

## Elucidating information processing in primate basal ganglia circuitry: a novel technique for pathway-selective ablation mediated by immunotoxin

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Employing a neuron-specific retrograde gene-transfer vector (NeuRet vector), we have recently developed a novel technique that achieves pathway-selective ablation in the primate brain. This technique is mediated by immunotoxin (IT) and eliminates a neuronal population that constitutes a particular pathway, leaving other pathways intact. By means of this technique, we have made an attempt to remove the hyperdirect pathway selectively from basal ganglia circuitry. The hyperdirect pathway links the motor cortex to the subthalamic nucleus (STN) directly and plays a crucial role in motor control. After electrical stimulation in the motor cortex, triphasic responses consisting of an early excitation, an inhibition, and a late excitation are usually elicited in the internal pallidal segment (GPi). Several pieces of pharmacophysiological evidence imply that the early excitation may be derived from the hyperdirect pathway. In our experiments, the NeuRet vector expressing human interleukin-2 receptor a-subunit was injected into the STN of macaque monkeys. Then, IT injections were performed into the supplementary motor area (SMA). When single neuron activity in the GPi was recorded in response to the SMA stimulation, it was found that the early excitation was significantly reduced with neither the inhibition nor the late excitation affected. The spontaneous firing rate and pattern of GPi neurons remained to be altered. This clearly indicates that IT-mediated tract targeting successfully eliminated the hyperdirect pathway with spontaneous activity of STN neurons unaffected. The electrophysiological findings were histologically confirmed by retrograde and anterograde neuronal labeling. The overall data define that the motor cortically driven early excitation in GPi neurons is conveyed through the hyperdirect pathway. The ITmediated pathway-selective ablation technique will provide a powerful tool for elucidating information processing in various neural networks.

Keywords: basal ganglia, hyperdirect pathway, information processing, immunotoxin, lentivirus, vectors, gene transfer, primates

## INTRODUCTION

To know about a variety of higher brain functions systematically, it is essential to elucidate the architecture of complex and elaborate neural networks. For clarifying the functional role of a given pathway, it is an effective way to explore behavioral and physiological changes due to ablation of a neuronal population that constitutes the target pathway. Neuronal targeting mediated by immunotoxin (IT) has been established in mice as a genetic method for eliminating a specific neuron group from a certain neural network (Kobayashi et al., 1995; Sano et al., 2003; Yasoshima et al., 2005). Recently, it has been revealed that the use of modified glycoprotein of rabies virus for preparing a pseudotyped lentiviral vector based on human immunodeficiency virus type 1 (HIV-1) can enhance the efficiency of gene transfer through retrograde transport of the vector (Kato et al., 2007, 2011a). This property of the pseudotyped lentiviral vector largely allows for gene transfer into cell bodies of neurons that are located remote from the injection site of the vector. For IT-mediated removal of a particular pathway, the highly efficient retrograde gene-transfer vector was produced to express human interleukin-2 receptor  $\alpha$ -subunit (IL-2R $\alpha$ ), a receptor molecule for the recombinant IT, in neuronal cell bodies via retrograde transport of the vector. In mice receiving injection of the IL-2R $\alpha$ -expressing vector into the striatum, IT injection into the thalamus successfully resulted in selective removal of the thalamostriatal pathway (Kato et al., 2011b).

In our recent work, we have applied the IT-mediated pathwayselective elimination technique to the primate brain, because the use of non-human primates as animal models is critical for investigating higher brain functions. Employing the nigrostriatal dopamine pathway as a test system, we have first established the basic methodology with a neuron-specific retrograde gene-transfer vector (NeuRet vector) that has newly been developed with improved neuron specificity (Kato et al., 2011c). Next, an attempt has been made to eliminate the cortico-subthalamic "hyperdirect" pathway selectively from basal ganglia circuitry in macaque monkeys (Inoue et al., 2012). The subthalamic nucleus (STN) receives major input from the motor cortex and, in turn, sends output to the internal segment of the globus pallidus (GPi), a main output station of the basal ganglia (Hartmann-von Monakow et al., 1978; Mink and Thach, 1993; Parent and Hazrati, 1995; Mink, 1996; Nambu et al., 1996, 1997, 2002a). It has been demonstrated that electrical stimulation in the motor cortex induces an early, short-latency excitation in GPi neurons, followed by an inhibition and then a late, long-latency excitation (Figure 1; Nambu et al., 2000, 2002a; Tachibana et al., 2008). According to several pharmacophysiological data, the early excitation is most likely to be conveyed through the cortico-STN-GPi pathway (Figure 1; Nambu et al., 2000, 2002a; Tachibana et al., 2008). However, no direct evidence has as yet been available. By means of IT-mediated pathway-selective ablation, we have successfully proven the contribution of the hyperdirect pathway to the emergence of the early excitation. Here we introduce the detailed data on this issue.



