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The contribution of mitochondria-associated endoplasmic reticulum membranes (MAMs) dysfunction in Alzheimer's disease and the potential countermeasure

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Alzheimer's disease (AD) is the most common neurodegenerative disease. There are many studies targeting extracellular deposits of amyloid β -peptide (A β) and intracellular neurofibrillary tangles (NFTs), however, there are no effective treatments to halt the progression. Mitochondria-associated endoplasmic reticulum membranes (MAMs) have long been found to be associated with various pathogenesis hypotheses of AD, such as A β deposition, mitochondrial dysfunction, and calcium homeostasis. However, there is a lack of literature summarizing recent advances in the mechanism and treatment studies. Accordingly, this article reviews the latest research involving the roles of MAM structure and tethering proteins in the pathogenesis of AD and summarizes potential strategies targeting MAMs to dissect treatment perspectives for AD.

KEYWORDS

Alzheimer's disease, mitochondria, endoplasmic reticulum, calcium homeostasis, lipid metabolism

Introduction

According to Alzheimer's disease (AD) International, 55 million people worldwide suffer from dementia. The most common type of dementia is AD. The main histopathological characteristics of AD are the accumulation of extracellular deposits of amyloid β -peptide (A β) and intracellular neurofibrillary tangles (NFTs). However, there are other biochemical and morphological characteristics present earlier in the course of AD, such as alterations in phospholipid metabolism, the elevation of circulating cholesterol levels, aberrant calcium regulation, reduction of brain glucose levels, and mitochondrial dysfunction. Plaques and tangles have received a lot of attention because they are observable physical entities, however, the other features may be the upstream factors and should not be overlooked (Area-Gomez and Schon, 2017).

Mitochondria-associated endoplasmic reticulum membranes (MAMs) are a special subdomain of the endoplasmic reticulum (ER) that physically and biologically connects mitochondria to ER. ER-mitochondrial communication and MAM functions are increased expressively in AD (Leal et al., 2020). MAMs are generated by the ER side-by-side to

the mitochondrial outer membrane linked by a series of tethering proteins (Degechisa et al., 2022). They are not the collection of membranes but the employment of the proteinaceous tethers (Rowland and Voeltz, 2012), which play a key role in many important metabolic processes including the transfer of calcium, mitochondrial dynamics, lipid synthesis, autophagy, apoptosis, and inflammation. MAM dysfunction is also central to the pathogenic mechanisms of AD, especially in the A β generation and deposition, mitochondrial dysfunction, imbalanced calcium homeostasis, abnormal lipids metabolism, and abnormal autophagy. This review highlights abnormal MAM structures and tethering proteins in the pathogenesis of AD from these aspects. In addition, we provide a summary of compounds and drugs that target MAM tethering proteins in AD models.

The role of MAMs in A β generation of AD

The A β peptide is the main component of the AD hallmark amyloid plaques, which is produced by the proteolysis of amyloid beta-precursor protein (APP) by two enzymes: β -site APP cleaving enzyme 1 (BACE1) and the γ -secretase complex. Many studies have shown that MAMs are the main site of $A\beta$ generation. Schreiner et al. (2015) reported that A^β may be generated directly in MAMs. APP, BACE1, and y-secretase have all been found in MAMs (Leal et al., 2020). BACE1 cleaves APP to produce sAPPß and C99. C99 is delivered to MAMs and cleaved to produce A β and APP intracellular domain (AICD) by γ -secretase (Montesinos et al., 2020). In addition, it has been reported that the main pathway for $A\beta$ entering mitochondria is MAMs (Del Prete et al., 2017). Knockdown of mitofusin-2 (Mfn2), which is involved in MAM tethering, leads to decreased contact between mitochondria and ER, resulting in lower y-secretase activity and decreased concentrations of intracellular and extracellular $A\beta_{40}$ and $A\beta_{42}$ (Leal et al., 2016). This proves that the increase of MAMs may promote mitochondrial Aß deposition. Axonal generation of AB also plays a key role in AD pathology. In AD models, swollen axons contain high levels of BACE1 (Gowrishankar et al., 2017). An average of 37 \pm 4% of the total $A\beta_{40}$ secreted from each mouse hippocampal neuron is secreted by axons (Niederst et al., 2015). Bhattacharyya et al. showed that down-regulation of MAM assembly by silencing MAM-resident sigma-1 receptor expression resulted in reduced palmitoylated APP cleavage by BACE1, thereby decreasing Aβ generation in neuronal processes and axons (Bhattacharyya et al., 2021).

