

Resistin as a Systemic Inflammation-Related Biomarker for Sarcopenia in Patients With Chronic Obstructive Pulmonary Disease

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Background: Sarcopenia is common in patients with chronic obstructive pulmonary disease (COPD) and is mainly caused by systemic inflammation. Resistin acts as a proinflammatory cytokine and is involved in the activation of multiple inflammatory signaling pathways. The aim of this study was to determine the relationship between resistin levels and systemic inflammation and to assess the clinical value of circulating resistin for sarcopenia in patients with COPD.

Methods: In this prospective observational study, we enrolled 235 patients with COPD who were divided into development and validation sets. The definition of sarcopenia followed the guidelines from the Asian Working Group for Sarcopenia. Serum concentrations of resistin and TNF- α were measured using an enzyme-linked immunosorbent assay (ELISA).

Results: In this study, higher serum resistin levels were significantly associated with lower skeletal muscle mass and muscular strength. The serum resistin levels in patients with sarcopenia were significantly higher than those in patients without sarcopenia. The serum resistin level had positive correlations with the serum TNF- α level (r = 0.250, p = 0.007). The predictive efficacy of the serum resistin level (AUC: 0.828) for sarcopenia was superior to that of the serum TNF- α level (AUC: 0.621). The cutoff point (7.138 ng/ml) for the serum resistin level was validated in the validation set (AUC: 0.818).

Conclusions: Serum resistin levels were associated with systemic inflammation and can be used accurately and easily to predict sarcopenia in patients with COPD.

Keywords: chronic obstructive pulmonary disease, sarcopenia, resistin, TNF-a, biomarker

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INTRODUCTION

Chronic obstructive pulmonary disease (COPD) is a chronic airway disease characterized by persistent respiratory symptoms and limited airflow that is often accompanied by multiple comorbidities involving the cardiovascular system, digestive system, hematologic system, musculoskeletal system, etc. (1, 2). Among the comorbidities, sarcopenia is associated with a poor prognosis and increased hospitalization and mortality rates (3). Patients with COPD have an increased risk of developing sarcopenia, with prevalence estimates ranging from 15 (4) to 55% (5). Sarcopenia had a consistently negative impact on a range of COPD-related clinical outcomes, including exercise capacity, balance, quadriceps, and handgrip strength, and physical activity levels (6-9). It was also associated with increased symptom burden and poorer quality of life. Early identification and intervention of sarcopenia is important to improve the quality of life and prognosis of patients with COPD. However, the diagnosis of sarcopenia requires simultaneous assessment of muscle mass, muscle strength, and physical function (10, 11). Therefore, it is time-consuming and difficult to assess sarcopenia in all COPD patients in clinical practice.

Inflammation and oxidative stress play a key role in the decline in respiratory function in COPD (12). More commonly, in patients with COPD, cigarette smoking-induced inflammation and oxidative stress in the lungs "spill over" into the circulation, manifesting as systemic inflammation (2). Persistent systemic inflammation is associated with an accelerated decline in lung function and is exacerbated during exacerbation episodes. Systemic inflammation is now a recognized risk factor for other complications commonly seen in patients with COPD (13). Several studies (14, 15) have suggested a possible association between systemic inflammation and comorbid sarcopenia in patients with COPD. TNF- α are cytokines related to chronic inflammatory metabolic disorders that can result in a decrease in protein synthesis in skeletal muscle and associated with a decrease in muscle mass and strength (16). Injections of exogenous TNF-a decreased muscle mass and its ability to regenerate in mouse model (17). Previous study suggested that sarcopenia is associated with increased TNF- α levels (18). Systemic ablation of TNF- α prevented sarcopenia (19). These results suggest that TNF- α may play an important role in the development of sarcopenia. Previous study in COPD shown that serum TNF- α levels are significantly associated with grip strength and skeletal muscle mass, which are important determinants of sarcopenia in patients with stable COPD (20, 21). These results suggest that systemic inflammation-related factors may be important biomarkers for patients with COPD in predicting sarcopenia.

