

Rapid Effects of Vagus Nerve Stimulation on Sensory Processing Through Activation of Neuromodulatory Systems

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Rodenkirch C, Carmel JB and Wang Q (2022) Rapid Effects of Vagus Nerve Stimulation on Sensory Processing Through Activation of Neuromodulatory Systems. Front. Neurosci. 16:922424. doi: 10.3389/fnins.2022.922424 After sensory information is encoded into neural signals at the periphery, it is processed through multiple brain regions before perception occurs (i.e., sensory processing). Recent work has begun to tease apart how neuromodulatory systems influence sensory processing. Vagus nerve stimulation (VNS) is well-known as an effective and safe method of activating neuromodulatory systems. There is a growing body of studies confirming VNS has immediate effects on sensory processing across multiple sensory modalities. These immediate effects of VNS on sensory processing are distinct from the more well-documented method of inducing lasting neuroplastic changes to the sensory pathways through repeatedly delivering a brief VNS burst paired with a sensory stimulus. Immediate effects occur upon VNS onset, often disappear upon VNS offset, and the modulation is present for all sensory stimuli. Conversely, the neuroplastic effect of pairing sub-second bursts of VNS with a sensory stimulus alters sensory processing only after multiple pairing sessions, this alteration remains after cessation of pairing sessions, and the alteration selectively affects the response properties of neurons encoding the specific paired sensory stimulus. Here, we call attention to the immediate effects VNS has on sensory processing. This review discusses existing studies on this topic, provides an overview of the underlying neuromodulatory systems that likely play a role, and briefly explores the potential translational applications of using VNS to rapidly regulate sensory processing.

Keywords: vagus nerve stimulation, sensory processing, neuromodulation, cholinergic system, dopaminergic system, serotonergic system, locus coereleus, noradrenergic system

INTRODUCTION

Accurate and detailed perception of tactile, auditory, and visual stimuli is critical for completing a large variety of tasks, including many necessary for daily life and independent living. Perceptual acuity is dependent upon both reliable transduction of sensory stimuli into neural signals at the periphery and high-fidelity processing of sensory information by the central nervous system. Once sensory information is transduced into neural activity by sensory receptors, it is processed through multiple stages of the sensory pathway before perception occurs (i.e., central sensory processing)

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(Wall and Dubner, 1972; Rodieck, 1979; Reid and Alonso, 1995; Chechik et al., 2006; Wang et al., 2010; Ollerenshaw et al., 2012). Developing methods that use neuromodulation of sensory processing to improve sensory acuity is of great interest as many significant clinical, commercial, and consumer problems stem from misperception or miscommunication. A growing body of evidence strongly suggests that vagus nerve stimulation (VNS) is a safe and effective method of neuromodulation (Collins et al., 2021). In this mini-review, we explore the effects of VNS on sensory processing. Multiple recent reviews have discussed in detail the ability of short VNS bursts repeatedly paired with sensory stimuli to catalyze neuroplastic reorganization of sensory pathways after multiple pairing sessions (Engineer et al., 2013; Hays, 2016; Engineer, 2019), likely via engagement of neuromodulatory systems including the acetylcholine system (Kilgard and Merzenich, 1998). Here, we instead specifically call attention to the immediate effects VNS has on sensory processing and discuss how they likely arise from VNS activating neuromodulatory systems that innervate sensory processing pathways.

Sensory processing is highly dependent upon behavioral states such as attention and arousal (Nicolelis and Fanselow, 2002; Carrasco et al., 2004; Niell and Stryker, 2010; Bennett et al., 2013; Reimer et al., 2014, 2016; McGinley et al., 2015; Vinck et al., 2015; Zheng et al., 2015; Schriver et al., 2018, 2020; Liu et al., 2021) as both are heavily influenced by the same global neuromodulatory systems, including the noradrenergic (Berridge and Waterhouse, 2003; Aston-Jones and Waterhouse, 2016; Liu et al., 2017; Chandler et al., 2019) and cholinergic systems (Pinto et al., 2013). For example, our laboratory recently demonstrated that activation of the locus coeruleus - norepinephrine system (LC-NE), a major neuromodulator of attention and arousal, rapidly enhanced somatosensory processing through NE-mediated suppression of burst spiking induced by calcium T-type channels (Rodenkirch et al., 2019). This NE-enhanced sensory processing increased accuracy of encoded information and improved perceptual sensitivity of awake rats performing tactile discrimination tasks.

LASTING ALTERATIONS TO SENSORY PROCESSING OCCUR OVER TIME WHEN A SENSORY STIMULUS IS REPEATEDLY PAIRED WITH PHASIC VAGUS NERVE STIMULATION

A large body of previous work has focused on using a short burst of VNS repeatedly paired with a brief sensory stimulus to induce reorganization of sensory pathways. This work was inspired by studies which found pairing an auditory tone with phasic activation of dopaminergic, cholinergic, or noradrenergic neuromodulatory systems resulted in a lasting shift of frequency selectivity for neurons in the auditory cortex that selectively favors the paired tone's frequency (Kilgard and Merzenich, 1998; Bao et al., 2001; Nichols et al., 2011; Martins and Froemke, 2015). We will not review these studies in detail here as they have already been well reviewed previously (Engineer et al., 2013; Hays, 2016; Engineer, 2019).

In general, these studies have delivered phasic VNS (e.g., 0.5 s, 30 Hz, 0.8 mA, 100 µs biphasic pulses) in pair with a specific sensory stimulus (e.g., a specific frequency auditory tone or tactile tap) repeatedly across multiple sessions (e.g., 300 times/day, 20 days). This alters sensory processing in a manner that facilitates detection of the specific paired stimulus (Martinez-Vargas et al., 2009; Engineer et al., 2011; Meyers et al., 2019; Darrow et al., 2020; Lai and David, 2021) and accordingly disfavors detection of non-paired stimuli. This mechanism of action can be strengthened over multiple sessions of pairing to produce long-term permanent reorganization of sensory pathways that alters perception. Taken together, these works suggest phasic VNS has great potential as a next generation neuromodulation technology for rehabilitative motor and sensory therapies (Neuhaus et al., 2007; Kreuzer et al., 2014; Engineer et al., 2015; Tyler et al., 2017; Vanneste et al., 2017; Kilgard et al., 2018; Adcock et al., 2020; Llanos et al., 2020; Thakkar et al., 2020; Altidor et al., 2021; Phillips et al., 2021).

TRANSIENT MODULATION OF SENSORY PROCESSING OCCURS RAPIDLY UPON VAGUS NERVE STIMULATION ONSET

The purpose of this review is to bring light to recent studies indicating VNS modulates sensory processing immediately upon onset. Here, we will discuss in detail studies investigating the immediate effects VNS has on the response properties of neurons along the sensory pathways.

Tonic Vagus Nerve Stimulation Drives a Rapid and Transient Enhancement of Tactile Processing

Our laboratory has recently demonstrated that VNS can be used to induce a rapid, general improvement of thalamic sensory processing (Figure 1). This is a continuation of our team's studies investigating the effects of the LC-NE system on thalamocortical circuitry (Rodenkirch et al., 2019), a critical stage for sensory processing and perception (Saalmann and Kastner, 2011; Stanley et al., 2012; Wang et al., 2012; Millard et al., 2013; Kelly et al., 2014; Ollerenshaw et al., 2014; Wimmer et al., 2015; Rikhye et al., 2018). These studies found that direct activation of the LC-NE system (electrical or optogenetic), in a continuous tonic fashion, optimized intrathalamic dynamics for sensory processing. Specifically, tonic LC stimulation (continuous, 5 Hz, 60 μ A, 500 μ s biphasic pulses) increased the efficiency and rate of sensory-related information transmitted by thalamocortical neurons (Rodenkirch et al., 2019). Further, the observed NEenhancement of sensory processing resulted in a significant improvement in perceptual sensitivity for rats tasked with discriminating between whisker stimuli of different frequencies. Through pharmacological manipulation it was determined that tonic LC activation improved thalamic sensory processing because a steady increase in NE concentration precludes priming,



and in turn activation, of thalamic T-type calcium channels. When active, T-type calcium channels introduced a nonlinear bursting response that degraded transmission of detailed sensory information.

Vagus nerve stimulation has been shown to activate the LC-NE system (Hulsey et al., 2017) and is accessible in a non-invasive manner, unlike the LC deep in the brainstem. Therefore, our team next investigated whether tonic VNS would drive similar rapid beneficial effects on sensory processing. Through testing the effects of multiple patterns of VNS on sensory processing, the beneficial effect was found to be highly transient (i.e., benefit begins to dissipate within seconds of ceasing VNS) (Rodenkirch and Wang, 2020). For example, duty-cycled VNS (30 s on/60 s off duty cycle, 30 Hz, 500 µs biphasic pulses) enhanced tactile sensory processing during the on cycle, but this enhancement rapidly dissipated during the off cycle, suggesting that cycling VNS on and off creates fluctuations in sensory processing that would likely be suboptimal for discrimination. This indicated that an uninterrupted pattern is required to produce a stable benefit. Indeed, continuous tonic VNS (continuous, 30 Hz, 500 µs biphasic pulses) induced a steady enhancement of sensory processing similar to that observed with direct tonic LC stimulation. This immediate enhancement of sensory processing during continuous, tonic VNS was found to be reliably present across recorded neurons. As each recorded neuron encoded for a unique kinetic feature of the whisker stimuli, this suggests the tonic VNS modulation provided a general enhancement of sensory processing regardless of stimulus input. This effect is distinct relative to the selective facilitation of responses to a specific sensory stimulus found after repeatedly pairing VNS bursts with that sensory stimulus.

Further, testing of various tonic VNS current levels and frequencies showed the beneficial effect of tonic VNS on sensory processing increased with intensity and frequency (10 vs. 30 Hz,

0.4 vs. 1 and 1.6 mA) and did not exhibit the inverted U-shape function of effect strength that has been observed with other types of VNS modulation (Morrison et al., 2019) (at least within the parameter ranges tested).

Vagus Nerve Stimulation Has Rapid Effects on Evoked Responses in the Auditory Cortex

Other research groups working with human subjects have published findings that suggest VNS has immediate beneficial effects on auditory processing. One study in humans who had been receiving chronic VNS (via implanted cuffs as a treatment for epilepsy), found VNS enhanced performance on a standard auditory oddball task when compared to performance after their VNS device was turned off (De Taeve et al., 2014). Specifically, during VNS (7 s on/18 s off duty cycle, 20-30 Hz, 0.75-3 mA, 250-500 µs pulses) both accuracy and response time were improved for participants tasked with responding to low frequency target audio tones while ignoring high frequency non-target tones. This same study analyzed auditory event-related potentials (AERP), measured via EEG, and found that during VNS, AERP amplitude was also increased. However, the effect on AERP was only significant in individuals whose epilepsy symptoms had positively responded to VNS treatment. A separate study investigating transcutaneous auricular vagus nerve stimulation (taVNS) (30 s on/30 s off duty cycle, 25 Hz, 250 µs pulses) in healthy adults found similar results. Specifically, taVNS increased the strength of AERPs during an oddball auditory task (Rufener et al., 2018). As this study used low frequency tones as nontargets and high frequency tones as targets, a reversal of the prior discussed oddball auditory task, taken together they suggest immediate VNS modulation of auditory response is not specific to low or high frequency audio tones. Another study delivering continuous taVNS (25 Hz, 500 μ s biphasic pulses) to healthy adults analyzed the neural response to auditory tones using magnetoencephalography (MEG) instead of EEG and found taVNS altered synchrony of brain activity (Hyvarinen et al., 2015). Further, recent studies using fMRI to monitor neural activity have shown taVNS rapidly affects auditory processing pathways. When taVNS (25 Hz, 0.1 to 1.8 mA, 500 μ s monophasic pulses) was delivered to male adults with chronic tinnitus, fMRI recordings exhibited altered activity of multiple brain regions involved with auditory processing (Yakunina et al., 2018). More recently, analysis of fMRI data from human subjects receiving taVNS indicated increased activity in the thalamus and auditory cortex (Peng et al., 2018), suggesting VNS rapidly modulates central auditory sensory processing in humans.

