



Sphingolipid Metabolism: A New Therapeutic Opportunity for Brain Degenerative Disorders

Alba Di Pardo and Vittorio Maglione*

IRCCS Neuromed, Pozzilli, Italy

Neurodegenerative diseases represent a class of fatal brain disorders for which the number of effective therapeutic options remains limited with only symptomatic treatment accessible. Multiple studies show that defects in sphingolipid pathways are shared among different brain disorders including neurodegenerative diseases and may contribute to their complex pathogenesis. In this mini review, we discuss the hypothesis that modulation of sphingolipid metabolism and their related signaling pathways may represent a potential therapeutic approach for those devastating conditions. The plausible "druggability" of sphingolipid pathways is greatly promising and represent a relevant feature that brings real advantage to the development of new therapeutic options for these conditions. Indeed, several molecules that selectively target sphingolipds are already available and many of them currently in clinical trial for human diseases. A deeper understanding of the "sphingolipid scenario" in neurodegenerative disorders would certainly enhance therapeutic perspectives for these conditions, by taking advantage from the already available molecules and by promoting the development of new ones.

OPEN ACCESS

Edited by:

Gian Carlo Bellenchi, Institute of Genetics and Biophysics (CNR), Italy

Reviewed by:

Nataliya G. Kolosova, Institute of Cytology and Genetics (RAS), Russia Riccardo Ghidoni, Università degli Studi di Milano, Italy

*Correspondence:

Vittorio Maglione vittorio.maglione@neuromed.it

Specialty section:

This article was submitted to Neurodegeneration, a section of the journal Frontiers in Neuroscience

Received: 29 October 2017 Accepted: 29 March 2018 Published: 17 April 2018

Citation:

Di Pardo A and Maglione V (2018) Sphingolipid Metabolism: A New Therapeutic Opportunity for Brain Degenerative Disorders. Front. Neurosci. 12:249. doi: 10.3389/fnins.2018.00249 Keywords: neurodegenerative diseases, sphingolipid metabolism, S1P, ceramide, FTY720, S1PRs

INTRODUCTION

Sphingolipids have long been viewed as merely ubiquitous components of the cell membrane, and exert a critical role in regulating vital cell functions and formation of membrane microdomain "lipid rafts" for integrating cell signaling (Gault et al., 2010; Olsen and Faergeman, 2017). Sphingolipids synthesis can occur via *de novo* biosynthetic pathway or the hydrolysis of sphingomyelin, or can also derive by the "salvage pathway" which determines the recovery of sphingosine, by the recycling of complex sphingolipids (gangliosides) through a coordinated action of several enzymes (see **Figure 1**) (Gault et al., 2010).

Enzymatic pathways result in the formation of several different and tightly regulated lipid mediators such as ceramide, ceramide-1-phosphate (C1P), sphingosine, and sphingosine-1-phosphate (S1P) (Gault et al., 2010; Goñi et al., 2014).

Ceramide is pivotal in the synthesis of sphingolipids (Mullen et al., 2012). It benefits early growth and development of neuronal cells and, at low levels it promotes cell survival and division (Schwarz and Futerman, 1997; Brann et al., 1999). Defects in its metabolism may have deleterious effects, often resulting in its abnormal accumulation that causes an increase of the programmed cell death in different cell types including neurons (Tohyama et al., 1999; Jana et al., 2009; Mullen and Obeid, 2012; Czubowicz and Strosznajder, 2014).



Direct phosphorylation of ceramide by ceramide kinase (CerK) is the major identified mechanism for generation of C1P in cells (Bajjalieh et al., 1989). The role of C1P has been mainly studied in blood cells in which it regulates proliferation, migration, and survival (Hoeferlin et al., 2013). The function of C1P in the brain is still poorly investigated. However, the presence of CerK in synaptic vesicles (Bajjalieh et al., 1989), along with the evidence that C1P regulates photoreceptor homeostasis (Miranda et al., 2011) as well as P-glycoprotein transport at the blood-brain barrier (Mesev et al., 2017), strongly suggests that C1P may be involved also in the homeostasis of the central nervous system (CNS).

Similarly, S1P is a potent signaling molecule that, beside governing essential physiological processes like vascular, bone formation (Hla et al., 2008; Xiong and Hla, 2014; Holmes, 2015) and inflammatory response (Huang et al., 2013; Aoki et al., 2016), regulates many of the molecular events crucial for brain development and neuronal survival (Mendelson et al., 2014; van Echten-Deckert et al., 2014). S1P acts either in the intracellular or in the extracellular compartments (Strub et al., 2010; Mendelson et al., 2014). Intracellularly, S1P may play different roles depending on its subcellular localization and normally regulates mitochondria function (Strub et al., 2011; Shen et al., 2014), gene expression by inhibiting histone deacetylases (HDACs) (Hait et al., 2009; Riccio, 2010) and ER stress (Lépine et al., 2011; Park et al., 2016).

Outside the cell, S1P acts as a high affinity agonist at five known G protein-coupled receptors, S1PR₁ -S1PR₅, which in the brain are expressed by many CNS cell types (Blaho and Hla, 2014; Martin and Sospedra, 2014) and have been shown to influence cell proliferation and migration, cell differentiation and survival as well as neurite outgrowth and neurogenesis (Toman et al., 2004; Anderson and Maes, 2014; Blaho and Hla, 2014; Martin and Sospedra, 2014; Ye et al., 2016).

Several studies showed that these sphingolipid mediators and their enzymes are likely to have an integral role in different cell processes including proliferation, inflammation, apoptosis, and migration (Zheng et al., 2006; Maceyka et al., 2012).

S1P metabolism involves a number of different highly specialized enzymes. S1P is normally synthesized by sphingosine kinase-1 and -2 (SPHK1 and 2) and degraded by sphingosine-1-phosphate phosphatase (SGPP) or lyase (SGPL1) (see **Figure 1**) (Le Stunff et al., 2002; Morozov et al., 2013). SPHK1 activity is mainly associated with cell survival (Le Stunff et al., 2002;

Morozov et al., 2013; Di Pardo et al., 2017a), while SPHK2 is widely described as being a dual-function protein whose activity may either guarantee the proper occurrence of physiological events like mitochondrial function and homeostasis as well as regulation of gene expression through inhibition of Class I HDACs or result detrimental mainly suppressing cell growth and promoting apoptosis (Maceyka et al., 2005; Hait et al., 2009; Riccio, 2010; Gomez et al., 2011; Strub et al., 2011).

On the other hand, the degradative enzymes SGPP and SGPL1 represent key regulators for the maintenance of balanced S1P levels and other sphingolipid intermediates that may control cell growth, proliferation and death (Serra and Saba, 2010). Uncontrolled up-regulation of SGPL1 typically results in a reduction of S1P availability and accumulation of hexadecenal, a biochemical condition previously reported to be cytotoxic (Kumar et al., 2011; Maceyka et al., 2012) that may likely contribute to neurodegenerative processes.