Resistin was originally described as an adipokine causing insulin resistance in mice that is produced by white adipose tissue (22). In humans, resistin also acts as a proinflammatory cytokine that is produced mainly by cells of the inflammatory system, such as monocytes, macrophages, and neutrophils (23). Previous studies have shown that resistin is an endogenous ligand for Toll-like receptor 4 (TLR4) and is involved in the activation of multiple inflammatory signaling pathways (24, 25). Resistin could disrupts myotube and nuclear fusion by activating the classical NF- κ B pathway (26). In additional, resistin inhibits myogenic myoblast differentiation (27). Resistin has been shown to impair insulin signaling (28), which is an important regulator mediating muscle growth and regeneration (29). These results suggest that resistin may play an important role in the development process of sarcopenia. A previous study (30) indicated that the mean resistin levels in patients with sarcopenia were 2-fold higher than those in healthy controls. In addition, significant negative correlations have been shown between the forced expiratory volume in 1 second (FEV₁) and resistin levels in patients with OOPD (30). The serum resistin level is associated with osteoporosis in patients with COPD (31). The accuracy of circulating resistin levels in predicting sarcopenia in patients with COPD has not been established.

The aim of this study was to assess the clinical value of circulating resistin in prospectively screening for sarcopenia in patients with COPD and to determine the cutoff value for use in clinical practice.

MATERIALS AND METHODS

Study Design and Subjects

This is a cross-sectional analysis that starts from a longitudinal study and was conducted at the First Hospital of China Medical University and the First Hospital of Dalian Medical University from August 2018 to December 2019. Patients with COPD who met the Global Initiative for Chronic Obstructive Lung Disease (GOLD) criteria were eligible for inclusion (32), and the predicted post-bronchodilator FEV₁% was divided into \geq 80%, 50-79%, 30-49%, and <30% according to GOLD stage. The exclusion criteria were as follows: acute exacerbation within the past 1 month; presence of severe cardiovascular disease, active lung disease, patients with edema or fluid overload, etc.; long-term systemic hormone therapy; and inability to read or understand the informed consent. The Medical Ethics Committee of the First Hospital of China Medical University approved this study (No. 2018-144-2), and all patients provided written informed consent. We included all patients with stable chronic obstructive pulmonary disease in the outpatient clinic who met the inclusion and exclusion criteria and who signed an informed consent during August 2018 to December 2019. The estimated total sample size used for univariate and multivariate logistic analyses to explore the clinical factors associated with sarcopenia in patients with COPD followed previous studies (33, 34). It was used for univariate and multivariate logistic analyses to explore the clinical factors associated with sarcopenia in patients with COPD. Given the prevalence of sarcopenia in patients with stable COPD of 25% (20) and 5 covariates in the logistic model, the sample size was estimated to be 200 according to the equation (N = 10 k/p) (33, 34). The final sample size was composed of 235 people. Diabetes was present in 43 (18.30%) patients. Hypertension was present in 56 (23.83%) patients. To obtain the desired level of statistical power for evaluating the accuracy of the diagnostic test, the minimal sample size required was calculated based on the estimated sensitivity and

specificity (35):

$$N_{Se} = \frac{Z\frac{\alpha^2}{2} \times \hat{Se} \times (1 - \hat{Se})}{d^2 \times Prev}; N_{Sp} = \frac{Z\frac{\alpha^2}{2} \times \hat{Sp} \times (1 - \hat{Sp})}{d^2 \times (1 - Prev)};$$

We estimated the sensitivity (Se) and specificity (Sp) for the serum resistin level to diagnose sarcopenia as 90 and 70%, respectively, according to our preliminary study; we estimated the prevalence of sarcopenia in patients with COPD as \sim 30% according to data from this study. For a maximum marginal error of the estimate not exceeding 10% with a 95% confidence level. The total required sample size was 115 patients which calculated using sensitivity. And, the total required sample size was 115 patients which calculated using specificity. Therefore, the total required sample size was 115 patients. A total of 117 patients with COPD who were recruited from the First Hospital of China Medical University were enrolled in the development set to assess the clinical value of resistin and determine the cutoff values. Another 118 patients with COPD who were recruited from the First Hospital of Dalian Medical University were enrolled in the validation set. And, the number of participants in development set and validation set is adequate to know the resistin cutoff point.