The role of MAMs in mitochondria dysfunction of AD

The neuronal activity must depend on the energy produced by mitochondria. Mitochondrial death can be observed before the histopathological features of AD appear. MAMs wrap around the location where mitochondria undergo fissuring by recruiting MAM protein inverted formin 2 (INF2) (Steffen and Koehler, 2018). The mitochondrial outer membrane protein FUNDC1 is a MAM protein that recruits dynamin 1 Like (DNM1L)/dynaminrelated protein 1 (DRP1) to drive mitochondrial fission (Wu et al., 2016). In an AD mouse model, alterations in MAMs precede changes in mitochondrial dynamics accompanied by aberrations in mitochondrial membrane potential (MMP) and ATP production (Leal et al., 2020). Moreover, the MAMs control mitochondrial Ca²⁺ intake. Reduced Ca²⁺ intake affects mitochondrial metabolism, leading to mitochondrial dysfunction. Fernandes et al. (2021) found a decrease in close ER-mitochondria contacts, a reduction of Ca²⁺ transfer from ER to mitochondria, and impaired mitochondrial function in APPswe cells. y-secretase activating protein (GSAP) fluorescence staining showed high colocalization with a MAM marker Facl4. Knockdown of GSAP significantly increased mitochondrial basal respiration and total ATP levels, which suggests that GSAP impairs mitochondria function (Xu et al., 2021). In an AD cell model, increased concentration of unprocessed C99 in the MAM region increased sphingolipid turnover and altered the lipid composition of mitochondrial membranes, which can interfere with the normal activity of the respiratory supercomplexes and thus may contribute to the bioenergetic defects in AD (Pera et al., 2017). Atlastin 2 (ATL2) is a protein associated with ER-mitochondria contacts whose expression was significantly increased both in $3 \times Tg$ mice and AD patients. While, knockout of ATL2 rescued the elevated ER-mitochondria contacts back to normal levels, reduced the abnormally elevated mitochondrial superoxide, and significantly increased the MMP (Han et al., 2021).

The role of MAMs in calcium homeostasis of AD

Calcium signaling in neurons is essential for neurotransmission and the maintenance of synaptic plasticity (Skobeleva et al., 2022). Dysregulation of calcium homeostasis disrupts neuronal and synaptic function in AD (McDaid et al., 2020). Ca²⁺ homeostasis depends on the Sarcoplasmic/endoplasmic reticulum Ca²⁺ ATPase (SERCA) pumps to regulate Ca²⁺ uptake from the cytoplasm to the ER and activated inositol 1,4,5-triphosphate receptors (IP3Rs) and ryanodine receptors (RyRs) to enable Ca²⁺ efflux from the ER (Kawamoto et al., 2012). The IP3R-glucoseregulated protein 75 (Grp75)-voltage-dependent anion channel (VDAC) Ca²⁺ channeling complex was concentrated in MAMs. Up-regulation of this complex results in increased calcium content. The accumulation of mitochondrial calcium could be prohibited by the IP3R inhibitor (Chen et al., 2021b). The RyR is a Ca^{2+} release channel protein located in the MAMs and sarcoplasmic reticulum. The RyR3 plays a dual character in AD pathology. Deletion of RyR3 in young APP/PS1 mice increases the excitability of hippocampal neuronal networks and accelerates AD progress. However, in aged APP/PS1 mice, the deletion of RyR3 restored network excitability (Liu et al., 2014). In the 5xFAD mouse model of AD, restricting RyR2 open time blocked the excessive activity of CA1 neurons (Yao et al., 2020). A small-molecule SERCA activator can increase ER Ca²⁺ and has shown efficacy in APP/PS1 mice, supporting SERCA activation as a therapeutic strategy for AD (Krajnak and Dahl, 2018).

The role of MAMs in lipid metabolism of AD

Lipid metabolism, especially cholesterol metabolism, has been implicated in the synaptic dysfunction of AD (Petrov et al., 2017). Ceramide and cholesterol have been found to increase in the brains of AD patients, normal aging mice, and neurons exposed to $A\beta_{1-42}$ (Cutler et al., 2004). MAMs are transient lipid rafts that are closely related to cholesterol and phospholipid metabolism (Pera et al., 2020). It has been proven that phospholipid transport between the ER and mitochondria is dependent on membrane integrity including MAMs rather than energy or MMP (Kojima et al., 2016). MAMs are where the enzymatic activities such as acetyl-CoA acetyltransferase 1 (ACAT1) that regulate cholesterol levels reside. ACAT1 gene ablation led to the amelioration of amyloid pathology and cognitive deficits in 3 \times Tg AD mice (Bryleva et al., 2010). The concentration of unprocessed C99 at the MAMs is increased in models of AD and cells from AD patients, subsequently associated with the increase in ceramide levels (Pera et al., 2017). GSAP enriched in MAMs regulates lipid homeostasis through the processing of APP (Xu et al., 2021). In a cortical impact mice model of AD, the injured cortex and hippocampus exhibited significant increases in MAM activity, phospholipid synthesis, sphingomyelinase (SMases) activity, and cholesterol turnover (Agrawal et al., 2022). When cholesterol concentrations exceed a certain threshold, SMases are activated and hydrolyze sphingomyelin (SM) to produce ceramides. The cholesterol affinity of APP is involved in limiting APP distribution, conversely, APP senses and balances the membrane cholesterol (DelBove et al., 2019). An increase in MAM-localized C99 triggers the upregulation of SMase activity. The increased cholesterol mobilization observed in AD cells may be an outcome of persistent SMase activity caused by increased MAM-C99, which disrupts cellular lipid homeostasis and causes the alterations in membrane lipid composition commonly happen in AD (Montesinos et al., 2020).

