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*CORRESPONDENCE Chengwei Zhou, Ø fyzhouchengwei@nbu.edu.cn

[†]These authors have contributed equally to this work

RECEIVED 08 March 2024 ACCEPTED 26 July 2024 PUBLISHED 08 August 2024

CITATION

Wu X, Chen H, Ge Z, Luo B, Pan H, Shen Y, Xie Z and Zhou C (2024), A novel mitochondria-related algorithm for predicting the survival outcomes and drug sensitivity of patients with lung adenocarcinoma. *Front. Mol. Biosci.* 11:1397281. doi: 10.3389/fmolb.2024.1397281

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A novel mitochondria-related algorithm for predicting the survival outcomes and drug sensitivity of patients with lung adenocarcinoma

Xianqiao Wu^{1†}, Hang Chen^{2†}, Zhen Ge¹, Binyu Luo², Hanbo Pan³, Yiming Shen⁴, Zuorun Xie^{2†} and Chengwei Zhou^{1*}

¹Department of Thoracic Surgery, The First Affiliated Hospital of Ningbo University, Ningbo, Zhejiang, China, ²Department of Thoracic Surgery, Ningbo Medical Center LiHuiLi Hospital, Ningbo, Zhejiang, China, ³Shanghai Lung Cancer Center, Shanghai Chest Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China, ⁴Department of Otology and Skull Base Surgery, Eye Ear Nose and Throat Hospital, Fudan University, Shanghai, China

Background: Mitochondria have always been considered too be closely related to the occurrence and development of malignant tumors. However, the bioinformatic analysis of mitochondria in lung adenocarcinoma (LUAD) has not been reported yet.

Methods: In the present study, we constructed a novel and reliable algorithm, comprising a consensus cluster analysis and risk assessment model, to predict the survival outcomes and tumor immunity for patients with terminal LUAD.

Results: Patients with LUAD were classified into three clusters, and patients in cluster 1 exhibited the best survival outcomes. The patients in cluster 3 had the highest expression of *PDL1* (encoding programmed cell death 1 ligand 11) and *HAVCR2* (encoding Hepatitis A virus cellular receptor 2), and the highest tumor mutation burden (TMB). In the risk assessment model, patients in the low-risk group tended to have a significantly better survival outcome. Furthermore, the risk score combined with stage could act as a reliable independent prognostic indicator for patients with LUAD. The prognostic signature is a novel and effective biomarker to select anti-tumor drugs. Low-risk patients tended to have a higher expression of *CTLA4* (encoding cytotoxic T-lymphocyte associated protein 4) and *HAVCR2*. Moreover, patients in the high-risk group were more sensitive to Cisplatin, Docetaxel, Erlotinib, Gemcitabine, and Paclitaxel, while low-risk patients would probably benefit more from Gefitinib.

Abbreviations: LUAD, Lung Adenocarcinoma; PDL1, Drogrammed cell Death Ligand 1; HAVCR2, Hepatitis A Virus Cellular Receptor 2; TMB, tumor mutation burden; CTLA4, Cytotoxic T-Lymphocyte Associated protein 4; OS, overall survival; TIME, Tumor Immune Microenvironment; ICIs, Immune Checkpoint Inhibitors; IncRNAs, Long Noncoding RNAs; GTF, Gene Transfer Format; GO, Gene Ontology; mrRNA, Mitochondria-Related IncRNAs; DEmrIncRNAs, Differentially Expressed mrIncRNAs; IPS, Immunophenoscore; IC₅₀, half maximal Inhibitory Concentration; CDF, Cumulative Distribution Function.

Conclusion: We constructed a novel and reliable algorithm comprising a consensus cluster analysis and risk assessment model to predict survival outcomes, which functions as a reliable guideline for anti-tumor drug treatment for patients with terminal LUAD.

KEYWORDS

mitochondria, tumor immunity, consensus cluster, risk assessment model, lung adenocarcinoma

1 Introduction

As the most common malignant tumor worldwide, lung cancer is famous for its high mortality and high heterogeneity among malignant tumors (Oliver, 2022). Lung cancer has shown the highest estimated incidence and mortality in the United States for years (Siegel et al., 2021; Siegel et al., 2022). Similarly, lung cancer has the highest incidence and mortality among patients in China (Xia et al., 2022). As the most common pathological classification of lung cancer, lung adenocarcinoma (LUAD) accounts for approximately 60% of lung cancer, and is considered to be closely related to heredity and gene mutations (Warth et al., 2012). Under appropriate conditions, surgical treatment, especially a video-assisted thoracic surgery, remains the gold standard for the treatment for LUAD, which could dramatically prolong the overall survival (OS) of patients (Merchant et al., 2018). Although the emergence of other therapeutic methods (e.g., molecular targeted therapy and immunotherapy) have improved the life quality of patients with terminal stage lung cancer (Pilotto et al., 2015; Dong et al., 2019), the 5-year survival rate of patients with distant metastasis is only 7% (American Cancer Society, 2024). Therefore, exploring the complex pathogenesis of LUAD and seeking novel and reliable biomarkers are important.

The mitochondrion is a membrane-enclosed structure that produces energy for fundamental cell activities (Tan et al., 2017), which is also involves in hepatic lipid metabolism and oxidative stress (André, 1994; Mansouri et al., 2018). Recently, increasing evidence has demonstrated the crucial role of mitochondria in the occurrence and development of malignant tumors, and mitochondria might be an effective target for patients with cancer (Zhao Y. et al., 2013; Boland et al., 2013; Ubah and Wallace, 2014). For instance, Zhao J. et al. (2013b) proposed that mitochondria could promote the invasion and migration of malignancy by providing a large amount of adenosine triphosphate for pseudopodia. Furthermore, Villa et al. (2017) reported that mitochondria mediated the sensitivity of LUAD cells to chemotherapy drugs by regulating the autophagy signaling pathway. Moreover, Chang et al. revealed that dihydroergotamine tartrate, a drug used to treat migraine, acted on mitochondria in LUAD cells, thereby promoting apoptosis and mitochondrial autophagy (Chang et al., 2016). Thus, mitochondria are involved in the biological behavior of malignant tumor cells, especially LUAD cells.

