



OPEN ACCESS

Edited by:

Robert W. Williams,
The University of Tennessee Health
Science Center (UTHSC),
United States

Reviewed by:

Rossen Donev,
MicroPharm Ltd., United Kingdom
Jun Xu,
Capital Medical University, China

***Correspondence:**

Lan Tan
dr.tanlan@163.com

[†]The data used in the preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in the analysis or writing of this report.

A complete listing of ADNI investigators can be found at: http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf

Specialty section:

This article was submitted to Neurogenomics, a section of the journal *Frontiers in Neuroscience*

Received: 14 September 2018

Accepted: 04 July 2019

Published: 17 July 2019

Citation:

Tian M-L, Ni X-N, Li J-Q, Tan C-C, Cao X-P and Tan L for the Alzheimer's Disease Neuroimaging Initiative (2019) A Candidate Regulatory Variant at the *TREM* Gene Cluster Confer Alzheimer's Disease Risk by Modulating Both Amyloid- β Pathology and Neuronal Degeneration. *Front. Neurosci.* 13:742. doi: 10.3389/fnins.2019.00742

A Candidate Regulatory Variant at the *TREM* Gene Cluster Confer Alzheimer's Disease Risk by Modulating Both Amyloid- β Pathology and Neuronal Degeneration

Mei-Ling Tian¹, Xiao-Neng Ni², Jie-Qiong Li¹, Chen-Chen Tan¹, Xi-Peng Cao³ and Lan Tan^{2,3*} for the Alzheimer's Disease Neuroimaging Initiative[†]

¹ Department of Neurology, Qingdao Municipal Hospital, Qingdao University, Qingdao, China, ² Department of Neurology, Qingdao Municipal Hospital, Nanjing Medical University, Nanjing, China, ³ Clinical Research Center, Qingdao Municipal Hospital, Qingdao University, Qingdao, China

Background: rs9357347 located at the triggering receptor expressed on myeloid cells (*TREM*) gene cluster could increase *TREM2* and *TREM*-like transcript 1 (*TREML1*) brain gene expression, which is considered to play a protective role against Alzheimer's disease (AD).

Objectives: To investigate the role of rs9357347 in AD pathogenesis by exploring the effects of rs9357347 on AD specific biomarkers.

Methods: This study analyzed the association of rs9357347 with AD-related cerebrospinal fluid (CSF) and neuroimaging markers from 201 cognitively normal (CN) older adults, 349 elders with mild cognitive impairment (MCI), and 172 elders with AD dementia from the Alzheimer's Disease Neuroimaging Initiative (ADNI). We next analyzed the association in 259 amyloid- β positive ($A\beta+$) elders and 117 amyloid- β negative ($A\beta-$) elders ($A\beta+$: CSF $A\beta_{1-42} \leq 192$ pg/ml; $A\beta-$: CSF $A\beta_{1-42} > 192$ pg/ml). Associations were tested using multiple linear regression models at baseline. Furthermore, multiple mixed-effects models were used in a longitudinal study which lasted 4 years.

Results: At baseline, we found that rs9357347 had association with CSF $A\beta_{1-42}$ in CN group ($\beta = 0.357$, $P = 0.009$). In AD group, rs9357347 was associated with total tau (T-tau) level ($\beta = -0.436$, $P = 0.007$). Moreover, the strong influence exerted by rs9357347 on T-tau was also seen in $A\beta+$ group ($\beta = -0.202$, $P = 0.036$). In the longitudinal study, rs9357347 was also found to be associated with $A\beta_{1-42}$ in CN group ($\beta = 0.329$, $P = 0.023$). In AD group, the mutation of rs9357347 was associated with slower accumulation of T-tau ($\beta = -0.472$, $P = 0.002$) and tau phosphorylated at

threonine 181 [P-tau 181 ($\beta = -0.330$, $P = 0.019$)]. Furthermore, the obvious influence exerted by rs9357347 on T-tau was also seen in A β + group ($\beta = -0.241$, $P = 0.013$).

Conclusion: This study suggested that rs9357347 reduced the risk of AD by modulating both amyloid- β pathology and neuronal degeneration.

Keywords: Alzheimer's disease, *TREM2* gene, *TREML1*, rs9357347, genetic mechanism, cerebrospinal fluid, amyloid- β pathology, neuronal degeneration

INTRODUCTION

Alzheimer's disease (AD) is a complex polygenetic disease characterized by the presence of extracellular deposits of the amyloid- β_{1-42} (A β_{1-42}) and intracellular twisted strands of the tau protein (Alzheimer's Association, 2011). In the genetic studies, the triggering receptor expressed on myeloid cells (*TREM*) gene cluster on chromosome 6p21.11 has been identified as significantly associated gene region with AD (Karch et al., 2014). Among the *TREM* genes, a rare variant rs75932628 (encoding p. Arg47His) in *TREM2* was reported to be associated with the highest risk of developing AD in Caucasians (Colonna, 2003; Piccio et al., 2007).

