Check for updates

OPEN ACCESS

EDITED BY Yi Li, Stanford University, United States

REVIEWED BY Kapil Gururangan, University of California, Los Angeles, United States Erfan Bahramnejad, University of Arizona, United States

*CORRESPONDENCE Marios Kaliakatsos Imarios.kaliakatsos@gosh.nhs.uk Sukhvir K. Wright Image: s.wright5@aston.ac.uk

RECEIVED 14 May 2024 ACCEPTED 29 July 2024 PUBLISHED 08 August 2024

CITATION

Champsas D, Zhang X, Rosch R, Ioannidou E, Gilmour K, Cooray G, Woodhall G, Pujar S, Kaliakatsos M and Wright SK (2024) NORSE/ FIRES: how can we advance our understanding of this devastating condition? *Front. Neurol.* 15:1426051. doi: 10.3389/fneur.2024.1426051

COPYRIGHT

© 2024 Champsas, Zhang, Rosch, Ioannidou, Gilmour, Cooray, Woodhall, Pujar, Kaliakatsos and Wright. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

NORSE/FIRES: how can we advance our understanding of this devastating condition?

Dimitrios Champsas^{1,2}, Xushuo Zhang², Richard Rosch^{3,4}, Evangelia Ioannidou¹, Kimberly Gilmour^{5,6,7}, Gerald Cooray^{8,9}, Gavin Woodhall², Suresh Pujar^{1,7}, Marios Kaliakatsos^{1,7*} and Sukhvir K. Wright^{2,10*}

¹Department of Neurology, Great Ormond Street Hospital (GOSH), London, United Kingdom, ²Institute of Health and Neurodevelopment, School of Health and Life Sciences, Aston University, Birmingham, United Kingdom, ³Department of Clinical Neurophysiology, King's College Hospital London NHS Foundation Trust, London, United Kingdom, ⁴Departments of Neurology and Pediatrics, Columbia University, New York, NY, United States, ⁵Department of Immunology, Great Ormond Street Hospital (GOSH), London, United Kingdom, ⁶Biomedical Research Centre (BRC), London, United Kingdom, ⁷Institute of Child Health, University College London, London, United Kingdom, ⁸Department of Neurophysiology, Great Ormond Street Hospital (GOSH), London, United Kingdom, ⁹Clinical Neuroscience, Karolinska Institutet, Stockholm, Sweden, ¹⁰Birmingham Women's and Children's Hospital NHS Trust, Birmingham, United Kingdom

Introduction: New onset refractory status epilepticus (NORSE) is a rare and devastating condition characterised by the sudden onset of refractory status epilepticus (RSE) without an identifiable acute or active structural, toxic, or metabolic cause in an individual without a pre-existing diagnosis of epilepsy. Febrile infection-related epilepsy syndrome (FIRES) is considered a subcategory of NORSE and presents following a febrile illness prior to seizure onset. NORSE/FIRES is associated with high morbidity and mortality in children and adults.

Methods and results: In this review we first briefly summarise the reported clinical, paraclinical, treatment and outcome data in the literature. We then report on existing knowledge of the underlying pathophysiology in relation to *in vitro* and *in vivo* pre-clinical seizure and epilepsy models of potential relevance to NORSE/FIRES.

Discussion: We highlight how pre-clinical models can enhance our understanding of FIRES/NORSE and propose future directions for research.

KEYWORDS

NORSE, FIRES, status epilepticus, immunomodulation, autoantibodies, animal models

Introduction

The term NORSE (New Onset Refractory Status Epilepticus) (1) was first used in 2005 to describe 7 cases (females, mean age 33 yrs., range 20–52 yrs) of status epilepticus (SE) refractory to standard treatment protocols with a poor outcome (5 died, 2 were in vegetative state) in whom a previous history of epilepsy or underlying cause was not identified. Five year later, "FIRES" (Febrile Infection-Related Epilepsy Syndrome) emerged as a clinical entity describing a case-series of 22 previously typically developing children (3–15 yrs) with prolonged or recurrent seizures occurring ~5 days after fever onset (2, 3). In 2018, an expert group released a Consensus definition of New-Onset Refractory Status Epilepticus (NORSE) (4) as a clinical presentation, not a specific diagnosis, in a patient without active epilepsy or other pre-existing relevant neurological disorder, with new onset of refractory status epilepticus (RSE) without a clear acute or

active structural, toxic, or metabolic cause but including patients with viral or autoimmune causes. FIRES was designated a subtype of NORSE where, following a minor febrile illness, seizures increase rapidly (100 s/day) and worsen to SRSE (4). Previously the term FIRES was used exclusively in children, however, it is now accepted that children can have NORSE and adults can have FIRES.

Despite progress in the clarification of NORSE/FIRES as a clinical syndrome, understanding of the underlying pathogenic mechanisms is limited. A cryptogenic cause accounts for up to 50% (5) of cases with these cases reported to have worst outcomes (6).

Treatment of FIRES/NORSE is challenging, and traditional antiseizure medicines (ASMs) used in status epilepticus often prove ineffective against the unrelenting ictal activity (7). All the available evidence for NORSE/FIRES treatment is from case-series, there have been no systematic clinical trials to date. As positive results from therapies are more likely to be published, the bias in treatment responsiveness needs acknowledgement. A Delphi expert consensus on a suggested treatment algorithm was recently published (2) stating initial treatment of SE and RSE with ASMs and anesthetic drugs as per published and local guidelines, and management of possible infections alongside diagnostic work-up and treatment for etiology if identified. If there is incomplete response then initiation of first-line immunological treatment (steroids, IVIG, PLEX) within the first 72 h of seizures is recommended. If ongoing unresponsiveness to treatment, children (and adults if possible) can be started on the ketogenic diet, followed by second-line immunological treatment [including IL-1Ra and IL-6 antagonists (if cryptogenic), and Rituximab (if neuronal autoantibody identified or autoimmune encephalitis suspected)]. Other treatments published in case reports and small case series include intrathecal dexamethasone (8, 9), cannabidiol (10, 11), and Janus kinase (JAK) inhibitors (12). Further studies are needed to determine the effectiveness of these treatments.

There is increasing interest in non-pharmacological treatment of SE in NORSE/FIRES, including Vagal Nerve Stimulation (VNS), deep brain stimulation (DBS) and electroconvulsive therapy (ECT). In a recent systematic review (13, 14), 20 patients with NORSE/FIRES treated with neuromodulation were identified. VNS was used in seven patients with cessation of SE in five, although two died. DBS was implanted in four patients (in one VNS first and then DBS), all achieving a good outcome for SE. ECT was performed in 10 patients, of which three were under 4-years-old with a diagnosis of FIRES (13). In 9 out 10 patients SE resolved after 4–8 sessions. Of the 17 patients that survived, 11 had continuing epilepsy, 12 had cognitive and/or motor dysfunction and one patient remained in a vegetative state. In rare cases, dependent on etiology, epilepsy surgery including focal resection may be indicated (15, 16).