Mitochondria can not only meet the energy demand of fundamental cellular activities, but also effectively regulate immune activities (Banoth and Cassel, 2018). Porporato et al. proposed that cancer cells could modify the tumor immune microenvironment

(TIME) and the immune response of the host by releasing dangerous signals and altering the metabolism of mitochondria (Porporato et al., 2018). Moreover, Klein et al. (2020) reported that mitochondrial oxidative phosphorylation inhibitors targeted cancer-related immune cells in the TIME, and played a crucial part in immune evasion in the occurrence and progression of cancer. Furthermore, Cloonan and Choi (2013) introduced the detailed role of mitochondria as sensors and mediators of innate immune receptor signaling. The precise coordination of oxidative stress between intracellular mitochondria and other organelles is crucial for cell survival. The dynamic balance of oxidative stress can not only coordinate complex cellular signaling events in cancer cells, but also affect other components of the tumor immune microenvironment (TIME). Immune cells, such as M2 macrophages, dendritic cells, and T cells, are the main components of immunosuppressive TMIE induced by oxidative stress (Kuo et al., 2022). Therefore, mitochondria are closely related to the immune activities of cells.

In recent years, immunotherapy has gradually become an effective tumor treatment strategy as an emerging tumor treatment strategy (Ribas and Wolchok, 2018). Unlike traditional radiotherapy and chemotherapy, immunotherapy is a treatment strategy that utilizes the human immune system to attack and eliminate cancer cells. It does not directly destroy tumor cells, but rather activates, enhances, or repairs the patient's own immune system to recognize and kill tumor cells. The most common immune checkpoint molecules are Programmed Death Ligand 1 (PD-L1) and Hepatitis A Virus Cellular Receiver 2 (HAVCR-2) (Pardoll, 2012). PD-L1 is an immune checkpoint protein that plays an important role in the immunotherapy of malignant tumors. It mainly inhibits T cell activity by binding to the Programmed Cell Death Protein 1 (PD-1) receptor, thereby reducing immune response and helping tumor cells evade immune surveillance. Therefore, inhibiting the interaction between PD-L1 or PD-1 can restore the activity of T cells and enhance the immune killing effect on tumor cells (Herbst et al., 2014; Sunshine and Taube, 2105). HAVCR-2, also known as T-cell Immunoglobulin and Mucin Domain 3 (TIM-3), is another important immune checkpoint molecule. It plays a crucial role in regulating the immune response process, especially in inhibiting T cell function. HAVCR-2 negatively regulates T cell activity by binding to its ligand, such as Galectin-9, and participates in regulating T cell depletion and immune tolerance phenomena. In immunotherapy, inhibition of HAVCR-2 is believed to enhance the anti-tumor effect of T cells, especially in patients who have failed treatment with PD-1/PD-L1 inhibitors and may play an important role. Therefore, HAVCR-2, as a potential target, is actively being studied and developed to expand and enhance the effectiveness

of immunotherapy (Sakuishi et al., 2010; Anderson et al., 2016; Das et al., 2017).

The potential relationships among mitochondria, tumor immunity, and LUAD have been reported. For example, Fei et al. (2022) found that mitochondrial topoisomerase I was closely related to immune cells and the expression of immune checkpoint inhibitors (ICIs) in patients with LUAD. However, there has been no bioinformatic study of consensus cluster analysis combined with prognostic signature for patients with LUAD. In addition, mitochondria also participate in the expression of PD-L1 and HAVCR-2 in tumor cells, demonstrating an undeniable role in tumor occurrence and development. The latest research indicates that mitochondria are involved in the localization regulation of PD-L1 protein on the outer membrane and mitochondria. Enhancing mitochondrial autophagy helps to degrade mitochondrial localization PD-L1, thereby overcoming the resistance of TNBC to chemotherapy and immunotherapy (Jabbarzadeh et al., 2020). The research results in this area are of great significance for enhancing the efficacy of targeted PD-1/PD-L1 therapy. In addition, studies have found that mitochondrial autophagy can enhance the therapeutic effect of ICI combined with paclitaxel by degrading mitochondrial distribution PD-L1, which can be inhibited by ATAD3A protein (Xie et al., 2023). Besides, in human colorectal cancer cancer cells, mitochondrial dysfunction inhibits the expression of HAVCR-2, thereby affecting the immune escape of tumor cells (Sakhnevych et al., 2019). Thus, fully understanding the role of mitochondria in the development of LUAD might provide theoretical guidance and new strategies for future mitochondrial targeted therapy.

In the present study, we established a novel and reliable algorithm compromising molecular subtypes and a risk assessment model to predict prognosis and select sensitive anti-tumor drugs for patients with LUAD.

2 Materials and methods

2.1 Data download

The gene expression at the transcriptome level and corresponding clinical information were downloaded from LUAD project of The Cancer Genome Atlas database (https://portal. gdc.cancer.gov/). Subsequently, the mRNAs and long noncoding RNAs (lncRNAs) were annotated using gene transfer format (GTF) files obtained from Ensembl. A list of mitochondria-related genes (mrgenes) were downloaded from The Gene Ontology Resource (GO, http://geneontology.org/) (The Gene Ontology Consortium, 2019). The mitochondria-related lncRNAs (mrlncRNAs) were identified by performing a Spearman correlation analysis between genes related to mitochondria and lncRNAs (|cor| > 0.4, P < 0.001). Differentially expressed mrlncRNAs (DEmrlncRNAs) were filtered using differential expression analysis ($|\log FC| > 1$, false discovery rate <0.001), and DEmrlncRNAs closely related to survival were screened using a univariate Cox analysis (P < 0.01), which were visualized using a volcano map and a forest map. We obtained mrlncRNAs that were closely related to the occurrence of LUAD and the OS of patients with LUAD, which were the foundation for the subsequent construction of the consensus cluster analysis and risk assessment model.

