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Sarcopenia: Molecular regulatory network for loss of muscle mass and function

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Skeletal muscle is the foundation of human function and plays a key role in producing exercise, bone protection, and energy metabolism. Sarcopenia is a systemic disease, which is characterized by degenerative changes in skeletal muscle mass, strength, and function. Therefore, sarcopenia often causes weakness, prolonged hospitalization, falls and other adverse consequences that reduce the quality of life, and even lead to death. In recent years, sarcopenia has become the focus of in-depth research. Researchers have suggested some molecular mechanisms for sarcopenia according to different muscle physiology. These mechanisms cover neuromuscular junction lesion, imbalance of protein synthesis and breakdown, satellite cells dysfunction, etc. We summarize the latest research progress on the molecular mechanism of sarcopenia in this review in order to provide new ideas for future researchers to find valuable therapeutic targets and develop relevant prevention strategies.

KEYWORDS

sarcopenia, molecular mechanism, neuromuscular junction lesion, imbalance of protein synthesis and breakdown, satellite cells dysfunction

1. Introduction

Sarcopenia was described as a syndrome by the European Working Group on Sarcopenia in Older People (EWGSOP) in 2010, which was characterized by degenerative changes in skeletal muscle mass and function throughout the body (1). However, sarcopenia is recognized as a muscle disease in the EWGSOP2 Consensus updated in 2019 (2). The guidelines (1, 2) state that decreased muscle strength as an important sign of sarcopenia should be examined first when a patient is suspected of sarcopenia. A patient is diagnosed with sarcopenia when both muscle strength and muscle mass are reduced, while a loss of muscle strength alone is pre-sarcopenia. In addition, muscle performance is an important indicator of the severity of disease.

Sarcopenia and cachexia are generally difficult to distinguish clinically in elderly cancer patients due to the common features of muscle atrophy (3). However, Berardi et al. (4) found that unlike cachexia, which features gradual loss of lean mass and adipose tissue in the absence of substantial muscle protein loss, sarcopenia is usually accompanied by chronic protein loss. Furthermore, systemic inflammation plays an important role in the occurrence and development of cachexia, which can be used to identify cachexia (5). In contrast, sarcopenia is often associated with age-related chronic low-grade inflammatory profile, and can occur even in patients without inflammation (4, 6). Therefore, the in-depth exploration of the mechanism of the occurrence and development of sarcopenia can contribute to the clinical differential diagnosis.

The quality and function of skeletal muscle are affected by a variety of molecular pathways such as aging, inflammation, malnutrition, oxidative stress, and mitochondrial dysfunction, thus accelerating the occurrence and development of sarcopenia (1, 2). However, the study of the molecular mechanism of sarcopenia is still limited due to the complexity and interrelation of these molecular pathways. It is now widely identified that the increase of age is a central element affecting the development and progression of physiological sarcopenia (7, 8). Age is not only a risk factor for diseases such as diabetes, cardiovascular disease, and cancer, but also a risk factor for sarcopenia (9-12). The main mechanism of skeletal muscle aging is muscle fiber atrophy and loss from the perspective of tissue level, while the muscle fiber atrophy is mainly caused by the imbalance between muscle protein synthesis and breakdown (13). Moreover, sarcopenia has recently been recognized as one of the criteria for the diagnosis of malnutrition by the Global Leadership Initiative on Malnutrition (GLIM) (14). Schneider et al. (15) have shown that sarcopenia can be used as a substitute for malnutrition and debilitating symptoms in patients, which further confirmed that sarcopenia was closely related to malnutrition.

Sarcopenia increases the mortality and disability rate of the elderly, affects the prognosis of patients, and increases the economic burden of patients (16). Herein, we discuss the fundamental molecular mechanisms of the occurrence and development of sarcopenia. Based on these studies, we are expecting to clarify its internal changes, develop prevention strategies, and identify molecular targets for future therapeutic interventions.

2. Neuromuscular junction lesion

Neuromuscular junction disease can lead to decreased muscle function through neuronal degeneration and muscle fiber denervation, which is one of the most important causes of sarcopenia (17). The neuromuscular junction (NMJ) is composed of three main components: the presynaptic motor nerve terminals; the synaptic cleft; and the endplate (18).

Agrin is released by motor neurons (19). Action potentials can be correctly transmitted from nerve endings to muscle through the agrin-muscle specific kinase (MuSk)-lipoprotein receptor-related protein 4 (Lrp4) signaling pathway, which is essential for maintaining normal neuromuscular function (20). Meanwhile, the agrin-MuSklrp4 signaling pathway mediates the aggregation of acetylcholine (ACh) receptors on the postsynaptic membrane (21). A growing number of research results have demonstrated this signaling pathway is dysfunction with the age increase, leading to changes in NMJ structure and function (18, 22, 23).

