#### Check for updates

#### **OPEN ACCESS**

EDITED BY Jian Wang, Zhengzhou University, China

#### REVIEWED BY

Berislav Zlokovic, University of Southern California, United States Zixuan Lin, Johns Hopkins University, United States Sandeepa Sur, Johns Hopkins Medicine, United States

\*CORRESPONDENCE Jianping Jia jjp@ccmu.edu.cn

#### SPECIALTY SECTION

This article was submitted to Neurodegeneration, a section of the journal Frontiers in Neuroscience

RECEIVED 20 May 2022 ACCEPTED 18 July 2022 PUBLISHED 08 August 2022

#### CITATION

Gong M and Jia J (2022) Contribution of blood-brain barrier-related blood-borne factors for Alzheimer's disease vs. vascular dementia diagnosis: A pilot study. *Front. Neurosci.* 16:949129. doi: 10.3389/fnins.2022.949129

#### COPYRIGHT

© 2022 Gong and Jia. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Contribution of blood-brain barrier-related blood-borne factors for Alzheimer's disease vs. vascular dementia diagnosis: A pilot study

### Min Gong<sup>1</sup> and Jianping Jia<sup>1,2,3,4,5</sup>\*

<sup>1</sup>Innovation Center for Neurological Disorders and Department of Neurology, Xuanwu Hospital, Capital Medical University, National Clinical Research Center for Geriatric Diseases, Beijing, China, <sup>2</sup>Beijing Key Laboratory of Geriatric Cognitive Disorders, Beijing, China, <sup>3</sup>Clinical Center for Neurodegenerative Disease and Memory Impairment, Capital Medical University, Beijing, China, <sup>4</sup>Center of Alzheimer's Disease, Beijing Institute of Brain Disorders, Collaborative Innovation Center for Brain Disorders, Capital Medical University, Beijing, China, <sup>5</sup>Key Laboratory of Neurodegenerative Diseases, Ministry of Education, Beijing, China

**Background:** Alzheimer's disease (AD) and vascular dementia (VaD) are the two most common types of neurodegenerative dementia among the elderly with similar symptoms of cognitive decline and overlapping neuropsychological profiles. Biological markers to distinguish patients with VaD from AD would be very useful. We aimed to investigate the expression of blood-brain barrier (BBB)-related blood-borne factors of soluble lowdensity lipoprotein receptor-related protein 1 (sLRP1), cyclophilin A (CyPA), and matrix metalloproteinase 9 (MMP9) and its correlation with cognitive function between patients with AD and VaD.

**Materials and methods:** Plasma levels of sLRP1, CyPA, and MMP9 were analyzed in 26 patients with AD, 27 patients with VaD, and 27 normal controls (NCs). Spearman's rank correlation analysis was used to explore the relationships among biomarker levels, cognitive function, and imaging references. Receiver operating characteristic (ROC) curve analysis was used to discriminate the diagnosis of AD and VaD.

**Results:** Among these BBB-related factors, plasma CyPA levels in the VaD group were significantly higher than that in the AD group (p < 0.05). Plasma sLRP1 levels presented an increasing trend in VaD while maintaining slightly low levels in patients with AD (p > 0.05). Plasma MMP9 in different diagnostic groups displayed the following trend: VaD group > AD group > NC group, but the difference was not statistically significant (p > 0.05). Furthermore, plasma sLRP1 levels were positively related to MoCA scores, and plasma CyPA levels were significantly correlated with MTA scores (p < 0.05) in the AD group. Plasma MMP9 levels were negatively correlated with MoCA scores (p < 0.05) in the VaD groups. No significant correlation was detected between the other factors and different cognitive scores (p > 0.05). ROC analysis showed

a good preference of plasma CyPA [AUC = 0.725, 95% CI (0.586-0.865); p = 0.0064] in diagnosis.

**Conclusion:** The plasma CyPA level is a reference index when distinguishing between an AD and subcortical ischemic vascular dementia (SIVD) diagnosis. Blood-derived factors associated with the BBB may provide new insights into the differential diagnosis of neurodegenerative dementia and warrant further investigation.

KEYWORDS

vascular dementia (VaD), Alzheimer's disease (AD), blood-brain barrier, low-density lipoprotein receptor-related protein, cyclophilin A, matrix metalloproteinase-9

## Introduction

Dementia manifests as global cognitive decline and significantly impairs daily activities, which has imposed a heavy burden on the public and healthcare systems as society ages (Jia et al., 2020). Alzheimer's disease (AD) and vascular dementia (VaD) are the two most common types of neurodegenerative dementia among the elderly. AD has a slow course characterized by gradual deterioration of cognitive function. VaD has a variable course presented by a stepwise worsening of executive function, which can have a sudden or slow onset (O'Brien and Thomas, 2015). Although these two types of dementia share many clinical features, including symptoms of cognitive decline and overlapping neuropsychological profiles, the underlying pathophysiological mechanisms are different. Efficient therapy depends on accurate diagnosis, thus it is crucial to distinguish between VaD and AD (Neto et al., 2015; Tachibana et al., 2016). Despite advances in molecular neuroimaging, the understanding of clinicopathological relevance and the development of novel biomarkers have been limited in the last decade (Park and Moon, 2016; Bjerke and Engelborghs, 2018). Moreover, imaging is relatively expensive and invasive and is usually not immediately available for such a diagnosis. As another option, lumbar puncture is an invasive procedure, which requires written informed consent. Clinicians still need reliable and noninvasive molecular markers for the differential diagnosis of neurodegenerative dementia. Blood testing is an economical, minimally invasive and more accessible procedure and is more suitable for investigating these pathological mechanisms and distinguishing between different forms, at least as a primary screening test.