2.2 Molecular subtypes according to DEmrlncRNAs

The patients with LUAD were classified into different molecular subtypes based on the expression of DEmrlncRNAs by running the ConsensusClusterPlus package (Wilkerson and Hayes, 2010). Then, the survival outcomes of the patients with different molecular subtypes were explored by performing a Kaplan-Meier survival analysis. According to the National Comprehensive Cancer Network guidelines, the expression of common ICIs could reflect the reactivity of patients with LUAD to immunotherapy approximately, which could benefit a large number of patients with a terminal stage tumor (Ettinger et al., 2021). The expression levels of common ICIs (e.g., PDL1 (encoding programmed cell death 1 ligand 1) and HAVCR2 [encoding Hepatitis A virus cellular receptor 2)] were compared between patients from different clusters, and a series of boxplots were generated for visualization, which were marked as: * * * P < 0.001; * * P < 0.01; and * P < 0.05. To better comprehend the relative abundance of stromal cells and immune cells in the TIME, the StromalScore, ImmuneScore, and ESTIMATEScore were calculated using the estimate package, which were subsequently compared between different clusters. The tumor mutation burden (TMB) represents the total number of mutations per million bases, which is used as a rough indicator of the efficacy of immunotherapy (Forde et al., 2018; Anagnostou et al., 2020; Marabelle et al., 2020). Several Wilcoxon rank-sum tests were performed to investigate whether there was a statistical difference in the TMB between patients from different clusters.

2.3 Risk assessment model based on DEmrlncRNAs

To better verify the predictive capability of the constructed risk assessment model, the patients with LUAD were equally divided into a training group and a test group randomly. To prevent over-fitting of the constructed model, the Least absolute shrinkage and selection operator regression combined with a multivariate Cox regression analysis were conducted on the DEmrlncRNAs of patients in the training group to construct a novel prognostic signature related to mitochondria. Then, the risk scores of patients in the training group were calculated using the following formula:

$$Riskscore = \sum_{i=1,}^{15} Coef(i) \times E(i),$$

Where E(i) and Coef (i) are the expression and the regression coefficients of the DEmrlncRNAs, respectively. The median value of the risk score for patients in the training group was used as the cut-off point to classify patients into high- and low-risk groups. Subsequently, the regression coefficients and cut-off point of patients in the test group were determined to be in full accordance with those of the patients in train group. Kaplan–Meier survival



analyses were conducted to exhibit the survival outcomes of patients in the different risk groups. To evaluate the predictive ability of the risk assessment model, the receiver operating characteristic curves were plotted and the area under the curve was calculated, respectively. To explore the potential relationship between the risk score and survival status, four scatter plots were plotted for visualization. Univariate and multivariate Cox regression analyses were conducted to investigate whether the risk assessment model could function as a reliable independent prognostic indicator for patients with LUAD, which was visualized using four forest maps. Furthermore, to filter the patients with LUAD whose prognosis could be predicted accurately using the risk assessment model, a series of Kaplan-Meier survival analyses were performed for validation. A clinical heatmap was used to exhibit the expression levels of 15 DEmrlncRNAs included in the modeling process, which revealed the potential relationships between the risk group and common clinicopathological characteristics [e.g., node (N), metastasis (M), tumor (T), stage, gender, age, ImmuneScore, and Cluster], in which the clinicopathological characteristics closely related to the risk groups were discussed in detail. Furthermore, to study the response of patients with LUAD to immunotherapy, the TMB and the expression levels of CTLA4 (encoding cytotoxic T-lymphocyte associated protein 4) and HAVCR2 in patients in the different risk groups were compared using Wilcoxon ranksum tests. Moreover, we used the immunophenoscore (IPS), which represents gene expression levels in immune cells closely related to the tumor, including lymphocytes and macrophages, which has been utilized to assess the response to immunotherapy targeting

PD-L1 and CTLA-4 (Charoentong et al., 2020). The IPS of each patient was downloaded from The Cancer Immunome Atlas (https://tcia.at/) (Prior et al., 2013), which were compared between different risk groups. The statistical differences in StromalScore, ImmuneScore, and ESTIMATEScore between the different risk groups were explored using Wilcoxon rank-sum tests. Singlesample gene-set enrichment analysis was used to quantify the relative abundance of common immune cells and the relative activity of common immune-related signaling pathways, which were compared between different risk groups. To explore the functions and signaling pathways closely related to the risk groups, two bar-plots were plotted for GO and Kyoto Encyclopedia of Genes and Genomes functional enrichment analyses for visualization. A Sankey diagram was generated to visualize the relationship between the molecular subtypes and the risk assessment model. The half maximal inhibitory concentration (IC₅₀) represents the concentration of an anti-tumor drugs that inhibits half of the tumors cells, which could effectively measure the reaction of patients with LUAD to anti-tumor drugs (Sebaugh, 2011). In the present study, the pRRophetic package was run to evaluate the IC₅₀ of common antitumor drugs including chemotherapy (e.g., Cisplatin, Docetaxel, Gemcitabine, and Paclitaxel) and molecular targeted therapy (e.g., Gefitinib and Erlotinib). A nomogram was plotted to display the calculation process of the risk scores for clinical patients, the predictive capability of which was evaluated using one-, three-, and five-year correction curves. Finally, a survival curve exhibited the survival outcomes of patients from different molecular subtypes and risk groups.