These findings in humans are further supported by multiple electrophysiological and behavioral work in animals that found VNS rapidly affects the response properties of neurons of the auditory pathway. In isoflurane-anesthetized rats, the responses of neurons along the auditory pathway were compared with and without VNS delivered via an implanted VNS cuff (30 s on/5 min off duty cycle, 10 Hz, 0.5 mA, 130 µs pulses). The baseline condition was recorded without any ongoing VNS. The VNS condition consisted of discontinuous duty-cycled VNS where auditory testing was performed only during the off periods of the VNS duty cycle. Here they found duty-cycled VNS weakened stimulus-specific adaptation in the cortex but not the thalamus (Shiramatsu et al., 2016), suggesting VNS may modulate thalamocortical transmission but not earlier stages of the auditory pathway. Further work by the same group, using the same paradigm, found VNS predominantly increased the amplitudes of auditory-evoked potentials in the sensory cortex (Takahashi et al., 2020).

Vagus Nerve Stimulation Modulates Olfactory and Gustatory Processing

The immediate effects of VNS on olfactory processing had been demonstrated as early as the 1980s. Specifically, a study in rats found that a single pulse of VNS from an implanted cuff (0.8-1.5 mA, 200 µs monophasic pulses) reliably evoked firing in the homolateral olfactory bulb (HOB) (Garcia-Diaz et al., 1984). Further evidence that VNS affects olfactory processing was found in more recent studies that used positron emission tomography (PET) to analyze the effects of VNS in awake rats. A PET scan conducted during the time period when the VNS cuff was switched on for the first time (30 s on/5 min off duty cycle, 30 Hz, 1.5 mA, 500 µs pulses) found VNS induced a significant increase in glucose metabolism in both olfactory bulbs (Dedeurwaerdere et al., 2005). However, another study in humans with implanted VNS cuffs for treatment of depression found that whether VNS (30 s on/5 min off duty cycle, 20 Hz, 1.25 mA) was on or off had no effect on subjects' ability to discriminate or detect olfactory stimuli (Sperling et al., 2011). Yet that same study did find that VNS significantly increased the intensity of the taste of sweet and bitter, suggesting that VNS may rapidly affect gustatory processing as well.

VAGUS NERVE STIMULATION ACTIVATES MULTIPLE NEUROMODULATORY SYSTEMS THAT RAPIDLY INFLUENCE THE RESPONSE PROPERTIES OF NEURONS ALONG SENSORY PATHWAYS

The ability of VNS to have immediate effects on sensory processing is likely due to VNS activating neuromodulatory systems (Figure 2). Here we briefly review studies of the effect of VNS on neuromodulatory systems in both human and animal models. Neurons in the neuromodulatory systems and sensory pathways discussed here can exhibit either tonic or burst spiking patterns (McCormick and Prince, 1988; Nuñez, 1996; Ramcharan et al., 2000; Devilbiss and Waterhouse, 2011; Rodenkirch et al., 2019). Tonic spiking refers to sustained firing of individual spikes at relatively slow rates compared to phasic. Phasic spiking refers to transient bursts of multiple spikes with short inter-spikeintervals. For neuromodulatory systems, the rate of continuous tonic spiking modulates brain state (e.g., attention and arousal) whereas phasic firing is linked with events (e.g., reward, sensory stimuli, and decision-making) and thought to regulate learning and behavior (Rajkowski et al., 1994; Parikh and Sarter, 2008). For sensory pathways, tonic encoding is favored during periods of increased attention and is thought to be more optimal for discrimination of sensory detail (Sherman, 2001a; Rodenkirch et al., 2019). Conversely, bursting responses to sensory stimuli are more likely when drowsy or inattentive and provide a strong encoding that facilitates detection, potentially serving as a wakeup call (Swadlow and Gusev, 2001; Weyand et al., 2001). It is important to note that neuromodulatory systems are well preserved over evolution, and the function of neuromodulatory systems are similar in humans and other mammals such as rodents (Avery and Krichmar, 2017). Indeed, the studies discussed earlier confirm VNS affects sensory processing in both rodents and humans.

Vagus Nerve Stimulation and the Noradrenergic System

The LC is the primary source of NE in the forebrain (Sara, 2009). The LC exhibits constant tonic firing (1–5 Hz) that regulates brain state (e.g., arousal) as well as intermediate phasic burst spiking events (2–5 spikes at 10–20 Hz per burst) that occur in response to salient sensory stimuli as well as when decisions or responses are made (Devilbiss and Waterhouse, 2011). These two firing modes have been shown to produce distinctly different modulations of the response properties of sensory neurons (Devilbiss and Waterhouse, 2011). The LC innervates multiple regions along the sensory pathway, including the sensory thalamus and cortex (Morrison and Foote, 1986; Simpson et al., 1997).

There is a large body of evidence showing that the LC-NE system modulates sensory processing and perceptual learning (Manunta and Edeline, 1997; Ego-Stengel et al., 2002; Hirata et al., 2006; Doucette et al., 2007; Martins and Froemke, 2015;



dorsal raphe nucleus; LC, locus coeruleus; LDT, laterodorsal tegmental nucleus; NTS, nucleus tractus solitaries; PPT, pedunculopontine tegmental nucleus; VTA, ventral tegmental area.

McBurney-Lin et al., 2019; Waterhouse and Navarra, 2019). Moreover, it is well documented that activation of the LC-NE system immediately modulates the response of sensory neurons. In vitro, NE has a depolarizing effect on auditory and visual thalamic relay neurons that coincides with a suppression of burst spiking (McCormick and Prince, 1988). This likely occurs because NE depolarization prevents the extended hyperpolarized periods needed to prime the calcium T-type channels responsible for bursts (Sherman, 2001a). In vivo, tonic LC activation has been found to reduce spontaneous activity of the somatosensory thalamus, while facilitating sensory evoked activity, resulting in an increase in signal to noise ratio (Hirata et al., 2006). Our team has shown how tonic LC-NE activation enhances the accuracy of encoded stimuli in the somatosensory thalamus by reducing the fluctuating influence of the calcium T-type channels responsible for bursting. Within the cortex, the LC-NE system can cause either facilitation or inhibition with resulting effect specific to the sensory modality, cell, and stimulation pattern (Devilbiss and Waterhouse, 2004; Videen et al., 1984; Sato et al., 1989; Vazey et al., 2018).

Vagus nerve stimulation's ability to activate the LC-NE system has long been hypothesized to underlie, in part, the clinical

benefits of VNS (Slater and Wang, 2021). VNS is thought to activate the LC via the vagus nerve's afferent projections to the nucleus tractus solitarius (NTS) (Van Bockstaele et al., 1999; Ruffoli et al., 2011). The NTS then sends an excitatory signal to the LC, likely via the nucleus paragigantocellularis (Ennis and Aston-Jones, 1988; Reyes and Van Bockstaele, 2006). Indeed, multiple studies have confirmed VNS readily activates the LC-NE system in both animals and humans. In rats, VNS delivered via an implanted cuff has been shown to increase the activity of LC neurons as confirmed by electrophysiological recordings under halothane (Groves et al., 2005), chloral hydrate (Dorr and Debonnel, 2006), equithesin (Manta et al., 2009a), and ketamine (Hulsey et al., 2017) as well as by immunohistochemical biomarkers of short-term neuronal activation (Cunningham et al., 2008). Similarly, multiple studies have found that microdialysis samples taken from rats receiving VNS exhibited increased NE concentration in the primary hippocampus (Raedt et al., 2011), basolateral amygdala (Hassert et al., 2004), and cortex (Roosevelt et al., 2006; Follesa et al., 2007; Manta et al., 2013). Finally, the findings in animals seem to be conserved in humans, as fMRI data from a study of adult males with tinnitus indicated taVNS activates the NTS and LC

(Yakunina et al., 2018). However, variations in VNS parameters may affect how reliably VNS drives the LC-NE system, as one study measuring NE concentration in the CSF of patients receiving VNS as a treatment for depression failed to detect a significant change (Carpenter et al., 2004).

In addition to direct evidence VNS activates the LC-NE system, many effects of VNS are blocked if the LC-NE system is impaired through either LC lesion or adrenergic receptor blockers. For example, the anticonvulsive effect of VNS is abrogated when hippocampal adrenergic receptors are blocked (Krahl et al., 1998; Raedt et al., 2011). Further, VNS enhancement of perforant path-CA3 synaptic transmission is blocked by either electrical lesions of the LC or an adrenergic receptor antagonist (timolol) (Shen et al., 2012). The antidepressant-like effects of VNS in rats, as measured by feeding and swim tests, have been shown to be blocked by lesion of noradrenergic neurons (Furmaga et al., 2011; Grimonprez et al., 2015). Immunotoxin depletion of norepinephrine was also found to prevent VNS-driven enhancement of motor cortex neuroplasticity (Hulsey et al., 2019).

Vagus Nerve Stimulation and Cholinergic Systems

Cholinergic nuclei of the basal forebrain (BF) project to the sensory processing regions of the thalamus (Kolmac and Mitrofanis, 1999) and cortex (Ballinger et al., 2016; Jimenez-Martin et al., 2021). Additionally, cholinergic nuclei of the pontomesencephalic area, including the laterodorsal tegmental nucleus (LDT) and pedunculopontine tegmental nucleus (PPT), are a major source of ACh to the thalamus (Schofield et al., 2011; Huerta-Ocampo et al., 2020). There are two distinct neuron populations of the BF that differentiate in exhibiting either a tonic (10–15 Hz) or a bursting (2–6 spikes/burst with bursting events occurring at 0.3–2 Hz) firing pattern (Nuñez, 1996) which influences arousal and attention. The response timing of both types of BF neurons is influenced by sensory stimuli (Laszlovszky et al., 2020) and linked with novelty, salience, and surprise (Zhang et al., 2019).

Extensive work has shown the cholinergic system strongly influences both sensory processing and perceptual learning across multiple sensory modalities (Murphy and Sillito, 1991; Kilgard and Merzenich, 1998; Verdier and Dykes, 2001; Linster and Cleland, 2002; Bentley et al., 2004; Wilson et al., 2004; Furey et al., 2008; Herrero et al., 2008; Pinto et al., 2013; Zhan et al., 2013; Rothermel et al., 2014; Kim et al., 2016; Gratton et al., 2017). Like the noradrenergic system, it is well documented that activation of the cholinergic systems has immediate effects on sensory processing. ACh applied in vitro to neurons of the thalamic reticular nucleus, a subthalamic region involved in sensory processing, causes hyperpolarization and induces burst spiking (McCormick and Prince, 1986), likely due to extended hyperpolarized periods priming the calcium T-type channels responsible for burst spiking (Sherman, 2001a). ACh applied to thalamic neurons of the primary visual and auditory pathways was found to increase firing rate (Sillito et al., 1983; McCormick and Prince, 1987), although a hyperpolarization effect has been observed in thalamic neurons of the secondary (non-lemniscal) auditory pathway (Mooney et al., 2004). Cholinergic modulation of the sensory cortex can cause either facilitation or inhibition with the resulting effect specific to the sensory modality, cell, and stimulation pattern (Donoghue and Carroll, 1987; Metherate and Weinberger, 1989; Metherate and Ashe, 1991; Jimenez-Martin et al., 2021). In the visual cortex, BF stimulation has been shown to enhance accurate encoding by inducing decorrelation and increased reliability (Goard and Dan, 2009).