The role of MAMs in abnormal autophagy in AD

Autophagy is a degradation mechanism of cells to remove damaged and senescent organelles and maintain cellular homeostasis. Autophagy plays a vital role in the generation and clearance of A β . Abnormal autophagy is involved in the development of AD (Yuan et al., 2022). Activation of autophagy alleviates pathological characters and cognitive deficits in APP/PS1 mice (Yang et al., 2023). The MAMs mark the starting point of autophagosome formation. Mitochondrial fusion protein Mfn2 is a MAM protein responsible for binding the ER and mitochondria. The energy sensor AMP-activated protein kinase (AMPK) interacts with Mfn2 and phosphorylates Mfn2 and induces autophagy (Hu et al., 2021). Autophagy initiation proteins such as autophagy and beclin 1 regulator 1 (AMBRA1) and Beclin 1 are recruited to the MAMs to regulate autophagy, demonstrating that MAM raft-like microdomains play a crucial part in the autophagosome formation (Garofalo et al., 2016). The sigma-1 receptor (Sig-1R) is a receptor with molecular chaperone activity clustered on MAMs. Autophagosome clearance is impaired in Sigma-1 KO cells, possibly due to impaired autophagosome-lysosomal organelle fusion mediated by the complex formed by Sig-1R with STX17 and ATG14. STX17 and ATG14 both appear at the ER-mitochondria contact site (Yang et al., 2019). The combination of vesicleassociated membrane protein-associated protein B (VAPB) and protein tyrosine phosphatase interacting protein 51 (PTPIP51) is identified as one of the MAM tethers. Manipulation of their expression to increase and decrease ER-mitochondrial junctions had significant effects on autophagy (Gomez-Suaga et al., 2017). The decreased expression of MAM tethering protein complexes including VAPB-PTPP51, BAP31-FIS1, and Mfn2-Mfn1 leads to abnormal autophagy, which leads to the decline of cognitive ability (Liu et al., 2022). The role of MAM tethering proteins in several major pathogeneses of AD is summarized in Figure 1 schematically.

Strategies targeting MAMs for treating AD

The functions of MAMs are mainly exerted by its numerous tethering proteins. The abnormalities of these proteins eventually lead to the pathological processes of AD through multiple mechanisms. The expression of MAM tethering proteins and the tight connection between mitochondria and ER can be one of the hallmarks of AD progression. The regulation of these proteins to improve the corresponding phenotypes can be used as a strategy and target for AD treatment.

Knockdown of MAM protein GSAP reduced Aß generation and plaque formation in an AD mouse model (He et al., 2010). Imatinib and imatinib methanesulfonate could prevent Aβ formation by inhibiting GSAP (He et al., 2010; Weintraub et al., 2013; Chu et al., 2014). Lithium treatment reduced abnormal IP3R-dependent ER Ca2+ signaling and enhanced synaptic plasticity in 3 \times Tg AD mice (Wiseman et al., 2023). Xestospongin C can ameliorate the Ca^{2+} overload of primary hippocampal neurons induced by $A\beta_{1-42}$ and improve the cognitive ability of APP/PS1 mice (Wang et al., 2019). Mecobalamin also reduces ER-mitochondrial calcium flux through IP3R and prevents mitochondrial dysfunction (Wang and Xu, 2019). RyR2 instability plays a vital role in the reduction of ER Ca²⁺ content, which alters synaptic transmission and plasticity mechanisms. While Dantrolene stabilizes RyR2 thereby reversing most AD-related phenotypes in AppNL-G-F mice (Nakamura et al., 2021). Compound 12a inhibited Ca^{2+} release and significantly accelerated the cognitive behavior of FAD mice in the Morris water maze test. Moreover, docking simulations testified that 12a could bind to the active site of RyR1 (Dai et al., 2021). A Sig-1R agonist (+) SKF-10,047 could significantly increase mitochondrial movement in cortical neurons of $3 \times Tg$ mice, which might be because it leads to the removal of Sig-1R from MAMs to mitochondria. (+) SKF-10,047 also leads to an increase in the number of mitochondria (Cavendish et al., 2019). Two other Sig-1R agonists ANAVEX2-73 and PRE-084 could prevent mitochondrial respiratory dysfunction in A β_{25-35} -injected mice (Lahmy et al., 2014). IRE1 α inhibitor 4µ8c can reduce ER-mitochondrial association and restore the normal function of MAMs by inhibiting the expression and interaction of IP3R, Grp75, and VDAC1, thereby restoring ATP content and MMP (Chu et al., 2021). Artesunate could reverse



cognitive impairment in APP/PS1 mice by regulating DRP1 to maintain mitochondrial dynamics (Qin et al., 2022). By increasing the expression of the MAM protein Mfn2, Biochanin A could reverse the imbalance of mitochondrial dynamics and abnormal mitophagy in APP/PS1 mice (Hou et al., 2022). Curcumin restores basal mitochondrial respiration and ATP production in thapsigargin-injured SH-SY5Y cells by targeting Mfn2 (Zhou et al., 2022). Besides, Myricetin, Selenomethionine, Leptin, Trans-ferulic acid, Ligustilide, and Albiflorin were also able to regulate Mfn2 and ameliorate mitochondrial dysfunction (Xu et al., 2018, 2019; Zafeer et al., 2019; Cheng et al., 2020; Zhu et al., 2020; Chen et al., 2021a; Yao et al., 2022). Cholesteryl ester produced in MAMs is involved in the pathogenesis of AD. Progesterone reduced the expression of ACAT1 through the ERK1/2 pathway, shortened the abnormally prolonged MAM length, inhibited cholesteryl ester accumulation in the cortex, and improved the cognitive function of APP/PS1 mice (Shi et al., 2021). We summarize potential compounds and drugs that exert anti-AD effects by regulating MAM tethering proteins in Table 1.