Although the etiology of AD and VaD may differ, the overall mechanisms of subsequent neurovascular dysfunction are similar, with defective blood-brain barrier (BBB) function. BBB failure is considered to be a core mechanism in vascularrelated diseases and neurodegenerative dementia, driving disease pathology and progression (Yamazaki and Kanekiyo, 2017; Cai et al., 2018; Ueno et al., 2019; Uemura et al., 2020). The breakdown of the BBB is caused by the degeneration of pericytes and endothelial cells, loss of tight junctions, and brain capillary leakages, which cause toxic molecules from the blood to enter the brain and initiate multiple neurodegenerative pathways (Bell et al., 2010; Zlokovic, 2011; Nelson et al., 2016; Storck et al., 2016; Montagne et al., 2020). Currently, it is speculated that low-density lipoprotein receptor-related protein 1 (LRP1), cyclophilin A (CyPA), and matrix metallopeptidase 9 (MMP9) are involved in the regulation of BBB permeability (Bell et al., 2012; Halliday et al., 2016). Soluble LRP1 (sLRP1) circulates freely in plasma and is primarily responsible for peripheral Aß clearance (Quinn et al., 1997). Several studies have reported a significant reduction in LRP1 expression in brain microvascular endothelial cells in AD (Shibata et al., 2000; Deane et al., 2004; Donahue et al., 2006). Circulating sLRP in the plasma binds AB and prevents brain reentry across the BBB, producing a peripheral sink that promotes the outflow of AB from the brain (Sagare et al., 2007, 2012; Deane et al., 2008). Plasma sLRP1 levels and A<sub>β</sub> binding to sLRP1 are significantly reduced due to increased levels of oxidized sLRP1, which does not bind A $\beta$ , resulting in an increase in free A $\beta$  levels in plasma to return to the brain *via* the receptor for advanced glycation end products (RAGE) in patients with AD (Deane et al., 2003; Sagare et al., 2007, 2011). CyPA is secreted by activated macrophages, lymphocytes, and platelets and mediates the harmful effects of pericytes on BBB disruption (Seizer et al., 2010, 2015, 2016; Pan et al., 2020). Previous studies have shown that the plasma CyPA level is a new biomarker for the diagnosis of coronary artery disease (CAD) and renal disease progression and is used as a prognostic factor in patients with ruptured intracranial aneurysms (Satoh et al., 2013; Ramachandran et al., 2014; Kao et al., 2015; El-Ebidi et al., 2020; Rath et al., 2020). MMP9 is involved in the increase of BBB permeability during AD, which accelerates its onset (Barr et al., 2010; Shackleton et al., 2019). These results provide support for the use of BBB-related blood-borne factors as sensitive predictors of neurodegenerative dementia (specifically BBB dysfunction-related cognitive decline) because they are tentatively present in biofluids and are involved in BBB function. Few studies have analyzed the correlation between BBB-related blood-borne factors (sLRP1, CyPA, and MMP9) and dementias.

In this clinical study, we aimed to measure plasma sLRP1, CyPA, and MMP9 levels in patients with AD, patients with VaD, and healthy controls and to evaluate their correlation with cognitive function to assist the discovery of new biomarkers.

## Materials and methods

### Study populations

A total of 80 participants were enrolled in our research who were admitted to the Department of Xuanwu Hospital, Capital Medical University from July 2019 to July 2021. More specifically, 26 patients with AD, 27 patients with subcortical ischemic vascular dementia (SIVD), and 27 agematched cognitive normal controls (NC) were recruited. The diagnosis of AD was performed according to the National Institute on Aging and the Alzheimer's Association (NIA-AA) criteria (Jack et al., 2011). The diagnosis of SIVD was performed according to the modified National Institute of Neurological Disorders and Stroke and the Association Internationale pour la Recherche et l'Enseignement en Neurosciences (NINDS-AIREN) and has evidence of ischemic lesions on brain magnetic resonance imaging (Erkinjuntti, 2003). Moreover, healthy individuals, not affected by neurodegenerative diseases, were recruited as NC. This study was approved by the ethics committee. Written informed consent was obtained before enrollment. The details of inclusion/exclusion criteria for AD and VD are shown in the Supplementary material.

## Plasma sample processing

Blood samples were taken in the morning after a 12-h fast. Notably, 6 ml of whole blood were drawn from each subject and stored in a polypropylene tube containing EDTA. Plasma separation was performed by centrifugation at 1,880  $\times$  *g* for 15 min. Finally, plasma was collected in centrifugal tubes and stored at  $-80^{\circ}$ C until analysis.

#### Measurements

The levels of sLRP1, CyPA, and MMP9 were measured in plasma using a commercially available enzyme-linked immunosorbent assay (ELISA) (LRP1: IC-LRP1-Hu, ImmunoClone, United States; CyPA: KE1726, immunoway, United States; and MMP9: KE1407, immunoway, United States) according to the manufacturer's instructions, and the dilution concentrations of sLPR1, CYPA, and MMP9 were 1:2,000,

1:10, and 1:50, respectively. The cognition of participants was assessed using the Mini-Mental State Examination (MMSE) and the Montreal Cognitive Assessment (MoCA). Total cholesterol (TC), triglyceride (TG), low-density lipoprotein cholesterol (LDL), and high-density lipoprotein cholesterol (HDL) were measured by the clinical levorotary testing center. Periventricular hyperintensity (PVH) and deep white matter hyperintense signals (DWMH) were evaluated using the Fazekas scale, and the scores of these two parts were summarized to obtain the total score (the lowest total score is 0, and the highest is 6) (Fazekas et al., 1987). A high Fazekas score is clinically associated with the diagnosis of individuals at high risk of cerebrovascular disease (Hilal et al., 2021). Concordantly, a high Fazekas score can successfully predict the cognitive function of patients with dementia in a clinical setting (Lam et al., 2021). Another, medial temporal lobe atrophy (MTA) visual rating scale has been shown to have high diagnostic accuracy for AD (Cavedo et al., 2014). It has also been reported in patients with VaD (Barber et al., 2000). In our study, ratings of PVH (0 = absence, 1 = "caps" or pencil-thin lining, 2 = smooth "halo," and 3 = irregular) and DWMH (0 = absence, 1 = punctate foci, 2 = beginning confluence of foci, and 3 = large confluent areas) were summarized into total Fazekas scores using the axial fluid-attenuated inversion recovery (FLAIR) images from a 3T MRI and were classified into low WMSA burden (Fazekas scores < 3) and high WMSA burden (Fazekas scores  $\geq$  3). MTA was dichotomized into groups of 0-1 (none to mild) vs. 2-4 (moderate to severe) by analyzing the width of the choroidal fissure, the width of the temporal horn of the lateral ventricle, and the height of the hippocampus on T1-weighted coronal sections. The total score of each patient was approved by two experienced neurologists who were blind to the clinical data. The typical images of MTA/white matter hyperintensity from patients have shown in Supplementary Figure 1.