It has long been hypothesized that VNS activates the BF-ACh system (Detari et al., 1983). VNS innervates the NTS (Ruffoli et al., 2011) and projections from the NTS activate the BF (Martin et al., 2022) in addition to the NTS projections that activate the LC (Ennis and Aston-Jones, 1988; Van Bockstaele et al., 1999; Reyes and Van Bockstaele, 2006). The LC also projects to the BF (Berridge et al., 2003), suggesting VNS activates the BF both directly through the NTS as well as indirectly through the LC. Indeed, two separate studies investigating the potential of VNS for inducing neuroprotection from cerebral ischemia found that VNS enhanced protein levels of the nicotinic acetylcholine receptor alpha7 subunit (a7nAchR) in the ischemic penumbra (Jiang et al., 2014; Lu et al., 2017). Recently, researchers performed in vivo calcium imaging of the auditory cortex and found VNS evoked activity of cholinergic axons innervating the region (Mridha et al., 2021). Further, they found the intensity of the evoked activity covaried with VNS intensity. In addition to this direct evidence that VNS rapidly activates the cholinergic system, multiple studies have shown ACh modulation of sensory pathways is a critical component underlying the plasticity effect induced by repeatedly pairing a burst of VNS with a sensory stimulus. For example, the effects of VNS on sensory processing in the auditory cortex were found to be blocked by a muscarinic antagonist (Nichols et al., 2011). Further, lesioning the NB in rats was shown to abrogate the well-documented ability of VNS pulses repeatedly paired with a movement to enhance motor cortex plasticity (Hulsey et al., 2016).

Vagus Nerve Stimulation and Serotonergic Systems

The dorsal raphe nucleus (DRN) is a major source of serotonin (5-HT) to the forebrain (Jacobs and Fornal, 1999). Neurons of the DRN consistently exhibit a continuous slow tonic firing rate (1-2 Hz) with little variation in inter-spike-interval (Trulson and Jacobs, 1979; Mlinar et al., 2016). Response of the DRN is related to both reward and punishment (Ranade and Mainen, 2009; Li et al., 2016; Ren et al., 2018) as well as linked to sensory input (Rasmussen et al., 1984; Waterhouse et al., 2004). The DRN innervates both cortical and subcortical regions of the sensory processing pathways (Kirifides et al., 2001). There is also a large body of work suggesting DRN activity modulates sensory processing and perception (Hurley and Pollak, 1999; Kähkönen et al., 2002; Dacks et al., 2009; Hurley and Hall, 2011; Jaber et al., 2014; Kapoor et al., 2016; Seillier et al., 2017). 5-HT has been shown to have instant effects on neurons of the sensory pathways. For example, 5-HT has been shown to cause excitation of thalamic perigeniculate and reticular nucleus neurons (McCormick and Wang, 1991; Funke and Eysel, 1993). In the inferior colliculus, an auditory region of the midbrain, 5-HT was found to modulate responses in both a cell and auditory stimulus specific manner (Hurley and Pollak, 1999). In the primary visual and auditory relay neurons of the visual and auditory pathways, 5-HT has been shown to have an inhibitory effect (Marks et al., 1987; Kayama et al., 1989; Monckton and McCormick, 2002). Additionally, activation of the DRN has been found to increase signal to noise ratio of the olfactory cortex (Lottem et al., 2016).

Vagus nerve stimulation may activate the DRN indirectly by first activating the LC which then projects to the DRN (Kim et al., 2004). This hypothesis is supported by a study in rats anesthetized with sodium pentobarbital that found VNS increased DRN neurons' firing rates, but this causal relationship was lost once the LC was lesioned (Manta et al., 2009a). Multiple studies have also shown that VNS increases DRN firing rate as measured via extracellular electrophysiological recordings (Dorr and Debonnel, 2006; Manta et al., 2009b). However, one study found only a subset of VNS patterns they tested increased DRN activity suggesting VNS activation of the DRN may be dependent on VNS parameters (Manta et al., 2012). In a followup work, the same group performed in vivo microdialysis in rats following chronic duty-cycled VNS and found increased 5-HT concentration in the DRN but not the hippocampus nor prefrontal cortex (PFC) (Manta et al., 2013). In contrast to these studies supporting VNS' ability to activate the DRN, another study analyzing microdialysis measurements in different brain regions of rats reported that neither vagotomy or chronic unilateral VNS had an effect on 5-HT levels in the ventral tegmental area (VTA), nucleus accumbens (NAc), PFC, and striatum (Ziomber et al., 2012). These conflicting findings could potentially be related to the fact that electrical stimulation was delivered to an abdominal branch of the vagus nerve in this study. Further suggesting a more complex interplay between the VNS and DRN, a study analyzing immunohistochemical biomarkers of both short-term and long-term neuronal activation suggests chronic VNS does not induce DRN activation until stimulation has occurred across multiple days (Cunningham et al., 2008).

In addition to direct evidence that VNS increases activity of the serotonergic system, functionality of serotonergic neurons has been shown to be critical for multiple documented effects of VNS. For example, the earlier-mentioned study on the antidepressant-like effects of VNS in rats, which used feeding and swim tests as indexes of depression, found the beneficial effects of VNS were also precluded by administration of a neurotoxin for serotonergic neurons (Furmaga et al., 2011). Additionally, a separate study found immunotoxin depletion of serotonin prevented the well-researched ability of repeatedly pairing a VNS burst with a movement to enhance motor cortex neuroplasticity (Hulsey et al., 2019).

Vagus Nerve Stimulation and Dopaminergic Systems

The VTA and Substantia Nigra pars Compacta (SNc) are primary sources of dopamine (DA) to the forebrain (Poulin et al., 2018)

and, respectively, they modulate cognition and movement (Mercuri et al., 1992). The VTA has been shown to innervate the sensory cortices (Hosp et al., 2019). The VTA exhibits both tonic (1–8 Hz) and burst firing (2–5 spike bursts with bursting events occurring at 0.1–1 Hz) with firing rates varying across cell types (Kiyatkin and Rebec, 1998; Hyland et al., 2002; Lodge and Grace, 2006). Tonic firing rate likely modulates brain state (e.g., motivation and arousal) and bursting events likely encode for salient stimuli (e.g., reward and sensory stimuli) (Dahan et al., 2007). Although the body of work investigating the effects of DA on sensory processing is limited, there is evidence it rapidly modulates sensory processing and response (Ungless, 2004; Govindaiah et al., 2010; Woolrych et al., 2021).

Although previous work demonstrated the LC projects to the VTA (Mejías-Aponte et al., 2009), many studies also suggest VNS effects on DA circuitry may be dependent on other factors besides VNS directly increasing VTA firing rates. For example, one study that performed in vivo microdialysis of rats following chronic duty-cycled VNS found an increase in DA in the PFC and NAc but a decrease in VTA neurons' firing rates as measured with electrophysiological recordings (Manta et al., 2013). A lack of VNS-induced changes in VTA firing and bursting rates was also reported in a separate study (Perez et al., 2014). Studies analyzing brain sections from rats that received chronic VNS have also reported varied results. One such study found decreased DA levels in the VTA, NAc, PFC, and striatum (Ziomber et al., 2012); however, to properly interpret these results it should be mentioned that electrical stimulation was delivered to an abdominal branch of the vagus nerve in this study. Two other studies performing a similar analysis found VNS induced changes to the elemental composition of dopamine-related brain structures (Szczerbowska-Boruchowska et al., 2012) and to the lipids and proteins within the VTA, NAc, SNc, striatum, dorsal motor nucleus of vagus, and the motor cortex (Surowka et al., 2015). A more recent study in awake rats found optogenetic VNS, which carries no risk of unintentional activation of surrounding nerves, increased the firing rate of dopaminergic VTA neurons as measured via in vivo imaging (Fernandes et al., 2020). This same study also found lesioning the hepatic branch of the vagus nerve abrogated the increase in VTA neuron activity usually observed following ingestion.

DISCUSSION: TRANSLATIONAL APPLICATIONS OF USING VAGUS NERVE STIMULATION TO RAPIDLY MODULATE SENSORY PROCESSING

Accurate perception is required for daily life and independent living. However, dysfunction or degradation of central sensory processing pathways can rapidly impair sensory ability. The studies referenced here implicate VNS as a potential tool for modulating sensory processing. Accordingly, VNS presents great potential as a targeted treatment for impaired senses arising from central sensory processing dysfunction. Many clinical causes of impaired central sensory processing exist including multiple

neurodegenerative conditions and neurological disorders. Impaired sensory processing reduces sensory acuity, increases likelihood of miscommunication, and causes misperceptions that potentially lead to costly human error. Further, the link between human performance and sensory ability suggests there may be commercial interest in enhancing sensory processing in addition to clinical. This translation potential has spurred clinical trials looking at the effect of VNS on auditory perception (e.g., NCT04812015 at www.clinicaltrials.gov). VNS methods of enhancing sensory processing have great translation possibility because cervical transcutaneous VNS (ctVNS) and taVNS have both been suggested to be safe and effective methods of noninvasively activating the vagus nerve in humans (Frangos and Komisaruk, 2017; Mwamburi et al., 2017; Reuter et al., 2019; McIntire et al., 2021). In light of this potential, our research team is currently conducting pilot clinical studies investigating the effects of continuous tonic VNS on vision, hearing, and touch.

Age-related impairment of central sensory processing is particularly devastating to the elderly as it interferes with their ability to communicate (Tun et al., 2012; Sardone et al., 2019), accelerates cognitive decline (Hewitt, 2017), and is linked with Alzheimer's disease (AD) (Panza et al., 2015). Treatments exist for age-related sensory receptor damage (Barriga-Rivera et al., 2017; Ferguson et al., 2017; Higuchi et al., 2017; Moshirfar et al., 2017). However, there is a stark lack of solutions addressing the co-occurring age-related impairment of central sensory processing (Humes et al., 2013; Humes, 2015; Engel-Yeger and Rosenblum, 2017; Lesica, 2018). For example, as evidence of this age-related decline in sensory processing, studies have shown that elderly individuals with normal audiograms, indicating normally functioning auditory receptors, still have decreased ability to discriminate detailed features of sensory stimuli, such as speech intelligibility over noise (Fullgrabe et al., 2014; Babkoff and Fostick, 2017). Similarly, aging is thought to degrade visual (Brannan, 1992; Wiegand et al., 2014) and tactile processing (Engel-Yeger et al., 2012). The ability to improve or restore sensory processing clarity with VNS, could therefore positively impact a large segment of society by helping them remain social and active through improving their ability to communicate clearly and walk safely. Many researchers share the belief that different forms of VNS could help elderly cognition

REFERENCES

- Adcock, K. S., Chandler, C., Buell, E. P., Solorzano, B. R., Loerwald, K. W., Borland, M. S., et al. (2020). Vagus nerve stimulation paired with tones restores auditory processing in a rat model of Rett syndrome. *Brain Stimul.* 13, 1494–1503. doi: 10.1016/j.brs.2020.08.006
- Altidor, L. K., Bruner, M. M., Deslauriers, J. F., Garman, T. S., Ramirez, S., Dirr, E. W., et al. (2021). Acute vagus nerve stimulation enhances reversal learning in rats. *Neurobiol. Learn. Mem.* 184:107498. doi: 10.1016/j.nlm.2021.107498
- Arnsten, A. F., and Dudley, A. G. (2005). Methylphenidate improves prefrontal cortical cognitive function through alpha2 adrenoceptor and dopamine D1 receptor actions: relevance to therapeutic effects in attention deficit hyperactivity disorder. *Behav. Brain Funct.* 1:2. doi: 10.1186/1744-9081-1-2
- Aston-Jones, G., and Waterhouse, B. (2016). Locus coeruleus: from global projection system to adaptive regulation of behavior. *Brain Res.* 1645, 75–78. doi: 10.1016/j.brainres.2016.03.001

and perception as suggested by the many ongoing clinical studies investigating that topic (e.g., Clinical Trials NCT04396249, NCT04276805, NCT03359902, NCT04908358, NCT04276805, and NCT03989375 at www.clinicaltrials.gov).