**FIGURE 1 | (A)** Peri-stimulus time histogram (PSTH; bin width of 1 ms, summed for 100 stimulus trials) showing triphasic responses of a neuron in the internal segment of the globus pallidus (GPi). Electrical stimulation in the motor cortex was given at time = 0 (arrow). Note that an early excitation (red), an inhibition (blue), and a late excitation (green) are derived respectively from the hyperdirect, direct, or indirect pathway depicted in **(B)**. **(B)** Three pathways of the basal ganglia. Cx, cerebral cortex; glu, glutamate; GPe, external segment of the globus pallidus; GNr, substantia nigra pars reticulata; STN, subthalamic nucleus; Str, striatum; Th, thalamus.

## **PREPARATION OF NeuRet VECTOR**

We have developed a new vector system that permits NeuRet by pseudotyping the HIV-1-based lentiviral vector with fusion glycoprotein C type (FuG-C) consisting of a hybrid of rabies virus glycoprotein (RV-G) and vesicular stomatitis virus glycoprotein (VSV-G; Kato et al., 2011c). Interestingly, the NeuRet vector exhibits high efficiency of retrograde gene transfer into various populations of neurons, while it markedly reduces gene transduction into dividing cells, including glial and neural stem/progenitor cells, around the vector injection site. The NeuRet vector is composed of the N-terminal segment of the extracellular domain (439 amino acids) of RV-G and the C-terminal segment





of the extracellular domain (16 amino acids) and transmembrane/cytoplasmic domains of VSV-G (Figure 2A). To verify the capability of the NeuRet vector for efficient retrograde gene transfer into the nigrostriatal pathway, we injected the vector encoding the green fluorescent protein (GFP) transgene into the striatum (caudate nucleus and putamen) of crab-eating monkeys. Intrastriatal injection of the NeuRet vector produced a large number of GFP-positive neurons in the nigra (Figure 2B). These neurons were immunostained for tyrosine hydroxylase, a key enzyme for dopamine biosynthesis (Figure 2C), indicating the transgene expression in the nigrostriatal dopamine neurons. Moreover, we assessed the extent of gene transfer with the NeuRet vector around the injection sites in the monkey striatum. The vector displayed a low level of gene transfer into neuronal cells, and the level of vector transfer into glial cells was also quite low in the striatum (Figures 2D,E). Therefore, the NeuRet vector mediates enhanced retrograde gene transfer into neuronal cells, whereas it reduces the efficiency of gene transfer into glial cells around the injection sites.

## **SELECTIVE ABLATION OF HYPERDIRECT PATHWAY**

The NeuRet vector expressing IL-2Ra was injected into electrophysiologically identified sites in the STN of Japanese monkeys. Following the vector injections into the STN, activity

of GPi neurons was recorded in response to electrical stimulation in the supplementary motor area (SMA). In most of the GPi neurons in which certain responses were induced, we observed a triphasic response pattern consisting of an early excitation, a subsequent inhibition, and a late excitation, as previously reported in normal monkeys (see **Figure 1**; Nambu et al., 2000; Tachibana et al., 2008). This indicated that the vector injections into the STN did not affect cortically evoked responses of GPi neurons.

