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SPECIALTY SECTION This article was submitted to Radiopharmacy and Radiochemistry, a section of the journal Frontiers in Nuclear Medicine

RECEIVED 07 June 2022 ACCEPTED 21 July 2022 PUBLISHED 16 August 2022

#### CITATION

Takalloobanafshi G, Kukreja A and Hicks JW (2022) Historical efforts to develop <sup>99m</sup>Tc-based amyloid plaque targeting radiotracers. *Front. Nucl. Med.* 2:963698. doi: 10.3389/fnume.2022.963698

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# Historical efforts to develop <sup>99m</sup>Tc-based amyloid plaque targeting radiotracers

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Imaging biomarkers have changed the way we study Alzheimer's disease and related dementias, develop new therapeutics to treat the disease, and stratify patient populations in clinical trials. With respect to protein aggregates comprised of amyloid- $\beta$  plagues and tau neurofibrillary tangles, Positron Emission Tomography (PET) has become the gold standard imaging modality for quantitative visualization. Due to high infrastructural costs, the availability of PET remains limited to large urban areas within high income nations. This limits access to leading edge medical imaging, and potentially access to new treatments, by millions of rural and remote residents in those regions as well as billions of people in middle- and low-income countries. Single Photon Emission Computed Tomography (SPECT) is a more widely available imaging alternative with lower infrastructural costs and decades of familiarity amongst nuclear medicine professionals. Recent technological advances have closed the gap in spatial resolution and quantitation between SPECT and PET. If effective SPECT radiotracers were available to visualize amyloid- $\beta$  plaques, geographic barriers to imaging could be circumvented. In this review, we will discuss past efforts to develop SPECT radiotracers targeting amyloid- $\!\beta$ plaques which incorporate the most used radionuclide in nuclear medicine: technetium-99m ( $^{99m}$ Tc;  $t_{1/2}$  = 6.01 h;  $\gamma$  = 140 keV). While reviewing the various chemical scaffolds and chelates employed, the focus will be upon the impact to the pharmacological properties of putative <sup>99m</sup>Tc-based amyloid-targeting radiotracers.

KEYWORDS

technetium, Alzheimer's disease, radiotracer development, rural and remote access, single photon emission computed tomography (SPECT)

# Introduction

Alzheimer's disease (AD) and related dementias have a significant socioeconomic impact on patients, their families, and communities. If the economic burden of AD were a country, it would be the 18th largest in the world, estimated to surpass two trillion US dollars this decade and affects nearly 55 million people worldwide (1, 2). This number is expected to double every 20 years with a new case diagnosed approximately every 3 s (3). Much of this growth will be in low- and middle-income counties where access

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to diagnoses and therapies may be hindered. Even in high income countries, barriers remain to rural and remote communities to accessing the best care. This is particularly dire as diagnosis and therapies transition from symptomatic (sitting with physician) to measuring biological and genetic biomarkers (fluid and imaging).

As knowledge of the underlying biological processes driving AD grows, researchers have been able to identify fluid and imaging biomarkers to detect changes in the brain before symptoms begin to manifest with a research framework from the National Institute on Aging and Alzheimer's Association (NIA-AA) recently redefining AD to include biomarkers (4). Although not intended for general clinical practice, the research framework does provide a strong scaffold for such guidance. Significant focus is placed upon the protein aggregates of amyloid beta (A $\beta$ ) as extracellular plaques, tau as neurofibrillary tangles, and neuronal injury such as metabolic or perfusion changes (so-called A/T/N classification) (5). Recent promising results with plasma biomarkers, particularly pTau217, may make large impacts on clinical diagnosis and disease staging (6, 7), however, they are limited to systemic and not direct measurements within the brain. Imaging has a powerful role to play in visualizing and quantifying these biomarkers non-invasively through application of positron emission tomography (PET) and single photon emission computed tomography (SPECT).

Recent advances in nuclear medicine imaging of AD and kindred dementias have taken center stage within the NIA-AA research framework. Several PET radiopharmaceuticals labeled with <sup>11</sup>C and <sup>18</sup>F ( $t_{1/2} = 20.4$  and 109.8 min) have been approved by regulatory bodies and licensed for clinical use (8). One such example is  $[^{18}F]$ florbetapir (9), which was recently used to demonstrate reduced amyloid burden following treatment with aducanumab (Aduhelm; the first approved anti-amyloid drug) (10). These promising results could change clinical management of AD patients; however, in order to receive this therapy, patient need to first have a PET scan to assess amyloid burden. High infrastructure and operating costs have limited availability of PET to large population centers. This in turn has hindered the broad implementation of PET beyond a research setting. Consequently, rural, remote, and developing populations without available PET biomarker imaging have difficulty accessing improved AD therapies.

The other nuclear medicine imaging technology, SPECT, has been widely available since the early 1990s. As of 2015 in Canada, SPECT scanners outnumber PET scanners by at least 40:1 (11, 12). In a vast, sparsely populated country, access to either of these highly sensitive modalities could improve patient management, reduce burden on patients and caregivers, and improve therapy outcomes (13). More broadly speaking, the International Atomic Energy Agency established the IMAGINE database in 2012 to illustrate the availability of medical imaging

and nuclear medicine worldwide (14). A similar trend to Canada is seen worldwide, with Figure 1 showing higher accessibility to SPECT compared to PET. Much of this increased access can be attributed to lower infrastructure costs (15). While a PET scanner costs circa \$2 million, SPECT systems can be purchased for around \$600,000. As discussed in more detail below, neither of these systems can operate without radiotracers. SPECT also has an advantage here with typically longer half-life (i.e., longer use per production and greater distribution range) and lower radionuclide production costs compared to cyclotron production required for PET. More information on access and availability of radiopharmaceuticals can be found in a recent review of the topic (16).

In neurology application of nuclear medicine, PET has dominated due to the plethora of small molecule radiotracers labeled with <sup>11</sup>C and <sup>18</sup>F which target proteins of interest in psychiatric and neurological disorders (17). Furthermore, it was assumed the spatial resolution and relative ease for quantification with PET would ensure its dominance over SPECT (18). In recent reviews focused on advancements to SPECT technology, brain SPECT received barely a mention (19, 20). Another perspective paper focused on the future direction of molecular imaging failed to even mention (21) or barely mentioned (22) SPECT as a modality. Many technological advances were made over the past 10 years, particularly cadmium zinc telleride detectors (23), novel collimator designs (24), improved reconstruction (25), and flexible ring gantries (26). These have drastically closed the gap in spatial resolution (2.5 mm on clinical or submillimeter on preclinical scanners) (23), quantitation ( $\pm 5\%$  of true radionuclide concentration) (27), and sensitivity [up to 850 count/s/MBq for CZTbased SPECT (28)]. These improvements mean SPECT is now matching or surpassing PET with respect to scanner performance. Even with older SPECT cameras, the modality has been shown to be more cost effective in some cases compared to PET (e.g., the SPARC trial) (29). It is time to reconsider assumptions regarding SPECT to improve access to molecular neuroimaging. To fully exploit these technological improvements, advancements are required in radio-pharmacy and -chemistry.

The two primary radionuclides used for neurological SPECT are iodine-123 ( $^{123}$ I;  $t_{1/2} = 13.2$ h) and technetium-99m ( $^{99m}$ Tc;  $t_{1/2} = 6.01$  h). Both have been used clinically to study neurodegenerative diseases. Hexamethyl propylene oxime (HMPAO) coordinated to  $^{99m}$ Tc has been used in brain perfusion imaging in AD for decades (30). These SPECT perfusion studies found changes in brain regions associated with neurodegeneration similar to [ $^{18}$ F]FDG-PET hypometabolism. Importantly, these areas of hypoperfusion were not impacted by blood glucose levels. Given co-morbidities between diabetes and AD, perfusion SPECT may be a more reliable biomarker in these patients (31). As for targeted radiotracers, tropane-based probes for dopamine transporters have had the most clinical success.



Although not used for imaging AD, [<sup>123</sup>I]FP-CIT (Datscan) is used frequently to image dopaminergic tone in Parkinson's disease (32). As <sup>123</sup>I is not readily available in low- and middle-income countries (33), an alternative tropane for dopamine transporters, [<sup>99m</sup>Tc]Trodat, represents the only clinically approved, targeted <sup>99m</sup>Tc-based radiotracer (34–36). The success of [<sup>99m</sup>Tc]Trodat holds promise that a <sup>99m</sup>Tc-based amyloid radiotracer is achievable.

The focus of this primer review will be on developments over the past two decades toward producing an effective <sup>99m</sup>Tcbased radiotracer targeting Aß aggregates. This topic has been previously reviewed as part of broader, metal-based probe discussions (37-40), or focused on Tc alone (41). Our goal is to update the field and provide the most comprehensive collection of in vitro and ex vivo data to date. Discussion will focus upon in vitro binding affinity and selectivity, bloodbrain barrier (BBB) penetration, and pharmacokinetic data in wild-type and transgenic animal models. Although no radiotracer has emerged for human use, it remains worthwhile to pursue a simple, inexpensive, and highly accessible imaging method for the non-invasive quantification of proteinopathies in neurodegeneration. Such a tool could have major repercussions for the understanding of the pathogenesis of AD and also for its diagnosis and treatment (42, 43).

# Radiotracers of A $\beta$ plaques based on <sup>99m</sup>Tc

Given the wider availability of SPECT technology, it is past time for expanding the library of readily available probes for neuroimaging. There are several examples of <sup>123</sup>I being successfully employed for neuro-SPECT (44) but logistical and chemical challenges remain (45, 46). As the most used radionuclide in nuclear medicine, radiopharmacies are familiar with the chelation chemistry and regularly handle <sup>99m</sup>Tc (47). The wide availability from a <sup>99</sup>Mo/<sup>99m</sup>Tc generator and ideal imaging properties for SPECT make <sup>99m</sup>Tc-based targeted probes desirable.

Getting a metal-based radiotracer through the blood-brain barrier is a non-trivial task. The chelates used to attach the metal to the targeting vector (i.e., antibody, peptide, small molecule, etc.) often have a large impact on in vivo properties. Choosing a targeting vector-chelate pair with appropriate lipophilicity, overall size, and polarity mitigates these impacts. The addition of the chelate should also have a minimal impact upon the affinity and selectivity of the radiotracer to maintain the biological properties of the targeting vector. Any putative Tc-based radiotracers must demonstrate an ability to differentiate AB binding in AD patients from age-matched controls. Off-target binding, tissue clearance, metabolism, and pharmacokinetics matching the physical and biological half-lives of the radiotracer also play a crucial role in obtaining an effective SPECT imaging agent. Additionally, a practical production method is required for either point of care formulation or large-scale production and distribution. These ideal properties have been reviewed extensively elsewhere and readers are referred to these for more information (44, 46, 48).

#### Earliest attempts

Prior to 2000, only three attempts to image A $\beta$  plaques with  $^{99m}$ Tc were conducted (Figure 2). It was believed that conjugating a  $^{99m}$ Tc chelate to a common histological dye would make a moderate radiotracer to image amyloid plaques. By



substituting pyridinyl rings for phenyl groups in the central portion of the dyes Congo Red in compound **1** and Chrysamine G in compound **2**, rhenium or technetium could be chelated (49). Previous studies showed this would have minimal impact on binding affinity, since binding is strongly driven by flat, extended pi systems fitting between A $\beta$  sheets (50). Han et al. labeled these modified dyes with <sup>99</sup>Tc ( $t_{1/2} = 2.1 \times 10^5$  years;  $\beta^-$ ) in 28 and 29% yields, respectively. Their binding affinities for A $\beta_{1-40}$  were determined to be 630 and 160 nM for **1** and **2**. Since both chelates were charged, it was assumed there was limited utility for brain imaging without assistance opening the BBB.

Congo Red and a derivative dye were later modified with a monoamine monoamide bis thiol (MAMA') chelates (3 and 4) (42). These chelates were conjugated with Re for *in vitro* testing and 3 was also labeled with ground state <sup>99</sup>Tc in 88% yield. The binding affinities for the isosteric Re congeners were comparable to Congo Red (K<sub>d</sub> = 1,500  $\pm$  0.005 nM) in the authors' assay.

When administered to brain tissue slices, amyloid in both blood vessels and plaques were visualized with both compounds, albeit with **Re-4** in 2.5 higher concentration. Additionally, the charged nature of **3**, large size (MW > 1,000 Da) and hydrophilicity make BBB penetration unlikely.

In a final twentieth century report, Chrysamine G was conjugated to <sup>99m</sup>Tc through a MAMA chelator by the Leuven group (51). With the help of microwave heating, <sup>99m</sup>Tc-5 was obtained in 75% yield. They were able to obtain promising *in vitro* properties (log P = 1.08, >4 h stability) along with some selectivity for A $\beta$  in autoradiography experiments with brain tissue slices from AD patients. When compared to <sup>125</sup>I-labeled human serum albumin, there was slightly higher brain:blood ratios (indicative of BBB permeability), however, biodistribution studies in healthy mice (male NMRI mice, no age given) were not promising. There was very low initial brain uptake (0.3  $\pm$  0.1 %ID/g at 2 min post-injection or p.i.) with no activity observed at 60 min p.i. The rapid clearance from blood and



increasing activity in the intestinal tract could be an indication of *in vivo* metabolism.