FIGURE 2

The Consensus cluster analysis based on DEmrIncRNAs. (A) 1,724 DEmrIncRNAs were filtered by differential expression analysis, among which 276 showed low expression, and 1,448 showed high expression in patients with LUAD. (B) 76 DEmrIncRNAs were considered to be closely related to prognosis using a univariate Cox analysis, 15 of which were included in the modeling process. (C–E) When the k value was 3, the variation of cumulative distribution function (CDF) was the smallest, and the relative change in area under the CDF curve was the highest. (F) Therefore, the patients with LUAD were classified into three 3 clusters, and patients in cluster 1 exhibited the best survival outcomes, which were statistically significant. (G, H) According to the expression of ICIs, the patients in cluster 3 had the highest PD-L1 and HAVCR-2 expression, followed by the patients in cluster 1, and the patients in cluster 2 had the lowest expression, which were all statistically significant. (I–K) The patients in cluster 2 had the lowest dhat they had the lowest abundance of stromal cells and immune cells, while there was no statistical significance between the remaining two groups.



FIGURE 3 Prognostic signature based on DEmrlncRNAs. (A) The patients in cluster 3 had the highest TMB. (B, C) 236 Patients were classified into the training group, while 232 patients were arranged into the test group. (D, E) The patients in the low-risk group tended to have a significantly better survival outcome. (F, G) The area under the curve value of the training group and the test group were 0.830 and 0.708, respectively. (H, I) Furthermore, with the accumulation of the risk score, the number of patients who died increased significantly.

 $\ensuremath{\mathsf{TABLE1}}$ The regression coefficients of mrlnRNAs included in the lasso regression.

Gene	Coef
AC090559.1	-0.0808422085604288
AL353152.1	-0.0128420568313749
MIR223HG	-0.0183021082479998
AC018647.1	-1.20900113082152
AC099850.4	0.011776662913192
AC021755.3	-0.0600844102402713
LINC01116	0.030971078922924
AC010999.2	-0.948844016068309
AL365181.2	0.0130238569092484
AL049836.1	0.0103501622344718
HIF1A-AS1	0.0823378698246388
LINC02323	0.124240788617215
TARID	0.454314559680914
LINC00578	-0.0348236259612791
AC027031.2	0.0131456798726987

3 Results

3.1 Identification of DEmrlncRNAs

As shown in Figure 1, a multi-step approach was carried out according to the flowchart. We downloaded a total of 551 samples (497 LUAD tissues and 54 normal tissues) from The Cancer Genome Atlas database. Then, we downloaded 1,838 mrgenes from the GO knowledgebase, and obtained 14,087 lncRNAs. After annotation, 3,546 lncRNAs were identified as mrlncRNAs by performing correlation analyses. Next, we identified 1,724 DEmrlncRNAs by differential expression analysis, of which 276 were downregulated, and 1,448 were upregulated in patients with LUADs (Figure 2A). Then, 76 DEmrlncRNAs were identified as DEmrlncRNAs closely related to prognosis using a univariate Cox analysis, 15 of which were included in the modeling process (Figure 2B).

3.2 The molecular subtype is a reliable indicator for tumor immunity

When the k value was three, the variation of the cumulative distribution function (CDF) was the smallest, and the relative change in the area under CDF curve was the highest (Figures 2C–E). Therefore, the patients with LUAD were classified into three clusters, and the patients in cluster 1 exhibited the best survival

outcomes among the clusters, with statistical significance (Figure 2F). According to the expression of ICIs, the patients in cluster 3 had the highest *PDL1* (Figure 2G) and *HAVCR2* (Figure 2H) expression, followed by the patients in cluster 1; the patients in cluster 2 had the lowest expression, which were all statistically significant. Furthermore, the patients in cluster 3 had the highest TMB (Figure 3A), which indicated that they might be most sensitive to immunotherapy targeting PD-L1 and HAVCR-2. The patients in cluster 2 had the lowest StromalScore, ImmuneScore, and ESTIMATEScore, which suggested that they had the lowest abundance of stromal cells and immune cells, while there was no statistical significance between the remaining two groups (Figure 2I–K).

3.3 The prognostic signature acts as a reliable biomarker for patients with LUAD

The risk score of patients with LUAD were calculated with the formula, and the regression coefficients of the DEmrlncRNAs were listed in Table 1. Patients (n = 236) were classified into the training group, while 232 patients were classified into the test group (Figures 3B, C), in which patients in the low-risk group tended to have a significantly better survival outcome (Figures 3D, E). The area under the curve values of the training group and the test group were 0.830 (Figure 3F) and 0.708 (Figure 3G), respectively, which suggested that the prognostic signature had a relatively better predictive capability for patients with LUAD. Furthermore, with increasing risk score, the number of patients who died increased significantly (Figures 3H, I). Moreover, the risk score (hazard ratio = 1.614 [confidence interval 1.383-1.884], P < 0.001) and stage (hazard ratio = 1.803 [confidence interval 1.456–2.231], *P* < 0.001), could act as reliable independent prognostic indicators for patients with LUAD based on univariate and multivariate Cox regression (Figures 4A-D). According to a series of survival analyses, the prognostic signature exhibited the best predictive ability among patients without distant metastasis (Figure 5D), regardless of age (Figure 4E), sex (Figure 4F), stage (Figure 5A), T (Figure 5B), and N (Figure 5C). The clinical heatmap (Figure 6A) revealed that the prognostic signature was closely related to N (Figure 6B), T (Figure 6C), stage (Figure 6D), ImmuneScore (Figure 6E), and molecular subtypes (Figure 6F). This indicated that high-risk patients tended to have a later stage of LUAD and always had relatively poor survival outcomes.

The prognostic signature is a novel and effectively biomarker to select anti-tumor drugs. The low-risk patients tended to have a higher expression of *CTLA4* (Figure 6G) and *HAVCR2* (Figure 6H) with a lower TMB (Figure 6I), suggesting that they might be more sensitive to immunotherapy targeting CTLA-4 and HAVCR-2. According to the IPS value, low-risk patients would always benefit from anti-PD-L1 therapy (Figure 7A), anti-CTLA-4 therapy (Figure 7B), and their combination (Figure 7C), with statistical significance. The low-risk patients possessed a higher abundance of stromal cells and immune cells, based on the estimate algorithm (Figures 7D–F). Furthermore, the low-risk patients had a higher content of common immune cells (Figure 7G) and more active immune-related signaling pathways (Figure 7H). Therefore, compared with the high-risk patients, the patients in low-risk group



tended to have stronger tumor immunity. According to the GO functional enrichment analysis, the mitochondria-related signature was closely related to the process of mitosis, including mitotic sister chromatid segregation, mitotic nuclear division, chromosome

segregation, and sister chromatid segregation (Figure 7I). Similarly, the mitosis-related signaling pathways were enriched in the risk assessment model, such as mitotic sister chromatid segregation, mitotic nuclear division, chromosome segregation, nuclear division,



and organelle fission (Figure 8A). The majority of high-risk patients were from cluster 2, the most of low-risk patients were from cluster 3, while 80% of the patients from cluster 1 were low-risk (Figure 8B).