This mini-review will shortly highlight the current knowledge of research that now supports the idea that sphingolipids are intimately involved in disease progression and, together with altered expression/activity of metabolizing enzymes and associated receptors, can provide effective drug targets for the treatment of pathological states.

Alteration of Sphingolipid Metabolism in Neurodegenerative Disorders

A fine balance between synthesis of sphingolipids and their degradation is normally required for many biological processes (Gault et al., 2010; Mullen et al., 2012), thus changes in their metabolism may profoundly affect brain homeostasis and function. Over the past few years, perturbed metabolism of the interconvertible bioactive sphingolipids, ceramide and S1P is increasingly becoming recognized as potential pathogenic factor in different neurodegenerative disorders (**Table 1**). Indeed, a plethora of new information identifying the importance of sphingolipids and their signaling pathways in these conditions has accumulated.

Alzheimer's Disease

Alzheimer's disease (AD) is the most common neurodegenerative disease and the leading cause of dementia in elderly people (Kumar et al., 2015). Progressive neurodegeneration in brain regions involved in learning and memory results in cognitive decline, loss of memory and changes of social and emotional behavior (Robins Wahlin and Byrne, 2011; Sona et al., 2013; Levenson et al., 2014). AD is characterized by extracellular accumulation of amyloid β -peptide (A β) toxic aggregates and intracellular deposits of abnormally phosphorylated tau protein (Blennow et al., 2006; Gouras et al., 2010).

A number of evidence demonstrates a key role of aberrant sphingolipid metabolism in the pathogenesis of the disease (Chakrabarti et al., 2016). Activity of ceramide synthetic enzyme 2 (CERS2), normally involved in the formation of long chain ceramide species (Levy and Futerman, 2010), has been recently found to be reduced in multiple brain regions of subjects at the preclinical stage of the disease (Couttas et al., 2016). Conversely, expression of genes involved in the *de novo* synthesis of sphingolipids is upregulated early in the disease progression (Katsel et al., 2007). Consistently, accumulation of ceramide has been reported in brain tissues from AD patients even at early stages of disease (Han et al., 2002; Cutler et al., 2004; He et al., 2010; Filippov et al., 2012) and may contribute to neurotoxic action of A β (Lee et al., 2004; Dinkins et al., 2015).

A number of evidence also indicates that, along with ceramide abnormalities, metabolism of other sphingolipids is affected in AD (Zheng et al., 2006; Mielke and Lyketsos, 2010). Also, alteration in the expression and/or in the activity of S1Pmetabolizing enzymes as well as reduced levels of S1P has been widely reported in AD human brains (Katsel et al., 2007; Ceccom et al., 2014a,b; Couttas et al., 2014).

In particular, loss of SPHK1 and reduced bioavailability of S1P have been found early in the pathogenesis of the disease even before clinical diagnosis (Couttas et al., 2014). Conversely, upregulation of SPHK2 has been described to modulate A β release (Takasugi et al., 2011). Further indication of global derangement of sphingolipid metabolism in AD comes from the evidence of concomitant reduction in the levels of SPHK1 and S1P receptor 1 with enhanced expression of SGPL1 in postmortem brains from human patients (Ceccom et al., 2014b).

Huntington's Disease

Huntington's disease (HD) is the most common dominantly inherited brain disorder, characterized by progressive striatal and cortical neurodegeneration which results in motor, cognitive and behavioral disturbance (Mccolgan and Tabrizi, 2018). The disease derives from the expansion of a polyglutamine stretch (polyQ) (> 36 repeats) in the N-terminal region of the protein huntingtin (Htt) (Jimenez-Sanchez et al., 2017). Although the function of this protein is not completely known, expansion of the polyQ stretch endows mutant Htt (mHtt) with toxic properties, resulting in the development of a number of deleterious effects in both neuronal and non-neuronal cells (Maglione et al., 2005, 2006a,b; Carroll et al., 2015; Jimenez-Sanchez et al., 2017).

Among all cellular dysfunctions and biochemical defects, classically associated with the disease, defective metabolism of sphingolipids seems to play a key role in its pathogenesis (Di Pardo et al., 2017a).

Expression of S1P-metabolizing enzymes was reported to be aberrant in multiple HD settings (Di Pardo et al., 2017a,b; Pirhaji et al., 2017). Levels of SPHK1 was found reduced in brain tissues form two fully manifest HD mouse models (R6/2 and YAC128 mice), and most importantly, in brain tissues from HD patients (Di Pardo et al., 2017a). Conversely, levels of SGPL1 were increased in the brain of multiple disease animal models and in early manifest R6/2 mice (Di Pardo et al., 2017a), indicating that defect in sphingolipid metabolism occurs early in the disease stage likely contributing to its pathogenic process. First signs of early defective sphingolipid metabolism in HD have been reported also (Di Pardo et al., 2017a; Pirhaji et al., 2017) and such further evidence corroborates the hypothesis that similar alterations may conceivably contribute to the pathogenesis of the disease.

The imbalance in S1P-metabolizing enzymes results in decreased bioavailability of S1P and increased levels of ceramide

Molecule	Alzheimer Disease	Huntington Disease	Parkinson Disease
CERS2	Downregulated (Couttas et al., 2016)	Not Available	Not Available
Ceramides	Increased levels (Han et al., 2002; Cutler et al., 2004; Lee et al., 2004; Dinkins et al., 2015)	Increased levels (Pirhaji et al., 2016, 2017; Di Pardo et al., 2017a)	Increased levels (Ferrazza et al., 2016)
SPHK1	Downregulated (Ceccom et al., 2014b; Couttas et al., 2014)	Downregulated (Di Pardo et al., 2017a)	Downregulated (Sivasubramanian and Tay Ssw, 2013; Pyszko and Strosznajder, 2014)
SPHK2	Upregulated (Takasugi et al., 2011)	Upregulated (Di Pardo et al., 2017a; Moruno-Manchon et al., 2017)	Downregulated (Sivasubramanian and Tay Ssw, 2013; Pyszko and Strosznajder, 2014 Sivasubramanian et al., 2015)
SGPL1	Upregulated (Ceccom et al., 2014b)	Upregulated (Di Pardo et al., 2017a; Pirhaji et al., 2017)	Not Available
S1P	Reduced levels (Couttas et al., 2014)	Reduced levels (Pirhaji et al., 2016, 2017; Di Pardo et al., 2017a)	Not Available

species as reported in HD models (Pirhaji et al., 2016, 2017; Di Pardo et al., 2017a).

Ultimately, synthesis of *de novo* sphingolipids is also affected in HD animals, even at early stage of the disease (Di Pardo et al., 2017b). This alteration determines a robust reduction of certain dihydroceramide species along with dihydrosphingosine and dihydroS1P (Di Pardo et al., 2017b).

