



Talking Convergence: Growing Evidence Links FOXP2 and Retinoic Acid in Shaping Speech-Related Motor Circuitry

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A commentary on

FOXP2 drives neuronal differentiation by interacting with retinoic acid signaling pathways by Devanna, P., Middelbeek, J., and Vernes, S. C. (2014). Front. Cell. Neurosci. 8:305. doi: 10.3389/fncel.2014.00305

FOXP2 was the first identified monogenic cause of a speech disorder (for review see Graham et al., 2015). However, it remains to be answered how it affects the development of speech related neuronal circuits and what the relevant molecular pathways may be. Devanna et al. (2014) reported a direct interaction between FOXP2 and Retinoic Acid (RA), an important signaling molecule in brain development and neuronal differentiation, in a cell model. What could this finding mean for our understanding of a language-ready brain?

Mounting evidence from diverse fields such as linguistics, genetics, brain development, and cellular neurophysiology suggests that the brain's motor circuitry might be the key to the conundrum of language disorders. Human patients with FOXP2 mutations show prominent motor learning deficits at the root of their developmental verbal dyspraxia diagnosis, which is reflected in mouse models of Foxp2 loss (for review see Deriziotis and Fisher, 2013; Graham et al., 2015). Interestingly, in mice with dysregulated RA signaling, similar motor learning impairments have been found (for review see Duester, 2013). Retinoic acid has several prominent roles in brain development and function, for example as a diffusible signaling molecule that changes gene expression via nuclear retinoic acid receptors (RARs, RORs, and RXRs; for review see Maden, 2007).

Devanna et al. (2014) demonstrated a direct interaction between FOXP2 and RA signaling in a human neuroblastoma cell line (SH-S5Y5), in which RA exposure initiates differentiation into a dopaminergic-neuron like state (Korecka et al., 2013). When the researchers compared FOXP2 overexpression to RA treatment they found an interesting convergence: In both cases, the cells differentiated from their previously proliferative state into a more neuron-like state and showed elevated expression of RA signaling-related genes, specifically the receptor RARß.

A recent review of the same group (van Rhijn and Vernes, 2015) suggests that the motor-learning deficits found in carriers of FoxP2 mutations might be mediated by defects in RA signaling, also in humans. Interestingly, FoxP2 and RA signaling have also been found (separately) to be involved in vocal production learning in a songbird model (Denisenko-Nehrbass et al., 2000; Olson et al., 2011; Roeske et al., 2014; Wohlgemuth et al., 2014). Even though direct evidence of synergies between FoxP2 and RA signaling is still lacking beyond cell models, the hypothesis that they are

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parts of the same regulatory network (e.g. Benítez-Burraco and Boeckx, 2014; van Rhijn and Vernes, 2015) would fit well with a number of findings, some of which we will address in the following commentary.

STRIATAL DEVELOPMENT: DO FoxP2 AND RA CONVERGE ON SPECIFIC CELL TYPES?

The canonical cortico-basal-ganglia-thalamic motor circuit is thought to be crucial for the motor aspects of vocal production. Interestingly, in this canonical circuit FoxP2 (refers to the gene across vertebrate species) and RA receptor expression overlap in several regions (**Figures 1A,B**). An especially illustrative region in this regard is the striatum, a hub of the motor circuitry that is known to be involved in motor sequencing in mammals (Kravitz and Kreitzer, 2012; Friend and Kravitz, 2014) and also in vocal production learning in songbirds (Bolhuis et al., 2010; Scharff and Petri, 2011). Available data indicates both FoxP2 expression and RA in the development and function of the striatum (van Rhijn and Vernes, 2015).