After IT injections into the SMA, especially its arm region, neuronal activity in the GPi was recorded in response to the SMA stimulation. Many of the recorded GPi neurons exhibited a biphasic pattern, an inhibition followed by a late excitation without an early excitation (Figure 3). Compared with a control condition (before the IT injections), the amplitude of the early excitation was largely (by almost 90% of the control) reduced after the IT injections. On the other hand, the amplitude of the inhibition and the late excitation remained relatively unchanged, although the late excitation slightly decreased with no significant change (Figure 3). In addition, virtually no alterations were found in the latency of the inhibition or the late excitation, or the duration of the inhibition or the late excitation. In the control monkey in which no vector injections were made into the STN, the response pattern of GPi neurons on SMA stimulation was essentially the same before and after IT injections into the SMA (Figure 4). Thus,



the IT injections into the SMA combined with the injections of the NeuRet vector expressing IL-2R $\alpha$  into the STN abolished the cortically evoked early excitation in the GPi without affecting either the inhibition or the late excitation.

Furthermore, the spontaneous firing rate and pattern were compared before and after the IT injections into the SMA. The spontaneous firing rate was left intact (**Figure 5**). Neurons in the GPi fired randomly at high frequency before the IT injections, and no apparent changes were observed after the IT injections (**Figure 5**). These results suggested that the firing rate and pattern of GPi neurons remained unchanged even after the removal of the cortico-STN projection.

In the monkeys subjected to the disappearance of the early excitation responding to the SMA stimulation, we performed retrograde and anterograde neuronal labeling by injecting Fluororuby (FR) into the STN and biotinylated dextran amine (BDA) into the SMA. After the FR injection into the STN, retrogradely labeled neurons in the SMA were much fewer in the arm region where the IT injections were primarily aimed than in the face and leg regions (**Figure 6**). Moreover, immunostaining for NeuN revealed that the IT injections into the SMA caused no marked tissue damage (**Figure 6**). After the BDA injections into the SMA forelimb region, anterogradely labeled axon terminals were so largely decreased in the STN, as compared to the control case (data not shown). In remarkable contrast, dense terminal labeling from the SMA was seen in the striatum, especially the putamen, as in the control case (data not shown). These anatomical data clearly indicated that cortico-STN projection originating from the SMA arm region was selectively eliminated without affecting either the corticostriatal projection or the cortico-STN projections from SMA regions with other representations than the arm.

We did not closely examine behavioral changes to be produced after elimination of the hyperdirect pathway, because our study was primarily designed to develop a new methodological approach to selective removal of a given pathway. As far as our experimental conditions were concerned, no apparent motor abnormalities were observed.

## **DISCUSSION**

Taking advantage of the NeuRet vector that allows for highly efficient retrograde gene-transfer with improved neuron specificity (Kato et al., 2011c), we have established IT-mediated pathwayselective ablation in the primate brain. Here we have applied this technique to the hyperdirect pathway (Inoue et al., 2012). In conjunction with the direct and indirect pathways (for reviews, see Albin et al., 1989; Alexander and Crutcher, 1990), the hyperdirect pathway is known to be among the key pathways of the basal ganglia, being involved in motor information processing in the basal



ganglia (Nambu et al., 1996, 2002a). This pathway connects the motor cortex to the GPi at short latency through the STN without relay at the striatum. When single neuron activity was recorded in the monkey GPi in response to electrical stimulation in the motor cortex, triphasic responses composed of an early (short-latency) excitation, an inhibition, and a late (long-latency) excitation were obtained. Based on the following pharmacophysiological data (see also Nambu et al., 2002a), it has been considered that the early excitation may be derived from the cortico-STN-GPi hyperdirect pathway: (1) Blockade of STN neuron activity by injection of the GABA<sub>A</sub> receptor agonist, muscimol, there into abolished the early as well as the late excitation of GPi neurons (Nambu et al., 2000); (2) Blockade of glutamatergic input from the STN to the GPi by local injection of an ionotropic glutamatergic receptor antagonist diminished the early as well as the late excitation of GPi neurons (Tachibana et al., 2008).

