# Serum-based protein biomarkers in blast-induced traumatic brain injury spectrum disorder

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Denes V. Agoston, Department of Anatomy, Physiology and Genetics, School of Medicine, Uniformed Services University, 4301 Jones Bridge Road, Bethesda, MD 20814, USA. e-mail: vagoston@usuhs.edu; denes.agoston@ki.se The biological consequences of exposure to explosive blast are extremely complex. Serum protein biomarkers in blast-induced traumatic brain injury (bTBI) can aid in determining injury severity, monitoring progress, and predicting outcome. Exposure to blast results in varying degrees of physical injury. Explosive blast can also induce psychological stress that can contribute to or amplify the extent of physical damage. Given the complexity, scale of injury, and variety of symptoms, bTBI may be best described as a spectrum disorder. In this focused review, we summarize the status of serum protein biomarkers in bTBI in the context of the classification and pathological changes of other forms of TBI. Finally, we recommend specific and easily implementable measures to accelerate serum protein biomarker discovery and validation in bTBI.

Keywords: blood, proteomics, traumatic, brain, injury, blast, biomarkers, serum

Serum protein biomarkers have long held promise in the treatment of traumatic brain injury (TBI). They can aid in diagnosing the disease, monitoring progress, predicting outcome, and providing pertinent molecular information about ongoing pathological changes for designing evidence-based therapeutic interventions (Kochanek et al., 2011). Serum protein biomarkers are of special importance in blast-induced TBI (bTBI) because they are typically associated with military operations with limited access to imaging and other diagnostic tools of hospitals (Agoston et al., 2009).

The physical and biological consequences of explosive blast are extremely complex. Blast generates high energy supersonic pressure waves, heat, toxic gases, electromagnetic pulses, etc. (Champion et al., 2009;Ramasamy et al., 2009a,b; Hicks et al., 2010; Nakagawa et al., 2011). How each of these forces, separately or in a combinatorial fashion interact with the brain and body is still poorly understood. While the cause of bTBI is exposure to blast, injury severity may range from mild to severe and result in outcomes that cover a wide set of symptoms (Mayorga, 1997; Guy et al., 2000; Elder and Cristian, 2009; Elder et al., 2010; Rosenfeld and Ford, 2010; Marion et al., 2011). Exposure to blast can also cause severe psychological stress that can contribute to or amplify the extent of physical damage (Kluger et al., 2004; Ling et al., 2009; Wallace, 2009; Wolf et al., 2009; Ling and Ecklund, 2011). Accordingly, bTBI may be best described as a spectrum disorder. Similar to other forms of TBI, the classification of bTBI is currently based on subjective neurobehavioral evaluations including the Glasgow Coma Scale (GCS) and the Military Acute Concussion Evaluation (MACE; Cernak et al., 1999; Secer et al., 2007; Bochicchio et al., 2008; Peleg and Savitsky, 2009; Rosenfeld and Ford, 2010; Tarmey et al., 2011). These functional assessments are only occasionally supplemented with information from neuroimaging techniques, such as computed tomography (CT) and magnetic resonance imaging (MRI; Ling and Ecklund, 2011).

In this paper, we provide a brief overview of the status of serum protein biomarkers in bTBI. Because of the limited information about protein biomarkers specific to blast injury, we will discuss them in the context of the classification and pathological changes of other forms of TBI.

## **EPIDEMIOLOGY OF TBI AND bTBI**

Traumatic brain injury is an enormous public health concern. The Centers for Disease Control and Prevention (CDC) estimates that  $\sim$ 1.7 million Americans sustain TBI every year (Coronado et al., 2011). TBI also contributes to about a third of all injury-related deaths, resulting in over 52,000 deaths a year (Coronado et al., 2011).

Improving TBI treatment for military personnel is especially pressing. The incidence of TBI for armed forces, even during peacetime, is greater than civilian populations (Ommaya et al., 1996). Also, due in part to the nature of modern combat, the

**Abbreviations:** AQP4, aquaporin 4; C-tau, cleaved-tau protein; GFAP, glial fibrillary acidic protein; HIF-1 $\alpha$ , hypoxia-inducible factor 1, alpha subunit; Hsp70, heat shock protein 70; IL-1 $\beta$ , interleukin-1 beta; IL-6, interleukin-6; IL-8, interleukin-8; MBP, myelin basic protein; MMP9, matrix metalloproteinase-9; NF-H, neurofilament-heavy chain; NSE, neuron specific enolase; pNF-H, phosphorylated neurofilament-heavy chain; S100 $\beta$ , S100 calcium binding protein B; SBPs, spectrin breakdown products; TNF- $\alpha$ , tumor necrosis factor alpha; UCH-L1, ubiquitin C-terminal hydrolase L1; VEGF, vascular endothelial growth factor; vWF, von Willebrand factor.

incidence rates of TBI have been increasing since 2000 (Sayer, 2012). Although it is difficult to assess exactly how many soldiers are victims of TBI (especially because many are not properly diagnosed), estimates from Veterans Affairs medical records report that about 7% of veterans from Iraq and Afghanistan received a TBI diagnosis (Taylor et al., 2012). Other survey studies suggest that 11–23% of military personnel deployed to Iraq or Afghanistan may have sustained at least mild TBI (Sayer, 2012). Studies from the Rand Corporation estimate that about of fifth of returning members suffer from TBI (Tenielian and Jaycox, 2008).

Blast injury from improvised explosive devices (IEDs) is an especially common form of TBI among military populations. According to the Joint Theater Trauma Registry, IEDs were the source of about 80% of all casualties of veterans from Iraq and Afghanistan between October 2001 and January 2005 (Owens et al., 2008). In another review of US Army casualties in Afghanistan and Iraq between 2001 and 2007, explosions were linked with 63% of all TBI diagnoses (Wojcik et al., 2010). Another similar survey of US Navy and Marine casualties in Iraq in 2004 found that 52% of all TBI cases involved explosions (Galarneau et al., 2008). Despite the variability of these studies, it is evident that TBI and bTBI especially affects a great deal of soldiers, thus requiring the need for improved diagnostics and treatments.

# CLASSIFICATION OF BLAST-INDUCED TRAUMATIC BRAIN INJURY

Individuals who experience less than 30 minutes of lost or altered levels of consciousness after exposure to blast are classified as having suffered mild bTBI (mibTBI; **Figure 1**; Trudeau et al., 1998; Hoge et al., 2008; Elder et al., 2010; Levin et al., 2010; Rosenfeld and Ford, 2010; Wilk et al., 2010). This initial period may be followed by post-injury amnesia that lasts no longer than 24 hours. There are typically no penetrating injuries to the head or other

organs, and neurological deficits (if any) are focal and transient in nature. Concurrently, the GCS score is nearly perfect at 13–15. The transient and mild neuropsychiatric deficits are typically followed by full recovery. However, similar to other types of mild TBI, a delayed onset of functional changes and long-term disabilities may still occur, especially after multiple exposures to mild blasts (Stern et al., 2011). Importantly, mibTBI shares symptoms and is often comorbid with post-traumatic stress disorder (PTSD; Hoge et al., 2008; Levin et al., 2010; Rosenfeld and Ford, 2010; Ursano et al., 2010). The combination of physical damage and psychological effects makes mibTBI especially difficult to diagnose. Thus, serum protein biomarkers that can distinguish between the physical and psychological components of the injury would be of special value (see also **Figures 2** and **3** and discussion below).

Blast-induced traumatic brain injury is classified as moderate if loss of consciousness is longer than 30 minutes, post-injury amnesia lasts longer than 24 hours, and the initial GCS score is between 9 and 12 (**Figure 1**; Thompson et al., 2008; Aarabi and Simard, 2009; Wolf et al., 2009; Drake et al., 2010). The long-term outcome of moderate bTBI (mobTBI) can include detectable and significant levels of cognitive and neuropsychiatric abnormalities. Moderate bTBI can also be comorbid with PTSD and injuries to other organs.