### Statistical analysis

Descriptive statistics were used to summarize the participant characteristics. For normally distributed data, including the variables of age, TC, HDL, MMSE scores, and MoCA scores, the means  $\pm$  standard deviation (SD) was used to describe the quantitative variables. The two groups were compared using *t*-test, among groups were compared using ANOVA, and the differences between the groups were statistically significant based on further pairwise comparison *t*-test with normal distributions. For non-normally distributed data, including the variables of TG, LDL, years of education, sLRP1, CyPA, and MMP9 levels, we used medians and interquartile ranges to describe the quantitative variables. Baseline characteristics were compared using the independent-sample Kruskal-Wallis

Characteristics	AD $(n = 26)$	SIVD ( <i>n</i> = 27)	NC ( <i>n</i> = 27)	Р
Age, Mean $\pm$ SD (year)	$66.65\pm9.53$	$65.37\pm9.93$	$65.04\pm 6.12$	0.708
Male, N (%)	11(42.3)	18 (66.7)	10 (37.0)	0.068
Education, median (Q25, Q75) (year)	12.00 (9.00, 15.00)	12.00 (9.00, 13.00)	12.00 (9.00, 12.00)	0.163
TC, Mean $\pm$ SD (mmol/L)	$4.50\pm1.09$	$4.07\pm1.46$	$4.06\pm0.87$	0.262
TG, median (Q25, Q75) (mmol/L)	1.20 (1.01, 1.74)	1.34 (1.03, 1.72)	1.02 (0.79, 1.38)	0.060
LDL, median (Q25, Q75) (mmol/L)	2.55 (1.92, 3.22)	1.77 (1.35, 3.18)	2.52 (1.83, 2.95)	0.414
HDL, Mean $\pm$ SD (mmol/L)	$1.32\pm0.38$	$1.20\pm0.43$	$1.25\pm0.38$	0.537
Hypertension, N (%)	11 (42.3)	17 (63.0)	20 (74.1)	0.057
Diabetes, N (%)	6 (23.1)	7 (25.9)	9 (33.3)	0.687
Hyperlipidemia, N (%)	3 (11.5)	5 (18.5)	9 (33.3)	0.139
MMSE, Mean $\pm$ SD (score)	$19.12 \pm 5.69^{\#}$	$21.09 \pm 8.32^{\#}$	$27.63 \pm 1.28$	< 0.001
MoCA, Mean $\pm$ SD (score)	$13.60 \pm 4.87^{\#}$	$16.27 \pm 7.32^{\#}$	$25.07 \pm 1.04$	< 0.001
Fazekas WMSA burden, $N$ (%)				< 0.001
High (3–6 scores)	1 (3.8)*	23 (85.7)	NA	
Low (0–2 scores)	25 (96.2)	4 (14.3)	NA	
MTA, N (%)				0.128
None to mild (0–1 scores)	8 (30.8)	14 (52.2)	NA	
Moderate to severe (2-4 scores)	18 (69.2)	13 (47.8)	NA	

TABLE 1 Participant characteristics.

Data are shown as mean  $\pm$  SD and N (%).

\*Compared with the subcortical ischemic vascular dementia (SIVD) group, the difference was significant (p < 0.05).

<sup>#</sup>Compared with the NC group, the difference was significant (p < 0.05).

TABLE 2 sLRP1, CyPA, and MMP9 values by diagnostic groups.

Characteristics	AD $(n = 26)$	SIVD ( <i>n</i> = 27)	NC ( <i>n</i> = 27)	Н	Р
sLRP1, median (Q25, Q75) (ng/ml)	2409.65 (1544.57, 3732.41)	2896.88 <sup>a</sup> (2353.69, 4694.35)	2639.14 (2042.29, 3564.04)	3.817	0.148
CyPA, median (Q25, Q75) (ng/ml)	18.95* <sup>b</sup> (10.46, 35.05)	36.57 (17.51, 73.32)	30.78 <sup>b</sup> (21.00, 49.15)	8.302	0.016
MMP9, median (Q25, Q75) (pg/ml)	15076.43 <sup>c</sup> (9873.41,20403.41)	17076.83 (10679.13, 26947.31)	10699.70* <sup>c</sup> (8450.16, 20368.37)	3.778	0.151

Data are shown as medians (IQR).

\*Compared with the subcortical ischemic vascular dementia (SIVD) group, the difference was significant (p < 0.05); <sup>*a*</sup>Delete 2 Outliers; <sup>*b*</sup>Delete 3 Outliers; <sup>*c*</sup>Delete 5 Outliers. \*p < 0.05.



test. Categorical data were presented as proportions, and among groups were compared using the chi-square test. The Spearman correlation analysis was performed to assess the correlation between two quantitative variables, adjusted for age, sex, and education. The diagnostic value of AD and SIVD was estimated by the area under the curve (AUC) using the receiver operating characteristic (ROC) curve. p < 0.05 was considered to indicate significant results. p-values were corrected using the Bonferroni method for multiple comparison corrections. All statistical analyses were performed using SPSS version 23.0 (SPSS Inc., Chicago, IL, United States).

## **Results**

#### Demographics

The demographic and clinical characteristics of each diagnostic group are described in **Table 1**. No significant difference was found between the groups in age, gender distribution, education, and others. The differences in the MMSE, MoCA scores, and Fazekas WMSA burden among the different groups were presented (refer to **Table 1**).

# Differences in sLRP1, CyPA, and MMP9 levels between diagnostic groups

Plasma sLRP1, CyPA, and MMP9 levels in the diagnostic group are shown in **Table 2**. Plasma sLRP1 levels in the AD group showed a decreasing trend compared with that in the NC group, but the difference was not statistically significant (H = 3.817, p = 0.148). The difference in plasma CyPA levels was statistically significant across the three groups (H = 8.302, p = 0.016). Further pairwise comparison showed that plasma CyPA levels in the SIVD group were significantly higher than that in the AD group (p = 0.018). The differences in the plasma MMP9 levels were not statistically significant (H = 3.778, p = 0.151). Figure 1 shows the plasma sLRP1, CyPA, and MMP9 levels in each group.

# The correlation of plasma biomarker levels with cognition score

Spearman correlation analysis found that plasma sLRP1 levels were positively related to MoCA (r = 0.414, p = 0.040) scores in the AD groups, and plasma MMP9 levels and MoCA (r = -0.528, p = 0.014) scores showed a significant negative correlation in the SIVD groups; however, CyPA levels were not significantly correlated with cognitive scores (**Table 3** and **Supplementary Figure 2**). Correlation analysis showed a significant correlation between plasma CyPA levels and MTA scores in the AD group (r = 0.464, p = 0.017). No significant correlation was detected between the other biomarkers and imaging parameters (p > 0.05) (**Table 3**). The correlation of plasma biomarkers with other factors is shown in the **Supplementary Tables 1–4**.

# Differential diagnostic value of CyPA concentrations

The ROC analysis was finally performed to assess the differential diagnosis value in patients with AD and SIVD



group using CyPA concentration; the performance was found to be good [AUC = 0.725, 95% CI (0.586–0.865); p = 0.0064]. ROC analysis identified a cutoff value for CyPA of 60.15 ng/ml (Figure 2).

## Discussion

Our study aimed to explore the levels of BBB-related blood-borne factors sLRP1, CyPA, and MMP9 in AD, SIVD, and NC. Furthermore, the association of BBBrelated factors with cognitive function and imaging parameters was evaluated. To the best of our knowledge, this study is the first to demonstrate that BBB-related blood-borne factors are associated with different types of dementia. Our results support the use of bloodborne factors as potential biomarkers for the differential diagnosis of dementia.