Parkinson's Disease

Parkinson's disease (PD) is a neurodegenerative movement disorder with a prevalence of approximately 1 to 2% of the population over 65 years which increases up to 5% in people over 85 years old (Fahn, 2003; Obeso et al., 2010).

The pathological hallmarks of PD include the loss of dopaminergic neurons in the *substantia nigra pars compacta* and the formation of Lewy bodies mainly composed of aggregated alpha-synuclein (a-syn) protein and other components, including lipids (Gai et al., 2000; Halliday et al., 2005; Dickson et al., 2009).

Several studies demonstrated that defective ceramide metabolism may contribute to the pathogenesis of PD (Bras et al., 2008; Haughey, 2010; Fabelo et al., 2011). Mutations in the gene encoding for Glucocerebrosidase (GBA), a lysosomal enzyme converting glucosyl-ceramides into ceramide (see also **Figure 1**), increase the risk of developing sporadic PD (O'Regan et al., 2017).

Alterations in the expression of ceramide synthase genes as well as in the levels of certain ceramide species have been reported in post-mortem brain tissues from sporadic PD patients (Abbott et al., 2014). Consistently, mice knock out for Leucine-rich repeat kinase 2 (LRRK2), whose gene mutations cause inherited PD (Li et al., 2014), show a significant increase in the content of brain ceramide (Ferrazza et al., 2016).

Although very little, defects in S1P metabolism have been also reported in PD. Levels of SPHK1 and 2 have been described aberrant in both *in-vitro* and *in-vivo* models highlighting a potential contribution of S1P metabolism to the pathogenesis of the disease (Sivasubramanian and Tay Ssw, 2013; Sivasubramanian et al., 2015). Consistently, further studies demonstrate altered expression of S1P metabolizing enzymes with reduced activity for both SPHK1 and 2 in an *in-vitro* PD model (neuronal dopaminergic SH-SY5Y cells) induced by 1methyl-4-phenylpyridinium (MPP+) (Pyszko and Strosznajder, 2014).

Sphingolipid Pathways as Target for the Development of Potential Therapeutic Interventions

The study of bioactive sphingolipids as novel therapeutic targets for the treatment of brain disorders is still relatively young, however efforts are being made to question how therapeutics may be designed to take advantage of sphingolipid signaling pathways. Over the past few years it has been established that modulation of sphingolipid metabolism can lead to improved efficacy in many brain disorders ranging from neurodevelopmental to neurodegenerative settings (Deogracias et al., 2012; Asle-Rousta et al., 2013a; Di Pardo et al., 2014; Miguez et al., 2015; Moruno Manchon et al., 2015; Pirhaji et al., 2017; Zhao et al., 2017).

Cell survival and proper functioning of neuronal circuits is critical to the effectiveness of therapies in brain disorders. Sphingolipids are key signaling molecules regulating many of these cellular processes, thus it is very likely that manipulation of sphingolipid metabolism may represent a great way to develop more effective and targeted therapeutic strategies.

Alzheimer's Disease

Very little is still known about the therapeutic effect that the modulation of sphingolipid metabolism may have in AD, however there is evidence demonstrating that stimulation of S1P receptors is beneficial in pre-clinical models of the diseases (Asle-Rousta et al., 2013a,b, 2014; Hemmati et al., 2013; Takasugi et al., 2013). Infusion of FTY720 (fingolimod), a non-selective S1P receptor modulator and FDA- approved drug for the treatment of Multiple Sclerosis, prevents spatial learning and memory impairments as well as A β induced-changes in the expression of some pro-apoptotic and inflammatory markers in AD animal models (Asle-Rousta et al., 2013a, 2014; Hemmati et al., 2013). Consistently, treatment with fingolimod leads to reduction of A β species in both *in-vitro* and *in-vivo* disease models (Takasugi et al., 2013). Activation of specific S1P receptors also exerts beneficial effect in animal models of the disease. Indeed, administration of SEW2871, a S1PR₁ selective agonist, ameliorates spatial memory impairments and attenuates the A β 1-induced hippocampal neuronal loss in an AD rat model (Asle-Rousta et al., 2013b).

Huntington's Disease

The effective therapeutic potential of the modulation of both levels and activity of S1P-metabolizing enzyme in HD is increasingly becoming evident in the last few years (Moruno Manchon et al., 2015; Di Pardo et al., 2017a; Pirhaji et al., 2017).

Recent evidence demonstrates that SPHK1 co-localizes with autophagosomes and its over expression favors autophagymediated clearance of mutant Htt exon-1 construct *in vitro* (Moruno Manchon et al., 2015). Coherently, stimulation of SPHK1, by the selective pharmacological activator K6PC-5 (Ji et al., 2015), significantly reduces apoptosis in mouse striatal derived HD cell lines and leads to the activation of pro-survival signaling pathways (Di Pardo et al., 2017a). The potential therapeutic validity of this pharmacological approach in HD is further supported by the beneficial effects that modulation of the kinase has in human HD iPSC-derived neurons (Di Pardo et al., 2017a).

Inhibition of SPHK2 or SGPL1 also represents a possible pharmacological strategy to increase survival in HD cell models (Di Pardo et al., 2017a; Moruno-Manchon et al., 2017; Pirhaji et al., 2017). Stimulation of S1P receptors is an additional therapeutically effective approach in this context. It stimulates the production of neurotrophins, activates neuronal pro-survival pathways and ultimately delays disease progression in HD mouse models (Di Pardo et al., 2014; Miguez et al., 2015).

Parkinson's Disease

Modulation of S1P pathways has been recently explored in experimental models of PD. Pharmacological inhibition of SPHK1 reduces cell survival and increases oxygen reactive species in MPP+ human dopaminergic neuronal cells (Pyszko and Strosznajder, 2014). Conversely, treatment with S1P significantly reduced apoptosis in the same experimental model by regulating the expression of S1PR₁ (Pyszko and Strosznajder, 2014).

Coherently, inhibition of SPHK2 leads to downregulation of mitochondrial-related genes such as proliferator-activated receptor γ coactivator-1 α (PGC-1 α) and its downstream targets nuclear respiratory factor 1 (NRF-1) and mitochondrial transcription factor A (TFAM) in multiple PD experimental models (Sivasubramanian et al., 2015). Furthermore, treatment with S1P enhance the expression of PGC-1 α and NRF-1 in a mouse model of the disease and exert neuroprotective effect of dopaminergic neurons via S1PR₁ (Sivasubramanian et al., 2015).