Deschenes et al. (24) proposed that acetylcholine release increased in aged muscles leading to a faster rate of neuromuscular fatigue. It has been suggested that decreasing ACh may be an effective remedy to treat structural changes in NMJ that can lead to sarcopenia (25). The same result was obtained in another study, which found that moderately reduced ACh levels could increase NMJ area, myofiber cross-sectional area, and satellite cells number in mice (26). Interestingly, Castets et al. (27) reported that the mechanistic target of rapamycin (mTOR) signal transduction also has a positive effect in the structural formation and functional maintenance of NMJ. This evidence may suggest that NMJ functional changes in sarcopenia are related to decreased protein synthesis. However, the molecular mechanism is still unclear and needs to further study.

3. Satellite cells dysfunction

As a key pathway for skeletal muscle satellite cell differentiation, wnt signaling pathway can stimulate skeletal muscle development and regeneration during the embryonic period, which is an extremely coordinated process (28). Studies targeting the typical wnt/ β -catenin pathway have shown that the number of proliferative satellite cells increases with activation of the pathway and that the pathway can inhibit skeletal muscle atrophy under the regulation of the zinc finger transcription factor Kruppel-like factor 5 (KLF5) (29, 30). However, several researchers have recently demonstrated that hyperactive wnt/ β -catenin signaling pathway can also lead to damage of skeletal muscle stem cells and inhibition of muscle regeneration (31, 32). These evidences suggest that balancing the expression of the canonical wnt pathway is an essential prerequisite for muscle regeneration.

Growing evidence indicated that the Type II muscle fiber atrophy leads to muscle mass loss in the sarcopenia, and the loss of satellite cells may be a significant cause of type II myofiber atrophy (33-35). Furthermore, Laura et al. found that the function and number of skeletal muscle stem cells decreased with age (36). A recent study suggests that the p53-p21CIP1 and P16INK4a-retinoblastoma (Rb) pathways may be the major signaling pathways for the activation of these stressors (37). Downstream studies on the two pathways have shown that cyclin-dependent kinase (CDK) 2 and CDK4/6 may act as downstream inhibitory molecules to promote over-phosphorylation of Rb protein, which can ultimately terminate cell division and thus enter the senescence state (37). The upstream signals of both pathways have been widely studied in satellite cells. A recent study demonstrated that loss of zinc-finger transcription factor Slug, which was a direct transcriptional repressor of p16^{InK4a}, could cause satellite cells to enter complete senescence under damage-induced stress (38). Furthermore, Myers et al. (39) reported that sirtuin 1 (SIRT1) and p53 have a synergistic effect in protecting skeletal muscle against fatigue damage, and type I fibers can also be protected from atrophy damage by this effect. Accordingly, SIRT1 may selectively inhibit the mechanism of p53 fiber damage, whereas further study needs to be done to confirm this conclusion. In contrast to the above views, Fry et al. (40) found that both mice injected with tamoxifen to clear satellite cells and control mice developed sarcopenia, and muscle fiber atrophy and grip strength were not changed by clearing satellite cells. Meanwhile, an increase in fibrosis was observed in skeletal muscle deprived of satellite cells. Therefore, the role of decreased regenerative capacity caused by senescence of satellite cells in senile sarcopenia needs further study.

4. Decreased protein synthesis

Currently, a large amount of studies have been done to explore the molecular mechanism of insulin-like growth factor 1 (IGF-1) participating in protein synthesis (41–45). The IGF-1 is an essential factor of metabolic molecular signaling, which is involved in a variety of skeletal muscle anabolic pathways. IGF-1 first binds to and *trans* phosphorylates the transmembrane tyrosine kinase receptor (IGFR) to form the docking site of insulin receptor substrate 1 (IRS-1). IGFR subsequently binds IRS-1 to activate the phosphoInositide-3 kinase (PI3K)-protein kinase B (Akt) pathway. Akt can activate mammalian target of rapamycin (mTOR) through phosphorylation and inactivates tuberous sclerosis complex-2. Subsequently, the 70 kDa ribosomal protein S6 kinase (P70S6K) is phosphorylated and activated by mTOR. Finally, eukaryotic initiation factor 4B (eIF4B) and programmed cell death 4 (PDCD4), which are two proteins that modulate cap-dependent mRNA translation are phosphorylated by activated p70S6K subsequently, resulting in increased mRNA translation either directly or indirectly. In addition, the IGF-1-PI3K-Akt pathway can also phosphorylate and inactivate O-type fork head box (FOXO) transcription factors to prevent protein degradation (Figure 1). FoxO1, FoxO3, and FoxO4 can promote the activation of protein breakdown mechanism with increasing the transcription of muscle ring finger protein-1 (MuRF-1) and amyotrophic F-box (MAFbx) genes, which are three isoforms expressed in skeletal muscle. It should be noted that Akt requires the simultaneous integration of FOXO1 and mTORC1 pathways to maintain oxidative metabolism and protein synthesis, which are not parallel (46).

Ascenzi et al. (47) found that molecular markers related to the autophagy pathway were significantly up-regulated in muscles of aged IGF-1Ea and IGF-1Eb mice. In addition, the two isoforms could also promote the expression of mitochondrial fusion proteins and mitochondrial fission process protein 1, which is a downstream target of a factor (PGC1- α) with antioxidant stress, to promote mitochondrial fusion and fission. The above evidence suggest that IGF-1 isoforms can counteracts sarcopenia by activating a series of molecular pathways that maintain skeletal muscle anabolism and regeneration. However, they found that only IGF-1Ea could promote skeletal muscle enlargement when both isoforms were overexpressed. This result implies that the two subtypes may mediate different molecular mechanisms in skeletal muscle, which needs further research to confirm.