Outcomes remain frustratingly poor in children and adults with NORSE/FIRES. Mortality is approximately 12% in children and higher in adults (up to 30%) (5, 6, 17). In a recent prospective study, only 25% of children with FIRES recovered baseline functions compared to 89% of patients with refractory status epilepticus (18). Another long-term follow-up study of 48 adult patients found only 4% of patients made a full neurological recovery (19). In terms of seizure outcomes a recent systematic review (including 280 adult and 587 pediatric cases) found refractory epilepsy persisted in 40% of adults and nearly 60% of children (20). Long-term cognitive data was available in approximately 70% of pediatric cases and up to one third were described as moderately or severely disabled. Overall, the

majority of patients did not return to a pre-morbid level of functioning (20).

There is an urgent need to develop pre-clinical models to understand the pathogenesis, and test effectiveness of specific therapies to improve outcomes in this catastrophic and life-changing condition.

In this review, we examine the potential relevance of existing pre-clinical seizure/epilepsy models and SE study approaches to NORSE/FIRES pathogenesis (Figure 1; Table 1) and propose future research directions.

Animal models of SE

In patients, when seizures in SE fails to respond to intravenous antiseizure medications (ASMs) it is termed refractory SE; superrefractory SE (SRSE), as seen in NORSE/FIRES, is defined when SE continues after 24h or more of anesthetic treatment (48). Frequent EEG analysis is of paramount importance for treatment decisions and prognostication.

When considering animal models of SE, the onset of SE, severity and length varies according to animal strain, sex, age, species, and method of SE induction (49, 50) (Table 1). Quantification of SE severity is primarily based on behavioral assessments using established severity scales [e.g., the Racine scale (51)] not always reflecting underlying ictal EEG changes which are less frequently recorded – in one model it was found that 50% of behavioral seizures did not correspond with underlying epileptiform EEG activity (52–54).

EEG findings in patients with NORSE/FIRES often reveal a pattern of continuous and diffuse slowing, along with focal, multifocal or generalized epileptiform discharges (4, 17). Continuous EEG findings have also been reported (55) and identified common pathognomonic elements. In 6/7 patients a specific seizure pattern of focal faster waveforms, characterized as "sparks," followed by a gradual appearance of well-formed rhythmic spike/spike and wave complexes was seen. "Ictal shifting" (shifting seizures with contralateral spread) was also described in 4 out of 7 patients (55). Given the significance of EEG in defining and mapping the evolution of SE and NORSE/FIRES, future animal models will need to provide this quantifiable data to ensure validity.

Recent work on EEG abnormalities in immune-mediated seizures has shown that computational models can explain observed EEG abnormalities and help identify putative pathophysiological mechanisms (56, 57). Dynamic Causal Modelling (DCM) of EEG data is an example of this computational approach that allows researchers to model the connectivity and interactions between brain regions, linking the physiology of integrated cortical circuits and their underlying synaptic constraints. DCM uses state-of-the-art Bayesian model inversion techniques to create in silico models that can be used to interrogate synaptic parameters that underlie specific dynamic brain states (58). In recent studies (56, 59), patient EEG recordings, in vivo animal experimental data, and in silico computational models have been combined to provide novel insights into the pathophysiology of immune-mediated epileptogenicity. In the context of NORSE/ FIRES, such computational EEG approaches combining relevant pre-clinical model and patient data could be instrumental in identifying microscopic changes in the cortex that initiate seizure resistance and persistence phenotypical of NORSE/FIRES.

In NORSE/FIRES patients the most common neuropathology findings include neuronal loss, microglial activation and reactive



gliosis (60, 61); 90% of children show brain abnormalities in longterm follow-up MRI scans, most commonly brain atrophy (74%, generalized in 58%) (62, 63). Given that many chemically or electrically-induced SE animal models go on to develop similar neuropathological changes following SE induction alongside chronic drug-resistant epilepsy and cognitive deficits, studying these longterm effects of NORSE/FIRES may be the most promising and relevant use of these models (64–66).

Animal models of immune-mediated epilepsy and seizures

Serum and CSF studies for neuronal autoantibodies are the most commonly performed diagnostic examination after infection work-up in NORSE/FIRES cases (120/907; 61%) (32). Antibodies identified include anti-MOG (myelin oligodendrocyte protein), anti-NMDAR (N-methyl-D-aspartate receptor) antibodies, paraneoplastic and non-paraneoplastic (17, 32, 67–69). In adults with symptomatic NORSE autoimmune encephalitis is the most commonly identified cause (32). Previous studies in autoimmune encephalitis models have successfully used CSF and other human-derived samples to recapitulate core features of the disease including behavioral change, cognitive deficits and seizures (31, 70, 71). More recently, rodent models of immune-mediated seizures in the context of autoimmune encephalitis have also been developed proving the direct epileptogenicity of antigen-specific antibodies (e.g., to the NMDAR, GABA_AR (gamma-aminobutyric acid receptor) and LGI1 (leucinerich glioma inactivated 1) protein) (31, 35, 70). Unlike NORSE/FIRES, status epilepticus was not frequently seen and seizures resolve after completion of intracerebroventricular antibody infusion. This passive transfer approach may be challenging for establishing causality in NORSE/FIRES with no universal specific biomarker (72). However, if seizures were to occur on intracerebral application of CSF from NORSE/FIRES patients this may support the hypothesis of epileptogenic molecules being present in the human-derived samples, particularly as in a recent study pro-inflammatory cytokines/ chemokines [e.g., IL-6 (interleukin-6), TNF-α (tumor necrosis factoralpha), IL-8 (interleukin-8), CCL2 (C-C Motif Chemokine Ligand 2), MIP-1α (macrophage inflammatory protein-1 alpha), and IL-12p70 (interleukin 12p70)] in serum and CSF were found to be significantly increased in patients with SE compared with control patients without SE. In a subset analysis comparing cryptogenic NORSE patients (n=51) and those with a known etiology for refractory SE (n=47), only serum innate immunity pro-inflammatory cytokines/chemokines (CXCL8, CCL2 and MIP-1a) were significantly higher. Elevated innate immunity serum and CSF cytokine/chemokine levels in patients with NORSE were associated with worse outcomes at discharge and at several months after cessation of SE (73). It is important to note that in patients with increased CSF innate immunity

TABLE 1 This table summarizes in vivo and in vitro seizures models, with specification of species, features, and relevance with NORSE and FIRES for
each individual model.