In the developing rodent striatum, two layers of progenitor cells in the ventricular and subventricular zone (SVZ) generate the medium spiny neurons (MSNs), the main cell type of the striatum (**Figure 1B**). Early during striatal development, a mixed population of progenitor cells in the SVZ starts producing RA, concomitant with the start of FoxP2 and RARß expression in the same region (Li et al., 2000; Ferland et al., 2003; Molotkova et al., 2007; Liao et al., 2008; Garcia-Calero et al., 2015). In general, RA signaling is involved very early in the spatial patterning of the

brain, starting with the formation of the telencephalic vesicle; however the precise extent has been controversially discussed (Siegenthaler et al., 2009; Chatzi et al., 2011, 2013; Duester, 2013). More importantly for motor circuit development, RA signaling is involved in the patterning of the striatum and striatonigral projections (Rataj-Baniowska et al., 2015) as well as migration of interneurons from the Ganglionic Eminences (that also produce MSNs) to the cortex (Crandall et al., 2011).

Within striatal MSNs, mouse studies imply an interesting convergence of FoxP2 and RA signaling onto a single cell population, namely D1R-MSNs ("direct pathway" neurons) of the dorsolateral striatum (Figure 1B). In mice lacking RARß, the progenitor population prematurely differentiates, specifically reducing the final cell numbers of D1R-MSNs (Rataj-Baniowska et al., 2015). Interestingly, RARβ was also one of the key RA receptors found to be affected in the human neuroblastoma cell study of Devanna et al. (2014). In mice lacking RARS, levels of the D1R-linked signal transduction protein Darpp-32 are greatly reduced (Liao et al., 2008). This is of interest not only because D1R-MSNs preferentially express FoxP2, but also because FoxP2 mutant mouse embryos likewise show strongly reduced Darpp-32 expression levels (Vernes et al., 2011, see supplementary material). One may therefore hypothesize that FoxP2 and RA signaling pathways converge in the development of the D1R-MSNs cell class in the dorsolateral striatum (Figure 1B), impacting the motor circuit's "direct pathway".

If these hypotheses are applied to a computational model of motor sequencing (Morita et al., 2012; Morita, 2014), several possible consequences emerge. On the circuit-function level, FoxP2 and RA disruptions would impair D1R-MSNs function, and hence the "go" signal in motor control (Sippy et al., 2015, **Figure 1B**). Furthermore, FoxP2 mutations may

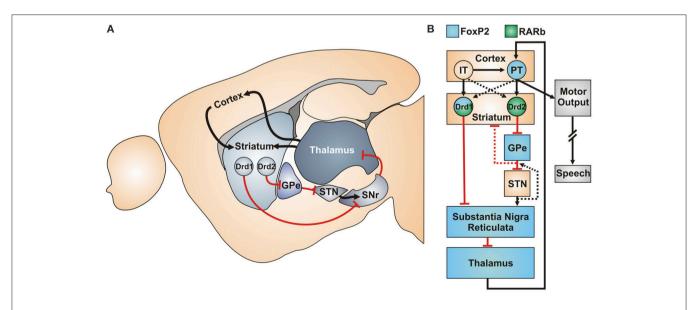


FIGURE 1 | Canonical cortico-basal-ganglia-thalamic circuit of the motor system (A) Sagittal view of the mouse motor system with excitatory (black) and inhibitory projections (red) connections. (B) Functional model (based on Morita 2014) of the motor sequencing circuitry with FoxP2 and RARb expression. IT, Intratelencephalic neurons of the cortical layer V; PT, Pyramidal tract "motor neurons" of layer Vb; Drd1/Drd2, Striatal dopamine-receptor 1/2 expressing MSNs; GPe, Globus Pallidus External; STN, Subthalamic nucleus; SNr, Substantia nigra, pars reticulata.

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affect inhibitory feedback to the striatum via a functionally defined population projecting from the Globus Pallidus Externa back to the striatum (Abdi et al., 2015; Dodson et al., 2015; Mallet et al., 2016), leading to additional defects in the "stop" motor control. Interestingly, a recent study found Foxp2 to regulate the development of the projection patterns of thalamic nuclei, another key integrative part of the motor circuitry (Ebisu et al., 2016).

DOWNSTREAM BEHAVIORAL EFFECTS: DO SIMILAR PHENOTYPES IMPLY CONVERGENT MECHANISMS?