The risk assessment model could function as a robust guideline for clinical medication using common chemotherapies and molecular targeted therapy. For example, patients in the high-risk group were more sensitive to Cisplatin (Figure 8C),



Docetaxel (Figure 8D), Erlotinib (Figure 8E), Gemcitabine (Figure 8G), and Paclitaxel (Figure 8H), while low-risk patients would probably benefit more from Gefitinib (Figure 8F). The nomogram simplified the calculation process of the risk score, and provided the corresponding approximate one-,

three-, and five-year survival rates based on the calculated risk score (Figure 9A), in which the nomogram exhibited the best predictive capability for 1-year survival (Figure 9B). According to the multi-survival curve, the high-risk patients in cluster 2 and cluster 3 exhibited poor survival outcomes,



FIGURE 7

The prognostic signature is a novel and effectively biomarker to select anti-tumor drugs (I). (A–C) Low-risk patients always benefitted from anti-PD-L1 therapy (A), anti-CTLA-4 therapy (B), and their combination (C), with statistical significance. (D–F) The low-risk patients possessed a higher abundance of stromal cells and immune cells based on the estimate algorithm. (G, H) The low-risk patients had a higher content of common immune cells (G), and more active immune-related signaling pathways (H). (I) Compared with the high-risk patients, the patients in the low-risk group tended to have stronger tumor immunity.

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The prognostic signature is a novel and effective biomarker to select anti-tumor drugs (II). (A) The mitotic-related signaling pathways were enriched in the risk assessment model, such as mitotic sister chromatid segregation, mitotic nuclear division, chromosome segregation, nuclear division, and organelle fission, etc. (B) The majority of high-risk patients were from cluster 2, the most low-risk patients were from cluster 3, while 80% of the patients from cluster 1 were low-risk. (C–F) The patients in high-risk group were more sensitive to Cisplatin (C), Docetaxel (D), Erlotinib (E), Gemcitabine (G), and Paclitaxel (H); while the low-risk patients probably benefitted more from Gefitinib (F).

while the low-risk patients in cluster 1 had a relative survival advantage (Figure 9C).

4 Discussion

Changes in mitochondrial homeostasis are closely related to many human diseases, such as cancer, neurodegenerative diseases [Parkinson's disease (Macdonald et al., 2018), Alzheimer's disease (Swerdlow, 2018), and Huntington's disease (Jodeiri Farshbaf and Ghaedi, 2017)], and myopathy (Kim, 2017; Xie et al., 2020; Andrieux et al., 2021). Recently, the bioinformatics-based construction of prognostic signatures to predict the prognosis and guide treatment for patients with the terminal stage of malignant tumors has become a research hotspot (Chen et al., 2021a; Chen et al., 2021b; Zhang et al., 2024). Zhuo et al. (2021) established a mitophagy-related signature to explore the survival outcomes, tumor immunity, mutation, and chemotherapy response in pancreatic cancer. In addition, Zhang et al. (2021) constructed a mitochondria-related signature to explore the TIME, infiltration of immune cells, and immunotherapy of patients with hepatocellular carcinoma.



In the present study, we identified DEmrlncRNAs related to prognosis, which were utilized for subsequent consensus cluster analysis and prognostic signature construction. The patients with LUAD were classified into three clusters, and patients in cluster 1 exhibited the best survival outcomes. The patients in cluster 3 had the highest expression of *PDL1* and *HAVCR2*, and the highest TMB, which indicated that they might be the most sensitive to immunotherapy. The patients in cluster 2 had the lowest StromalScore, ImmuneScore, and ESTIMATEScore. According to the risk assessment model, patients in the low-risk group tended to have a significantly better survival outcome than the patients in the other groups. Furthermore, the risk score and stage could act as reliable independent prognostic indicators for patients with LUAD. The prognostic signature exhibited an excellent predictive ability among patients without distant metastasis, regardless of age, sex, stage, T, and N. The clinical heatmap revealed that the prognostic signature was closely related to N, T, stage, ImmuneScore, and molecular subtypes.

The prognostic signature is a novel and effective biomarker to select anti-tumor drugs. The low-risk patients tended to have a higher expression of CTLA4 and HAVCR2, with a lower TMB. According to the IPS value, the low-risk patients would always benefit from anti-PD-L1 therapy, anti-CTLA4 therapy, and their combination, which was significantly different. Furthermore, compared with the high-risk patients, the patients in the low-risk group tended to have a stronger tumor immunity. Furthermore, the risk assessment model could function as a robust guideline to select clinical medication comprising common chemotherapy and molecular targeted therapy. For example, patients in the high-risk group were more sensitive to Cisplatin, Docetaxel, Erlotinib, Gemcitabine, and Paclitaxel, while the low-risk patients would probably benefit more from Gefitinib. The high-risk patients in cluster 2 and cluster 3 exhibited poor survival outcomes, while the low-risk patients in cluster 1 showed a relative survival advantage.

The present study was the first to carry out bioinformatic analyses closely related to mitochondria and patients with LUAD. Furthermore, compared with traditional modeling process, we established a novel mitochondria-related algorithm containing a consensus cluster analyses and risk assessment model, in which patients with LUAD were classified into six groups that received different treatment strategies based on the corresponding groups. Moreover, least absolute shrinkage and selection operator regression analysis was carried out together with a multivariate Cox regression analysis to avoid overfitting of the model.