Attention deficit hyperactivity disorder (ADHD) has been linked with impaired sensory processing evidenced by poor frequency discrimination ability (Sutcliffe et al., 2006; Shimizu et al., 2014). Moreover, inattention is linked with increased bursting activity in the sensory thalamus, a type of neural activity our team's research has found is suboptimal for encoding details and features of sensory stimuli therefore causing loss of sensory acuity (Rodenkirch et al., 2019). Further, thalamocortical bursting in response to sensory stimuli is thought to serve as a "wake-up-call" in response to salient stimuli, suggesting bursts are distracting (Sherman, 2001b). Recently, poor intrathalamic processing due to abnormal TRN responses has been suggested as a cause of ADHD (Wells et al., 2016). ADHD treatments (including stimulants) work, in part, via amplifying NE effects (Arnsten and Dudley, 2005; De Crescenzo et al., 2018; Schneider et al., 2019). Methylphenidate, a common treatment for ADHD, has been shown to enhance early-stage sensory processing through increasing DA and NE concentration in the brain (Navarra et al., 2017). Previous work shows that VNS activates the locus coeruleus-norepinephrine (LC-NE) system (Hulsey et al., 2017), and our work shows VNS suppresses noisy bursting activity along sensory pathways. Taken together, these findings suggest VNS could be potentially used to treat the sensory processing dysfunction linked with ADHD.

AUTHOR CONTRIBUTIONS

CR and QW wrote the manuscript with input from JC. All authors contributed to the article and approved the submitted version.

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- Avery, M. C., and Krichmar, J. L. (2017). Neuromodulatory systems and their interactions: a review of models, theories, and experiments. *Front. Neural Circ.* 11:108. doi: 10.3389/fncir.2017.00108
- Babkoff, H., and Fostick, L. (2017). Age-related changes in auditory processing and speech perception: cross-sectional and longitudinal analyses. *Eur. J. Ageing* 14, 269–281. doi: 10.1007/s10433-017-0410-y
- Ballinger, E. C., Ananth, M., Talmage, D. A., and Role, L. W. (2016). Basal forebrain cholinergic circuits and signaling in cognition and cognitive decline. *Neuron* 91, 1199–1218. doi: 10.1016/j.neuron.2016.09.006
- Bao, S., Chan, V. T., and Merzenich, M. M. (2001). Cortical remodelling induced by activity of ventral tegmental dopamine neurons. *Nature* 412, 79–83. doi: 10.1038/35083586
- Barriga-Rivera, A., Bareket, L., Goding, J., Aregueta-Robles, U. A., and Suaning, G. J. (2017). Visual prosthesis: interfacing stimulating electrodes with retinal neurons to restore vision. *Front. Neurosci.* 11:620. doi: 10.3389/fnins.2017. 00620

- Bennett, C., Arroyo, S., and Hestrin, S. (2013). Subthreshold mechanisms underlying state-dependent modulation of visual responses. *Neuron* 80, 350– 357. doi: 10.1016/j.neuron.2013.08.007
- Bentley, P., Husain, M., and Dolan, R. J. (2004). Effects of cholinergic enhancement on visual stimulation, spatial attention, and spatial working memory. *Neuron* 41, 969–982. doi: 10.1016/S0896-6273(04)00145-X
- Berridge, C. W., Isaac, S. O., and España, R. A. (2003). Additive wake-promoting actions of medial basal forebrain noradrenergic alpha1- and beta-receptor stimulation. *Behav. Neurosci.* 117, 350–359. doi: 10.1037/0735-7044.117.2.350
- Berridge, C. W., and Waterhouse, B. D. (2003). The locus coeruleus–noradrenergic system: modulation of behavioral state and state-dependent cognitive processes. *Brain Res. Rev.* 42, 33–84. doi: 10.1016/s0165-0173(03)00143-7
- Brannan, J. R. (1992). "Changes in temporal visual processing in normal aging," in Advances in Psychology, ed. J. R. Brannan (Amsterdam: North-Holland), 119–134. doi: 10.1016/S0166-4115(08)61351-5
- Carpenter, L. L., Moreno, F. A., Kling, M. A., Anderson, G. M., Regenold, W. T., Labiner, D. M., et al. (2004). Effect of vagus nerve stimulation on cerebrospinal fluid monoamine metabolites, norepinephrine, and gamma-aminobutyric acid concentrations in depressed patients. *Biol. Psychiatry* 56, 418–426. doi: 10.1016/ j.biopsych.2004.06.025
- Carrasco, M., Ling, S., and Read, S. (2004). Attention alters appearance. *Nat. Neurosci.* 7, 308–313. doi: 10.1038/nn1194
- Chandler, D. J., Jensen, P., McCall, J. G., Pickering, A. E., Schwarz, L. A., and Totah, N. K. (2019). Redefining noradrenergic neuromodulation of behavior: impacts of a modular locus coeruleus architecture. *J. Neurosci.* 39:8239. doi: 10.1523/JNEUROSCI.1164-19.2019
- Chechik, G., Anderson, M. J., Bar-Yosef, O., Young, E. D., Tishby, N., and Nelken, I. (2006). Reduction of information redundancy in the ascending auditory pathway. *Neuron* 51, 359–368. doi: 10.1016/j.neuron.2006.06.030
- Collins, L., Boddington, L., Steffan, P. J., and McCormick, D. (2021). Vagus nerve stimulation induces widespread cortical and behavioral activation. *Curr. Biol.* 31, 2088.e3–2098.e3. doi: 10.1016/j.cub.2021.02.049
- Cunningham, J. T., Mifflin, S. W., Gould, G. G., and Frazer, A. (2008). Induction of c-Fos and DeltaFosB immunoreactivity in rat brain by Vagal nerve stimulation. *Neuropsychopharmacology* 33, 1884–1895. doi: 10.1038/sj.npp.1301570
- Dacks, A. M., Green, D. S., Root, C. M., Nighorn, A. J., and Wang, J. W. (2009). Serotonin modulates olfactory processing in the antennal lobe of *Drosophila*. *J. Neurogenet.* 23, 366–377. doi: 10.3109/01677060903085722
- Dahan, L., Astier, B., Vautrelle, N., Urbain, N., Kocsis, B., and Chouvet, G. (2007). Prominent burst firing of dopaminergic neurons in the ventral tegmental area during paradoxical sleep. *Neuropsychopharmacology* 32, 1232–1241. doi: 10. 1038/sj.npp.1301251
- Darrow, M. J., Mian, T. M., Torres, M., Haider, Z., Danaphongse, T., Rennaker, R. L. Jr., et al. (2020). Restoration of somatosensory function by pairing vagus nerve stimulation with tactile rehabilitation. *Ann. Neurol.* 87, 194–205. doi: 10.1002/ana.25664
- De Crescenzo, F., Ziganshina, L. E., Yudina, E. V., Kaplan, Y. C., Ciabattini, M., Wei, Y., et al. (2018). Noradrenaline reuptake inhibitors (NRIs) for attention deficit hyperactivity disorder (ADHD) in adults. *Cochrane Database Syst. Rev.* 2018:CD013044. doi: 10.1002/14651858.CD013044
- De Taeye, L., Vonck, K., van Bochove, M., Boon, P., Van Roost, D., Mollet, L., et al. (2014). The P3 event-related potential is a biomarker for the efficacy of vagus nerve stimulation in patients with epilepsy. *Neurotherapeutics* 11, 612–622. doi: 10.1007/s13311-014-0272-3
- Dedeurwaerdere, S., Cornelissen, B., Van Laere, K., Vonck, K., Achten, E., Slegers, G., et al. (2005). Small animal positron emission tomography during vagus nerve stimulation in rats: a pilot study. *Epilepsy Res.* 67, 133–141. doi: 10.1016/ j.eplepsyres.2005.09.008
- Detari, L., Juhasz, G., and Kukorelli, T. (1983). Effect of stimulation of vagal and radial nerves on neuronal activity in the basal forebrain area of anaesthetized cats. *Acta Physiol. Hung* 61, 147–154.
- Devilbiss, D. M., and Waterhouse, B. D. (2004). The effects of tonic locus ceruleus output on sensory-evoked responses of ventral posterior medial thalamic and barrel field cortical neurons in the awake rat. J. Neurosci. 24, 10773–10785. doi: 10.1523/JNEUROSCI.1573-04.2004
- Devilbiss, D. M., and Waterhouse, B. D. (2011). Phasic and tonic patterns of locus coeruleus output differentially modulate sensory network function in the awake rat. J. Neurophysiol. 105, 69–87. doi: 10.1152/jn.00445.2010

- Donoghue, J. P., and Carroll, K. L. (1987). Cholinergic modulation of sensory responses in rat primary somatic sensory cortex. *Brain Res.* 408, 367–371. doi: 10.1016/0006-8993(87)90407-0
- Dorr, A. E., and Debonnel, G. (2006). Effect of vagus nerve stimulation on serotonergic and noradrenergic transmission. J. Pharmacol. Exp. Ther. 318, 890–898. doi: 10.1124/jpet.106.104166
- Doucette, W., Milder, J., and Restrepo, D. (2007). Adrenergic modulation of olfactory bulb circuitry affects odor discrimination. *Learn. Mem.* 14, 539–547. doi: 10.1101/lm.606407
- Ego-Stengel, V., Bringuier, V., and Shulz, D. E. (2002). Noradrenergic modulation of functional selectivity in the cat visual cortex: an *in vivo* extracellular and intracellular study. *Neuroscience* 111, 275–289. doi: 10.1016/s0306-4522(02) 00011-8
- Engel-Yeger, B., Hus, S., and Rosenblum, S. (2012). Age effects on sensoryprocessing abilities and their impact on handwriting. *Can. J. Occup. Ther. Rev. Can. D'ergother.* 79, 264–274. doi: 10.2182/CJOT.2012.79.5.2
- Engel-Yeger, B., and Rosenblum, S. (2017). The relationship between sensoryprocessing patterns and occupational engagement among older persons. *Can. J. Occup. Ther. Rev. Can. D'ergother.* 84, 10–21. doi: 10.1177/000841741769 0415
- Engineer, C. (2019). Vagus nerve stimulation as a strategy to augment auditory rehabilitation. *Brain Stimul.* 12:420. doi: 10.1016/j.brs.2018.12.360
- Engineer, C. T., Engineer, N. D., Riley, J. R., Seale, J. D., and Kilgard, M. P. (2015). Pairing speech sounds with vagus nerve stimulation drives stimulus-specific cortical plasticity. *Brain Stimul.* 8, 637–644. doi: 10.1016/j.brs.2015.01.408
- Engineer, N. D., Moller, A. R., and Kilgard, M. P. (2013). Directing neural plasticity to understand and treat tinnitus. *Hear Res.* 295, 58–66. doi: 10.1016/j.heares. 2012.10.001
- Engineer, N. D., Riley, J. R., Seale, J. D., Vrana, W. A., Shetake, J. A., Sudanagunta, S. P., et al. (2011). Reversing pathological neural activity using targeted plasticity. *Nature* 470, 101–104. doi: 10.1038/nature09656
- Ennis, M., and Aston-Jones, G. (1988). Activation of locus coeruleus from nucleus paragigantocellularis: a new excitatory amino acid pathway in brain. J. Neurosci. 8, 3644–3657. doi: 10.1523/JNEUROSCI.08-10-03644.1988
- Ferguson, M. A., Kitterick, P. T., Chong, L. Y., Edmondson-Jones, M., Barker, F., and Hoare, D. J. (2017). Hearing aids for mild to moderate hearing loss in adults. *Cochr. Database Syst. Rev.* 9:CD012023. doi: 10.1002/14651858. CD012023.pub2
- Fernandes, A. B., Alves da Silva, J., Almeida, J., Cui, G., Gerfen, C. R., Costa, R. M., et al. (2020). Postingestive modulation of food seeking depends on vagusmediated dopamine neuron activity. *Neuron* 106, 778.e6–788.e6. doi: 10.1016/j. neuron.2020.03.009
- Follesa, P., Biggio, F., Gorini, G., Caria, S., Talani, G., Dazzi, L., et al. (2007). Vagus nerve stimulation increases norepinephrine concentration and the gene expression of BDNF and bFGF in the rat brain. *Brain Res.* 1179, 28–34. doi: 10.1016/j.brainres.2007.08.045
- Frangos, E., and Komisaruk, B. R. (2017). Access to vagal projections via cutaneous electrical stimulation of the neck: fMRI evidence in healthy humans. Brain Stimul. 10, 19–27. doi: 10.1016/j.brs.2016.10.008
- Fullgrabe, C., Moore, B. C., and Stone, M. A. (2014). Age-group differences in speech identification despite matched audiometrically normal hearing: contributions from auditory temporal processing and cognition. *Front. Aging Neurosci.* 6:347. doi: 10.3389/fnagi.2014.00347
- Funke, K., and Eysel, U. T. (1993). Modulatory effects of acetylcholine, serotonin and noradrenaline on the activity of cat perigeniculate neurons. *Exp. Brain Res.* 95, 409–420. doi: 10.1007/BF00227133
- Furey, M. L., Pietrini, P., Haxby, J. V., and Drevets, W. C. (2008). Selective effects of cholinergic modulation on task performance during selective attention. *Neuropsychopharmacology* 33, 913–923. doi: 10.1038/sj.npp.1301461
- Furmaga, H., Shah, A., and Frazer, A. (2011). Serotonergic and noradrenergic pathways are required for the anxiolytic-like and antidepressant-like behavioral effects of repeated vagal nerve stimulation in rats. *Biol. Psychiatry* 70, 937–945. doi: 10.1016/j.biopsych.2011.07.020
- Garcia-Diaz, D. E., Aguilar-Baturoni, H. U., Guevara-Aguilar, R., and Wayner, M. J. (1984). Vagus nerve stimulation modifies the electrical activity of the olfactory bulb. *Brain Res. Bull.* 12, 529–537. doi: 10.1016/0361-9230(84)90168-0
- Goard, M., and Dan, Y. (2009). Basal forebrain activation enhances cortical coding of natural scenes. Nat. Neurosci. 12, 1444–1449. doi: 10.1038/nn.2402