In mice, Foxp2 loss impairs vocalizations after birth (e.g. Castellucci et al., 2016; Chen et al., 2016). However, mouse vocalizations are thought to lack a learning component, a key factor in human speech (see French and Fisher, 2014 for a review). In male zebra finches, a vocalization learning model, FoxP2 is expressed throughout the song system (Haesler et al., 2004, 2007; Scharff and Petri, 2011; Mendoza et al., 2015). Expression is especially high in the song-learning related area X of the striatum at the time when songs are acquired (Haesler et al., 2004; Thompson et al., 2013), and FoxP2 knockdown during this period disrupts social modulation of song variability (Murugan et al., 2013). Similar to the mouse striatal network, FoxP2 knockdown in the juvenile zebra finch leads to specific reductions of D1R and Darpp-32 expression in the dorsolateral striatum, specifically in area X (Denisenko-Nehrbass et al., 2000; Haesler et al., 2004, 2007; Olson et al., 2011; Murugan et al., 2013; Roeske et al., 2014). It should be noted that in the songbird striatum, D1R and D2R expression in MSNs overlap to a greater degree than in the mouse (Kubikova et al., 2010). Furthermore, striatal RA signaling plays an important role in the maintenance of birdsong: RA-producing enzymes are located in projection axons to area X, where the MSN-like neurons in turn express RA receptors. Functionally, RA may be important for balancing the input to area X neurons from vocal motor pathway (stereotyped song) versus anterior forebrain pathway (variable song) vocalization centers (Roeske et al., 2014; Wohlgemuth et al., 2014).

REFERENCES

- Abdi, A., Mallet, N., Mohamed, F. Y., Sharott, A., Dodson, P. D., Nakamura, K. C., et al. (2015). Prototypic and arkypallidal neurons in the dopamine-intact external globus pallidus. *J. Neurosci.* 35, 6667–6688. doi: 10.1523/JNEUROSCI. 4662-14.2015
- Benítez-Burraco, A., and Boeckx, C. (2014). FOXP2, retinoic acid, and language: a promising direction. Front. Cell. Neurosci. 8:387. doi: 10.3389/fncel.2014. 00387
- Bolhuis, J. J., Okanoya, K., and Scharff, C. (2010). Twitter evolution: converging mechanisms in birdsong and human speech. *Nat. Rev. Neurosci.* 11, 747–759. doi: 10.1038/nrn2931
- Castellucci, G. A., McGinley, M. J., and McCormick, D. A. (2016). Knockout of Foxp2 disrupts vocal development in mice. Sci. Rep. 6:23305. doi: 10.1038/srep 23305

FoxP2 AND RA: SYNERGY IN A LARGER REGULATORY NETWORK FOR NEURONAL CONNECTIVITY?

On the molecular level, the gene networks regulated by FoxP2 and RA signaling might overlap in specific parts of the motor circuitry. It is important to separate developmental effects from on-line functional effects: Developmental defects might lead to an impaired network setup (e.g. a smaller and miswired MSN population), while functional effects might impair the learning and function even of healthy networks (e.g. failure to balance synaptic inputs to MSNs). Furthermore, it should be noted that both FoxP2 and RA are broad regulators of large gene networks, also outside the motor circuitry.

In this context, a recent study is of particular interest: Chen et al. (2016) found that Foxp2 loss was associated with defects in dendritic spine formation in striatal neurons via de-repression of the autism risk gene Mef2C, leading to defects in corticostriatal connectivity. Research in a different murine cell model suggests that RA treatment indirectly interacts with Mef2C to specify a neuronal fate, via the transcription factor Sp1 (Elmi et al., 2007). It would therefore be highly interesting to see whether Foxp2, RA, Mef2C, and Sp1 are all part of the same regulatory network and influence the development of MSN connectivity.