In recent years, decreased levels of IGF-1 have been found to be associated with sarcopenia in various disease states (48– 50). Xu et al. (51) found that patients with sarcopenia were older (P < 0.001) and had significantly lower IGF-1 levels (P < 0.01) through measuring the hormone levels of patients, suggesting that aging may affect IGF-1 levels. This is the same result as the study by Naro et al. (52). However, Naro et al. found the expression of genes in muscles of the elderly that could cause muscle fiber atrophy were also decreased significantly. The above evidences suggest that reduced protein anabolic metabolism is closely associated with sarcopenia, while it seems that it can play a compensatory role by downregulating atrophic factors in the elderly, thereby partially restoring protein stability.

5. Ubiquitin proteasome

Ubiquitin proteasomes (UPS) and autophagic lysosomes are two proteolytic systems that control protein degradation in skeletal muscle (53). Proteins are ubiquitinated by ubiquitin activator enzyme (E1), ubiquitin-binding enzyme (E2), and ubiquitin ligase (E3), and then degraded into short peptides and amino acids in the 26S/30s proteasome (42) (Figure 2). E3 is a key enzyme in this pathway, which can identification specific target protein sequences and precisely regulate the degradation rate of the target protein. Atrogin-1 (MAFbx) and MURF-1 are two important E3 ubiquitin ligases, which are regarded as markers of skeletal muscle atrophy. Researchers have observed their expression in a variety of diseases that can result in skeletal muscle atrophy (54).

Recently, there are increased evidences have demonstrated that the level of oxidative stress and chronic inflammation is an essential factor in inducing sarcopenia in the elderly (55, 56). Some studies have shown that the p38 mitogen-activated protein kinase (MAPK) and nuclear factor-Kappa B (NF-KB) paths are two basic pathways for UPS activation, which can be activated and regulated by oxidative stress and inflammatory cytokines, such as tumor necrosis factor- α (TNF- α) And interleukin-1 (IL-1) (57). In another study, excessive reactive oxygen species (ROS) and inflammatory factors



FIGURE 1

Insulin-like growth factor 1 (IGF-1)-mediated signaling pathways relevant to protein synthesis. IGF-1, insulin-like growth factor 1; IRS-1, insulin receptor substrate 1; PI3K, phospholnositide-3 kinase; Akt, protein kinase B; mTOR, mechanistic target of rapamycin; p70S6K, 70-kDa ribosomal protein S6 kinase; eIF4B, eukaryotic initiation factor 4B; PDCD4, programmed cell death 4; FOXO, O-type fork head box; MuRF-1, muscle RING-finger protein-1; MAFbx, muscle atrophy F-Box. Arrow indicates activation of expression; lines with a perpendicular line at the end indicate inhibition of expression.



can activate UPS and autophagy processes through phosphorylation signal transducers and transcriptional activators 3 (58).

Wu et al. (59) found that the expression of MAFbx or MuRF-1 gene in normal young muscle was similar to the elderly. However, Bodine and Baehr (60) found that proteasome and other components of UPS increased in muscle with researching sarcopenia rats. The study of Zhang et al. (61) also obtained the same result, which found that MuRF-1 expression and protein ubiquitination were higher in patients with sarcopenia than in non-sarcopenia patients, while atrogin-1 expression was not significantly different. Meanwhile, Sharma et al. (62) found that T. cordifolia water extract could reduce oxidative stress and inflammation by inhibiting MURF-1 and calpain, which can reduce proteolysis. The above evidence indicates that UPS activated mainly in patients with severe muscle atrophy, and MuRF-1 plays a major role in skeletal muscle degradation. UPS has a promising future as a therapeutic target for sarcopenia.

The proteolysis effect of UPS in inducing muscle mass loss has been widely confirmed. However, studies have also shown that UPS plays an important role in maintaining muscle mass. Three isogenies of MuRF, MuRF-1, MURF-2, and MuRF-3, are expressed in myocardial and skeletal muscle as the key factor E3 of UPS (63). Perera et al. (64) showed myogenic differentiation and delayed myotube formation after MURF-2 knockout in mice. Lodka et al. (65) found that the muscle strength of soleus and extensor digitalis longus in MuRF-2 and MuRF-3 double-knocked mice was weakened, and the fiber type changed to type I. Moriscot et al. (66) found significant loss of sodium type ii fibers in mice with MuRF1 and MuRF2 knocked out and showed that the maintenance effects of MuRF-1 and MuRF-2 on type ii fibers were closely related to myozenin-1. The above evidence suggests that the MuRF family plays an important role in the maintenance of early differentiation, structure, and function of skeletal muscle. It remains to be seen whether the UPS-related gene knockout alone can benefit patients with sarcopenia.