For selective removal of the hyperdirect pathway, the NeuRet vector expressing IL-2R $\alpha$  was injected into the STN, and, subsequently, IT was injected into the SMA in our experimental protocol. Our histological examination clearly indicated that cortical neurons in the arm region of the SMA projecting to the STN were selectively ablated. In such model monkeys, GPi neuron activity was recorded in response to electrical stimulation in the SMA. The SMA stimulation yields selective activation of SMArecipient zones in the basal ganglia (Nambu et al., 2002b). It was found that out of the triphasic responses, only the early excitation was largely suppressed without either the inhibition or the late excitation affected. This indicates that IT-mediated tract targeting successfully eliminated the hyperdirect pathway selectively from basal ganglia circuitry. Our results define that the cortically driven early excitation of GPi neurons is derived from the cortico-STN projection. It has also been revealed that the firing rate and pattern of GPi neurons remain unchanged even after the removal of the cortico-STN projection. This implies that the cortico-STN projection conveys phasic activity changes from the SMA to the GPi, but does not contribute to maintenance of tonic activity of GPi neurons. In contrast to the early excitation, the inhibition in the GPi was not affected by the elimination of the cortico-STN projection, as it can be considered that the inhibition is mediated through the cortico-striato-GPi direct pathway (Tachibana et al., 2008). On the other hand, the late excitation in the GPi was slightly diminished though not significant. This late excitation is ascribable to the late excitation in the STN and is probably mediated by the cortico-striato-external pallidal segment (GPe)-STN-GPi indirect pathway. However, it has also been suggested that the late excitation in the STN is part of the prolonged excitation induced by the cortico-STN projection, which may explain a slight decrease in the late excitation in the GPi after the elimination of the cortico-STN projection (Tachibana et al., 2008). The SMA neurons giving rise to the cortico-STN projection are likely to issue axon collaterals to cortical and/or subcortical (other than the STN) regions. Thus, it cannot be ruled out that no such possible collateral projections may be affected by IT injected into the SMA.

According to the cortically driven triphasic response pattern elicited in GPi neurons, the hyperdirect pathway conveys excitatory signals from the motor cortex toward the GPi, bypassing the striatum, with shorter conduction time than signals via the striatum that arise from both the direct and the indirect pathways (see



Nambu et al., 2000, 2002a). In favor of a dynamic "center-surround model" of basal ganglia function that was first proposed by Mink and Thach (1993), the functional role of the hyperdirect pathway has been implicated in the control of voluntary limb movements (see also Mink, 1996; Hikosaka et al., 2000; Nambu et al., 2002a). When a voluntary movement is about to be initiated by the motor cortical mechanism, a corollary signal conveyed through the cortico-STN-GPi hyperdirect pathway first inhibits large areas of the thalamic and cortical target structures that are related not only to a desired motor program, but also to other competing programs. Then, another corollary signal through the cortico-striato-GPi direct pathway disinhibits part of the thalamic and cortical target areas and releases the desired motor program alone. Finally, the third corollary signal conveyed by way of the cortico-striato-GPe-STN-GPi indirect pathway again inhibits the thalamic and cortical target areas extensively. By virtue of such sequential motor information processing, only the desired motor program is initiated, executed, and terminated at appropriate timings, whereas other competing programs are canceled. Thus, it is most likely that the hyperdirect pathway exerts a powerful excitatory effect on the GPi to suppress involuntary and unnecessary movements prior to the selected motor action. This notion is substantiated by the following issues: (1) Lesions or blockade of STN neuron activity induced involuntary movements, hemiballism (Carpenter et al., 1950; Hamada and DeLong, 1992; Nambu et al., 2000), suggesting that both the hyperdirect and indirect pathways might inhibit unnecessary movements; (2) According to functional magnetic resonance imaging studies using human subjects, the cortico-STN projection conveys stop signals to inhibit motor responses (Aron and Poldrack, 2006; Jahfari et al., 2011); (3) It is also suggested that the cortico-STN projection may inhibit automatic movements and switch to volitionally controlled movement (Isoda and Hikosaka, 2008).