This study showed that plasma CyPA levels presented a decreasing trend in the AD group, and an increasing trend in the SIVD group compared with the NC group, and the plasma CyPA level in the SIVD group was significantly higher than that in the AD group. We used plasma CyPA as a reference index for the differential diagnosis of SIVD and AD. To date, studies have shown that CyPA secretion is increased in patients with central nervous system diseases, such as cerebrovascular disease, brain trauma, and obstructive sleep apnoea with mild cognitive impairment (MCI) (Redell et al., 2007; Chang et al., 2018, 2020; Li et al., 2021). These studies have confirmed that CyPA is critical for brain damage. Furthermore, CyPA levels in endothelial cells and pericytes of the brain are elevated in patients with AD (Halliday et al., 2016). Associations between serum CyPA levels and regional gray matter volume indicated that blood levels of CyPA may reflect the pathological mechanism of AD in

Factors	sLRP1		СуРА		MMP9	
	r	Р	r	Р	r	Р
The whole sample						
MMSE	0.167	0.152	0.066	0.571	-0.128	0.276
MoCA	0.169	0.149	0.077	0.514	-0.182	0.123
Fazekas	0.089	0.543	0.185	0.203	0.151	0.309
MTA	-0.010	0.945	-0.086	0.558	-0.007	0.961
The AD groups						
MMSE	0.303	0.141	0.085	0.685	0.108	0.606
MoCA	0.414	0.040*	0.237	0.254	-0.017	0.937
Fazekas	0.053	0.797	0.259	0.201	0.167	0.416
MTA	0.346	0.083	0.464	0.017*	0.000	1.000
The SIVD groups						
MMSE	0.314	0.145	-0.139	0.527	-0.325	0.140
MoCA	0.228	0.308	-0.397	0.067	-0.528	0.014*
Fazekas	0.001	0.998	0.195	0.398	-0.248	0.278
MTA	0.197	0.367	-0.157	0.476	0.142	0.529

TABLE 3 Correlation of plasma biomarker levels with cognitive scores and imaging parameters.

Adjusted for age, sex, and education.

\*The correlation of plasma biomarker levels with cognitive scores and imaging parameters, the difference was significant (p < 0.05).

the brain (Choi et al., 2021). Given our study of CyPA, we postulate that plasma CyPA exerts a crucial association with dementia, especially when comparing AD vs. VaD. The molecular mechanisms underlying the effects of CyPA on AD and SIVD are not yet fully understood. In vitro studies have shown that CyPA induces endothelial dysfunction, the proliferation of vascular smooth muscle cells, and the migration of inflammatory cells and promotes the development of atherosclerosis (Nigro et al., 2011; Satoh et al., 2011). Pan et al. (2020) demonstrated that CyPA mediates the destruction of brain BBB by pericytes through CD147/NF-кB/MMP9 signaling and junction protein degradation, which may provide a new insights into the management by targeting CyPA and pericytes. APOE protein can control BBB integrity via the inhibition of CypA-MMP9 signaling cascades (Halliday et al., 2016; Montagne et al., 2020, 2021). Bell et al. also found that the level of CyPA in the cerebral microvessels of APOE  $\epsilon 4$  and APOE-/- mice was five to six times higher than that in the control group of APOE &2 and APOE &3 mice. Knocking out CyPA could eliminate BBB damage caused by APOE  $\varepsilon 4$  and APOE-/- mice. Moreover, treating APOE  $\varepsilon 4$ mice with low doses of cyclosporine was shown to eliminate BBB damage, indicating that the change in CyPA on the BBB is reversible, and CyPA may be a therapeutic target that causes BBB destruction (Bell et al., 2012; Montagne et al., 2021). Taken together, these results indicate that CyPA controls cerebrovascular integrity. In conjunction with our study, although the number of patients in this study was small, the differences between patients with VaD and AD were significant. These results suggest that plasma CyPA measurements, while not diagnostic, might be combined with psychometric and imaging references to improve the early differentiation between VaD and AD, which may help to better select patients in future clinical trials. To the best of our knowledge, this is the first study to describe elevated plasma CyPA levels in patients with VaD.

Our results provide evidence of differences in plasma CyPA in AD vs. VaD. We also found that plasma CyPA levels were associated with MTA scores in the AD group; however, CyPA levels were not significantly correlated with cognitive scores. The breakdown of the BBB initially occurs in the hippocampus during normal aging, which is a key area for memory. Disruption of the BBB in the hippocampus is associated with MCI, which is in turn associated with damage to pericytes (Montagne et al., 2015). Individuals with early cognitive impairment, regardless of changes in A\beta and/or tau biomarkers of AD, develop brain capillary damage and disruption of the hippocampal BBB, indicating that BBB disruption is an early biomarker of human cognitive dysfunction independent of  $A\beta$  and tau (Nation et al., 2019). These results indicate that CyPA is involved in the brain structure of the hippocampus via the BBB, which may be an early event in the aging human brain that begins in the hippocampus and is reflected by CyPA. Thus, CyPA may predict early BBB dysfunction in the hippocampus in AD; however, the mechanism requires further exploration. Another study showed that CyPA and MMP9 levels in serum were associated with cognitive impairment and white matter signal abnormalities, which is controversial in our study (Li et al., 2021). The

correlation between CyPA levels and cognitive function requires further verification.

Lipoprotein receptor-related protein 1, a major transporter in the brain-to-blood clearance of AB across the BBB, is associated with cognitive decline in AD (Deane et al., 2004; Storck et al., 2016; Ma et al., 2018). Our study found that lower plasma sLRP1 levels were associated with cognitive decline in patients with AD. A previous study showed that cannabinoid treatment enhanced AB transfer at the BBB, accompanied by increased brain and plasma sLRP1 levels (Bachmeier et al., 2013). This finding is consistent with those of this study. Another study found that BBB-associated pericytes cleared AB aggregation through LRP1/APOE subtype-specific mechanisms, supporting the role of the LRP1/APOE interaction as a potential therapeutic target for controlling AB clearance in AD (Ma et al., 2018). Masaya et al. identified LRP1 as the potential molecular mechanism by which APOE £4 in patients with AD intensifies the deposition of  $A\beta$  protein in the brain, which was seen both in a mouse model of AD and in autopsies of patients with AD (Tachibana et al., 2019). Halliday et al. also suggested that APOE £4 leads to the accelerated loss of the LRP1dependent CyPA-MMP9 BBB degradation pathway in pericytes and endothelial cells (Halliday et al., 2016; Nikolakopoulou et al., 2021). A recent study suggested that the loss of brain endothelial LRP1 results in the loss of BBB integrity, neuronal loss, and cognitive deficits in mice, which could be reversed by endothelial-specific LRP1 gene therapy (Nikolakopoulou et al., 2021). Thus, increased cerebral LRP1 and plasma sLRP may explain the increased AB BBB transport and may provide an effective strategy to reduce the  $A\beta$  burden in the AD brain. At present, many studies have focused on the inhibition of  $A\beta$ production. However, the occurrence and development of AD are believed to be due to the reduction in A $\beta$ , rather than the excessive production of AB (Mawuenyega et al., 2010; Xiang et al., 2015; Wang et al., 2017). Our study focused on Aβ clearance-related proteins and enzymes in AD, which can help discover their potential in the development of AD drugs and provide an optimistic prospect for future therapeutic targets. Furthermore, previous studies found downregulation of LRP1 protein and mRNA expression in VaD rats via the IKK/NFκB signaling pathway (Cai et al., 2020; Wang et al., 2020). Nevertheless, Mercedes et al. proposed that serum sLRP1 can serve as a candidate marker to differentiate AD from mixed dementia phenotypes, with a significant increase in sLRP1 serum protein levels in subjects with mixed dementia and relatively normal levels in AD (Lachén-Montes et al., 2021). Our study also showed that plasma sLRP1 levels presented an increasing trend in VaD while maintaining slightly low levels in patients with AD. This inconsistency may be attributed to the relatively small sample size, different samples, and methods used. Further research is needed to determine whether and how sLRP1 in the blood affects cognitive preference in patients with dementia.