According to the current classification system, bTBI is severe if the GCS score is less than 9 (**Figure 1**; Ling and Marshall, 2008; Ling et al., 2009; Ling and Ecklund, 2011). In the severe form of bTBI (sbTBI), polytrauma, i.e., injuries to other parts of the body, most frequently to the extremities, abdomen, and lungs can significantly contribute to and modify the pathology and outcome of brain injury. In addition to its comorbidity with polytrauma, sbTBI is often the result of multiple types of brain injury. Beyond the damage caused by the primary components of blast, various objects, debris, and shrapnel may break through the



Biomarker	Cell Specificity	Origin/	Function	bTBI Experimental	bTBI Clinical	TBI Experimental	TBI Clinical
biomarker	cen specificity	Localization	- runcuon	brorexperimental	Biblicinical	rbicxperimental	rbi clinicar
5100 β	Glia, astrocytes, muscle	Cytoplasm, also secreted extra- cellularly	Intracellular signaling, Ca2+ binding, regulating PKC phosphorylation	Yes <sup>1</sup>		Yes <sup>3, 4</sup>	Yes <sup>5-11</sup>
NSE	Neurons	Neuronal cyto- plasm	Glycolytic enzyme, regulation of intracellular chloride level	Yes <sup>2, 14, 15</sup>		Yes <sup>3, 16, 17,</sup>	Yes <sup>5, 8, 9, 11, 18</sup>
GFAP	Astroglia	Cytoskeleton	Structural filament	Yes 1, 2, 15, 50		Yes 17	Yes 11, 20 - 22
MBP	Oligodendrocytes	White matter, axons	Involved in myelination	Yes <sup>1</sup>		Yes 23	Yes <sup>2, 24</sup>
pNF -H	Neurons, neuroendo- crine tissue	Cytoskeleton, axons	Structural filament			Yes 25, 26	Yes 11, 27, 28, 51
NF -H	Neurons, neuroendo- crine tissue	Cytoskeleton, axons	Structural filament	Yes <sup>1, 15</sup>			Yes 7
Hsp70	Ubiquitous	Nucleus, cyto- plasm	Transcribed upon cellular stress to protect cell				Yes 11, 29
Secretagogin	Neurons, neuroendo- crine tissue	Nucleus, cytosol	Ca2+ binding protein				Yes 11
IL -1β	Glia, immune cells	Serum, cell receptors	Released as part of inflammatory response			Yes 30	Yes 11, 31, 32
IL -6	Glia, immune cells	Serum, cell receptors	Released as part of inflammatory response				Yes 11, 31, 32
IL -8	Glia, immune cells	Serum, cell receptors	Released as part of inflammatory response		Yes 33		Yes 11, 31, 32
TNF -α	Activated macro- phages	Serum, cell receptors	Involved in inflammatory response, apoptosis		Yes 33	Yes <sup>34</sup>	Yes 31, 32
UCH -L1	Neurons, neuroendo- crine tissue	Cytoplasm	Ubiquitin hydrolysis	Yes <sup>2</sup>		Yes 35	Yes <sup>27, 36, 37</sup>
C-tau	Neurons, low level in astrocytes and oli- godendrocytes	Microtubules	Axoplasmic flow, microtubule for- mation			Yes 38, 39	Yes <sup>8, 11, 40</sup>
Alpha II -Spectrin	Neuron, axon	Cytoskeletal membrane	Structural component of neuronal/ axonal cytoskeleton			Yes 41	Yes 27
SBPs	Neuron, axon	Serum	Cleaved by calpain - and caspase - derived proteases			Yes 42-44	Yes 27
Ceruloplasmin	Ubiquitous	Serum	Copper -carrier protein involved in copper and iron metabolism				Yes 45,46
vWF	Mainly endothelial cells, other cell types	Serum, cell receptors	Glycoprotein involved in hemostasis	Yes 48			Yes 47
VEGF	Ubiquitous	Serum, cell receptors	Promotes angiogenesis	Yes 48		Yes 15	
Claudin -5	Mainly endothelial cells	Cell membrane, tight junctions	Involved in forming tight junctions, regulating paracellular transport	Yes <sup>48</sup>			
AQP4	Mainly astrocytes	Cell membrane	Water channel			Yes 49	
MMP9	Ubiquitous	Extracellular matrix	Involved in breaking down the extra- cellular matrix			Yes 49	
HIF-1α	Ubiquitous	Nucleus, Cyto- plasm	Induced by hypoxia, activates cell protection response			Yes 49	

#### FIGURE 2 | Candidate protein biomarkers for blood-based diagnostics

**in traumatic brain injury.** <sup>1</sup>Gyorgy et al. (2011), <sup>2</sup>Svetlov et al. (2010), <sup>3</sup>Hardemark et al. (1989), <sup>4</sup>Rothoerl et al. (2000), <sup>5</sup>Bellander et al. (2011), <sup>6</sup>Gonzclez-Mao et al. (2011), <sup>7</sup>Haqqani et al. (2007), <sup>8</sup>Begaz et al. (2006), <sup>9</sup>Pleines et al. (2001), <sup>10</sup>Townend et al. (2006), <sup>11</sup>Zurek and Fedora (2012), <sup>12</sup>Honda et al. (2010), <sup>13</sup>Berger et al. (2005), <sup>14</sup>Cheng et al. (2010), <sup>15</sup>Kwon et al. (2011), <sup>16</sup>Pineda et al. (2004), <sup>17</sup>Woertgen et al. (2002), <sup>18</sup>Graham et al. (2011), <sup>19</sup>Hergenroeder et al. (2008), <sup>20</sup>Papa et al. (2012), <sup>21</sup>Vos et al. (2010), <sup>22</sup>Pelinka et al. (2004), <sup>23</sup>Liu et al. (2006), <sup>24</sup>Berger (2006), <sup>25</sup>Petzold (2005),

skull causing penetrating TBI. Also, it is not uncommon for the victim to be physically thrown from the mechanical force of explosion, causing further injury (acceleration-deceleration TBI). The combined damage from blast, penetrating injury, and acceleration-deceleration types of forces cause severe brain damage that leads to complex and debilitating long-term neurological and neuropsychiatric deficits, if not death (see **Figures 2** and **3**; Discussion below; Ling and Marshall, 2008; Ling et al., 2009; Ling and Ecklund, 2011). With respect to biomarkers, measuring changes in specific serum proteins indicative of the extent of neuronal and glial cell loss, axonal, and vascular damage, and damage to other organs can significantly add to the current diagnostic palette of neurobehavioral tests (see also **Figures 2** and **3** and discussion below).

<sup>26</sup>Anderson et al. (2008), <sup>27</sup>Siman et al. (2009), <sup>28</sup>Sandler et al. (2010), <sup>29</sup>da
Rocha et al. (2005), <sup>30</sup>Kinoshita et al. (2002), <sup>31</sup>Stein et al. (2011),
<sup>32</sup>Hayakata et al. (2004), <sup>33</sup>Surbatovic et al. (2007), <sup>34</sup>Vitarbo et al. (2004),
<sup>35</sup>Liu et al. (2010), <sup>36</sup>Berger et al. (2012), <sup>37</sup>Papa et al. (2010), <sup>38</sup>Zemlan et al. (2002), <sup>39</sup>Gabbita et al. (2005), <sup>40</sup>Bulut et al. (2006), <sup>41</sup>Pike et al. (2001),
<sup>42</sup>Newcomb et al. (1997), <sup>43</sup>Ringger et al. (2004), <sup>44</sup>Saatman et al. (2010),
<sup>45</sup>Dash et al. (2010), <sup>46</sup>Young et al. (1988), <sup>47</sup>De Oliveira et al. (2007),
<sup>48</sup>Ahmed et al. (in preparation), <sup>49</sup>Higashida et al. (2011), <sup>50</sup>Garman et al. (2011), <sup>51</sup>Zurek et al. (2011), <sup>52</sup>Brophy et al. (2009).

