cholesterol as a therapeutic target in cancers

Structural biology studies of SMO have demonstrated that the extracellular cysteine rich domain (CRD) and the transmembrane domain (TMD) of Smo directly binds to cholesterol to activate SHH signaling (Nedelcu et al., 2013; Huang et al., 2018; Deshpande et al., 2019). In the absence of SHH ligand, PTCH1 can bind and efflux cholesterol from cells, thereby restricting cholesterol supply to Smo (Bidet et al., 2011). When SHH ligand binds to PTCH1 to inhibit its cholesterol efflux activity, a specific fraction of cholesterol in the plasma membrane called accessible cholesterol increases to bind to the CRD and TMD of Smo, thereby activating the smo and its downstream molecules (Kinnebrew et al., 2019). Other fraction of cholesterol termed sequestered cholesterol consists of cholesterol bound to phospholipids such as sphingomyelin (SM). Interestingly, depletion of SM by a antibiotics myricion, which inhibits the enzyme of SM synthesis, increases the amount of accessible cholesterol to enhance the Smo activation (Kinnebrew et al., 2019). Depletion of cellular cholesterol with methyl-β-cyclodextrin inhibits SHH signaling, suggesting that lowering the level of cholesterol promotes anti-cancer effects. Indeed, it was reported that statins reduce cancer recurrence and mortality by prohibiting the outgrowth of dormant cancer cells (Beckwitt et al., 2018). In breast cancer, estrogen receptor (ER)-negative breast cancer is particularly correlated with statin sensitivity, and fluvastatin has been clinically used to enhance tumor cell apoptosis in ER-negative breast cancer (Garwood et al., 2010). Clinical trials of combined treatment of standard anti-cancer drugs and statins are also underway in various other cancer types including head and neck, pulmonary, gastric, colorectal, hepatic, pancreatic, renal, prostatic, endometrial, and ovarian cancers, malignant lymphoma and multiple myeloma; and glioblastoma (Longo et al., 2020; Jiang et al., 2021). Because of the side effects of statins, which include rhabdomyolysis and liver damage in some cases, more specific cell membrane cholesterol-lowering technology could provide safer cancer therapies.

Ciliary cholesterol is a potential target for inhibiting oncogenic signaling. Our previous report demonstrated that peroxisomes move along the microtubules to supply cholesterol to the ciliary membrane, and that peroxisome biogenesis disorder Zellweger syndrome patient cells show defects in ciliary cholesterol levels and the SHH signal response. In the context of peroxisomemediated ciliary cholesterol trafficking, small G protein Rab10, its GTP exchange factor Rabin8, and a kinesin molecule KIFC3 that moves to the minus end of microtubules, form a complex on the peroxisomal membrane (Miyamoto et al., 2020). Inhibitors of peroxisome dynamics might act more specifically to lower ciliary cholesterol. Mass spectrometry of the membranes of isolated cilia from sea urchin embryos revealed that several oxysterols synthesized by HSD11 $\beta$ 2 at the endoplasmic reticulum are enriched to stimulate SMO (Raleigh et al., 2018). Importantly, genetic ablation of *HSD11B2* reduced the size of medulloblastoma in the SmoM transgenic mouse model, suggesting that HSD11 $\beta$ 2 inhibitors might be applied as cilia-related cancer therapies (Daggubati et al., 2021).

Whether ciliary cholesterol regulates other oncogenic signalings is an important open question. WNT signal is a strong candidate of ciliary cholesterol-mediated oncogenic signal. Recently, it was reported that the WNT co-receptor LRP6 localizes to ciliary membrane to meditate the signal (Zhang et al., 2023). Cholesterol in the plasma membrane is enriched around the LRP6 and its binding partner Fz. WNT receptors and Cholesterol complex ensures the membrane recruitment and activation of DVL for the WNT/ $\beta$ -catenin signal transduction (Sheng et al., 2014). These findings suggest that ciliary membrane mediates the WNT/ $\beta$ -catenin signal for the proliferation of cancer cells and maintenance of cancer stem cells. Taken together, further investigation of ciliary cholesterol levels will enable exploration of new drugs against cancer.

# **5** Conclusion

In general, primary cilia have been considered to be absent in malignant and proliferative tumors. However, morphological observation of a wide variety of cancer tissues has revealed that there are carcinomas with primary cilia, which contribute to tumor formation and progression and the maintenance of cancer stemness through cilia-related signaling. Therefore, it is suggested that ciliary signal inhibition may be a therapeutic target and that statin therapy may be effective in inhibiting tumor progression. Ciliary cholesterol is involved in SHH signaling in skin basal cell carcinoma and medulloblastoma, while further investigations are needed to clarify which cilia-related signaling pathways are dependent on ciliary cholesterol for the application of statins and ciliary cholesterol agents as cancer therapeutics.