Matrix metallopeptidase 9 is a major protein related to brain disorders and the BBB (Rempe et al., 2016). Inhibition of MMP prevented tight junction protein loss, suggesting that MMP interferes with barrier integrity by degrading tight junction proteins (Yang et al., 2007). Previous studies have found that plasma MMP-9 levels of patients with AD are higher than those in normal controls through an increase in BBB permeability (Lorenzl et al., 2003, 2008). However, Horstmann et al. (2010) showed that plasma MMP9 activity was decreased by 41% in patients with AD compared with that in normal controls. Another study verified that the damage of MMP9 to neurovascular units is thought to be related to vascular cognitive dysfunction by increasing the BBB opening (Candelario-Jalil et al., 2011). The level of MMP9 in the CSF of patients with VaD was higher than that in the AD and normal control groups (Adair et al., 2004). In this study, plasma MMP9 in different diagnostic groups displayed the following trend: SIVD group > AD group > NC group, and higher plasma MMP9 was correlated with cognitive decline, which was broadly consistent with previous studies. MMP9 may increase the permeability of the BBB, activate inflammatory factors, cause various inflammatory responses in cells, and accelerate the onset of disease. Thus, MMP9 may contribute to the course of senile neurodegenerative dementia and may play a role in predicting cognitive function in the periphery. Further studies are required to explore the role of MMP in AD progression.

Our study has some limitations. First, due to the relatively small number of experimental studies and limited research participants, the quality of the statistical analysis may be affected; second, cross-sectional studies have limited causality, and a longitudinal follow-up study is needed to elucidate the relationship between CyPA, sLRP1, and MMP9 levels and disease progression; third, the diagnostic value of a single biomarker is limited, and combinations with other biomarkers should be considered in the future to be used to predict the diagnosis of neurodegenerative dementia; fourth, considering the effect of APOE on neurovascular injury and neuronal dysfunction, clarifying the relationship of BBB-related bloodborne factors with APOE genotypes is required in the future; fifth, as the assessment of imaging parameters uses visual rating, adding some quantitative volumetric analysis such as hippocampal volume would be informative in future validation studies; the assessment of scales for other cognitive domains and their correlations with biomarkers should also be considered; and finally, we did not have additional information to clarify the mechanism of BBB dysfunction.

In summary, our study suggests that the plasma CyPA level is a reference index when distinguishing between an AD and SIVD diagnosis. sLRP1 and MMP-9 may be ideal biomarkers of cognitive decline. Our findings highlight that blood-derived factors associated with the BBB may provide new insights into the differential diagnosis of neurodegenerative dementia and warrant further investigation.

## Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

## **Ethics statement**

The studies involving human participants were reviewed and approved by the Ethics Review Board of the Xuanwu Hospital. The patients/participants provided their written informed consent to participate in this study.

## Author contributions

JJ supervised and obtained funding for this study. MG did the statistical analysis. Both authors designed the study, critically revised and drafted the manuscript, contributed to the collection, analysis, and interpretation of data, and approved the final version of the manuscript.

## Funding

This study was supported by the Key Project of the National Natural Science Foundation of China (U20A20354), the Beijing Brain Initiative from Beijing Municipal Science & Technology Commission (Z201100005520016 and Z201100005520017), the National Major R&D Projects of China-Scientific Technological Innovation 2030 (2021ZD0201802), the National Key Scientific Instrument and Equipment Development Project (31627803),

### References

Adair, J. C., Charlie, J., Dencoff, J. E., Kaye, J. A., Quinn, J. F., Camicioli, R. M., et al. (2004). Measurement of gelatinase B (MMP-9) in the cerebrospinal fluid of patients with vascular dementia and Alzheimer disease. *Stroke* 35, e159–62. doi: 10.1161/01.Str.0000127420.10990.76

Bachmeier, C., Beaulieu-Abdelahad, D., Mullan, M., and Paris, D. (2013). Role of the cannabinoid system in the transit of beta-amyloid across the blood-brain barrier. *Mol. Cell. Neurosci.* 56, 255–262. doi: 10.1016/j.mcn.2013.06.004

Barber, R., Ballard, C., McKeith, I. G., Gholkar, A., and O'Brien, J. T. (2000). MRI volumetric study of dementia with Lewy bodies: A comparison with AD and vascular dementia. *Neurology* 54, 1304–1309. doi: 10.1212/wnl.54.6.1304

Barr, T. L., Latour, L. L., Lee, K. Y., Schaewe, T. J., Luby, M., Chang, G. S., et al. (2010). Blood-brain barrier disruption in humans is independently associated with increased matrix metalloproteinase-9. *Stroke* 41, e123–8. doi: 10.1161/strokeaha. 109.570515

Bell, R. D., Winkler, E. A., Sagare, A. P., Singh, I., LaRue, B., Deane, R., et al. (2010). Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron* 68, 409–427. doi: 10.1016/j.neuron. 2010.09.043

Bell, R. D., Winkler, E. A., Singh, I., Sagare, A. P., Deane, R., Wu, Z., et al. (2012). Apolipoprotein E controls cerebrovascular integrity via cyclophilin A. *Nature* 485, 512–516. doi: 10.1038/nature11087 and the Key Project of the National Natural Science Foundation of China (81530036).