## PATHOLOGICAL MECHANISMS AND ASSOCIATED SERUM PROTEIN BIOMARKERS IN bTBI

The initial interactions between the physical forces of blast and the brain trigger pathological responses called the primary injury process or mechanism. The pathological components of the primary injury mechanism are largely influenced by whether the insult results in open (penetrating) or closed head injury. Penetrating head injury not only causes substantial direct tissue damage, but also instantaneously breaks down existing biological barriers, generating massive pathological responses to toxic molecules and cellular debris. In contrast, closed head injury typically causes metabolic changes and axonal damage of various degrees. In response to the typically short-lasting primary injury mechanism, there is a second wave of long-lasting pathological changes

Events	Mechanism	Candidate Serum Biomarkers			
		Mild TBI	Moderate TBI	Severe TBI	
	Activation of factors triggering	Neuronal: NSE, C- tau, SBP, All- Spectrin	Neuronal: NSE, pNF-H, NF-H, NMDAR, Hsp70, UCH-L1, C-tau, All- spectrin, SBP, Secretagogin	Neuronal: NSE, pNF-H, NF-H NMDAR, Hsp70, UCH-L1, C- tau, All-spectrin, SBP, Secretagogin	
Gilai Celi Deali	necrosis and/or apoptosis	Glial: S100β , GFAP, MBP, C-tau	Glial: S100β, GFAP, MBP, NMDAR, Hsp70, IL-1β, IL-6, IL-8, TN- α, C-tau, AQP4	Glial: S100β, GFAP, MBP, NMDAR, Hsp70, IL-1β, IL-6, IL-8, TN- α, C -tau, AQP4	
Vasospasm	Dysregulation of vascular constriction and relaxation	-	Hsp70, TNF-α, VEGF, Claudin-5, vWF	Hsp70, TNF-α, VEGF, Claudin-5, vWF	
Edema	Vasogenic and cytotoxic events caused by toxic and inflammatory factors	-	Hsp70, IL-1β, IL -6, IL- 8, VEGF, Claudin-5, vWF, AQP4, MMP9	Hsp70, IL-1β, IL -6, IL-8, VEGF, Claudin-5, vWF, AQP4, MMP9	
Axonal Injury	Mechanical injury; Neuronal degeneration	S100β, NSE, C-tau, MBP, SBP, All- Spectrin	S100β, MBP, NSE, PNF-H, NMDAR, Hsp70, C-tau, All- spectrin, SBP	S100β, MBP, NSE, PNF-H, NMDAR, Hsp70, C-tau, All- spectrin, SBP	
Inflammation	Cytokine release and cellular stress	IL-1β, IL -6, IL-8, TNF-α, IFN -γ	Hsp70,IL-1β, IL-6, IL-8, TNF-α, IFN -γ	Hsp70,IL-1β, IL -6, IL-8, TNF- α, IFN -γ	
Metabolic Changes	Hypoxia; altered energy demand, ion homeostasis and neurotransmission; increased repair processes	-	Ceruloplasmin, HIF-1α	Ceruloplasmin, HIF-1a	

FIGURE 3 | Candidate serum protein biomarkers associated with injury severity and selected pathological mechanisms.

called the secondary injury mechanism. These pathologies include metabolic changes, neuroinflammation, axonal injury, vascular abnormalities, and neuronal and glial cell death (Ghirnikar et al., 1998; Lenzlinger et al., 2001; Vink et al., 2001; Morganti-Kossmann et al., 2002; Nortje and Menon, 2004; Warden et al., 2006; Cernak and Noble-Haeusslein, 2010; Donkin and Vink, 2010). Metabolic changes include abnormal levels of oxygenation (hypoxia), altered cell metabolism (e.g., glucose utilization), disrupted energy levels and utilization (leading to ionic imbalance, excitotoxicity, etc.), systemic hormonal secretion, and an upregulation of inflammatory activity (Cook et al., 2008; Feng et al., 2012). Inflammation is almost always a result of injury, and occurs in response to damaging stimuli, triggering the release and activation of cytokines and chemokines and the activation and proliferation of microglia (and astroglia) in the CNS. A propagating immune response may promote neurotoxicity and vascular changes (Morganti-Kossmann et al., 2007; Ziebell and Morganti-Kossmann, 2010; Brown et al., 2011). Vascular abnormalities are marked by aberrations in the water content of the brain parenchyma, dysregulation of water channels, and a compromised blood-brain barrier (BBB). Vascular abnormalities can be triggered by cyto- and vasogenic factors leading to edema, vasospasm, and altered rates of perfusion. Diffuse axonal injury is also common, and entails a loss of membrane integrity, altered axonal architecture, Wallerian-type axonal degeneration, metabolic disruption leading to degeneration, and increased serum levels of axonal proteins and filaments (Meythaler et al., 2001). Finally, neuronal and glial cell loss results from necrotic and apoptotic cell death during primary and secondary injury, and may lead to an increase in various neuron- and gliaspecific proteins in serum (Stoica and Faden, 2010). Compounded together, these primary and secondary injury processes may lead to a range of neuropsychiatric symptoms, including various forms of memory and learning deficits, anxiety, and depression (Arciniegas, 2011).

According to our current understanding, the various forms of TBI can share common pathological "components" during both the primary and the secondary injury processes. What likely distinguishes the various forms of TBIs are the onsets and relative contributions of these individual components to the overall pathological cascades. Earlier works in different forms of TBI have identified candidate biomarkers associated with the various pathological changes (Figures 2 and 3). Many of these markers are neuron- and glia-specific and reflect damage to the different cellular components of the brain. Others are more ubiquitous and may indicate generalized metabolic changes, inflammation, etc. Changes in the serum levels of some of these markers have also been found in bTBI. The onset, intensity, and temporal patterns of the various pathological components likely depend on the severity of the injury (Figure 4). For example, one of the distinguishing features of sbTBI is the unusually early onset (within hours after exposure) and extent (high severity) of edema, whereas vasospasm is unusually delayed (up to 2 weeks postinjury; Ling and Marshall, 2008; Ling et al., 2009; Ling and Ecklund, 2011). It should also be noted that hemorrhage is not associated with mibTBI, but is highly characteristic of sbTBI (Ling and Marshall, 2008; Ling et al., 2009; Ling and Ecklund, 2011).

The temporal aspects of injury, like the onset of the various pathologies, are especially important in dynamically changing diseases like bTBI. An experimental study investigating the temporal



FIGURE 4 |The onset and the extent of selected pathological mechanisms of bTBI by injury severity. Hypothetical model. The colored horizontal bars intend to illustrate the approximate onset and extent of the individual pathological changes following injury. The color intensities and bold type reflect an increased severity and contribution of the individual pathologies, whereas lighter colors, question marks, and strikethroughs represent weaker, relatively transient, or nearly absent conditions. Abbreviations: hr, hour(s); wk., week(s); mo., month(s).

pattern of changes in serum levels of four of the most commonly used clinical and experimental biomarkers,  $S100\beta$ , NSE, MBP, and NF-H showed that the temporal pattern of changes can reflect injury severity. Using a swine model of explosive blast and monitoring changes in the serum levels of the four markers up to 2 weeks post-injury, Gyorgy et al. (2011) found increases over time in the serum levels of all four markers. Importantly, the temporal pattern of changes in the serum levels of NF-H showed that in sbTBI, serum NF-H levels peaked early (within 6 hours after injury). The temporal pattern of changes of the other three markers showed no correlation with injury severity. This study illustrates how monitoring the temporal pattern of changes (e.g., "time to peak") of serum biomarkers can be useful for identifying injury severity and outcome. This study also underlines the importance of monitoring changes in serum levels of several markers as they can reflect the dynamics of distinct but important pathologies, e.g., glial response (GFAP and S100 $\beta$ ) vs. axonal damage (NF-H).