Discussion

Drug development for AD has continuously been challenged. The most popular AD hypotheses are amyloid cascade, hyperphosphorylation of tau, and mitochondrial cascade. However, numerous studies targeting these hypotheses have not been able to fully elucidate the mechanism of AD or retard its progression. The interaction between different organelles, especially mitochondria and ER in cells has gradually emerged in the study of various diseases (Theurey and Rieusset, 2017; Luan et al., 2021). Cellular dysfunction in the early stage of AD, including Ca^{2+} homeostasis, mitochondrial dysfunction, oxidative stress, and abnormal autophagy, are all associated with MAM function (Tapella et al., 2022).

As for the amyloid cascade, MAMs are the sites where C99 is cleaved to AB. Inhibition of MAMs protein expression and function may reduce AB production and extracellular deposition. MAMs recruit proteins such as INF2 and DRP1 to participate in mitochondrial fission, thereby affecting MMP and ATP production. Upregulation of some MAM-resident proteins causes enhanced mitochondria-ER contacts, along with mitochondrial damage such as increased mitochondrial superoxide. The MAM has both protein pumps that allow Ca²⁺ to flow from the cytoplasm to the ER and Ca²⁺ to flow from the ER, thus regulating Ca²⁺ homeostasis. Ca²⁺ flux also plays a dual role in AD progression. For instance, excessive Ca²⁺ influx leads to increased ROS production and cell death due to caspase activation; however, attenuated Ca²⁺ signaling may also be detrimental to ATP production (Filadi and Greotti, 2021; Arnst et al., 2022). Modulation of MAM-resident enzymes such as ACAT1 and MAM-C99 content affects cholesterol levels and lipid homeostasis in AD, along with amyloid synthesis and synaptic transmission. MAMs mark the starting point of autophagy. Decreased expressions of MAM tethering proteins, such as Sig-1R, VAPB-PTPP51, and Mfn2-Mfn1, lead to abnormal autophagy. All these indicate that MAM is an important target that should not be ignored in the study of AD pathogenesis.

Although MAMs have been shown to be closely related to the progression of AD, the molecular pathways are still not fully understood. At present, the research targets of AD drugs targeting MAM proteins are still limited. The mechanisms of interest are mainly in A β production, mitochondrial function, calcium

| Compound/drug | Target | Mechanism | Model | References |
|---------------------------|------------------------|--|--|-----------------------------------|
| Imatinib | GSAP | Aβ generation | $3 \times Tg$ mice N2a-APP695 cells | He et al., 2010; Chu et al., 2014 |
| Imatinib methanesulfonate | GSAP | Aβ generation | LPS-induced inflammation | Weintraub et al., 2013 |
| Lithium | IP3R | Ca ²⁺ signaling abnormalities | $3 \times Tg$ mice | Wiseman et al., 2023 |
| Xestospongin C | IP3R | Ca ²⁺ homeostasis | APP/PS1 mice, $A\beta_{1-42}$ -treated primary hippocampal neurons | Wang et al., 2019 |
| Methyl vitamin B12 | IP3R | Ca ²⁺ homeostasis, mitochondrial dysfunction | Aβ-treated PC12 cells | Wang and Xu, 2019 |
| Dantrolene | RyR2 | Ca ²⁺ homeostasis | App ^{NL-G-F} mice | Nakamura et al., 2021 |
| 12a | RyR1 | Ca ²⁺ homeostasis | FAD mice | Dai et al., 2021 |
| (+) SKF-10,047 | Sig-1R | Mitochondrial movement and number | Primary hippocampal neurons from 3 × Tg mice | Cavendish et al., 2019 |
| ANAVEX2-73, PRE-084 | Sig-1R | Mitochondrial respiratory dysfunction | $A\beta_{25-35}$ -injected mice | Lahmy et al., 2014 |
| 4µ8c | IP3R, Grp75, and VDAC1 | Mitochondrial dysfunction | Aβ-treated SH-SY5Y cells | Chu et al., 2021 |
| Artesunate | DRP1 | Mitochondrial dynamics | APP/PS1 mice | Qin et al., 2022 |
| Biochanin A | Mfn2 | Mitochondrial dynamics and mitophagy | APP/PS1 mice | Hou et al., 2022 |
| Curcumin | Mfn2 | Mitochondrial dysfunction | Thapsigargin-treated SH-SY5Y cells | Zhou et al., 2022 |
| Myricetin | Mfn2 | Mitochondrial dysfunction | N2a-APP695-Swedish cells | Yao et al., 2022 |
| Selenomethionine | Mfn2 | Mitochondrial dysfunction | N2a-APP695-Swedish cells, $3 \times Tg$ mice | Chen et al., 2021a |
| Leptin | Mfn2 | Mitochondrial dysfunction | Aβ1-42-treated SH-SY5Y cells | Cheng et al., 2020 |
| Trans-ferulic acid | Mfn2 | Mitochondrial dysfunction | Streptozocin injection | Zafeer et al., 2019 |
| Ligustilide | Mfn1, Mfn2 | Mitochondrial dysfunction | SAMP8 mice APP/PS1 mice | Xu et al., 2018; Zhu et al., 2020 |
| Albiflorin | Mfn1, Mfn2 | Mitochondrial dysfunction | APP/PS1 mice | Xu et al., 2019 |
| Progesterone | ACAT1 | Cholesterol metabolism | APP/PS1 mice | Shi et al., 2021 |

TABLE 1 Compounds and drugs targeting MAM tethering proteins for treating AD.

homeostasis, and lipid metabolism. Currently, there are few studies on the interaction between MAMs and tau phosphorylation. Besides, although tethering proteins are the main manifestation of MAM function, the observation of MAM structure should not be ignored despite the technical difficulties in the research of AD treatment strategies. We look forward to new studies to further explore the role of the structure and functions of MAMs in the pathogenesis and treatment of AD.