<i>In vivo</i> model	Epilepsy/seizure induction method	Species	Features of the model	Relevance with NORSE/FIRES
Kainic acid – induced status epilepticus (SE) model (21)	Intracranial (intra-amygdala/ intrahippocampal) microinjection	8-week-old C57BL/6 mice	Intra-amygdala more severe SE and higher mortality (44%) than intrahippocampal model. Absent or short latent period from SE to spontaneous recurrence seizures	Prolonged (12–18h) SE leading quickly to spontaneous recurrence seizures (SRS)
Pilocarpine – induced SE rat model (22)	A single acute dose of pilocarpine (380 mg/kg, i.p.) injected to induce SE	Adult male Wistar rats	One of the most prevalent models of human temporal lobe epilepsy, which exhibits physiological, behavioral, electroencephalographic and seizure patterns resembling those of temporal lobe epilepsy	Pilocarpine model has upregulation of CCR2 and C-C motif chemokine ligand 2 (CCL2) within glial and neuronal cells in the hippocampus, while NORSE has elevated levels of IL-6, TNF- α , IL-2, and IL-4 in the CSF, and elevated levels of IL-6 and TNF- α in the periphery (23). The innate immune system neuroinflammation is similar between NORSE/FIRES and pilocarpine model.
The pentylenetetrazol model of epilepsy (24)	Pentylenetetrazol intraperitoneally injected	Adult male Wistar rats	Pentylenetetrazol can be used to develop both acute and chronic models of epilepsy.	Relevant to resistant epilepsy (25)
The rapid kindling protocol (RKP) in rat pups (26)	Electrical stimuli to precipitate seizures with Saline or LPS (50 µg/ kg i.p.)	Wistar rats pups (postnatal D14)	LPS increased the number of severe seizures and the mean duration of the seizures compared to the saline control group	LPS increased hippocampal baseline excitability, this can be reversed by IL-1 receptor antagonist, similar to drug effects in some patients with NORSE/FIRES. Neuroinflammation worsens outcome in this model of SE.
Hyperthermic seizures	Rats: 200 µg/kg LPS, 2.5 h after being placed in a 30°C incubator, \geq 41.5°C induced seizures and \geq 39.0°C was maintained for 30 min; mice: 100 µg/ kg LPS, 2 h before hyperthermic seizures induction (27)	P14 Long Evans rats; P14 C57Bl/6 mice	LPS increases the rodents' susceptibility to hyperthermic seizures; Priming LPS before seizures induction increases cytokine production and microglia activation; LPS also decreases the temperature threshold in seizures induction	Prolonged febrile seizures, cytokine production as seen in NORSE/FIRES.
Acute encephalopathy mouse model (28)	Low-dose LPS (50 or 100 mg/kg) intraperitoneally injected 2 h before hyperthermia treatment (41.5°C, 30 min).	P8 ICR mice of both genders	Exacerbation of BBB disruption; Microglia activation and small ischemic lesions in the cerebral cortexes were observed in some (29)	Displays symptoms of cytokine storm- induced acute encephalopathy.
Pediatric Rat Model of Organophosphate- Induced Refractory Status Epilepticus (30)	DFP (1.4x LD50 mg/kg, s.c.) administered to induce persistent SE. Animals were pretreated with pyridostigmine bromide (0.026 mg/ kg, i.m.) 30 min before DFP injection.	Male Sprague–Dawley rats at postnatal day 21 (P21) (25–40 g).	Persistent refractory-like SE in P21 rats; progressive decline in learning and memory functions; progressive increase in anxiety-like symptoms and disruption in mood-related disturbances; progression of epileptogenesis with recurring spontaneous seizure episodes; widespread neurodegeneration of principal cells and inhibitory interneurons; persistent neuroinflammation with an extensive reactive microglia response; and aberrant mossy fiber sprouting in the hippocampus.	P21 rats are at a stage comparable to 2-to 4-year-old children. Features of behavioral change, neuropathology and inflammation consistent with FIRES/NORSE.

(Continued)

TABLE 1 (Continued)

<i>In vivo</i> model		Species	Features of the model	Relevance with NORSE/FIRES
	induction method			
NMDAR antibody passive transfer rat model (31)	10 mg of NMDAR human monoclonal antibody, or 8 µL of NMDAR antibody or IgG (derived from patient's plasma) via intracerebroventricular injection	Postnatal day 21 (P21) Wistar rats, male, with weights between 50–58 g	NMDAR antibodies cause spontaneous epileptiform activity including the sustained and repetitive hyperexcitable behavior (myoclonic twitches, jerks and jumps)	Identified etiology of NMDAR-Ab encephalitis in FIRES/NORSE patients (32)
IgG from anti- NMDAR encephalitis osmotic pump infusion (33)	Osmotic pump insertion to infuse polyclonal antibodies against the N-terminal domain of human GluN1protein (pooled from anti- NMDAR encephalitis and seizure patient)	male C57BL/6 mice that were 8–10 weeks old	CSF from patients with anti-NMDAR encephalitis induces seizures in mice, with 2% associated with only minimal behavior change	Identified etiology of NMDAR-Ab encephalitis in FIRES/NORSE patients (34)
Intrathecal osmotic pump infusions of GABA _A receptor monoclonal antibodies (mAbs) (35)	Cerebroventricular infusion with $GABA_A$ receptor mAbs, in mice and rats	Male C57BL/6 mice, age:13–14 week; male Wistar rats at postnatal age of 21 days (weighing 50–58 g)	Spontaneous epileptiform activities including detectable ictal events. One died of status epilepticus with higher dose of infused antibody.	Refractive status epilepticus and GABA _A R- Abs found in cases of NORSE/FIRES.
Viral triggered SE	Theiler's murine encephalomyelitis virus injected intracerebrally (temporal region right hemisphere) to recapitulate epilepsy secondary to CNS viral infection	Male/female C57/BL6 mice from age 5-6 weeks	Acute and chronic seizures. Susceptible to handling-induced acute seizures.	Most observable seizures occur during the acute infection period of most relevance to initial phase of NORSE/FIRES, only a small proportion develop chronic spontaneous seizures (36).
Methyl- azoxymethanol- acetate/pilocarpine model (37)	Pregnant rats received 2 doses of methyl-azoxymethanol-acetate 12 h apart, at embryonic day 15. The young adult (2–3 months old litters) received pilocarpine treatment (270 mg/kg i.p.).	Pregnant Sprague– Dawley rats	Severe seizures with mortality in some. Severe CNS neuroinflammation with encephalitis- like features, brain malformation.	This model can be considered a model of autoimmune-associated epilepsy, and show activation of adaptive immunity as seen in NORSE/FIRES (37).
In vitro model	Epilepsy/seizure induction method	Species	Features of the model	Relevance with NORSE/FIRES
Temporal cortex/ hippocampal brain slices with LPS preincubation and/ or 4-AP perfusion (38)	Brain slices preincubated for 30 min with the addition of $10 \mu g/mL$ lipopolysaccharide (LPS) before perfusion for 40 min with aCSF containing zero (0) Mg ²⁺ and 100 μ M 4-aminopyridine (4-AP)	28 to 30-day old male C57BL6/N mice	LPS exacerbates epileptiform activity in slices, reproducing the cytokine storm that precedes seizure precipitation in FIRES, with 4-AP inducing the refractive epileptiform activities	This model mimics the unresponsiveness o epileptiform events to antiseizure medications, as observed in NORSE/FIRES patients
Drganotypic nippocampal slice cultures (39)	Isolated hippocampi culture from 350 μm slices.	P8 Sprague–Dawley rat pups	Progression in epileptiform activity, starting with interictal spike discharges around 14–17 days and transitioning to mostly ictal-like electrographic activity by day 25–30. Ictal events lasted greater than 3 min in duration in 80% of slices	Applicable to epileptogenesis in NORSE/ FIRES (40)
4-aminopyridine (4-AP) model (41)	Rodent brain slices of 400 µm. 4-AP (100 µM) used for inducing seizure like events	Adult, 7–11-week-old, Wistar Han rats (200– 350 g)	The model is generated via 4-AP, a non-selective potassium channel blocker and a potent proconvulsant compound in the limbic system	In vitro resistant seizures
Organotypic hippocampal slices culture and acute slices of temporal cortex (42)	0 Mg ²⁺ exposure for 3 h at 28–30°C to hippocampal and temporal cortex cultures slices	GAD2-cre-tdTomato mice (C57BL/6 background, JAX lab) or Wistar rats; 7-day-old	With extended periods of 0 Mg ²⁺ , distinct ictal events are replaced by recurrent epileptiform discharges.	<i>In vitro</i> status epilepticus which strongly resemble clinical EEG recordings of convulsive status epilepticus.