Experimental data derived from a rodent model of bTBI has shown that in addition to neuronal and glial cell damage, there are also vascular abnormalities that occur in mibTBI (Kovesdi et al., 2011; Kwon et al., 2011). Elevated serum levels of neuronor glia-specific proteins (NF-H, NSE, CK-BB; GFAP, MBP, S100 $\beta$ ) indicate increased permeability of the BBB (in addition to neuronal and glial cell damage or loss). In the same study, the authors found more direct evidence of vascular abnormalities. Serum levels of VEGF, a protein associated with regulating complex vascular functions including vascular permeability (Neufeld et al., 1999; Croll et al., 2004; Rosenstein and Krum, 2004), were significantly elevated. This study has the limitation of measuring serum levels of the protein markers only at a single, terminal time point. However, the terminal time point of the elevated serum protein markers was taken more than 2 months after injury. This finding indicates that there may be long-lasting ongoing pathological changes, even after a single exposure to mild blast. These findings can have great clinical relevance if they can be repeated. Given the large number of soldiers exposed to a single mild blast, and our very limited knowledge about the long-term consequences of blast, a longitudinal study focusing on a few protein biomarkers should be considered. The data from such a study can be correlated with long-term neurobehavioral assessments in order to identify individuals with increased vulnerabilities and also result in a better understanding of the pathobiology of mbTBI.

In addition to neuronal and glial damage, there is also experimental evidence of neuroinflammation and vascular changes as pathological responses to bTBI (Agoston et al., 2009; Kamnaksh et al., 2011; Kovesdi et al., 2011; Kwon et al., 2011). Along with elevated serum levels of VEGF, there is data showing increased levels of Claudin-5 and vWF in rodents after repeated exposures to mild blasts (Ahmed et al., in preparation). The temporal pattern of changes in their serum levels and the correlation between injury severity and the temporal patterns are currently being investigated (Ahmed et al., in preparation).

Evidence from the study discussed above also showed that animals that were exposed to stressful conditions (in order to simulate battlefield conditions and trigger PTSD) but were not injured had no increase in the serum levels of the previously mentioned protein biomarkers(Kwon et al., 2011). Despite this, they had increased serum corticosterone (CORT) levels and displayed behavioral pathologies like increased anxiety. These findings indicate that one may be able to design objective, serum-based differential diagnostics to distinguish between mibTBI and PTSD. Such a test would be especially important because miBTBI is the most frequent form of blast-induced neurotrauma, accounting for approximately 70% of all bTBIs (Trudeau et al., 1998; Thompson et al., 2008; Elder and Cristian, 2009; Elder et al., 2010; Rosenfeld and Ford, 2010). Moreover, PTSD has emerged as one of the most frequent and lasting consequences of recent military conflicts (Rosenfeld and Ford, 2010; Belanger et al., 2011; Luethcke et al., 2011). Mild bTBI and PTSD have overlapping neurobehavioral symptoms, clinically as well as experimentally. Based on current knowledge, exposure to blast can trigger cellular damage, thus requiring different therapeutic interventions than those used for PTSD (Thompson et al., 2008; Elder et al., 2010; Luethcke et al., 2011). Soldiers showing no functional deficit based on current neurobehavioral assessments (e.g., MACE) after exposure to mild levels of explosive blast return to duty and often become re-exposed to additional blasts (Hayes et al., 2012). As early studies implicate, additional exposures can have severe consequences, including an increased risk of developing long-term neuropsychiatric abnormalities (Okie, 2005; Peota, 2005; Aarabi and Simard, 2009; Jaffee and Meyer, 2009; Cernak et al., 2011; Plurad, 2011; Hayes et al., 2012). Serum protein biomarkers that can indicate the extent of individual vulnerability are of major value. For example, advanced "bio-dosimeters" can be developed for soldiers by using a combination of serumbased health information and physical parameters provided by helmet-mounted sensors (accelerometers). Such a personalized tool can indicate the real-time vulnerability of a soldier to any additional blast. Similar personalized "dosimeters" can also be developed for athletes with a high risk of repeated TBI, such as NFL players.

There are several conceptual, logistical, and technical problems associated with developing serum biomarkers as a diagnostic tool in neuronal insults like TBI. Technical problems include selecting the best proteomics method for serum biomarker discovery (Agoston et al., 2009). To facilitate the antibody-based validation of serum protein biomarkers in bTBI, we listed potential markers as a function of their association with different pathologies and severities of TBI (Figure 3). Some of the markers ("the usual suspects") have been well studied, established, and analyzed in bTBI (Bauman et al., 2009; Gyorgy et al., 2011; Kovesdi et al., 2011; Kwon et al., 2011). Unfortunately, only a few of the listed markers have been verified by clinical studies to show changes in their serum levels specifically due to injury to the brain (and not to other organs). Even fewer markers have been evaluated in clinical settings and correlated with functional and neurobehavioral changes (e.g., GCS and other neurobehavioral tests) routinely used in clinical settings (Agoston et al., 2012).

In addition to aiding in the diagnosis and assessment of injury severity, serum protein biomarkers in mTBI can provide critical information for designing individualized treatment and for monitoring disease progression and treatment effectiveness. Simple versions of such an approach are already in use at neurointensive care units, where serum C-reactive Protein levels are monitored to assess general inflammation and treatment effectiveness. However, the current lack of clinical evidence about how changes in serum levels of protein biomarkers correlate to pathomechanisms and functional outcomes in TBI is a major hindrance. Concentrated, large scale, and preferably international research efforts are needed in order to generate reliable and clinically useful information for aiding in the evidence-based treatment of TBI.

## **SUMMARY AND FUTURE DIRECTIONS**

We would like to conclude this focused review by suggesting three relatively easy and implementable measures that can speed up both serum protein biomarker discovery and validation in bTBI. First: blood should be obtained at multiple time points (serial sampling) in both experimental and clinical studies in order to enable the longitudinal analysis of changes in serum levels of protein markers in bTBI. This approach can reveal temporal patterns of changes that may be of vital diagnostic and therapeutic value. Second: changes in the serum levels of a whole panel of proteins rather than a single protein need to be analyzed (at multiple time points). Multiplex assays, already in use in cancer biology, can provide substantially improved diagnostic precision, and especially so if combined with a systems biology analysis. Third: changes in serum protein biomarkers should be analyzed in relation to functional and neurobehavioral changes in both clinical and experimental settings. Such a combined analysis would radically improve the diagnostic and prognostic value of serum protein biomarkers by facilitating a much more direct understanding of how serum changes relate to functional deficits.

In addition to proteins, there are other potential biomarkers such as microRNAs. While microRNAs have some advantages (e.g.,

stability) compared to proteins, there is currently much less known about their functions. However, as our knowledge increases about their involvement in the various pathological processes their value as serum biomarkers will also increase.

In summary, serum-based protein biomarkers have the enormous potential to fundamentally change our understanding of bTBI and ultimately can – and will – be of major help in designing

### REFERENCES

- Aarabi, B., and Simard, J. M. (2009). Traumatic brain injury. Curr. Opin. Crit. Care 15, 548–553.
- Agoston, D. V., Gyorgy, A., Eidelman, O., and Pollard, H. B. (2009). Proteomic biomarkers for blast neurotrauma: targeting cerebral edema, inflammation, and neuronal death cascades. *J. Neurotrauma* 26, 901–911.
- Agoston, D. V., Risling, M., and Bellander, B. M. (2012). Benchto-bedside and bedside back to the bench; coordinating clinical and experimental traumatic brain injury studies. *Front. Neurol.* 3:3. doi:10.3389/fneur.2012.00003
- Anderson, K. J., Scheff, S. W., Miller, K. M., Roberts, K. N., Gilmer, L. K., Yang, C., and Shaw, G. (2008). The phosphorylated axonal form of the neurofilament subunit NF-H (pNF-H) as a blood biomarker of traumatic brain injury. *J. Neurotrauma* 25, 1079–1085.
- Arciniegas, D. B. (2011). Addressing neuropsychiatric disturbances during rehabilitation after traumatic brain injury: current and future methods. *Dialogues Clin. Neurosci.* 13, 325–345.
- Bauman, R. A., Ling, G., Tong, L., Januszkiewicz, A., Agoston, D., Delanerolle, N., Kim, Y., Ritzel, D., Bell, R., Ecklund, J., Armonda, R., Bandak, F., and Parks, S. (2009). An introductory characterization of a combat-casualty-care relevant swine model of closed head injury resulting from exposure to explosive blast. *J. Neurotrauma* 26, 841–860.
- Begaz, T., Kyriacou, D. N., Segal, J., and Bazarian, J. J. (2006). Serum biochemical markers for postconcussion syndrome in patients with mild traumatic brain injury. J. Neurotrauma 23, 1201–1210.
- Belanger, H. G., Proctor-Weber, Z., Kretzmer, T., Kim, M., French, L. M., and Vanderploeg, R. D. (2011). Symptom complaints following reports of blast versus non-blast mild TBI: does mechanism of injury matter? *Clin. Neuropsychol.* 25, 702–715.
- Bellander, B. M., Olafsson, I. H., Ghatan, P. H., Bro Skejo, H. P., Hansson,