The potential role of S1PR₁ stimulation in PD has been further explored. Treatment with FTY720 attenuates motor deficit and prevent dopaminergic neuronal loss in two chemical-induced PD models (Zhao et al., 2017). The neuroprotective effect of the drug is associated with the activation of pro-survival kinase ERK in

both *in-vitro* and *in-vivo* PD models (Zhao et al., 2017). The benefit of FTY720 was abolished by the co-treatment with W146, a S1PR₁ selective antagonist, indicating that action of fingolimod was mediated, at least in part, by its binding at S1PR₁.

CONCLUDING REMARKS AND FUTURE PERSPECTIVES

In the last few years, the recognition of aberrant sphingolipid metabolism is becoming more evident in neurodegenerative disorders and, its deeper investigation may either strongly contribute to a better understanding of the disease pathogenesis or support the development of novel and more targeted therapeutic approaches.

Defects in this metabolism may profoundly affect the CNS and may interfere with selective biological pathways, whose dysregulation can explain some of the molecular and cellular alterations underlying neurodegeneration.

The hypothesis of candidating sphingolipid pathways as attractive therapeutic targets is strongly supported by evidence that demonstrates that modulation of such pathways has beneficial effects in different neurodegenerative conditions. To date, what makes sphingolipid metabolism a promising target with a real potential to be successfully translated into clinical practice of brain disorders is the evidence that some of the drugs targeting S1P and its receptors are already marketed or in advanced phases of clinical development for the treatment of human disorders (Hatoum et al., 2017; O'Sullivan and Dev, 2017). This could certainly allow to take advantage from the already available molecules and eventually to promote the development of new ones.

Takin into account the diverse functions that some sphingolipids may have, reducing ceramide accumulation and/or targeting its conversion into other sphingolipid species may represent an alternative approach for brain disorders (Brodowicz et al., 2018). In this context, the feasibility of lowering ceramide levels and its effectiveness is supported by a number of pre-clinical studies and by the evidence that some drugs that target ceramide production are currently approved for clinical use (Kornhuber et al., 2010). On the other hand, boosting ceramide metabolism toward the synthesis of C1P and/or S1P, through the use of specific kinase activators (Kwon et al., 2007; Tada et al., 2010), may also represent a fascinating approach.

AUTHOR CONTRIBUTIONS

AD and VM co-wrote the manuscript.

FUNDING

This research was supported by Fondazione Neuromed and funded by Italian Ministry of Health Ricerca Corrente funding program granted to VM.

REFERENCES

- Abbott, S. K., Li, H., Muñoz, S. S., Knoch, B., Batterham, M., Murphy, K. E., et al. (2014). Altered ceramide acyl chain length and ceramide synthase gene expression in Parkinson's disease. *Mov. Disord.* 29, 518–526. doi: 10.1002/mds.25729
- Anderson, G., and Maes, M. (2014). Reconceptualizing adult neurogenesis: role for sphingosine-1-phosphate and fibroblast growth factor-1 in co-ordinating astrocyte-neuronal precursor interactions. CNS Neurol. Disord. Drug Targets 13, 126–136. doi: 10.2174/18715273113126660132
- Aoki, M., Aoki, H., Ramanathan, R., Hait, N. C., and Takabe, K. (2016). Sphingosine-1-phosphate signaling in immune cells and inflammation: roles and therapeutic potential. *Mediators Inflamm.* 2016:8606878. doi: 10.1155/2016/8606878
- Asle-Rousta, M., Kolahdooz, Z., Dargahi, L., Ahmadiani, A., and Nasoohi, S. (2014). Prominence of central sphingosine-1-phosphate receptor-1 in attenuating abeta-induced injury by fingolimod. *J. Mol. Neurosci.* 54, 698–703. doi: 10.1007/s12031-014-0423-3
- Asle-Rousta, M., Kolahdooz, Z., Oryan, S., Ahmadiani, A., and Dargahi, L. (2013a). FTY720 (fingolimod) attenuates beta-amyloid peptide (Abeta42)induced impairment of spatial learning and memory in rats. J. Mol. Neurosci. 50, 524–532. doi: 10.1007/s12031-013-9979-6
- Asle-Rousta, M., Oryan, S., Ahmadiani, A., and Rahnema, M. (2013b). Activation of sphingosine 1-phosphate receptor-1 by SEW2871 improves cognitive function in Alzheimer's disease model rats. *EXCLI J.* 12, 449–461.
- Bajjalieh, S. M., Martin, T. F., and Floor, E. (1989). Synaptic vesicle ceramide kinase. A calcium-stimulated lipid kinase that co-purifies with brain synaptic vesicles. J. Biol. Chem. 264, 14354–14360.
- Blaho, V. A., and Hla, T. (2014). An update on the biology of sphingosine 1-phosphate receptors. J. Lipid Res. 55, 1596–1608. doi: 10.1194/jlr.R046300
- Blennow, K., De Leon, M. J., and Zetterberg, H. (2006). Alzheimer's disease. *Lancet* 368, 387–403. doi: 10.1016/S0140-6736(06)69113-7
- Brann, A. B., Scott, R., Neuberger, Y., Abulafia, D., Boldin, S., Fainzilber, M., et al. (1999). Ceramide signaling downstream of the p75 neurotrophin receptor mediates the effects of nerve growth factor on outgrowth of cultured hippocampal neurons. J. Neurosci. 19, 8199–8206.
- Bras, J., Singleton, A., Cookson, M. R., and Hardy, J. (2008). Emerging pathways in genetic Parkinson's disease: potential role of ceramide metabolism in Lewy body disease. *FEBS J.* 275, 5767–5773. doi: 10.1111/j.1742-4658.2008.06709.x
- Brodowicz, J., Przegalinski, E., Müller, C. P., and Filip, M. (2018). Ceramide and its related neurochemical networks as targets for some brain disorder therapies. *Neurotox. Res.* 33, 474–484. doi: 10.1007/s12640-017-9798-6
- Carroll, J. B., Bates, G. P., Steffan, J., Saft, C., and Tabrizi, S. J. (2015). Treating the whole body in Huntington's disease. *Lancet Neurol.* 14, 1135–1142. doi: 10.1016/S1474-4422(15)00177-5
- Ceccom, J., Delisle, M. B., and Cuvillier, O. (2014a). [Sphingosine 1phosphate as a biomarker for Alzheimer's disease? *Med. Sci.* 30, 493–495. doi: 10.1051/medsci/20143005006
- Ceccom, J., Loukh, N., Lauwers-Cances, V., Touriol, C., Nicaise, Y., Gentil, C., et al. (2014b). Reduced sphingosine kinase-1 and enhanced sphingosine 1-phosphate lyase expression demonstrate deregulated sphingosine 1phosphate signaling in Alzheimer's disease. Acta Neuropathol. Commun. 2:12. doi: 10.1186/2051-5960-2-12
- Chakrabarti, S. S., Bir, A., Poddar, J., Sinha, M., Ganguly, A., and Chakrabarti, S. (2016). Ceramide and sphingosine-1-phosphate in cell death pathways: relevance to the pathogenesis of Alzheimer's disease. *Curr. Alzheimer Res.* 13, 1232–1248. doi: 10.2174/1567205013666160603004239
- Couttas, T. A., Kain, N., Daniels, B., Lim, X. Y., Shepherd, C., Kril, J., et al. (2014). Loss of the neuroprotective factor Sphingosine 1-phosphate early in Alzheimer's disease pathogenesis. *Acta Neuropathol. Commun.* 2:9. doi: 10.1186/2051-5960-2-9
- Couttas, T. A., Kain, N., Suchowerska, A. K., Quek, L. E., Turner, N., Fath, T., et al. (2016). Loss of ceramide synthase 2 activity, necessary for myelin biosynthesis, precedes tau pathology in the cortical pathogenesis of Alzheimer's disease. *Neurobiol. Aging* 43, 89–100. doi: 10.1016/j.neurobiolaging.2016.03.027
- Cutler, R. G., Kelly, J., Storie, K., Pedersen, W. A., Tammara, A., Hatanpaa, K., et al. (2004). Involvement of oxidative stress-induced abnormalities in ceramide and