Recently, Carrasquillo et al. (2017) re-analyzed whole genome and exome sequencing data to test which common AD-related variants within the *TREM* gene cluster influence AD through gene expression. rs9357347, located downstream from *TREML2* and upstream from *TREM2*, was found to be associated with AD risk (Carrasquillo et al., 2017). And the locus was demonstrated to influence *TREML1* and *TREM2* expression in the temporal cortex (Ford and McVicar, 2009). Meanwhile, they found that only *TREM2* and *TREML1* had reliable expression in the brain region. Thus, they point out that rs9357347^{CC} exerts protective effects on AD through increasing *TREML1* and *TREM2* brain expression levels. However, the specific pathogenic effect rs9357347 exerts on AD remains unclear. Therefore, the current study was to test associations of rs9357347 with AD-related cerebrospinal fluid (CSF) and neuroimaging markers in the population from Alzheimer's Disease Neuroimaging Initiative (ADNI).

MATERIALS AND METHODS

Study Design and Participants

Data used in this study were obtained from the ADNI database led by Principal Investigator Michael W. Weiner, MD, which is a public-private partnership, launched in 2003. The main goal of ADNI has been to test whether clinical and cognitive assessment, positron emission tomography (PET), CSF, serial magnetic resonance imaging (MRI), and other biological markers can be combined to evaluate the progression of mild cognitive impairment (MCI) and early AD. Written informed consent was obtained from all participants or their guardians before test samples being drawn. The institutional review boards of all sites participating in the ADNI provided review and approval of the ADNI data collection protocol. For more details, see www.adni-info.org.

Our ADNI cohort consisted of available baseline and longitudinal AD-related marker samples from all cognitively normal (CN) elders, patients with MCI, and patients with AD dementia. Inclusion or exclusion criteria are described in detail on pages 20–22 of the online ADNI protocol. Briefly, all subjects were aged from 55 to 90 years old, kept contact with their study partners 10 h per week or more, had completed at least six grades of education or had a good employment history, spoke English or Spanish fluently, and were removed of any significant neurological disease other than AD (Petersen et al., 2010). Any history of head trauma or brain lesions, any serious neurological disease other than probable AD, and any psychoactive medication use that could otherwise account for the deterioration in memory and related symptoms must be excluded (Jagust et al., 2009; Weigand et al., 2011).

We used two classification criteria for the 722 ADNI samples to study the association of rs9357347 allele with AD dementia. One is clinical classification (AD, MCI, CN groups) which includes 201 CN older adults, 349 elders with MCI, and 172 elders with AD dementia. And another is pathological classification classifying the subjects into two groups according to previously established cutoff. It has been shown that individuals with CSF A β_{1-42} levels less than 192 pg/ml in the ADNI cohort have evidence of A β_{1-42} deposition in the brain, as detected by PET-PIB (Fagan et al., 2007; Mattsson et al., 2009). Individuals with CSF A β_{1-42} levels below these thresholds (CSF A $\beta_{1-42} \leq 192$ pg/ml) could be classified as A β -positive (A β +), otherwise (CSF A $\beta_{1-42} > 192$ pg/ml) they will be classified as A β -negative (A β -) (Olsson et al., 2005; Jack et al., 2008; Shaw et al., 2009). Thus, 259 A β + elders and 117 A β - elders were selected from ADNI database.

CSF Biomarkers

The CSF data used in this study were obtained from ADNI dataset. CSF sampled by lumbar puncture was gathered into collection tubes, and then transferred to polypropylene conveying tubes. After collection, the samples were frozen on dry ice within 60 min and transported immediately to the ADNI Biomarker Core laboratory at the University of Pennsylvania Medical Center. Preparation of aliquots (0.5 ml) from these samples was done after unfreezing (60 min) at room temperature and gentle mixing (Weigand et al., 2011). The aliquots were stored in bar code-labeled polypropylene vials at -80°C (Shaw et al., 2009). The multiplex platform (xMAP; Luminex Corporation) with a kit (INNO-BIA AlzBio3; Fujirebio Europe) was used for simultaneous measurement of the CSF protein biomarkers such as A β_{1-42} , total tau (T-tau), and tau

phosphorylated at threonine 181 (P-tau 181). More details for CSF acquisition and measurement have been reported previously (Olsson et al., 2005).

Brain Structures on MRI

The volumes of brain structures in MRI used in our study were from the UCSF data in ADNI dataset¹. Firstly, we downloaded preprocessed MRI data from Laboratory of Neuroimaging (LONI) IDA² with 1.5T or 3T data available. Then cerebral image analysis and segmentation were performed with the Free Surfer version 5.1 including corrected for motion, averaged, normalized for intensity. The technical details of these procedures are described in prior publications (Jack et al., 2008). Here, we selected the most associated brain regions with AD, such as hippocampus, ventricle and middle temporal as our regions of interest (ROI).