Data Collection and Measurements

Age, sex, smoking status, and smoking pack-years were recorded. Spirometry measurements were performed as recommended by the American Thoracic Society and the European Respiratory Society guidelines using a Jaeger MasterScreen system (Viasys Healthcare GmbH, Hoechberg, Germany). Dyspnea and health status were assessed based on the Chinese version of the mMRC dyspnea score (36) and the CAT (37). Exercise capacity was determined using the 6-min walking test following the 2002 American Thoracic Society (ATS) guidelines (38). In short, a closed, long, and straight 30-m corridor was selected indoors, and the patient was told to walk as far as possible. Standard language was used to encourage the patient. After 6 min, the patient was told "time is up" and asked to stop walking. The testing personnel recorded the distance traveled in meters. Venous blood was obtained in the fasted state. Blood samples were collected into serum separator tubes, left to stand for 2 h at room temperature to clot, and then centrifuged at 3,000 rpm for 15 min at room temperature. The supernatant liquid was obtained and kept in a -80° C refrigerator. TNF- α was assessed by high-sensitivity ELISA kits (#HSTA00E; R&D, Minneapolis, MN, USA) following the manufacturer's instructions. The resistin level was determined using a human resistin Quantikine ELISA kit (#DRSN00; R&D, Minneapolis, MN, USA) according to the manufacturer's instructions.

Measurement Thickness and Cross-Sectional Area of the Rectus Femoris

Measurement of quadriceps rectus femoris thickness and crosssectional area followed previous studies (39, 40). Grayscale ultrasound was used with a 4- to 15-MHz linear-array transducer (SuperSonic Imagine, Aix-en-Provence, France). Two physicians from ultrasonography performed the ultrasound examinations.

The patient requested refrain from strenuous exercise for 72 h prior to the test, rested quietly for 30 min and then lay on the back on the operating bed to relax the muscles of the whole body. The researchers set up a bracket to fix the ultrasound probe to reduce muscle deformation due to external forces and placed the ultrasound probe perpendicular to the patient's dominant leg. The transducer was positioned perpendicular to the long axis of the dominant leg (precisely at 3/5 of the distance from the anterior superior iliac spine to the superior patellar border). The scanning depth was set to where the femur could be discerned for orientation. Gentle contraction-relaxation maneuveres were employed to delineate muscle septa prior to image acquisition. RF_{thick} and RF_{csa} were calculated after the inner echogenic line of the rectus femoris was outlined by a movable cursor on a frozen image. RF_{thick} and RF_{csa} were taken as averages of three consecutive measurements within 10%.

Assessment of Sarcopenia

The definition of sarcopenia followed the guidelines from the AWGS (11), as follows: low muscle mass [bioelectrical impedance (males, <7.0 kg/m²; females, <5.7 kg/m²)]; low muscle strength [handgrip strength (males, <28 kg; females, <18 kg)]; and/or poor physical performance [5-time chair stand test (5STS), \geq 12 s]. Muscle mass was evaluated using bioelectrical impedance analysis [BIA] (InBody770; InBody, Seoul, South Korea). Muscle strength was assessed by handgrip strength using a JAMAR[®] Plus⁺ hand dynamometer (Sammons Preston, Bolingbrook, IL, USA). The 5STS was performed according to the procedure described in a previous study (41). Specifically, the patient sits on a chair (48 cm in height) without armrests, with feet on the ground, the back supported by the back of the chair, and hands folded in front of the chest. After hearing the test start command, the participant was asked to complete 5 standing and sitting movements as fast as possible; the time was recorded. During the test, the patient was instructed to cross the hands on the chest without separation. The knee joints were an equal distance from the ground when in the standing position. The participant was given verbal encouragement during the test. The test was performed 3 times, with 1-min rest intervals. The average of 3 test times was taken as the result.

Statistical Analyses

Statistical analyses were performed using SPSS 13.0 software (IBM, Armonk, NY, USA). To check the distribution status of serum resistin levels, the Shapiro–Wilk test, histograms, and Q-Q (quantile–quantile) plots were used. Pearson's correlation coefficient was calculated to determine the association between continuous variables. Univariate and multivariate logistic analyses were used to explore the clinical factors associated with sarcopenia. Results were presented as OR and 95%CI (42). *P*-values < 0.05 were considered statistically significant. Variables with a *P*-value of <0.05 in the univariate logistic analysis were included in the multivariate logistic to identify corresponding risk factors. For the continuous variables, differences between two groups were assessed using a *t*-test (normally distributed data) or the Mann–Whitney test (non-normal distribution). *P*-values < 0.05 were considered statistically significant.