There was little chance for success penetrating the BBB using these large and/or charged dye molecules. When it comes to brain radiotracers, BBB permeability is considered a crucial factor. In addition to affinity and selectivity for Aß aggregates, candidate SPECT radiotracers must also optimize lipohilicity, polar surface area, size, and proton donor/acceptor pairs to pass through the BBB. Several types of molecular transport mechanisms have been identified at the BBB (52). It's commonly accepted that non-polar, lipidsoluble molecules can passively diffuse through the cell membranes of endothelial cells across the BBB and enter the brain. Second, receptor-mediated transport mechanism, allowing polar nutrients (i.e., carbohydrates, metal ions, and amino acids) and molecular carriers (such as transferrin) to enter the brain. Conversely, efflux transporters work to remove exogenous molecules from the brain. Testing for efflux transporter affinity can rule out this mechanism for poor brain uptake.

Following the early success of PET radiotracers based on smaller, neutral dyes such as thioflavin S, benzofurans, hydrazines, etc., <sup>99m</sup>Tc-based probes began following a similar trend. The following sections are divided based upon chelating class—conjugated or integrated. Each of these is broken down further by targeting vector into various groups: (1) benzo-furan and (2) benzo-thiazoles (3) stilbenes, (4) chalcones, and (5) miscellaneous. Several additional compounds using bivalent ligand structures are also included.

# **Conjugated probes**

There are two main chelator classifications which group the putative SPECT radiotracers. The first is based on the classical tethered <sup>99m</sup>Tc-chelate with the targeting vector and metal complex separated by a linker. It is thought that by separating the bulky metal-ligand complex, the affinity to target will be unaffected by the added volume (53). As consequence of adding carbon or ethoxy chains, the mass and lipophilicity are altered. Although most examples below maintained adequate affinity for A $\beta$  aggregates *in vitro*, the added mass and often higher lipophilicity values may have negatively contributed to poor biophysical attributes.

## **Benzothiazoles**

#### For cerebral Aß detection

With the surge of PET radiotracer development culminating in the success of [<sup>11</sup>C]Pittsburgh Compound B (PiB), there was seemly a hiatus from <sup>99m</sup>Tc-based radiotracer development from 2000 to 2006. In 2007, the Leuven group reported the first <sup>99m</sup>Tc benzothiazole candidate (Figure 3; Table 1) (54). They chelated <sup>99m</sup>Tc *via* a bis-amino-bis-thiol (BAT) conjugated to the aniline of a thioflavin-T derivative in a mean yield of 86%. By adding carrier <sup>99</sup>Tc, the group was able to obtain mass spectroscopic data confirming the presence of **6** (m/z = 559.2), confirming the complexation of  $[Tc(V)O]^{3+}$ . The affinity for A $\beta$  plaques was assessed qualitatively using postmortem brain slices with and without the presence of thioflavin-T (1  $\mu$ M). Any

							% Injected dose/gram (%ID/g)			
Compound		n	R	Х	<i>K</i> <sub>1</sub> ( <b>n</b> M)	Log P	2 min	60 min	Ratio <sup>a</sup>	
6					n.d.	$1.92\pm0.012$	0.09	0.03	3.00	
7					n.d.	$2.90\pm0.13$	$1.34\pm0.16$	$0.65\pm0.10$	2.01	
8					10.0	3.26		n.d.		
9	a	3	Н		$143\pm26$	$3.51\pm0.014$	$0.50\pm0.10$	$0.18\pm0.07$	2.78	
	b	5	Н		$76\pm37$	$3.17\pm0.04$	$0.36 \pm 0.07$	$0.19\pm0.02$	1.89	
	с	3	Me		$64\pm14$	$3.55 \pm 0.10$	$0.26\pm0.04$	$0.11\pm0.02$	2.36	
	d	5	Me		$24\pm 8$	$3.26\pm0.06$	$0.37\pm0.08$	$0.14\pm0.03$	2.64	
10	a	3	Н	CH	$19\pm3$	$2.69\pm0.08$	$1.06\pm0.16$	$0.39\pm0.06$	2.72	
	b	3	Me	CH	$13\pm5$	$2.99\pm0.12$	$0.70\pm0.14$	$0.17\pm0.03$	4.12	
	с	3	Н	Ν	$204\pm34$	$2.81\pm0.02$	$0.69\pm0.08$	$0.31\pm0.12$	2.23	
	d	3	Me	Ν	$134\pm25$	$2.81\pm0.02$	$0.54\pm0.10$	$0.10\pm0.03$	5.40	
1	a	4	Me	CH	$16\pm 6$	n.d.	$0.69\pm0.16$	$0.46\pm0.12$	1.50	
	b	5	Me	CH	$34\pm18$	n.d.	$0.46\pm0.09$	$0.40\pm0.09$	1.15	
	с	6	Me	CH	$8\pm1$	n.d.	$0.59\pm0.12$	$0.43\pm0.15$	1.37	
	d	4	Н	CH	$9\pm2$	n.d.	$2.11\pm0.11$	$0.62\pm0.08$	3.40	
	e	5	Н	CH	$29\pm14$	n.d.	$0.92\pm0.09$	$0.63\pm0.15$	1.46	
	f	5	Me	Ν	$18\pm13$	n.d.	$0.47\pm0.07$	$0.19\pm0.03$	2.47	
	g	6	Me	Ν	$24\pm3$	n.d.	$0.60\pm0.05$	$0.29\pm0.05$	2.07	
12	a			CH	$31\pm 6$	$2.88\pm0.04$	$1.57\pm0.15$	$0.54\pm0.21$	2.91	
	b			Ν	$103\pm31$	$2.48\pm0.06$	$0.92\pm0.24$	$0.15\pm0.03$	6.13	
13					n.d.	$-0.42\pm0.02$	$0.28\pm0.08$	$0.20\pm0.10$	1.40	
14					n.d.	$0.81\pm0.01$	$0.38\pm0.07$	$0.017\pm0.05$	2.24	
15					n.d.	$1.1\pm0.02$	$0.07\pm0.02$	$0.03\pm0.01$	2.33	
16	a	3			$162\pm43$	n.d.		n.d.		
	b	5			$37\pm17$	$2.80\pm0.25$	$0.18\pm0.02$	$0.08\pm0.02$	2.25	
17	a	4	Н		$45\pm12$	n.d.	$0.80\pm0.17$	$0.03\pm0.01$	26.67	
	b	6	Н		$62\pm8$	n.d.	$0.61\pm0.08$	$0.18\pm0.01$	3.39	
	с	4	Me		$51\pm13$	n.d.	$0.88\pm0.14$	$0.14\pm0.02$	6.29	
	d	6	Me		$42\pm11$	n.d.	$1.21\pm0.22$	$0.06\pm0.01$	20.17	

TABLE 1 In vitro and ex vivo properties of <sup>99m</sup>Tc-labeled benzothiazole aniline derivatives for imaging cerebral Aβ plaques.

n.d., not determined.

<sup>a</sup>Clearance ratio of 60 min/2 min.

blocking of the binding was indicative of similar binding sites between **6** and thioflavin-T. Along with the low mass (<600 Da) and moderate lipophilicity, **6** was investigated in normal mice (stain, sex, and age not given) for brain uptake. The early uptake was low (0.9 and 0.11% ID in the cerebrum and cerebellum, respectively) and the compound was deemed unsuitable for detecting amyloid deposition in living brains. Moreover, by analyzing the mouse plasma it was found that by 2 min p.i., 85% of the injected activity was metabolized to a polar metabolite, leaving little intact **6** to cross the BBB.

The following year, a group at Beijing Normal tethered a MAMA ligand to the hydroxy on the benzothiazole and evaluated 7 as a potential  $\beta$ -amyloid SPECT probe (55). Following on earlier computational work, this site was selected to minimize impact on binding affinity if the linker is sufficiently

long (56). The Re congener was shown to bind to brain tissue from transgenic mice (C57 APP, 12 months, no sex given) and a human AD patient (male, 57 yo). Once complexed with <sup>99m</sup>Tc, the biodistribution of 7 in normal mice (IRC, no age or sex given) was investigated. Moderate initial uptake ( $1.34 \pm 0.16$  %ID/g) with slow wash-out was observed (0.65 %ID/g at 60 min p.i.). As no A $\beta$  plaques were expected in these normal mice, a roughly 50% reduction in brain uptake points to a high level of non-specific binding. Furthermore, the high amount of 7 remaining in blood even at 2 h p.i. ( $4.71 \pm 1.24$  %ID/g) is problematic for imaging.

In 2009, the originators of PiB prepared six Re complexes of 2-phenylbenzothiazoles which were never radiolabeled (57). These compounds were conjugated through pendant group (8) or integrated into the molecular structure to reduce the overall size (43–45; see below). Biological evaluation of the conjugated compound 8 showed strong binding affinity ( $K_i = 10 \text{ nM}$ ) and reasonable lipophilicity (Log P = 3.26) for permeating the BBB. Although 8 had the highest affinity in this group of putative SPECT radiotracers, the three-carbon tether added to the molecular weight (>600 g/mol compared to < 500 for integrated examples below) and the lipophilicity (integrated were all < 2.60 compared to 3.26). Analogous compounds 11 and 12 were radiolabeled by other groups, as discussed below.

Hoping to minimize the impact the chelator had upon brain uptake, Jia et al. elected to use a piano-stool cyclopentadienyl Tc(I) tricarbonyl core for compounds 9 and 10 (58, 59). The CpM(CO)<sub>3</sub> ligand has several advantages including low molecular weight, compact size, minimal steric impact, and moderate lipophilicity. Importantly, these half-sandwich complexes are highly stable. In their first study, the linker length and number of methyl groups of 9 were varied, with slighty lower affinity compared to a common assay standard  $[^{125}I]$ IMPY (K<sub>i</sub> = 12.5 ± 2.8 nM in this report) (58). Tissue from 12-month-old, female, C57BL6 APPswe/PSEN1 transgenic mice was use for fluorescent staining and autoradiographic experiments which showed co-localization to thioflavin-S (or the absence with age-matched controls). In normal mice (ddY, 5-week-old, male) there was unsatisfactory initial brain uptake (0.26-0.50 %ID/g) and poor wash-out (2 min/60 min %ID/g ranged from 1.9 to 2.8). This poor in vivo performance was attributed to high overall molecular weight and the amide bond of the linker which is available to form hydrogen bond. The follow-up study with 10 changed the link to an ester and added a heteroatom to the aniline moiety (59). These changes had a positive impact on binding affinity but did not significantly improve the brain penetration or clearance. Although this group argues the conjugated approach, tethering the chelate through a linker, is needed to achieve high affinity, later work will demonstrate the benefits of integrating the CpM(CO)<sub>3</sub> group into the targeting vector (vide infra).

Sticking with the conjugated approach, the Beijing group next reported further derivatization of the 2-anilinobenzothiazole core (11; Figure 3) (60). Switching back to BAT for chelating <sup>99m</sup>Tc, a subset of these compounds had as good or better binding affinity compared to [<sup>125</sup>I]IMPY for  $A\beta_{1-42}$ aggregates. Those with the lowest Ki were radiolabeled and evaluated ex vivo in normal mice (ICR, male, no age given). Fluorescent staining with the Re congeners distributed with a similar pattern to thioflavin-S with AD patient and transgenic mouse (C57BL6 APPswe/PSEN1, male, 12-months-old) brain slices. This gave some evidence for specificity. Compound 11d had the highest initial brain uptake reported to date with reasonable clearance. The study's authors propose the lower lipophilicity (as assigned by HPLC retention times) of 11d led to less plasma protein binding. They concede there are other factors which could have led to higher brain uptake which were not explored in the present study. Nonetheless, the encouraging *ex vivo* results led to the first *in vivo* use of a <sup>99m</sup>Tc radiotracer candidate in non-human primates. Semi-quantitative analysis of the SPECT/CT images revealed low initial brain uptake of **11d** (1.23 and 0.78 %ID in a healthy male and female rhesus monkey, respectively) with slow clearance (0.88 and 0.64 %ID at 40 min p.i., respectively). As one of the most comprehensive studies reviewed, this exemplary paper should act as a guide for anyone entering this field.

As part of a follow up study, Zhang et al. altered the linker moiety of **12** to an oligoethyleneoxy to further modify the lipophilicity (61). This did not have the desired effect on binding affinity as the values increased compared to the alkyl chain. There was also a diminished performance in *ex vivo* biodistribution experiments (ICR mice, male, no age given) with lower uptake compared to the promising **11d** (Table 1). Given the better properties of a single methyl on the aniline, it was surprising that these derivatives were not prepared.