### References

Aros, C. J., Pantoja, C. J., and Gomperts, B. N. (2021). Wnt signaling in lung development, regeneration, and disease progression. *Commun. Biol.* 4 (1), 601. doi:10. 1038/s42003-021-02118-w

Athar, M., Li, C., Kim, A. L., Spiegelman, V. S., and Bickers, D. R. (2014). Sonic hedgehog signaling in Basal cell nevus syndrome. *Cancer Res.* 74 (18), 4967–4975. doi:10.1158/0008-5472.CAN-14-1666

Barakat, M. T., Humke, E. W., and Scott, M. P. (2013). Kif3a is necessary for initiation and maintenance of medulloblastoma. *Carcinogenesis* 34 (6), 1382–1392. doi:10.1093/ carcin/bgt041

Basten, S. G., and Giles, R. H. (2013). Functional aspects of primary cilia in signaling, cell cycle and tumorigenesis. *Cilia* 2 (1), 6. doi:10.1186/2046-2530-2-6

Beckwitt, C. H., Brufsky, A., Oltvai, Z. N., and Wells, A. (2018). Statin drugs to reduce breast cancer recurrence and mortality. *Breast Cancer Res.* 20 (1), 144. doi:10.1186/s13058-018-1066-z

Bidet, M., Joubert, O., Lacombe, B., Ciantar, M., Nehme, R., Mollat, P., et al. (2011). The hedgehog receptor patched is involved in cholesterol transport. *PLoS One* 6 (9), e23834. doi:10.1371/journal.pone.0023834

Byrne, E. F. X., Sircar, R., Miller, P. S., Hedger, G., Luchetti, G., Nachtergaele, S., et al. (2016). Structural basis of Smoothened regulation by its extracellular domains. *Nature* 535 (7613), 517–522. doi:10.1038/nature18934

# Author contributions

SK, ToM, KH, and TaM wrote the manuscript. HI, TY and TaM supervised.

## Funding

This work was supported by: Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan (to T.M.: 20K21845 and 21H02718); AMED-PRIME from the Japan Agency for Medical Research and Development, AMED (to T.M.: JP18gm5910011h0004); JST COI-NEXT (to T.M.: JPMJPF 2010); Chugai Foundation for Innovative Drug Discovery Science (to T.M.); and Astellas Foundation for Research on Metabolic Disorders (to T.M.).

## Acknowledgments

We thank Drs. Takeshi Itabashi and Hiroko Kishi for helpful discussion.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Castiella, T., Munoz, G., Luesma, M. J., Santander, S., Soriano, M., and Junquera, C. (2013). Primary cilia in gastric gastrointestinal stromal tumours (GISTs): An ultrastructural study. *J. Cell Mol. Med.* 17 (7), 844–853. doi:10.1111/jcmm.12067

Chae, W. J., and Bothwell, A. L. M. (2018). Canonical and non-canonical Wnt signaling in immune cells. *Trends Immunol.* 39 (10), 830–847. doi:10.1016/j.it.2018. 08.006

Daggubati, V., Hochstelter, J., Bommireddy, A., Choudhury, A., Krup, A. L., Kaur, P., et al. (2021). Smoothened-activating lipids drive resistance to CDK4/6 inhibition in Hedgehog-associated medulloblastoma cells and preclinical models. *J. Clin. Invest* 131 (6), e141171. doi:10.1172/JCI141171

Deshpande, I., Liang, J., Hedeen, D., Roberts, K. J., Zhang, Y., Ha, B., et al. (2019). Smoothened stimulation by membrane sterols drives Hedgehog pathway activity. *Nature* 571 (7764), 284–288. doi:10.1038/s41586-019-1355-4

Dvorak, J., Hadzi Nikolov, D., Dusek, L., Filipova, A., Richter, I., Buka, D., et al. (2016). Prognostic significance of the frequency of primary cilia in cells of small bowel and colorectal adenocarcinoma. *J. BUON* 21 (5), 1233–1241.

