



# Mechanism of Action and Clinical Potential of Fingolimod for the Treatment of Stroke

Wentao Li1, Haoliang Xu2 and Fernando D. Testai1\*

<sup>1</sup> Department of Neurology and Rehabilitation, University of Illinois College of Medicine, Chicago, IL, USA, <sup>2</sup>Department of Pathology, University of Illinois College of Medicine, Chicago, IL, USA

# Fingolimod (FTY720) is an orally bio-available immunomodulatory drug currently approved by the FDA for the treatment of multiple sclerosis. Currently, there is a significant interest in the potential benefits of FTY720 on stroke outcomes. FTY720 and the sphingolipid signaling pathway it modulates has a ubiquitous presence in the central nervous system and both rodent models and pilot clinical trials seem to indicate that the drug may improve overall functional recovery in different stroke subtypes. Although the precise mechanisms behind these beneficial effects are yet unclear, there is evidence that FTY720 has a role in regulating cerebrovascular responses, blood–brain barrier permeability, and cell survival in the event of cerebrovascular insult. In this article, we critically review the data obtained from the latest laboratory findings and clinical trials involving both ischemic and hemorrhagic stroke, and attempt to form a cohesive picture of FTY720's mechanisms of action in stroke.

Keywords: stroke, neuroprotection, vascular endothelium, fingolimod, FTY720, hemorrhagic stroke, sphingosine-1-phosphate

# INTRODUCTION

Fingolimod (FTY720) is an orally bio-available immunomodulatory drug unique for its reversible leukocyte sequestration properties. In 2010, it was approved by the FDA for the treatment of multiple sclerosis (MS) (1, 2).

Given the current understanding of the role of the immune system in the pathophysiology of brain injury in cerebrovascular diseases, there is now significant interest in the potential benefits of FTY720 on stroke outcomes. Several groups have independently evaluated its effects in rodent models of brain ischemia and intracerebral hemorrhage (ICH). More recently, pilot clinical trials have been conducted demonstrating promising results, albeit in small populations (3–5). Taken together, these studies seem to indicate that administration of FTY720 results in overall improved functional recovery in different stroke subtypes.

The precise mechanisms behind these beneficial effects are still under investigation. FTY720 is a partial sphingosine-1-phosphate (S1P) agonist with immunomodulatory properties that regulates cerebrovascular responses, blood-brain barrier (BBB) permeability, and central nervous system (CNS) cell survival (6, 7). In this article, we will organize these elements and attempt to form a cohesive picture of FTY720's mechanisms of action in stroke, and critically review the data obtained from recent clinical trials.

#### OPEN ACCESS

#### Edited by:

M-Alain Babi, Duke University, USA

## Reviewed by:

Jason D. Hinman, University of California Los Angeles Medical Center, USA Roshini Prakash, University of California Los Angeles, USA Andrew Martin Price, Duke University Hospital, USA

> \*Correspondence: Fernando D. Testai testai@uic.edu

#### Specialty section:

This article was submitted to Neurocritical and Neurohospitalist Care, a section of the journal Frontiers in Neurology

Received: 20 June 2016 Accepted: 15 August 2016 Published: 26 August 2016

#### Citation:

Li W, Xu H and Testai FD (2016) Mechanism of Action and Clinical Potential of Fingolimod for the Treatment of Stroke. Front. Neurol. 7:139. doi: 10.3389/fneur.2016.00139

1

# ROLE OF THE IMMUNE SYSTEM IN STROKE PROGRESSION

Immunomodulation is a well-characterized effect of FTY720 and is thought to mediate some of the beneficial effects seen in stroke models. In order to understand the extent to which the immune system impacts the evolution of stroke outcomes, we will sketch a proposed model of the immune cascade following ischemic insult.

In the immediate aftermath of an ischemic event, complement activation, clot formation, and oxidative stress result in direct damage to local vasculature. Endothelial cells die and detach, interrupting the integrity of the BBB and exposing sub-endothelial antigens. Immune cells adhere to the vessel wall and upregulate the expression of chemoattractant and adhesion molecules that lead to the infiltration of the brain parenchyma by the innate immune system.

A combination of neutrophils, monocytes, and macrophages, this innate immune system further contributes to vascular compromise and early inflammation. One well-documented process is through the release of matrix metalloproteinases (MMPs) by the immune cells, particularly MMP-9, which contributes to the breakdown of the BBB with the resultant edema and growth of the infarcted area (8).

In the parenchyma, glial cells are also activated by the inflammation and damage-associated molecular patterns (DAMPs) released from dying neurons. These reactive astrocytes and microglia further stimulate the recruitment of leukocytes, which release their own pro-inflammatory chemokines, perpetuating a cycle of vascular damage, inflammation, and cell death (9).

The second, adaptive phase of the immune response is mediated predominantly by effector T cells, which are stimulated by DAMPS and brain-specific antigens released upon neuronal cell death (10). These T cells mobilize to the injured regions of the brain, infiltrating a compromised BBB to release proinflammatory cytokines, including IFN- $\gamma$ , resulting in a delayed neurotoxic effect (11, 12). Of note, brain-specific antigens were identified in cervical nodes and palatine tonsils of animals with cerebral ischemia and stroke survivors. Interestingly, some of these antigens were associated with infarct volume and survival. While these studies need to be replicated in larger cohorts, they suggest the participation of peripheral lymphoid tissue in strokeassociated inflammation and outcomes.

Lastly, the inflammatory process is brought to an end via a combination of B cells and regulatory T cells. The latter acts through IL-10, which in combination with TGF- $\beta$  produced by local macrophages, suppresses further helper T-cell-induced inflammation, and promotes the regeneration of remaining viable neurons (13, 14).

A reduction in various components of the innate and adaptive immune response have been associated with better stroke outcomes. Clinically, a lower ratio of CD14+ pro-inflammatory monocytes to CD16+ reparative monocytes has been correlated with better acute and long-term functional outcomes (15). Similarly, decreased complement activation, specifically the reduced expression of C3, C4, and C-reactive protein, has been associated with better recovery at 3 months post-stroke (16). Experimentally, inhibition of CD8+ and CD4+ T-cell migration into the CNS and direct disruption of CD8+ cytotoxicity has led to reduced infarct volume and post-ischemic inflammation (17). Disruption of the DAMPs-activated  $\gamma\delta T$  cells have also resulted in decreased infarct size and better functional recovery in mice (18). Finally, direct delivery of B cells and IL-10 to the brain in animal models have resulted in a reduction of inflammatory cytokines produced by effector T cells and a reduction of infarct size (19, 20).