Glucose Metabolism on FDG-PET

The information related to glucose metabolism was from the UC Berkeley and Lawrence Berkeley National Laboratory (Landau et al., 2011). The brief routine processes were as follows. First of all, PET data were downloaded from LONI website³. Then the mean counts from the meta ROIs (left and right inferior/middle temporal gyrus, left and right angular gyri, and bilateral posterior cingulate gyrus) for each subject's FDG scans at each time point were extracted and the intensity values were calculated with SPM5 subroutines. Finally, mean FDG uptake was extracted for each of the five ROIs and normalized by dividing it by pons/vermis reference region mean (Landau et al., 2011). Total FDG uptake was calculated as a mean of the five individual Meta ROIs⁴.

Statistical Analysis

Associations between diagnosis and demographic, clinical factors were tested at baseline. We examined differences in continuous variables (education years, age, volume, etc.) using Kruskal–Wallis test. We tested differences in Categorical data (gender, APOE $\epsilon 4$ status) using chi-square test. The correlations between rs9357347 and various endophenotypes (CSF proteins, MRI and FDG-PET) were estimated using linear regression models at baseline. Furthermore, association between rs9357347 and the above phenotypes in the longitudinal study were tested using linear mixed-effects models. Of note, we chosen the 4-year follow-up data for all three existing genotypes to make analysis. We used mixed linear models that specified a random subject-specific intercept and a random subject-specific slope. All outcome variables in linear regression models and linear mixed-effects models were standardized to z scores to facilitate comparisons between genotypes. Difference with a P -value < 0.05 was considered to be statistically significant. All regression analyses were corrected for age,

gender, APOE $\epsilon 4$ genotype and educational level, and the regression analysis of brain structure volume was also corrected for intracranial volume. All statistical analyses were performed by R3.2.0⁵.

RESULTS

Baseline Characteristics of Study Participants

The study population was composed of 201 CN elders, 349 elders with MCI, and 172 elders with AD. Demographic and clinical characteristics at the baseline were summarized in the **Table 1**. As expected, the frequency of the APOE $\epsilon 4$ allele in AD group was significantly higher than those in MCI and CN group. Among the three groups, the diagnosis was found to be correlated with gender, education level, volume of brain structure, and FDG. The diagnosis did not differ by age. Individuals in AD and MCI cohorts exhibited typical CSF biomarker phenotype of AD with elevated mean levels of T-tau and P-tau181 and lower level of $A\beta_{1-42}$.

Impacts of rs9357347 on CSF Markers

At baseline, we analyzed the correlations between rs9357347 and concentration of CSF proteins in three clinical groups (CN, MCI, and AD) and two pathological groups ($A\beta+$, $A\beta-$). In CN group, the significant correlation was seen with CSF $A\beta_{1-42}$ ($\beta = 0.36$, $P = 0.009$) (**Figure 1A**). In AD group, rs9357347 was found to be correlated with CSF T-tau ($\beta = -0.44$, $P = 0.007$) (**Figure 1B**). Meanwhile, we discovered possible relation between rs9357347 and CSF P-tau 181 in AD group ($\beta = -0.31$, $P = 0.051$) (**Figure 1C**). As for the pathological group, we also found the obvious association between rs9357347 and CSF T-tau in $A\beta+$ group ($\beta = -0.20$, $P = 0.036$) (**Figure 1D**).

In longitudinal study, the findings were similar to the results at baseline after adjusting for age, gender, education level, APOE $\epsilon 4$. In CN group, CSF $A\beta_{1-42}$ was found to be correlated with the locus ($\beta = 0.33$, $P = 0.023$) (**Figure 1E**). Furthermore, in AD group, rs9357347 was found to exert a strong effect on CSF T-tau ($\beta = -0.47$, $P = 0.002$) (**Figure 1F**) and P-tau 181 ($\beta = -0.33$, $P = 0.019$) (**Figure 1G**), while we failed to discover similar correlation in MCI and CN groups. And when the samples were stratified by pathological status, the C allele of rs9357347 was found to decrease the CSF T-tau ($\beta = -0.24$, $P = 0.013$) (**Figure 1H**) in $A\beta+$ group.

Impacts of rs9357347 on MRI Structure and Glucose Metabolism

Association of rs9357347 with AD-related neuroimaging measures and FDG-PET imaging were investigated at baseline and longitudinally. Neither AD-related brain structures (right and left hippocampus, middle temporal gyrus and ventricle) nor cerebral metabolism rate of glucose (CMRgl) on FDG-PET imaging was found to have obvious association with rs9357347.