Author contributions

YY and HL conceived the idea. ZL wrote the manuscript. YC, HP, and LM revised the manuscript. All authors contributed to the work and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

Agrawal, R. R., Larrea, D., Xu, Y., Shi, L., Zirpoli, H., Cummins, L. G., et al. (2022). Alzheimer's-associated upregulation of mitochondria-associated ER membranes after traumatic brain injury. *Cell Mol. Neurobiol.* doi: 10.1007/s10571-022-01299-0 [Epub ahead of print].

Area-Gomez, E., and Schon, E. A. (2017). On the pathogenesis of Alzheimer's disease: the MAM hypothesis. FASEB J. 31, 864–867. doi: 10.1096/fj.201601309

Arnst, N., Redolfi, N., Lia, A., Bedetta, M., Greotti, E., and Pizzo, P. (2022). Mitochondrial Ca(2+) signaling and bioenergetics in Alzheimer's disease. *Biomedicines* 10:3025. doi: 10.3390/biomedicines10123025

Bhattacharyya, R., Black, S. E., Lotlikar, M. S., Fenn, R. H., Jorfi, M., Kovacs, D. M., et al. (2021). Axonal generation of amyloid-β from palmitoylated APP in mitochondria-associated endoplasmic reticulum membranes. *Cell Rep.* 35:109134. doi: 10.1016/j.celrep.2021.109134

Bryleva, E. Y., Rogers, M. A., Chang, C. C., Buen, F., Harris, B. T., Rousselet, E., et al. (2010). ACAT1 gene ablation increases 24(S)-hydroxycholesterol content in the brain and ameliorates amyloid pathology in mice with AD. *Proc. Natl. Acad. Sci. U. S. A.* 107, 3081–3086. doi: 10.1073/pnas.0913828107

Cavendish, J. Z., Sarkar, S. N., Colantonio, M. A., Quintana, D. D., Ahmed, N., White, B. A., et al. (2019). Mitochondrial movement and number deficits in embryonic cortical neurons from 3xTg-AD mice. *J. Alzheimers Dis.* 70, 139–151. doi: 10.3233/jad-190143

Chen, C., Chen, Y., Zhang, Z. H., Jia, S. Z., Chen, Y. B., Huang, S. L., et al. (2021a). Selenomethionine improves mitochondrial function by upregulating mitochondrial selenoprotein in a model of Alzheimer's disease. *Front. Aging Neurosci.* 13:750921. doi: 10.3389/fnagi.2021.750921

Chen, L. T., Xu, T. T., Qiu, Y. Q., Liu, N. Y., Ke, X. Y., Fang, L., et al. (2021b). Homocysteine induced a calcium-mediated disruption of mitochondrial function and dynamics in endothelial cells. *J. Biochem. Mol. Toxicol.* 35:e22737. doi: 10.1002/jbt. 22737

Cheng, Y., Buchan, M., Vitanova, K., Aitken, L., Gunn-Moore, F. J., Ramsay, R. R., et al. (2020). Neuroprotective actions of leptin facilitated through balancing mitochondrial morphology and improving mitochondrial function. *J. Neurochem.* 155, 191–206. doi: 10.1111/jnc.15003

Chu, B., Li, M., Cao, X., Li, R., Jin, S., Yang, H., et al. (2021). IRE1 α -XBP1 affects the mitochondrial function of A β 25-35-treated SH-SY5Y cells by regulating mitochondria-associated endoplasmic reticulum membranes. *Front. Cell Neurosci.* 15:614556. doi: 10.3389/fncel.2021.614556

Chu, J., Lauretti, E., Craige, C. P., and Praticò, D. (2014). Pharmacological modulation of GSAP reduces amyloid- β levels and tau phosphorylation in a mouse model of Alzheimer's disease with plaques and tangles. *J. Alzheimers Dis.* 41, 729–737. doi: 10.3233/jad-140105

Cutler, R. G., Kelly, J., Storie, K., Pedersen, W. A., Tammara, A., Hatanpaa, K., et al. (2004). Involvement of oxidative stress-induced abnormalities in ceramide and cholesterol metabolism in brain aging and Alzheimer's disease. *Proc. Natl. Acad. Sci. U.S.A.* 101, 2070–2075. doi: 10.1073/pnas.0305799101

Dai, B., Ma, X., Tang, Y., Xu, L., Guo, S., Chen, X., et al. (2021). Design, synthesis, and biological activity of novel semicarbazones as potent Ryanodine receptor1 inhibitors of Alzheimer's disease. *Bioorg. Med. Chem.* 29:115891. doi: 10.1016/j.bmc.2020.115891

Degechisa, S. T., Dabi, Y. T., and Gizaw, S. T. (2022). The mitochondrial associated endoplasmic reticulum membranes: a platform for the pathogenesis of inflammation-mediated metabolic diseases. *Immun. Inflamm. Dis.* 10:e647. doi: 10.1002/iid3.647