Variable	Development set	Validation set	P-value	
	(<i>n</i> = 117)	(<i>n</i> = 118)		
Demographics				
Age, years	64.9 ± 12.0	63.9 ± 9.2	0.110	
Sex, m/f	83/34	77/41	0.349	
Pulmonary function	n			
FEV1, L	1.6 ± 0.6	1.5 ± 0.5	0.370	
FEV1, %predicted	59.8 ± 21.0	56.2 ± 20.7	0.368	
FVC, L	2.8 ± 0.9	2.7 ± 0.8	0.452	
FVC, % predicted	82.5 ± 23.7	84.5 ± 20.0	0.700	
FEV1/FVC, %	55.2 ± 9.9	53.1 ± 11.1	0.232	
GOLD stage			0.986	
1	19	21		
2	59	57		
3	30	31		
4	9	9		
Physical function				
6MWD, m	369.0 ± 74.2	359.1 ± 78.1	0.370	
5STS, s	7.8 ± 3.3	7.4 ± 2.3	0.624	
Body composition				
BMI, kg/m ²	23.9 ± 3.9	23.6 ± 3.8	0.532	
Body fat (%)	28.9 ± 6.3	29.0 ± 6.4	0.907	
FFM (kg)	47.0 ± 9.3	45.0 ± 9.1	0.148	
FFMI (kg/m ²)	16.84 ± 2.4	16.7 ± 2.3	0.750	
SMM (kg)	17.7 ± 4.4	15.8 ± 4.6	0.137	
SMMI (kg/m ²)	6.4 ± 1.1	5.9 ± 1.4	0.143	
HGS (kg)	26.6 ± 8.1	27.0 ± 1.1	0.787	

TABLE 1 | Baseline characteristics of subjects.

RESULTS

Baseline Characteristics

The baseline characteristics of patients with COPD are shown in **Table 1**.

Relationships of the Serum Resistin Level With the Clinical Features of Patients With COPD

First, the results (**Supplementary Figure 1**) from histograms, Q-Q plots, and Shapiro–Wilk's test suggested that serum resistin expression data obeyed a normal distribution (p = 0.429). Next, to verify the clinical value of resistin, the serum levels of resistin were detected using ELISA. First, the expression of resistin increased in patients with GOLD stages 1 and 2 compared to patients with GOLD stages 3 and 4 (**Figure 1A**). Next, we analyzed the relationship between the serum resistin level and pulmonary function, which showed that the serum resistin level was negatively correlated with the forced expiratory volume in the first second percentage predicted (FEV₁%predicted; r =-0.193, p = 0.037) and forced expiratory volume in the first second/forced vital capacity percentage predicted (FEV₁/FVC, r= -0.206, p = 0.026; **Figure 1B**). The mMRC and CAT scores are widely used to assess the clinical symptoms of patients with

COPD. As shown in Figure 1C, the serum resistin levels were positively correlated with the mMRC (r = 0.288, p = 0.002) and CAT scores (r = 0.215, p = 0.023). Reduced exercise tolerance is one of the main clinical characteristics of patients with COPD and is associated with a poor prognosis. The 6MWD (6-min walking distance) is a widely used tool to assess exercise tolerance. As shown in Figure 1D, the serum resistin levels were negatively correlated with the 6MWD (r = -0.527, p < 0.001). In addition, malnutrition has negative effects on lung function, as well as an increase in exacerbations and mortality. Body mass index (BMI), body fat ratio, and fat-free mass index (FFMI) are used to assess nutritional status in COPD patients. As shown in Figure 1E, the serum resistin levels were negatively correlated with BMI (r = -0.274, p = 0.003) and FFMI (r = -0.284, p = 0.014) but unrelated to the body fat ratio. Overall, these results suggested that resistin is an important biomarker in patients with COPD.