# FINGOLIMOD AND STROKE-RELATED MECHANISMS OF ACTION

#### Pharmacology

Isaria sinclairii, otherwise known as "winter-insect and summerplant," is a fungus that has been used in traditional Chinese medicine for over 300 years. Classically prescribed as a panacea for multiple ailments, it produces an atypical amino acid myriocin (ISP-1) that blocks the synthesis of sphingolipids. This chemical compound has since been modified into fingolimod, also known as FTY720 (21). In Western Medicine, FTY720 initially showed promise in preventing ischemic-reperfusion injury following organ transplant, but failed clinical trials due to the development of acute macular edema in some patients (22, 23). In 2010, it became the first oral disease modifying drug approved for treatment of MS (6). In its base form, FTY720 is an orally bio-available, lipophilic molecule that readily crosses the BBB and steadily accumulates in the CNS white and gray matter (24). It bears a structural similarity to sphingosine and is reversibly phosphorylated primarily by Sphingosine-Kinase 2 (SphK2) and, to a lesser extent, by SphK1 (25). In its activated form, FTY720phosphate is a sphingosine-1-phosphate (S1P) analog that binds to cell membrane G-coupled S1P receptors (S1PR) S1PR1, 3, 4, and 5, but not S1P2 (26). With the exception of S1PR4, these receptors are ubiquitously distributed in the CNS (Table 1). In addition to its action on cell surface receptors, FTY720 regulates the synthesis of different bioactive sphingolipids and, together with S1P, regulates gene expression via epigenetic mechanisms (Figure 1). The half-life of FTY720 averages ~9 days and its

Type of receptor	Function
S1P1 = S1P3 > S1P2 = S1P5	Neurogenesis, neural progenitor migration, cell survival, and neurotransmission
S1P5 > S1P1 = S1P2 > S1P3	Oligodendrocyte precursor cell survival, migration, differentiation, and morphology
S1P3 > S1P1 > S1P2 > S1P5	Proliferation, migration, gap function communication, and growth factor production
S1P1 > S1P2 > S1P3 = S1P5	Pro-inflammatory cytokine production
	S1P1 = S1P3 > S1P2 = S1P5 S1P5 > S1P1 = S1P2 > S1P3 S1P3 > S1P1 > S1P2 > S1P5



**FIGURE 1** | **Overview of sphingosine-1-phosphate (S1P) and fingolimod (FTY720) effects**. S1P signals through five different G protein-coupled cell surface receptors (S1PR 1-5). S1PR1–3 are ubiquitously expressed throughout the body. In comparison, S1PR4 is predominantly expressed in the immune system and S1PR5 in the central nervous system (CNS) and the spleen. These receptors are associated with different profiles of G $\alpha$  subunits, including G<sub>i</sub>, G<sub>et</sub>, and G<sub>12/13</sub>. The profile of S1PR and G $\alpha$  subunit expression enable cells to activate different downstream signaling pathways and to orchestrate complex responses in response to single agonist. The activated form of FTY720, FTY720-phosphate (FTY720-P), is a partial agonist of all the S1PRs, with the exception of S1PR2. Downstream, S1PRs regulate oxidative-stress-induced cell survival, endothelial barrier function, leukocyte migration and taxis, and oligodendrocyte survival and proliferation in the CNS (26, 30, 31). In the presence of FTY720-P, S1PR1 is downregulated from through non-receptor-related epigenetic mechanisms related to the inhibition of histone deacetylases (HDAC). *Insert:* metabolism of sphingolipids (33). Ceramide, a central hub in the metabolism of sphingolipids, has been associated with cell death, quiescence, and differentiation. It can be generated by the breakdown of sphingonyelin by the acid or neutral sphingonyelinases, or synthesized *de novo* from serine and palmitoyl-CoA by serine-palmitoyl transferase and ceramide synthase. Ceramide can be metabolized into sphingolipids, ischer Shory (34). In addition to its effect on cell membrane receptors and HDAC, FTY720-P also response to response to sphingolipids (35). S1P is broken down into hexadecenal and phosphotethanolamine by the S1P-lyase. FTY720 is phosphorylated primarily by SphK2. Both S1P and FTY720-P are transported to the extracellular space by the membrane transporter Spns2 (34). In addition to its effect on cell membrane receptors and HDAC, FTY720 also inhibits several key enz

pharmacology is not sensibly affected by age, weight, sex, or ethnicity (27).

## Immunomodulation

Immunomodulation, particularly peripheral lymphocyte sequestration, is a well-documented mechanism of FTY720. S1P1receptors are expressed on CCR-7 positive lymphocytes and play a key role in inflammation, stimulating lymphocyte release from peripheral lymphoid tissues in response to endogenous S1P gradients (35, 36). Lymphocyte S1P1-receptor binding by FTY720 results in a functional antagonism via involution and degradation of this cell surface receptor, ultimately inhibiting lymphocyte recirculation (37). In rodent models using 1 mg/kg of FTY720 at 1-, 24-, and 48-h intervals, 60% peripheral lymphopenia can be achieved within 6 h of first administration with peak suppression lasting for 7–9 days (38). By reducing the quantity of infiltrating lymphocytes, FTY720 effectively decreases the direct neurotoxic effects of the adaptive immune system upon the CNS and reduces secondary ischemic damage caused by cytokine-induced inflammation of the surrounding microvasculature (39–41).

Fingolimod may also attenuate the neuroinflammatory response by acting directly on reactive glial cells, which are activated within minutes following ischemic injury (42). Studies done in rodent models of cerebral ischemia show that FTY720 ameliorates the influx of leukocytes in the ischemic brain and reduces the expression of pro-inflammatory cytokines (IL-1ß and IFN- $\gamma$ ) and adhesion molecules, such as intercellular adhesion molecule-1 (ICAM-1) (38, 41). Studies done in vivo in our own laboratory confirm that FTY720 reduces leukocyte adhesion to pial vessels and ameliorates astrogliosis in cerebral ischemia-reperfusion models (Figures 2 and 3). In non-stroke models, FTY720 both suppressed TNF-α-induced inflammatory genes and stimulated the production of neurotrophic mediators via S1P1 and S1P3 receptors on astrocytes (43). In particular, FTY720 decreased ceramide production, a pro-inflammatory lipid that causes loss of BBB integrity and increased infiltration of immune cells into the CNS (44). In support of FTY720's direct CNS effects, direct infusion of FTY720 into rat brains in MS models has been shown to decrease disease severity in the absence of peripheral lymphopenia (7).

#### Vasoprotection

It is possible that FTY720 also provides a vasoprotective effect in a manner related to its immunomodulatory action. There is some evidence that FTY720 can induce granulocyte and macrophage colony-stimulating factor (GM-CSF) release from astrocytes, which may limit endothelial cell death after exposure to TNF- $\alpha$ and IFN- $\gamma$  (47). The presence of astrocyte-derived GM-CSF also suppresses ICAM-1 expression in endothelial cells (48). Decreased surface ICAM-1 reduces leukocyte adhesion to the vessel walls and local platelet activation, which leads to reduced thrombosis and associated inflammation, improving microvascular function (39, 49).

Furthermore, S1P1-receptors are expressed on endothelial cells and administration of FTY720 may directly enhance BBB

integrity. First, FTY720 has been shown to directly decrease endothelial ICAM-1 expression in stroke models, which may further contribute to ameliorating the no-reflow phenomenon (41). Second, in non-stroke-related models, FTY720 can stimulate endothelial cells to recruit proteins for adherence junction assembly, reducing vascular permeability and neutrophil infiltration (50). Finally, there is evidence that administration of FTY720 blocks VEGF-induced permeability in human umbilical vein endothelial cells and regulates endothelial cell barrier capacity and vascular permeability in lungs (51). A similar process may also help to maintain the structural integrity of the BBB.