6. Autophagy

In recent years, autophagy, which is a protein degradation mechanism other than UPS, has been extensively studied in relation to sarcopenia (67, 68). Pigna et al. (69) have demonstrated that maintaining a balance of autophagy production and clearance guarantees adequate autophagy fluxes in muscles, and this balance is disrupted in colon cancer mice, leading to incorrect autophagy fluxes and promoting muscle consumption. This result suggests that the disruption of autophagy balance may induce sarcopenia.

Aging may be an important cause of autophagy defects among the factors that break the balance. A recent study has shown that the clearance capacity of autophagy lysosome system (ALS) decreases with aging, resulting in insufficient clearance of accumulated wastes by cells (70). Moreover, studies have found that autophagy defects in aged stem cells could reduce the regenerative ability of satellite cells in sarcopenia, and the activation of autophagy clears senescent cells in time to promote muscle regeneration (71, 72).

Excessive autophagy caused by disease states such as cancer and malnutrition may be another important cause of autophagy imbalance. Zhang et al. (61) confirmed that the protein expressions of Beclin-1 (the autophagy key regulatory protein complex) and LC3B (LC3-II and LC3-I) were up-regulated in sarcopenia patients with or without gastric cancer, suggesting ALS is activated. Furthermore, it has demonstrated that malnutrition can inhibit the PI3K-Akt signaling pathway in mouse muscles, which can subsequently induce autophagy through reducing mTOR and activating FOXO3, resulting in excessive autophagy (53).

In addition to affecting muscle mass, muscle function can also be impaired by the breakdown of autophagy balance due to the dysfunction of autophagy in the process of mitochondrial clearance (73). Mitophagy selectively degrades mitochondria through the Pten-induced putative kinase 1 (PINK1)/Parkin pathway and BCL2 adenovirus E1B 19-KDA-Interacting protein 3 (BNIP3)/BNIP3like (BNIP3L, also known as NIX) pathway, thereby eliminating dysfunction and redundant mitochondria to maintain energy homeostasis. The BNIP3/NIX pathway directly interacts with LC3-II through the connexin p62 (74–76). Urine-varela et al. (74) found that the molecular expression of mitochondrial autophagy signaling pathway decreased in the process of aging, leading to mitochondrial dysfunction and ros induced biomolecular damage increased. Meanwhile, Davuluri et al. (76) demonstrated that mitochondrial phagocytosis increases with age, resulting in a decrease in mitochondrial content in aging muscle. According to the above evidence, maintaining equilibrium and efficient autophagy is crucial for preserving muscle homeostasis.

7. Nutrition

Endoplasmic unfolded protein response (UPR) is the general name of the signaling pathway induced by endoplasmic reticulum (ER) stress (77). Hulmi et al. (78) found that UPR/ER pathway indicators such as activated transcription factor-6 (ATF6), inositol requirement protein (IRE) 1 α , and protein kinase R (PKR) -like endoplasmic reticulum kinase (PERK) protein were increased and muscle strength was decreased in a mouse model of malnutrition. Meanwhile, levels of multiple UPR markers in skeletal muscle of fasting mice were found to be decreased with the loss of TNF receptor-associated factor 6(TRAF6), an E3 ubiquitin ligase (79). These studies suggest that UPR may be involved in skeletal muscle atrophy caused by nutritional deficiency. However, the factors that activate UPR in malnutrition and the specific mechanisms of the various branches of UPR in skeletal muscle atrophy remain unclear and require further investigation.

Amino acids are primarily stored in skeletal muscle. Previous studies have shown that basic amino acid metabolism may not be

affected during aging, while the ability to stimulate anabolic responses is reduced in older adults (80). Essential amino acids (EAA) and branched-chain amino acids can stimulate protein synthesis in the elderly. Leucine can directly initiate mRNA translation and play a key role in regulating human and rat muscle protein synthesis (81). However, the effect appears to diminish in older adults. Similar to this argument, Drummond et al. (82) found that the muscle protein synthesis (MPS) response of the elderly was slower than that of the young after resistance exercise and EAA intake, which may be related to the non-response of ERK1/2 signal and AMPK activation in the aging muscles. This anabolic resistance is closely associated with loss of muscle mass, strength, and function (83). However, further research is needed to elucidate the upstream drivers of this anabolic resistance. In addition, Brook et al. (84) have shown that muscles need to provide other tissues with amino acids and other substrates due to the huge energy requirements of patients with nutrient deficiencies, which are mainly manifested as accelerated catabolism and reduced synthesis of muscle protein, while protein synthesis of liver increases. This means that skeletal muscle tissue may be continuously depleted and reduced in some disease states due to disruption of amino acid balance in the body.