Emoto, K., Masugi, Y., Yamazaki, K., Effendi, K., Tsujikawa, H., Tanabe, M., et al. (2014). Presence of primary cilia in cancer cells correlates with prognosis of pancreatic ductal adenocarcinoma. *Hum. Pathol.* 45 (4), 817–825. doi:10.1016/j.humpath.2013. 11.017

Fabbri, L., Bost, F., and Mazure, N. M. (2019). Primary cilium in cancer hallmarks. *Int. J. Mol. Sci.* 20 (6), 1336. doi:10.3390/ijms20061336

Fu, W., Asp, P., Canter, B., and Dynlacht, B. D. (2014). Primary cilia control hedgehog signaling during muscle differentiation and are deregulated in rhabdomyosarcoma. *Proc. Natl. Acad. Sci. U S A* 111 (25), 9151–9156. doi:10.1073/pnas.1323265111

Garwood, E. R., Kumar, A. S., Baehner, F. L., Moore, D. H., Au, A., Hylton, N., et al. (2010). Fluvastatin reduces proliferation and increases apoptosis in women with high grade breast cancer. *Breast Cancer Res. Treat.* 119 (1), 137–144. doi:10.1007/s10549-009-0507-x

Guen, V. J., Chavarria, T. E., Kroger, C., Ye, X., Weinberg, R. A., and Lees, J. A. (2017). EMT programs promote basal mammary stem cell and tumor-initiating cell stemness by inducing primary ciliogenesis and Hedgehog signaling. *Proc. Natl. Acad. Sci. U S A* 114 (49), E10532–E10539. doi:10.1073/pnas.1711534114

Harfe, B. D., Scherz, P. J., Nissim, S., Tian, H., McMahon, A. P., and Tabin, C. J. (2004). Evidence for an expansion-based temporal Shh gradient in specifying vertebrate digit identities. *Cell* 118 (4), 517–528. doi:10.1016/j.cell.2004.07.024

Haseeb, M., Pirzada, R. H., Ain, Q. U., and Choi, S. (2019). Wnt Signaling in the Regulation of Immune Cell and Cancer Therapeutics. *Cells* 8 (11). doi:10.3390/ cells8111380

Hassounah, N. B., Bunch, T. A., and McDermott, K. M. (2012). Molecular pathways: The role of primary cilia in cancer progression and therapeutics with a focus on hedgehog signaling. *Clin. Cancer Res.* 18 (9), 2429–2435. doi:10.1158/1078-0432.CCR-11-0755

Hassounah, N. B., Nagle, R., Saboda, K., Roe, D. J., Dalkin, B. L., and McDermott, K. M. (2013). Primary cilia are lost in preinvasive and invasive prostate cancer. *PLoS One* 8 (7), e68521. doi:10.1371/journal.pone.0068521

He, S., Lu, Y., Liu, X., Huang, X., Keller, E. T., Qian, C. N., et al. (2015). Wnt3a: Functions and implications in cancer. *Chin. J. Cancer* 34 (12), 554–562. doi:10.1186/ s40880-015-0052-4

Huang, P., Zheng, S., Wierbowski, B. M., Kim, Y., Nedelcu, D., Aravena, L., et al. (2018). Structural basis of smoothened activation in hedgehog signaling. *Cell* 175 (1), 295–297. doi:10.1016/j.cell.2018.09.003

Izawa, I., Goto, H., Kasahara, K., and Inagaki, M. (2015). Current topics of functional links between primary cilia and cell cycle. *Cilia* 4, 12. doi:10.1186/s13630-015-0021-1

Jain, S., Song, R., and Xie, J. (2017). Sonidegib: Mechanism of action, pharmacology, and clinical utility for advanced basal cell carcinomas. *Onco Targets Ther.* 10, 1645–1653. doi:10.2147/OTT.S130910

Jiang, W., Hu, J. W., He, X. R., Jin, W. L., and He, X. Y. (2021). Statins: A repurposed drug to fight cancer. J. Exp. Clin. Cancer Res. 40 (1), 241. doi:10.1186/s13046-021-02041-2

Kim, S., Lee, K., Choi, J. H., Ringsrad, N., and Dynlacht, B. D. (2015). Nek2 activation of Kif24 ensures cilium disassembly during the cell cycle. *Nat. Commun.* 6, 8087. doi:10. 1038/ncomms.9087

Kinnebrew, M., Iverson, E. J., Patel, B. B., Pusapati, G. V., Kong, J. H., Johnson, K. A., et al. (2019). Cholesterol accessibility at the ciliary membrane controls hedgehog signaling. *Elife* 8, e50051. doi:10.7554/eLife.50051

Kinzel, D., Boldt, K., Davis, E. E., Burtscher, I., Trumbach, D., Diplas, B., et al. (2010). Pitchfork regulates primary cilia disassembly and left-right asymmetry. *Dev. Cell* 19 (1), 66–77. doi:10.1016/j.devcel.2010.06.005