L. O., Wanecek, M., and Svensson, M. A. (2011). Secondary insults following traumatic brain injury enhance complement activation in the human brain and release of the tissue damage marker S100B. *Acta Neurochir* (*Wien*) 153, 90–100

- Berger, R. P. (2006). The use of serum biomarkers to predict outcome after traumatic brain injury in adults and children. J. Head Trauma Rehabil. 21, 315–333.
- Berger, R. P., Adelson, P. D., Pierce, M. C., Dulani, T., Cassidy, L. D., and Kochanek, P. M. (2005). Serum neuron-specific enolase, S100B, and myelin basic protein concentrations after inflicted and noninflicted non-inflicted traumatic brain injury in children. J. Neurosurg. 103, 61–68.
- Berger, R. P., Hayes, R. L., Richichi, R., Beers, S. R., and Wang, K. K. (2012). Serum concentrations of ubiquitin C-terminal hydrolase-L1 and alphaII-spectrin breakdown product 145 kDa correlate with outcome after pediatric TBI. *J. Neurotrauma* 29, 162–167.
- Bochicchio, G. V., Lumpkins, K., O'Connor, J., Simard, M., Schaub, S., Conway, A., Bochicchio, K., and Scalea, T. M. (2008). Blast injury in a civilian trauma setting is associated with a delay in diagnosis of traumatic brain injury. *Am. Surg.* 74, 267–270.
- Brophy, G. M., Pineda, J. A., Papa, L., Lewis, S. B., Valadka, A. B., Hannay, H. J., Heaton, S. C., Demery, J. A., Liu, M. C., Tepas, J. J. III, Gabrielli, A., Robicsek, S., Wang, K. K., Robertson, C. S., and Hayes, R. L. (2009). AlphaII-Spectrin breakdown product cerebrospinal fluid exposure metrics suggest differences in cellular injury mechanisms after severe traumatic brain injury. J. Neurotrauma 26, 471–479.
- Brown, T. S., Hawksworth, J. S., Sheppard, F. R., Tadaki, D. K., and Elster, E. (2011). Inflammatory response is associated with critical colonization in combat wounds. *Surg. Infect.* (*Larchmt*) 12, 351–357.
- Bulut, M., Koksal, O., Dogan, S., Bolca, N., Ozguc, H., Korfali, E., Ilcol, Y. O., and Parklak, M. (2006). Tau protein

evidence-based treatments for individuals suffering from the consequences of blast injury.

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as a serum marker of brain damage in mild traumatic brain injury: preliminary results. *Adv. Ther.* 23, 12–22.

- Cernak, I., Merkle, A. C., Koliatsos, V. E., Bilik, J. M., Luong, Q. T., Mahota, T. M., Xu, L., Slack, N., Windle, D., and Ahmed, F. A. (2011). The pathobiology of blast injuries and blastinduced neurotrauma as identified using a new experimental model of injury in mice. *Neurobiol. Dis.* 41, 538–551.
- Cernak, I., and Noble-Haeusslein, L. J. (2010). Traumatic brain injury: an overview of pathobiology with emphasis on military populations. J. Cereb. Blood Flow Metab. 30, 255–266.
- Cernak, I., Savic, J., Zunic, G., Pejnovic, N., Jovanikic, O., and Stepic, V. (1999). Recognizing, scoring, and predicting blast injuries. *World J. Surg*, 23, 44–53.
- Champion, H. R., Holcomb, J. B., and Young, L. A. (2009). Injuries from explosions: physics, biophysics, pathology, and required research focus. *J. Trauma* 66, 1468–1477; discussion 1477.
- Cheng, J., Gu, J., Ma, Y., Yang, T., Kuang, Y., Li, B., and Kang, J. (2010). Development of a rat model for studying blast-induced traumatic brain injury. J. Neurol. Sci. 294, 23–28.
- Cook, A. M., Peppard, A., and Magnuson, B. (2008). Nutrition considerations in traumatic brain injury. *Nutr. Clin. Pract.* 23, 608–620.
- Coronado, V. G., Xu, L., Basavaraju, S. V., Mcguire, L. C., Wald, M. M., Faul, M. D., Guzman, B. R., and Hemphill, J. D. (2011). Surveillance for traumatic brain injury-related deaths – United States, 1997–2007. MMWR Surveill. Summ. 60, 1–32.
- Croll, S. D., Goodman, J. H., and Scharfman, H. E. (2004). Vascular endothelial growth factor (VEGF) in seizures: a double-edged sword. Adv. Exp. Med. Biol. 548, 57–68.
- da Rocha, A. B., Zanoni, C., De Freitas, G. R., Andre, C., Himelfarb, S., Schneider, R. F., Grivicich, I., Borges, L., Schwartsmann, G., Kaufmann, M., and Regner, A. (2005). Serum Hsp70 as an early predictor of fatal outcome after severe traumatic brain

injury in males. J. Neurotrauma 22, 966–977.

- Dash, P. K., Redell, J. B., Hergenroeder, G., Zhao, J., Clifton, G. L., and Moore, A. (2010). Serum ceruloplasmin and copper are early biomarkers for traumatic brain injuryassociated elevated intracranial pressure. J. Neurosci. Res. 88, 1719–1726.
- De Oliveira, C. O., Reimer, A. G., Da Rocha, A. B., Grivicich, I., Schneider, R. F., Roisenberg, I., Regner, A., and Simon, D. (2007). Plasma von Willebrand factor levels correlate with clinical outcome of severe traumatic brain injury. *J. Neurotrauma* 24, 1331–1338.
- Donkin, J. J., and Vink, R. (2010). Mechanisms of cerebral edema in traumatic brain injury: therapeutic developments. *Curr. Opin. Neurol.* 23, 293–299.
- Drake, A. I., Meyer, K. S., Cessante, L. M., Cheung, C. R., Cullen, M. A., Mcdonald, E. C., and Holland, M. C. (2010). Routine TBI screening following combat deployments. *NeuroRehabilitation* 26, 183–189.
- Elder, G. A., and Cristian, A. (2009). Blast-related mild traumatic brain injury: mechanisms of injury and impact on clinical care. *Mt. Sinai J. Med.* 76, 111–118.
- Elder, G. A., Mitsis, E. M., Ahlers, S. T., and Cristian, A. (2010). Blast-induced mild traumatic brain injury. *Psychiatr. Clin. North Am.* 33, 757–781.
- Feng, J. F., Zhao, X., Gurkoff, G. G., Van, K. C., Shahlaie, K., and Lyeth, B. G. (2012). Post-traumatic hypoxia exacerbates neuronal cell death in the hippocampus. *J. Neurotrauma* 29, 1167–1179.
- Gabbita, S. P., Scheff, S. W., Menard, R. M., Roberts, K., Fugaccia, I., and Zemlan, F. P. (2005). Cleaved-tau: a biomarker of neuronal damage after traumatic brain injury. *J. Neurotrauma* 22, 83–94.
- Galarneau, M. R., Woodruff, S. I., Dye, J. L., Mohrle, C. R., and Wade, A. L. (2008). Traumatic brain injury during Operation Iraqi Freedom: findings from the United States Navy-Marine Corps Combat Trauma Registry. J. Neurosurg. 108, 950–957.