Del Prete, D., Suski, J. M., Oulès, B., Debayle, D., Gay, A. S., Lacas-Gervais, S., et al. (2017). Localization and processing of the amyloid- β protein precursor in mitochondria-associated membranes. *J. Alzheimers Dis.* 55, 1549–1570. doi: 10.3233/jad-160953

DelBove, C. E., Strothman, C. E., Lazarenko, R. M., Huang, H., Sanders, C. R., and Zhang, Q. (2019). Reciprocal modulation between amyloid precursor protein and synaptic membrane cholesterol revealed by live cell imaging. *Neurobiol. Dis.* 127, 449–461. doi: 10.1016/j.nbd.2019.03.009

Fernandes, T., Resende, R., Silva, D. F., Marques, A. P., Santos, A. E., Cardoso, S. M., et al. (2021). Structural and functional alterations in mitochondria-associated membranes (MAMs) and in mitochondria activate stress response mechanisms in an *in vitro* model of Alzheimer's disease. *Biomedicines* 9:881. doi: 10.3390/biomedicines9080881

Filadi, R., and Greotti, E. (2021). The yin and yang of mitochondrial Ca(2+) signaling in cell physiology and pathology. *Cell Calcium* 93:102321. doi: 10.1016/j.ceca. 2020.102321

Garofalo, T., Matarrese, P., Manganelli, V., Marconi, M., Tinari, A., Gambardella, L., et al. (2016). Evidence for the involvement of lipid rafts localized at the ERmitochondria associated membranes in autophagosome formation. *Autophagy* 12, 917–935. doi: 10.1080/15548627.2016.1160971

Gomez-Suaga, P., Paillusson, S., Stoica, R., Noble, W., Hanger, D. P., and Miller, C. C. J. (2017). The ER-mitochondria tethering complex VAPB-PTPIP51 regulates autophagy. *Curr. Biol.* 27, 371–385. doi: 10.1016/j.cub.2016.12.038

Gowrishankar, S., Wu, Y., and Ferguson, S. M. (2017). Impaired JIP3-dependent axonal lysosome transport promotes amyloid plaque pathology. *J. Cell Biol.* 216, 3291–3305. doi: 10.1083/jcb.201612148

Han, J., Park, H., Maharana, C., Gwon, A. R., Park, J., Baek, S. H., et al. (2021). Alzheimer's disease-causing presenilin-1 mutations have deleterious effects on mitochondrial function. *Theranostics* 11, 8855–8873. doi: 10.7150/thno.59776

He, G., Luo, W., Li, P., Remmers, C., Netzer, W. J., Hendrick, J., et al. (2010). Gamma-secretase activating protein is a therapeutic target for Alzheimer's disease. *Nature* 467, 95–98. doi: 10.1038/nature09325

Hou, Y., Zhao, W., Yu, H., Zhang, F., Zhang, H. T., and Zhou, Y. (2022). Biochanin A alleviates cognitive impairment and hippocampal mitochondrial damage in ovariectomized APP/PS1 mice. *Phytomedicine* 100:154056. doi: 10.1016/j.phymed. 2022.154056

Hu, Y., Chen, H., Zhang, L., Lin, X., Li, X., Zhuang, H., et al. (2021). The AMPK-MFN2 axis regulates MAM dynamics and autophagy induced by energy stresses. *Autophagy* 17, 1142–1156. doi: 10.1080/15548627.2020.1749490

Kawamoto, E. M., Vivar, C., and Camandola, S. (2012). Physiology and pathology of calcium signaling in the brain. *Front. Pharmacol.* 3:61. doi: 10.3389/fphar.2012.00061

Kojima, R., Endo, T., and Tamura, Y. (2016). A phospholipid transfer function of ER-mitochondria encounter structure revealed *in vitro*. *Sci. Rep.* 6:30777. doi: 10.1038/ srep30777

Krajnak, K., and Dahl, R. (2018). A new target for Alzheimer's disease: a small molecule SERCA activator is neuroprotective *in vitro* and improves memory and cognition in APP/PS1 mice. *Bioorg. Med. Chem. Lett.* 28, 1591–1594. doi: 10.1016/j. bmcl.2018.03.052

Lahmy, V., Long, R., Morin, D., Villard, V., and Maurice, T. (2014). Mitochondrial protection by the mixed muscarinic/ σ 1 ligand ANAVEX2-73, a tetrahydrofuran derivative, in A β 25-35 peptide-injected mice, a nontransgenic Alzheimer's disease model. *Front. Cell Neurosci.* 8:463. doi: 10.3389/fncel.2014.00463

Leal, N. S., Dentoni, G., Schreiner, B., Naia, L., Piras, A., Graff, C., et al. (2020). Amyloid B-peptide increases mitochondria-endoplasmic reticulum contact altering mitochondrial function and autophagosome formation in Alzheimer's disease-related models. *Cells* 9:2552. doi: 10.3390/cells9122552

Leal, N. S., Schreiner, B., Pinho, C. M., Filadi, R., Wiehager, B., Karlström, H., et al. (2016). Mitofusin-2 knockdown increases ER-mitochondria contact and decreases amyloid β -peptide production. *J. Cell Mol. Med.* 20, 1686–1695. doi: 10.1111/jcmm. 12863

Liu, J., Supnet, C., Sun, S., Zhang, H., Good, L., Popugaeva, E., et al. (2014). The role of ryanodine receptor type 3 in a mouse model of Alzheimer disease. *Channels* 8, 230–242. doi: 10.4161/chan.27471