¹<https://ida.loni.usc.edu/pages/access/studyData.jsp>

²<https://ida.loni.usc.edu>

³<http://loni.usc.edu/>

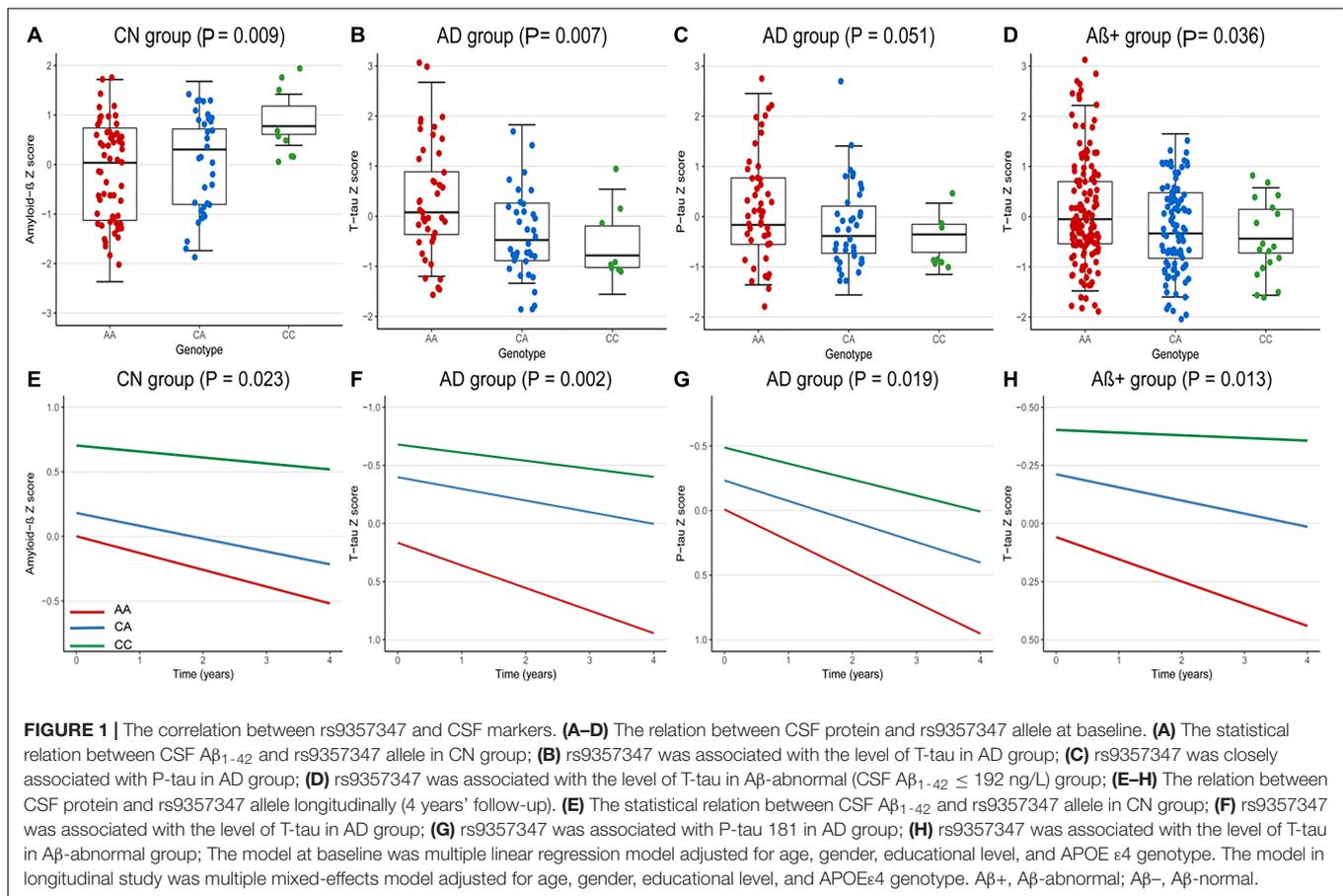
⁴www.adni-info.org

⁵<http://www.r-project.org/>

TABLE 1 | The characteristics of the subjects in clinical group at baseline.