Although the algorithm might function as a guideline for clinical medication, there are also several limitations. Firstly, we conducted the internal verification using The Cancer Genome Atlas database, rather than other databases (e.g., GEO datasets). Secondly, all analyses were confined to bioinformatic analyses, and the study lacks validation of clinical specimens and molecular biological experiments, which are necessary for the clinical application of this algorithm.

Finally, based on the results of this manuscript, we have provided prospects for future potential research directions in this field. In recent years, the additional molecular pathways and incorporating multi-omics data integrations may play an increasingly important role in exploring the potential mechanisms of LUAD progression and treatment response. Nowadays, more and more research focuses on transcriptome sequencing. For example, studies based on transcriptome sequencing have demonstrated through *in vitro* and *in vivo* functional and mechanistic experiments that B4GALT1 promotes immune escape at both transcriptional and post transcriptional levels, thereby promoting the progression of LUAD (Cui et al., 2023). In addition, there are also studies based on single-cell RNA sequencing analysis data, which calculate the immunogenic cell death value of cells to construct a set of prognostic models that can predict the prognosis of LUAD patients and immunotherapy, and to some extent guide the clinical treatment of LUAD patients (Zhang et al., 2023a). More interestingly, researchers explored the role of Treg cells in Esophageal squamous cell carcinoma by combining single-cell RNA sequencing and bulk RNA-seq analysis, in order to predict patient prognosis and immune therapy responsiveness as a prognostic model (Zhang et al., 2023b).

In the present study, we constructed a novel and reliable algorithm comprising a consensus cluster analysis and risk assessment model to predict the survival outcomes, which function as a reliable guideline to select anti-tumor drugs to treat patients with terminal LUAD, which might provide a theoretical foundation for customized individualized treatment.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/supplementary material.

Author contributions

XW: Validation, Writing-review and editing. HC: Formal Analysis, Software, Writing-original draft, Writing-review and editing. ZG: Software, Writing-original draft. BL: Software, Writing-original draft. HP: Validation, Writing-original draft. YS: Validation, Writing-original draft. ZX: Validation, Writing-original draft. CZ: Conceptualization, Writing-original draft, Funding acquisition.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. The study was funded by Basic public welfare project of Ningbo (Grant No. 2022S042).

Acknowledgments

We thank the TCGA database for generously sharing a large amount of data.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

American Cancer Society (2024). Non-small cell lung cancer survival rates by stage[OL]. Available at: www.cancer.org/cancer/non-small-cell-lung-cancer/detection-diagnosis-staging/survival-rates.html (Date last updated: 04/20/21).

Anagnostou, V., Bruhm, D., Niknafs, N., White, J., Shao, X., Sidhom, J., et al. (2020). Integrative tumor and immune cell multi-omic analyses predict response to immune checkpoint blockade in melanoma. *Cell Rep. Med.* 1 (8), 100139. doi:10.1016/j.xcrm.2020.100139

Anderson, A., Joller, N., and Kuchroo, V. (2016). Lag-3, tim-3, and TIGIT: Coinhibitory receptors with specialized functions in immune regulation. *Immunity* 44 (5), 989–1004. doi:10.1016/j.immuni.2016.05.001

André, J. (1994). Mitochondria. *Biol. Cell* 80, 103–106. doi:10.1111/j.1768-322x.1994.tb00915.x

Andrieux, P., Chevillard, C., Cunha-Neto, E., and Nunes, J. (2021). Mitochondria as a cellular hub in infection and inflammation. *Int. J. Mol. Sci.* 22 (21), 11338. doi:10.3390/ijms222111338

Banoth, B., and Cassel, S. (2018). Mitochondria in innate immune signaling. *Transl. Res.* 202, 52–68. doi:10.1016/j.trsl.2018.07.014

Boland, M., Chourasia, A., and Macleod, K. (2013). Mitochondrial dysfunction in cancer. *Front. Oncol.* 3, 292. doi:10.3389/fonc.2013.00292

Chang, S., Lee, A., Yu, K., Park, J., Kim, K., and Cho, M. (2016). Dihydroergotamine tartrate induces lung cancer cell death through apoptosis and mitophagy. *Chemotherapy* 61 (6), 304–312. doi:10.1159/000445044

Charoentong, P., Finotello, F., Angelova, M., Mayer, C., Efremova, M., Rieder, D., et al. (2020). Pan-cancer immunogenomic analyses reveal genotype-immunophenotype relationships and predictors of response to checkpoint blockade. *Cell Rep.* 18 (1), 248–262. doi:10.1016/j.celrep.2016.12.019

Chen, H., Hu, Z., Sang, M., Ni, S., Lin, Y., Wu, C., et al. (2021b). Identification of an autophagy-related lncRNA prognostic signature and related tumor immunity research in lung adenocarcinoma. *Front. Genet.* 12, 767694. doi:10.3389/ fgene.2021.767694

Chen, H., Shen, W., Ni, S., Sang, M., Wu, S., Mu, Y., et al. (2021a). Construction of an immune-related lncRNA signature as a novel prognosis biomarker for LUAD. *Aging (Albany NY)* 13 (16), 20684–20697. doi:10.18632/aging.203455

Cloonan, S., and Choi, A. (2013). Mitochondria: sensors and mediators of innate immune receptor signaling. *Curr. Opin. Microbiol.* 16 (3), 327-338. doi:10.1016/j.mib.2013.05.005

Cui, Y., Li, J., Zhang, P., Yin, D., Wang, Z., Dai, J., et al. (2023). B4GALT1 promotes immune escape by regulating the expression of PD-L1 at multiple levels in lung adenocarcinoma. *J. Exp. Clin. Cancer Res.* 42, 146. doi:10.1186/s13046-023-02711-3

Das, M., Zhu, C., and Kuchroo, V. (2017). Tim-3 and its role in regulating anti-tumor immunity. *Immunol. Rev.* 276 (1), 97–111. doi:10.1111/imr.12520