Taken together, these immunomodulative and vasoprotective mechanisms may contribute to FTY720's ability to decrease secondary edema formation following cerebral ischemia (41). Such a process would also be consistent with our rat model of middle cerebral artery (MCA) ischemia–reperfusion, where FTY720 treatment resulted in decreased ipsilateral brain edema compared to the vehicle group (**Figure 4**). Vasoprotection may also account for reduction in hemorrhagic transformation after delayed recombinant tissue plasminogen activator (tPA or alteplase) administration (at 3 h post-stroke), at least in moderate sized thromboembolic strokes (52, 53).

### **Direct Neuroprotection**

In addition to glial cells, S1P-receptors are ubiquitous within the CNS, with multiple receptor subtypes present on neurons and oligodendrocytes (28). Experimental evidence has shown that fingolimod has direct effects on astrocytes, microglia, oligodendrocytes, neurons, and BBB endothelial cells, which are all affected after stroke (Table 2) (7). There is also increasing evidence that FTY720 may achieve some of its protective effects by directly interacting with neuronal S1P1 and S1P3 receptors, promoting production of anti-apoptotic factors and increasing resilience against ischemic injury. One proposed mechanism of direct neuroprotection is via the S1P/Phosphatidylinositol-3-Kinase (PI3K)/Akt/FOXO3a axis. Administration of FTY720 results in phosphorylation of Akt, which in turn prevents activation of FOXO3a, a transcription factor critical in mediating oxidative stress-induced cell death. In ischemic conditions, this suppression of FOXO3a by FTY720 may contribute to continued neuronal survival (54). FTY720 may also provide a neuroprotective effect







control, and FTY/20-treated animals. (B) Quantification of leukocyte adhesion, expressed as the percentage of the vessel area occupied by adherent leukocytes measured 1 h after leukocyte labeling. The MCAo-vehicle group demonstrated a significant increase in vascular leukocyte adhesion at 24 h post reperfusion compared to sham. The treatment with FTY720 decreased the adherence of leukocytes to pial vessels by almost 60%. Significance determined using an unpaired *t*-test. Means  $\pm$  SD (n = 5).

through the ERK/bcl-2 pathway. Administration of FTY720 results in persistent ERK phosphorylation in the face of ischemic insult, resulting in enhanced levels of the anti-apoptotic protein Bcl-2, thus rescuing neurons from apoptosis (55).

Pretreatment with FTY720 at 48 h before MCA ischemiareperfusion also leads to an ischemia tolerant phenotype in mice models mimicking the effect of hypoxic preconditioning and decreasing subsequent brain ischemia volume by 33%. This mechanism is hypothesized to act via a hypoxia-inducible factor (HIF), SphK2/S1P1-receptor, and chemokine (C–C motif) ligand-2 (CCL2) signaling pathway, ultimately resulting in increased CCL2 expression in neurons (57). CCL2 is a leukocyte-derived pro-inflammatory chemokine shown to be protective against ischemia in the myocardium (58). It is possible that prophylactic treatment with FTY720 leads to early exposure to low doses of CCL2, which may prime neurons against future ischemic damage, however, the exact mechanism remains to be fully elucidated (59).



(A) Representative coronal brain cuts obtained 24 h after MCAo. The arrows highlight areas of cerebral infarct volume was determined as previously described (56). Brain edema was determined by measuring hemispheric water content (41). (A) Representative coronal brain cuts obtained 24 h after MCAo. The arrows highlight areas of cerebral infarction. Brain cuts were digitalized and infarct volume determined by using the image analysis software (MetaMorph; Universal Imaging Corp., Downingtown, PA, USA). (B) Brain edema determined in the ipsilateral and contralateral hemisphere. Compared to vehicle animals, the treatment with 0.5 mg/kg FTY720 reduced infarct volume and brain edema. Significance determined using an unpaired *t*-test. Means  $\pm$  SD (n = 5).

TABLE 2 | Central nervous system effect of fingolimod (7).

Cell type	Function
Neuron	Protects from excitotoxic death Restores neuronal function
Oligodendrocyte	Increases survival of oligodendrocyte precursor cells Enhances remyelination Regulates migration, differentiation, and process dynamics
Astrocyte	Inhibits pro-inflammatory cytokine production Inhibits astrogliosis Regulates cell migration
Microglia	Reduces microglial activation Ameliorates microgliosis
Blood-brain barrier	Reduces blood-brain barrier leakiness

Finally, S1P and FTY720 have intracellular effects that are not mediated by cell surface receptors (60). One identified mechanism is the epigenetic regulation of histone acetylation. Both agents are class I histone deacetylase (HDAC) inhibitors and elevated levels of intranuclear S1P or phosphorylated-FTY720 increase acetylated histone H3 (61). This epigenetic phenomenon has been associated with alterations in hippocampal neurons and rescuing memory deficits independent of FTY720's immunosuppressive actions (62). The beneficial effect of fingolimod on cognition has also been shown in experimental models of neurodegeneration and neuroinflammation, and in memory performance after cerebral ischemia–reperfusion (63–66). However, whether this effect is related to epigenetic mechanisms is yet to be determined.

# FINGOLIMOD IN CLINICAL TRIALS

Given the consistent benefits demonstrated in laboratory stroke models and the favorable safety profile observed in clinical trials involving MS patients (67), some research groups have started to assess FTY720 in stroke subjects. So far, the results of three smallsized studies have been published by the Tianjin Neurological Institute looking at patients with anterior circulation acute ischemic strokes (AIS) and ICH (3–5).

# **Ischemic Stroke**

Two small clinical trials investigated the effect of FTY720 in acute cerebral ischemia. The first study published in 2014 was an openlabel, evaluator-blinded, non-randomized study that examined the effect of FTY720 on ischemic stroke compared to standard of care alone. The inclusion criteria included age >18 years, National Institute of Health Stroke Score (NIHSS) score at baseline  $\geq$ 5, anterior circulation ischemic stroke, and time of onset to admission within 4.5-72 h. Major exclusion criteria included history of bradyarrhythmia or atrioventricular blocks, use of antineoplastic or immunomodulating therapies, and macular edema. A total of three consecutive oral doses of FTY720 0.5 mg daily were administered to the study group. The first dose of the study drug was given 1 h after the baseline MRI and no later than 72 h after symptoms onset. The primary endpoint was lesion size at 7 days and microvascular permeability and clinical improvement. Clinical outcomes included (a) NIHSS which is a stroke severity score that ranges from 0 to 42, with higher numbers representing increased stroke severity; (b) modified Barthel index (mBI) which is a validated score of dependency; this score runs from 0 to 100 with higher numbers denoting better outcome with increasing independency to perform activities of daily living; and (c) modified Rankin Scale (mRS) which is a tool that assesses disability; this scale ranges from 0 to 6, with 0 denoting perfect health and 6 death.