Relationship of the Serum Resistin Level With Skeletal Muscle Function in Patients With COPD

Next, we analyzed the relationship between skeletal muscle function and serum resistin levels in patients with COPD. As shown in Figure 2A, the serum resistin levels were negatively correlated with the SMMI (skeletal muscle mass index; r =-0.273, p = 0.010). Next, we analyzed the relationship between resistin levels and muscular strength. As shown in Figure 2B, the serum resistin levels were negatively correlated with quadriceps muscular strength (QMS; r = -0.332, p = 0.004) and handgrip strength (HGS; r = -0.190, p = 0.040). Reduced physical performance is a common feature of patients with COPD and is caused by poor skeletal muscle function. The 5STS is used to assess physical performance; a higher 5STS indicates lower physical performance (43). As shown in Figure 2C, the serum resistin levels were positively correlated with the 5STS (r = 0.212, p = 0.047). Finally, we analyzed the relationship between resistin and the rectus femoris. An abnormal muscle quantity in the lower limbs is a manifestation of skeletal muscle dysfunction in patients with COPD (44). The thickness and cross-sectional area of the rectus femoris in patients with COPD were significantly reduced compared with those in healthy controls (45). Based on Figure 2D, the serum resistin levels were negatively correlated with RF_{thick} (r = -0.440, p < 0.001) and RF_{csa} (r = -0.445, p < 0.001). These findings suggest that resistin is an important skeletal muscle biomarker in patients with COPD.

Clinical Factors Associated With Sarcopenia in Patients With COPD

Univariate and multivariate logistic analyses were used to explore the clinical factors associated with sarcopenia (**Table 2**). The results of univariate analysis showed that old age [odds ratio (OR): 1.084, 95% CI: 1.028–1.144, p = 0.003], high BMI (OR: 0.844, 95% CI: 0.752–0.948, p = 0.004), high FEV₁/FVC (OR: 0.957, 95% CI: 0.920–0.995, p = 0.028), and resistin (OR: 5.147, 95% CI: 3.934–11.584, p < 0.001) were associated with sarcopenia in patients with



FIGURE 1 The relationships of serum resistin levels with the clinical features of patients with COPD. (A) Difference in serum resistin level in patients with global initiative for chronic obstructive lung disease (GOLD) A and GOLD B and in patients with GOLD C and GOLD D; (B) The relationship between serum resistin level and FEV₁% predicted, FEV₁/FVC; (C) The relationship between serum resistin level and mMRC score, CAT score; (D) The relationship between serum resistin level and 6MWD; (E) The relationship between serum resistin level and BMI, body fat ratio, FFMI. p < 0.05.

COPD. Multivariate analysis showed that old age (OR: 1.115, 95% CI: 1.032–1.234, p = 0.008), high BMI (OR: 0.800, 95% CI: 0.683–0.937, p = 0.006), high FEV₁/FVC (OR: 0.947, 95% CI: 0.912–0.987, p = 0.031), and serum resistin level (OR: 4.417, 95% CI: 1.645–5.029, p = 0.007) were independent factors.

Clinical Value of Serum Resistin Levels for Predicting Sarcopenia in Patients With COPD

Previous studies indicated that a systemic inflammatory factor (TNF- α) was associated with sarcopenia in patients with COPD

(20), and resistin is involved in the activation of multiple inflammatory signaling pathways (46). Therefore, we next analyzed the relationship between resistin and TNF- α expression in patients with COPD and the predictive value of the resistin level for sarcopenia. The serum levels of resistin and TNF- α were significantly higher in patients with sarcopenia than in patients without sarcopenia (**Figures 3A,B**). Furthermore, the serum levels of resistin were shown to be positively correlated with the serum levels of TNF- α (r = 0.250, p = 0.007) (**Figure 3C**). The sensitivity and specificity of the serum level of resistin for predicting sarcopenia were 93.02 and 62.16%, respectively [the cutoff point was 7.138 ng/ml, and the AUC value was 0.828 (95% CI: 0.753–0.903)] (**Figure 3D**).



TABLE 2 | Clinical factors associated with sarcopenia in patients with COPD.