- Govindaiah, G., Wang, Y., and Cox, C. L. (2010). Dopamine enhances the excitability of somatosensory thalamocortical neurons. *Neuroscience* 170, 981– 991. doi: 10.1016/j.neuroscience.2010.08.043
- Gratton, C., Yousef, S., Aarts, E., Wallace, D. L., D'Esposito, M., and Silver, M. A. (2017). Cholinergic, but not dopaminergic or noradrenergic, enhancement sharpens visual spatial perception in humans. J. Neurosci. 37, 4405–4415. doi: 10.1523/JNEUROSCI.2405-16.2017
- Grimonprez, A., Raedt, R., Portelli, J., Dauwe, I., Larsen, L. E., Bouckaert, C., et al. (2015). The antidepressant-like effect of vagus nerve stimulation is mediated through the locus coeruleus. *J. Psychiatr. Res.* 68, 1–7. doi: 10.1016/j.jpsychires. 2015.05.002
- Groves, D. A., Bowman, E. M., and Brown, V. J. (2005). Recordings from the rat locus coeruleus during acute vagal nerve stimulation in the anaesthetised rat. *Neurosci. Lett.* 379, 174–179. doi: 10.1016/j.neulet.2004.12.055
- Hassert, D. L., Miyashita, T., and Williams, C. L. (2004). The effects of peripheral vagal nerve stimulation at a memory-modulating intensity on norepinephrine output in the basolateral amygdala. *Behav. Neurosci.* 118, 79–88. doi: 10.1037/ 0735-7044.118.1.79
- Hays, S. A. (2016). Enhancing rehabilitative therapies with vagus nerve stimulation. Neurotherapeutics 13, 382–394. doi: 10.1007/s13311-015-0417-z
- Herrero, J. L., Roberts, M. J., Delicato, L. S., Gieselmann, M. A., Dayan, P., and Thiele, A. (2008). Acetylcholine contributes through muscarinic receptors to attentional modulation in V1. *Nature* 454, 1110–1114. doi: 10.1038/ nature07141
- Hewitt, D. (2017). Age-related hearing loss and cognitive decline: you haven't heard the half of it. Front. Aging Neurosci. 9:112. doi: 10.3389/fnagi.2017.00112
- Higuchi, A., Kumar, S. S., Benelli, G., Alarfaj, A. A., Munusamy, M. A., Umezawa, A., et al. (2017). Stem cell therapies for reversing vision loss. *Trends Biotechnol.* 35, 1102–1117. doi: 10.1016/j.tibtech.2017.06.016
- Hirata, A., Aguilar, J., and Castro-Alamancos, M. A. (2006). Noradrenergic activation amplifies bottom-up and top-down signal-to-noise ratios in sensory thalamus. J. Neurosci. 26, 4426–4436. doi: 10.1523/JNEUROSCI.5298-05.2006
- Hosp, J. A., Coenen, V. A., Rijntjes, M., Egger, K., Urbach, H., Weiller, C., et al. (2019). Ventral tegmental area connections to motor and sensory cortical fields in humans. *Brain Struct. Funct.* 224, 2839–2855. doi: 10.1007/s00429-019-01939-0
- Huerta-Ocampo, I., Hacioglu-Bay, H., Dautan, D., and Mena-Segovia, J. (2020). Distribution of midbrain cholinergic axons in the thalamus. *eNeuro* 7:ENEURO.0454-19.2019. doi: 10.1523/ENEURO.0454-19.2019
- Hulsey, D. R., Hays, S. A., Khodaparast, N., Ruiz, A., Das, P., Rennaker, R. L. II, et al. (2016). Reorganization of motor cortex by vagus nerve stimulation requires cholinergic innervation. *Brain Stimul.* 9, 174–181. doi: 10.1016/j.brs.2015.12. 007
- Hulsey, D. R., Riley, J. R., Loerwald, K. W., Rennaker, R. L. II, Kilgard, M. P., and Hays, S. A. (2017). Parametric characterization of neural activity in the locus coeruleus in response to vagus nerve stimulation. *Exp. Neurol.* 289, 21–30. doi: 10.1016/j.expneurol.2016.12.005
- Hulsey, D. R., Shedd, C. M., Sarker, S. F., Kilgard, M. P., and Hays, S. A. (2019). Norepinephrine and serotonin are required for vagus nerve stimulation directed cortical plasticity. *Exp. Neurol.* 320:112975. doi: 10.1016/j.expneurol. 2019.112975
- Humes, L. E. (2015). Age-related changes in cognitive and sensory processing: focus on middle-aged adults. *Am. J. Audiol.* 24, 94–97. doi: 10.1044/2015_AJA-14-0063
- Humes, L. E., Busey, T. A., Craig, J., and Kewley-Port, D. (2013). Are agerelated changes in cognitive function driven by age-related changes in sensory processing? *Atten. Percept. Psychophys.* 75, 508–524. doi: 10.3758/s13414-012-0406-9
- Hurley, L. M., and Hall, I. C. (2011). Context-dependent modulation of auditory processing by serotonin. *Hear. Res.* 279, 74–84. doi: 10.1016/j.heares.2010.12. 015
- Hurley, L. M., and Pollak, G. D. (1999). Serotonin differentially modulates responses to tones and frequency-modulated sweeps in the inferior colliculus. J. Neurosci. 19, 8071–8082. doi: 10.1523/JNEUROSCI.19-18-08071. 1999
- Hyland, B. I., Reynolds, J. N. J., Hay, J., Perk, C. G., and Miller, R. (2002). Firing modes of midbrain dopamine cells in the freely moving rat. *Neuroscience* 114, 475–492. doi: 10.1016/S0306-4522(02)00267-1

- Hyvarinen, P., Yrttiaho, S., Lehtimaki, J., Ilmoniemi, R. J., Makitie, A., Ylikoski, J., et al. (2015). Transcutaneous vagus nerve stimulation modulates tinnitusrelated beta- and gamma-band activity. *Ear Hear.* 36, e76–e85. doi: 10.1097/ AUD.00000000000123
- Jaber, L., Zhao, F. L., Kolli, T., and Herness, S. (2014). A physiologic role for serotonergic transmission in adult rat taste buds. *PLoS One* 9:e112152. doi: 10.1371/journal.pone.0112152
- Jacobs, B. L., and Fornal, C. A. (1999). Activity of serotonergic neurons in behaving animals. *Neuropsychopharmacology* 21, 9S–15S. doi: 10.1016/S0893-133X(99) 00012-3
- Jiang, Y., Li, L., Liu, B., Zhang, Y., Chen, Q., and Li, C. (2014). Vagus nerve stimulation attenuates cerebral ischemia and reperfusion injury *via* endogenous cholinergic pathway in rat. *PLoS One* 9:e102342. doi: 10.1371/journal.pone. 0102342
- Jimenez-Martin, J., Potapov, D., Potapov, K., Knöpfel, T., and Empson, R. M. (2021). Cholinergic modulation of sensory processing in awake mouse cortex. *Sci. Rep.* 11:17525. doi: 10.1038/s41598-021-96696-8
- Kähkönen, S., Ahveninen, J., Pennanen, S., Liesivuori, J., Ilmoniemi, R. J., and Jääskeläinen, I. P. (2002). Serotonin modulates early cortical auditory processing in healthy subjects. Evidence from MEG with acute tryptophan depletion. *Neuropsychopharmacology* 27, 862–868. doi: 10.1016/S0893-133X(02)00357-3
- Kapoor, V., Provost, A. C., Agarwal, P., and Murthy, V. N. (2016). Activation of raphe nuclei triggers rapid and distinct effects on parallel olfactory bulb output channels. *Nat. Neurosci.* 19, 271–282. doi: 10.1038/nn.4219
- Kayama, Y., Shimada, S., Hishikawa, Y., and Ogawa, T. (1989). Effects of stimulating the dorsal raphe nucleus of the rat on neuronal activity in the dorsal lateral geniculate nucleus. *Brain Res.* 489, 1–11. doi: 10.1016/0006-8993(89) 90002-4
- Kelly, S. T., Kremkow, J., Jin, J., Wang, Y., Wang, Q., Alonso, J.-M., et al. (2014). The role of thalamic population synchrony in the emergence of cortical feature selectivity. *PLoS Comput. Biol.* 10:e1003418. doi: 10.1371/journal.pcbi.1003418
- Kilgard, M. P., and Merzenich, M. M. (1998). Cortical map reorganization enabled by nucleus basalis activity. *Science* 279, 1714–1718. doi: 10.1126/science.279. 5357.1714
- Kilgard, M. P., Rennaker, R. L., Alexander, J., and Dawson, J. (2018). Vagus nerve stimulation paired with tactile training improved sensory function in a chronic stroke patient. *NeuroRehabilitation* 42, 159–165. doi: 10.3233/NRE-172273
- Kim, J. H., Jung, A. H., Jeong, D., Choi, I., Kim, K., Shin, S., et al. (2016). Selectivity of neuromodulatory projections from the basal forebrain and locus ceruleus to primary sensory cortices. *J. Neurosci.* 36, 5314–5327. doi: 10.1523/JNEUROSCI. 4333-15.2016
- Kim, M. A., Lee, H. S., Lee, B. Y., and Waterhouse, B. D. (2004). Reciprocal connections between subdivisions of the dorsal raphe and the nuclear core of the locus coeruleus in the rat. *Brain Res.* 1026, 56–67. doi: 10.1016/j.brainres. 2004.08.022
- Kirifides, M. L., Simpson, K. L., Lin, R. C., and Waterhouse, B. D. (2001). Topographic organization and neurochemical identity of dorsal raphe neurons that project to the trigeminal somatosensory pathway in the rat. J. Comp. Neurol. 435, 325–340. doi: 10.1002/cne.1033
- Kiyatkin, E. A., and Rebec, G. V. (1998). Heterogeneity of ventral tegmental area neurons: single-unit recording and iontophoresis in awake, unrestrained rats. *Neuroscience* 85, 1285–1309. doi: 10.1016/s0306-4522(98)00054-2
- Kolmac, C., and Mitrofanis, J. (1999). Organization of the basal forebrain projection to the thalamus in rats. *Neurosci. Lett.* 272, 151–154. doi: 10.1016/ s0304-3940(99)00614-x
- Krahl, S. E., Clark, K. B., Smith, D. C., and Browning, R. A. (1998). Locus coeruleus lesions suppress the seizure-attenuating effects of vagus nerve stimulation. *Epilepsia* 39, 709–714. doi: 10.1111/j.1528-1157.1998.tb01155.x
- Kreuzer, P. M., Landgrebe, M., Resch, M., Husser, O., Schecklmann, M., Geisreiter, F., et al. (2014). Feasibility, safety and efficacy of transcutaneous vagus nerve stimulation in chronic tinnitus: an open pilot study. *Brain Stimul.* 7, 740–747. doi: 10.1016/j.brs.2014.05.003
- Lai, J., and David, S. V. (2021). Short-term effects of vagus nerve stimulation on learning and evoked activity in auditory cortex. *eNeuro* 8:ENEURO.0522-20.2021. doi: 10.1523/ENEURO.0522-20.2021
- Laszlovszky, T., Schlingloff, D., Hegedüs, P., Freund, T. F., Gulyás, A., Kepecs, A., et al. (2020). Distinct synchronization, cortical coupling and behavioral

function of two basal forebrain cholinergic neuron types. Nat. Neurosci. 23, 992–1003. doi: 10.1038/s41593-020-0648-0