Kuonen, F., Huskey, N. E., Shankar, G., Jaju, P., Whitson, R. J., Rieger, K. E., et al. (2019). Loss of primary cilia drives switching from hedgehog to ras/MAPK pathway in resistant basal cell carcinoma. *J. Invest Dermatol* 139 (7), 1439–1448. doi:10.1016/j.jid. 2018.11.035

Lee, J., Park, K. C., Sul, H. J., Hong, H. J., Kim, K. H., Kero, J., et al. (2021). Loss of primary cilia promotes mitochondria-dependent apoptosis in thyroid cancer. *Sci. Rep.* 11 (1), 4181. doi:10.1038/s41598-021-83418-3

Lee, K. H. (2020). Involvement of Wnt signaling in primary cilia assembly and disassembly. *FEBS J.* 287 (23), 5027–5038. doi:10.1111/febs.15579

Liu, H., Kiseleva, A. A., and Golemis, E. A. (2018). Ciliary signalling in cancer. Nat. Rev. Cancer 18 (8), 511–524. doi:10.1038/s41568-018-0023-6

Liu, L., Sheng, J. Q., Wang, M. R., Gan, Y., Wu, X. L., Liao, J. Z., et al. (2019). Primary cilia blockage promotes the malignant behaviors of hepatocellular carcinoma via induction of autophagy. *Biomed. Res. Int.* 2019, 5202750. doi:10.1155/2019/5202750

Longo, J., van Leeuwen, J. E., Elbaz, M., Branchard, E., and Penn, L. Z. (2020). Statins as anticancer agents in the era of precision medicine. *Clin. Cancer Res.* 26 (22), 5791–5800. doi:10.1158/1078-0432.CCR-20-1967

Marino, S. (2022). Subgroup-specific roles of primary cilia in medulloblastoma. *Genes Dev.* 36 (11-12), 650–651. doi:10.1101/gad.349856.122

Meiss, F., and Zeiser, R. (2014). Vismodegib. Recent Results Cancer Res. 201, 405–417. doi:10.1007/978-3-642-54490-3 25

Miyamoto, T., Hosoba, K., Akutsu, S. N., and Matsuura, S. (2022). Imaging of the ciliary cholesterol underlying the sonic hedgehog signal transduction. *Methods Mol. Biol.* 2374, 49–57. doi:10.1007/978-1-0716-1701-4\_5

Miyamoto, T., Hosoba, K., Itabashi, T., Iwane, A. H., Akutsu, S. N., Ochiai, H., et al. (2020). Insufficiency of ciliary cholesterol in hereditary Zellweger syndrome. *EMBO J.* 39 (12), e103499. doi:10.15252/embj.2019103499

Miyamoto, T., Hosoba, K., Ochiai, H., Royba, E., Izumi, H., Sakuma, T., et al. (2015). The microtubule-depolymerizing activity of a mitotic kinesin protein KIF2A drives primary cilia disassembly coupled with cell proliferation. *Cell Rep.* 10 (5), 664–673. doi:10.1016/j.celrep.2015.01.003

Nedelcu, D., Liu, J., Xu, Y., Jao, C., and Salic, A. (2013). Oxysterol binding to the extracellular domain of Smoothened in Hedgehog signaling. *Nat. Chem. Biol.* 9 (9), 557–564. doi:10.1038/nchembio.1290

Niewiadomski, P., Niedziolka, S. M., Markiewicz, L., Uspienski, T., Baran, B., and Chojnowska, K. (2019). Gli proteins: Regulation in development and cancer. *Cells* 8 (2), 147. doi:10.3390/cells8020147

Nishimura, Y., Yamakawa, D., Uchida, K., Shiromizu, T., Watanabe, M., and Inagaki, M. (2021). Primary cilia and lipid raft dynamics. *Open Biol.* 11 (8), 210130. doi:10.1098/ rsob.210130

Nobutani, K., Shimono, Y., Yoshida, M., Mizutani, K., Minami, A., Kono, S., et al. (2014). Absence of primary cilia in cell cycle-arrested human breast cancer cells. *Genes cells*. 19 (2), 141–152. doi:10.1111/gtc.12122

Park, H. S., Papanastasi, E., Blanchard, G., Chiticariu, E., Bachmann, D., Plomann, M., et al. (2021). ARP-T1-associated Bazex-Dupre-Christol syndrome is an inherited basal cell cancer with ciliary defects characteristic of ciliopathies. *Commun. Biol.* 4 (1), 544. doi:10.1038/s42003-021-02054-9

Phesse, T., Flanagan, D., and Vincan, E. (2016). Frizzled7: A promising achilles' heel for targeting the Wnt receptor complex to treat cancer. *Cancers (Basel)* 8 (5), 50. doi:10. 3390/cancers8050050