Dong, J., Li, B., Lin, D., Zhou, Q., and Huang, D. (2019). Advances in targeted therapy and immunotherapy for non-small cell lung cancer based on accurate molecular typing. *Front. Pharmacol.* 10, 230. doi:10.3389/fphar.2019.00230

Ettinger, D., Wood, D., Aisner, D., Akerley, W., Bauman, J., Bharat, A., et al. (2021). NCCN guidelines insights: non-small cell lung cancer, version 2.2021. J. Natl. Compr. Canc Netw. 19 (3), 254–266. doi:10.6004/jnccn.2021.0013

Fei, L., Lu, Z., Xu, Y., and Hou, G. (2022). A comprehensive pan-cancer analysis of the expression characteristics, prognostic value, and immune characteristics of *TOP1MT*. *Front. Genet.* 13, 920897. doi:10.3389/fgene.2022.920897

Forde, P., Chaft, J., Smith, K., Anagnostou, V., Cottrell, T., Hellmann, M., et al. (2018). Neoadjuvant PD-1 blockade in resectable lung cancer. *N. Engl. J. Med.* 378 (21), 1976–1986. doi:10.1056/NEJMoa1716078

Herbst, R., Soria, J., Kowanetz, M., Fine, G., Hamid, O., Gordon, M., et al. (2014). Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. *Nature* 515 (7528), 563–567. doi:10.1038/nature14011

Jabbarzadeh, K., Salimian, F., Aghapour, S., Xiang, S., Zhao, Q., Li, M., et al. (2020). Akt-targeted therapy as a promising strategy to overcome drug resistance in breast cancer - a comprehensive review from chemotherapy to immunotherapy. *Pharmacol. Res.* 156, 104806. doi:10.1016/j.phrs.2020.104806

Jodeiri Farshbaf, M., and Ghaedi, K. (2017). Huntington's disease and mitochondria. *Neurotox. Res.* 32 (3), 518–529. doi:10.1007/s12640-017-9766-1 organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Kim, J. (2017). Myopathy, drugs, and mitochondria. J. Korean Med. Sci. 32 (11), 1732–1733. doi:10.3346/jkms.2017.32.11.1732

Klein, K., He, K., Younes, A., Barsoumian, H., Chen, D., Ozgen, T., et al. (2020). Role of mitochondria in cancer immune evasion and potential therapeutic approaches. *Front. Immunol.* 11, 573326. doi:10.3389/fmmu.2020.573326

Kuo, C., Babuharisankar, A. P., Lin, Y., Lien, H., Lo, Y., Chou, H., et al. (2022). Mitochondrial oxidative stress in the tumor microenvironment and cancer immunoescape: foe or friend? *J. Biomed. Sci.* 29 (1), 74. doi:10.1186/s12929-022-00859-2

Macdonald, R., Barnes, K., Hastings, C., and Mortiboys, H. (2018). Mitochondrial abnormalities in Parkinson's disease and Alzheimer's disease: can mitochondria be targeted therapeutically? *Biochem. Soc. Trans.* 46 (4), 891–909. doi:10.1042/BST20170501

Mansouri, A., Gattolliat, C., and Asselah, T. (2018). Mitochondrial dysfunction and signaling in chronic liver diseases. *Gastroenterology* 155 (3), 629–647. doi:10.1053/j.gastro.2018.06.083

Marabelle, A., Fakih, M., Lopez, J., Shah, M., Shapira-Frommer, R., Nakagawa, K., et al. (2020). Association of tumour mutational burden with outcomes in patients with advanced solid tumours treated with pembrolizumab: prospective biomarker analysis of the multicohort, open-label, phase 2 KEYNOTE-158 study. *Lancet Oncol.* 21 (10), 1353–1365. doi:10.1016/S1470-2045(20)30445-9

Merchant, N., McKenna, R., and Onugha, O. (2018). Is there a role for VATS sleeve lobectomy in lung cancer? *Surg. Technol. Int.* 32, 225–229.

Oliver, A. (2022). Lung cancer: epidemiology and screening. Surg. Clin. N. Am. 102 (3), 335–344. doi:10.1016/j.suc.2021.12.001

Pardoll, D. (2012). The blockade of immune checkpoints in cancer immunotherapy. *Nat. Rev. Cancer* 12 (4), 252–264. doi:10.1038/nrc3239

Pilotto, S., Molina-Vila, M., Karachaliou, N., Carbognin, L., Viteri, S., González-Cao, M., et al. (2015). Integrating the molecular background of targeted therapy and immunotherapy in lung cancer: a way to explore the impact of mutational landscape on tumor immunogenicity. *Transl. Lung Cancer Res.* 4 (6), 721–727. doi:10.3978/j.issn.2218-6751.2015.10.11

Porporato, P., Filigheddu, N., Pedro, J., Kroemer, G., and Galluzzi, L. (2018). Mitochondrial metabolism and cancer. *Cell Res.* 28 (3), 265–280. doi:10.1038/cr.2017.155

Prior, F., Clark, K., Commean, P., Freymann, J., Jaffe, C., Kirby, J., et al. (2013). TCIA: an information resource to enable open science. *Annu. Int. Conf. IEEE Eng. Med. Biol. Soc.* 2013, 1282–1285. doi:10.1109/EMBC.2013.6609742

Ribas, A., and Wolchok, J. (2018). Cancer immunotherapy using checkpoint blockade. Science 359 (6382), 1350–1355. doi:10.1126/science.aar4060

Sakhnevych, S., Yasinska, I., Fasler-Kan, E., and Sumbayev, V. (2019). Mitochondrial defunctionalization supresses tim-3-galectin-9 secretory pathway in human colorectal cancer cells and thus can possibly affect tumor immune escape. *Front. Pharmacol.* 10, 342. doi:10.3389/fphar.2019.00342