- Lesica, N. A. (2018). Why do hearing aids fail to restore normal auditory perception? *Trends Neurosci.* 41, 174–185. doi: 10.1016/j.tins.2018.01.008
- Li, Y., Zhong, W., Wang, D., Feng, Q., Liu, Z., Zhou, J., et al. (2016). Serotonin neurons in the dorsal raphe nucleus encode reward signals. *Nat. Commun.* 7:10503. doi: 10.1038/ncomms10503
- Linster, C., and Cleland, T. A. (2002). Cholinergic modulation of sensory representations in the olfactory bulb. *Neural Netw.* 15, 709–717. doi: 10.1016/ S0893-6080(02)00061-8
- Liu, Y., Narasimhan, S., Schriver, B. J., and Wang, Q. (2021). Perceptual behavior depends differently on pupil-linked arousal and heartbeat dynamics-linked arousal in rats performing tactile discrimination tasks. *Front. Syst. Neurosci.* 14:614248. doi: 10.3389/fnsys.2020.614248
- Liu, Y., Rodenkirch, C., Moskowitz, N., Schriver, B., and Wang, Q. (2017). Dynamic lateralization of pupil dilation evoked by locus coeruleus activation results from sympathetic, not parasympathetic, contributions. *Cell Rep.* 20, 3099–3112. doi: 10.1016/j.celrep.2017.08.094
- Llanos, F., McHaney, J. R., Schuerman, W. L., Yi, H. G., Leonard, M. K., and Chandrasekaran, B. (2020). Non-invasive peripheral nerve stimulation selectively enhances speech category learning in adults. *NPJ Sci. Learn.* 5:12. doi: 10.1038/s41539-020-0070-0
- Lodge, D. J., and Grace, A. A. (2006). The laterodorsal tegmentum is essential for burst firing of ventral tegmental area dopamine neurons. *Proc. Natl. Acad. Sci.* U.S.A. 103, 5167–5172. doi: 10.1073/pnas.0510715103
- Lottem, E., Lörincz, M. L., and Mainen, Z. F. (2016). Optogenetic activation of dorsal raphe serotonin neurons rapidly inhibits spontaneous but not odor-evoked activity in olfactory cortex. J. Neurosci. 36, 7–18. doi: 10.1523/ JNEUROSCI.3008-15.2016
- Lu, X. X., Hong, Z. Q., Tan, Z., Sui, M. H., Zhuang, Z. Q., Liu, H. H., et al. (2017). Nicotinic acetylcholine receptor Alpha7 subunit mediates vagus nerve stimulation-induced neuroprotection in acute permanent cerebral ischemia by a7nAchR/JAK2 pathway. *Med. Sci. Monit.* 23, 6072–6081. doi: 10.12659/msm. 907628
- Manta, S., Dong, J., Debonnel, G., and Blier, P. (2009a). Enhancement of the function of rat serotonin and norepinephrine neurons by sustained vagus nerve stimulation. J. Psychiatry Neurosci. 34, 272–280.
- Manta, S., Dong, J., Debonnel, G., and Blier, P. (2009b). Optimization of vagus nerve stimulation parameters using the firing activity of serotonin neurons in the rat dorsal raphe. *Eur. Neuropsychopharmacol.* 19, 250–255. doi: 10.1016/j. euroneuro.2008.12.001
- Manta, S., El Mansari, M., and Blier, P. (2012). Novel attempts to optimize vagus nerve stimulation parameters on serotonin neuronal firing activity in the rat brain. *Brain Stimul.* 5, 422–429. doi: 10.1016/j.brs.2011.04.005
- Manta, S., El Mansari, M., Debonnel, G., and Blier, P. (2013). Electrophysiological and neurochemical effects of long-term vagus nerve stimulation on the rat monoaminergic systems. *Int. J. Neuropsychopharmacol.* 16, 459–470. doi: 10. 1017/S1461145712000387
- Manunta, Y., and Edeline, J. M. (1997). Effects of noradrenaline on frequency tuning of rat auditory cortex neurons. *Eur. J. Neurosci.* 9, 833–847. doi: 10.1111/ j.1460-9568.1997.tb01433.x
- Marks, G. A., Speciale, S. G., Cobbey, K., and Roffwarg, H. P. (1987). Serotonergic inhibition of the dorsal lateral geniculate nucleus. *Brain Res.* 418, 76–84. doi: 10.1016/0006-8993(87)90964-4
- Martin, K. A., Papadoyannis, E. S., Schiavo, J. K., Fadaei, S. S., Temiz, N. Z., McGinley, M. J., et al. (2022). Vagus nerve stimulation recruits the central cholinergic system to enhance perceptual learning. *bioRxiv* [Preprint]. doi: 10.1101/2022.01.28.478197v1
- Martinez-Vargas, D., Valdes-Cruz, A., Magdaleno-Madrigal, V. M., Almazan-Alvarado, S., and Fernandez-Mas, R. (2009). Effects of electrical stimulation of the vagus nerve on the development of visual habituation in the cat. *Behav. Brain Res.* 205, 45–49. doi: 10.1016/j.bbr.2009.06.029
- Martins, A. R., and Froemke, R. C. (2015). Coordinated forms of noradrenergic plasticity in the locus coeruleus and primary auditory cortex. *Nat. Neurosci.* 18, 1483–1492. doi: 10.1038/nn.4090
- McBurney-Lin, J., Lu, J., Zuo, Y., and Yang, H. (2019). Locus coeruleusnorepinephrine modulation of sensory processing and perception: a focused

review. Neurosci. Biobehav. Rev. 105, 190-199. doi: 10.1016/j.neubiorev.2019. 06.009

- McCormick, D. A., and Prince, D. A. (1986). Acetylcholine induces burst firing in thalamic reticular neurones by activating a potassium conductance. *Nature* 319, 402–405. doi: 10.1038/319402a0
- McCormick, D. A., and Prince, D. A. (1987). Actions of acetylcholine in the guineapig and cat medial and lateral geniculate nuclei, in vitro. *J. Physiol.* 392, 147–165. doi: 10.1113/jphysiol.1987.sp016774
- McCormick, D. A., and Prince, D. A. (1988). Noradrenergic modulation of firing pattern in guinea pig and cat thalamic neurons, in vitro. J. Neurophysiol. 59, 978–996. doi: 10.1152/jn.1988.59.3.978
- McCormick, D. A., and Wang, Z. (1991). Serotonin and noradrenaline excite GABAergic neurones of the guinea-pig and cat nucleus reticularis thalami. *J. Physiol.* 442, 235–255. doi: 10.1113/jphysiol.1991.sp018791
- McGinley, M. J., Vinck, M., Reimer, J., Batista-Brito, R., Zagha, E., Cadwell Cathryn, R., et al. (2015). Waking state: rapid variations modulate neural and behavioral responses. *Neuron* 87, 1143–1161. doi: /10.1016/j.neuron.2015.09. 012
- McIntire, L. K., McKinley, R. A., Goodyear, C., McIntire, J. P., and Brown, R. D. (2021). Cervical transcutaneous vagal nerve stimulation (ctVNS) improves human cognitive performance under sleep deprivation stress. *Commun. Biol.* 4:634. doi: 10.1038/s42003-021-02145-7
- Mejías-Aponte, C. A., Drouin, C., and Aston-Jones, G. (2009). Adrenergic and noradrenergic innervation of the midbrain ventral tegmental area and retrorubral field: prominent inputs from medullary homeostatic centers. *J. Neurosci.* 29, 3613–3626. doi: 10.1523/JNEUROSCI.4632-08.2009
- Mercuri, N. B., Calabresi, P., and Bernardi, G. (1992). The electrophysiological actions of dopamine and dopaminergic drugs on neurons of the substantia nigra pars compacta and ventral tegmental area. *Life Sci.* 51, 711–718. doi: 10.1016/0024-3205(92)90479-9
- Metherate, R., and Ashe, J. H. (1991). Basal forebrain stimulation modifies auditory cortex responsiveness by an action at muscarinic receptors. *Brain Res.* 559, 163–167. doi: 10.1016/0006-8993(91)90301-b
- Metherate, R., and Weinberger, N. M. (1989). Acetylcholine produces stimulusspecific receptive field alterations in cat auditory cortex. *Brain Res.* 480, 372–377. doi: 10.1016/0006-8993(89)90210-2
- Meyers, E. C., Kasliwal, N., Solorzano, B. R., Lai, E., Bendale, G., Berry, A., et al. (2019). Enhancing plasticity in central networks improves motor and sensory recovery after nerve damage. *Nat. Commun.* 10:5782. doi: 10.1038/s41467-019-13695-0
- Millard, D. C., Wang, Q., Gollnick, C. A., and Stanley, G. B. (2013). System identification of the nonlinear dynamics in the thalamocortical circuit in response to patterned thalamic microstimulation in vivo. *J. Neural Eng.* 10:066011. doi: 10.1088/1741-2560/10/6/066011
- Mlinar, B., Montalbano, A., Piszczek, L., Gross, C., and Corradetti, R. (2016). Firing properties of genetically identified dorsal raphe serotonergic neurons in brain slices. *Front. Cell. Neurosci.* 10:195. doi: 10.3389/fncel.2016.00195
- Monckton, J. E., and McCormick, D. A. (2002). Neuromodulatory role of serotonin in the ferret thalamus. J. Neurophysiol. 87, 2124–2136. doi: 10.1152/jn.00650. 2001
- Mooney, D. M., Zhang, L., Basile, C., Senatorov, V. V., Ngsee, J., Omar, A., et al. (2004). Distinct forms of cholinergic modulation in parallel thalamic sensory pathways. *Proc. Natl. Acad. Sci. U.S.A.* 101, 320–324. doi: 10.1073/pnas. 0304445101
- Morrison, J. H., and Foote, S. L. (1986). Noradrenergic and serotoninergic innervation of cortical, thalamic, and tectal visual structures in Old and New World monkeys. J. Comp. Neurol. 243, 117–138. doi: 10.1002/cne.902430110
- Morrison, R. A., Hulsey, D. R., Adcock, K. S., Rennaker, R. L. II, Kilgard, M. P., and Hays, S. A. (2019). Vagus nerve stimulation intensity influences motor cortex plasticity. *Brain Stimul.* 12, 256–262. doi: 10.1016/j.brs.2018.10.017
- Moshirfar, M., Jehangir, N., Fenzl, C. R., and McCaughey, M. (2017). LASIK enhancement: clinical and surgical management. J. Refract. Surg. 33, 116–127. doi: 10.3928/1081597X-20161202-01
- Mridha, Z., de Gee, J. W., Shi, Y., Alkashgari, R., Williams, J., Suminski, A., et al. (2021). Graded recruitment of pupil-linked neuromodulation by parametric stimulation of the vagus nerve. *Nat. Commun.* 12:1539. doi: 10.1038/s41467-021-21730-2