Characteristics	CN (N = 201)	MCI (N = 349)	AD (N = 172)	P value
Age (years)	75.80(4.84)	74.76(7.34)	75.42(7.40)	0.56
Gender (male/female)	107/94	229/120	91/81	<0.01
Education (years)	16.08(2.82)	15.64(3.08)	14.64(3.17)	<0.01
ApoE ϵ 4 (0/1/2)	147/50/4	156/151/42	57/84/31	<0.01
A β (pg/ml)	206.33(54.35)	162.86(52.31)	143.39(38.09)	<0.01
T-tau (pg/ml)	69.85(30.25)	103.04(58.31)	121.07(56.80)	<0.01
P-tau (pg/ml)	25.39(14.80)	36.17(19.26)	41.92(20.27)	<0.01
FDG	1.29(0.12)	1.20(0.12)	1.09(0.13)	<0.01
Hippocampus (mm ³)	7279.21(888.23)	6381.58(1074.03)	5596.41(1063.70)	<0.01
Middle temporal (mm ³)	19921.81(2790.99)	18589.17(2912.60)	16827.31(3131.44)	<0.01
Ventricular (mm ³)	35088.02(19446.31)	44700.22(23944.76)	49504.77(25596.41)	<0.01

CN, cognition normal; MCI, mild cognitive impairment; AD, Alzheimer's disease; A β , β -amyloid; T-tau, total tau; P-tau, tau phosphorylated at threonine; FDG, F18-fluorodeoxyglucose. N indicates the number of subjects from each group with at least 1 follow-up measure available. Data are mean (standard error). p values for continuous variables are from Kruskal–Wallis test. p values for categorical data are from chi-square test.



DISCUSSION

In this study, we aimed to characterize the pathogenic effect rs9357347 exerted on AD based on the premise that rs9357347, a candidate regulatory variant at the *TREM* gene cluster, associates with decreased AD risk and increased *TREML1* and *TREM2* brain gene expression (Carrasquillo et al., 2017). We found

statistically significant associations with both A β_{1-42} and tau in CSF (at baseline and longitudinally). Specifically, we reported that rs9357347 was mainly associated with CSF A β_{1-42} in CN participants; in AD participants, the mutation was detected to be negatively associated with CSF T-tau, which is consistent with the result in A β + group; the association between the locus and CSF P-tau 181 was also found in AD group longitudinally.

To sum up, our results were in line with the hypothetical model that relates disease stage to AD biomarkers, in which CSF $A\beta_{1-42}$ biomarkers become abnormal first and then the neurodegenerative biomarkers (Hardy and Selkoe, 2002; Dennis and Selkoe, 2016). This suggests that there is an association of rs9357347 with brain amyloidosis and tau pathology of AD. This finding may supply clues to the protective role of rs9357347 against AD risk.

Previous studies have proposed a mechanism by which genetic variation in *TREM2* alters amyloid pathology. First of all, *TREM2* is a cell surface receptor of the immunoglobulin superfamily, which is expressed nearly exclusively on microglia within the central nervous system (CNS) (Zheng et al., 2016). Second, in CNS, *TREM2* signaling is intimately linked with the adapter protein, DAP12 (also known as TYRO protein tyrosine kinase binding protein, TYROBP) (Li and Zhang, 2018). *TREM2* deficiency is associated with decreased bacterial clearance and increased pro-inflammatory cytokine production, thus suggesting the anti-inflammatory and protective functions of *TREM2* (Zheng et al., 2016). The number of myeloid cells around amyloid plaques was decreased in *TREM2* hemizygous mutations showed that a loss of *TREM2* function results in reduced $A\beta_{1-42}$ uptake (Yuan et al., 2016). Further, a previous study found that microglia became more active to remove amyloid plaques after *TREM2* overexpression, and the density of amyloid plaques in the brain reduced in middle-aged APP/ PS1 mice (7–8 months old) (Jiang et al., 2014). Condello et al. (2018) postulated that the microglia around the amyloid surface limits fibril outgrowth and plaque-associated toxicity.

Many of studies suggested that *TREM2* participates in AD pathogenesis through intraneuronal deposition of phosphorylated tau besides $A\beta_{1-42}$ deposition and clearance (Jay et al., 2015; Jiang et al., 2015, 2018). Tau pathology in the CNS, current is involved in a series of neurodegenerative disorders including AD (Medina and Avila, 2014). Therefore, regulation of tau pathology may be an impactful approach to delaying the AD development. Mutations in *TREM2* have been suggested to be correlated with tau pathology in AD. Lill et al. (2015) reported that a variant of *TREM2* (rs75932628) significantly increased the accumulation of CSF T-tau in a European population, which suggested that *TREM2*'s role in AD may involve neuronal degeneration. additionally, the overexpression of *TREM2* has been found to significantly enhance hyperphosphorylation of tau proteins (Lill et al., 2015). Consequently, *TREM2* overexpression significantly improves neuronal loss and may play a role in the phosphorylation of tau protein, thereby improving the incidence of AD. Of note, inflammatory stimuli in the brain have also been proved to be associated with accelerating tau phosphorylation (Jiang et al., 2016). Since tau phosphorylation would be accelerated resulting from reduced *TREM2* levels suggesting that *TREM2* act as an anti-inflammatory factor. In the light of our result, we can hypothesize that the upregulation of *TREM2* driven by the rs9357347-C allele serves as a compensatory response to $A\beta_{1-42}$ and tau and subsequently protects against AD progression.