(Continued)

<i>In vivo</i> model	Epilepsy/seizure induction method	Species	Features of the model	Relevance with NORSE/FIRES
Human brain tissue	None required	6 patients (F:M 9:7),	Spontaneous in vitro epileptiform	Spontaneous epileptiform activity in human
resected from epilepsy surgery patients (43)		median age 10.5 years (range 3–18 years)	activity	tissue from drug-resistant pediatric epilepsy patients; treatment response to KD (44)
Hippocampal slices (44, 45)	Superfusion of the high-K+ (8.5 mM) external solution	Wistar rats (postnatal days 9–19, weights from18 g – 49 g)	Epileptiform discharges in the stratum pyramidale of hippocampal slices	Tonic-firing pattern of spike events allowed reliable quantification of antiepileptic drugs to study focal seizures (46).
Intact cortical hippocampal formation (47)	Low-Mg ²⁺ aCSF	Wistar rats at postnatal days 7 and 8	Spontaneous seizures were synchronized in hippocampal and cortical regions	Recurrent ictal-like events of infantile epilepsy that are resistant to commonly used antiepileptic drugs (47)

TABLE 1 (Continued)

BBB, blood brain barrier; KD, ketogenic diet; LPS, lipopolysaccharide; SE, status epilepticus.

cytokines, serial measurements frequently showed normalization of the values within days, and it is unclear if this is associated with immunomodulatory treatment. Further studies are needed with collection of acute and chronic CSF/serum samples and accurate documentation of immunotherapy and seizure frequency to establish cause and effect with respect to these immune mediators in NORSE/ FIRES (67).

In animal models the activation and role of the innate immune system in the pathogenesis of status epilepticus has been shown in pre-clinical models and recently comprehensively reviewed by Vezzani et al. (74). In brief, there is strong evidence that neuronal cells release pro-inflammatory molecules after SE onset that initiate neuroinflammatory changes further influencing neuronal excitability and excitotoxicity (75). The fact that these changes are seen in normal rodent brains after SE induction is relevant to NORSE/FIRES and encouraging that anti-inflammatory treatments alter the progression to chronic epilepsy. A recently described in vitro model of FIRES using rodent brain slices and lipopolysaccharide (LPS) to induce inflammation demonstrated increased mRNA levels of relevant cytokines and inhibition of ictal activity with two specific immunomodulatory drugs, however the relatively acute time course of SE in vitro before treatment (10 min) and pre-incubation with these drugs may limit the translatability of the results to the clinical scenario (38).

Neuropathological findings in NORSE/FIRES also include perivascular T-cell infiltration, i.e., further involvement of the adaptive immune system beyond neuronal autoantibodies (60). A recent study highlighted the marginal involvement of adaptive immunity in the classical model of pilocarpine-induced epilepsy in normal rats when compared to their MethylAzoxyMethanol (MAM)/pilocarpine (MP) model that display malformed brains, status epilepticus and subsequent spontaneous recurrent seizures (37). This model of CNS autoimmune-associated epilepsy has potential to provide crucial insights into the complexities of the acute and chronic neuroimmunological response in NORSE/FIRES and the role of disease-modifying treatments.

Further work is needed to translate all these pre-clinical findings to NORSE/FIRES patients, specifically: the causal role of immune system mediators, identifying the optimal time in SE for immunodulatory intervention and if this intervention should be targeted to the innate and/or adaptive system.

Genetic epilepsy models

In a recent systematic retrospective analysis of FIRES patients, genetic testing performed in 23 of 25 patients was non-diagnostic. Interestingly, when a broader cohort of 959 individuals with RSE was analyzed, the highest proportion of individuals with genetic RSE had onset within the first 3 months of life, whereas individuals with FIRES typically have onset in later childhood, with an age of onset ranging from 7.6 months to 18.7 years (69). Similar negative findings for a genetic etiology have been reported in previous studies (76, 77).

This lack of a single causative gene in NORSE/FIRES may hinder the development of a genetic epilepsy model but genetic therapy designed to suppress seizure activity, for example by modulating a specific channel (78), may be considered as a therapeutic approach that could be tested in pre-clinical models (79).