Variable	Sarcopenia					
	Univariate analysis		Multivariate analysis			
	OR (95%CI)	P-value	OR (95%CI)	P-value		
Age	1.084 (1.028–1.144)	0.003	1.115 (1.032–1.234)	0.008		
Sex	0.588 (0.255–1.357)	0.214				
BMI	0.844 (0.752–0.948)	0.004	0.800 (0.683–0.937)	0.006		
FEV1/FVC	0.957 (0.920–0.995)	0.028	0.947 (0.912–0.987)	0.031		
Serum resistin level	5.147 (3.934–11.584)	<0.001	4.417 (1.645-5.029)	0.007		

In addition, the sensitivity and specificity of the serum level of TNF- α for predicting sarcopenia were 74.42 and 47.3%, respectively [the cutoff point was 2.637 ng/ml, and the AUC value was 0.621 (95% CI: 0.511–0.731)] (**Figure 3D**). Because the diagnostic efficacy of resistin was significantly better than that of TNF- α , we next validated the diagnostic efficacy of resistin in the validation set. As shown in **Figure 3E**, the AUC value for predicting sarcopenia based on resistin was 0.818 (95% CI: 0.711–0.926, p < 0.001). Collectively, these results suggest that resistin may be associated with systemic inflammation and that serum resistin expression may serve as a biomarker for predicting sarcopenia in patients with COPD.

DISCUSSION

This is the first study to identify resistin for predicting sarcopenia in patients with COPD. In addition, we determined the cutoff value of serum resistin level that distinguished sarcopenia and validated the cutoff value in an independent validation set. Previous studies (47, 48) identified several biomarkers to predict sarcopenia in patients with COPD. Compared to these biomarkers, the strengths of this study were the large sample size, the high predictive power of serum resistin levels, and the validation of an external independent data set.

Resistin is a hormone expressed and secreted by immune cells, especially macrophages, and is associated with inflammatory



responses (49). Extensive studies have shown that resistin is a key inflammatory cytokine involved in many chronic diseases, such as atherosclerosis (25), tumors (50), and diabetic vascular disease (51). We found that the serum levels of resistin were increased with increasing GOLD stage in patients with COPD and significantly correlated with pulmonary function, clinical symptoms, nutritional status, and exercise tolerance. The data reported in this study are consistent with previous reports (30, 52). The serum resistin level did not correlate with the body fat ratio in this study. In the rodent model, resistin was initially suggested to be related to the increase in fat mass, which was due to resistin being derived almost exclusively from adipose tissue in rodents (53, 54). However, resistin is expressed primarily in inflammatory cells (monocytes, macrophages, etc.) in humans and play an important role in inflammation (55, 56). BMI (weight adjusted for stature, kg/m²) is used to describe underweight, normal, overweight and three classes of obesity, as defined by the World Health Organization (WHO) (57). Although BMI is a good marker of overall body fat content (58, 59), it cannot discern the composition of body mass, which

is defined as the proportions and distribution of bone, muscle and fat tissues in the human body. In this study, we found that serum resistin levels were associated with BMI and did not correlate with body fat ratio. This difference may be due to the serum resistin level being associated with FFMI, which represents the lean component (muscle, bone, etc.) of the body devoid of fat. Importantly, this study showed, for the first time, that serum resistin levels were negatively correlated with skeletal muscle mass and function in patients with COPD. In addition, the serum resistin levels correlated with systemic inflammation, as indicated by the serum TNF- α level, in patients. TNF- α is an important pro-inflammatory cytokine (60). The elevated expression of TNF-α in muscle or circulatory system is associated with loss of muscle strength and muscle mass (61). TNF- α leads to the degradation of skeletal muscle protein, and leads to an impaired muscle regenerative capacity. Mechanistically, the activation of NF-κB by TNF-α causes the loss of MyoD mRNA, and MyoD and myogenin are degraded via ubiquitinproteasome pathway and autophagy system. In additional, TNFα could bound to TNFR, leading to downstream activation of the caspase cascade (caspase-8 and -10) which stimulates nuclear apoptosis of skeletal muscle. Overall, TNF- α was implicated in the pathogenesis of sarcopenia. In conclusion, this study suggested that resistin is a potential biological marker in patients with COPD.