Combining a nutritional strategy of amino acid supplementation with exercise may be the best treatment strategy for people with agerelated sarcopenia (85). Leucine supplementation may aid in muscle protein synthesis in the elderly due to its important role in protein synthesis. Xu et al. (86) have shown that leucine supplementation can ameliorate the sluggish muscle response to amino acid intake in older adults and contribute to increased protein synthesis after meals. Furthermore, a recent study suggests that higher intake of branched chain amino acids may help prevent sarcopenia in older adults (87). Studies have shown that physical activity can regulate oxidative stress and satellite cell activity, increase muscle's ability to adapt to oxidative



FIGURE 3

Molecular mechanism of sarcopenia. The agrin-MuSk-Lrp4 signaling pathway may be dysregulated with the age increase, resulting in the neuromuscular junction lesion. Slug and SIRT1 inhibit cell senescence by inhibiting the two signaling pathways of p53-p21CIP1 and p16^{InK4a}-Rb. The absence of two factors accelerates the entry of satellite cells into a state of complete senescence under damage-induced stress. The KLF5 can regulate skeletal muscle atrophy through wnt/ β -Catenin signaling pathway. The decrease of IGF-1 leads to a decrease in the activation of P13K-Akt-mTOR signaling pathway, which result in the decrease in protein synthesis. Oxidative stress and inflammatory cytokines can activate and regulate MAPK and NF-KB pathways, which can activate the UPS. Malnutrition can inhibit the P13K-Akt signaling pathway, which subsequently activates autophagy by activating FOXO3. Arrow indicates activation of expression; lines with a perpendicular line at the end indicate inhibition of expression.

stress, and promote the proliferation of satellite cells, thus delaying the development of sarcopenia (88). In addition, resistance training can increase the cross-sectional area of type II muscle fibers and improve aging muscle outcomes and function (29).

Caloric restriction (CR) may be a measure to prevent the adverse effects of aging for the elderly. CR and maintenance of adequate micronutrient supply have been found to play a positive role in extending life span and alleviating aging-induced sarcopenia in mouse models (89, 90). However, caloric restriction should probably be used to limit calorie intake while providing adequate protein since malnutrition is common in many elderly patients with sarcopenia, and increased amino acid intake may delay sarcopenia (43).

8. Inflammation and metabolism

Studies have shown that age-related muscle loss is often associated with chronic low-grade inflammation (6). This effect is often associated with inflammatory factors that can cause imbalances in muscle protein synthesis and breakdown, which can lead to muscle atrophy (91, 92). In multiple studies of the relationship between inflammation and sarcopenia, levels of the inflammatory markers C-reactive protein, TNF- α , and IL-6 have been negatively correlated with muscle strength and mass (93, 94).

Age-related sarcopenia may be accompanied by an increase in fat content along with a decline in muscle strength and mass (95). In response to aging and chronic inflammation, the increased fat content in the body is redistributed, resulting in increased visceral fat content and infiltration of skeletal muscle fat (96). The increase of fat infiltration in and between myocytes can induce mitochondrial dysfunction, resulting in the increase of ROS produced by lipid oxidation, and promote a series of adverse reactions such as lipid toxicity, insulin resistance (IR), and local inflammation (97). Under the combined action of ROS and local inflammation, oxidative stress is enhanced and IGF-1-PI3K-mTOR pathway is inhibited, leading to reduced protein synthesis while mitochondrial function is further damaged, forming a vicious cycle (97). Meanwhile, proinflammatory factors released by local inflammation can induce infiltration of macrophages and other immune cells, further release a large number of inflammatory factors, promote the breakdown of muscle protein, expand the range of inflammation, and aggravate the damage of IR and lipotoxicity (98). In addition, lipid toxicity has been shown to inhibit protein synthesis and promote anabolic resistance by inhibiting phosphorylation of Akt and mTOR through ceramides (99).

Studies have shown that IR leads to increased gluconeogenesis, which promotes increased triglyceride production and transport to skeletal muscle and liver, resulting in increased lipid accumulation in skeletal muscle and liver (97, 100). In addition, IR can cause compensatory hyperinsulinemia with increased myostatin production and inhibition of the IGF-1 axis, resulting in increased muscle protein breakdown, and decreased synthesis (97, 100).

9. Conclusion

Sarcopenia is a common disease leading to poor prognosis in the elderly, which occurrence is the result of multiple factors. The essence of sarcopenia is a reduction in the cross-sectional area of muscle fibers and the imbalance of protein synthesis and degradation. Currently, the molecular mechanism of sarcopenia has not been fully understood. However, the molecular mechanism of sarcopenia can be distinguished on the basis of etiology due to the in-depth research on the factors affecting the occurrence and development of sarcopenia in recent years (Figure 3).

In conclusion, the mechanism of sarcopenia involves several aspects, including neuromuscular connection damage, satellite cell dysfunction, reduced protein synthesis, increased protein breakdown, and nutrient deficiency. Precisely, aging is considered to be a key factor in sarcopenia. The protein degradation increases with aging, resulting in an imbalance between protein synthesis and breakdown that promotes the loss of muscle mass.

Taking corresponding measures (such as exercise, nutritional supplements, antioxidants, etc.) to intervene in sarcopenia-related molecular mechanism targets is the fundamental direction of future research on the prevention and treatment of sarcopenia. For example, an increasing number of studies indicate that interfering with the IGF-1 pathway can increasing protein synthesis, which may improve sarcopenia (101, 102). Further elucidation of the mechanism of sarcopenia will be helpful in determining effective lifestyle and therapeutic interventions.