Human brain tissue and epilepsy models

Human brain tissue is an invaluable resource that is being increasingly used in neuroscience research of direct relevance to NORSE/FIRES. For example, donated human tissue samples from pediatric epilepsy patients undergoing surgery have been used to demonstrate the therapeutic effectiveness of neurosteroids and decanoic acid, a major medium-chain fatty acid provided in the medium-chain triglyceride ketogenic diet, in *in vitro* electrophysiology studies (31, 43). Recent recommendations to integrate human tissuebased *in vitro* models into pre-clinical studies to develop anti-seizure therapies will benefit NORSE/FIRES research (44).

Using CSF and brain tissue samples, further insights into underlying immunopathology can be gained from single cell genomics as shown in pediatric epilepsy patients. Kumar et al., found in a study using samples from 6 pediatric epilepsy patients a pro-inflammatory microenvironment in the resected tissue, with extensive activation of microglia and infiltration of other pro-inflammatory immune cells (80). Of particular interest was clear demonstration of a direct interaction between T cells and microglia inside epileptic brain tissue similar to multiple sclerosis. One must caveat these findings with the knowledge that not all the transcriptional-level information will be translated to the protein level. Further validation is required before direct translation to management but nevertheless this advanced genomic analysis should be used in NORSE/FIRES to potentially identify relevant subpopulations of aberrant inflammatory and/or neuronal cells to investigate for pathogenicity.

To maximize the opportunities these rare samples can provide for research and increased understanding of NORSE/FIRES disease pathogenesis it is imperative that standardized procedures for specimen collection and biobanking are followed (67).

Figure 1 summarizes the contribution that existing pre-clinical epilepsy and seizure models could make to the understanding of NORSE/FIRES pathophysiology while also highlighting the research gap for a model that recapitulates the full clinical phenotype.

Discussion and call to action

NORSE/FIRES is a devastating clinical syndrome characterized by acute intractable seizures evolving to treatmentresistant super-refractory status epilepticus, and mostly inevitable adverse cognitive impact in the long-term survivors. There is increasing worldwide collaboration in terms of disease definition, treatment guidelines and establishment of biorepositories underpinned by the unwavering support of dedicated affected family member advocates, parents, guardians, and carers. The role of an aberrant immune system response appears to be gaining evidence in terms of underlying pathophysiology and possible treatments. However, most of the published studies, case-reports and case series are comprised of small patient numbers and often retrospective in nature making it difficult to estimate treatment efficacy in the wider population. In this review we have highlighted how we can use existing pre-clinical models and human-derived samples to further NORSE/FIRES understanding as well as identifying promising future research areas. We call for experts from neurology, neurophysiology, computational biology, genetics, neuroscience, and immunology to collaborate and drive these promising areas of research forward to improve outcomes for affected patients.

References

1. Wilder-Smith E, Lim E, Teoh H, Sharma V, Tan J, Chan B, et al. The NORSE (newonset refractory status epilepticus) syndrome: defining a disease entity. *Annals-Academy Med Singapore*. (2005) 34:417.

2. Wickstrom R, Taraschenko O, Dilena R, Payne ET, Specchio N, Nabbout R, et al. International consensus recommendations for management of new onset refractory status epilepticus including febrile infection-related epilepsy syndrome: statements and supporting evidence. *Epilepsia*. (2022) 63:2840–64. doi: 10.1111/epi.17397

3. van Baalen A, Häusler M, Boor R, Rohr A, Sperner J, Kurlemann G, et al. Febrile infection-related epilepsy syndrome (FIRES): a nonencephalitic encephalopathy in childhood. *Epilepsia*. (2010) 51:1323–8. doi: 10.1111/j.1528-1167.2010.02535.x

4. Hirsch LJ, Gaspard N, van Baalen A, Nabbout R, Demeret S, Loddenkemper T, et al. Proposed consensus definitions for new-onset refractory status epilepticus (NORSE), febrile infection-related epilepsy syndrome (FIRES), and related conditions. *Epilepsia*. (2018) 59:739–44. doi: 10.1111/epi.14016

5. Tharmaraja T, Ho JSY, Neligan A, Rajakulendran S. The etiology and mortality of new-onset refractory status epilepticus (NORSE) in adults: a systematic review and meta-analysis. *Epilepsia*. (2023) 64:1113–24. doi: 10.1111/epi.17523

6. Gaspard N, Foreman BP, Alvarez V, Cabrera Kang C, Probasco JC, Jongeling AC, et al. New-onset refractory status epilepticus: etiology, clinical features, and outcome. *Neurology*. (2015) 85:1604–13. doi: 10.1212/WNL.000000000001940

Author contributions

DC: Conceptualization, Writing – original draft, Writing – review & editing. XZ: Writing – original draft, Writing – review & editing. RR: Writing – original draft, Writing – review & editing. EI: Writing – original draft, Writing – review & editing. KG: Writing – original draft, Writing – review & editing. GC: Writing – original draft, Writing – review & editing. GW: Writing – original draft, Writing – review & editing. SP: Writing – original draft, Writing – review & editing. SP: Writing – original draft, Writing – review & editing. SW: Conceptualization, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. SW was funded through Wellcome Trust Fellowship DC and XZ supported by Epilepsy Research Institute Endeavor Project grant.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

7. Lin W-S. The nuances of immunotherapy for NORSE/FIRES. *Epilepsia*. (2022) 63:3212–4. doi: 10.1111/epi.17439

8. Mehta NP, Sawdy R, Maloney K, Overlee B, Johnson RK, Howe CL, et al. Intrathecal dexamethasone in febrile infection-related epilepsy syndrome: a case report. *Neurol. Clin. Practice.* (2023) 13:e200153. doi: 10.1212/CPJ.0000000000200153

9. Horino A, Kuki I, Inoue T, Nukui M, Okazaki S, Kawawaki H, et al. Intrathecal dexamethasone therapy for febrile infection-related epilepsy syndrome. *Ann. Clin. Transl. Neurol.* (2021) 8:645–55. doi: 10.1002/acn3.51308

10. Fetta A, Crotti E, Campostrini E, Bergonzini L, Cesaroni CA, Conti F, et al. Cannabidiol in the acute phase of febrile infection-related epilepsy syndrome (FIRES). *Epilepsia Open.* (2023) 8:685–91. doi: 10.1002/epi4.12740

11. Gofshteyn JS, Wilfong A, Devinsky O, Bluvstein J, Charuta J, Ciliberto MA, et al. Cannabidiol as a potential treatment for febrile infection-related epilepsy syndrome (FIRES) in the acute and chronic phases. *J Child Neurol*. (2017) 32:35–40. doi: 10.1177/0883073816669450

12. Goh Y, Tay SH, Yeo LLL, Rathakrishnan R. Bridging the gap: tailoring an approach to treatment in febrile infection-related epilepsy syndrome. *Neurology*. (2023) 100:1151–5. doi: 10.1212/WNL.000000000207068