Liu, J., Wang, L., Ge, L., Sun, W., Song, Z., Lu, X., et al. (2022). Lanthanum decreased VAPB-PTPP51, BAP31-FIS1, and MFN2-MFN1 expression of mitochondriaassociated membranes and induced abnormal autophagy in rat hippocampus. *Food Chem. Toxicol.* 161:112831. doi: 10.1016/j.fct.2022.112831

Luan, Y., Luan, Y., Yuan, R. X., Feng, Q., Chen, X., and Yang, Y. (2021). Structure and function of mitochondria-associated endoplasmic reticulum membranes (MAMs) and their role in cardiovascular diseases. *Oxid. Med. Cell Longev.* 2021:4578809. doi: 10.1155/2021/4578809

McDaid, J., Mustaly-Kalimi, S., and Stutzmann, G. E. (2020). Ca(2+) dyshomeostasis disrupts neuronal and synaptic function in Alzheimer's disease. *Cells* 9:2655. doi: 10.3390/cells9122655

Montesinos, J., Pera, M., Larrea, D., Guardia-Laguarta, C., Agrawal, R. R., Velasco, K. R., et al. (2020). The Alzheimer's disease-associated C99 fragment of APP regulates cellular cholesterol trafficking. *Embo J.* 39:e103791. doi: 10.15252/embj.2019103791

Nakamura, Y., Yamamoto, T., Xu, X., Kobayashi, S., Tanaka, S., Tamitani, M., et al. (2021). Enhancing calmodulin binding to ryanodine receptor is crucial to limit neuronal cell loss in Alzheimer disease. *Sci. Rep.* 11:7289. doi: 10.1038/s41598-021-86822-x

Niederst, E. D., Reyna, S. M., and Goldstein, L. S. (2015). Axonal amyloid precursor protein and its fragments undergo somatodendritic endocytosis and processing. *Mol. Biol. Cell* 26, 205–217. doi: 10.1091/mbc.E14-06-1049

Pera, M., Larrea, D., Guardia-Laguarta, C., Montesinos, J., Velasco, K. R., Agrawal, R. R., et al. (2017). Increased localization of APP-C99 in mitochondria-associated ER membranes causes mitochondrial dysfunction in Alzheimer disease. *Embo J.* 36, 3356–3371. doi: 10.15252/embj.201796797

Pera, M., Montesinos, J., Larrea, D., Agrawal, R. R., Velasco, K. R., Stavrovskaya, I. G., et al. (2020). MAM and C99, key players in the pathogenesis of Alzheimer's disease. *Int. Rev. Neurobiol.* 154, 235–278. doi: 10.1016/bs.irn.2020.03.016

Petrov, A. M., Kasimov, M. R., and Zefirov, A. L. (2017). Cholesterol in the pathogenesis of Alzheimer's, Parkinson's diseases and autism: link to synaptic dysfunction. *Acta Naturae* 9, 26–37.

Qin, Y. R., Ma, C. Q., Jiang, J. H., Wang, D. P., Zhang, Q. Q., Liu, M. R., et al. (2022). Artesunate restores mitochondrial fusion-fission dynamics and alleviates neuronal injury in Alzheimer's disease models. J. Neurochem. 162, 290–304. doi: 10.1111/jnc. 15620

Rowland, A. A., and Voeltz, G. K. (2012). Endoplasmic reticulum-mitochondria contacts: function of the junction. *Nat. Rev. Mol. Cell Biol.* 13, 607–625. doi: 10.1038/nrm3440

Schreiner, B., Hedskog, L., Wiehager, B., and Ankarcrona, M. (2015). Amyloid- β peptides are generated in mitochondria-associated endoplasmic reticulum membranes. *J. Alzheimers Dis.* 43, 369–374. doi: 10.3233/jad-132543

Shi, W., Wu, H., Liu, S., Wu, Z., Wu, H., Liu, J., et al. (2021). Progesterone suppresses cholesterol esterification in APP/PS1 mice and a cell model of Alzheimer's disease. *Brain Res. Bull* 173, 162–173. doi: 10.1016/j.brainresbull.2021.05.020

Skobeleva, K., Shalygin, A., Mikhaylova, E., Guzhova, I., Ryazantseva, M., and Kaznacheyeva, E. (2022). The STIM1/2-regulated calcium homeostasis is impaired in hippocampal neurons of the 5xFAD mouse model of Alzheimer's disease. *Int. J. Mol. Sci.* 23:14810. doi: 10.3390/ijms232314810

Steffen, J., and Koehler, C. M. (2018). ER-mitochondria contacts: actin dynamics at the ER control mitochondrial fission via calcium release. *J. Cell Biol.* 217, 15–17. doi: 10.1083/jcb.201711075

Tapella, L., Dematteis, G., Moro, M., Pistolato, B., Tonelli, E., Vanella, V. V., et al. (2022). Protein synthesis inhibition and loss of homeostatic functions in astrocytes from an Alzheimer's disease mouse model: a role for ER-mitochondria interaction. *Cell Death Dis.* 13:878. doi: 10.1038/s41419-022-05324-4

Theurey, P., and Rieusset, J. (2017). Mitochondria-associated membranes response to nutrient availability and role in metabolic diseases. *Trends Endocrinol. Metab.* 28, 32–45. doi: 10.1016/j.tem.2016.09.002