Author contributions

QZ: conception and design, and administrative support. JW, PD, PY, HG, and YT: provision of study materials and patients. PD, PY, YT, and HG: collection and assembly of data. JW and PD: data analysis and interpretation. All authors wrote and approved the final version of manuscript.

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Conflict of interest

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References

1. Cruz-Jentoft A, Baeyens J, Bauer J, Boirie Y, Cederholm T, Landi F, et al. Sarcopenia: European consensus on definition and diagnosis: report of the european working group on sarcopenia in older people. *Age Ageing*. (2010) 39:412–23. doi: 10.1093/ageing/afq034

2. Cruz-Jentoft A, Bahat G, Bauer J, Boirie Y, Bruyère O, Cederholm T, et al. Sarcopenia: revised European consensus on definition and diagnosis. *Age Ageing*. (2019) 48:16–31. doi: 10.1093/ageing/afy169

3. Meza-Valderrama D, Marco E, Dávalos-Yerovi V, Muns M, Tejero-Sánchez M, Duarte E, et al. Sarcopenia, malnutrition, and cachexia: adapting definitions and terminology of nutritional disorders in older people with Cancer. *Nutrients.* (2021) 13:761. doi: 10.3390/nu13030761

4. Berardi E, Madaro L, Lozanoska-Ochser B, Adamo S, Thorrez L, Bouche M, et al. A pound of flesh: what cachexia is and what it is not. *Diagnostics (Basel)*. (2021) 11:116. doi: 10.3390/diagnostics11010116

5. Muscaritoli M, Anker S, Argilés J, Aversa Z, Bauer J, Biolo G, et al. Consensus definition of sarcopenia, cachexia and pre-cachexia: joint document elaborated by Special Interest Groups (SIG) "cachexia-anorexia in chronic wasting diseases" and "nutrition in geriatrics". *Clin Nutr.* (2010) 29:154–9. doi: 10.1016/j.clnu.2009.12.004

6. Beyer I, Mets T, Bautmans I. Chronic low-grade inflammation and age-related sarcopenia. *Curr Opin Clin Nutr Metab Care*. (2012) 15:12–22. doi: 10.1097/MCO. 0b013e32834dd297

7. Cruz-Jentoft A, Sayer A. Sarcopenia. Lancet. (2019) 393:2636–46. doi: 10.1016/S0140-6736(19)31138-9

8. Picca A, Calvani R. Molecular mechanism and pathogenesis of sarcopenia: an overview. Int J Mol Sci. (2021) 22:3032. doi: 10.3390/ijms22063032

9. Yuan C, Kim J, Wang Q, Lee A, Babic A, PanScan/PanC4 I-III consortium. et al. The age-dependent association of risk factors with pancreatic cancer. *Ann Oncol.* (2022) 33:693–701. doi: 10.1016/j.annonc.2022.03.276

10. Basina M, Maahs D. Age at type 1 diabetes onset: a new risk factor and call for focused treatment. *Lancet.* (2018) 392:453–4. doi: 10.1016/S0140-6736(18)31811-7

11. Wallace R, Twomey L, Custaud M, Turner J, Moyna N, Cummins P, et al. The role of epigenetics in cardiovascular health and ageing: a focus on physical activity and nutrition. *Mech Ageing Dev.* (2018) 174:76–85. doi: 10.1016/j.mad.2017.11.013

12. Cruz-Jentoft A, Landi F, Schneider S, Zúñiga C, Arai H, Boirie Y, et al. Prevalence of and interventions for sarcopenia in ageing adults: a systematic review. report of the international sarcopenia initiative (EWGSOP and IWGS). Age Ageing. (2014) 43:748–59. doi: 10.1093/ageing/afu115

13. Wilkinson D, Piasecki M, Atherton P. The age-related loss of skeletal muscle mass and function: measurement and physiology of muscle fibre atrophy and muscle fibre loss in humans. *Ageing Res Rev.* (2018) 47:123–32. doi: 10.1016/j.arr.2018.07.005

14. Cederholm T, Jensen G, Correia M, Gonzalez M, Fukushima R, Higashiguchi T, et al. GLIM criteria for the diagnosis of malnutrition - a consensus report from the global clinical nutrition community. *Clin Nutr.* (2019) 38:1–9. doi: 10.1016/j.clnu.2018.08.002

15. Schneider M, Hübner M, Becce F, Koerfer J, Collinot J, Demartines N, et al. Sarcopenia and major complications in patients undergoing oncologic colon surgery. J Cachexia Sarcopenia Muscle. (2021) 12:1757–63. doi: 10.1002/jcsm.12771

16. Xie W, He M, Yu D, Wu Y, Wang X, Lv S, et al. Mouse models of sarcopenia: classification and evaluation. *J Cachexia Sarcopenia Muscle*. (2021) 12:538–54. doi: 10. 1002/jcsm.12709

17. Narici M, Maffulli N. Sarcopenia: characteristics, mechanisms and functional significance. *Br Med Bull*. (2010) 95:139–59. doi: 10.1093/bmb/ldq008