13. Stavropoulos I, Pak HL, Alarcon G, Valentin A. Neuromodulation techniques in children with super-refractory status epilepticus. *Brain Sci.* (2023) 13:1527. doi: 10.3390/brainsci13111527

14. Stavropoulos I, Khaw JH, Valentin A. Neuromodulation in new-onset refractory status epilepticus. *Front Neurol.* (2023) 14:1195844. doi: 10.3389/fneur.2023.1195844

15. Mamaril-Davis J, Vessell M, Ball T, Palade A, Shafer C, Aguilar-Salinas P, et al. Combined responsive Neurostimulation and focal resection for super refractory status epilepticus: a systematic review and illustrative case report. *World Neurosurg.* (2022) 167:195–204.e7. doi: 10.1016/j.wneu.2022.07.141

16. Marashly A, Lew S, Koop J. Successful surgical management of new onset refractory status epilepticus (NORSE) presenting with gelastic seizures in a 3 year old girl. *Epilepsy Behav Case Reports*. (2017) 8:18–26. doi: 10.1016/j.ebcr.2017.05.002

17. Kramer U, Chi CS, Lin KL, Specchio N, Sahin M, Olson H, et al. Febrile infectionrelated epilepsy syndrome (FIRES): pathogenesis, treatment, and outcome: a multicenter study on 77 children. *Epilepsia*. (2011) 52:1956–65. doi: 10.1111/j.1528-1167.2011.03250.x

18. Sculier C, Barcia Aguilar C, Gaspard N, Gaínza-Lein M, Sánchez Fernández I, Amengual-Gual M, et al. Clinical presentation of new onset refractory status epilepticus in children (the pSERG cohort). *Epilepsia*. (2021) 62:1629–42. doi: 10.1111/epi.16950

19. Costello DJ, Matthews E, Aurangzeb S, Doran E, Stack J, Wesselingh R, et al. Clinical outcomes among initial survivors of cryptogenic new-onset refractory status epilepsy (NORSE). *Epilepsia*. (2024) 65:1581–8. doi: 10.1111/epi.17950

20. Taraschenko O, Pavuluri S, Schmidt CM, Pulluru YR, Gupta N. Seizure burden and neuropsychological outcomes of new-onset refractory status epilepticus: systematic review. *Front Neurol.* (2023) 14:14. doi: 10.3389/fneur.2023.1095061

21. Welzel L, Schidlitzki A, Twele F, Anjum M, Loscher W. A face-to-face comparison of the intra-amygdala and intrahippocampal kainate mouse models of mesial temporal lobe epilepsy and their utility for testing novel therapies. *Epilepsia*. (2020) 61:157–70. doi: 10.1111/epi.16406

22. Heysieattalab S, Sadeghi L. Dynamic structural neuroplasticity during and after epileptogenesis in a pilocarpine rat model of epilepsy. *Acta Epileptologica*. (2021) 3:37. doi: 10.1186/s42494-020-00037-7

23. Wesselingh R, Butzkueven H, Buzzard K, Tarlinton D, O'Brien TJ, Monif M. Seizures in autoimmune encephalitis: kindling the fire. *Epilepsia*. (2020) 61:1033–44. doi: 10.1111/epi.16515

24. Mahmoudi T, Lorigooini Z, Rafieian-Kopaei M, Arabi M, Rabiei Z, Bijad E, et al. Effect of *Curcuma zedoaria* hydro-alcoholic extract on learning, memory deficits and oxidative damage of brain tissue following seizures induced by pentylenetetrazole in rat. *Behav Brain Funct*. (2020) 16:7. doi: 10.1186/s12993-020-00169-3

25. Singh T, Mishra A, Goel RK. PTZ kindling model for epileptogenesis, refractory epilepsy, and associated comorbidities: relevance and reliability. *Metab Brain Dis.* (2021) 36:1573–90. doi: 10.1007/s11011-021-00823-3

26. Auvin S, Shin D, Mazarati A, Sankar R. Inflammation induced by LPS enhances epileptogenesis in immature rat and may be partially reversed by IL1RA. *Epilepsia*. (2010) 51:34–8. doi: 10.1111/j.1528-1167.2010.02606.x

27. Eun BL, Abraham J, Mlsna L, Kim MJ, Koh S. Lipopolysaccharide potentiates hyperthermia-induced seizures. *Brain Behav.* (2015) 5:e00348. doi: 10.1002/brb3.348

28. Kurata H, Saito K, Kawashima F, Ikenari T, Oguri M, Saito Y, et al. Developing a mouse model of acute encephalopathy using low-dose lipopolysaccharide injection and hyperthermia treatment. *Exp Biol Med (Maywood)*. (2019) 244:743–51. doi: 10.1177/1535370219846497

29. Koh S, Dupuis N, Auvin S. Ketogenic diet and Neuroinflammation. *Epilepsy Res.* (2020) 167:106454. doi: 10.1016/j.eplepsyres.2020.106454

30. Singh T, Ramakrishnan S, Wu X, Reddy DS. A pediatric rat model of organophosphate-induced refractory status epilepticus: characterization of long-term epileptic seizure activity, neurologic dysfunction and neurodegeneration. *J Pharmacol Exp Ther.* (2024) 388:416–31. doi: 10.1124/jpet.123.001794

31. Wright SK, Rosch RE, Wilson MA, Upadhya MA, Dhangar DR, Clarke-Bland C, et al. Multimodal electrophysiological analyses reveal that reduced synaptic excitatory neurotransmission underlies seizures in a model of NMDAR antibody-mediated encephalitis. *Commun. Biol.* (2021) 4:1106. doi: 10.1038/s42003-021-02635-8

32. Lattanzi S, Leitinger M, Rocchi C, Salvemini S, Matricardi S, Brigo F, et al. Unraveling the enigma of new-onset refractory status epilepticus: a systematic review of aetiologies. *Eur J Neurol.* (2022) 29:626–47. doi: 10.1111/ene.15149

33. Taraschenko O, Fox HS, Pittock SJ, Zekeridou A, Gafurova M, Eldridge E, et al. A mouse model of seizures in anti-N-methyl-d-aspartate receptor encephalitis. *Epilepsia*. (2019) 60:452–63. doi: 10.1111/epi.14662

34. Vogrig A, Gigli GL, Nilo A, Pauletto G, Seizures VM. Epilepsy, and NORSE secondary to autoimmune encephalitis: a practical guide for clinicians. *Biomedicines*. (2022) 11:44. doi: 10.3390/biomedicines11010044