#### For cerebrovascular Aß detection

Three compounds, 13-15 (Figure 3; Table 1), were reported as candidates for detecting systemic amyloidosis which may find use in imaging cerebral amyloid angiopathy (CAA) (62). All three bore a 2-phenylbenzothiazole core with different bifunctional chelators used: a mercaptoacetyltriglycine (MAG3; 8), a hydrazinonicotinic acid (HYNIC)/tricine (9), and an imino-diacetic acid (10). The first and third were chosen for their negative charge which was hypothesized to limit the hepatobiliary uptake in favor of renal secretion. When used with a tricine co-ligand, Tc-HYNIC complexes are also polar and shown to have similar favorable excretion characteristic, as seen with <sup>99m</sup>Tc-octreotide derivatives (63). The three candidates were radiolabeled in high yields (80, 70, and 64%, respectively) and evaluated for biodistribution in normal mice (male NMRI, no age given). Given their hydrophilic nature, these compounds unsurprisingly showed low brain uptake (Table 1). Unlike the previous studies, this paper did not report any affinity information or brain tissue binding, likely due to the intended peripheral application. Even so, it was concluded that 13-15 had less than ideal pharmacokinetic characteristics, postulated to be from rapid in vivo metabolism.

In 2014 the Beijing Normal team produced positively charged dipyridylamine-[<sup>99m</sup>Tc]Tc(CO)<sub>3</sub> compounds **16** (64). Both compounds, with 3 or 5 carbon chain linkers, had moderate *in vitro* affinity to  $A\beta_{1-42}$  but  $A\beta_{1-40}$ , the predominate peptide in CAA, was not tested. Given a four-fold higher affinity, only **16b** was radiolabeled. It was hypothesized that the larger chelate required more distance from the 2-phenylthiazole moiety for binding. *Ex vivo* biodistribution in normal mice (ICR, male, 5 weeks) indeed showed little brain uptake at 2 or 60 min. As such, **16b** may be a potential SPECT radiotracer for CAA.

Concurrent with their above efforts, the group from Beijing Normal also explored imino-diacetic- $^{99m}$ Tc chelates 17 for use



with cerebrovascular amyloid (65). Possessing a negative charge will prohibit BBB entry, thus, visualizing only the vascular  $A\beta$  deposits is another way to differentiate between AD and cerebral amyloid angiopathy. This polar chelate had a negative impact on binding affinity which was minimally improved by moving the group further from the targeting vector. Although the goal was not to show good brain penetration, surprisingly these compounds did exhibit some of the best clearance in mouse brains (ICR, 5-week-old, male). Compounds 17a and 17d both had comparable initial uptake at 2 min p.i. vs. the other compounds in Table 1 but the washout was much greater. This may be due to the wider margin of error (circa 20%) at the early timepoints as the amount of radioactivity present in blood was high (22 and 32 %ID/g for 15a and 15d). The blood volume present for each sample at the time of counting may play a significant role in the variation.

#### Benzoxazole and benzofuran

Slightly deviating from the thioflavin dyes, Ono et al. prepared two benzofuran-based <sup>99m</sup>Tc probes with BAT and MAMA (**18a** and **18b**; Figure 4) chelates (66). In an [<sup>125</sup>I]IMPY based inhibition assay, both compounds bound to  $A\beta_{1-42}$  aggregates with the K<sub>i</sub> of **Re-18a** determined to be lower than **Re-18b**. This affinity was also seen in autoradiographic experiments with **18a** using sections of Tg2576 mouse brains. Observed binding was comparable to staining adjacent slices with thioflavin-S. When investigated for biodistribution in wild-type mice (ddY, 5 weeks, no sex given), **18b** had very low and slow uptake with no washout, making it unviable for SPECT imaging. Although **18a** had moderate initial uptake, the washout was still only about 60% after an hour.

In follow up work on these benzofurans, Cheng et al. changed the aniline to a pyridine and varied the number of methyl groups of compound **19** (67). This structure was based upon previous success by the Kyoto group with [<sup>18</sup>F]FPYBF-2 as a selective PET radiotracer for A $\beta$  plaques (68). The group chose the use a BAT chelator and forgo MAMA as the former

is known to have good BBB penetration (69). From Figure 4, comparing the derivatives of 18 and 20 reinforces this decision. Three Re derivatives of **19** were assessed for binding to  $A\beta_{1-42}$ using an [<sup>125</sup>I]IMPY assay. Only **19c** retained similar affinity  $(K_i = 13.6 \pm 1.60 \, \text{nM})$  to IMPY and PiB  $(K_i = 10.5 \pm 1.05$ and 9.00  $\pm$  1.31 nM, respectively). These results were consistent with increasing affinity correlated to increasing methylation seen in previous studies, however, they were somewhat surprisingly higher than the fluorinated versions (68). The radiolabeled analogs underwent autoradiographic analysis using tissue from Tg2576 and healthy age-matched mice (male, 27-months-old). By comparing to co-stained (thioflavin-S) slices, selectivity for Aß plaques was shown. This higher affinity and selectivity did not translate to high brain uptake in normal mice (ddY, male, 5-weeks-old). For comparison, [<sup>18</sup>F]florbetapir has >7 %ID/g in the mouse brain (70). More disheartening is the poor washout after 60 min, with 45% remaining in the best case of 19b.

Structurally similar benzoxazoles 20ab were reported by Wang et al. albeit with longer linker between the BAT and MAMA chelates (71). The Re congeners were again used to obtain the  $A\beta_{1-42}$  binding affinity information. Much like **18a**, the BAT derivative 20a had slightly stronger binding compared to 20b (Figure 4) and both were comparable to the value obtain for IMPY ( $K_i = 10.1 \pm 1.9 \text{ nM}$ ) in the same assay. Selectivity for amyloid plaques was determined using brain tissue slices from 12-month-old C57BL6, APPswe/PSEN1 transgenic mice (no sex given). Both compounds clearly stained A<sup>β</sup> plaques with a similar staining pattern to thioflavin-S. To assess the initial brain uptake and clearance, normal 5-week-old mice (no strain or sex given) were used in biodistribution experiments. Neither derivative of 20 had good initial uptake but the prospects of 20b were further harmed by practically no wash-out. These attempts are therefore not suitable for further evaluation as amyloid targeting SPECT radiotracers.

Follow up studies by the same group led to **21** to improve the biological properties of the benzoxazoles (72). Fluorescent staining with the four Re complexes and thioflavin-S showed consistent co-localization in brain sections from AD patient



tissue and transgenic mice (C57BL6 APPswe/PSEN1, 11months-old, male). Autoradiography with the mouse tissue and the four <sup>99m</sup>Tc-labeled **21** derivatives also revealed similar distribution of A $\beta$  plaques to thioflavin-S stained sections. For *in vitro* binding affinity, only one compound, **21c**, was comparable to [<sup>125</sup>I]IMPY (K<sub>i</sub> = 15.9 vs. 11.5 nM). By comparing four **21** derivatives, the presence of the pyridine group improved the binding affinity. Despite this affinity and reasonable lipophilicity, no derivative had sufficient brain uptake, however, the washout of the pyridinyl derivatives **21c** and **21d** was greatly improved with almost 90% of the latter cleared from the mouse brain. Interestingly, two methyl groups on the amine lowered the affinity whereas higher order substitution led to an increase with **19**.

### Miscellaneous fused rings—Flavones, aurones, napthalenes, and phenylquinoxalines

Beyond the fused heterocycles above,  $^{99m}$ Tc-labeled small molecules based upon other planar, extended pi systems were also explored over the past two decades (Figure 5; Table 2). Two early examples, **22** and **23**, come from the Kyoto group (73). These  $^{99m}$ Tc complexes based upon aurone and flavone employed BAT chelators to hold the radiometal. Both compounds bound to A $\beta_{1-42}$  aggregates in a dose dependent

manner with reported low non-specific binding (<2%). No  $K_i$  values were determined but the % bound at various concentrations were reported. To confirm 22 and 23 had affinity for A $\beta$  plaques, histochemical staining with their Re congeners and thioflavin-S were carried out with mouse brain sections (Tg2567, 30-months-old, female). The separate staining experiments labeled the A $\beta$  plaques in a similar pattern. The two radiolabeled compounds were then evaluated in *ex vivo* biodistribution experiments in normal mice (ddY, 5-weeks-old, no sex given). Both 22 and 23 were minimally brain penetrate but the little that did enter were mostly cleared by 60 min. Particularly, 84% of 23 was cleared and represents one of the best 2:60 min ratios at that point in time. Despite this promising pharmacokinetic property, this class of compounds was never revisited.

Using a unique, neutral  $[^{99m}Tc(CO)_2NO]$  core, Yang et al. labeled the flavonol moiety with iminodiacetate chelators to give **24** and **25** (74). This technetium core has a formal 2+ charge and forms neutral complexes with iminodiacetate. These two compounds were not evaluated beyond autoradiography as results using transgenic mouse tissue (C57 APP/PS1, 12 months, no sex given) found no binding to amyloid.

In another report from Beijing Normal University, two naphthylethylidene derivatives selected for radiolabeling with  $^{99m}$ Tc due to similarities to PET radiotracers [ $^{18}$ F]FDDNP and [ $^{18}$ F]FENE (75). Choosing this core is surprising as these radiotracers are known to have affinity for both A $\beta$  and neurofibrillary tangles. Moreover, the  $^{18}$ F-labeled compounds

						% Injected dose/gram (%ID/g)			
Compou	nd	n	R	K <sub>i</sub> (nM)	Log P	2 min	60 min	<b>Ratio</b> <sup>a</sup>	
22				n.d.	$2.77\pm0.04$	$0.64\pm0.07$	$0.23\pm0.23$	2.78	
23				n.d.	$2.23\pm0.04$	$0.79\pm0.12$	$0.11\pm0.04$	7.18	
24				n.d.	n.d.		n.d.		
25				n.d.	n.d.		n.d.		
26				$3.65\pm0.49\mu\mathrm{M}$	$1.70\pm0.02^{\rm b}$	$0.65\pm0.09$	$0.36\pm0.06$	1.81	
27				$4.64\pm0.77\mu\mathrm{M}$	$1.89\pm0.04^{\rm b}$	$0.28\pm0.03$	$0.12\pm0.02$	2.33	
28	a		NH <sub>2</sub>	n.d.	n.d.		n.d.		
	b		NHCH <sub>3</sub>	296	n.d.		n.d.		
	c		$NH(CH_3)_2$	53.7	2.45	0.88	0.25	3.52	
29	a	2		33.2 <sup>c</sup>	$1.08\pm0.02$	$0.78\pm0.07$	$0.09\pm0.03$	8.67	
	b	3		102.5 <sup>c</sup>	$1.10\pm0.06$	$0.86\pm0.07$	$0.12\pm0.02$	7.17	

TABLE 2 In vitro and ex vivo analysis of fused (hetero)cyclic radiotracer candidates in Figure 5.

n.d., not determined.

<sup>a</sup>Clearance ratio of 60 min/2 min.

°IC<sub>50</sub>.

have low affinity for A $\beta$  aggregates with K<sub>i</sub> values in the 400– 470 nM range. The group also chose to use a MAMA chelator despite precedent of higher non-specific binding and lower brain uptake. After coordinating Re, the binding affinity of **26** and **27** were found to be 10-fold lower (3–5  $\mu$ M range) but retained some selectivity for A $\beta$  in brain tissue slices of transgenic mice (C57BL6 APP<sub>swe</sub>/PSEN1, 12 months, no sex given). Low molecular weight and lipophilicities in the ideal range held promise for brain uptake but virtually none was observed in *ex vivo* biodistribution (ICR mice, no age or sex given). High non-specific binding, as indicated by poor clearance at 60 min p.i., was also seen. Unsurprisingly, no follow up studies were conducted using the DDNP or ENE backbones.

Building upon early success with [<sup>123</sup>I]ABC577, the Kyoto group reported three phenylquinazoline derivatives conjugated with BAT-<sup>99m</sup>Tc chelates (76). This scaffold was report to have higher binding affinity and more selectivity for A $\beta$  than other commonly employed PET probes (i.e., PiB, florbetapir). Adding the three carbon linker and Re chelate did lower the binding affinity significantly. Amyloid aggregate binding remained for the dimethyl derivative **28c** but no binding was observed for the other two. Although autoradiographic experiments showed promising co-staining with **28c** and thioflavin-S, low initial brain uptake and moderate washout in healthy male DDY mice (no age given). The authors hypothesized the high stomach uptake pointed to decomposition of **28c** as contributing to low brain uptake.