Currently, TREML1 (also named as TLT-1) has been identified as a myeloid receptor expressed exclusively in the α -granules of megakaryocytes and platelets (Washington et al., 2004). TREML1 was proved to antagonize proinflammatory activation of TREM1 by competing with its ligand (Derive et al., 2012). Besides, soluble TREML1 has also been shown to play an anti-inflammatory role. Meanwhile, *TREML1* levels in the brain have reported to have association with decreased AD risk in humans (Carrasquillo et al., 2017) suggesting that upregulated expression of *TREML1* may be protective against AD. Nevertheless, as we all know, no variant in TREML1 is currently exerted to associated with AD (Derive et al., 2012). For that reason, our results that rs9357347 exerted a significant effect on AD by modulating CSF biomarkers including $A\beta_{1-42}$, T-tau, and P-tau 181 has thrown light on the hypothesis about the mechanism through which TREML1 modifies AD risk, which still needs further investigation.

Our findings were consistent with the results of Carrasquillo et al. (2017) that rs9357347 has functional influence on AD and may indicated the role that rs9357347 played on protection from AD. The study demonstrated that the locus was associated with the level of $A\beta$ and tau. Thus far, it has been identified that both $A\beta$ and tau pathology could lead to neuronal dysfunction and neurodegeneration (Jack et al., 2010; Wang et al., 2015). Many studies suggested that *TREM2*'s role in AD may involve tau dysfunction and $A\beta$ deposition (Lill et al., 2015; Zheng et al., 2016). All the above evidence, along with our findings, supported that rs9357347 mediated AD risk by modulating the alteration of the amyloid- β pathology and neuronal degeneration biomarkers. Although our results showed evidence that $A\beta_{1-42}$ and tau response were in part mediated through rs9357347, there remained a possibility that tau effects may be also derived from an imbalance between $A\beta_{1-42}$ production and clearance (Hardy and Selkoe, 2002; Selkoe and Hardy, 2016). A detailed analysis of the potential significance of secondary changes is indeed interesting, so future studies are needed to robustly identify the locus for the associations with $A\beta_{1-42}$ and tau pathology.

CONCLUSION

In summary, our findings confirmed that the rs9357347^{CC} exerted a protective effect on AD by decreasing the $A\beta_{1-42}$ accumulation and tau burden. These findings further supported the hypothesis that *TREM2* and *TREML1* may modulate amyloid- β pathology and neuronal degeneration to influence the risk of AD.

AUTHOR CONTRIBUTIONS

J-QL and C-CT contributed to the conception and design of the study. M-LT and X-NN performed the statistical analysis. M-LT wrote the manuscript. X-PC, J-QL, C-CT, and LT contributed to the manuscript revision, read and approved the submitted version of the manuscript.

FUNDING

This work was supported by grants from the National Key R&D Program of China (2016YFC1305803), the National Natural Science Foundation of China (81771148, 81471309, 81571245, 81501103, and 81701253), the Shandong Provincial Outstanding Medical Academic Professional Program, Taishan Scholars Program of Shandong Province (ts201511109, tsqn20161078, and tsqn20161079), Qingdao Key Health Discipline Development Fund, Qingdao Outstanding Health Professional Development Fund, Shandong Provincial Collaborative Innovation Center for Neurodegenerative Disorders, Research Award Fund for Outstanding Young and Middle-Aged Scientists of Shandong Province (BS2015SW006), and Projects of Medical and Health Technology Development Program in Shandong Province (2015WSA02054). Data collection and sharing for this project was funded by the ADNI (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie; Alzheimer's Association;

Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd., and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC; Johnson & Johnson Pharmaceutical Research & Development LLC; Lumosity; Lundbeck; Merck & Co., Inc.; Meso Scale Diagnostics, LLC; NeuroRxResearch; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Therapeutic Research Institute at the University of Southern California. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California.