35. Kreye J, Wright SK, van Casteren A, Stoffler L, Machule ML, Reincke SM, et al. Encephalitis patient-derived monoclonal GABAA receptor antibodies cause epileptic seizures. *J Exp Med.* (2021) 218. doi: 10.1084/jem.20210012

36. Batot G, Metcalf CS, Bell LA, Pauletti A, Wilcox KS, Bröer S. A model for epilepsy of infectious etiology using theiler's murine encephalomyelitis virus. *JoVE.* (2022) 184:e63673. doi: 10.3791/63673

37. Costanza M, Ciotti A, Consonni A, Cipelletti B, Cattalini A, Cagnoli C, et al. CNS autoimmune response in the MAM/pilocarpine rat model of epileptogenic cortical

malformation. Proc Natl Acad Sci USA. (2024) 121:e2319607121. doi: 10.1073/ pnas.2319607121

38. Cerovic M, Di Nunzio M, Craparotta I, Vezzani A. An *in vitro* model of drugresistant seizures for selecting clinically effective antiseizure medications in febrile infection-related epilepsy syndrome. *Front Neurol.* (2023) 14:1129138. doi: 10.3389/ fneur.2023.1129138

39. Dyhrfjeld-Johnsen J, Berdichevsky Y, Swiercz W, Sabolek H, Staley KJ. Interictal spikes precede ictal discharges in an organotypic hippocampal slice culture model of epileptogenesis. *J Clin Neurophysiol.* (2010) 27:418–24. doi: 10.1097/WNP.0b013e3181fe0709

40. Wong M. Epilepsy in a dish: an *in vitro* model of Epileptogenesis. *Epilepsy Curr*. (2011) 11:153–4. doi: 10.5698/1535-7511-11.5.153

41. Heuzeroth H, Wawra M, Fidzinski P, Dag R, Holtkamp M. The 4-Aminopyridine model of acute Seizures *in vitro* elucidates efficacy of new antiepileptic drugs. *Front Neurosci.* (2019) 13:677. doi: 10.3389/fnins.2019.00677

42. Burman RJ, Selfe JS, Lee JH, van den Berg M, Calin A, Codadu NK, et al. Excitatory GABAergic signalling is associated with benzodiazepine resistance in status epilepticus. *Brain*. (2019) 142:3482–501. doi: 10.1093/brain/awz283

43. Wright SK, Wilson MA, Walsh R, Lo WB, Mundil N, Agrawal S, et al. Abolishing spontaneous epileptiform activity in human brain tissue through AMPA receptor inhibition. *Ann Clin Transl Neurol.* (2020) 7:883–90. doi: 10.1002/acn3.51030

44. Morris G, Avoli M, Bernard C, Connor K, de Curtis M, Dulla CG, et al. Can *in vitro* studies aid in the development and use of antiseizure therapies? A report of the ILAE/AES joint translational task force. *Epilepsia.* (2023) 64:2571–85. doi: 10.1111/epi.17744

45. Stephen F, Traynelis RD. Potassium-induced-spontaneous-electrographicseizures-in-the-rat-hippocampal-slice. J Neurophysiol. (1988) 59:259–76.

46. Taing KD, O'Brien TJ, Williams DA, French CR. Anti-epileptic drug combination efficacy in an *in vitro* seizure model – phenytoin and valproate, lamotrigine and valproate. *PLoS One.* (2017) 12:e0169974. doi: 10.1371/journal.pone.0169974

47. Quilichini PP, Diabira D, Chiron C, Milh M, Ben-Ari Y, Gozlan H. Effects of antiepileptic drugs on refractory seizures in the intact immature corticohippocampal formation *in vitro. Epilepsia.* (2003) 44:1365–74. doi: 10.1046/j.1528-1157.2003.19503.x

48. Dubey D, Kalita J, Misra UK. Status epilepticus: refractory and super-refractory. *Neurol India*. (2017) 65:S12-7. doi: 10.4103/neuroindia.NI_958_16

49. Reddy DS, Kuruba R. Experimental models of status epilepticus and neuronal injury for evaluation of therapeutic interventions. *Int J Mol Sci.* (2013) 14:18284–318. doi: 10.3390/ijms140918284

50. Sharma S, Puttachary S, Thippeswamy A, Kanthasamy AG, Thippeswamy T. Status epilepticus: behavioral and electroencephalography seizure correlates in Kainate experimental models. *Front Neurol.* (2018) 9:7. doi: 10.3389/fneur.2018.00007

51. Racine RJ, Burnham WM, Gartner JG, Levitan D. Rates of motor seizure development in rats subjected to electrical brain stimulation: strain and inter-stimulation interval effects. *Electroencephalogr Clin Neurophysiol.* (1973) 35:553–6. doi: 10.1016/0013-4694(73)90033-3

52. Tse K, Puttachary S, Beamer E, Sills GJ, Thippeswamy T. Advantages of repeated low dose against single high dose of kainate in C57BL/6J mouse model of status epilepticus: behavioral and electroencephalographic studies. *PLoS One*. (2014) 9:e96622. doi: 10.1371/journal.pone.0096622

53. Lehmkuhle MJ, Thomson KE, Scheerlinck P, Pouliot W, Greger B, Dudek FE. A simple quantitative method for analyzing electrographic status epilepticus in rats. *J Neurophysiol.* (2009) 101:1660–70. doi: 10.1152/jn.91062.2008

54. Puttachary S, Sharma S, Tse K, Beamer E, Sexton A, Crutison J, et al. Immediate Epileptogenesis after Kainate-induced status epilepticus in C57BL/6J mice: evidence from long term continuous video-EEG telemetry. *PLoS One.* (2015) 10:e0131705. doi: 10.1371/journal.pone.0131705

55. Farias-Moeller R, Bartolini L, Staso K, Schreiber JM, Carpenter JL. Early ictal and interictal patterns in FIRES: the sparks before the blaze. *Epilepsia*. (2017) 58:1340–8. doi: 10.1111/epi.13801

56. Cooray GK, Sengupta B, Douglas P, Englund M, Wickstrom R, Friston K. Characterising seizures in anti-NMDA-receptor encephalitis with dynamic causal modelling. *NeuroImage*. (2015) 118:508–19. doi: 10.1016/j.neuroimage.2015.05.064

57. Symmonds M, Moran CH, Leite MI, Buckley C, Irani SR, Stephan KE, et al. Ion channels in EEG: isolating channel dysfunction in NMDA receptor antibody encephalitis. *Brain.* (2018) 141:1691–702. doi: 10.1093/brain/awy107