Sakuishi, K., Apetoh, L., Sullivan, J., Blazar, B., Kuchroo, V., and Anderson, A. (2010). Targeting Tim-3 and PD-1 pathways to reverse T cell exhaustion and restore anti-tumor immunity. *J. Exp. Med.* 207 (10), 2187–2194. doi:10.1084/jem.20100643

Sebaugh, J. (2011). Guidelines for accurate EC50/IC50 estimation. *Pharm. Stat.* 10 (2), 128–134. doi:10.1002/pst.426

Siegel, R., Miller, K., Fuchs, H., and Jemal, A. (2021). Cancer statistics, 2021. CA-Cancer J. Clin. 71 (1), 7-33. doi:10.3322/caac.21654

Siegel, R., Miller, K., Fuchs, H., and Jemal, A. (2022). Cancer statistics, 2022. CA-Cancer J. Clin. 72 (1), 7–33. doi:10.3322/caac.21708

Sunshine, J., and Taube, J. (2105). PD-1/PD-L1 inhibitors. Curr. Opin. Pharmacol. 23, 32–38. doi:10.1016/j.coph.2015.05.011

Swerdlow, R. (2018). Mitochondria and mitochondrial cascades in Alzheimer's disease. J. Alzheimers Dis. 62 (3), 1403-1416. doi:10.3233/JAD-170585

Tan, K., Li, C., Li, Y., Fei, J., Yang, B., Fu, Y., et al. (2017). Real-time monitoring atp in mitochondrion of living cells: a specific fluorescent probe for atp by dual recognition sites. *Anal. Chem.* 89 (3), 1749–1756. doi:10.1021/acs.analchem.6b04020

The Gene Ontology Consortium (2019). The gene Ontology resource: 20 years and still GOing strong. *Nucleic Acids Res.* 47, D330–D338. doi:10.1093/nar/gky1055

Ubah, O., and Wallace, H. (2014). Cancer therapy: targeting mitochondria and other sub-cellular organelles. *Curr. Pharm. Des.* 20 (2), 201–222. doi:10.2174/13816128113199990031

Villa, E., Proïcs, E., Rubio-Patiño, C., Obba, S., Zunino, B., Bossowski, J., et al. (2017). Parkin-independent mitophagy controls chemotherapeutic response in cancer cells. *Cell Rep.* 20 (12), 2846–2859. doi:10.1016/j.celrep. 2017.08.087

Warth, A., Muley, T., Meister, M., Stenzinger, A., Thomas, M., Schirmacher, P., et al. (2012). The novel histologic International Association for the Study of Lung Cancer/American Thoracic Society/European Respiratory Society classification system of lung adenocarcinoma is a stage-independent predictor of survival. J. Clin. Oncol. 30 (13), 1438–1446. doi:10.1200/JCO.2011.37.2185

Wilkerson, M., and Hayes, D. (2010). ConsensusClusterPlus: a class discovery tool with confidence assessments and item tracking. *Bioinformatics* 26 (12), 1572–1573. doi:10.1093/bioinformatics/btq170

Xia, C., Dong, X., Li, H., Cao, M., Sun, D., He, S., et al. (2022). Cancer statistics in China and United States, 2022: profiles, trends, and determinants. *Chin. Med. J-Peking* 135 (5), 584–590. doi:10.1097/CM9.00000000002108

Xie, X., Yang, Y., Wang, Q., Liu, H., Fang, X., Li, C., et al. (2023). Targeting ATAD3A-PINK1-mitophagy axis overcomes chemoimmunotherapy resistance by redirecting PD-L1 to mitochondria. *Cell Res.* 33 (3), 215–228. doi:10.1038/s41422-022-00766-z

Xie, Y., Liu, J., Kang, R., and Tang, D. (2020). Mitophagy receptors in tumor biology. *Front. Cell Dev. Biol.* 8, 594203. doi:10.3389/fcell.2020.594203 Zhang, P., Dong, S., Sun, W., Zhong, W., Xiong, J., Gong, X., et al. (2023b). Deciphering Treg cell roles in esophageal squamous cell carcinoma: a comprehensive prognostic and immunotherapeutic analysis. *Front. Mol. Biosci.* 10, 1277530. doi:10.3389/fmolb.2023.1277530

Zhang, P., Pei, S., Zhou, G., Zhang, M., Zhang, L., and Zhang, Z. (2024). Purine metabolism in lung adenocarcinoma: a single-cell analysis revealing prognostic and immunotherapeutic insights. *J. Cell Mol. Med.* 28, e18284. doi:10.1111/jcmm.18284

Zhang, P., Zhang, H., Tang, J., Ren, Q., Zhang, J., Chi, H., et al. (2023a). The integrated single-cell analysis developed an immunogenic cell death signature to predict lung adenocarcinoma prognosis and immunotherapy. *Aging (Albany NY)* 15, 10305–10329. doi:10.18632/aging.205077

Zhang, T., Nie, Y., Gu, J., Cai, K., Chen, X., Li, H., et al. (2021). Identification of mitochondrial-related prognostic biomarkers associated with primary bile acid biosynthesis and tumor microenvironment of hepatocellular carcinoma. *Front. Oncol.* 11, 587479. doi:10.3389/fonc.2021.587479

Zhao, J., Zhang, J., Yu, M., Xie, Y., Huang, Y., Wolff, D., et al. (2013b). Mitochondrial dynamics regulates migration and invasion of breast cancer cells. *Oncogene* 32 (40), 4814–4824. doi:10.1038/onc.2012.494

Zhao, Y., Butler, E., and Tan, M. (2013a). Targeting cellular metabolism to improve cancer therapeutics. *Cell Death Dis.* 4 (3), e532. doi:10.1038/cddis.2013.60

Zhuo, Z., Lin, H., Liang, J., Ma, P., Li, J., Huang, L., et al. (2021). Mitophagyrelated gene signature for prediction prognosis, immune scenery, mutation, and chemotherapy response in pancreatic cancer. *Front. Cell Dev. Biol.* 9, 802528. doi:10.3389/fcell.2021.802528