- Garman, R. H., Jenkins, L. W., Switzer, R. C. III, Bauman, R. A., Tong, L. C., Swauger, P. V., Parks, S. A., Ritzel, D. V., Dixon, C. E., Clark, R. S., Bayir, H., Kagan, V., Jackson, E. K., and Kochanek, P. M. (2011). Blast exposure in rats with body shielding is characterized primarily by diffuse axonal injury. *J. Neurotrauma* 28, 947–959.
- Ghirnikar, R. S., Lee, Y. L., and Eng, L. F. (1998). Inflammation in traumatic brain injury: role of cytokines and chemokines. *Neurochem. Res.* 23, 329–340.
- Gonzclez-Mao, M. C., Reparaz-Andrade, A., Del Campo-Perez, V., Alvarez-Garcia, E., Vara-Perez, C., and Andrade-Olivie, M. A. (2011). Model predicting survival/exitus after traumatic brain injury: biomarker S100B 24h. *Clin. Lab.* 57, 587–597.
- Graham, M. R., Myers, T., Evans, P., Davies, B., Cooper, S. M., Bhattacharya, K., Grace, F. M., and Baker, J. S. (2011). Direct hits to the head during amateur boxing is associated with a rise in serum biomarkers for brain injury. *Int. J. Immunopathol. Pharmacol.* 24, 119–125.
- Guy, R. J., Glover, M. A., and Cripps, N. P. (2000). Primary blast injury: pathophysiology and implications for treatment. Part III: injury to the central nervous system and the limbs. J. R. Nav. Med. Serv. 86, 27–31.
- Gyorgy, A., Ling, G., Wingo, D., Walker, J., Tong, L., Parks, S., Januszkiewicz, A., Baumann, R., and Agoston, D. V. (2011). Time-dependent changes in serum biomarker levels after blast traumatic brain injury. *J. Neurotrauma* 28, 1121–1126.
- Haqqani, A. S., Hutchison, J. S., Ward, R., and Stanimirovic, D. B. (2007). Biomarkers and diagnosis; protein biomarkers in serum of pediatric patients with severe traumatic brain injury identified by ICAT-IC-MS/MS. J. Neurotrauma 24, 54–74.
- Hardemark, H. G., Ericsson, N., Kotwica, Z., Rundstrom, G., Mendel-Hartvig, I., Olsson, Y., Pahlman, S., and Persson, L. (1989). S-100 protein and neuron-specific enolase in CSF after experimental traumatic or focal ischemic brain damage. J. Neurosurg. 71, 727–731.
- Hayakata, T., Shiozaki, T., Tasaki, O., Ikegawa, H., Inoue, Y., Toshiyuki, F., Hosotubo, H., Kieko, F., Yamashita, T., Tanaka, H., Shimazu, T., and Sugimoto, H. (2004). Changes in CSF S100B and cytokine concentrations in early-phase severe traumatic brain injury. *Shock* 22, 102–107.

- Hayes, J. P., Morey, R. A., and Tupler, L. A. (2012). A case of frontal neuropsychological and neuroimaging signs following multiple primaryblast exposure. *Neurocase* 18, 258– 269.
- Hergenroeder, G. W., Redell, J. B., Moore, A. N., and Dash, P. K. (2008). Biomarkers in the clinical diagnosis and management of traumatic brain injury. *Mol. Diagn. Ther.* 12, 345–358.
- Hicks, R. R., Fertig, S. J., Desrocher, R. E., Koroshetz, W. J., and Pancrazio, J. J. (2010). Neurological effects of blast injury. J. Trauma 68, 1257–1263.
- Higashida, T., Kreipke, C. W., Rafols, J. A., Peng, C., Schafer, S., Schafer, P., Ding, J. Y., Dornbos, D. III, Li, X., Guthikonda, M., Rossi, N. F., and Ding, Y. (2011). The role of hypoxiainducible factor-1alpha, aquaporin-4, and matrix metalloproteinase-9 in blood-brain barrier disruption and brain edema after traumatic brain injury. *J. Neurosurg.* 114, 92–101.
- Hoge, C. W., Mcgurk, D., Thomas, J. L., Cox, A. L., Engel, C. C., and Castro, C. A. (2008). Mild traumatic brain injury in U.S. Soldiers returning from Iraq. *N. Engl. J. Med.* 358, 453–463.
- Honda, M., Tsuruta, R., Kaneko, T., Kasaoka, S., Yagi, T., Todani, M., Fujita, M., Izumi, T., and Maekawa, T. (2010). Serum glial fibrillary acidic protein is a highly specific biomarker for traumatic brain injury in humans compared with S-100B and neuron-specific enolase. *J. Trauma* 69, 104–109.
- Jaffee, M. S., and Meyer, K. S. (2009). A brief overview of traumatic brain injury (TBI) and post-traumatic stress disorder (PTSD) within the Department of Defense. *Clin. Neuropsychol.* 23, 1291–1298.
- Kamnaksh, A., Kovesdi, E., Kwon, S. K., Wingo, D., Ahmed, F., Grunberg, N. E., Long, J., and Agoston, D. V. (2011). Factors affecting blast traumatic brain injury. *J. Neurotrauma* 28, 2145–2153.
- Kinoshita, K., Chatzipanteli, K., Vitarbo, E., Truettner, J. S., Alonso, O. F., and Dietrich, W. D. (2002). Interleukinlbeta messenger ribonucleic acid and protein levels after fluidpercussion brain injury in rats: importance of injury severity and brain temperature. *Neurosurgery* 51, 195–203: discussion 203.
- Kluger, Y., Kashuk, J., and Mayo, A. (2004). Terror bombingmechanisms, consequences and implications. *Scand. J. Surg.* 93, 11–14.

- Kochanek, P. M., Bramlett, H., Dietrich, W. D., Dixon, C. E., Hayes, R. L., Povlishock, J., Tortella, F. C., and Wang, K. K. (2011). A novel multicenter preclinical drug screening and biomarker consortium for experimental traumatic brain injury: operation brain trauma therapy. J. Trauma 71, S15–S24.
- Kovesdi, E., Gyorgy, A. B., Kwon, S. K., Wingo, D. L., Kamnaksh, A., Long, J. B., Kasper, C. E., and Agoston, D. V. (2011). The effect of enriched environment on the outcome of traumatic brain injury; a behavioral, proteomics, and histological study. *Front. Neurosci.* 5:42. doi:10.3389/fnins.2011.00042
- Kwon, S. K., Kovesdi, E., Gyorgy, A. B., Wingo, D., Kamnaksh, A., Walker, J., Long, J. B., and Agoston, D. V. (2011). Stress and traumatic brain injury: a behavioral, proteomics, and histological study. *Front. Neurol.* 2:12. doi:10.3389/fneur.2011.00012
- Lenzlinger, P. M., Morganti-Kossmann, M. C., Laurer, H. L., and Mcintosh, T. K. (2001). The duality of the inflammatory response to traumatic brain injury. *Mol. Neurobiol.* 24, 169–181.
- Levin, H. S., Wilde, E., Troyanskaya, M., Petersen, N. J., Scheibel, R., Newsome, M., Radaideh, M., Wu, T., Yallampalli, R., Chu, Z., and Li, X. (2010). Diffusion tensor imaging of mild to moderate blast-related traumatic brain injury and its sequelae. J. Neurotrauma 27, 683–694.
- Ling, G., Bandak, F., Armonda, R., Grant, G., and Ecklund, J. (2009). Explosive blast neurotrauma. J. Neurotrauma 26, 815–825.
- Ling, G. S., and Ecklund, J. M. (2011). Traumatic brain injury in modern war. *Curr. Opin. Anaesthesiol.* 24, 124–130.
- Ling, G. S., and Marshall, S. A. (2008). Management of traumatic brain injury in the intensive care unit. *Neurol. Clin.* 26, 409–426; viii.
- Liu, M. C., Akinyi, L., Scharf, D., Mo, J., Larner, S. F., Muller, U., Oli, M. W., Zheng, W., Kobeissy, F., Papa, L., Lu, X. C., Dave, J. R., Tortella, F. C., Hayes, R. L., and Wang, K. K. (2010). Ubiquitin C-terminal hydrolase-L1 as a biomarker for ischemic and traumatic brain injury in rats. *Eur. J. Neurosci.* 31, 722–732.
- Liu, M. C., Akle, V., Zheng, W., Kitlen, J., O'Steen, B., Larner, S. F., Dave, J. R., Tortella, F. C., Hayes, R. L., and Wang, K. K. (2006). Extensive degradation of myelin basic protein isoforms by calpain following traumatic brain injury. J. Neurochem. 98, 700–712.