Thus, with their finding of synergies between FOXP2 and RA in a cell model Devanna et al. (2014) made an important first step. Other studies provide indirect evidence that FoxP2 and RA signaling genes could be part of the same regulatory network and co-influence development and function of the speech-motor control circuits, especially the striatum. However, a direct proof beyond cell models is still lacking. Hence future studies in adequate animal models will have to confirm this synergy on the neuronal circuit level, also considering the more extensive regulatory molecular network affecting the development and functioning of speech-motor control circuits.

AUTHOR CONTRIBUTIONS

DS and MN both reviewed the current literature, wrote the manuscript and prepared the figures with equal contributions.

- Chatzi, C., Brade, T., and Duester, G. (2011). Retinoic acid functions as a key GABAergic differentiation signal in the basal ganglia. *PLoS Biol.* 9:e1000609. doi: 10.1371/journal.pbio.1000609
- Chatzi, C., Cunningham, T. J., and Duester, G. (2013). Investigation of retinoic acid function during embryonic brain development using retinaldehyderescued Rdh10 knockout mice. *Dev. Dyn.* 242, 1056–1065. doi: 10.1002/dvdy. 23000
- Chen, Y. C., Kuo, H. Y., Bornschein, U., Takahashi, H., Chen, S. Y., Lu, K. M., et al. (2016). Foxp2 controls synaptic wiring of corticostriatal circuits and vocal communication by opposing Mef2c. Nat. Neurosci. 19, 1513–1522. doi: 10.1038/nn.4380
- Crandall, J. E., Goodman, T., McCarthy, D. M., Duester, G., Bhide, P. G., Dräger, U. C., et al. (2011). Retinoic acid influences neuronal migration from the ganglionic eminence to the cerebral cortex. *J. Neurochem.* 119, 723–735. doi:10.1111/j.1471-4159.2011.07471.x