A total of 22 patients were assigned to the control or the FTY720 groups. The treatment with fingolimod was initiated at an average of 20 h after stroke onset. Both the control (n = 11) and the study groups (n = 11) had comparable baseline characteristics. Compared to the lesion volume and location-matched control group, there was no significant decrease in lesion size at 7 days on FLAIR imaging (70  $\pm$  26 vs. 85  $\pm$  30 ml, P = 0.69). However, lesion expansion was more commonly seen in the controls (9  $\pm$  3 vs. 27  $\pm$  8 ml, P = 0.0494) and infarct volume extension ratio, defined as the ratio of infarct volume change from baseline to day 7/infarct volume at baseline, was significantly decreased in the FTY720 group ( $15.2 \pm 4.2$  vs.  $41.6 \pm 4.5$ , P = 0.0003). In addition, microvascular permeability, measured as parenchymal enhancement contrast-enhanced T1-weighted imaging, was 2× higher in controls compared to the FTY720-treated group (p = 0.005). In terms of functional outcomes, study patients demonstrated significant improvement in neurological functions within the first week. This was again observed at 90 days post stroke, with significantly lower NIHSS scores  $(1.7 \pm 0.8 \text{ vs. } 5.8 \pm 1.7, P = 0.02)$ and significantly higher mBI (62  $\pm$  11 vs. 87  $\pm$  8, P = 0.0049). In addition, decreased neurological disability, defined as a mRS score of 0-1 at 90 days, was more commonly seen in the FTY720 groups than in controls (0 vs. 73%; P = 0.009). In terms of safety, the treatment with FTY720 resulted in a sustained lymphopenia that persisted for more than 7 days and resolved by day 30. Significantly, the rate of complications and adverse reactions, including bradycardia, macular edema, or infections, were similar in the control and the study groups.

In 2015, a second randomized, open-label, evaluator-blinded study was conducted to assess the efficacy of FTY720 in conjunction with tPA. The inclusion criteria included age 18–80 years, first-ever hemispheric stroke due to anterior or middle cerebral artery occlusion (MCAo), and initial NIHSS score  $\geq$ 5. Patients were excluded from the study if they had contraindications to tPA, pre-existing disability, history of bradyarrhythmia or atrioventricular blocks, concurrent use with antineoplastic or immunomodulating therapies, and macular edema. The first dose of fingolimod was given before alteplase administration. Similar to the previous study, a total of three doses of 0.5 mg oral fingolimod were given in consecutive days. Lesion size, hemorrhagic volume, and NIHSS scores at 24 h were the primary endpoints; also assessed were lesion size at day 7 and clinical improvement up to day 90.

A total of 47 patients were randomly assigned to receive tPA alone or tPA plus FTY720. The mean time from stroke onset to initiation of treatment was  $\sim$ 3 h. The baseline characteristics of the control and the study groups were comparable with the exception of atrial fibrillation that was more commonly seen in the FTY720 group. Recanalization rates were the same in both groups, but compared to the control group, the combined

treatment with FTY720 and alteplase resulted in decreased infarct volume expansion  $(10.1 \pm 1.2 \text{ vs. } 34.3 \pm 10.4 \text{ ml}; P = 0.04)$ , decreased hemorrhagic transformation volume (1.2  $\pm$  0.4 vs.  $4.4 \pm 1.1$  ml; P = 0.01), lower rate of parenchymal hemorrhage (0 vs. 36%; P = 0.002), and greater improvements in NIHSS scores (4 vs. 2; P = 0.02), all at 24 h. At day 7, the combination therapy also demonstrated decreased lesion volume compared to alteplase treatment alone  $(-2.3 \pm 2.7 \text{ vs. } 12.1 \pm 3.7 \text{ ml}; P < 0.01)$ and greater improvement in the NIHSS score from the presenting stroke (2.5 vs. 1; P < 0.01). At 3 months, the rate of patients with good neurological outcome, defined as mRS 0-1, was higher in the group of patients that received FTY720 plus tPA (73 vs. 32%; P = 0.01). From the safety stand point, malignant cerebral edema requiring craniotomy occurred in only one patient receiving tPA monotherapy but in none of the subjects in the tPA plus alteplase group. Aside from the expected lymphopenia resulting from the use of FTY720, safety outcomes were comparable in both groups.

During ischemia, lymphocytes interact with endothelial cells and platelets to induce microvascular dysfunction and enhanced thrombosis. This process, referred to as no reflow, occurs within a few hours after the ictus and results in the obliteration of small capillaries. In addition, the occurrence of ischemia is associated with a profound local inflammatory response. The term thromboinflammation refers to the active interplay that occurs between thrombotic and inflammatory mechanisms. Data obtained in experimental models of cerebral ischemia/reperfusion demonstrate that FTY720 ameliorates the number of lymphocytes in the intracranial vascular compartment and reduces ischemia-associated vascular dysfunction and microvascular thrombosis. Thus, it is conceivable that FTY720 contributes to improved outcome in cerebral ischemia by reducing thromboinflammation (39). In addition, enhanced inflammation has been linked to BBB dysfunction and increased risk of hemorrhagic transformation. Studies done in animal models of MCA thromboembolic occlusion show that FTY720 attenuates BBB dysfunction and hemorrhagic transformation with delayed tPA administration at 3 h post-stroke (53).

#### **Intracerebral Hemorrhage**

Intracerebral hemorrhage constitutes a neurological emergency that carries a mortality of ~50%. Hematoma volume and location, age, and the presence of intraventricular extension are established predictors of outcome. Perihematomal edema occurs within the first 72 h post ictus and contributes to both morbidity and mortality (68). The breakdown of the hematoma attracts immune cells that infiltrate the parenchyma and secrete pro-inflammatory cytokines and MMPs. These mediators disrupt the BBB and facilitate the recruitment of additional leukocytes that results in a vicious cycle that contributes to secondary brain injury. Peripheral immune cells, including T-lymphocytes, have been identified as active contributors to perihematomal edema. Studies done in experimental models show that the treatment with FTY720 ameliorates ICH-associated infiltration of lymphocytes and the expression of adhesion molecules and other cytokines, including intercellular adhesion molecule-1 (ICAM- 1), interferon-y (INF- γ), and interleukin-17 (IL-17) at 72 h after ICH. This reduction in neuroinflammation was associated with reduced cerebral edema, neuronal apoptosis, and brain atrophy and improved neurological outcome (69–71). These findings supported the notion that fingolimod may be protective in ICH.

The Tianjin group investigated the use of FTY720 in non-traumatic ICH using an open-label evaluator-blinded case-control study. The inclusion criteria included supratentorial cerebral hemorrhages between 5 and 30 ml, Glasgow Coma Scales >6, and symptom onset within 72 h prior to admission. Patient were excluded from the study if they had dysphagia, nausea or vomiting, hematoma expansion, pre-existing disability (defined as mRS > 1), bradycardia or atrioventricular block, secondary ICH, macular edema, or concomitant use of antineoplastic or immunomodulatory treatment. Patient were matched by NIHSS as well as hematoma volume and location and then randomly assigned to the FTY720 or control groups. Primary endpoints included perihematomal edema volume at 7 and 14 days (defined by MR imaging), and functional recovery at 90 days. Clinical management was left to the discretion of the treating physician. A total of 23 subjects met the inclusion and exclusion criteria, 11 of whom received 0.5 mg of FTY720 orally daily for three consecutive days. Mean time from stroke onset to intervention was 20 h, with FTY720 administered roughly 1 h after initial CT evaluation.