- Luethcke, C. A., Bryan, C. J., Morrow, C. E., and Isler, W. C. (2011). Comparison of concussive symptoms, cognitive performance, and psychological symptoms between acute blast-versus nonblastnonblast-induced mild traumatic brain injury. J. Int. Neuropsychol. Soc. 17, 36–45.
- Marion, D. W., Curley, K. C., Schwab, K., and Hicks, R. R. (2011). Proceedings of the military mTBI Diagnostics Workshop, St. Pete Beach, August 2010. J. Neurotrauma 28, 517–526.
- Mayorga, M. A. (1997). The pathology of primary blast overpressure injury. *Toxicology* 121, 17–28.
- Meythaler, J. M., Peduzzi, J. D., Eleftheriou, E., and Novack, T. A. (2001). Current concepts: diffuse axonal injury-associated traumatic brain injury. Arch. Phys. Med. Rehabil. 82, 1461–1471.
- Morganti-Kossmann, M. C., Rancan, M., Stahel, P. F., and Kossmann, T. (2002). Inflammatory response in acute traumatic brain injury: a double-edged sword. *Curr. Opin. Crit. Care* 8, 101–105.
- Morganti-Kossmann, M. C., Satgunaseelan, L., Bye, N., and Kossmann, T. (2007). Modulation of immune response by head injury. *Injury* 38, 1392–1400.
- Nakagawa, A., Manley, G. T., Gean, A. D., Ohtani, K., Armonda, R., Tsukamoto, A., Yamamoto, H., Takayama, K., and Tominaga, T. (2011). Mechanisms of primary blast-induced traumatic brain injury: insights from shock-wave research. J. Neurotrauma 28, 1101–1119.
- Neufeld, G., Cohen, T., Gengrinovitch, S., and Poltorak, Z. (1999). Vascular endothelial growth factor (VEGF) and its receptors. *FASEB J.* 13, 9–22.
- Newcomb, J. K., Kampfl, A., Posmantur, R. M., Zhao, X., Pike, B. R., Liu, S. J., Clifton, G. L., and Hayes, R. L. (1997). Immunohistochemical study of calpain-mediated breakdown products to alpha-spectrin following controlled cortical impact injury in the rat. *J. Neurotrauma* 14, 369–383.
- Nortje, J., and Menon, D. K. (2004). Traumatic brain injury: physiology, mechanisms, and outcome. *Curr. Opin. Neurol.* 17, 711–718.
- Okie, S. (2005). Traumatic brain injury in the war zone. *N. Engl. J. Med.* 352, 2043–2047.
- Ommaya, A. K., Dannenberg, A. L., and Salazar, A. M. (1996). Causation, incidence, and costs of traumatic brain injury in the U.S.

military medical system. J. Trauma 40, 211–217.

- Owens, B. D., Kragh, J. F. Jr., Wenke, J. C., Macaitis, J., Wade, C. E., and Holcomb, J. B. (2008). Combat wounds in operation Iraqi Freedom and operation Enduring Freedom. J. Trauma 64, 295–299.
- Papa, L., Akinyi, L., Liu, M. C., Pineda, J. A., Tepas, J. J. III, Oli, M. W., Zheng, W., Robinson, G., Robicsek, S. A., Gabrielli, A., Heaton, S. C., Hannay, H. J., Demery, J. A., Brophy, G. M., Layon, J., Robertson, C. S., Hayes, R. L., and Wang, K. K. (2010). Ubiquitin C-terminal hydrolase is a novel biomarker in humans for severe traumatic brain injury. *Crit. Care Med.* 38, 138–144.
- Papa, L., Lewis, L. M., Falk, J. L., Zhang, Z., Silvestri, S., Giordano, P., Brophy, G. M., Demery, J. A., Dixit, N. K., Ferguson, I., Liu, M. C., Mo, J., Akinyi, L., Schmid, K., Mondello, S., Robertson, C. S., Tortella, F. C., Hayes, R. L., and Wang, K. K. (2012). Elevated levels of serum glial fibrillary acidic protein breakdown products in mild and moderate traumatic brain injury are associated with intracranial lesions and neurosurgical intervention. Ann. Emerg. Med. 59, 471–483.
- Peleg, K., and Savitsky, B. (2009). Terrorism-related injuries versus road traffic accident-related trauma: 5 years of experience in Israel. *Disaster Med. Public Health Prep.* 3, 196–200.
- Pelinka, L. E., Kroepfl, A., Leixnering, M., Buchinger, W., Raabe, A., and Redl, H. (2004). GFAP versus S100B in serum after traumatic brain injury: relationship to brain damage and outcome. *J. Neurotrauma* 21, 1553–1561.
- Peota, C. (2005). Invisible wounds. Minn. Med. 88, 13–14.
- Petzold, A. (2005). Neurofilament phosphoforms: surrogate markers for axonal injury, degeneration and loss. *J. Neurol. Sci.* 233, 183–198.
- Pike, B. R., Flint, J., Dutta, S., Johnson, E., Wang, K. K., and Hayes, R. L. (2001). Accumulation of non-erythroid alpha IIspectrin and calpain-cleaved alpha II-spectrin breakdown products in cerebrospinal fluid after traumatic brain injury in rats. J. Neurochem. 78, 1297–1306.
- Pineda, J. A., Wang, K. K., and Hayes, R. L. (2004). Biomarkers of proteolytic damage following traumatic brain injury. *Brain Pathol.* 14, 202–209.
- Pleines, U. E., Morganti-Kossmann, M. C., Rancan, M., Joller, H., Trentz, O., and Kossmann, T. (2001). S-100

beta reflects the extent of injury and outcome, whereas neuronal specific enolase is a better indicator of neuroinflammation in patients with severe traumatic brain injury. *J. Neurotrauma* 18, 491–498.

- Plurad, D. S. (2011). Blast injury. Mil. Med. 176, 276–282.
- Ramasamy, A., Hill, A. M., and Clasper, J. C. (2009a). Improvised explosive devices: pathophysiology, injury profiles and current medical management. J. R. Army. Med. Corps. 155, 265–272.
- Ramasamy, A., Hill, A. M., Hepper, A. E., Bull, A. M., and Clasper, J. C. (2009b). Blast mines: physics, injury mechanisms and vehicle protection. *J. R. Army. Med. Corps.* 155, 258–264.
- Ringger, N. C., O'Steen, B. E., Brabham, J. G., Silver, X., Pineda, J., Wang, K. K., Hayes, R. L., and Papa, L. (2004). A novel marker for traumatic brain injury: CSF alphaII-spectrin breakdown product levels. *J. Neurotrauma* 21, 1443–1456.
- Rosenfeld, J. V., and Ford, N. L. (2010). Bomb blast, mild traumatic brain injury and psychiatric morbidity: a review. *Injury* 41, 437–443.
- Rosenstein, J. M., and Krum, J. M. (2004). New roles for VEGF in nervous tissue – beyond blood vessels. *Exp. Neurol.* 187, 246–253.
- Rothoerl, R. D., Brawanski, A., and Woertgen, C. (2000). S-100B protein serum levels after controlled cortical impact injury in the rat. *Acta Neurochir.* (*Wien*) 142, 199–203.
- Saatman, K. E., Creed, J., and Raghupathi, R. (2010). Calpain as a therapeutic target in traumatic brain injury. *Neurotherapeutics* 7, 31–42.
- Sandler, S. J., Figaji, A. A., and Adelson, P. D. (2010). Clinical applications of biomarkers in pediatric traumatic brain injury. *Childs Nerv. Syst.* 26, 205–213.
- Sayer, N. A. (2012). Traumatic brain injury and its neuropsychiatric sequelae in war veterans. Annu. Rev. Med. 63, 405–419.
- Secer, H. I., Gonul, E., and Izci, Y. (2007). Head injuries due to landmines. Acta Neurochir. (Wien) 149, 777–781; discussion 782.
- Siman, R., Toraskar, N., Dang, A., Mcneil, E., Mcgarvey, M., Plaum, J., Maloney, E., and Grady, M. S. (2009). A panel of neuron-enriched proteins as markers for traumatic brain injury in humans. *J. Neurotrauma* 26, 1867–1877.
- Stein, D. M., Lindell, A., Murdock, K. R., Kufera, J. A., Menaker, J., Keledjian, K., Bochicchio, G. V., Aarabi, B., and Scalea, T. M. (2011). Relationship of serum and cerebrospinal

fluid biomarkers with intracranial hypertension and cerebral hypoperfusion after severe traumatic brain injury. *J. Trauma* 70, 1096–1103.