Two novel 2-aryl imidazole[2,1-b]benzothiazole (IBT; **29**) derivatives were synthesized and successfully labeled with  $^{99m}$ Tc radionuclide at high radiochemical purity using fac- $[^{99m}$ Tc(CO)<sub>3</sub>(H<sub>2</sub>O)<sub>3</sub>]<sup>+</sup> synthon (77). Autoradiography and staining with Congo Red confirmed that these complexes bind

to AB plaques in brain sections of diseased rat models (male Wistar, stereotactic injection with AB oligomers at 3 months), but the age of the rats at the time of tissue collection is not given. Both candidates exhibited moderate affinity toward βamyloid aggregates with IC50 values of 33.2 and 102.5 nM in Aß aggregates. Unlike most of the studies presented in this review, blocking studies were conducted with  $A\beta_{1-42}$  aggregate binding to demonstrate some selectivity. In the inhibition assay results, 27b was found to have higher binding and to be more selectivity. It should be noted that the higher  $IC_{50}$ values represent displacement of a fixed amount of 99mTclabeled compounds by BTA-1 (benzothiazole derivative akin to thioflavin-S) and therefore cannot be directly compared to the Ki values obtained from previous studies. As the length of the linker (alkyl chain) increased, so did the IC50 value, the lipophilicity, and the initial brain uptake. Although both 29ab stable in saline after 2h (92% of both remained intact), no histidine or cysteine challenges were completed. Nevertheless, both 29a and **b** showed little initial brain uptake and low non-specific binding in biodistribution studies (BALB-C, no sex or age given) which was indicated by the clearance ratios of 8.67 and 7.17, respectively. The relatively high hydrophilicity and molecular weight (>600 Da) may have led to the blunted brain uptake.

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# Stilbenes, chalcones, and curcumin derivatives

Following the FDA approval of florbetapir in 2012, investigators embarked on developing <sup>99m</sup>Tc-labeled stilbene analogs (Figure 6; Table 3). The first attempt from Jia et al. gave

<sup>&</sup>lt;sup>b</sup>Log D.

**30a** and **30b** as the second half of a 2014 study, which included **16** above (64). Only the latter had a strong binding affinity to  $A\beta_{1-42}$  aggregates and was radiolabeled. As the aim of this study was to develop a BBB impermeable radiotracer for CAA, it was favorable that **30b** was found to have low initial brain uptake with minor non-specific binding after 60 min. With a reasonably high binding affinity and desired low brain uptake, it is somewhat surprising no follow up studies have been reported.

A year later, a team from Australia prepared analogous complexes 30c, 30d, and 31 (78). A chemistry focused paper, there was little in vitro evaluation of these four candidates but there was a rare example of ex vivo biodistribution data from a transgenic mouse model (APP/PS1, 10 months, no sex given). There was little brain uptake of 31b with no statistical difference between wild type and transgenic mice. The exchange of a pyridyl for an acetate group eliminated the positive charge of compounds 30 but low BBB permeability persisted. This paper also reported the first example of small animal SPECT imaging of <sup>99m</sup>Tc-based AD radiotracers. With 30d and transgenic mice (APP/PS1, 15 months, no sex given), the activity was found to mostly accumulate in the bladder, liver, and kidney. No brain uptake was observed and there was some accumulation in the thyroid, pointing to possible oxidative degradation to pertechnetate.

In the same studying reporting 12, Zhang et al. introduced an oligoethyleneoxy linker between 99mTc/Re-BAT and a βamyloid binding styrene scaffold (61). They intended to improve brain uptake by decreasing the hydrophobicity and replacing the alkyl linker between the targeting vector and chelator with PEG chains. Compound 32a had the highest binding affinity to A $\beta$  plaques (K<sub>i</sub> = 13.4 nM). Furthermore, the capability of both 32 analogs to bind A\beta plaques was positively verified by in vitro fluorescent staining with transgenic mice (C57BL6 APPswe/PSEN1, 12 months, no sex given) and AD patient tissue samples. Similarly, ex vivo autoradiography showcased amyloid binding in transgenic mice (but not wild type), matching thioflavin S staining on adjacent slices. Compounds 32a had high binding affinity and low clearance ratio, while compounds 32b had lower binding affinity but higher clearance ratios (for reference,  $[^{125}I]$ IMPY had a K<sub>i</sub> of  $12.5 \pm 1.4$  nM). Extending the linker length therefore had a mixed result, further illustrating the difficulty in attaining a delicate balance between the pharmacokinetic parameters. Furthermore, the biodistribution in healthy mice (ICR, male, no age given) showed less uptake than compounds 12 on average, but the clearance ratios were better, possibly indicating less non-specific binding. Compound 32b had a higher uptake than the previous study (with a clearance ratio of 7.33), indicating some potential as a SPECT probe for imaging AD. As with 11d, 32a was used in vivo with SPECT imaging of a non-human primate (rhesus monkeys, n= 6, 3/3 male/female, 4–30 years old). Brains were scanned for 60 min to show moderate brain uptake with little washout. The activity peaked at 20 min with 2-2.6 %ID/g (semi-quantitative)

with clearance ratios around 1.4. This uptake is higher than for **11d** but much lower than the PET analogs. That said, there were no A $\beta$  plaques present in these animals and further investigation into these promising candidates may be warranted.

Published in 2010 following their first flavone study, Ono et al. reported four chalcone derivatives with **33** as a scaffold (79). These were conjugated to BAT and MAMA through either a three or five methylene linker (Figure 6; Table 3). Of the four, **33d** had the lowest binding to  $A\beta_{1-42}$  aggregates but had by far the best pharmacokinetics (highest initial uptake, best clearance). Using the same mice as their previous study (Tg2567, 30-months-old, female), brain sections were stained with similar intensity for all four derivatives (matched thioflavin-S staining pattern). Given the *ex vivo* and *in vitro* results, there appears to be a disconnect between solution  $A\beta_{1-42}$  aggregate binding affinity and the ideal pharmacokinetics for imaging. Perhaps this was why the authors did not determine the K<sub>i</sub> values for these compounds or for **23** and **24**.

Four <sup>99m</sup>Tc-labeled dibenzylideneacetone derivatives (80) were successfully synthesized and biologically evaluated. This backbone was selected due to its similarity to curcumin (81). Previous work had shown that the para position of the phenoxy ring had a high tolerance for bulky substitutions. Both BAT and MAMA chelators were conjugated and radiolabeled. Complexes 34 showed high to moderate affinity for  $A\beta_{1-42}$  aggregates (K<sub>i</sub>  $= 13.6 - 120.9 \text{ nM}; [^{125}\text{I}]\text{IMPY} = 11.5 \pm 2.5 \text{ nM})$ , and selectively stained β-amyloid plaques on brain sections of transgenic mice (C57BL6 APPswe/PSEN1, 11 months, no sex given). In vitro fluorescent staining confirmed the presence and distribution of β-amyloid plaques using thioflavin-S as well as absence of staining in age-matched control mice tissue. Biodistribution in normal mice (ICR, 5 weeks, male) revealed that 34b and d exhibit moderate clearance ratios, while 34a and 34c exhibit higher clearance ratios. Once again, the MAMA based ligand showed a higher non-specific binding profile compared to BAT. Unfortunately, the initial uptake was low for all compounds tested. Additional refinements are needed to improve the penetration through BBB, but these promising results indicate that 34a may be a potential SPECT probe. It has a high affinity for  $\beta$ -amyloid plaques and an excellent clearance ratio; therefore, it is the most suitable derivative from this series for further attention.

### Bivalent Tc-99m ligands for AD

High binding affinity is seen as a prerequisite for developing useful molecular imaging probes, although some results within this review may challenge this assumption, at least for imaging A $\beta$  aggregates. Nonetheless, multivalent ligands are known to have enhanced binding compared with monovalent counterparts (82). The enhanced binding of bivalent ligands may arise from rebinding of the ligand prior to complete dissociation.



TABLE 3	In vitro and	ex vivo evaluation	of stilbenes 30-32.
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				% In	jected dose/gram	(%ID/g)		
Compound		n	R	K <sub>i</sub> (nM)	Log P	2 min	60 min	Ratio <sup>a</sup>
30	a	3	N(CH <sub>3</sub> ) <sub>2</sub>	$366\pm70$	n.d.		n.d.	
	b	5	$N(CH_3)_2$	$78\pm13$	$1.79\pm0.14$	$0.24\pm0.02$	$0.10\pm0.04$	2.40
	c	1	Н	n.d.	1.8		n.d.	
	d	1	$N(CH_3)_2$	n.d.	1.0		n.d.	
31	а	1	Н	n.d.	2.1		n.d.	
	b	1	$N(CH_3)_2$	n.d.	1.7	$0.25\pm0.05$	$0.21\pm0.02$	1.19 <sup>b</sup>
						$0.24\pm0.02$	$0.19\pm0.01$	1.26 <sup>b,c</sup>
32	a	1		$13.4\pm3.3$	$3.04\pm0.02$	$2.10\pm0.22$	$0.61\pm0.08$	3.44
	b	2		$142.1\pm24.4$	$3.15\pm0.01$	$1.10\pm0.11$	$0.15\pm0.02$	7.33
33	а	5	СО	n.d.	$2.55\pm0.19$	$0.22\pm0.05$	$0.11\pm0.01$	2.00
	b	5	CH2	n.d.	$2.73\pm0.16$	$0.78\pm0.16$	$0.16\pm0.01$	4.88
	с	3	CO	n.d.	$1.51\pm0.09$	$0.62\pm0.27$	$0.16\pm0.08$	3.88
	d	3	CH3	n.d.	$2.51\pm0.05$	$1.48\pm0.44$	$0.17\pm0.06$	8.71
34	a	3	$CH_2$	$24.7\pm 6.1$	$3.36\pm0.08$	$0.49\pm0.08$	$0.08\pm0.01$	6.13
	b	5	$CH_2$	$13.6\pm7.8$	$3.17\pm0.10$	$0.47\pm0.11$	$0.12\pm0.02$	3.92
	с	3	СО	$120.9\pm4.3$	$3.52\pm0.10$	$0.48\pm0.06$	$0.09\pm0.01$	5.33
	d	5	СО	$59.1\pm24.0$	$3.57\pm0.15$	$0.31\pm0.06$	$0.15\pm0.02$	2.07

n.d., not determined.

<sup>a</sup>Clearance ratio of 60 min/2 min.

<sup>b</sup>30/2 min.

<sup>c</sup>APP/PS1 transgenic mice.

Another explanation is multisite binding, however the distance between targeting vectors may be too short in the cases reviewed herein. The Kyoto group built upon their previous work with <sup>99m</sup>Tc-hydroxamamide complexes, shown to have high kinetic stability, to prepare 1:2 metal to ligand complexes **35–39** (Figure 7; Table 4) (83). These were easily radiolabeled in high yield and two isomers were isolated as previously reported with <sup>99m</sup>Tc-hydroxamamides. The absolute configurations were not determined but the isomers with shorter retention times on the HPLC were designated A and the latter as B. In the solution based A $\beta_{1-42}$  aggregate binding assay, only the A isomers were used. These assays showed the bivalent ligands had stronger binding with **36** and **38** having greater affinity compared to **35** and **37**, respectively. In the case where no amyloid targeting vector was present (**39**), no binding was observed.

They next evaluated the inhibitory binding by displacing **35–39** from A $\beta_{1-42}$  aggregates with unlabeled PiB, thus higher values indicate stronger binding. Trends from the assay results



matched their previous assay, with no binding of **39** seen. Interestingly, the A isomer of **35** and **36** showed stronger binding whereas the opposite was observed for **37** and **38**. Finally, *in vitro* autoradiography comparing radiotracer binding in wild-type (no strain given, female, 28 months) and transgenic mice (Tg2576, female, 28 months) showed (a) higher uptake for all eight compounds in the latter group, (b) good correlation to thioflavin-S staining, and (c) appreciable blocking with unlabeled PiB in transgenic animals. Preliminary biodistribution studies with the highest affinity compound, **36**, showed very low initial uptake (0.28 %ID/g).

Given the low brain uptake with the bivalent compounds, the same group published two more studies investigating their utility for imaging CAA (84, 85). As  $A\beta_{1-40}$  is the predominate subtype of amyloid in this disease, Iikuni et al. measured the affinity of **35–38** to these aggregates (84). After assessing brain permeability compared to a prototypical PET agent, [<sup>18</sup>F]florbetapir, low uptake led to the conclusion **35–38** would be well suited for this type of amyloidosis. More of this approach, directly comparing putative SPECT radiotracers to clinical PET agents, should be conducted with future studies. Another rare experiment from this study was *in vivo* SPECT/CT with Tg2576

mice and wild-type mice (female, 27 or 30 months) 30 min p.i. Although specific binding was observed in autoradiography studies with brain tissue from these rodents, there was no differential distribution observed *in vivo* between transgenic and wild-type mice with **36A**. This could be due to high blood:brain ratio observed in the *ex vivo* studies (50:1–60:1). The contribution from the non-specific binding in plasma could be overwhelming the small signal from cerebral amyloid. Further optimization is required for *in vivo* imaging cerebral amyloid angiopathy.

To avoid obtaining two geometric isomers upon chelation of  $^{99m}$ Tc, the authors used a methylated hydroxamamide (85). Previous studies also noted this alteration increased stability (86). Herein, methylating **38** led to **40** with a marked increase of stability in mouse plasma (ddY, male, 5 weeks) from 86 to 93% radiochemical purity at 2 h. Removing the hydrogen bonding amine group did not improve brain permeability. Both **38** and **40** had circa 0.35 %ID/g in ddY mice at 2 min p.i. Binding affinity was negatively impacted by the methyl groups with the IC<sub>50</sub> values of PiB displacement dropping to 0.56  $\mu$ M. *In vitro* autoradiography revealed significantly higher nonspecific binding to Tg2576 (female, 28 months) brain sections.