Wang, M., and Xu, T. (2019). Methyl B12 protects PC12 cells against cytotoxicity induced by A β (25-35). J. Cell Biochem. 120, 11921–11930. doi: 10.1002/jcb.28475

Wang, Z. J., Zhao, F., Wang, C. F., Zhang, X. M., Xiao, Y., Zhou, F., et al. (2019). Xestospongin C, a reversible IP3 receptor antagonist, alleviates the cognitive and pathological impairments in APP/PS1 mice of Alzheimer's disease. *J. Alzheimers Dis.* 72, 1217–1231. doi: 10.3233/jad-190796

Weintraub, M. K., Bisson, C. M., Nouri, J. N., Vinson, B. T., Eimerbrink, M. J., Kranjac, D., et al. (2013). Imatinib methanesulfonate reduces hippocampal amyloid-β and restores cognitive function following repeated endotoxin exposure. *Brain Behav. Immun.* 33, 24–28. doi: 10.1016/j.bbi.2013.05.002

Wiseman, A. L., Briggs, C. A., Peritt, A., Kapecki, N., Peterson, D. A., Shim, S. S., et al. (2023). Lithium provides broad therapeutic benefits in an alzheimer's disease mouse model. *J. Alzheimers Dis.* 91, 273–290. doi: 10.3233/jad-220758

Wu, W., Li, W., Chen, H., Jiang, L., Zhu, R., and Feng, D. (2016). FUNDC1 is a novel mitochondrial-associated-membrane (MAM) protein required for hypoxiainduced mitochondrial fission and mitophagy. *Autophagy* 12, 1675–1676. doi: 10.1080/ 15548627.2016.1193656 Xu, P., Chang, J. C., Zhou, X., Wang, W., Bamkole, M., Wong, E., et al. (2021). GSAP regulates lipid homeostasis and mitochondrial function associated with Alzheimer's disease. *J. Exp. Med.* 218:e20202446. doi: 10.1084/jem.202 02446

Xu, Y. J., Mei, Y., Qu, Z. L., Zhang, S. J., Zhao, W., Fang, J. S., et al. (2018). Ligustilide ameliorates memory deficiency in APP/PS1 transgenic mice via restoring mitochondrial dysfunction. *Biomed. Res. Int.* 2018:4606752. doi: 10.1155/2018/ 4606752

Xu, Y. J., Mei, Y., Shi, X. Q., Zhang, Y. F., Wang, X. Y., Guan, L., et al. (2019). Albiforin ameliorates memory deficits in APP/PS1 transgenic mice via ameliorating mitochondrial dysfunction. *Brain Res.* 1719, 113–123. doi: 10.1016/j.brainres.2019. 05.037

Yang, C. Z., Wang, S. H., Zhang, R. H., Lin, J. H., Tian, Y. H., Yang, Y. Q., et al. (2023). Neuroprotective effect of astragalin via activating PI3K/Akt-mTOR-mediated autophagy on APP/PS1 mice. *Cell Death Discov*. 9:15. doi: 10.1038/s41420-023-0 1324-1

Yang, H., Shen, H., Li, J., and Guo, L. W. (2019). SIGMAR1/Sigma-1 receptor ablation impairs autophagosome clearance. *Autophagy* 15, 1539–1557. doi: 10.1080/15548627.2019.1586248

Yao, J., Sun, B., Institoris, A., Zhan, X., Guo, W., Song, Z., et al. (2020). Limiting RyR2 open time prevents Alzheimer's disease-related neuronal hyperactivity and memory loss but not β -amyloid accumulation. *Cell Rep.* 32:108169. doi: 10.1016/j. celrep.2020.108169

Yao, X., Zhang, J., Lu, Y., Deng, Y., Zhao, R., and Xiao, S. (2022). Myricetin restores $A\beta$ -induced mitochondrial impairments in N2a-SW cells. *ACS Chem. Neurosci.* 13, 454–463. doi: 10.1021/acschemneuro.1c00591

Yuan, M., Wang, Y., Huang, Z., Jing, F., Qiao, P., Zou, Q., et al. (2022). Impaired autophagy in amyloid-beta pathology: a traditional review of recent Alzheimer's research. *J. Biomed. Res.* 37, 30–46. doi: 10.7555/jbr.36.20 220145

Zafeer, M. F., Firdaus, F., Anis, E., and Mobarak Hossain, M. (2019). Prolong treatment with Trans-ferulic acid mitigates bioenergetics loss and restores mitochondrial dynamics in streptozotocin-induced sporadic dementia of Alzheimer's type. *Neurotoxicology* 73, 246–257. doi: 10.1016/j.neuro.2019. 04.006

Zhou, H. Y., Sun, Y. Y., Chang, P., and Huang, H. C. (2022). Curcumin inhibits cell damage and apoptosis caused by thapsigargin-induced endoplasmic reticulum stress involving the recovery of mitochondrial function mediated by mitofusin-2. *Neurotox Res.* 40, 449–460. doi: 10.1007/s12640-022-00481-y

Zhu, W. L., Zheng, J. Y., Cai, W. W., Dai, Z., Li, B. Y., Xu, T. T., et al. (2020). Ligustilide improves aging-induced memory deficit by regulating mitochondrial related inflammation in SAMP8 mice. *Aging* 12, 3175–3189. doi: 10.18632/aging. 102793