- Murphy, P. C., and Sillito, A. M. (1991). Cholinergic enhancement of direction selectivity in the visual cortex of the cat. *Neuroscience* 40, 13–20. doi: 10.1016/ 0306-4522(91)90170-S
- Mwamburi, M., Liebler, E. J., and Tenaglia, A. T. (2017). Review of non-invasive vagus nerve stimulation (gammaCore): efficacy, safety, potential impact on comorbidities, and economic burden for episodic and chronic cluster headache. *Am. J. Manag. Care* 23, S317–S325.
- Navarra, R. L., Clark, B. D., Gargiulo, A. T., and Waterhouse, B. D. (2017). Methylphenidate enhances early-stage sensory processing and rodent performance of a visual signal detection task. *Neuropsychopharmacology* 42, 1326–1337. doi: 10.1038/npp.2016.267
- Neuhaus, A. H., Luborzewski, A., Rentzsch, J., Brakemeier, E. L., Opgen-Rhein, C., Gallinat, J., et al. (2007). P300 is enhanced in responders to vagus nerve stimulation for treatment of major depressive disorder. J. Affect. Disord. 100, 123–128. doi: 10.1016/j.jad.2006.10.005
- Nichols, J. A., Nichols, A. R., Smirnakis, S. M., Engineer, N. D., Kilgard, M. P., and Atzori, M. (2011). Vagus nerve stimulation modulates cortical synchrony and excitability through the activation of muscarinic receptors. *Neuroscience* 189, 207–214. doi: 10.1016/j.neuroscience.2011.05.024
- Nicolelis, M. A. L., and Fanselow, E. E. (2002). Thalamocortical optimization of tactile processing according to behavioral state. *Nat. Neurosci.* 5, 517–523. doi: 10.1038/nn0602-517
- Niell, C. M., and Stryker, M. P. (2010). Modulation of visual responses by behavioral state in mouse visual cortex. *Neuron* 65, 472–479. doi: 10.1016/j. neuron.2010.01.033
- Nuñez, A. (1996). Unit activity of rat basal forebrain neurons: relationship to cortical activity. *Neuroscience* 72, 757–766. doi: 10.1016/0306-4522(95)00 582-X
- Ollerenshaw, D. R., Bari, B. A., Millard, D. C., Orr, L. E., Wang, Q., and Stanley, G. B. (2012). Detection of tactile inputs in the rat vibrissa pathway. *J. Neurophysiol.* 108, 479–490. doi: 10.1152/jn.00004.2012
- Ollerenshaw, D. R., Zheng, H. J. V., Millard Daniel, C., Wang, Q., and Stanley Garrett, B. (2014). The adaptive trade-off between detection and discrimination in cortical representations and behavior. *Neuron* 81, 1152–1164. doi: 10.1016/j. neuron.2014.01.025
- Panza, F., Solfrizzi, V., and Logroscino, G. (2015). Age-related hearing impairmenta risk factor and frailty marker for dementia and AD. *Nat. Rev. Neurol.* 11, 166–175. doi: 10.1038/nrneurol.2015.12
- Parikh, V., and Sarter, M. (2008). Cholinergic mediation of attention. Ann. N.Y. Acad. Sci. 1129, 225–235. doi: 10.1196/annals.1417.021
- Peng, L., Mu, K., Liu, A., Zhou, L., Gao, Y., Shenoy, I. T., et al. (2018). Transauricular vagus nerve stimulation at auricular acupoints Kindey (CO10), Yidan (CO11), Liver (CO12) and Shenmen (TF4) can induce auditory and limbic cortices activation measured by fMRI. *Hear. Res.* 359, 1–12. doi: 10.1016/ j.heares.2017.12.003
- Perez, S. M., Carreno, F. R., Frazer, A., and Lodge, D. J. (2014). Vagal nerve stimulation reverses aberrant dopamine system function in the methylazoxymethanol acetate rodent model of schizophrenia. *J. Neurosci.* 34, 9261–9267. doi: 10.1523/JNEUROSCI.0588-14.2014
- Phillips, I., Calloway, R. C., Karuzis, V. P., Pandza, N. B., O'Rourke, P., and Kuchinsky, S. E. (2021). Transcutaneous auricular vagus nerve stimulation strengthens semantic representations of foreign language tone words during initial stages of learning. *J. Cogn. Neurosci.* 34, 127–152. doi: 10.1162/jocn_a_ 01783
- Pinto, L., Goard, M. J., Estandian, D., Xu, M., Kwan, A. C., Lee, S.-H., et al. (2013). Fast modulation of visual perception by basal forebrain cholinergic neurons. *Nat. Neurosci.* 16, 1857–1863. doi: 10.1038/nn.3552
- Poulin, J. F., Caronia, G., Hofer, C., Cui, Q., Helm, B., Ramakrishnan, C., et al. (2018). Mapping projections of molecularly defined dopamine neuron subtypes using intersectional genetic approaches. *Nat. Neurosci.* 21, 1260–1271. doi: 10. 1038/s41593-018-0203-4
- Raedt, R., Clinckers, R., Mollet, L., Vonck, K., El Tahry, R., Wyckhuys, T., et al. (2011). Increased hippocampal noradrenaline is a biomarker for efficacy of vagus nerve stimulation in a limbic seizure model. *J. Neurochem.* 117, 461–469. doi: 10.1111/j.1471-4159.2011.07214.x
- Rajkowski, J., Kubiak, P., and Aston-Jones, G. (1994). Locus coeruleus activity in monkey: phasic and tonic changes are associated with altered vigilance. *Brain Res. Bull.* 35, 607–616. doi: 10.1016/0361-9230(94)90175-9

- Ramcharan, E. J., Gnadt, J. W., and Sherman, S. M. (2000). Burst and tonic firing in thalamic cells of unanesthetized, behaving monkeys. *Vis. Neurosci.* 17, 55–62. doi: 10.1017/s0952523800171056
- Ranade, S. P., and Mainen, Z. F. (2009). Transient firing of dorsal raphe neurons encodes diverse and specific sensory, motor, and reward events. J. Neurophysiol. 102, 3026–3037. doi: 10.1152/jn.00507.2009
- Rasmussen, K., Heym, J., and Jacobs, B. L. (1984). Activity of serotonin-containing neurons in nucleus centralis superior of freely moving cats. *Exp. Neurol.* 83, 302–317. doi: 10.1016/S0014-4886(84)90100-6
- Reid, C. R., and Alonso, J.-M. (1995). Specificity of monosynaptic connections from thalamus to visual cortex. *Nature* 378, 281–284. doi: 10.1038/378281a0
- Reimer, J., Froudarakis, E., Cadwell Cathryn, R., Yatsenko, D., Denfield George, H., and Tolias Andreas, S. (2014). Pupil fluctuations track fast switching of cortical states during quiet wakefulness. *Neuron* 84, 355–362. doi: 10.1016/j.neuron. 2014.09.033
- Reimer, J., McGinley, M. J., Liu, Y., Rodenkirch, C., Wang, Q., McCormick, D. A., et al. (2016). Pupil fluctuations track rapid changes in adrenergic and cholinergic activity in cortex. *Nat. Commun.* 7:13289. doi: 10.1038/ ncomms13289
- Ren, J., Friedmann, D., Xiong, J., Liu, C. D., Ferguson, B. R., Weerakkody, T., et al. (2018). Anatomically defined and functionally distinct dorsal raphe serotonin sub-systems. *Cell* 175, 472.e20–487.e20. doi: 10.1016/j.cell.2018.07.043
- Reuter, U., McClure, C., Liebler, E., and Pozo-Rosich, P. (2019). Non-invasive neuromodulation for migraine and cluster headache: a systematic review of clinical trials. J. Neurol. Neurosurg. Psychiatry 90, 796–804. doi: 10.1136/jnnp-2018-320113
- Reyes, B. A., and Van Bockstaele, E. J. (2006). Divergent projections of catecholaminergic neurons in the nucleus of the solitary tract to limbic forebrain and medullary autonomic brain regions. *Brain Res.* 1117, 69–79. doi: 10.1016/j.brainres.2006.08.051
- Rikhye, R. V., Wimmer, R. D., and Halassa, M. M. (2018). Toward an integrative theory of thalamic function. Annu. Rev. Neurosci. 41, 163–183. doi: 10.1146/ annurev-neuro-080317-062144
- Rodenkirch, C., Liu, Y., Schriver, B. J., and Wang, Q. (2019). Locus coeruleus activation enhances thalamic feature selectivity via norepinephrine regulation of intrathalamic circuit dynamics. Nat. Neurosci. 22, 120–133. doi: 10.1038/ s41593-018-0283-1
- Rodenkirch, C., and Wang, Q. (2020). Rapid and transient enhancement of thalamic information transmission induced by vagus nerve stimulation. *J. Neural Eng.* 17:e026027. doi: 10.1088/1741-2552/ab6b84
- Rodieck, R. W. (1979). Visual pathways. Annu. Rev. Neurosci. 2, 193–225. doi: 10.1146/annurev.ne.02.030179.001205
- Roosevelt, R. W., Smith, D. C., Clough, R. W., Jensen, R. A., and Browning, R. A. (2006). Increased extracellular concentrations of norepinephrine in cortex and hippocampus following vagus nerve stimulation in the rat. *Brain Res.* 1119, 124–132. doi: 10.1016/j.brainres.2006.08.048
- Rothermel, M., Carey, R. M., Puche, A., Shipley, M. T., and Wachowiak, M. (2014). Cholinergic inputs from Basal forebrain add an excitatory bias to odor coding in the olfactory bulb. *J. Neurosci.* 34, 4654–4664. doi: 10.1523/JNEUROSCI.5026-13.2014
- Rufener, K. S., Geyer, U., Janitzky, K., Heinze, H. J., and Zaehle, T. (2018). Modulating auditory selective attention by non-invasive brain stimulation: differential effects of transcutaneous vagal nerve stimulation and transcranial random noise stimulation. *Eur. J. Neurosci.* 48, 2301–2309. doi: 10.1111/ejn. 14128
- Ruffoli, R., Giorgi, F. S., Pizzanelli, C., Murri, L., Paparelli, A., and Fornai, F. (2011). The chemical neuroanatomy of vagus nerve stimulation. *J. Chem. Neuroanat.* 42, 288–296. doi: 10.1016/j.jchemneu.2010.12.002
- Saalmann, Y. B., and Kastner, S. (2011). Cognitive and perceptual functions of the visual thalamus. *Neuron* 71, 209–223. doi: 10.1016/j.neuron.2011.06.027
- Sara, S. J. (2009). The locus coeruleus and noradrenergic modulation of cognition. Nat. Rev. Neurosci. 10, 211–223. doi: 10.1038/nrn2573
- Sardone, R., Battista, P., Panza, F., Lozupone, M., Griseta, C., Castellana, F., et al. (2019). The age-related central auditory processing disorder: silent impairment of the cognitive ear. *Front. Neurosci.* 13:619. doi: 10.3389/fnins.2019.00619
- Sato, H., Fox, K., and Daw, N. W. (1989). Effect of electrical stimulation of locus coeruleus on the activity of neurons in the cat visual cortex. *J. Neurophysiol.* 62, 946–958. doi: 10.1152/jn.1989.62.4.946