18. Dobrowolny G, Barbiera A, Sica G, Scicchitano B. Age-Related alterations at neuromuscular junction: role of oxidative stress and epigenetic modifications. *Cells.* (2021) 10:1307. doi: 10.3390/cells10061307

19. Moreira-Pais A, Ferreira R, Oliveira P, Duarte JA. A neuromuscular perspective of sarcopenia pathogenesis: deciphering the signaling pathways involved. *Geroscience*. (2022) 44:1199–213. doi: 10.1007/s11357-021-00510-2

20. Ohno K, Ohkawara B, Ito M. Agrin-LRP4-MuSK signaling as a therapeutic target for myasthenia gravis and other neuromuscular disorders. *Expert Opin Ther Targets.* (2017) 21:949–58. doi: 10.1080/14728222.2017.1369960

21. Zong Y, Jin R. Structural mechanisms of the agrin-LRP4-MuSK signaling pathway in neuromuscular junction differentiation. *Cell Mol Life Sci.* (2013) 70:3077–88. doi: 10.1007/s00018-012-1209-9

22. Li L, Xiong W, Mei L. Neuromuscular junction formation, aging, and disorders. *Annu Rev Physiol.* (2018) 80:159–88.

23. Ding Q, Kesavan K, Lee K, Wimberger E, Robertson T, Gill M, et al. Impaired signaling for neuromuscular synaptic maintenance is a feature of motor neuron disease. *Acta Neuropathol Commun.* (2022) 10:61. doi: 10.1186/s40478-022-01360-5

24. Deschenes M, Flannery R, Hawbaker A, Patek L, Mifsud M. Adaptive remodeling of the neuromuscular junction with aging. *Cells.* (2022) 11:1150. doi: 10.3390/cells11071150

25. Sugita S, Fleming L, Wood C, Vaughan S, Gomes M, Camargo W, et al. VAChT overexpression increases acetylcholine at the synaptic cleft and accelerates aging of neuromuscular junctions. *Skelet Muscle.* (2016) 6:31. doi: 10.1186/s13395-016-0105-7

26. Vaughan S, Sutherland N, Valdez G. Attenuating cholinergic transmission increases the number of satellite cells and preserves muscle mass in old age. *Front Aging Neurosci.* (2019) 11:262. doi: 10.3389/fnagi.2019.00262

27. Castets P, Ham D, Rüegg M. The TOR pathway at the neuromuscular junction: more than a metabolic player? *Front Mol Neurosci.* (2020) 13:162. doi: 10.3389/fnmol.2020. 00162

28. von Maltzahn J. Regulation of muscle stem cell function. Vitam Horm. (2021) 116:295-311. doi: 10.1016/bs.vh.2021.02.012

29. Fujimaki S, Hidaka R, Asashima M, Takemasa T, Kuwabara T. Wnt proteinmediated satellite cell conversion in adult and aged mice following voluntary wheel running. *J Biol Chem.* (2014) 289:7399–412. doi: 10.1074/jbc.M113.539247

30. Zhang D, Yin H, Li J, Wang Y, Yang C, Jiang X, et al. KLF5 regulates chicken skeletal muscle atrophy via the canonical Wnt/ β -catenin signaling pathway. *Exp Anim.* (2020) 69:430–40. doi: 10.1538/expanim.20-0046

31. Rudolf A, Schirwis E, Giordani I, Parisi A, Lepper C, Taketo M, et al. β -Catenin activation in muscle progenitor cells regulates tissue repair. *Cell Rep.* (2016) 15:1277–90. doi: 10.1016/j.celrep.2016.04.022

32. Girardi F, Le Grand F. Wnt signaling in skeletal muscle development and regeneration. *Prog Mol Biol Transl Sci.* (2018) 153:157–79. doi: 10.1016/bs.pmbts.2017. 11.026

33. McPhee J, Cameron J, Maden-Wilkinson T, Piasecki M, Yap M, Jones D, et al. The contributions of fiber atrophy, fiber loss, in situ specific force, and voluntary activation to weakness in sarcopenia. *J Gerontol A Biol Sci Med Sci.* (2018) 73:1287–94. doi: 10.1093/gerona/gly040

34. Nilwik R, Snijders T, Leenders M, Groen B, van Kranenburg J, Verdijk L, et al. The decline in skeletal muscle mass with aging is mainly attributed to a reduction in type II muscle fiber size. *Exp Gerontol.* (2013) 48:492–8. doi: 10.1016/j.exger.2013.02.012

35. Verdijk L, Koopman R, Schaart G, Meijer K, Savelberg H, van Loon L. Satellite cell content is specifically reduced in type II skeletal muscle fibers in the elderly. *Am J Physiol Endocrinol Metab.* (2007) 292:E151–7. doi: 10.1152/ajpendo.00278.2006

36. García-Prat L, Sousa-Victor P, Muñoz-Cánoves P. Functional dysregulation of stem cells during aging: a focus on skeletal muscle stem cells. *FEBS J.* (2013) 280:4051–62. doi: 10.1111/febs.12221