- Stern, R. A., Riley, D. O., Daneshvar, D. H., Nowinski, C. J., Cantu, R. C., and Mckee, A. C. (2011). Long-term consequences of repetitive brain trauma: chronic traumatic encephalopathy. *Platin. Met. Rev.* 3, S460–S467.
- Stoica, B. A., and Faden, A. I. (2010). Cell death mechanisms and modulation in traumatic brain injury. *Neurotherapeutics* 7, 3–12.
- Surbatovic, M., Filipovic, N., Radakovic, S., Stankovic, N., and Slavkovic, Z. (2007). Immune cytokine response in combat casualties: blast or explosive trauma with or without secondary sepsis. *Mil. Med.* 172, 190–195.
- Svetlov, S. I., Prima, V., Kirk, D. R., Gutierrez, H., Curley, K. C., Hayes, R. L., and Wang, K. K. (2010). Morphologic and biochemical characterization of brain injury in a model of controlled blast overpressure exposure. J. Trauma 69, 795–804.
- Tarmey, N. T., Park, C. L., Bartels, O. J., Konig, T. C., Mahoney, P. F., and Mellor, A. J. (2011). Outcomes following military traumatic cardiorespiratory arrest: a prospective observational study. *Resuscitation* 82, 1194–1197.
- Taylor, B. C., Hagel, E. M., Carlson, K. F., Cifu, D. X., Cutting, A., Bidelspach, D. E., and Sayer, N. A. (2012). Prevalence and costs of co-occurring traumatic brain injury with and without psychiatric disturbance and pain among Afghanistan and Iraq War Veteran V.A. users. *Med. Care* 50, 342–346.
- Tenielian, T., and Jaycox, L. H. (2008). Invisible Wounds of War: Psychological and Cognitive Injuries, their Consequences, and Services to Assist Recovery. Los Angeles: RAND Corporation, The RAND Centre for Military Health Policy Research.
- Thompson, J. M., Scott, K. C., and Dubinsky, L. (2008). Battlefield brain: unexplained symptoms and blast-related mild traumatic brain injury. *Can. Fam. Physician* 54, 1549–1551.
- Townend, W., Dibble, C., Abid, K., Vail, A., Sherwood, R., and Lecky, F. (2006). Rapid elimination of protein S-100B from serum after minor head trauma. *J. Neurotrauma* 23, 149–155.
- Trudeau, D. L., Anderson, J., Hansen, L. M., Shagalov, D. N., Schmoller, J., Nugent, S., and Barton, S. (1998). Findings of mild traumatic brain injury in combat veterans with

PTSD and a history of blast concussion. *J. Neuropsychiatry Clin. Neurosci.* 10, 308–313.

- Ursano, R. J., Goldenberg, M., Zhang, L., Carlton, J., Fullerton, C. S., Li, H., Johnson, L., and Benedek, D. (2010). Posttraumatic stress disorder and traumatic stress: from bench to bedside, from war to disaster. Ann. N. Y. Acad. Sci. 1208, 72–81.
- Vink, R., Nimmo, A. J., and Cernak, I. (2001). An overview of new and novel pharmacotherapies for use in traumatic brain injury. *Clin. Exp. Pharmacol. Physiol.* 28, 919–921.
- Vitarbo, E. A., Chatzipanteli, K., Kinoshita, K., Truettner, J. S., Alonso, O. F., and Dietrich, W. D. (2004). Tumor necrosis factor alpha expression and protein levels after fluid percussion injury in rats: the effect of injury severity and brain temperature. *Neurosurgery* 55, 416–424; discussion 424–415.
- Vos, P. E., Jacobs, B., Andriessen, T. M., Lamers, K. J., Borm, G. F., Beems, T., Edwards, M., Rosmalen, C. F., and Vissers, J. L. (2010). GFAP and S100B are biomarkers of traumatic brain injury: an observational cohort study. *Neurology* 75, 1786–1793.
- Wallace, D. (2009). Improvised explosive devices and traumatic brain injury: the military experience in Iraq and Afghanistan. Australas. Psychiatry 17, 218–224.
- Warden, D. L., Gordon, B., Mcallister, T. W., Silver, J. M., Barth, J. T., Bruns, J., Drake, A., Gentry, T., Jagoda, A., Katz, D. I., Kraus, J., Labbate, L. A., Ryan, L. M., Sparling, M. B., Walters, B., Whyte, J., Zapata, A., and Zitnay, G. (2006). Guidelines for the pharmacologic treatment of neurobehavioral sequelae of traumatic brain injury. J. Neurotrauma 23, 1468–1501.
- Wilk, J. E., Thomas, J. L., Mcgurk, D. M., Riviere, L. A., Castro, C. A., and Hoge, C. W. (2010). Mild traumatic brain injury (concussion) during combat: lack of association of blast mechanism with persistent postconcussive symptoms. *J. Head Trauma Rehabil.* 25, 9–14.
- Woertgen, C., Rothoerl, R. D., Wiesmann, M., Missler, U., and Brawanski, A. (2002). Glial and neuronal serum markers after controlled cortical impact injury in the rat. Acta Neurochir. Suppl. 81, 205–207.
- Wojcik, B. E., Stein, C. R., Bagg, K., Humphrey, R. J., and Orosco, J. (2010). Traumatic brain injury hospitalizations of U.S. army soldiers deployed to Afghanistan and Iraq. *Am. J. Prev. Med.* 38, S108–S116.

- Wolf, S. J., Bebarta, V. S., Bonnett, C. J., Pons, P. T., and Cantrill, S. V. (2009). Blast injuries. *Lancet* 374, 405–415.
- Young, A. B., Ott, L. G., Beard, D., Dempsey, R. J., Tibbs, P. A., and Mcclain, C. J. (1988). The acute-phase response of the braininjured patient. *J. Neurosurg.* 69, 375–380.
- Zemlan, F. P., Jauch, E. C., Mulchahey, J. J., Gabbita, S. P., Rosenberg, W. S., Speciale, S. G., and Zuccarello, M. (2002). C-tau biomarker of neuronal damage in severe brain injured patients: association with elevated intracranial pressure

and clinical outcome. *Brain Res.* 947, 131–139.

- Ziebell, J. M., and Morganti-Kossmann, M. C. (2010). Involvement of pro- and anti-inflammatory cytokines and chemokines in the pathophysiology of traumatic brain injury. *Neurotherapeutics* 7, 22–30.
- Zurek, J., Bartlova, L., and Fedora, M. (2011). Hyperphosphorylated neurofilament NF-H as a predictor of mortality after brain injury in children. *Brain Inj.* 25, 221–226.
- Zurek, J., and Fedora, M. (2012). The usefulness of S100B, NSE, GFAP,

NF-H, secretagogin and Hsp70 as a predictive biomarker of outcome in children with traumatic brain injury. *Acta Neurochir. (Wien).* 154, 93–103; discussion 103.

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