- Schneider, J., Patterson, M., and Jimenez, X. F. (2019). Beyond depression: other uses for tricyclic antidepressants. *Cleveland Clin. J. Med.* 86, 807–814. doi: 10.3949/ccjm.86a.19005
- Schofield, B. R., Motts, S. D., and Mellott, J. G. (2011). Cholinergic cells of the pontomesencephalic tegmentum: connections with auditory structures from cochlear nucleus to cortex. *Hear. Res.* 279, 85–95. doi: 10.1016/j.heares.2010. 12.019
- Schriver, B., Bagdasarov, S., and Wang, Q. (2018). Pupil-linked arousal modulates behavior in rats performing a whisker deflection direction discrimination task. *J. Neurophysiol.* 120, 1655–1670. doi: 10.1152/jn.00290.2018
- Schriver, B. J., Perkins, S. M., Sajda, P., and Wang, Q. (2020). Interplay between components of pupil-linked phasic arousal and its role in driving behavioral choice in Go/No-Go perceptual decision-making. *Psychophysiology* 57:e13565. doi: 10.1111/psyp.13565
- Seillier, L., Lorenz, C., Kawaguchi, K., Ott, T., Nieder, A., Pourriahi, P., et al. (2017). Serotonin decreases the gain of visual responses in awake macaque V1. *J. Neurosci.* 37, 11390–11405. doi: 10.1523/JNEUROSCI.1339-17.2017
- Shen, H., Fuchino, Y., Miyamoto, D., Nomura, H., and Matsuki, N. (2012). Vagus nerve stimulation enhances perforant path-CA3 synaptic transmission via the activation of beta-adrenergic receptors and the locus coeruleus. *Int. J. Neuropsychopharmacol.* 15, 523–530. doi: 10.1017/S1461145711000708
- Sherman, S. M. (2001a). Tonic and burst firing: dual modes of thalamocortical relay. *Trends Neurosci.* 24, 122–126.
- Sherman, S. M. (2001b). A wake-up call from the thalamus. Nat. Neurosci. 4, 344–346.
- Shimizu, V. T., Bueno, O. F., and Miranda, M. C. (2014). Sensory processing abilities of children with ADHD. Brazil. J. Phys. Ther. 18, 343–352.
- Shiramatsu, T. I., Hitsuyu, R., Ibayashi, K., Kanzaki, R., Kawai, K., and Takahashi, H. (2016). Effect of vagus nerve stimulation on neural adaptation in thalamocortical system in rats. Annu. Int. Conf. IEEE Eng. Med. Biol. Soc. 2016, 1834–1837. doi: 10.1109/EMBC.2016.7591076
- Sillito, A. M., Kemp, J. A., and Berardi, N. (1983). The cholinergic influence on the function of the cat dorsal lateral geniculate nucleus (dLGN). *Brain Res.* 280, 299–307.
- Simpson, K. L., Altman, D. W., Wang, L., Kirifides, M. L., Lin, R. C., and Waterhouse, B. D. (1997). Lateralization and functional organization of the locus coeruleus projection to the trigeminal somatosensory pathway in rat. *J. Comp. Neurol.* 385, 135–147.
- Slater, C., and Wang, Q. (2021). Alzheimer's disease: an evolving understanding of noradrenergic involvement and the promising future of electroceutical therapies. *Clin. Transl. Med.* 11:e397. doi: 10.1002/ctm2.397
- Sperling, W., Biermann, T., Spannenberger, R., Clepce, M., Padberg, F., Reulbach, U., et al. (2011). Changes in gustatory perceptions of patients with major depression treated with vagus nerve stimulation (VNS). *Pharmacopsychiatry* 44, 67–71. doi: 10.1055/s-0030-1268427
- Stanley, G. B., Jin, J., Wang, Y., Desbordes, G., Wang, Q., Black, M. J., et al. (2012). Visual orientation and directional selectivity through thalamic synchrony. *J. Neurosci.* 32, 9073–9088. doi: 10.1523/JNEUROSCI.4968-11.2012
- Surowka, A. D., Krygowska-Wajs, A., Ziomber, A., Thor, P., Chrobak, A. A., and Szczerbowska-Boruchowska, M. (2015). Peripheral vagus nerve stimulation significantly affects lipid composition and protein secondary structure within dopamine-related brain regions in rats. *Neuromol. Med.* 17, 178–191. doi: 10. 1007/s12017-015-8349-7
- Sutcliffe, P. A., Bishop, D. V., Houghton, S., and Taylor, M. (2006). Effect of attentional state on frequency discrimination: a comparison of children with ADHD on and off medication. J. Speech Lang. Hear. Res. 49, 1072–1084. doi: 10.1044/1092-4388(2006/076)
- Swadlow, H. A., and Gusev, A. G. (2001). The impact of 'bursting' thalamic impulses at a neocortical synapse. *Nat. Neurosci.* 4, 402–408. doi: 10.1038/86054
- Szczerbowska-Boruchowska, M., Krygowska-Wajs, A., Ziomber, A., Thor, P., Wrobel, P., Bukowczan, M., et al. (2012). The influence of electrical stimulation of vagus nerve on elemental composition of dopamine related brain structures in rats. *Neurochem. Int.* 61, 156–165. doi: 10.1016/j.neuint.2012.04.018
- Takahashi, H., Shiramatsu, T. I., Hitsuyu, R., Ibayashi, K., and Kawai, K. (2020). Vagus nerve stimulation (VNS)-induced layer-specific modulation of evoked responses in the sensory cortex of rats. *Sci. Rep.* 10:8932. doi: 10.1038/s41598-020-65745-z

- Thakkar, V. J., Engelhart, A. S., Khodaparast, N., Abadzi, H., and Centanni, T. M. (2020). Transcutaneous auricular vagus nerve stimulation enhances learning of novel letter-sound relationships in adults. *Brain Stimul.* 13, 1813–1820. doi: 10.1016/j.brs.2020.10.012
- Trulson, M. E., and Jacobs, B. L. (1979). Raphe unit activity in freely moving cats: correlation with level of behavioral arousal. *Brain Res.* 163, 135–150.
- Tun, P. A., Williams, V. A., Small, B. J., and Hafter, E. R. (2012). The effects of aging on auditory processing and cognition. Am. J. Audiol. 21, 344–350.
- Tyler, R., Cacace, A., Stocking, C., Tarver, B., Engineer, N., Martin, J., et al. (2017). Vagus nerve stimulation paired with tones for the treatment of tinnitus: a prospective randomized double-blind controlled pilot study in humans. *Sci. Rep.* 7:11960. doi: 10.1038/s41598-017-12178-w

Ungless, M. A. (2004). Dopamine: the salient issue. Trends Neurosci. 27, 702-706.

- Van Bockstaele, E. J., Peoples, J., and Telegan, P. (1999). Efferent projections of the nucleus of the solitary tract to peri-locus coeruleus dendrites in rat brain: evidence for a monosynaptic pathway. J. Comp. Neurol. 412, 410–428. doi: 10.1002/(sici)1096-9861(19990927)412:3<410::aid-cne3>3.0.co;2-f
- Vanneste, S., Martin, J., Rennaker, R. L. II, and Kilgard, M. P. (2017). Pairing sound with vagus nerve stimulation modulates cortical synchrony and phase coherence in tinnitus: an exploratory retrospective study. *Sci. Rep.* 7:17345. doi: 10.1038/s41598-017-17750-y
- Vazey, E. M., Moorman, D. E., and Aston-Jones, G. (2018). Phasic locus coeruleus activity regulates cortical encoding of salience information. *Proc. Natl. Acad. Sci. U.S.A.* 115, E9439–E9448. doi: 10.1073/pnas.1803716115
- Verdier, D., and Dykes, R. W. (2001). Long-term cholinergic enhancement of evoked potentials in rat hindlimb somatosensory cortex displays characteristics of long-term potentiation. *Exp. Brain Res.* 137, 71–82. doi: 10.1007/s002210 000646
- Videen, T. O., Daw, N. W., and Rader, R. K. (1984). The effect of norepinephrine on visual cortical neurons in kittens and adult cats. J. Neurosci. 4, 1607–1617.
- Vinck, M., Batista-Brito, R., Knoblich, U., and Cardin, J. A. (2015). Arousal and locomotion make distinct contributions to cortical activity patterns and visual encoding. *Neuron* 86, 740–754. doi: 10.1016/j.neuron.2015.03.028
- Wall, P. D., and Dubner, R. (1972). Somatosensory pathways. Annu. Rev. Physiol. 34, 315–336.
- Wang, Q., Millard, D. C., Zheng, H. J. V., and Stanley, G. B. (2012). Voltagesensitive dye imaging reveals improved topographic activation of cortex in response to manipulation of thalamic microstimulation parameters. *J. Neural Eng.* 9:026008. doi: 10.1088/1741-2560/9/2/026008
- Wang, Q., Webber, R., and Stanley, G. B. (2010). Thalamic synchrony and the adaptive gating of information flow to cortex. *Nat. Neurosci.* 13, 1534–1541. doi: 10.1038/nn.2670
- Waterhouse, B. D., Devilbiss, D., Seiple, S., and Markowitz, R. (2004). Sensorimotor-related discharge of simultaneously recorded, single neurons in the dorsal raphe nucleus of the awake, unrestrained rat. *Brain Res.* 1000, 183–191. doi: 10.1016/j.brainres.2003.11.030
- Waterhouse, B. D., and Navarra, R. L. (2019). The locus coeruleus-norepinephrine system and sensory signal processing: a historical review and current perspectives. *Brain Res.* 1709, 1–15. doi: 10.1016/j.brainres.2018.08.032
- Wells, M. F., Wimmer, R. D., Schmitt, L. I., Feng, G., and Halassa, M. M. (2016). Thalamic reticular impairment underlies attention deficit in Ptchd1(Y/-) mice. *Nature* 532, 58–63. doi: 10.1038/nature17427
- Weyand, T. G., Boudreaux, M., and Guido, W. (2001). Burst and tonic response modes in thalamic neurons during sleep and wakefulness. J. Neurophysiol. 85, 1107–1118.
- Wiegand, I., Tollner, T., Dyrholm, M., Muller, H. J., Bundesen, C., and Finke, K. (2014). Neural correlates of age-related decline and compensation in visual attention capacity. *Neurobiol. Aging* 35, 2161–2173. doi: 10.1016/j. neurobiolaging.2014.02.023
- Wilson, D. A., Fletcher, M. L., and Sullivan, R. M. (2004). Acetylcholine and olfactory perceptual learning. *Learn. Mem.* 11, 28–34.
- Wimmer, R. D., Schmitt, L. I., Davidson, T. J., Nakajima, M., Deisseroth, K., and Halassa, M. M. (2015). Thalamic control of sensory selection in divided attention. *Nature* 526, 705–709.
- Woolrych, A., Vautrelle, N., Reynolds, J. N. J., and Parr-Brownlie, L. C. (2021). Throwing open the doors of perception: the role of dopamine in visual processing. *Eur. J. Neurosci.* 54, 6135–6146. doi: 10.1111/ejn.15408

- Yakunina, N., Kim, S. S., and Nam, E. C. (2018). BOLD fMRI effects of transcutaneous vagus nerve stimulation in patients with chronic tinnitus. *PLoS One* 13:e0207281. doi: 10.1371/journal.pone.0207281
- Zhan, X., Yin, P., and Heinbockel, T. (2013). The basal forebrain modulates spontaneous activity of principal cells in the main olfactory bulb of anesthetized mice. *Front. Neural Circ.* 7:148. doi: 10.3389/fncir.2013.00148
- Zhang, K., Chen, C. D., and Monosov, I. E. (2019). Novelty, salience, and surprise timing are signaled by neurons in the basal forebrain. *Curr. Biol.* 29, 134.e3– 142.e3. doi: 10.1016/j.cub.2018.11.012
- Zheng, H. J. V., Wang, Q., and Stanley, G. B. (2015). Adaptive shaping of cortical response selectivity in the vibrissa pathway. J. Neurophysiol. 113, 3850–3865.
- Ziomber, A., Thor, P., Krygowska-Wajs, A., Zalecki, T., Moskala, M., Romanska, I., et al. (2012). Chronic impairment of the vagus nerve function leads to inhibition of dopamine but not serotonin neurons in rat brain structures. *Pharmacol. Rep.* 64, 1359–1367. doi: 10.1016/s1734-1140(12)70933-7

Conflict of Interest: All authors have financial interest in Sharper Sense, a company developing methods of enhancing sensory processing with VNS.

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