## Acknowledgments

We thank the patients for their participation.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

# Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fnins.2022.949129/full#supplementary-material

Bjerke, M., and Engelborghs, S. (2018). Cerebrospinal Fluid Biomarkers for Early and Differential Alzheimer's Disease Diagnosis. *J. Alzheimers Dis.* 62, 1199– 1209. doi: 10.3233/jad-170680

Cai, H., Cai, T., Zheng, H., Liu, L., Zhou, L., Pang, X., et al. (2020). The Neuroprotective Effects of Danggui-Shaoyao San on Vascular Cognitive Impairment: Involvement of the Role of the Low-Density Lipoprotein Receptor-Related Protein. *Rejuvenation Res.* 23, 420–433. doi: 10.1089/rej.2019.2182

Cai, Z., Qiao, P. F., Wan, C. Q., Cai, M., Zhou, N. K., and Li, Q. (2018). Role of Blood-Brain Barrier in Alzheimer's Disease. *J. Alzheimers Dis.* 63, 1223–1234. doi: 10.3233/jad-180098

Candelario-Jalil, E., Thompson, J., Taheri, S., Grossetete, M., Adair, J. C., Edmonds, E., et al. (2011). Matrix metalloproteinases are associated with increased blood-brain barrier opening in vascular cognitive impairment. *Stroke* 42, 1345–1350. doi: 10.1161/strokeaha.110.600825

Cavedo, E., Pievani, M., Boccardi, M., Galluzzi, S., Bocchetta, M., Bonetti, M., et al. (2014). Medial temporal atrophy in early and late-onset Alzheimer's disease. *Neurobiol. Aging* 35, 2004–2012. doi: 10.1016/j.neurobiolaging.2014.03.009

Chang, C. S., Kuo, C. L., Huang, C. S., Cheng, Y. S., Lin, S. S., and Liu, C. S. (2020). Association of cyclophilin A level and pulse pressure in predicting recurrence of cerebral infarction. *Kaohsiung J. Med. Sci.* 36, 122–128. doi: 10.1002/kjm2.12143

Chang, C. S., Su, S. L., Kuo, C. L., Huang, C. S., Tseng, W. M., Lin, S. S., et al. (2018). Cyclophilin A: A Predictive Biomarker of Carotid Stenosis in Cerebral Ischemic Stroke. *Curr. Neurovasc. Res.* 15, 111–119. doi: 10.2174/1567202615666180516120959

Choi, H. I., Kim, K., Lee, J., Chang, Y., Rhee, H. Y., Park, S., et al. (2021). Relationship between Brain Tissue Changes and Blood Biomarkers of Cyclophilin A, Heme Oxygenase-1, and Inositol-Requiring Enzyme 1 in Patients with Alzheimer's Disease. *Diagnostics* 11:740. doi: 10.3390/diagnostics11050740

Deane, R., Du Yan, S., Submamaryan, R. K., LaRue, B., Jovanovic, S., Hogg, E., et al. (2003). RAGE mediates amyloid-beta peptide transport across the bloodbrain barrier and accumulation in brain. *Nat. Med.* 9, 907–913. doi: 10.1038/ nm890

Deane, R., Sagare, A., and Zlokovic, B. V. (2008). The role of the cell surface LRP and soluble LRP in blood-brain barrier Abeta clearance in Alzheimer's disease. *Curr. Pharm. Des.* 14, 1601–1605. doi: 10.2174/138161208784705487

Deane, R., Wu, Z., Sagare, A., Davis, J., Du Yan, S., Hamm, K., et al. (2004). LRP/amyloid beta-peptide interaction mediates differential brain efflux of Abeta isoforms. *Neuron* 43, 333–344. doi: 10.1016/j.neuron.2004.07.017

Donahue, J. E., Flaherty, S. L., Johanson, C. E., Duncan, J. A. III, Silverberg, G. D., Miller, M. C., et al. (2006). RAGE, LRP-1, and amyloid-beta protein in Alzheimer's disease. *Acta Neuropathol.* 112, 405–415. doi: 10.1007/s00401-006-0115-3

El-Ebidi, A. M., Saleem, T. H., Saadi, M. G. E., Mahmoud, H. A., Mohamed, Z., and Sherkawy, H. S. (2020). Cyclophilin A (CyPA) as a Novel Biomarker for Early Detection of Diabetic Nephropathy in an Animal Model. *Diabetes Metab. Syndr. Obes.* 13, 3807–3819. doi: 10.2147/dmso.S260293

Erkinjuntti, T. (2003). Subcortical ischemic vascular disease and dementia. *Int. Psychogeriatr.* 15, 23–26. doi: 10.1017/s1041610203008925

Fazekas, F., Chawluk, J. B., Alavi, A., Hurtig, H. I., and Zimmerman, R. A. (1987). MR signal abnormalities at 1.5 T in Alzheimer's dementia and normal aging. *AJR Am. J. Roentgenol.* 149, 351–356. doi: 10.2214/ajr.149.2.351

Halliday, M. R., Rege, S. V., Ma, Q., Zhao, Z., Miller, C. A., Winkler, E. A., et al. (2016). Accelerated pericyte degeneration and blood-brain barrier breakdown in apolipoprotein E4 carriers with Alzheimer's disease. *J. Cereb. Blood Flow Metab.* 36, 216–227. doi: 10.1038/jcbfm.2015.44

Hilal, S., Liu, S., Wong, T. Y., Vrooman, H., Cheng, C. Y., Venketasubramanian, N., et al. (2021). White matter network damage mediates association between cerebrovascular disease and cognition. *J. Cereb. Blood Flow Metab.* 41, 1858–1872. doi: 10.1177/0271678x21990980

Horstmann, S., Budig, L., Gardner, H., Koziol, J., Deuschle, M., Schilling, C., et al. (2010). Matrix metalloproteinases in peripheral blood and cerebrospinal fluid in patients with Alzheimer's disease. *Int. Psychogeriatr.* 22, 966–972. doi: 10.1017/s1041610210000827

Jack, C. R. Jr., Albert, M. S., Knopman, D. S., McKhann, G. M., Sperling, R. A., Carrillo, M. C., et al. (2011). Introduction to the recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement.* 7, 257–262. doi: 10.1016/j. jalz.2011.03.004

Jia, L., Quan, M., Fu, Y., Zhao, T., Li, Y., Wei, C., et al. (2020). Dementia in China: Epidemiology, clinical management, and research advances. *Lancet Neurol.* 19, 81–92. doi: 10.1016/s1474-4422(19)30290-x

Kao, H. W., Lee, K. W., Chen, W. L., Kuo, C. L., Huang, C. S., Tseng, W. M., et al. (2015). Cyclophilin A in Ruptured Intracranial Aneurysm: A Prognostic Biomarker. *Medicine* 94:e1683. doi: 10.1097/md.000000000001683

Lachén-Montes, M., Íñigo-Marco, I., Cartas-Cejudo, P., Fernández-Irigoyen, J., and Santamaría, E. (2021). Olfactory Bulb Proteomics Reveals Widespread Proteostatic Disturbances in Mixed Dementia and Guides for Potential Serum Biomarkers to Discriminate Alzheimer Disease and Mixed Dementia Phenotypes. J. Pers. Med. 11:503. doi: 10.3390/jpm11060503