cholesterol metabolism in brain aging and Alzheimer's disease. *Proc. Natl. Acad. Sci. U.S.A.* 101, 2070–2075. doi: 10.1073/pnas.0305799101

- Czubowicz, K., and Strosznajder, R. (2014). Ceramide in the molecular mechanisms of neuronal cell death. The role of sphingosine-1-phosphate. *Mol. Neurobiol.* 50, 26–37. doi: 10.1007/s12035-013-8606-4
- Deogracias, R., Yazdani, M., Dekkers, M. P., Guy, J., Ionescu, M. C., Vogt, K. E., et al. (2012). Fingolimod, a sphingosine-1 phosphate receptor modulator, increases BDNF levels and improves symptoms of a mouse model of Rett syndrome. *Proc. Natl. Acad. Sci. U.S.A.* 109, 14230–14235. doi: 10.1073/pnas.1206093109
- Dickson, D. W., Fujishiro, H., Orr, C., Delledonne, A., Josephs, K. A., Frigerio, R., et al. (2009). Neuropathology of non-motor features of Parkinson disease. *Parkinsonism Relat. Disord.* 15(Suppl. 3), S1–S5. doi: 10.1016/S1353-8020(09)70769-2
- Dinkins, M. B., Dasgupta, S., Wang, G., Zhu, G., He, Q., Kong, J. N., et al. (2015). The 5XFAD mouse model of Alzheimer's disease exhibits an age-dependent increase in anti-ceramide IgG and exogenous administration of ceramide further increases anti-ceramide titers and amyloid plaque burden. *J. Alzheimers. Dis.* 46, 55–61. doi: 10.3233/JAD-150088
- Di Pardo, A., Amico, E., Basit, A., Armirotti, A., Joshi, P., Neely, D. M., et al. (2017a). Defective sphingosine-1-phosphate metabolism is a druggable target in Huntington's disease. *Sci. Rep.* 7:5280. doi: 10.1038/s41598-017-05709-y
- Di Pardo, A., Amico, E., Favellato, M., Castrataro, R., Fucile, S., Squitieri, F., et al. (2014). FTY720 (fingolimod) is a neuroprotective and disease-modifying agent in cellular and mouse models of Huntington disease. *Hum. Mol. Genet.* 23, 2251–2265. doi: 10.1093/hmg/ddt615
- Di Pardo, A., Basit, A., Armirotti, A., Amico, E., Castaldo, S., Pepe, G., et al. (2017b). *De novo* synthesis of sphingolipids is defective in experimental models of Huntington's Disease. *Front. Neurosci.* 11:698. doi: 10.3389/fnins.2017.00698
- Fabelo, N., Martín, V., Santpere, G., Marin, R., Torrent, L., Ferrer, I., et al. (2011). Severe alterations in lipid composition of frontal cortex lipid rafts from Parkinson's disease and incidental Parkinson's disease. *Mol. Med.* 17, 1107–1118. doi: 10.2119/molmed.2011.00119
- Fahn, S. (2003). Description of Parkinson's disease as a clinical syndrome. *Ann. N.Y. Acad. Sci.* 991, 1–14. doi: 10.1111/j.1749-6632.2003.tb07458.x
- Ferrazza, R., Cogo, S., Melrose, H., Bubacco, L., Greggio, E., Guella, G., et al. (2016). LRRK2 deficiency impacts ceramide metabolism in brain. *Biochem. Biophys. Res. Commun.* 478, 1141–1146. doi: 10.1016/j.bbrc.2016.08.082
- Filippov, V., Song, M. A., Zhang, K., Vinters, H. V., Tung, S., Kirsch, W. M., et al. (2012). Increased ceramide in brains with Alzheimer's and other neurodegenerative diseases. J. Alzheimers. Dis. 29, 537–547. doi: 10.3233/JAD-2011-111202
- Gai, W. P., Yuan, H. X., Li, X. Q., Power, J. T., Blumbergs, P. C., and Jensen, P. H. (2000). *In situ* and *in vitro* study of colocalization and segregation of alpha-synuclein, ubiquitin, and lipids in lewy bodies. *Exp. Neurol.* 166, 324–333. doi: 10.1006/exnr.2000.7527
- Gault, C. R., Obeid, L. M., and Hannun, Y. A. (2010). An overview of sphingolipid metabolism: from synthesis to breakdown. *Adv. Exp. Med. Biol.* 688, 1–23. doi: 10.1007/978-1-4419-6741-1_1
- Gomez, L., Paillard, M., Price, M., Chen, Q., Teixeira, G., Spiegel, S., et al. (2011). A novel role for mitochondrial sphingosine-1-phosphate produced by sphingosine kinase-2 in PTP-mediated cell survival during cardioprotection. *Basic Res. Cardiol.* 106, 1341–1353. doi: 10.1007/s00395-011-0223-7
- Goñi, F. M., Sot, J., and Alonso, A. (2014). Biophysical properties of sphingosine, ceramides and other simple sphingolipids. *Biochem. Soc. Trans.* 42, 1401–1408. doi: 10.1042/BST20140159
- Gouras, G. K., Tampellini, D., Takahashi, R. H., and Capetillo-Zarate, E. (2010). Intraneuronal beta-amyloid accumulation and synapse pathology in Alzheimer's disease. *Acta Neuropathol.* 119, 523–541. doi: 10.1007/s00401-010-0679-9
- Hait, N. C., Allegood, J., Maceyka, M., Strub, G. M., Harikumar, K. B., Singh, S. K., et al. (2009). Regulation of histone acetylation in the nucleus by sphingosine-1phosphate. *Science* 325, 1254–1257. doi: 10.1126/science.1176709
- Halliday, G. M., Ophof, A., Broe, M., Jensen, P. H., Kettle, E., Fedorow, H., et al. (2005). Alpha-synuclein redistributes to neuromelanin lipid in the substantia nigra early in Parkinson's disease. *Brain* 128, 2654–2664. doi: 10.1093/brain/awh584