REFERENCES

- Alzheimer's Association (2011). 2011 Alzheimer's disease facts and figures. *Alzheimers Dement.* 7, 208–244. doi: 10.1016/j.jalz.2011.02.004
- Carrasquillo, M. M., Allen, M., Burgess, J. D., Wang, X., Strickland, S. L., Aryal, S., et al. (2017). A candidate regulatory variant at the TREM gene cluster associates with decreased alzheimer's disease risk and increased TREML1 and TREM2 brain gene expression. *Alzheimers Dement.* 13, 663–673. doi: 10.1016/j.jalz.2016.10.005
- Colonna, M. (2003). TREMs in the immune system and beyond. *Nat. Rev. Immunol.* 3, 445–453. doi: 10.1038/nri1106
- Condello, C., Yuan, P., and Grutzendler, J. (2018). Microglia-mediated neuroprotection, trem2, and alzheimer's disease: evidence from optical imaging. *Biol. Psychiatry* 83, 377–387. doi: 10.1016/j.biopsych.2017.10.007
- Dennis, J., and Selkoe, J. H. (2016). The amyloid hypothesis of alzheimer's disease at 25 years. *EMBO Mol. Med.* 8, 595–608. doi: 10.15252/emmm.2016.06210
- Derive, M., Bouazza, Y., Sennoun, N., Marchionni, S., Quigley, L., Washington, V., et al. (2012). Soluble TREM-like transcript-1 regulates leukocyte activation and controls microbial sepsis. *J. Immunol.* 188, 5585–5592. doi: 10.4049/jimmunol.1102674
- Fagan, A. M., Roe, C. M., Xiong, C., Mintun, M. A., Morris, J. C., and Holtzman, D. M. (2007). Cerebrospinal fluid tau/beta-amyloid(42) ratio as a prediction of cognitive decline in nondemented older adults. *Arch. Neurol.* 64, 343–349. doi: 10.1001/archneur.64.3.noc60123
- Ford, J. W., and McVicar, D. W. (2009). TREM and TREM-like receptors in inflammation and disease. *Curr. Opin. Immunol.* 21, 38–46. doi: 10.1016/j.coi.2009.01.009
- Hardy, J., and Selkoe, D. J. (2002). The amyloid hypothesis of alzheimer's disease: progress and problems on the road to therapeutics. *Science* 297, 353–356. doi: 10.1126/science.1072994
- Jack, C. R. Jr., Bernstein, M. A., Fox, N. C., Thompson, P., Alexander, G., Harvey, D., et al. (2008). The alzheimer's disease neuroimaging initiative (adni): mri methods. *J. Magn. Reson. Imaging* 27, 685–691. doi: 10.1002/jmri.21049
- Jack, C. R. Jr., Knopman, D. S., Jagust, W. J., Shaw, L. M., Aisen, P. S., Weiner, M. W., et al. (2010). Hypothetical model of dynamic biomarkers of the alzheimer's pathological cascade. *Lancet Neurol.* 9, 119–128. doi: 10.1016/S1474-4422(09)70299-6
- Jagust, W. J., Landau, S. M., Shaw, L. M., Trojanowski, J. Q., Koeppe, R. A., Reiman, E. M., et al. (2009). Relationships between biomarkers in aging and dementia. *Neurology* 73, 1193–1199. doi: 10.1212/WNL.0b013e3181bc010c
- Jay, T. R., Miller, C. M., Cheng, P. J., Graham, L. C., Bemiller, S., Broihier, M. L., et al. (2015). TREM2 deficiency eliminates TREM2+ inflammatory macrophages and ameliorates pathology in alzheimer's disease mouse models. *J. Exp. Med.* 212, 287–295. doi: 10.1084/jem.20142322
- Jiang, T., Tan, L., Zhu, X. C., Zhang, Q. Q., Cao, L., Tan, M. S., et al. (2014). Upregulation of TREM2 ameliorates neuropathology and rescues spatial cognitive impairment in a transgenic mouse model of alzheimer's disease. *Neuropsychopharmacology* 39, 2949–2962. doi: 10.1038/npp.2014.164
- Jiang, T., Tan, L., Zhu, X. C., Zhou, J. S., Cao, L., Tan, M. S., et al. (2015). Silencing of TREM2 exacerbates tau pathology, neurodegenerative changes, and spatial learning deficits in P301S tau transgenic mice. *Neurobiol. Aging* 36, 3176–3186. doi: 10.1016/j.neurobiolaging.2015.08.019
- Jiang, T., Zhang, Y. D., Chen, Q., Gao, Q., Zhu, X. C., Zhou, J. S., et al. (2016). TREM2 modifies microglial phenotype and provides neuroprotection in P301S tau transgenic mice. *Neuropharmacology* 105, 196–206. doi: 10.1016/j.neuropharm.2016.01.028
- Jiang, T., Zhang, Y. D., Gao, Q., Ou, Z., Gong, P. Y., Shi, J. Q., et al. (2018). TREM2 ameliorates neuronal tau pathology through suppression of microglial inflammatory response. *Inflammation* 41, 811–823. doi: 10.1007/s10753-018-0735-5
- Karch, C. M., Cruchaga, C., and Goate, A. M. (2014). Alzheimer's disease genetics: from the bench to the clinic. *Neuron* 83, 11–26. doi: 10.1016/j.neuron.2014.05.041
- Landau, S. M., Harvey, D., Madison, C. M., Koeppe, R. A., Reiman, E. M., Foster, N. L., et al. (2011). Associations between cognitive, functional, and FDG-PET measures of decline in AD and MCI. *Neurobiol. Aging* 32, 1207–1218. doi: 10.1016/j.neurobiolaging.2009.07.002
- Li, J. T., and Zhang, Y. (2018). TREM2 regulates innate immunity in alzheimer's disease. *J. Neuroinflammation* 15:107. doi: 10.1186/s12974-018-1148-y
- Lill, C. M., Rengmark, A., Pihlstrom, L., Fogh, I., Shatunov, A., Sleiman, P. M., et al. (2015). The role of TREM2 R47H as a risk factor for alzheimer's disease, frontotemporal lobar degeneration, amyotrophic lateral sclerosis, and parkinson's disease. *Alzheimers Dement.* 11, 1407–1416. doi: 10.1016/j.jalz.2014.12.009
- Mattsson, N., Zetterberg, H., Hansson, O., Andreasen, N., Parnetti, L., Jonsson, M., et al. (2009). CSF biomarkers and incipient alzheimer disease in patients