Since no explicit motor abnormalities were seen in our experimental conditions, changes in the activity of GPi neurons in response to the cortical stimulation following a limited amount of elimination of the hyperdirect pathway may not be enough to elicit behavioral alterations. Accordingly, there is a need to determine the relationship between the extent of the elimination of the selected pathway and the expression of altered behaviors.

The IT-mediated tract targeting achieves selective ablation of a given pathway in primates. This novel technique will provide a potent strategy to explore not only specific functional roles

### **REFERENCES**

- Albin, R. L., Young, A. B., and Penney, J. B. (1989). The functional anatomy of basal ganglia disorders. *Trends Neurosci.* 12, 366–375. doi: 10.1016/0166-2236(89)90074-X
- Alexander, G. E., and Crutcher, M. D. (1990). Functional architecture of basal ganglia circuits: neural substrates of parallel processing. *Trends Neurosci.* 13, 266–271. doi: 10.1016/0166-2236(90)90107-L
- Aron, A. R., and Poldrack, R. A. (2006). Cortical and subcortical contributions to stop signal response inhibition: role of the subthalamic nucleus. *J. Neurosci.* 26, 2424– 2433. doi: 10.1523/JNEUROSCI. 4682-05.2006
- Carpenter, M. B., Whittier, J. R., and Mettler, F. A. (1950). Analysis of choreoid hyperkinesia in the rhesus monkey; surgical and pharmacological analysis of hyperkinesia resulting from lesions in the subthalamic nucleus of Luys. J. Comp. Neurol. 92, 293–331. doi: 10.1002/cne.900920303
- Hamada, I., and DeLong, M. R. (1992). Excitotoxic acid lesions of the primate subthalamic nucleus result in transient dyskinesias of the contralateral limbs. J. Neurophysiol. 68, 1850– 1858.
- Hartmann-von Monakow, K., Akert, K., and Künzle, H. (1978). Projections of the precentral motor cortex and other cortical areas of the frontal lobe to the subthalamic nucleus in the monkey. *Exp. Brain Res.* 33, 395–403. doi: 10.1007/BF00235561
- Hikosaka, O., Takikawa, Y., and Kawagoe, R. (2000). Role of the basal ganglia in the control of purposive saccadic eye movements. *Physiol. Rev.* 80, 953–978.
- Inoue, K., Koketsu, D., Kato, S., Kobayashi, K., Nambu, A., and Takada, M. (2012). Immunotoxinmediated tract targeting in the primate brain: selective elimination of the cortico–subthalamic "hyperdirect" pathway. *PLoS ONE* 7:e39149. doi: 10.1371/journal.pone.0039149
- Isoda, M., and Hikosaka, O. (2008). Role for subthalamic nucleus neurons in switching

from automatic to controlled eye movement. J. Neurosci. 28, 7209– 7218. doi: 10.1523/JNEUROSCI. 0487-08.2008

- Jahfari, S., Waldorp, L., van den Wildenberg, W. P., Scholte, H. S., Ridderinkhof, K. R., and Forstmann, B. U. (2011). Effective connectivity reveals important roles for both the hyperdirect (frontosubthalamic) and the indirect (fronto-striatal-pallidal) fronto-basal ganglia pathways during response inhibition. J. Neurosci. 31, 6891– 6899. doi: 10.1523/JNEUROSCI. 5253-10.2011
- Kato, S., Inoue, K., Kobayashi, K., Yasoshima, Y., Miyachi, S., Inoue, S., et al. (2007). Efficient gene transfer via retrograde transport in rodent and primate brains by an HIV-1-based vector pseudotyped with rabies virus glycoprotein. *Hum. Gene Ther.* 18, 1141–1151. doi: 10.1089/hum.2007.082
- Kato, S., Kobayashi, K., Inoue, K., Kuramochi, M., Okada, T., Yaginuma, H., et al. (2011a). A lentiviral strategy for highly efficient retrograde gene transfer by pseudotyping with fusion envelope glycoprotein. *Hum. Gene Ther.* 22, 197–206. doi: 10.1089/hum.2009.179
- Kato, S., Kuramochi, M., Kobayashi, K., Fukabori, R., Okada, K., Uchigashima, M., et al. (2011b). Selective neural pathway targeting reveals key roles of thalamostriatal projection in the control of visual discrimination. J. Neurosci. 31, 17169– 17179. doi: 10.1523/JNEUROSCI. 4005-11.2011
- Kato, S., Kuramochi, M., Takasumi, K., Kobayashi, K., Inoue, K., Takahara, D., et al. (2011c). Neuronspecific gene transfer through retrograde transport of lentiviral vector pseudotyped with a novel type of fusion envelope glycoprotein. *Hum. Gene Ther.* 22, 1511–1523. doi: 10.1089/hum.2011.111
- Kobayashi, K., Morita, S., Sawada, H., Mizuguchi, T., Yamada, K., Nagatsu, I., et al. (1995). Immunotoxinmediated conditional disruption of specific neurons in transgenic mice. *Proc. Natl. Acad.*