Lam, S., Lipton, R. B., Harvey, D. J., Zammit, A. R., and Ezzati, A. (2021). White matter hyperintensities and cognition across different Alzheimer's biomarker profiles. J. Am. Geriatr. Soc. 69, 1906–1915. doi: 10.1111/jgs.17173

Li, M., Sun, H., Shen, T., Xue, S., Zhao, Y., Leng, B., et al. (2021). Increased serum levels of cyclophilin a and matrix metalloproteinase-9 are associated with cognitive impairment in patients with obstructive sleep apnea. *Sleep Med.* 93, 75–83. doi: 10.1016/j.sleep.2021.10.009

Lorenzl, S., Albers, D. S., Relkin, N., Ngyuen, T., Hilgenberg, S. L., Chirichigno, J., et al. (2003). Increased plasma levels of matrix metalloproteinase-9 in patients with Alzheimer's disease. *Neurochem. Int.* 43, 191–196. doi: 10.1016/s0197-0186(03)00004-4

Lorenzl, S., Buerger, K., Hampel, H., and Beal, M. F. (2008). Profiles of matrix metalloproteinases and their inhibitors in plasma of patients with dementia. *Int. Psychogeriatr.* 20, 67–76. doi: 10.1017/s1041610207005790

Ma, Q., Zhao, Z., Sagare, A. P., Wu, Y., Wang, M., Owens, N. C., et al. (2018). Blood-brain barrier-associated pericytes internalize and clear aggregated amyloid-β42 by LRP1-dependent apolipoprotein E isoform-specific mechanism. *Mol. Neurodegener.* 13:57. doi: 10.1186/s13024-018-0286-0

Mawuenyega, K. G., Sigurdson, W., Ovod, V., Munsell, L., Kasten, T., Morris, J. C., et al. (2010). Decreased clearance of CNS beta-amyloid in Alzheimer's disease. *Science* 330:1774. doi: 10.1126/science.1197623

Montagne, A., Barnes, S. R., Sweeney, M. D., Halliday, M. R., Sagare, A. P., Zhao, Z., et al. (2015). Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* 85, 296–302. doi: 10.1016/j.neuron.2014.12.032

Montagne, A., Nation, D. A., Sagare, A. P., Barisano, G., Sweeney, M. D., Chakhoyan, A., et al. (2020). APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline. *Nature* 581, 71–76. doi: 10.1038/s41586-020-2 247-3

Montagne, A., Nikolakopoulou, A. M., Huuskonen, M. T., Sagare, A. P., Lawson, E. J., Lazic, D., et al. (2021). APOE4 accelerates advanced-stage vascular and neurodegenerative disorder in old Alzheimer's mice via cyclophilin A independently of amyloid- $\beta$ . *Nat. Aging* 1, 506–520. doi: 10.1038/s43587-021-00073-z

Nation, D. A., Sweeney, M. D., Montagne, A., Sagare, A. P., D'Orazio, L. M., Pachicano, M., et al. (2019). Blood-brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nat. Med.* 25, 270–276. doi: 10.1038/s41591-018-0297-y

Nelson, A. R., Sweeney, M. D., Sagare, A. P., and Zlokovic, B. V. (2016). Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease. *Biochim. Biophys. Acta* 1862, 887–900. doi: 10.1016/j.bbadis.2015.12.016

Neto, E., Allen, E. A., Aurlien, H., Nordby, H., and Eichele, T. (2015). EEG Spectral Features Discriminate between Alzheimer's and Vascular Dementia. *Front. Neurol.* 6:25. doi: 10.3389/fneur.2015.00025

Nigro, P., Satoh, K., O'Dell, M. R., Soe, N. N., Cui, Z., Mohan, A., et al. (2011). Cyclophilin A is an inflammatory mediator that promotes atherosclerosis in apolipoprotein E-deficient mice. *J. Exp. Med.* 208, 53–66. doi: 10.1084/jem. 20101174

Nikolakopoulou, A. M., Wang, Y., Ma, Q., Sagare, A. P., Montagne, A., Huuskonen, M. T., et al. (2021). Endothelial LRP1 protects against neurodegeneration by blocking cyclophilin A. *J. Exp. Med.* 218:e20202207. doi: 10.1084/jem.20202207

O'Brien, J. T., and Thomas, A. (2015). Vascular dementia. *Lancet* 386, 1698–1706. doi: 10.1016/s0140-6736(15)00463-8

Pan, P., Zhao, H., Zhang, X., Li, Q., Qu, J., Zuo, S., et al. (2020). Cyclophilin a signaling induces pericyte-associated blood-brain barrier disruption after subarachnoid hemorrhage. *J. Neuroinflammation* 17:16. doi: 10.1186/s12974-020-1699-6

Park, M., and Moon, W. J. (2016). Structural MR Imaging in the Diagnosis of Alzheimer's Disease and Other Neurodegenerative Dementia: Current Imaging Approach and Future Perspectives. *Korean J. Radiol.* 17, 827–845. doi: 10.3348/ kjr.2016.17.6.827

Quinn, K. A., Grimsley, P. G., Dai, Y. P., Tapner, M., Chesterman, C. N., and Owensby, D. A. (1997). Soluble low density lipoprotein receptor-related protein (LRP) circulates in human plasma. *J. Biol. Chem.* 272, 23946–23951. doi: 10.1074/ jbc.272.38.23946

Ramachandran, S., Venugopal, A., Kutty, V. R., Chitrasree, V., Mullassari, A., Pratapchandran, N. S., et al. (2014). Plasma level of cyclophilin A is increased in patients with type 2 diabetes mellitus and suggests presence of vascular disease. *Cardiovasc. Diabetol.* 13:38. doi: 10.1186/1475-2840-13-38

Rath, D., von Ungern-Sternberg, S., Heinzmann, D., Sigle, M., Monzien, M., Horstmann, K., et al. (2020). Platelet surface expression of cyclophilin A is associated with increased mortality in patients with symptomatic coronary artery disease. *J. Thromb. Haemost.* 18, 234–242. doi: 10.1111/jth.14635

Redell, J. B., Zhao, J., and Dash, P. K. (2007). Acutely increased cyclophilin a expression after brain injury: A role in blood-brain barrier function and tissue preservation. *J. Neurosci. Res.* 85, 1980–1988. doi: 10.1002/jnr.2 1324

Rempe, R. G., Hartz, A. M. S., and Bauer, B. (2016). Matrix metalloproteinases in the brain and blood-brain barrier: Versatile breakers and makers. *J. Cereb. Blood Flow Metab.* 36, 1481–1507. doi: 10.1177/0271678x16655551