					% Injected dose/gram (%ID/g)			
Compound <sup>a</sup>		R	х	IC <sub>50</sub> (μM)	2 min	60 min	Ratio <sup>b</sup>	
35	Α			$0.72\pm0.10$	$0.37\pm0.05$	$0.08\pm0.00$	4.62	
	В			$0.38\pm0.08$		n.d.		
36	Α			$16.40\pm2.47$	$0.28\pm0.03$	$0.11\pm0.01$	2.55	
	В			$2.55\pm0.45$		n.d.		
37	Α			$0.26\pm0.02$		n.d.		
	В			$0.47\pm0.05$	$0.36\pm0.03$	$0.08\pm0.02$	4.50	
38	Α			$2.80\pm0.32$		n.d.		
	В			$5.78\pm0.53$	$0.37\pm0.11$	$0.05\pm\!0.01$	7.40	
39				n.d.		n.d.		
40				$0.56\pm0.08$	$0.35\pm0.04$	$0.11\pm0.01$	3.18	
41	a	CH <sub>3</sub>	CH <sub>3</sub>	n.d.		n.d.		
	b	$C_2H_4N(CH_3)_2$	CH <sub>3</sub>	n.d.		n.d.		
	с	CH <sub>3</sub>	Н	n.d.		n.d.		
	d	$C_2H_4N(CH_3)_2$	Н	n.d.		n.d.		
42	a	CH <sub>3</sub>		n.d.		n.d.		
	b	$C_2H_4N(CH_3)_2$		n.d.		n.d.		

TABLE 4 In vitro and ex vivo data for bivalent Aβ binding SPECT radiotracers.

n.d., not determined.

<sup>a</sup>Capital letter apply to undetermined isomers of 35-39 with "A" eluting first on HPLC.

<sup>b</sup>Clearance ratio of 60/2 min.

This contrasted with distinctive labeling of  $A\beta$  plaques with **38**. Similar outcomes were observed with human CAA patient brain tissues.

In a report focused on chemistry and radiolabeling, Fletcher et al. provided details for six bivalent thiosemicarbazide ligands **41** and **42** (87). The sole biological evaluation was the interaction of the ligands with  $A\beta_{1-42}$  in human tissue samples. The fluorescent signal from Thioflavin-T would decrease with respect to increased ligand binding. It was found that the **41d** had the greatest signal reduction, followed by **42b**, **41c**, **41b**, **42a**, and the lowest reduction was **41a**. It appears the phenolic and amino groups present on the benzofuran and stilbene groups had a positive benefit to binding affinity. Only the stilbenes (**42**) were radiolabeled with <sup>99m</sup>Tc but no further biological evaluation was conducted.

### Integrated Tc-99m Aβ probes

Radiometalated PET or SPECT probes fall into three categories: (1) colloids, (2) conjugated, such as those discuss above, and (3) metal-centric or integrated small molecules. By making the metal an integral part of the scaffold and target binding motif, the molecular weight, volume, and lipophilicity (from linkers and chelators) can be minimized. The examples below include the most promising <sup>99m</sup>Tc-based amyloid radiotracers reported with respect to initial brain uptake and clearance after 60 or 90 min p.i. They often possess lower

*in vitro* binding affinity, but this may not be such as important factor for *in vivo* interactions with Aβ-sheets.

#### Benzo-thiazole and -imidazole

Starting from 2009, a study from the inventors of [<sup>11</sup>C]PiB (first PET amyloid radiotracer to be widely used), several 99m Tcbased candidates, 43-48, were reported (Figure 8; Table 5) (57, 88). The integrated approach was adopted for these as a conjugated analog, 8, was thought to be too lipophilic (Log P = 3.26) and heavy (MW = 617) for good BBB permeability. Integrating the tetradentate chelators into the pharmacophore would address these concerns. Additionally, by placing these groups at the 6 and 4' positions, their electron donating effects will promote higher binding affinity. The weights were decreased to <550 Da and the lipophilicities now ranged from 1.21 to 2.59; within the ideal range. In the initial study (43a, 44ab, and 45ab) the K<sub>i</sub> values were reasonable for imaging amyloid (29.7-88.6 nM), albeit lower than the 8 (10 nM). The followup study in 2013 was by far the most extensive structure activity relationship investigation in this area of research. In total, an additional 21 compounds were prepared and evaluated for lipophilicity and binding affinity (Table 5). By varying the heteroatoms of the chelator, the lipophilicity was finetuned. For example, switching from an SN<sub>2</sub>O chelator to SN<sub>2</sub>S raised the lipophilicity of the Re chelates an average of 0.75 (e.g.,



**45a** vs. **45b**). With respect to binding affinity, it was generally found that including a fluorine or methoxy donating group enhanced binding affinity. The position of the chelators also had a significant impact on affinity. The study authors found a 2–7-fold increase when a semi-rigid chelator was attached in the 4' vs. 3' position (**45** vs. **47**). Although one of the largest efforts in this area, no Tc labeling was completed along with no *ex vivo* biodistribution experiments to confirm brain uptake.

Hoping to take advantage of the kinetic inertness of the <sup>99m</sup>Tc(CO)<sub>3</sub> core (89), Sapati et al. produced the neutral compound 49. Radiolabeling was highly efficient (95% radiochemical yield) and was found to be stable in serum for up to 6 h. Histidine and cysteine challenges likewise caused no degradation of 49. No  $IC_{50}$  or  $K_i$  values were given for this compound. By mixing 0.05 nM of radiotracer with  $20\,\mu M$  of  $A\beta_{1-40}$  peptide, 4.6  $\pm$  0.3% of 49 was found to specifically bind. To assess specificity, thioflavin T was used to inhibition roughly 25% of that initial binding at 10 and 30 mM. With the lipophilicity near the bottom of the ideal range, 49 was evaluated in normal mice (no strain, age, or sex given). Little change to uptake in the gastrointestinal tract after 2 h p.i. further demonstrated the stability of 49. Unfortunately, brain uptake was minimal and clearance after 1h was poor (Table 5).

As previously mentioned for **9**, the cyclopentadienyl pianostool chelates are compact, moderately lipophilic groups (90). A preliminary study affixing this chelate to the anilino group showed excellent binding affinity despite the metal center being integrated into the pharmacophore (91). The rhenium congener of 50 was also observed to co-stain areas of human AD brain tissue with thioflavin S using confocal microscopy. After demonstrating good stability in saline and under transchelation challenges (>95 and >90% after 6 h, respectively), brain uptake was evaluated. Biodistributions studies in normal mice (Swiss albino, 5 weeks, no sex given) showed low initial uptake and poor clearance after 90 min. In a rare ex vivo biodistribution experiment in transgenic animals (C57BL6 5xFAD, 7 months, no sex given), the initial uptake was the same but there was a 4-fold increase in uptake after 90 min. Static gamma camera images also clearly showed brain accumulation of 50. Increase uptake was attributed to the presence of confirmed amyloid plaques (costaining post-mortem tissue). This study demonstrated that an effective SPECT radiotracer will depend on an intricate balance of factors and the importance of including disease models in their evaluation.

Building on their promising study, the group from Demokritos Institute prepared another set of candidates wherein the anilino group was replaced the  $Cp^{99m}Tc(CO)_3$  core (so-called cytectrenes) (92). Compounds 51 showed remarkable brain uptake and clearance in normal mice (Swiss albino, 5 weeks, male). These were by far the highest uptake observed at 2 min p.i. and greatest clearance (Table 5). Initial uptake of **51a** was on par with [<sup>18</sup>F]florbetapir (7.33 %ID/g in mice) (70) and was greater than [<sup>18</sup>F]florbetaben (4.77 %ID/g) (93). More remarkable still was the high affinity for **51b** and **51c**, further emphasizing the minimal impact this <sup>99m</sup>Tc chelate has on biological activity. Further biodistribution studies of **51a** in transgenic mice (C57BL6 5xFAD, 11 months, no sex) showed higher accumulation at 15 min p.i. compared

					% In	jected dose/gram (%	oID/g)
Compound		X/R	K <sub>i</sub> (nM)	Log P	2 min	60 min	Ratio <sup>a</sup>
43	a	N(CH <sub>3</sub> ) <sub>2</sub>	30	2.59		n.d.	
	b	Н	556	2.54		n.d.	
	с	F	617	2.67		n.d.	
	d	OCH3	85	2.37		n.d.	
14	a	OH	89	1.21		n.d.	
	b	OCH <sub>3</sub> <sup>b</sup>	87	2.52		n.d.	
	с	Н	378	2.65		n.d.	
	d	F	118	2.84		n.d.	
15	a	O/OCH <sub>3</sub>	30	1.65		n.d.	
	ь	S/OCH <sub>3</sub>	43	2.30		n.d.	
	с	O/H	109	1.70		n.d.	
	d	O/F	64	1.90		n.d.	
	e	O/OCH <sub>3</sub>	30	1.65		n.d.	
	f	S/H	38	2.41		n.d.	
	g	S/F	31	2.60		n.d.	
	h	S/OCH <sub>3</sub>	43	2.30		n.d.	
6	a	N/H	90	2.333		n.d.	
	ь	N/F	113	2.50		n.d.	
	с	N/OCH <sub>3</sub>	61	2.21		n.d.	
7	a	O/H	280	1.68		n.d.	
	ь	O/F	226	1.87		n.d.	
	с	O/OCH <sub>3</sub>	140	1.59		n.d.	
	d	S/H	264	2.49		n.d.	
	e	S/F	93	2.65		n.d.	
	f	S/OCH <sub>3</sub>	132	2.45		n.d.	
8	a	Н	200	2.35		n.d.	
	b	F	148	3.53		n.d.	
	с	OCH <sub>3</sub>	178	3.25		n.d.	
9			$> 30  \mathrm{mM}$	$1.03\pm0.01$	$0.25\pm0.04$	$0.11\pm0.01$	2.27
60			$13.6\pm4.8$	$2.35\pm0.08$	$0.53\pm0.11$	$0.25\pm0.08^{c}$	2.12 <sup>c</sup>
					$0.52 \pm 0.08^d$	$1.94\pm0.25^{c,d}$	
51	a	S	$65.8 \pm 21.3$	$2.52\pm0.14$	$7.94 \pm 1.46$	$0.20\pm0.03^{c}$	39.7 <sup>c</sup>
	b	NH	$7.0\pm2.9$	$1.84\pm\!0.17$	$3.99\pm0.60$	$0.04\pm0.01^{\rm c}$	99.8 <sup>c</sup>
	с	NCH <sub>3</sub>	$5.7 \pm 2.9$	$1.50\pm0.12$	$5.36\pm0.65$	$0.09\pm0.02^{c}$	59.6 <sup>c</sup>

## TABLE 5 In vitro and ex vivo data for integrated <sup>99m</sup>Tc probes in Figure 8.

n.d., not determined.

<sup>a</sup>Clearance ratio of 60/2 min.

<sup>b</sup>Repeated in Pan et al. (88).

<sup>c</sup>90 min.

<sup>d</sup>In Tg mice.

to their wild-type littermates ( $3.90 \pm 0.19$  vs.  $2.68 \pm 0.06$ , respectively). Staining of brain tissue from the transgenic group with thioflavin S confirmed the presence of amyloid plaques, however, no co-staining or autoradiography with **51a** was reported. Anti-amyloid aggregation activity of the three Re congeners was also evaluated but is outside of the scope of this review.

#### Miscellaneous integrated radiotracers

Seven other integrated compounds with various pharmacophores were reported (Figure 9; Table 6). The biphenyl **52** was an early example from the University of Pennsylvania. They incorporated a MAMA– or BAT–like ligand into one anilino ring but these were not able to



TABLE 6	In vit	tro and	er viv	o data	for	comr	ounds	in	Figure	٩
IADLE 0	111 VIG	.ro anu	CX VIV	o uata	101	COIIIL	ounus		FIGULE	э.

					% Injected dose/gram (%ID/g)			
Compound		X/R	K <sub>i</sub> (nM)	Log P	2 min	60 min	Ratio <sup>a</sup>	
52	a	СО	n.d.	2.23	$0.29\pm0.04$	$0.19\pm0.03$	1.53	
	b	CH <sub>2</sub>	n.d.	2.57	$1.18\pm0.20$	$0.46\pm0.08$	2.57	
53			11.16 <sup>b</sup>	n.d.	$0.48\pm0.05$	$0.44\pm0.08$	1.09	
54	a	$H_2O$	n.d.	n.d.		n.d.		
	b	imidazole	n.d.	n.d.		n.d.		
	с	Isocyanocyclo-hexane	n.d.	n.d.		n.d.		
55	a	n = 1	$899\pm78$	$2.89\pm0.09$	$4.10\pm0.38$	$0.50\pm0.08$	8.20	
	b	n = 2	$211\pm19$	$3.61\pm0.04$	$2.30\pm0.27$	$0.55\pm0.08$	4.18	
	с	n = 3	$108\pm16$	$3.45\pm0.09$	$1.11\pm0.34$	$0.51\pm0.08$	2.18	
6			n.d.	$1.44\pm0.02$		n.d.		
57			855	$1.87\pm0.02$		n.d.		
58	a	$X=X'=CH_2$	$260\pm82$	$1.54\pm0.03$	$0.15\pm0.06$	$0.17\pm0.01$	0.88	
	ь	$X=CH_2; X'=CO$	$241\pm46$	$2.04\pm0.07$	$0.36\pm0.09$	$0.15\pm0.02$	2.4	
	с	X=X <sup>′</sup> =CO	$260\pm73$	n.d.		n.d.		

n.d., not determined.