of individual pathways constituting a particular neural network, but also large-scale operative mechanisms underlying the entire network.

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*Sci. U.S.A.* 92, 1132–1136. doi: 10.1073/pnas.92.4.1132

- Mink, J. W. (1996). The basal ganglia: focused selection and inhibition of competing motor programs. *Prog. Neurobiol.* 50, 381–425. doi: 10.1016/S0301-0082(96)00042-1
- Mink, J. W., and Thach, W. T. (1993). Basal ganglia intrinsic circuits and their role in behavior. *Curr. Opin. Neurobiol.* 3, 950–957. doi: 10.1016/0959-4388(93)90167-W
- Nambu, A., Takada, M., Inase, M., and Tokuno, H. (1996). Dual somatotopical representations in the primate subthalamic nucleus: evidence for ordered but reversed body-map transformations from the primary motor cortex and the supplementary motor area. *J. Neurosci.* 16, 2671– 2683.
- Nambu, A., Tokuno, H., Hamada, I., Kita, H., Imanishi, M., Akazawa, T., et al. (2000). Excitatory cortical inputs to pallidal neurons via the subthalamic nucleus in the monkey. J. Neurophysiol. 84, 289–300.
- Nambu, A., Tokuno, H., Inase, M., and Takada, M. (1997). Corticosubthalamic input zones from forelimb representations of the dorsal and ventral divisions of the premotor cortex in the macaque monkey: comparison with the input zones from the primary motor area. *Neurosci. Lett.* 239, 13–16. doi: 10.1016/S0304-3940(97)00877-X
- Nambu, A., Tokuno, H., and Takada, M. (2002a). Functional significance of the cortico–subthalamo–pallidal 'hyperdirect' pathway. *Neurosci. Res.* 43, 111–117. doi: 10.1016/S0168-0102(02)00027-5
- Nambu, A., Kaneda, K., Tokuno, H., and Takada, M. (2002b). Organization of corticostriatal motor inputs in monkey putamen. *J. Neurophysiol.* 88, 1830–1842.
- Parent, A., and Hazrati, L.-N. (1995). Functional anatomy of the basal ganglia. II. The place of subthalamic nucleus and external pallidum in basal ganglia circuitry. *Brain Res. Brain Res. Rev.* 20, 128– 154. doi: 10.1016/0165-0173(94) 00008-D

- Sano, H., Yasoshima, Y., Matsushita, N., Kaneko, T., Kohno, K., Pastan, I., et al. (2003). Conditional ablation of striatal neuronal types containing dopamine D2 receptor disturbs coordination of basal ganglia function. *J. Neurosci.* 23, 9078–9088.
- Tachibana, Y., Kita, H., Chiken, S., Takada, M., and Nambu, A. (2008).
  Motor cortical control of internal pallidal activity through glutamatergic and GABAergic inputs in awake monkeys. *Eur. J. Neurosci.* 27, 238– 253. doi: 10.1111/j.1460-9568.2007. 05990.x
- Yasoshima, Y., Kai, N., Yoshida, S., Shiosaka, S., Koyama, Y., Kayama, Y., et al. (2005). Subthalamic neurons coordinate basal ganglia function through differential neural pathways. *J. Neurosci.* 25, 7743–7753. doi: 10.1523/JNEUROSCI.1904-05. 2005

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