58. Moran R, Pinotsis DA, Friston K. Neural masses and fields in dynamic causal modeling. *Front Comput Neurosci.* (2013) 7:57. doi: 10.3389/fncom.2013.00057

59. Rosch RE, Wright S, Cooray G, Papadopoulou M, Goyal S, Lim M, et al. NMDAreceptor antibodies alter cortical microcircuit dynamics. *Proc Natl Acad Sci.* (2018) 115:E9916–25. doi: 10.1073/pnas.1804846115

60. Hanin A, Cespedes J, Huttner A, Strelnikov D, Gopaul M, DiStasio M, et al. Neuropathology of new-onset refractory status epilepticus (NORSE). *J Neurol.* (2023) 270:3688–702. doi: 10.1007/s00415-023-11726-x

61. Penner J, Hassell J, Brown JR, Mankad K, Storey N, Atkinson L, et al. Translating metagenomics into clinical practice for complex paediatric neurological presentations. *J Infect*. (2023) 87:451–8. doi: 10.1016/j.jinf.2023.08.002

62. Moreno-Brauer D, Häusler M, Kluger G, Hensler J, van Baalen A. Spectrum, evolution, and clinical relationship of magnetic resonance imaging in 31 children with febrile infection-related epilepsy syndrome. *Neuropediatrics*. (2023) 55:009–15. doi: 10.1055/s-0043-1774318

63. Culleton S, Talenti G, Kaliakatsos M, Pujar S, D'Arco F. The spectrum of neuroimaging findings in febrile infection-related epilepsy syndrome (FIRES): a literature review. *Epilepsia*. (2019) 60:585–92. doi: 10.1111/epi.14684

64. Kyriatzis G, Bernard A, Bôle A, Khrestchatisky M, Ferhat L. In the rat Hippocampus, pilocarpine-induced status epilepticus is associated with reactive glia and concomitant increased expression of CD31, PDGFR β , and collagen IV in endothelial cells and Pericytes of the blood-brain barrier. *Int J Mol Sci.* (2024) 25. doi: 10.3390/ ijms25031693

65. Luna-Munguia H, Marquez-Bravo L, Concha L. Longitudinal changes in gray and white matter microstructure during epileptogenesis in pilocarpine-induced epileptic rats. *Seizure*. (2021) 90:130–40. doi: 10.1016/j.seizure.2021.02.011

66. Korotkov A, Mills JD, Gorter JA, van Vliet EA, Aronica E. Systematic review and meta-analysis of differentially expressed miRNAs in experimental and human temporal lobe epilepsy. *Sci Rep.* (2017) 7:11592. doi: 10.1038/s41598-017-11510-8

67. Hanin A, Cespedes J, Pulluru Y, Gopaul M, Aronica E, Decampo D, et al. Review and standard operating procedures for collection of biospecimens and analysis of biomarkers in new onset refractory status epilepticus. *Epilepsia.* (2023) 64:1444–57. doi: 10.1111/epi.17600

68. Li E-C, Zheng Y, Cai M-T, Lai Q-L, Fang G-L, Du B-Q, et al. Seizures and epilepsy in multiple sclerosis, aquaporin 4 antibody-positive neuromyelitis optica spectrum disorder, and myelin oligodendrocyte glycoprotein antibody-associated disease. *Epilepsia*. (2022) 63:2173–91. doi: 10.1111/epi.17315

69. deCampo DXJ, Karlin A, Sullivan KR, Ruggiero SMGP, Ramos M, Abend NS, et al. Investigating the genetic contribution in febrile infection-related epilepsy syndrome and refractory status epilepticus. *Front Neurol.* (2023) 14:1161161. doi: 10.3389/ fneur.2023.1161161 70. Upadhya M, Kirmann T, Wilson MA, Simon CM, Dhangar D, Geis C, et al. Peripherally-derived LGI1-reactive monoclonal antibodies cause epileptic seizures *in vivo. Brain.* (2024). doi: 10.1093/brain/awae129

71. Petit-Pedrol M, Sell J, Planagumà J, Mannara F, Radosevic M, Haselmann H, et al. LGI1 antibodies alter Kv1.1 and AMPA receptors changing synaptic excitability, plasticity and memory. *Brain*. (2018) 141:3144–59. doi: 10.1093/brain/awy253

72. Rose NR, Bona C. Defining criteria for autoimmune diseases (Witebsky's postulates revisited). *Immunol Today*. (1993) 14:426–30. doi: 10.1016/0167-5699(93)90244-F

73. Hanin A, Cespedes J, Dorgham K, Pulluru Y, Gopaul M, Gorochov G, et al. Cytokines in new-onset refractory status epilepticus predict outcomes. *Ann Neurol.* (2023) 94:75–90. doi: 10.1002/ana.26627

74. Vezzani A, Di Sapia R, Kebede V, Balosso S, Ravizza T. Neuroimmunology of status epilepticus. *Epilepsy Behav.* (2023) 140:109095. doi: 10.1016/j.yebeh.2023.109095

75. Vezzani A, Balosso S, Ravizza T. Neuroinflammatory pathways as treatment targets and biomarkers in epilepsy. *Nat Rev Neurol.* (2019) 15:459–72. doi: 10.1038/s41582-019-0217-x

76. Helbig I, Barcia G, Pendziwiat M, Ganesan S, Mueller SH, Helbig KL, et al. Wholeexome and HLA sequencing in febrile infection-related epilepsy syndrome. *Ann Clin Transl Neurol.* (2020) 7:1429–35. doi: 10.1002/acn3.51062

77. Appenzeller S, Helbig I, Stephani U, Häusler M, Kluger G, Bungeroth M, et al. Febrile infection-related epilepsy syndrome (FIRES) is not caused by SCN1A, POLG, PCDH19 mutations or rare copy number variations. *Dev Med Child Neurol.* (2012) 54:1144–8. doi: 10.1111/j.1469-8749.2012.04435.x

78. Snowball A, Chabrol E, Wykes RC, Shekh-Ahmad T, Cornford JH, Lieb A, et al. Epilepsy gene therapy using an engineered Potassium Channel. *J Neurosci.* (2019) 39:3159–69. doi: 10.1523/JNEUROSCI.1143-18.2019

79. Street JS, Qiu Y, Lignani G. Are genetic therapies for epilepsy ready for the clinic? *Epilepsy Curr.* (2023) 23:245–50. doi: 10.1177/15357597231176234

80. Kumar P, Lim A, Hazirah SN, Chua CJH, Ngoh A, Poh SL, et al. Single-cell transcriptomics and surface epitope detection in human brain epileptic lesions identifies proinflammatory signaling. *Nat Neurosci.* (2022) 25:956–66. doi: 10.1038/s41593-022-01095-5