<sup>a</sup>Clearance ratio of 60 min/2 min.

<sup>b</sup>K<sub>d</sub>.

adequately enter the brain of mice (ICR, 2–3 months, male). The clearance after 60 min was also low, indicating some non-specific binding. Another detriment to **52** was a side reaction upon coordination. An imine formed a much more lipophilic product which, although easily removed, decreased the radiochemical yield.

Along with conjugated examples 24 and 25, Yang et al. prepared 53. This flavone-like complex had similar binding affinity to reported fluorescent dyes and, unlike the former two candidates, 53 was able to stain A $\beta$  aggregates in tissue samples from transgenic mice (C57 APP-PS1, 12 months, no sex given). In biodistribution studies, the low initial uptake

was not cleared from the brain (Table 6). Both the poor uptake and high non-specific binding could be due to the labile water ligand <sup>99m</sup>Tc. If replaced in the blood pool, **53** may become charged and not enter the brain. Likewise, binding to nucleophilic peptide residues could lead to high retention to parenchyma.

Combining the stable  $d^6$  core of  ${}^{99m}$ Tc(CO)<sub>3</sub> with a curcumin pharmacophore led to candidate radiotracer 54. Sagnou et al. labeled the  $\beta$ -diketone with various monodentate co-ligands. The initial water complex 54a was readily prepared (90%) with moderate and excellent ligand exchange for imidazole (25–35%) and Takalloobanafshi et al.

isocyanocyclohexane (90%), respectively. As a chemistry focused paper, little biological evaluation was completed. Staining of adjacent slices from human post-mortem tissue showed co-localization of all three derivatives with curcumin. No further studies with this compound class were reported.

Returning to the piano stool Cp<sup>99m</sup>Tc(CO)<sub>3</sub> core, the group at Beijing Normal University prepared chalcone derivatives 55 with 1-3 unsaturated groups. Although still an extended pi system, the flexibility of these groups vs. aryl rings may have contributed to the poor binding affinity ranging from 100 to 900 nM. Each of the three 55 derivatives selectively stained  $A\beta$  aggregates in tissue from transgenic mice (C57BL6 APPswe/PSEN1, 11 months, male) and a human AD patient. Moreover, measured lipophilicity values were within or near the ideal range for brain permeation. Biodistribution in normal mice (ICR, 5 weeks, male) showed an inverse relationship between brain uptake and molecular weight or lipophilicity. The uptake and clearance are remarkably higher than bifunctional conjugate analogs like 33. Unfortunately, blocking of PgP with cyclosporin A increased the brain uptake (1.5-1.6-fold), indicating 55 may be a substrate for this efflux transporter.

Compounds 56 and 57 were reported by Hayne et al. to evaluate neutral [TcO]<sup>3+</sup> core integrated into a styrylpyridine pharmacophore. The Re analogs were crystallized and x-ray crystallographic analysis showed a distorted square-pyramidal environment around the metal center. As planar structures are desired for high Aß aggregate binding, this orientation may harm affinity. Nonetheless, both compounds were radiolabeled with  $^{99m}$ Tc in >90 and 60% radiochemical yield for 56 and 57, respectively. The stability of 57 was not reported and 56 was shown to be stable to glutathione exchange (<5% decomposition after 1 h) but not histidine (16% hydrophilic species after 1 h). As a chemistry focused paper, the biological evaluation was limited to A $\beta$  aggregate binding and confocal microscopy of 57. As with other compounds, co-staining of adjacent slices of post-mortem tissue samples from confirmed AD patients was observed. More biological evaluation may not be justified given the low stability to the histidine challenge, poor radiochemical yield, and low binding affinity.

In the most recent report covered in this review, Spyrou et al. outlined the preparation of three derivatives of **58**. These incorporated the stilbene backbone with an N<sub>3</sub>S chelator for oxotechnetium (V) but replaced the phenolate group of **57** with a thiolate to improve the stability and lower lipophilicity. Both amide and amine donors were prepared with improved stability of both (>94% parent of both after 30 min). All three Re complexes of **58** had similar binding affinity for  $A\beta_{1-42}$ . Compound **58c** could not be isolated in sufficient purity for *ex vivo* experiments, however, neither **58a** or **58b** has sufficient initial brain uptake or good clearance.

### Future perspective

A diverse array of over fifty 99mTc-labeled scaffolds has been reported, however, with few promising candidates for clinical translation. There is minimal correlation between binding affinity and favorable pharmacokinetics from the reviewed scaffolds. For example, looking at 51 and 55, there is in fact an inverse relationship between affinity and uptake/clearance properties. Although in vitro binding affinity experiments are easy to complete, their utility for selecting candidates for radiolabeling and in vivo/ex vivo experiments may be overvalued. Furthermore, co-staining experiments with validated histochemical amyloid dyes and novel radiotracer candidates (Re-congener fluorescence or autoradiography) rarely disagreed. While this in vitro data remains valuable, to significantly advance 99mTc-based radiotracers for brain SPECT imaging, "failing faster" in whole organisms (e.g., animals) may be warranted. The addition of predictive in silico methods for predicting brain permeability may offer the most cost effective, early screen for BBB permeability (94, 95). Better understanding of the structure-activity relationship for both BBB permeability and target affinity is required to build those models and should be a priority over cherry-picking. Trends from the compounds discussed herein point to the integrated approach finding more success.

The focus also needs to shift away from ex vivo studies evaluating initial uptake and clearance in healthy animals. Whole body imaging in AD disease model animals and age-matched controls would better and more rapidly screen potential candidates. Adding dynamic imaging with proper plasma metabolite analysis, like PET radiotracer development, will provide more details into the pharmacokinetics and assist with future iterations of SPECT radiotracer development. New SPECT systems designed for animal imaging have improved spatial resolution and sensitivity such that longitudinal studies in transgenic rodents, particularly rat models (96), should be possible to track sensitivity of 99mTc-based radiotracers to changes in amyloid plaques. Clinical translation could be further accelerated by shifting away from rodents into porcine models (less costly and fewer ethic concerns than non-human primates) (97). Their larger brains can be imaged using clinical SPECT scanners (easy translation to humans) and will permit more granular assessment of radiotracer distribution. Continued confirmation of target specificity through histochemistry and autoradiography in animal and human tissues samples will affirm in vivo results. While there was improvement over time to reporting species and sex of animals, studies with both sexes need to become standard practice (98).

A final concern translating SPECT probes into the clinic could be white matter binding. Not only does this "non-specific" binding decrease signal to noise, the partial volume effect from white matter will confound quantitative analysis. Fluorine-18 radiotracer displayed higher subcortical white matter uptake compared to [<sup>11</sup>C]PiB, potentially due to increased lipophilicity (99). This reiterates the need for optimizing physiochemical properties during radiotracer development beyond affinity. For amyloid imaging, this uptake may be unavoidable as the so-called non-specific binding has been tied to myelin coating on axons (100). It does point to a potential additional application of <sup>99m</sup>Tc-based radiotracers for quantifying white matter lesions. With no clear lead candidate radiotracer approaching clinical translation, it could be premature to worry about optimizing signal to noise ratios, but it bears keeping in mind when designing and developing putative <sup>99m</sup>Tc-based radiotracers.

For SPECT neuroimaging, the 99mTc-based radiotracer development field needs to continue growth to keep pace with other SPECT technological advances. Even in a review extoling the benefits of SPECT/CT (20), there is hardly a mention of neurological applications. Separately, a perspective article on the direction of molecular imaging has no mention of SPECT (21). By changing the perspective of SPECT as an inferior neuroimaging modality, disparate access to leading-edge molecular imaging can be overcome. Indeed, the more widespread and lower cost of SPECT/CT is a wasted resource if only used from the neck down. To move out of the shadow cast by PET for imaging the brain, effective gamma emitting radiotracers are critically needed. By applying some lessons outlined in this review, particularly with remarkable brain penetration of integrated probes, to other targets would expand the radiotracer library for neuro-SPECT (i.e., tau, demyelination, monoamine receptors, etc.). Thus, barriers to accessing nuclear imaging modalities in rural or underdeveloped areas can be removed. Improved access to diagnostic imaging then generates improved health outcomes, further lowering the societal cost burden. This all starts with radiotracer development.

## Author contributions

JH conceived of the project and supervised GT and AK. AK conducted the initial literature review drafting part of her senior undergraduate and as project. GT proofread, completed the thorough literature review, and wrote the manuscript. All authors contributed to the article and approved the submitted version.

## Funding

This work was provided by Western University Internal Research Funds for AK Fourth Year Project, New Frontier's in Research Fund—Exploration (NFRF-E 2020-01021) funds GT training.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## References

1. Wimo A, Guerchet M, Ali G-C, Wu Y-T, Prina AM, Winblad B, et al. The worldwide costs of dementia 2015 and comparisons with 2010. *Alzheimers Dement.* (2017) 13: 1–7. doi: 10.1016/j.jalz.2016.07.150

2. El-Hayek YH, Wiley RE, Khoury CP, Daya RP, Ballard C, Evans AR, et al. Tip of the iceberg: assessing the global socioeconomic costs of Alzheimer's disease and related dementias and strategic implications for stakeholders. *J Alzheimer's Dis.* (2019) 70:323–41. doi: 10.3233/JAD-190426

3. Alzheimer's Disease International. *World Alzheimer Report 2015: The Global Impact of Dementia.* London: Alzheimer's Disease International (2015).

4. Jack CR, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, et al. NIA-AA research framework: toward a biological definition of Alzheimer's disease. *Alzheimer's Dement.* (2018) 14:535–62. doi: 10.1016/j.jalz.2018.02.018

5. Jack CR, Bennett DA, Blennow K, Carrillo MC, Feldman HH, Frisoni GB, et al. A/T/N: an unbiased descriptive classification scheme for Alzheimer disease biomarkers. *Neurology*. (2016) 87:539–47. doi: 10.1212/WNL.00000000002923

6. Barthélemy NR, Horie K, Sato C, Bateman RJ. Blood plasma phosphorylatedtau isoforms track CNS change in Alzheimer's disease. *J Exp Med.* (2020) 217:e20200861. doi: 10.1084/jem.20200861 7. Thijssen EH, Verberk IMW, Kindermans J, Abramian A, Vanbrabant J, Ball AJ, et al. Differential diagnostic performance of a panel of plasma biomarkers for different types of dementia. *Alzheimers Dement (Amst).* (2022) 14:e12285. doi: 10.1002/dad2.12285

8. Koychev I, Hofer M, Friedman N. Correlation of Alzheimer disease neuropathologic staging with amyloid and Tau scintigraphic imaging biomarkers. *J Nucl Med.* (2020) 61:1413–8. doi: 10.2967/jnumed.119.230458

9. Sevigny J, Chiao P, Bussière T, Weinreb PH, Williams L, Maier M, et al. The antibody aducanumab reduces A $\beta$  plaques in Alzheimer's disease. *Nature*. (2016) 537:50–6. doi: 10.1038/nature19323

10. Schneider L. A resurrection of aducanumab for Alzheimer's disease. Lancet Neurol. (2020) 19:111-2. doi: 10.1016/S1474-4422(19)30480-6

11. Positron Emission Tomography in Canada. CADTH.ca (2015). Available online at: https://www.cadth.ca/positron-emission-tomography-canada-2015 (accessed October 15, 2020).

12. SPECT-Related Technologies in Canada. *CADTH.ca*. Available online at: https://www.cadth.ca/spect-related-technologies-canada (accessed October 15, 2020).

13. Honer M, Gobbi L, Martarello L, Comley R. A radioligand development for molecular imaging of the central nervous system with positron emission tomography. *Drug Discovery Today.* (2014) 19:1936–44. doi: 10.1016/j.drudis.2014.08.012

14. Human Health Campus - Database & Statistics. Available online at: https://humanhealth.iaea.org/HHW/DBStatistics/IMAGINE.html (accessed July 22, 2021).