Both the control and the FTY720 groups had similar baseline characteristics, with the exception of a higher representation of females in the FTY720 arm. The circulating levels of CD4+ T cells, CD8+ T cells, and CD19+ B cells dropped abruptly after the third dose of FTY720. The initial hematoma volume in both groups was comparable (15.4 vs. 16.7 ml, P = 0.68). At 7 days post ictus, the perihematomal edema volume in the FTY720-treated group was significantly lower than that in the control group (47 vs. 108 ml, P = 0.04). At 14 days, the FTY720 group continued to have a smaller perihematomal edema volume than the control group but this was not statistically significant (55 vs. 124 ml, P = 0.07). The loss of significance at 14 days post ICH may represent a floor effect or lack of power. However, it is important to highlight that 100% of FTY720 patients had a striking functional recovery with a GCS of 15 by day 7 compared to 50% of the controls. The rate of patients with good clinical outcome, defined as an mRS of 0 or 1, was 63% in the FTY720 group and 0% in the control group (P = 0.001). In addition, 63% of the subjects treated with FTY720 had reduced dependency after ICH, defined by an mBI score of 95-100, as compared to 0% in the control group. In terms of safety outcomes, a single case of transient asymptomatic bradycardia was reported in the group treated with FTY720. Otherwise, both groups had no major differences in the frequency of infections or cardiac arrhythmias.

It is plausible that the reduction in perihematomal edema observed in ICH patients treated with FTY720 relates to its immunomodulatory effects. However, other mechanisms may contribute to this phenomenon. By inducing cytoskeletal rearrangement and increasing the expression of adhesion molecules, including VE-cadherin and  $\beta$ -catenin, S1P has been shown to enhance endothelial barrier permeability and leukocyte entry (50). In addition, the participation of S1P-signaling pathways in endothelial integrity and BBB function has been described in different scenarios. In the rodent model of experimental autoimmune encephalomyelitis, for example, treatment with fingolimod modified the expression of MMPs resulting in a proteolytic

balance favoring BBB integrity; and in experimental cerebral malaria, treatment increased serum levels of angiopoietin-1, a marker of endothelium quiescence and stability (72, 73). In addition, FTY720 was shown to block VEGF-induced permeability in human umbilical vein endothelial cells and to regulate endothelial cell barrier capacity and vascular permeability in lungs (51, 52). Whether these mechanisms contributed to the improved outcome observed in ICH patients treated with FTY720 is still to be determined.

## DISCUSSION

Of the small-scale pilot studies described above, larger sample sizes in the setting of double-blinded, fully randomized design would be the next step in validating the results. Better gender balance, racial diversity, as well as homogeneity in lesion location, and neurologic deficit are also worth consideration. Compared to laboratory studies, clinical trials have not shown significant reductions in primary infarct size. Differences in modes of delivery and dosing regimens beyond the 3 days may help assess the ability of FTY720 to achieve reductions in primary lesion size in humans. A larger phase 2 study looking the efficacy and safety of FTY720 in AIS is currently underway. We anticipate that the results of this trial will provide additional valuable data to determine the role of fingolimod in the treatment of this condition (74).

Regarding the concern of suppressing an already compromised immune system during the subacute stage of stroke, stroke induced immunodepression is a well-characterized phenomenon thought to contribute to the high prevalence and morbidity of common infections during ICU and hospital stay (10, 13). So far, however, no significant increase in spontaneous bacterial infections has been reported in either laboratory studies or clinical trials (75).

Finally, in the laboratory studies described above, some contradictory data have not been fully accounted for. Wei et al. were unable to reduce glutamate or hydrogen peroxide-induced cytotoxicity through administration of FTY720 or S1P, which could argue against the direct neuroprotective effects of FTY720 (41). Also, CCL2, which is elevated in neurons by pretreatment with FTY720 and implicated in the development of an ischemia tolerant phenotype in stroke models, is shown to be deleterious when expressed by reactive astrocytes in MS models (76). These and other discrepancies, which will require future verification, may prove to be critical in fully understanding FTY720 and S1P's mechanism in stroke-related injuries.

# REFERENCES

- Kappos L, Radue EW, O'Connor P, Polman C, Hohlfeld R, Calabresi P, et al. A placebo-controlled trial of oral fingolimod in relapsing multiple sclerosis. N Engl J Med (2010) 362(5):387–401. doi:10.1056/NEJM oa0909494
- Chun J, Hartung H-P. Mechanism of action of oral fingolimod (FTY720) in multiple sclerosis. *Clin Neuropharmacol* (2011) 33(2):91–101. doi:10.1097/ WNF.0b013e3181cbf825
- Fu Y, Zhang N, Ren L, Yan Y, Sun N, Li YJ, et al. Impact of an immune modulator fingolimod on acute ischemic stroke. *Proc Natl Acad Sci U S A* (2014) 111(51):18315–20. doi:10.1073/pnas.1416166111

# CONCLUSION

We are just now beginning to understand the role that FTY720 plays in stroke recovery. Taken together, current laboratory and clinical data suggest a beneficial role of FTY720 in preventing secondary progression of tissue injury and promoting short- and long-term rehabilitation, both in AIS with and without tPA, and in ICH. There is also growing evidence supporting the protective effect of fingolimod in subarachnoid hemorrhage (77). Although the exact mechanism is still elusive, that these results are at least in part due to the peripheral sequestration of lymphocytes has been largely accepted. Evidence also exists for other forms of immunomodulation through attenuating reactive astrocytes, vasoprotection (both immune-related and direct), and direct neuroprotection through surface and intracellular S1P receptors.

The sphingolipid signaling pathway modulated by fingolimod have a ubiquitous presence in the CNS, affecting various disease states from Parkinsonism to epilepsy and brain tumors (6, 78). The data presented in this review are predominantly gathered from stroke-related research but includes MS models as well. It is likely that as progress is made in uncovering FTY720 and S1P's role in other diseases, mechanisms pertaining to stroke will also come to light. In the meantime, we appear to have a drug that continues to show promise in ameliorating the effects of stroke and one that may change the future prognosis and management of cerebrovascular diseases.

# **AUTHOR CONTRIBUTIONS**

Dr. WL, HX, and FT participated in the data analysis and interpretation as well as the conception, drafting, and correction of the manuscript. Dr. HX and FT, in addition, participated in data collection. All the authors have revised the manuscript, approved its final version, and confirm the accuracy and integrity of the data included.

# ACKNOWLEDGMENTS

The authors dedicate this manuscript to the memory of Professor Dale Pelligrino who was a dedicated neuroscience scientist, encouraging mentor, and good friend.

# FUNDING

This study was funded by the University of Illinois recruitment package for FDT.