Synergies between FOXP2 and RA

- Denisenko-Nehrbass, N. I., Jarvis, E., Scharff, C., Nottebohm, F., and Mello, C. V. (2000). Site-specific retinoic acid production in the brain of adult songbirds. Neuron 27, 359–370. doi: 10.1016/S0896-6273(00)00043-X
- Deriziotis, P., and Fisher, S. E. (2013). Neurogenomics of speech and language disorders: the road ahead. *Genome Biol.* 14:204. doi: 10.1186/gb-2013-14-4-204
- Devanna, P., Middelbeek, J., and Vernes, S. C. (2014). FOXP2 drives neuronal differentiation by interacting with retinoic acid signaling pathways. Front. Cell. Neurosci. 8:305. doi: 10.3389/fncel.2014.00305
- Dodson, P. D., Larvin, J. T., Duffell, J. M., Garas, F. N., Doig, N. M., Kessaris, N., et al. (2015). Distinct developmental origins manifest in the specialized encoding of movement by adult neurons of the external globus pallidus. *Neuron* 86, 501–513. doi: 10.1016/j.neuron.2015.03.007
- Duester, G. (2013). Retinoid signaling in control of progenitor cell differentiation during mouse development. Semin. Cell Dev. Biol. 24, 694–700. doi: 10.1016/j.semcdb.2013.08.001
- Ebisu, H., Iwai-Takekoshi, L., Fujita-Jimbo, E., Momoi, T., and Kawasaki, H. (2016). Foxp2 regulates identities and projection patterns of thalamic nuclei during development. *Cereb. Cortex*. doi: 10.1093/cercor/bhw187. [Epub ahead of Print].
- Elmi, M., Faigle, R., Yang, W., Matsumoto, Y., Rosenqvist, E., and Funa, K. (2007). Mechanism of MASH1 induction by ASK1 and ATRA in adult neural progenitors. *Mol. Cell. Neurosci.* 36, 248–259. doi: 10.1016/j.mcn.2007. 07 001
- Ferland, R. J., Cherry, T. J., Preware, P. O., Morrisey, E. E., and Walsh, C. A. (2003). Characterization of Foxp2 and Foxp1 mRNA and protein in the developing and mature brain. J. Comp. Neurol. 460, 266–279. doi: 10.1002/cne.10654
- French, C. A., and Fisher, S. E. (2014). What can mice tell us about Foxp2 function? Curr. Opin. Neurobiol. 28C, 72–79. doi: 10.1016/j.conb.2014.07.003
- Friend, D. M., and Kravitz, A. V. (2014). Working together: basal ganglia pathways in action selection. *Trends Neurosci.* 37, 301–303. doi: 10.1016/j.tins. 2014.04.004
- Garcia-Calero, E., Botella-Lopez, A., Bahamonde, O., Perez-Balaguer, A., and Martinez, S. (2015). FoxP2 protein levels regulate cell morphology changes and migration patterns in the vertebrate developing telencephalon. *Brain Struct. Funct.* 221, 2905–2917. doi: 10.1007/s00429-015-1079-7
- Graham, S. A., Deriziotis, P., and Fisher, S. E. (2015). Insights into the genetic foundations of human communication. *Neuropsychol. Rev.* 25, 3–26. doi:10.1007/s11065-014-9277-2
- Haesler, S., Rochefort, C., Georgi, B., Licznerski, P., Osten, P., and Scharff, C. (2007). Incomplete and inaccurate vocal imitation after knockdown of FoxP2 in songbird basal ganglia nucleus area X. PLoS Biol. 5:e321. doi: 10.1371/journal.pbio.0050321
- Haesler, S., Wada, K., Nshdejan, A., Morrisey, E. E., Lints, T., Jarvis, E. D., et al. (2004). FoxP2 expression in avian vocal learners and non-learners. *J. Neurosci.* 24, 3164–3175. doi: 10.1523/JNEUROSCI.4369-03.2004
- Korecka, J. A., van Kesteren, R. E., Blaas, E., Spitzer, S. O., Kamstra, J. H., Smit, A. B., et al. (2013). Phenotypic characterization of retinoic acid differentiated SH-SY5Y cells by transcriptional profiling. *PLoS ONE* 8:e63862. doi: 10.1371/journal.pone.0063862
- Kubikova, L., Wada, K., and Jarvis, E. D. (2010). Dopamine receptors in a songbird brain. J. Comp. Neurol. 518, 741–769. doi: 10.1002/cne.22255
- Li, H., Wagner, E., McCaffery, P., Smith, D., Andreadis, A., and Dräger, U. C. (2000). A retinoic acid synthesizing enzyme in ventral retina and telencephalon of the embryonic mouse. *Mech. Dev.* 95, 283–289. doi: 10.1016/S0925-4773(00)00352-X
- Liao, W. L., Tsai, H. C., Wang, H. F., Chang, J., Lu, K. M., Wu, H. L., et al. (2008). Modular patterning of structure and function of the striatum by retinoid receptor signaling. *Proc. Natl. Acad. Sci. U.S.A.* 105, 6765–6770. doi: 10.1073/ pnas.0802109105
- Maden, M. (2007). Retinoic acid in the development, regeneration and maintenance of the nervous system. Nat. Rev. Neurosci. 8, 755–765. doi:10.1038/nrn2212
- Mallet, N., Schmidt, R., Leventhal, D., Chen, F., Amer, N., Boraud, T., et al. (2016). Arkypallidal cells provide a Stop signal to Striatum. *Neuron* 89, 308–316. doi: 10.1016/j.neuron.2015.12.017