15. Paez D, Giammarile F, Orellana, P. Nuclear medicine: a global perspective. *Clin Transl Imaging.* (2020) 8:51–3. doi: 10.1007/s40336-020-00359-z

16. Cutler CS, Bailey EA, Kumar V, Schwarz SW, Bom HH-S, Hatazawa J, et al. Global issues of radiopharmaceutical access and availability: a nuclear medicine global initiative project. *J Nucl Med.* (2020) 62:jnumed.120.247197. doi: 10.2967/jnumed.120.247197

17. Gunn RN, Slifstein M, Searle GE, Price JC. Quantitative imaging of protein targets in the human brain with PET. *Phys Med Biol.* (2015) 60:R363. doi: 10.1088/0031-9155/60/22/R363

18. Alavi A, Basu S. Planar and SPECT imaging in the era of PET and PET-CT: can it survive the test of time? *Eur J Nucl Med Mol Imaging*. (2008) 35:1554. doi: 10.1007/s00259-008-0813-2

19. Israel O, Pellet O, Biassoni L, De Palma D, Estrada-Lobato E, Gnanasegaran G, et al. Two decades of SPECT/CT PEthe coming of age of a technology: an updated review of literature evidence. *Eur J Nucl Med Mol Imaging.* (2019) 46:1990–2012. doi: 10.1007/s00259-019-04404-6

20. Van den Wyngaert T, Elvas F, De Schepper S, Kennedy JA, Israel O. SPECT/CT: standing on the shoulders of giants, it is time to reach for the sky! *J Nucl Med.* (2020) 61:1284–91. doi: 10.2967/jnumed.119.236943

21. Grimm J, Kiessling F, Pichler BJ. Quo Vadis, molecular imaging? J Nucl Med. (2020) 61:1428–34. doi: 10.2967/jnumed.120.241984

22. Barthel H, Villemagne VL, Drzezga A. Future directions in molecular imaging of neurodegenerative disorders. J Nucl Med. (2022) 63(Suppl. 1):68S-74S. doi: 10.2967/jnumed.121.263202

23. Matsunari I, Miyazaki Y, Kobayashi M, Nishi K, Mizutani A, Kawai K, et al. Performance evaluation of the EXplore SpeCZT preclinical imaging system. *Ann Nucl Med.* (2014) 28:484–97. doi: 10.1007/s12149-014-0828-7

24. Van Audenhaege K, Van Holen R, Vandenberghe S, Vanhove C, Metzler SD, Moore SC. Review of SPECT collimator selection, optimization, and fabrication for clinical and preclinical. *Imaging Med Phys.* (2015) 42:4796-813. doi: 10.1118/1.4927061

25. Sibille L, Chambert B, Alonso S, Barrau C, D'Estanque E, Tabaa YA, et al. Impact of the adaptive statistical iterative reconstruction technique on radiation dose and image quality in bone SPECT/CT. J Nucl Med. (2016) 57:1091– 5. doi: 10.2967/jnumed.115.164772

26. Desmonts C, Bouthiba MA, Enilorac B, Nganoa C, Agostini D, Aide N. Evaluation of a new multipurpose whole-body CzT-based camera: comparison with a dual-head anger camera and first clinical images. *EJNMMI Physics*. (2020) 7:18. doi: 10.1186/s40658-020-0284-5

27. De Schepper S, Ritt P, Van den Wyngaert T, Kuwert T. Quantitative radionuclide imaging of bone metastases. *Q J Nucl Med Mol Imaging*. (2019) 63:129–35. doi: 10.23736/s1824-4785.19.03204-7

28. Imbert L, Poussier S, Franken PR, Songy B, Verger A, Morel O, et al. Compared performance of high-sensitivity cameras dedicated to myocardial perfusion SPECT: a comprehensive analysis of phantom and human images. *J Nucl Med.* (2012) 53:1897–903. doi: 10.2967/jnumed.112.107417

29. Hlatky MA, Shilane D, Hachamovitch R, DiCarli MF. Economic outcomes in the study of myocardial perfusion and coronary anatomy imaging roles in coronary artery disease registry: the SPARC study. *J Am Coll Cardiol.* (2014) 63:1002–8. doi: 10.1016/j.jacc.2013.11.038

30. Henderson TA. The diagnosis and evaluation of dementia and mild cognitive impairment with emphasis on SPECT perfusion neuroimaging. *CNS Spectr.* (2012) 17:176–206. doi: 10.1017/S1092852912000636

31. Valotassiou V, Angelidis G, Psimadas D, Tsougos I, Georgoulias P. In the era of FDG PET, is it time for brain perfusion SPECT to gain a place in Alzheimer's disease imaging biomarkers? *Eur J Nucl Med Mol Imaging.* (2021) 48:969–71. doi: 10.1007/s00259-020-05077-2

32. Bega D, Kuo PH, Chalkidou A, Grzeda MT, Macmillan T, Brand C, et al. Clinical utility of DaTscan in patients with suspected Parkinsonian syndrome: a systematic review and meta-analysis. *NPJ Parkinsons Dis.* (2021) 7:1–8. doi: 10.1038/s41531-021-00185-8

33. Ferrando R, Damian A. Brain SPECT as a biomarker of neurodegeneration in dementia in the era of molecular imaging: still a valid option? *Front Neurol.* (2021) 12:629442. doi: 10.3389/fneur.2021.629442

34. Kung HF, Kung M-P, Wey S-P, Lin K-J, Yen T-C. Clinical acceptance of a molecular imaging agent: a long march with [<sup>99m</sup>Tc]TRODAT. *Nucl Med Biol.* (2007). 34 (7), 787–789. doi: 10.1016/j.nucmedbio.2007.03.010

35. Shih MC, Franco de Andrade LA, Amaro E, Jr., Felicio AC, Ferraz HB, Wagner J, et al. A higher nigrostriatal dopamine neuron loss in early than late onset Parkinson's disease?oa [<sup>99m</sup>Tc]-TRODAT-1 SPECT. *Study Mov Disord.* (2007) 22:863–6. doi: 10.1002/mds.21315

36. Fallahi B, Esmaeili A, Beiki D, Oveisgharan S, Noorollahi-Moghaddam H, Erfani M, et al. Evaluation of <sup>99m</sup>Tc-TRODAT-1 SPECT in the diagnosis of Parkinson's disease versus other progressive movement disorders. *Ann Nucl Med.* (2016) 30:153–62. doi: 10.1007/s12149-015-1042-y

37. Hayne DJ, Lim S, Donnelly PS. Metal complexes designed to bind to amyloid- $\beta$  for the diagnosis and treatment of Alzheimer's disease. *Chem Soc Rev.* (2014) 43:6701–15. doi: 10.1039/C4CS00026A

38. Chen K, Cui M. Recent progress in the development of metal complexes as βsamyloid imaging probes in the brain. *Med Chem Commun.* (2017) 8:1393–407. doi: 10.1039/C7MD00064B

39. Sedgwick A, Brewster J, Harvey P, Iovan DA, Smith G, He X-P, et al. Metalbased imaging agents: progress towards interrogating neurodegenerative disease. *Chem Soc Rev.* (2020) 49:2886–915. doi: 10.1039/C8CS00986D

40. Gomes LMF, Bataglioli JC, Storr T. Metal complexes that bind to the amyloid- $\beta$  peptide of relevance to Alzheimer's disease coordination. Chem Rev. (2020) 412:213255. doi: 10.1016/j.ccr.2020.213255

41. Molavipordanjani S, Emami S, Hosseinimehr SJ. <sup>99m</sup>Tc-labeled small molecules for diagnosis of Alzheimer's disease: past, recent and future perspectives. *CMC*. (2019) 26:2166–89. doi: 10.2174/0929867325666180410104023

42. Zhen W, Han H, Anguiano M, Lemere CA, Cho C-G, Lansbury PT. Synthesis and amyloid binding properties of rhenium complexes: preliminary progress toward a reagent for SPECT IMAGING of Alzheimer's disease brain. *J Med Chem.* (1999) 42:2805–15. doi: 10.1021/jm990103w

43. Kaur A, New EJ, Sunde M. Strategies for the molecular imaging of amyloid and the value of a multimodal approach. ACS Sens. (2020) 5:2268-82. doi: 10.1021/acssensors.0c01101

44. Adak S, Bhalla R, Raj KKV, Mandal S, Pickett R, Luthra SK. Radiotracers for SPECT imaging: current scenario and future prospects. *Radiochimica Acta*. (2012) 100:95–107. doi: 10.1524/ract.2011.1891

45. Kung HF, Kung M-P, Choi SR. Radiopharmaceuticals for single-photon emission computed tomography brain imaging seminars in nuclear. *Medicine*. (2003) 33:2–13. doi: 10.1053/snuc.2003.127296

46. Akbar MU, Ahmad MR, Shaheen A, Mushtaq S. A review on evaluation of technetium-99m labeled radiopharmaceuticals. *J Radioanal Nucl Chem.* (2016) 310:477–93. doi: 10.1007/s10967-016-5019-7

47. Eckelman W. Review of new clinical applications of radiometal labeled radiopharmaceuticals. *Nucl Med Biol.* (2011) 38:613–6. doi: 10.1016/j.nucmedbio.2010.12.009

48. Duatti A. Review on <sup>99m</sup>Tc radiopharmaceuticals with emphasis on new advancements. *Nucl Med Biol.* (2021) 92:202– 16. doi: 10.1016/j.nucmedbio.2020.05.005

49. Han H, Cho C-G, Lansbury PT. Technetium complexes for the quantitation of brain amyloid. J Am Chem Soc. (1996) 118:4506–7. doi: 10.1021/ja960207l

50. Ashburn TT, Han H, McGuinness BF, Lansbury PT. Amyloid probes based on congo red distinguish between fibrils comprising different peptides. *Chem Biol.* (1996) 3:351–8. doi: 10.1016/S1074-5521(96)90118-0

51. Dezutter NA, Dom RJ, de Groot TJ, Bormans GM, Verbruggen A. M <sup>99m</sup>Tc-MAMA-chrysamine G, a probe for beta-amyloid protein of Alzheimer's disease. *Eur J Nucl Med.* (1999) 26:1392–9. doi: 10.1007/s002590050470

52. Barichello T, Collodel A, Hasbun R, Morales R. An overview of the bloodbrain barrier. In: *Blood-Brain Barrier - A Neuromethods Book Series, Vol. 142,* New York, NY: Springer New York (2018). p. 1–8.

53. Banerjee SR, Foss CA, Castanares M, Mease RC, Byun Y, Fox JJ, et al. Synthesis and Evaluation of Technetium-99m- and Rhenium-Labeled Inhibitors of the Prostate-Specific Membrane Antigen (PSMA). ACS Publications. Available online at: https://pubs.acs.org/doi/pdf/10.1021/jm800111u (accessed May 3, 2022).

54. Serdons K, Verduyckt T, Cleynhens J, Terwinghe C, Mortelmans L, Bormans G, et al. Synthesis and evaluation of a  $^{99m}$ Tc-BAT-phenylbenzothiazole conjugate as a potential *in vivo* tracer for visualization of amyloid  $\beta$ . *Bioorg Med Chem Lett.* (2007) 17:6086–90. doi: 10.1016/j.bmcl.2007.09.055

55. Chen X, Yu P, Zhang L, Liu B. Synthesis and biological evaluation of  $^{99m}$ Tc, re-monoamine-monoamide conjugated to 2-(4-aminophenyl)benzothiazole as potential probes for  $\beta$ mamyloid plaques in the brain. *Bioorg Med Chem Lett.* (2008) 18:1442–5. doi: 10.1016/j.bmcl.2007.12.071

56. Chen X. QSAR and primary docking studies of trans-stilbene (TSB) series of imaging agents for  $\beta$ -amyloid plaques. *J Mol Struct THEOCHEM*. (2006) 763:83–9. doi: 10.1016/j.theochem.2006.01.028

57. Lin K-S, Debnath ML, Mathis CA, Klunk WE. Synthesis and  $\beta$ -amyloid binding properties of rhenium 2-phenylbenzothiazoles. *Bioorg Med Chem Lett.* (2009) 19:2258–62. doi: 10.1016/j.bmcl.2009.02.096

58. Jia J, Cui M, Dai J, Liu B. 2-Phenylbenzothiazole conjugated with cyclopentadienyl tricarbonyl [CpM(CO)3] (M = Re,  $^{99m}Tc$ ) complexes as potential imaging probes for  $\beta$ -amyloid plaques. Dalton Trans. (2015) 44:6406–15. doi: 10.1039/C5DT00023H

59. Jia J, Zhou K, Dai J, Liu B, Cui, M. 2-arylbenzothiazoles labeled with [CpRe/^99mTc(CO)3] and evaluated as  $\beta$ -amyloid imaging probes. Eur J Med Chem. (2016) 124:763–72. doi: 10.1016/j.ejmech.2016.09.001

60. Zhang X, Yu P, Yang Y, Hou Y, Peng C, Liang Z, et al. <sup>99m</sup>Tc-labeled 2-arylbenzothiazoles: A $\beta$  imaging probes with favorable brain pharmacokinetics for single-photon emission computed tomography. *Bioconjugate Chem.* (2016) 27:2493–504. doi: 10.1021/acs.bioconjchem.6b00444

61. Zhang X, Hou Y, Peng C, Wang C, Wang X, Liang Z, et al. Oligoethyleneoxymodified <sup>99m</sup>Tc-labeled  $\beta$ -amyloid imaging probes with improved brain pharmacokinetics for single-photon emission computed tomography. *J Med Chem.* (2018) 61:1330–9. doi: 10.1021/acs.jmedchem.7b01834