Pugacheva, E. N., Jablonski, S. A., Hartman, T. R., Henske, E. P., and Golemis, E. A. (2007). HEF1-dependent Aurora A activation induces disassembly of the primary cilium. *Cell* 129 (7), 1351–1363. doi:10.1016/j.cell.2007.04.035

Qi, X., Schmiege, P., Coutavas, E., Wang, J., and Li, X. (2018). Structures of human Patched and its complex with native palmitoylated sonic hedgehog. *Nature* 560 (7716), 128–132. doi:10.1038/s41586-018-0308-7

Raleigh, D. R., Sever, N., Choksi, P. K., Sigg, M. A., Hines, K. M., Thompson, B. M., et al. (2018). Cilia-associated oxysterols activate smoothened. *Mol. Cell* 72 (2), 316–327. doi:10.1016/j.molcel.2018.08.034

Ribes, V., and Briscoe, J. (2009). Establishing and interpreting graded sonic hedgehog signaling during vertebrate neural tube patterning: The role of negative feedback. *Cold Spring Harb. Perspect. Biol.* 1 (2), a002014. doi:10.1101/ cshperspect.a002014

Riscal, R., Skuli, N., and Simon, M. C. (2019). Even cancer cells watch their cholesterol. Mol. Cell 76 (2), 220-231. doi:10.1016/j.molcel.2019.09.008

Rozsypalova, A., Rosova, B., Filipova, A., Nikolov, D. H., Chloupkova, R., Richter, I., et al. (2019). Negative prognostic significance of primary cilia, CD8+ tumor infiltrating lymphocytes and PD1+ cells expression in clear cell renal cancer. *J. BUON* 24 (4), 1644–1651.

Seeley, E. S., Carriere, C., Goetze, T., Longnecker, D. S., and Korc, M. (2009). Pancreatic cancer and precursor pancreatic intraepithelial neoplasia lesions are devoid of primary cilia. *Cancer Res.* 69 (2), 422–430. doi:10.1158/0008-5472.CAN-08-1290

Sheng, R., Kim, H., Lee, H., Xin, Y., Chen, Y., Tian, W., et al. (2014). Cholesterol selectively activates canonical Wnt signalling over non-canonical Wnt signalling. *Nat. Commun.* 5, 4393. doi:10.1038/ncomms5393

Singla, V., and Reiter, J. F. (2006). The primary cilium as the cell's antenna: Signaling at a sensory organelle. *Science* 313 (5787), 629–633. doi:10.1126/science.1124534

Wang, J., Sinha, T., and Wynshaw-Boris, A. (2012). Wnt signaling in mammalian development: Lessons from mouse genetics. *Cold Spring Harb. Perspect. Biol.* 4 (5), a007963. doi:10.1101/cshperspect.a007963

Wang, W., Shiraishi, R., and Kawauchi, D. (2022). Sonic hedgehog signaling in cerebellar development and cancer. *Front. Cell Dev. Biol.* 10, 864035. doi:10.3389/fcell. 2022.864035

WHO Classification of Tumours Editorial Board (2021). Central nervous system tumours. 5th ed. Lyon, France: International Agency for Research on Cancer.

Wilson, M. M., Callens, C., Le Gallo, M., Mironov, S., Ding, Q., Salamagnon, A., et al. (2021). An EMT-primary cilium-GLIS2 signaling axis regulates mammogenesis and claudin-low breast tumorigenesis. *Sci. Adv.* 7 (44), eabf6063. doi:10.1126/sciadv.abf6063

Yang, J., Li, C., Shen, Y., Zhou, H., Shao, Y., Zhu, W., et al. (2020). Impact of statin use on cancer-specific mortality and recurrence: A meta-analysis of 60 observational studies. *Med. Baltim.* 99 (14), e19596. doi:10.1097/MD. 000000000019596

Youn, Y. H., Hou, S., Wu, C. C., Kawauchi, D., Orr, B. A., Robinson, G. W., et al. (2022). Primary cilia control translation and the cell cycle in medulloblastoma. *Genes Dev.* 36 (11-12), 737–751. doi:10.1101/gad.349596.122

Zhang, K., Da Silva, F., Seidl, C., Wilsch-Brauninger, M., Herbst, J., Huttner, W. B., et al. (2023). Primary cilia are WNT-transducing organelles whose biogenesis is controlled by a WNT-PP1 axis. *Dev. Cell* 58 (2), 139–154. e8. doi:10.1016/j.devcel. 2022.12.006