- Mendoza, E., Tokarev, K., Düring, D. N., Retamosa, E. C., Weiss, M., Arpenik, N., et al. (2015). Differential coexpression of FoxP1, FoxP2, and FoxP4 in the Zebra Finch (*Taeniopygia guttata*) song system. *J. Comp. Neurol.* 523, 1318–1340. doi: 10.1002/cne.23731
- Molotkova, N., Molotkov, A., and Duester, G. (2007). Role of retinoic acid during forebrain development begins late when Raldh3 generates retinoic acid in the ventral subventricular zone. *Dev. Biol.* 303, 601–610. doi: 10.1016/j.ydbio.2006.11.035
- Morita, K. (2014). Differential cortical activation of the striatal direct and indirect pathway cells: reconciling the anatomical and optogenetic results by using a computational method. J. Neurophysiol. 112, 120–146. doi: 10.1152/jn. 00625.2013
- Morita, K., Morishima, M., Sakai, K., and Kawaguchi, Y. (2012). Reinforcement learning: computing the temporal difference of values via distinct corticostriatal pathways. *Trends Neurosci.* 35, 457–467. doi: 10.1016/j.tins.2012.04.009
- Murugan, M., Harward, S., Scharff, C., and Mooney, R. (2013). Diminished FoxP2 levels affect dopaminergic modulation of corticostriatal signaling important to song variability. *Neuron* 80, 1464–1476. doi: 10.1016/j.neuron.2013.09.021
- Olson, C. R., Rodrigues, P. V., Jeong, J. K., Prahl, D. J., and Mello, C. V. (2011). Organization and development of zebra finch HVC and paraHVC based on expression of zRalDH, an enzyme associated with retinoic acid production. J. Comp. Neurol. 519, 148–161. doi: 10.1002/cne.22510
- Rataj-Baniowska, M., Niewiadomska-Cimicka, A., Paschaki, M., Szyszka-Niagolov, M., Carramolino, L., Torres, M., et al. (2015). Retinoic acid receptor controls development of striatonigral projection neurons through FGF-dependent and Meis1-dependent mechanisms. J. Neurosci. 35, 14467–14475. doi: 10.1523/JNEUROSCI.1278-15.2015
- Roeske, T. C., Scharff, C., Olson, C. R., Nshdejan, A., and Mello, C. V. (2014). Long-distance retinoid signaling in the zebra finch brain. *PLoS ONE* 9:e111722. doi: 10.1371/journal.pone.0111722
- Scharff, C., and Petri, J. (2011). Evo-devo, deep homology and FoxP2: implications for the evolution of speech and language. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 366, 2124–2140. doi: 10.1098/rstb.2011.0001
- Siegenthaler, J. A., Ashique, A. M., Zarbalis, K., Patterson, K. P., Hecht, J. H., Kane, M. A., et al. (2009). Retinoic acid from the meninges regulates cortical neuron generation. *Cell* 139, 597–609. doi: 10.1016/j.cell.2009.10.004
- Sippy, T., Lapray, D., Crochet, S., and Petersen, C. C. (2015). Cell-type-specific sensorimotor processing in striatal projection neurons during goal-directed behavior. *Neuron* 88, 298–305. doi: 10.1016/j.neuron.2015.08.039
- Thompson, C. K., Schwabe, F., Schoof, A., Mendoza, E., Gampe, J., Rochefort, C., et al. (2013). Young and intense: FoxP2 immunoreactivity in Area X varies with age, song stereotypy, and singing in male zebra finches. *Front. Neural Circuits* 7:24. doi: 10.3389/fncir.2013.00024
- van Rhijn, J. R., and Vernes, S. C. (2015). Retinoic acid signaling: A new piece in the spoken language puzzle. *Front. Psychol.* 6:1816. doi: 10.3389/fpsyg.2015.01816
- Vernes, S. C., Oliver, P. L., Spiteri, E., Lockstone, H. E., Puliyadi, R., Taylor, J. M., et al. (2011). Foxp2 regulates gene networks implicated in neurite outgrowth in the developing brain. *PLoS Genet.* 7:e1002145. doi:10.1371/journal.pgen.1002145
- Kravitz, A. V., and Kreitzer, A. C. (2012). Striatal Mechanisms Underlying Movement, Reinforcement, and Punishment. *Physiology* 27, 167–177. doi:10.1152/physiol.00004.2012
- Wohlgemuth, S., Adam, I., and Scharff, C. (2014). FoxP2 in songbirds. Curr. Opin. Neurobiol. 28, 86–93. doi: 10.1016/j.conb.2014.06.009
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