- Zhu Z, Fu Y, Tian D, Sun N, Han W, Chang G, et al. Combination of the immune modulator fingolimod with alteplase in acute ischemic stroke: a pilot trial. *Circulation* (2015) 132(12):1104–12. doi:10.1161/CIRCULATIONAHA.115.016371
- Fu Y, Hao J, Zhang N, Ren L, Sun N, Li YJ, et al. Fingolimod for the treatment of intracerebral hemorrhage: a 2-arm proof-of-concept study. *JAMA Neurol* (2014) 71(9):1–10. doi:10.1001/jamaneurol.2014.1065
- Brunkhorst R, Vutukuri R, Pfeilschifter W. Fingolimod for the treatment of neurological diseases-state of play and future perspectives. *Front Cell Neurosci* (2014) 8:283. doi:10.3389/fncel.2014.00283
- Hunter SF, Bowen JD, Reder AT. The direct effects of fingolimod in the central nervous system: implications for relapsing multiple sclerosis. CNS Drugs (2016) 30(2):135–47. doi:10.1007/s40263-015-0297-0

- Chaturvedi M, Kaczmarek L. MMP-9 inhibition: a therapeutic strategy in ischemic stroke. *Mol Neurobiol* (2014) 49(1):563–73. doi:10.1007/s12035-013-8538-z
- Iadecola C, Anrather J. The immunology of stroke: from mechanisms to translation. Nat Med (2011) 17(7):796–808. doi:10.1038/nm.2399
- Chamorro Á, Meisel A, Planas AM, Urra X, van de Beek D, Veltkamp R. The immunology of acute stroke. *Nat Rev Neurol* (2012) 8(7):401–10. doi:10.1038/ nrneurol.2012.98
- Yilmaz G, Arumugam TV, Stokes KY, Granger DN. Role of T lymphocytes and interferon-gamma in ischemic stroke. *Circulation* (2006) 113(17):2105–12. doi:10.1161/CIRCULATIONAHA.105.593046
- Planas AM, Gómez-Choco M, Urra X, Gorina R, Caballero M, Chamorro Á. Brain-derived antigens in lymphoid tissue of patients with acute stroke. *J Immunol* (2012) 188(5):2156–63. doi:10.4049/jimmunol.1102289
- Kamel H, Iadecola C. Brain-immune interactions and ischemic stroke: clinical implications. Arch Neurol (2012) 69(5):576–81. doi:10.1001/archneurol. 2011.3590
- Ziv Y, Ron N, Butovsky O, Landa G, Sudai E, Greenberg N, et al. Immune cells contribute to the maintenance of neurogenesis and spatial learning abilities in adulthood. *Nat Neurosci* (2006) 9(2):268–75. doi:10.1038/nn1629
- Urra X, Villamor N, Amaro S, Gómez-Choco M, Obach V, Oleaga L, et al. Monocyte subtypes predict clinical course and prognosis in human stroke. J Cereb Blood Flow Metab (2009) 29(5):994–1002. doi:10.1038/jcbfm. 2009.25
- Cervera A, Planas AM, Justicia C, Urra X, Jensenius JC, Torres F, et al. Genetically-defined deficiency of mannose-binding lectin is associated with protection after experimental stroke in mice and outcome in human stroke. *PLoS One* (2010) 5(2):e8433. doi:10.1371/journal.pone.0008433
- Liesz A, Zhou W, Mracskó É, Karcher S, Bauer H, Schwarting S, et al. Inhibition of lymphocyte trafficking shields the brain against deleterious neuroinflammation after stroke. *Brain* (2011) 134(3):704–20. doi:10.1093/brain/ awr008
- Konoeda F, Shichita T, Yoshida H, Sugiyama Y, Muto G, Hasegawa E, et al. Therapeutic effect of IL-12/23 and their signaling pathway blockade on brain ischemia model. *Biochem Biophys Res Commun* (2010) 402(3):500–6. doi:10.1016/j.bbrc.2010.10.058
- Ooboshi H, Ibayashi S, Shichita T, Kumai Y, Takada J, Ago T, et al. Postischemic gene transfer of interleukin-10 protects against both focal and global brain ischemia. *Circulation* (2005) 111(7):913–9. doi:10.1161/01. CIR.0000155622.68580.DC
- Ren X, Akiyoshi K, Dziennis S, Vandenbark AA, Herson PS, Hurn PD, et al. Regulatory B cells limit CNS inflammation and neurologic deficits in murine experimental stroke. *J Neurosci* (2011) 31(23):8556–63. doi:10.1523/ JNEUROSCI.1623-11.2011
- Fujita T, Yoneta M, Hirose R, Sasaki S, Inoue K, Kiuchi M, et al. Simple compounds, 2-alkyl-2-amino-1,3-propanediols have potent immunosuppressive activity. *Bioorg Med Chem Lett* (1995) 5(8):847–52. doi:10.1016/ 0960-894X(95)00126-E
- Martin M, Mory C, Prescher A, Wittekind C, Fiedler M, Uhlmann D. Protective effects of early CD4(+) T cell reduction in hepatic ischemia/reperfusion injury. J Gastrointest Surg (2010) 14(3):511–9. doi:10.1007/s11605-009-1104-3
- Delbridge MS, Shrestha BM, Raftery AT, El Nahas AM, Haylor JL. Reduction of ischemia-reperfusion injury in the rat kidney by FTY720, a synthetic derivative of sphingosine. *Transplantation* (2007) 84(2):187–95. doi:10.1097/01. tp.0000269794.74990.da
- Tamagnan G, Tavares A, Barret O, Alagille D, Seibyl J, Marek K, et al. Brain distribution of BZM055, an analog of fingolimod (FTY720), in human. *Mult Scler* (2012) 18(4):379.
- Paugh SW, Payne SG, Barbour SE, Milstien S, Spiegel S. The immunosuppressant FTY720 is phosphorylated by sphingosine kinase type 2. *FEBS Lett* (2003) 554(1–2):189–93. doi:10.1016/S0014-5793(03)01168-2
- Brinkmann V, Davis MD, Heise CE, Albert R, Cottens S, Hof R, et al. The immune modulator FTY720 targets sphingosine 1-phosphate receptors. *J Biol Chem* (2002) 277(24):21453–7. doi:10.1074/jbc.C200176200
- Kovarik JM, Hsu CH, Skerjanec A, Riviere GJ, Schmouder R. FTY720 disposition is not affected by demographic or clinical factors in de novo kidney transplant patients [abstract no:705]. *Am J Transplant* (2003) 3(Suppl 5):332–3.