- Han, X., D, M.H., Mckeel, D. W. Jr., Kelley, J., and Morris, J. C. (2002). Substantial sulfatide deficiency and ceramide elevation in very early Alzheimer's disease: potential role in disease pathogenesis. *J. Neurochem.* 82, 809–818. doi: 10.1046/j.1471-4159.2002.00997.x
- Hatoum, D., Haddadi, N., Lin, Y., Nassif, N. T., and McGowan, E. M. (2017). Mammalian sphingosine kinase (SphK) isoenzymes and isoform expression: challenges for SphK as an oncotarget. *Oncotarget* 8, 36898–36929. doi: 10.18632/oncotarget.16370
- Haughey, N. J. (2010). Sphingolipids in neurodegeneration. *Neuromole. Med.* 12, 301–305. doi: 10.1007/s12017-010-8135-5
- He, X., Huang, Y., Li, B., Gong, C. X., and Schuchman, E. H. (2010). Deregulation of sphingolipid metabolism in Alzheimer's disease. *Neurobiol. Aging* 31, 398–408. doi: 10.1016/j.neurobiolaging.2008.05.010
- Hemmati, F., Dargahi, L., Nasoohi, S., Omidbakhsh, R., Mohamed, Z., Chik, Z., et al. (2013). Neurorestorative effect of FTY720 in a rat model of Alzheimer's disease: comparison with memantine. *Behav. Brain Res.* 252, 415–421. doi: 10.1016/j.bbr.2013.06.016
- Hla, T., Venkataraman, K., and Michaud, J. (2008). The vascular S1P gradientcellular sources and biological significance. *Biochim. Biophys. Acta* 1781, 477–482. doi: 10.1016/j.bbalip.2008.07.003
- Hoeferlin, L. A., Wijesinghe, D. S., and Chalfant, C. E. (2013). The role of ceramide-1-phosphate in biological functions. *Handb. Exp. Pharmacol.* 215, 153–166. doi: 10.1007/978-3-7091-1368-4_8
- Holmes, D. (2015). Bone: key role for S1P in bone remodelling. Nat. Rev. Endocrinol. 11:3. doi: 10.1038/nrendo.2014.190
- Huang, W. C., Nagahashi, M., Terracina, K. P., and Takabe, K. (2013). Emerging role of Sphingosine-1-phosphate in inflammation, cancer, and lymphangiogenesis. *Biomolecules* 3, 408–434. doi: 10.3390/biom3030408
- Jana, A., Hogan, E. L., and Pahan, K. (2009). Ceramide and neurodegeneration: susceptibility of neurons and oligodendrocytes to cell damage and death. J. Neurol. Sci. 278, 5–15. doi: 10.1016/j.jns.2008.12.010
- Ji, F., Mao, L., Liu, Y., Cao, X., Xie, Y., Wang, S., et al. (2015). K6PC-5, a novel sphingosine kinase 1 (SphK1) activator, alleviates dexamethasone-induced damages to osteoblasts through activating SphK1-Akt signaling. *Biochem. Biophys. Res. Commun.* 458, 568–575. doi: 10.1016/j.bbrc.2015.02.007
- Jimenez-Sanchez, M., Licitra, F., Underwood, B. R., and Rubinsztein, D. C. (2017). Huntington's disease: mechanisms of pathogenesis and therapeutic strategies. *Cold Spring Harb. Perspect. Med.* 7:a024240. doi: 10.1101/cshperspect.a024240
- Katsel, P., Li, C., and Haroutunian, V. (2007). Gene expression alterations in the sphingolipid metabolism pathways during progression of dementia and Alzheimer's disease: a shift toward ceramide accumulation at the earliest recognizable stages of Alzheimer's disease? *Neurochem. Res.* 32, 845–856. doi: 10.1007/s11064-007-9297-x
- Kornhuber, J., Tripal, P., Reichel, M., Mühle, C., Rhein, C., Muehlbacher, M., et al. (2010). Functional Inhibitors of Acid Sphingomyelinase (FIASMAs): a novel pharmacological group of drugs with broad clinical applications. *Cell. Physiol. Biochem.* 26, 9–20. doi: 10.1159/000315101
- Kumar, A., Byun, H. S., Bittman, R., and Saba, J. D. (2011). The sphingolipid degradation product trans-2-hexadecenal induces cytoskeletal reorganization and apoptosis in a JNK-dependent manner. *Cell. Signal.* 23, 1144–1152. doi: 10.1016/j.cellsig.2011.02.009
- Kumar, A., Singh, A., and Ekavali (2015). A review on Alzheimer's disease pathophysiology and its management: an update. *Pharmacol. Rep.* 67, 195–203. doi: 10.1016/j.pharep.2014.09.004
- Kwon, Y. B., Kim, C. D., Youm, J. K., Gwak, H. S., Park, B. D., Lee, S. H., et al. (2007). Novel synthetic ceramide derivatives increase intracellular calcium levels and promote epidermal keratinocyte differentiation. *J. Lipid Res.* 48, 1936–1943. doi: 10.1194/jlr.M700185-JLR200
- Lee, J. T., Xu, J., Lee, J. M., Ku, G., Han, X., Yang, D. I., et al. (2004). Amyloid-beta peptide induces oligodendrocyte death by activating the neutral sphingomyelinase-ceramide pathway. *J. Cell Biol.* 164, 123–131. doi: 10.1083/jcb.200307017
- Lépine, S., Allegood, J. C., Park, M., Dent, P., Milstien, S., and Spiegel, S. (2011). Sphingosine-1-phosphate phosphohydrolase-1 regulates ER stress-induced autophagy. *Cell Death Differ*. 18, 350–361. doi: 10.1038/cdd.2010.104
- Le Stunff, H., Peterson, C., Liu, H., Milstien, S., and Spiegel, S. (2002). Sphingosine-1-phosphate and lipid phosphohydrolases. *Biochim. Biophys. Acta* 1582, 8–17. doi: 10.1016/S1388-1981(02)00132-4