62. Serdons K, Verduyckt T, Cleynhens J, Bormans G, Verbruggen A. Development of <sup>99m</sup>Tc-thioflavin-t derivatives for detection of systemic amyloidosis. *J Label Comp Radiopharm.* (2008) 51:357–67. doi: 10.1002/jlcr.1536

63. Decristoforo C, Mather SJ, Cholewinski W, Donnemiller E, Riccabona G, Moncayo, R. <sup>99m</sup>Tc-EDDA/HYNIC-TOC: a new <sup>99m</sup>Tc-labelled radiopharmaceutical for imaging somatostatin receptor-positive tumours: first clinical results and intra-patient comparison with <sup>111</sup>In-labelled octreotide derivatives. *Eur J Nucl Med.* (2000) 27:1318–25. doi: 10.1007/s002590000289

64. Jia J, Cui M, Dai J, Wang X, Ding Y-S, Jia H, et al.  $^{99m}$ Tc-labeled benzothiazole and stilbene derivatives as imaging agents for A $\beta$  plaques in cerebral amyloid angiopathy. *Med Chem Commun.* (2014) 5:153–8. doi: 10.1039/C3MD00195D

65. Jia J, Cui M, Dai J, Liu B. <sup>99m</sup>Tc(CO)<sub>3</sub>-labeled benzothiazole derivatives preferentially bind cerebrovascular amyloid: potential use as imaging agents for cerebral amyloid angiopathy. *Mol Pharm.* (2015) 12:2937–46. doi: 10.1021/acs.molpharmaceut.5b00209

66. Ono M, Fuchi Y, Fuchigami T, Kobashi N, Kimura H, Haratake M, et al. Novel benzofurans with  $^{99m}$ Tc complexes as probes for imaging cerebral  $\beta$ -amyloid plaques. ACS Med Chem Lett. (2010) 1:443–7. doi: 10.1021/ml100140d

67. Cheng Y, Ono M, Kimura H, Ueda M, Saji H. Technetium-99m labeled pyridyl benzofuran derivatives as single photon emission computed tomography imaging probes for  $\beta$  amyloid plaques in Alzheimer's brains. *J Med Chem.* (2012) 55:2279–86. doi: 10.1021/jm201513c

68. Ono M, Cheng Y, Kimura H, Cui M, Kagawa S, Nishii R, et al. Novel <sup>18</sup>Flabeled benzofuran derivatives with improved properties for positron emission tomography (PET) imaging of  $\beta$ -amyloid plaques in Alzheimer's brains. J Med Chem. (2011) 54:2971–9. doi: 10.1021/jm200057u

69. Oya S, Plössl K, Kung M-P, Stevenson DA, Kung HF. Small and neutral TcvO BAT, bisaminoethanethiol (N2S2) complexes for developing new brain imaging agents. *Nuclear Med Biol.* (1998) 25:135–40. doi: 10.1016/S0969-8051(97)00153-4

70. Choi SR, Golding G, Zhuang Z, Zhang W, Lim N, Hefti F, et al. Preclinical properties of 18F-AV-45: a PET agent for Ao plaques in the brain. *J Nucl Med.* (2009) 50:1887–94. doi: 10.2967/jnumed.109.065284

71. Wang X, Cui M, Yu P, Li Z, Yang Y, Jia H, et al. Synthesis and biological evaluation of novel technetium-99m labeled phenylbenzoxazole derivatives as potential imaging probes for  $\beta$ -amyloid plaques in brain. *Bioorg Med Chem Lett.* (2012) 22:4327–31. doi: 10.1016/j.bmcl.2012.05.010

72. Wang X, Cui M, Jia J, Liu, B. <sup>99m</sup>Tc-labeled-2-arylbenzoxazole derivatives as potential A $\beta$  imaging probes for single-photon emission computed tomography. *Eur J Med Chem.* (2015) 89:331–9. doi: 10.1016/j.ejmech.2014.10.046

73. Ono M, Ikeoka R, Watanabe H, Kimura H, Fuchigami T, Haratake M, et al.  $^{99m}Tc/Re$  complexes based on flavone and aurone as SPECT probes for imaging cerebral  $\beta$ -amyloid plaques. Bioorg Med Chem Lett. (2010) 20:5743–8. doi: 10.1016/j.bmcl.2010.08.004

74. Yang Y, Zhu L, Cui M, Tang R, Zhang H. Preparation of classical Re/ $^{99m}Tc(CO)_3^+$  and novel  $^{99m}Tc(CO)_2(NO)_2^+$  cores complexed with flavonol derivatives and their binding characteristics for A\beta(1-40) aggregates. Bioorg Med Chem Lett. (2010) 20:5337–44. doi: 10.1016/j.bmcl.2010. 04.026

75. Cui M, Tang R, Li Z, Ren H, Liu, B.  $^{99m}\text{Tc-}$  and re-labeled 6-dialkylamino-2-naphthylethylidene derivatives as imaging probes for  $\beta$ -amyloid plaques. Bioorg Med Chem Lett. (2011) 21:1064–8. doi: 10.1016/j.bmcl.2010.11.096

76. likuni S, Ono M, Tanimura K, Watanabe H, Yoshimura M, Saji H, et al. Synthesis and biological evaluation of novel technetium-99m-labeled phenylquinoxaline derivatives as single photon emission computed tomography imaging probes targeting βcamyloid plaques in Alzheimer's disease. *RSC Adv.* (2017) 7:20582–90. doi: 10.1039/C6RA28395K

77. Molavipordanjani S, Emami S, Mardanshahi A, Talebpour Amiri F, Noaparast Z. Hosseinimehr, et al. J novel <sup>99m</sup>Tc-2-arylimidazo[2,1-b]benzothiazole derivatives as SPECT imaging agents for amyloid-m plaques. *Eur J Med Chem.* (2019) 175:149–61. doi: 10.1016/j.ejmech.2019.04.069

78. Hayne DJ, North AJ, Fodero-Tavoletti M, White JM, Hung LW, Rigopoulos A, et al. Rhenium and technetium complexes that bind to amyloid- $\beta$  plaques. *Dalton Trans.* (2015) 44:4933–44. doi: 10.1039/C4DT02969K

79. Ono M, Ikeoka R, Watanabe H, Kimura H, Fuchigami T, Haratake M, et al. Synthesis and evaluation of novel chalcone derivatives with  $^{99m}$ Tc/Re complexes as potential probes for detection of  $\beta$ -amyloid plaques. ACS Chem Neurosci. (2010) 1:598–607. doi: 10.1021/cn100042d

80. Yang Y, Cui M, Jin B, Wang X, Li Z, Yu P, et al.  $^{99m}$ Tc-labeled dibenzylideneacetone derivatives as potential SPECT probes for *in vivo* imaging of β-amyloid plaque. *Eur J Med Chem.* (2013) 64:90–8. doi: 10.1016/j.ejmech.2013.03.057

81. Cui M, Ono M, Kimura H, Liu B, Saji H. Synthesis and structure–affinity relationships of novel dibenzylideneacetone derivatives as probes for  $\beta$ namyloid plaques. *J Med Chem.* (2011) 54:2225–40. doi: 10.1021/jm101404k

82. Johansson SMC, Arnberg N, Elofsson M, Wadell G, Kihlberg J. Multivalent HSA conjugates of 3'-sialyllactose are potent inhibitors of adenoviral cell attachment and infection. *Chembiochem.* (2005) 6:358–64. doi: 10.1002/cbic.200400227

83. Iikuni S, Ono M, Watanabe H, Matsumura K, Yoshimura M, Harada N, et al. Enhancement of binding affinity for amyloid aggregates by multivalent interactions of <sup>99m</sup>Tc-hydroxamamide complexes. *Mol Pharmaceutics*. (2014) 11:1132–9. doi: 10.1021/mp400499y

84. Iikuni S, Ono M, Watanabe H, Matsumura K, Yoshimura M, Kimura H, et al. Imaging of cerebral amyloid angiopathy with bivalent <sup>99m</sup>Tc-hydroxamamide complexes. *Sci Rep.* (2016) 6:25990. doi: 10.1038/srep25990

85. Iikuni S, Ono M, Watanabe H, Yoshimura M, Ishibashi-Ueda H, Ihara M, et al. Novel bivalent <sup>99m</sup>Tc-complex with N-methyl-substituted hydroxamamide as probe for imaging of cerebral amyloid angiopathy. *PLoS ONE.* (2016) 11:e0163969. doi: 10.1371/journal.pone.0163969

86. Thipyapong K, Uehara T, Tooyama Y, Braband H, Alberto R, Arano Y, et al. Insight into technetium amidoxime complex: oxo technetium(V) complex of Nsubstituted benzamidoxime as new basic structure for molecular imaging. *Inorg Chem.* (2011) 50:992–8. doi: 10.1021/ic101714q

87. Fletcher SP, Noor A, Hickey JL, McLean CA, White JM, Donnelly PS. Rhenium and technetium complexes of thioamide derivatives of pyridylhydrazine that bind to amyloid- $\beta$  plaques. J Biol Inorg Chem. (2018) 23:1139–51. doi: 10.1007/s00775-018-1590-4

88. Pan J, Mason NS, Debnath ML, Mathis CA, Klunk WE, Lin K-S. Design, synthesis and structure-activity relationship of rhenium 2-arylbenzothiazoles as  $\beta$ samyloid plaque binding agents. *Bioorg Med Chem Lett.* (2013) 23:1720-6. doi: 10.1016/j.bmcl.2013.01.068

89. Morais M, Paulo A, Gano L, Santos I, Correia JDG. Target-specific Tc(CO)<sub>3</sub>-complexes for *in vivo* imaging. *J Organomet Chem.* (2013) 744:125-39. doi: 10.1016/j.jorganchem.2013.05.050

90. Lengacher R, Alberto R. Bioorganometallics: <sup>99m</sup>Tc cytectrenes, syntheses and applications in nuclear medicine coordination. *Chem Rev.* (2021) 437:213869. doi: 10.1016/j.ccr.2021.213869

91. Kiritsis C, Mavroidi B, Shegani A, Palamaris L, Loudos G, Sagnou M, et al. 2-(4'-Aminophenyl)benzothiazole labeled with  $^{99m}$ Tc-cyclopentadienyl for imaging  $\beta$ -amyloid plaques. ACS Med Chem Lett. (2017) 8:1089–92. doi: 10.1021/acsmedchemlett.7b00294

92. Sagnou M, Mavroidi B, Shegani A, Paravatou-Petsotas M, Raptopoulou C, Psycharis V, et al. Remarkable brain penetration of cyclopentadienyl  $M(CO)_3$  ( $M = ^{99m}$ Tc, Re) derivatives of benzothiazole and benzimidazole paves the way for their application as diagnostic, with single-photon-emission computed tomography (SPECT), and therapeutic agents for Alzheimer's disease. *J Med Chem.* (2019) 62:2638–50. doi: 10.1021/acs.jmedchem.8b01949

93. Kung HF, Choi SR, Qu W, Zhang W, Skovronsky D.  $^{18}F$  stilbenes and styrylpyridines for PET Imaging of A $\beta$  plaques in alzheimer's disease: a miniperspective. J Med Chem. (2010) 53:933–41 doi: 10.1021/jm901039z

94. Wager TT, Hou X, Verhoest PR, Villalobos A. Moving beyond rules: the development of a central nervous system multiparameter optimization (CNS MPO) approach to enable alignment of druglike properties. *ACS Chem Neurosci.* (2010) 1:435–49. doi: 10.1021/cn100008c

95. Jackson IM, Webb EW, Scott PJH, James ML. *In silico* approaches for addressing challenges in CNS radiopharmaceutical design. *ACS Chem Neurosci.* (2022) 13:1675–83. doi: 10.1021/acschemneuro.2c00269

96. Weishaupt N, Liu Q, Shin S, Singh R, Agca Y, Agca C, et al. APP21 transgenic rats develop age-dependent cognitive impairment and microglia accumulation within white matter tracts. *J Neuroinflam.* (2018) 15:241. doi: 10.1186/s12974-018-1273-7

97. Hoffe B, Holahan MR. The use of pigs as a translational model for studying neurodegenerative diseases. *Front Physiol.* (2019) 10:838. doi: 10.3389/fphys.2019.00838

98. Clayton JA. Studying both sexes: a guiding principle for biomedicine. *FASEB J.* (2016) 30:519–24. doi: 10.1096/fj.15-279554

99. Heurling K, Buckley C, Vandenberghe R, Laere KV, Lubberink M. Separation of  $\beta$ -amyloid binding and white matter uptake of <sup>18</sup>F-flutemetamol using spectral analysis. *Am J Nucl Med Mol Imaging.* (2015) 5:515–26.

100. Zeydan B, Schwarz CG, Przybelski SA, Lesnick TG, Kremers WK, et al. Comparison of <sup>11</sup>C-pittsburgh compound-B and <sup>18</sup>F-flutemetamol white matter binding in PET. *J Nucl Med.* (2021) 63:1239–44. doi: 10.2967/jnumed.121. 263281