Resistin, as an adipokine, affects skeletal muscle mass or function in a variety of ways, such as via endocrine, inflammation, and energy metabolism (26, 62). A previous study (63) found that resistin levels are inversely correlated with muscle mass and quadricep strength. Resistin inhibits the differentiation of skeletal muscle satellite cells into skeletal muscle cells and promotes differentiation into adipocytes, leading to a decrease in skeletal muscle mass (64). Resistin effects are mediated by the Toll-like receptor 4 (TLR4) receptor, resulting in the activation of the p38 mitogen-activated protein kinase (MAPK), and nuclear factor-kB (NF-kB) pathways, involving in the occurrence and development process of muscle atrophy and muscle dysfunction. In additional, resistin induce ectopic deposition of lipids in mouse skeletal muscle cell C2C12 through miR-696 (65). Ectopic fat deposition of skeletal muscle has been recognized as an important component of sarcopenia (66, 67). These results suggest that resistin level is closely associated with the development of sarcopenia. Another study involving 2,000 subjects based on abdominal CT images showed that abdominal muscle density was negatively correlated with resistin levels and that targeted exercise of abdominal muscles helped prevent the development of sarcopenia (68). Whether resistin levels predict sarcopenia has not been established. Our results from the development and validation sets indicate that higher serum resistin levels (>7.138 ng/ml) have good predictive ability for sarcopenia. This study was the first to clarify the predictive value of the serum resistin level for sarcopenia in patients with COPD. Importantly, we determined the cutoff point of the serum resistin level, which will contribute to clinical application.

In this study, we identified TNF- α and resistin as predictors of sarcopenia in patients with COPD. And, the predictive efficacy of resistin was significantly better than that of TNF- α . TNF- α and resistin are pro-inflammatory cytokines (69, 70). Chronic systemic inflammation has been postulated as a mechanism in the loss of skeletal muscle in COPD. TNF-a has been recognized as systemic inflammatory molecules acting synchronously on lung parenchyma and the skeletal muscle in lung disease, is a key cytokine in the regulation of muscle mass signaling via a number of pathways (71, 72). Unlike other inflammatory factors, resistin is an adipokine. Resistin can be involved in lipid ectopic deposition by regulating fatty acid metabolism in skeletal muscle cells (73). Fatty infiltrations in skeletal muscles, resulting in decreased overall strength and functionality, leading occurrence of sarcopenia (74). This may represent the reason for the better predictive efficacy of Resistin in sarcopenia.

It is time-consuming and difficult to assess sarcopenia in all COPD patients in clinical practice, especially in community hospitals. More patients could be identified as having sarcopenia based on our prediction model, which would allow clinicians to detect and intervene with more patients at risk of sarcopenia. Therefore, serum biomarker may help clinicians diagnose sarcopenia at early admission, prehospital admission, or in the community. Sarcopenia is an age-related disease, and with the aging of the human body, there is inevitably a gradual decline in muscle mass, quality, and strength (75). In this study, we demonstrated that the association of serum resistin levels with sarcopenia in patients with COPD was independent of age. In the future, we will try to analyze the temporality of the variables and the order of occurrence of each variable through a multicenter, prospective longitudinal study.

COPD is often combined with other chronic inflammatory diseases [such as cardiovascular disease (76) and asthma (52)], which may influence serum resistin levels. In addition, resistin levels were significantly increased in patients with acute exacerbation COPD (30). Therefore, the subjects of this study were patients with stable COPD and patients with clear comorbidities (presence of severe cardiovascular disease, active lung disease, etc.) were excluded. In the future, we will try to collect more clinical samples through multicenter, prospective longitudinal studies to further analyze the clinical value of serum resistin levels in patients with different comorbidities and different disease states of COPD.

This study also had several limitations. The main limitation of this study is that our study employed a cross-sectional design, and assessments were implemented only at the time of study entry. We need to consider the fluctuations in muscle mass and resistin levels over time. Additional prospective studies are needed to support the use of resistin levels as a predictor of risk for developing sarcopenia. The BIA that we used to measure muscle mass is widely used for the diagnosis of sarcopenia but is not a good surrogate of skeletal muscle mass. In addition, the study was limited to stable COPD patients, and whether the findings can be applied to patients with acute exacerbations of COPD and those undergoing pulmonary rehabilitation is not known.

In conclusion, serum resistin levels were shown to be associated with systemic inflammation and can be used accurately and easily to predict sarcopenia in patients with COPD.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Research Ethics Committee of China Medical University. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

Conceptualization, writing—original draft, and writing—review and editing: GH and MD. Data curation: MD, JG, YL,

YY, and XZ. Methodology: QZ. Project administration: XZ. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnut.2022. 921399/full#supplementary-material

 $\label{eq:supplementary Figure 1 | The histograms (A), Q-Q plots (B) of serum resistin data.$

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