- with mild cognitive impairment. *JAMA* 302, 385–393. doi: 10.1001/jama.2009.1064
- Medina, M., and Avila, J. (2014). New perspectives on the role of tau in alzheimer's disease. *Biochem. Pharmacol.* 88, 540–547. doi: 10.1016/j.bcp.2014.01.013
- Olsson, A., Vanderstichele, H., Andreasen, N., De Meyer, G., Wallin, A., Holmberg, B., et al. (2005). Simultaneous measurement of beta-amyloid(1-42), total tau, and phosphorylated tau (Thr181) in cerebrospinal fluid by the xMAP technology. *Clin. Chem.* 51, 336–345. doi: 10.1373/clinchem.2004.03.9347
- Petersen, R. C., Aisen, P. S., Beckett, L. A., Donohue, M. C., Gamst, A. C., Harvey, D. J., et al. (2010). Alzheimer's disease neuroimaging initiative (ADNI): clinical characterization. *Neurology* 74, 201–209. doi: 10.1212/WNL.0b013e3181cb3e25
- Piccio, L., Buonsanti, C., Mariani, M., Cella, M., Gilfillan, S., Cross, A. H., et al. (2007). Blockade of TREM-2 exacerbates experimental autoimmune encephalomyelitis. *Eur. J. Immunol.* 37, 1290–1301. doi: 10.1002/eji.20063.6837
- Selkoe, D. J., and Hardy, J. (2016). The amyloid hypothesis of alzheimer's disease at 25 years. *EMBO Mol. Med.* 8, 595–608. doi: 10.15252/emmm.201606210
- Shaw, L. M., Vanderstichele, H., Knapik-Czajka, M., Clark, C. M., Aisen, P. S., Petersen, R. C., et al. (2009). Cerebrospinal fluid biomarker signature in alzheimer's disease neuroimaging initiative subjects. *Ann. Neurol.* 65, 403–413. doi: 10.1002/ana.21610
- Wang, L., Benzinger, T., Hassenstab, J., Blazey, T., Owen, C., Liu, J., et al. (2015). Spatially distinct trophic links between beta-amyloid and tau in preclinical alzheimer disease. *Neurology* 84, 1254–1260. doi: 10.1212/WNL.0000000000001401
- Washington, A. V., Schubert, R. L., Quigley, L., Disipio, T., Feltz, R., Cho, E. H., et al. (2004). A trem family member, TLT-1, is found exclusively in the alpha-granules of megakaryocytes and platelets. *Blood* 104, 1042–1047. doi: 10.1182/blood-2004-01-0315
- Weigand, S. D., Vemuri, P., Wiste, H. J., Senjem, M. L., Pankratz, V. S., Aisen, P. S., et al. (2011). Transforming cerebrospinal fluid abeta42 measures into calculated pittsburgh Compound b units of brain abeta amyloid. *Alzheimers Dement.* 7, 133–141. doi: 10.1016/j.jalz.2010.08.230
- Yuan, P., Condello, C., Keene, C. D., Wang, Y., Bird, T. D., Paul, S. M., et al. (2016). TREM2 haploinsufficiency in mice and humans impairs the microglia barrier function leading to decreased amyloid compaction and severe axonal dystrophy. *Neuron* 90, 724–739. doi: 10.1016/j.neuron.2016.05.003
- Zheng, H., Liu, C. C., Atagi, Y., Chen, X. F., Jia, L., Yang, L., et al. (2016). Opposing roles of the triggering receptor expressed on myeloid cells 2 and triggering receptor expressed on myeloid cells-like transcript 2 in microglia activation. *Neurobiol. Aging* 42, 132–141. doi: 10.1016/j.neurobiolaging.2016.03.004

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Tian, Ni, Li, Tan, Cao and Tan for the Alzheimer's Disease Neuroimaging Initiative. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.