Sagare, A., Deane, R., Bell, R. D., Johnson, B., Hamm, K., Pendu, R., et al. (2007). Clearance of amyloid-beta by circulating lipoprotein receptors. *Nat. Med.* 13, 1029–1031. doi: 10.1038/nm1635

Sagare, A. P., Deane, R., Zetterberg, H., Wallin, A., Blennow, K., and Zlokovic, B. V. (2011). Impaired lipoprotein receptor-mediated peripheral binding of plasma amyloid-β is an early biomarker for mild cognitive impairment preceding Alzheimer's disease. *J Alzheimers Dis.* 24, 25–34. doi: 10.3233/jad-2010-101248

Sagare, A. P., Deane, R., and Zlokovic, B. V. (2012). Low-density lipoprotein receptor-related protein 1: A physiological A $\beta$  homeostatic mechanism with multiple therapeutic opportunities. *Pharmacol. Ther.* 136, 94–105. doi: 10.1016/j.pharmthera.2012.07.008

Satoh, K., Fukumoto, Y., Sugimura, K., Miura, Y., Aoki, T., Nochioka, K., et al. (2013). Plasma cyclophilin A is a novel biomarker for coronary artery disease. *Circ. J.* 77, 447–455. doi: 10.1253/circj.cj-12-0805

Satoh, K., Nigro, P., Zeidan, A., Soe, N. N., Jaffré, F., Oikawa, M., et al. (2011). Cyclophilin A promotes cardiac hypertrophy in apolipoprotein E-deficient mice. *Arterioscler. Thromb. Vasc. Biol.* 31, 1116–1123. doi: 10.1161/atvbaha.110.21 4601

Seizer, P., Fuchs, C., Ungern-Sternberg, S. N., Heinzmann, D., Langer, H., Gawaz, M., et al. (2016). Platelet-bound cyclophilin A in patients with stable coronary artery disease and acute myocardial infarction. *Platelets* 27, 155–158. doi: 10.3109/09537104.2015.1051466

Seizer, P., Schönberger, T., Schött, M., Lang, M. R., Langer, H. F., Bigalke, B., et al. (2010). EMMPRIN and its ligand cyclophilin A regulate MT1-MMP, MMP-9 and M-CSF during foam cell formation. *Atherosclerosis* 209, 51–57. doi: 10.1016/j.atherosclerosis.2009.08.029

Seizer, P., Ungern-Sternberg, S. N., Schönberger, T., Borst, O., Münzer, P., Schmidt, E. M., et al. (2015). Extracellular cyclophilin A activates platelets via EMMPRIN (CD147) and PI3K/Akt signaling, which promotes platelet adhesion and thrombus formation in vitro and in vivo. Arterioscler. Thromb. Vasc. Biol. 35, 655–663. doi: 10.1161/atvbaha.114.305112

Shackleton, B., Ringland, C., Abdullah, L., Mullan, M., Crawford, F., and Bachmeier, C. (2019). Influence of Matrix Metallopeptidase 9 on Beta-Amyloid Elimination Across the Blood-Brain Barrier. *Mol. Neurobiol.* 56, 8296–8305. doi: 10.1007/s12035-019-01672-z

Shibata, M., Yamada, S., Kumar, S. R., Calero, M., Bading, J., Frangione, B., et al. (2000). Clearance of Alzheimer's amyloid-ss(1-40) peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier. *J. Clin. Invest.* 106, 1489–1499. doi: 10.1172/jci10498

Storck, S. E., Meister, S., Nahrath, J., Meißner, J. N., Schubert, N., Di Spiezio, A., et al. (2016). Endothelial LRP1 transports amyloid- $\beta$ (1-42) across the blood-brain barrier. *J. Clin. Invest.* 126, 123–136. doi: 10.1172/jci81108

Tachibana, H., Washida, K., Kowa, H., Kanda, F., and Toda, T. (2016). Vascular Function in Alzheimer's Disease and Vascular Dementia. *Am. J. Alzheimers Dis. Other Demen.* 31, 437–442. doi: 10.1177/1533317516653820

Tachibana, M., Holm, M. L., Liu, C. C., Shinohara, M., Aikawa, T., Oue, H., et al. (2019). APOE4-mediated amyloid- $\beta$  pathology depends on its neuronal receptor LRP1. J. Clin. Invest. 129, 1272–1277. doi: 10.1172/jci124853

Uemura, M. T., Maki, T., Ihara, M., Lee, V. M. Y., and Trojanowski, J. Q. (2020). Brain Microvascular Pericytes in Vascular Cognitive Impairment and Dementia. *Front. Aging Neurosci.* 12:80. doi: 10.3389/fnagi.2020.00080

Ueno, M., Chiba, Y., Murakami, R., Matsumoto, K., Fujihara, R., Uemura, N., et al. (2019). Disturbance of Intracerebral Fluid Clearance and Blood-Brain Barrier in Vascular Cognitive Impairment. *Int. J. Mol. Sci.* 20:2600. doi: 10.3390/ ijms20102600

Wang, J., Gu, B. J., Masters, C. L., and Wang, Y. J. (2017). A systemic view of Alzheimer disease - insights from amyloid- $\beta$  metabolism beyond the brain. *Nat. Rev. Neurol.* 13, 612–623. doi: 10.1038/nrneurol.2017.111

Wang, J. J., Wu, S. B., Cheng, H. L., Yang, C., Chen, Y., and Yang, J. (2020). [Effect of moxibustion on expression of RAGE and LRP-1 and neuronal ultrastructure of frontal cortex and hippocampus in vascular dementia rats]. *Zhen Ci Yan Jiu* 45, 33–39. doi: 10.13702/j.1000-0607.1902956

Xiang, Y., Bu, X. L., Liu, Y. H., Zhu, C., Shen, L. L., Jiao, S. S., et al. (2015). Physiological amyloid-beta clearance in the periphery and its therapeutic potential for Alzheimer's disease. *Acta Neuropathol.* 130, 487–499. doi: 10.1007/s00401-015-1477-1

Yamazaki, Y., and Kanekiyo, T. (2017). Blood-Brain Barrier Dysfunction and the Pathogenesis of Alzheimer's Disease. *Int. J. Mol. Sci.* 18:1965. doi: 10.3390/ ijms18091965

Yang, Y., Estrada, E. Y., Thompson, J. F., Liu, W., and Rosenberg, G. A. (2007). Matrix metalloproteinase-mediated disruption of tight junction proteins in cerebral vessels is reversed by synthetic matrix metalloproteinase inhibitor in focal ischemia in rat. *J. Cereb. Blood Flow Metab.* 27, 697–709. doi: 10.1038/sj.jcbfm. 9600375

Zlokovic, B. V. (2011). Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat. Rev. Neurosci.* 12, 723-738. doi: 10.1038/nrn3114