- Soliven B, Miron V, Chun J. The neurobiology of sphingosine 1-phosphate signaling and sphingosine 1-phosphate receptor modulators. *Neurology* (2011) 76(8 Suppl 3):S9–14. doi:10.1212/WNL.0b013e31820d9507
- Aktas O, Küry P, Kieseier B, Hartung H-P. Fingolimod is a potential novel therapy for multiple sclerosis. *Nat Rev Neurol* (2010) 6(7):373–82. doi:10.1038/ nrneurol.2010.76
- Xu M, Waters CL, Hu C, Wysolmerski RB, Vincent PA, Minnear FL. Sphingosine 1-phosphate rapidly increases endothelial barrier function independently of VE-cadherin but requires cell spreading and Rho kinase. *Am J Physiol Cell Physiol* (2007) 293(4):C1309–18. doi:10.1152/ajpcell. 00014.2007
- Nussbaum C, Bannenberg S, Keul P, Gräler MH, Gonçalves-de-Albuquerque CF, Korhonen H, et al. Sphingosine-1-phosphate receptor 3 promotes leukocyte rolling by mobilizing endothelial P-selectin. *Nat Commun* (2015) 6:6416. doi:10.1038/ncomms7416
- 32. Fukuhara S, Simmons S, Kawamura S, Inoue A, Orba Y, Tokudome T, et al. The sphingosine-1-phosphate transporter Spns2 expressed on endothelial cells regulates lymphocyte trafficking in mice. *J Clin Invest* (2012) 122(4):1416–26. doi:10.1172/JCI60746
- Hannun YA, Obeid LM. Principles of bioactive lipid signalling: lessons from sphingolipids. Nat Rev Mol Cell Biol (2008) 9(2):139–50. doi:10.1038/nrm2329
- Hisano Y, Kobayashi N, Kawahara A, Yamaguchi A, Nishi T. The sphingosine 1-phosphate transporter, SPNS2, functions as a transporter of the phosphorylated form of the immunomodulating agent FTY720. *J Biol Chem* (2011) 286(3):1758–66. doi:10.1074/jbc.M110.171116
- Matloubian M, Lo CG, Cinamon G, Lesneski MJ, Xu Y, Brinkmann V, et al. Lymphocyte egress from thymus and peripheral lymphoid organs is dependent on S1P receptor 1. *Nature* (2004) 427(6972):355–60. doi:10.1038/ nature02284
- Thangada S, Khanna KM, Blaho VA, Oo ML, Im DS, Guo C, et al. Cell-surface residence of sphingosine 1-phosphate receptor 1 on lymphocytes determines lymphocyte egress kinetics. *J Exp Med* (2010) 207(7):1475–83. doi:10.1084/ jem.20091343
- Pham THM, Okada T, Matloubian M, Lo CG, Cyster JG. S1P1 receptor signaling overrides retention mediated by G alpha I-coupled receptors to promote T cell egress. *Immunity* (2008) 28(1):122–33. doi:10.1016/j.immuni.2007. 11.017
- Liesz A, Sun L, Zhou W, Schwarting S, Mracsko E, Zorn M, et al. FTY720 reduces post-ischemic brain lymphocyte influx but does not improve outcome in permanent murine cerebral ischemia. *PLoS One* (2011) 6(6):e21312. doi:10.1371/journal.pone.0021312
- Kraft P, Göb E, Schuhmann MK, Göbel K, Deppermann C, Thielmann I, et al. FTY720 ameliorates acute ischemic stroke in mice by reducing thrombo-inflammation but not by direct neuroprotection. *Stroke* (2013) 44(11):3202–10. doi:10.1161/STROKEAHA.113.002880
- Czech B, Pfeilschifter W, Mazaheri-Omrani N, Strobel MA, Kahles T, Neumann-Haefelin T, et al. The immunomodulatory sphingosine 1-phosphate analog FTY720 reduces lesion size and improves neurological outcome in a mouse model of cerebral ischemia. *Biochem Biophys Res Commun* (2009) 389(2):251–6. doi:10.1016/j.bbrc.2009.08.142
- Wei Y, Yemisci M, Kim HH, Yung LM, Shin HK, Hwang SK, et al. Fingolimod provides long-term protection in rodent models of cerebral ischemia. *Ann Neurol* (2011) 69(1):119–29. doi:10.1002/ana.22186
- Barreto G, White RE, Ouyang Y, Xu L, Giffard RG. Astrocytes: targets for neuroprotection in stroke. *Cent Nerv Syst Agents Med Chem* (2011) 11(2):164–73. doi:10.2174/187152411796011303
- Hoffmann FS, Hofereiter J, Rübsamen H, Melms J, Schwarz S, Faber H, et al. Fingolimod induces neuroprotective factors in human astrocytes. *J Neuroinflammation* (2015) 12:184. doi:10.1186/s12974-015-0393-6
- 44. van Doorn R, Nijland PG, Dekker N, Witte ME, Lopes-Pinheiro MA, van het Hof B, et al. Fingolimod attenuates ceramide-induced blood-brain barrier dysfunction in multiple sclerosis by targeting reactive astrocytes. Acta Neuropathol (2012) 124(3):397–410. doi:10.1007/s00401-012-1014-4
- Watcharotayangul J, Mao L, Xu H, Vetri F, Baughman VL, Paisansathan C, et al. Post-ischemic vascular adhesion protein-1 inhibition provides neuroprotection in a rat temporary middle cerebral artery occlusion model. *J Neurochem* (2012) 123(Suppl):116–24. doi:10.1111/j.1471-4159.2012.07950.x
- 46. Xu H-L, Baughman VL, Pelligrino DA. Estrogen replacement treatment in diabetic ovariectomized female rats potentiates postischemic leukocyte

adhesion in cerebral venules. Stroke (2004) 35(8):1974-8. doi:10.1161/01. STR.0000135016.24349.9F

- 47. Spampinato SF, Obermeier B, Cotleur A, Love A, Takeshita Y, Sano Y, et al. Sphingosine 1 phosphate at the blood brain barrier: can the modulation of S1P receptor 1 influence the response of endothelial cells and astrocytes to inflammatory stimuli? *PLoS One* (2015) 10(7):e0133392. doi:10.1371/journal. pone.0133392
- Tisato V, Secchiero P, Rimondi E, Gianesini S, Menegatti E, Casciano F, et al. GM-CSF exhibits anti-inflammatory activity on endothelial cells derived from chronic venous disease patients. *Mediators Inflamm* (2013) 2013:561689. doi:10.1155/2013/561689
- Kleinschnitz C, Kraft P, Dreykluft A, Hagedorn I, Göbel K, Schuhmann MK, et al. Regulatory T cells are strong promoters of acute ischemic stroke in mice by inducing dysfunction of the cerebral microvasculature. *Blood* (2013) 121(4):679–91. doi:10.1182/blood-2012-04-426734
- Prager B, Spampinato SF, Ransohoff RM. Sphingosine 1-phosphate signaling at the blood-brain barrier. *Trends Mol Med* (2015) 21(6):354–63. doi:10.1016/j. molmed.2015.03.006
- Camp SM, Bittman R, Chiang ET, Moreno-Vinasco L, Mirzapoiazova T, Sammani S, et al. Synthetic analogs of FTY720 [2-amino-2-(2-[4-octylphenyl]ethyl)-1,3-propanediol] differentially regulate pulmonary vascular permeability in vivo and in vitro. *J Pharmacol Exp Ther* (2009) 331(1):54–64. doi:10.1124/jpet.109.153544
- Sanchez T, Estrada-Hernandez T, Paik JH, Wu MT, Venkataraman K, Brinkmann V, et al. Phosphorylation and action of the immunomodulator FTY720 inhibits vascular endothelial cell growth factor-induced vascular permeability. *J Biol Chem* (2003) 278(47):47281–90. doi:10.1074/jbc. M306896200
- Campos F, Qin T, Castillo J, Seo JH, Arai K, Lo EH, et al. Fingolimod reduces hemorrhagic transformation associated with delayed tissue plasminogen activator treatment in a mouse thromboembolic model. *Stroke* (2013) 44(2):505–11. doi:10.1161/STROKEAHA.112.679043
- Safarian F, Khallaghi B, Ahmadiani A, Dargahi L. Activation of S1P1 receptor regulates PI3K/Akt/FoxO3a pathway in response to oxidative stress in PC12 cells. J Mol Neurosci (2015) 56(1):177–87. doi:10.1007/s12031-014-0478-1
- Hasegawa Y, Suzuki H, Sozen T, Rolland W, Zhang JH. Activation of sphingosine 1-phosphate receptor-1 by FTY720 is neuroprotective after ischemic stroke in rats. *Stroke* (2010) 41(2):368–74. doi:10.1161/STROKEAHA.109. 568899
- Goldlust EJ, Paczynski RP, He YY, Hsu CY, Goldberg MP. Automated measurement of infarct size with scanned images of triphenyltetrazolium chloride-stained rat brains. *Stroke* (1996) 27(9):1657–62. doi:10.1161/01. STR.27.9.1657
- Wacker BK, Perfater JL, Gidday JM. Hypoxic preconditioning induces stroke tolerance in mice via a cascading HIF, sphingosine kinase, and CCL2 signaling pathway. J Neurochem (2012) 123(6):954–62. doi:10.1111/jnc.12047
- Martire A, Fernandez B, Buehler A, Strohm C, Schaper J, Zimmermann R, et al. Cardiac overexpression of monocyte chemoattractant protein-1 in transgenic mice mimics ischemic preconditioning through SAPK/JNK1/2 activation. *Cardiovasc Res* (2003) 57(2):523–34. doi:10.1016/S0008-6363(02)00697-1
- Stowe AM, Wacker BK, Cravens PD, Perfater JL, Li MK, Hu R, et al. CCL2 upregulation triggers hypoxic preconditioning-induced protection from stroke. J Neuroinflammation (2012) 9(1):33. doi:10.1186/1742-2094-9-33
- Pfeilschifter W, Czech-Zechmeister B, Sujak M, Mirceska A, Koch A, Rami A, et al. Activation of sphingosine kinase 2 is an endogenous protective mechanism in cerebral ischemia. *Biochem Biophys Res Commun* (2011) 413(2):212–7. doi:10.1016/j.bbrc.2011.08.070
- Hait NC, Allegood J, Maceyka M, Strub GM, Harikumar KB, Singh SK, et al. Regulation of histone acetylation in the nucleus by sphingosine-1-phosphate. *Science* (2009) 325(5945):1254–7. doi:10.1126/science.1176709
- Hait NC, Wise LE, Allegood JC, O'Brien M, Avni D, Reeves TM, et al. Active, phosphorylated fingolimod inhibits histone deacetylases and facilitates fear extinction memory. *Nat Neurosci* (2014) 17(7):971–80. doi:10.1038/ nn.3728
- Asle-Rousta M, Kolahdooz Z, Oryan S, Ahmadiani A, Dargahi L. FTY720 (fingolimod) attenuates beta-amyloid peptide (Aβ42)-induced impairment of spatial learning and memory in rats. *J Mol Neurosci* (2013) 50(3):524–32. doi:10.1007/s12031-013-9979-6