- Levenson, R. W., Sturm, V. E., and Haase, C. M. (2014). Emotional and behavioral symptoms in neurodegenerative disease: a model for studying the neural bases of psychopathology. *Annu. Rev. Clin. Psychol.* 10, 581–606. doi: 10.1146/annurev-clinpsy-032813-153653
- Levy, M., and Futerman, A. H. (2010). Mammalian ceramide synthases. *IUBMB Life* 62, 347–356. doi: 10.1002/iub.319
- Li, J. Q., Tan, L., and Yu, J. T. (2014). The role of the LRRK2 gene in Parkinsonism. Mol. Neurodegener. 9:47. doi: 10.1186/1750-1326-9-47
- Maceyka, M., Harikumar, K. B., Milstien, S., and Spiegel, S. (2012). Sphingosine-1-phosphate signaling and its role in disease. *Trends Cell Biol.* 22, 50–60. doi: 10.1016/j.tcb.2011.09.003
- Maceyka, M., Sankala, H., Hait, N. C., Le Stunff, H., Liu, H., Toman, R., et al. (2005). SphK1 and SphK2, sphingosine kinase isoenzymes with opposing functions in sphingolipid metabolism. *J. Biol. Chem.* 280, 37118–37129. doi: 10.1074/jbc.M502207200
- Maglione, V., Cannella, M., Gradini, R., Cislaghi, G., and Squitieri, F. (2006a). Huntingtin fragmentation and increased caspase 3, 8 and 9 activities in lymphoblasts with heterozygous and homozygous Huntington's disease mutation. *Mech. Ageing Dev.* 127, 213–216. doi: 10.1016/j.mad.2005.09.011
- Maglione, V., Cannella, M., Martino, T., De Blasi, A., Frati, L., and Squitieri, F. (2006b). The platelet maximum number of A2A-receptor binding sites (Bmax) linearly correlates with age at onset and CAG repeat expansion in Huntington's disease patients with predominant chorea. *Neurosci. Lett.* 393, 27–30. doi: 10.1016/j.neulet.2005.09.037
- Maglione, V., Giallonardo, P., Cannella, M., Martino, T., Frati, L., and Squitieri, F. (2005). Adenosine A2A receptor dysfunction correlates with age at onset anticipation in blood platelets of subjects with Huntington's disease. Am. J. Med. Genet. B Neuropsychiatr. Genet. 139B, 101–105. doi: 10.1002/ajmg.b.30223
- Martin, R., and Sospedra, M. (2014). Sphingosine-1 phosphate and central nervous system. *Curr. Top. Microbiol. Immunol.* 378, 149–170. doi: 10.1007/978-3-319-05879-5_7
- Mccolgan, P., and Tabrizi, S. J. (2018). Huntington's disease: a clinical review. *Eur. J. Neurol.* 25, 24–34. doi: 10.1111/ene.13413
- Mendelson, K., Evans, T., and Hla, T. (2014). Sphingosine 1-phosphate signalling. Development 141, 5–9. doi: 10.1242/dev.094805
- Mesev, E. V., Miller, D. S., and Cannon, R. E. (2017). Ceramide 1phosphate increases P-Glycoprotein transport activity at the blood-brain barrier via prostaglandin E2 signaling. *Mol. Pharmacol.* 91, 373–382. doi: 10.1124/mol.116.107169
- Mielke, M. M., and Lyketsos, C. G. (2010). Alterations of the sphingolipid pathway in Alzheimer's disease: new biomarkers and treatment targets? *Neuromolecular Med.* 12, 331–340. doi: 10.1007/s12017-010-8121-y
- Miguez, A., García-Díaz, G., Brito, V., Straccia, M., Giralt, A., Ginés, S., et al. (2015). Fingolimod (FTY720) enhances hippocampal synaptic plasticity and memory in Huntington's disease by preventing p75NTR up-regulation and astrocyte-mediated inflammation. *Hum. Mol. Genet.* 24, 4958–4970. doi: 10.1093/hmg/ddv218
- Miranda, G. E., Abrahan, C. E., Agnolazza, D. L., Politi, L. E., and Rotstein, N. P. (2011). Ceramide-1-phosphate, a new mediator of development and survival in retina photoreceptors. *Invest. Ophthalmol. Vis. Sci.* 52, 6580–6588. doi: 10.1167/iovs.10-7065
- Morozov, V. I., Sakuta, G. A., and Kalinski, M. I. (2013). Sphingosine-1-phosphate: distribution, metabolism and role in the regulation of cellular functions. Ukr. Biokhim. Zh. 85, 5–21. doi: 10.15407/ubj85.01.005
- Moruno-Manchon, J. F., Uzor, N. E., Blasco-Conesa, M. P., Mannuru, S., Putluri, N., Furr-Stimming, E. E., et al. (2017). Inhibiting sphingosine kinase 2 mitigates mutant Huntingtin-induced neurodegeneration in neuron models of Huntington disease. *Hum. Mol. Genet.* 26, 1305–1317. doi: 10.1093/hmg/ddx046
- Moruno Manchon, J. F., Uzor, N. E., Dabaghian, Y., Furr-Stimming, E. E., Finkbeiner, S., and Tsvetkov, A. S. (2015). Cytoplasmic sphingosine-1-phosphate pathway modulates neuronal autophagy. *Sci. Rep.* 5:15213. doi: 10.1038/srep15213
- Mullen, T. D., Hannun, Y. A., and Obeid, L. M. (2012). Ceramide synthases at the centre of sphingolipid metabolism and biology. *Biochem. J.* 441, 789–802. doi: 10.1042/BJ20111626
- Mullen, T. D., and Obeid, L. M. (2012). Ceramide and apoptosis: exploring the enigmatic connections between sphingolipid metabolism and

programmed cell death. Anticancer. Agents Med. Chem. 12, 340-363. doi: 10.2174/187152012800228661