- Omidbakhsh R, Rajabli B, Nasoohi S, Khallaghi B, Mohamed Z, Naidu M, et al. Fingolimod affects gene expression profile associated with LPS-induced memory impairment. *Exp Brain Res* (2014) 232(11):3687–96. doi:10.1007/ s00221-014-4052-4
- Hemmati F, Dargahi L, Nasoohi S, Omidbakhsh R, Mohamed Z, Chik Z, et al. Neurorestorative effect of FTY720 in a rat model of Alzheimer's disease: comparison with memantine. *Behav Brain Res* (2013) 252:415–21. doi:10.1016/j. bbr.2013.06.016
- Nazari M, Keshavarz S, Rafati A, Namavar MR, Haghani M. Fingolimod (FTY720) improves hippocampal synaptic plasticity and memory deficit in rats following focal cerebral ischemia. *Brain Res Bull* (2016) 124:95–102. doi:10.1016/j.brainresbull.2016.04.004
- Willis MA, Cohen JA. Fingolimod therapy for multiple sclerosis. Semin Neurol (2013) 33(1):37–44. doi:10.1055/s-0033-1343794
- Adeoye O, Broderick JP. Advances in the management of intracerebral hemorrhage. Nat Rev Neurol (2010) 6(11):593–601. doi:10.1038/nrneurol.2010.146
- Rolland WB II, Manaenko A, Lekic T, Hasegawa Y, Ostrowski R, Tang J, et al. FTY720 is neuroprotective and improves functional outcomes after intracerebral hemorrhage in mice. *Acta Neurochir Suppl* (2011) 111:213–7. doi:10.1007/978-3-7091-0693-8\_36
- Rolland WB, Lekic T, Krafft PR, Hasegawa Y, Altay O, Hartman R, et al. Fingolimod reduces cerebral lymphocyte infiltration in experimental models of rodent intracerebral hemorrhage. *Exp Neurol* (2013) 241(1):45–55. doi:10.1016/j.expneurol.2012.12.009
- Lu L, Barfejani AH, Qin T, Dong Q, Ayata C, Waeber C. Fingolimod exerts neuroprotective effects in a mouse model of intracerebral hemorrhage. *Brain Res* (2014) 1555:89–96. doi:10.1016/j.brainres.2014.01.048
- 72. Foster CA, Mechtcheriakova D, Storch MK, Balatoni B, Howard LM, Bornancin F, et al. FTY720 rescue therapy in the dark agouti rat model of experimental autoimmune encephalomyelitis: expression of central nervous system genes and reversal of blood-brain-barrier damage. *Brain Pathol* (2009) 19(2):254–66. doi:10.1111/j.1750-3639.2008.00182.x
- Finney CA, Hawkes CA, Kain DC, Dhabangi A, Musoke C, Cserti-Gazdewich C, et al. S1P is associated with protection in human and experimental cerebral malaria. *Mol Med* (2011) 17(7–8):717–25. doi:10.2119/molmed.2010.00214
- 74. Shi F-D. *Efficacy and Safety of FTY720 for Acute Stroke*. (2016). Available from: https://clinicaltrials.gov/ct2/show/study/NCT02002390#contacts
- Pfeilschifter W, Czech-Zechmeister B, Sujak M, Foerch C, Wichelhaus TA, Pfeilschifter J. Treatment with the immunomodulator FTY720 does not promote spontaneous bacterial infections after experimental stroke in mice. *Exp Transl Stroke Med* (2011) 3:2. doi:10.1186/2040-7378-3-2
- Van Doorn R, Van Horssen J, Verzijl D, Witte M, Ronken E, Van Het Hof B, et al. Sphingosine 1-phosphate receptor 1 and 3 are upregulated in multiple sclerosis lesions. *Glia* (2010) 58(12):1465–76. doi:10.1002/glia.21021
- Xu H-L, Pelligrino DA, Paisansathan C, Testai FD. Protective role of fingolimod (FTY720) in rats subjected to subarachnoid hemorrhage. *J Neuroinflammation* (2015) 12(1):16. doi:10.1186/s12974-015-0234-7
- Sivasubramanian M, Kanagaraj N, Dheen ST, Tay SSW. Sphingosine kinase 2 and sphingosine-1-phosphate promotes mitochondrial function in dopaminergic neurons of mouse model of Parkinson's disease and in MPP+treated MN9D cells in vitro. *Neuroscience* (2015) 290:636–48. doi:10.1016/j. neuroscience.2015.01.032

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer AP and handling editor declared their shared affiliation, and the handling editor states that the process nevertheless met the standards of a fair and objective review.

Copyright © 2016 Li, Xu and Testai. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.