- Obeso, J. A., Rodriguez-Oroz, M. C., Goetz, C. G., Marin, C., Kordower, J. H., Rodriguez, M., et al. (2010). Missing pieces in the Parkinson's disease puzzle. *Nat. Med.* 16, 653–661. doi: 10.1038/nm.2165
- Olsen, A. S. B., and Færgeman, N. J. (2017). Sphingolipids: membrane microdomains in brain development, function and neurological diseases. *Open Biol.* 7:170069. doi: 10.1098/rsob.170069
- O'Regan, G., Desouza, R. M., Balestrino, R., and Schapira, A. H. (2017). Glucocerebrosidase mutations in Parkinson disease. *J. Parkinsons. Dis.* 7, 411–422. doi: 10.3233/JPD-171092
- O'Sullivan, S., and Dev, K. K. (2017). Sphingosine-1-phosphate receptor therapies: advances in clinical trials for CNS-related diseases. *Neuropharmacology* 113, 597–607. doi: 10.1016/j.neuropharm.2016.11.006
- Park, K., Ikushiro, H., Seo, H. S., Shin, K. O., Kim, Y. I., Kim, J. Y., et al. (2016). ER stress stimulates production of the key antimicrobial peptide, cathelicidin, by forming a previously unidentified intracellular S1P signaling complex. *Proc. Natl. Acad. Sci. U.S.A.* 113, E1334–E1342. doi: 10.1073/pnas.1504555113
- Pirhaji, L., Milani, P., Dalin, S., Wassie, B. T., Dunn, D. E., Fenster, R. J., et al. (2017). Identifying therapeutic targets by combining transcriptional data with ordinal clinical measurements. *Nat. Commun.* 8:623. doi: 10.1038/s41467-017-00353-6
- Pirhaji, L., Milani, P., Leidl, M., Curran, T., Avila-Pacheco, J., Clish, C. B., et al. (2016). Revealing disease-associated pathways by network integration of untargeted metabolomics. *Nat. Methods* 13, 770–776. doi: 10.1038/nmeth.3940
- Pyszko, J. A., and Strosznajder, J. B. (2014). The key role of sphingosine kinases in the molecular mechanism of neuronal cell survival and death in an experimental model of Parkinson's disease. *Folia Neuropathol.* 52, 260–269. doi: 10.5114/fn.2014.45567
- Riccio, A. (2010). New endogenous regulators of class I histone deacetylases. *Sci. Signal.* 3:pe1. doi: 10.1126/scisignal.3103pe1
- Robins Wahlin, T. B., and Byrne, G. J. (2011). Personality changes in Alzheimer's disease: a systematic review. Int. J. Geriatr. Psychiatry 26, 1019–1029. doi: 10.1002/gps.2655
- Schwarz, A., and Futerman, A. H. (1997). Distinct roles for ceramide and glucosylceramide at different stages of neuronal growth. J. Neurosci. 17, 2929–2938. doi: 10.1523/JNEUROSCI.17-09-02929.1997
- Serra, M., and Saba, J. D. (2010). Sphingosine 1-phosphate lyase, a key regulator of sphingosine 1-phosphate signaling and function. Adv. Enzyme Regul. 50, 349–362. doi: 10.1016/j.advenzreg.2009.10.024
- Shen, Z., Liu, C., Liu, P., Zhao, J., and Xu, W. (2014). Sphingosine 1-phosphate (S1P) promotes mitochondrial biogenesis in Hep G2 cells by activating Peroxisome proliferator-activated receptor gamma coactivator 1alpha (PGC-1alpha). Cell Stress Chaperones 19, 541–548. doi: 10.1007/s12192-013-0480-5
- Sivasubramanian, M., and Tay Ssw D. S. (2013). Alteration in the sphingolipid metabolism leads to activation of the apoptotic cascade in the MPTP induced mouse model of Parkinson's disease *FASEB J.* 27, 533–538.
- Sivasubramanian, M., Kanagaraj, N., Dheen, S. T., and Tay, S. S. (2015). Sphingosine kinase 2 and sphingosine-1-phosphate promotes mitochondrial function in dopaminergic neurons of mouse model of Parkinson's disease and in MPP+ -treated MN9D cells *in vitro*. *Neuroscience* 290, 636–648. doi: 10.1016/j.neuroscience.2015.01.032
- Sona, A., Ellis, K. A., and Ames, D. (2013). Rapid cognitive decline in Alzheimer's disease: a literature review. *Int. Rev. Psychiatry* 25, 650–658. doi: 10.3109/09540261.2013.859128

- Strub, G. M., Maceyka, M., Hait, N. C., Milstien, S., and Spiegel, S. (2010). Extracellular and intracellular actions of sphingosine-1-phosphate. Adv. Exp. Med. Biol. 688, 141–155. doi: 10.1007/978-1-4419-6741-1_10
- Strub, G. M., Paillard, M., Liang, J., Gomez, L., Allegood, J. C., Hait, N. C., et al. (2011). Sphingosine-1-phosphate produced by sphingosine kinase 2 in mitochondria interacts with prohibitin 2 to regulate complex IV assembly and respiration. *FASEB J.* 25, 600–612. doi: 10.1096/fj.10-167502
- Tada, E., Toyomura, K., Nakamura, H., Sasaki, H., Saito, T., Kaneko, M., et al. (2010). Activation of ceramidase and ceramide kinase by vanadate via a tyrosine kinase-mediated pathway. *J. Pharmacol. Sci.* 114, 420–432. doi: 10.1254/jphs.10181FP
- Takasugi, N., Sasaki, T., Ebinuma, I., Osawa, S., Isshiki, H., Takeo, K., et al. (2013). FTY720/fingolimod, a sphingosine analogue, reduces amyloid-beta production in neurons. *PLoS ONE* 8:e64050. doi: 10.1371/journal.pone.0064050
- Takasugi, N., Sasaki, T., Suzuki, K., Osawa, S., Isshiki, H., Hori, Y., et al. (2011). BACE1 activity is modulated by cell-associated sphingosine-1-phosphate. J. Neurosci. 31, 6850–6857. doi: 10.1523/JNEUROSCI.6467-10.2011
- Tohyama, J., Oya, Y., Ezoe, T., Vanier, M. T., Nakayasu, H., Fujita, N., et al. (1999). Ceramide accumulation is associated with increased apoptotic cell death in cultured fibroblasts of sphingolipid activator protein-deficient mouse but not in fibroblasts of patients with Farber disease. J. Inherit. Metab. Dis. 22, 649–662. doi: 10.1023/A:1005590316064
- Toman, R. E., Payne, S. G., Watterson, K. R., Maceyka, M., Lee, N. H., Milstien, S., et al. (2004). Differential transactivation of sphingosine-1-phosphate receptors modulates NGF-induced neurite extension. J. Cell Biol. 166, 381–392. doi: 10.1083/jcb.200402016
- van Echten-Deckert, G., Hagen-Euteneuer, N., Karaca, I., and Walter, J. (2014). Sphingosine-1-phosphate: boon and bane for the brain. *Cell. Physiol. Biochem.* 34, 148–157. doi: 10.1159/000362991
- Xiong, Y., and Hla, T. (2014). S1P control of endothelial integrity. Curr. Top. Microbiol. Immunol. 378, 85–105. doi: 10.1007/978-3-319-05879-5_4
- Ye, Y., Zhao, Z., Xu, H., Zhang, X., Su, X., Yang, Y., et al. (2016). Activation of sphingosine 1-phosphate receptor 1 enhances hippocampus neurogenesis in a rat model of Traumatic brain injury: an involvement of MEK/Erk signaling pathway. *Neural Plast.* 2016:8072156. doi: 10.1155/2016/80 72156
- Zhao, P., Yang, X., Yang, L., Li, M., Wood, K., Liu, Q., et al. (2017). Neuroprotective effects of fingolimod in mouse models of Parkinson's disease. *FASEB J.* 31, 172–179. doi: 10.1096/fj.201600751R
- Zheng, W., Kollmeyer, J., Symolon, H., Momin, A., Munter, E., Wang, E., et al. (2006). Ceramides and other bioactive sphingolipid backbones in health and disease: lipidomic analysis, metabolism and roles in membrane structure, dynamics, signaling and autophagy. *Biochim. Biophys. Acta* 1758, 1864–1884. doi: 10.1016/j.bbamem.2006.08.009

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Di Pardo and Maglione. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.