37. Mankhong S, Kim S, Moon S, Kwak H, Park D, Kang J. Experimental models of sarcopenia: bridging molecular mechanism and therapeutic strategy. *Cells.* (2020) 9:1385. doi: 10.3390/cells9061385

38. Zhu P, Zhang C, Gao Y, Wu F, Zhou Y, Wu W. The transcription factor Slug represses p16^{Ink4a} and regulates murine muscle stem cell aging. *Nat Commun.* (2019) 10:2568. doi: 10.1038/s41467-019-10479-4

39. Myers M, Shepherd D, Durr A, Stanton D, Mohamed J, Hollander J, et al. The role of SIRT1 in skeletal muscle function and repair of older mice. *J Cachexia Sarcopenia Muscle*. (2019) 10:929–49. doi: 10.1002/jcsm.12437

40. Fry C, Lee J, Mula J, Kirby T, Jackson J, Liu F, et al. Inducible depletion of satellite cells in adult, sedentary mice impairs muscle regenerative capacity without affecting sarcopenia. *Nat Med.* (2015) 21:76–80. doi: 10.1038/nm. 3710

41. Glass D. Molecular mechanisms modulating muscle mass. *Trends Mol Med.* (2003) 9:344–50. doi: 10.1016/s1471-4914(03)00138-2

42. Sakuma K, Aoi W, Yamaguchi A. Molecular mechanism of sarcopenia and cachexia: recent research advances. *Pflugers Arch.* (2017) 469:573–91. doi: 10.1007/s00424-016-1933-3

43. Wiedmer P, Jung T, Castro J, Pomatto L, Sun P, Davies K, et al. Sarcopenia - molecular mechanisms and open questions. *Ageing Res Rev.* (2021) 65:101200. doi: 10.1016/j.arr.2020.101200

44. Javanmardifard Z, Shahrbanian S, Mowla S. MicroRNAs associated with signaling pathways and exercise adaptation in sarcopenia. *Life Sci.* (2021) 285:119926. doi: 10.1016/j.lfs.2021.119926

45. Gordon B, Kelleher A, Kimball S. Regulation of muscle protein synthesis and the effects of catabolic states. *Int J Biochem Cell Biol*. (2013) 45:2147–57. doi: 10.1016/j.biocel. 2013.05.039

46. Jaiswal N, Gavin M, Loro E, Sostre-Colón J, Roberson P, Uehara K, et al. controls protein synthesis and oxidative metabolism via combined mTORC1 and FOXO1 signalling to govern muscle physiology. *J Cachexia Sarcopenia Muscle*. (2022) 13:495–514. doi: 10.1002/jcsm.12846

47. Ascenzi, F, Barberi L, Dobrowolny G, Villa Nova Bacurau A, Nicoletti C, Rizzuto E, et al. Effects of IGF-1 isoforms on muscle growth and sarcopenia. *Aging Cell.* (2019) 18:e12954. doi: 10.1111/acel.12954

48. Hata S, Mori H, Yasuda T, Irie Y, Yamamoto T, Umayahara Y, et al. A low serum IGF-1 is correlated with sarcopenia in subjects with type 1 diabetes mellitus: findings from a post-hoc analysis of the iDIAMOND study. *Diabetes Res Clin Pract.* (2021) 179:108998. doi: 10.1016/j.diabres.2021.108998

49. Hou L, Liu Y, Li X, Huo C, Jia X, Yang J, et al. Changes and risk factors of skeletal muscle mass and strength in patients with type 2 diabetes over 60 years old: a cross-sectional study from China. *J Diabetes Res.* (2020) 2020:9815485. doi: 10.1155/2020/9815485

50. Saeki C, Kanai T, Nakano M, Oikawa T, Torisu Y, Saruta M, et al. Low serum branched-chain amino acid and insulin-like growth factor-1 levels are associated with sarcopenia and slow gait speed in patients with liver cirrhosis. *J Clin Med.* (2020) 9:3239. doi: 10.3390/jcm9103239

51. Xu B, Guo Z, Jiang B, Zhang K, Zhu W, Lian X, et al. Factors affecting sarcopenia in older patients with chronic diseases. *Ann Palliat Med.* (2022) 11:972–83. doi: 10.21037/apm-22-201

52. Naro F, Venturelli M, Monaco L, Toniolo L, Muti E, Milanese C, et al. Skeletal muscle fiber size and gene expression in the oldest-old with differing degrees of mobility. *Front Physiol.* (2019) 10:313. doi: 10.3389/fphys.2019.00313

53. Park S, Seo Y, Kwon K. Sarcopenia targeting with autophagy mechanism by exercise. *BMB Rep.* (2019) 52:64–9.

54. Altun M, Besche H, Overkleeft H, Piccirillo R, Edelmann M, Kessler B, et al. Muscle wasting in aged, sarcopenic rats is associated with enhanced activity of the ubiquitin proteasome pathway. *J Biol Chem.* (2010) 285:39597–608. doi: 10.1074/jbc.M110.129718

55. Budui S, Rossi A, Zamboni M. The pathogenetic bases of sarcopenia. *Clin Cases Miner Bone Metab.* (2015) 12:22–6. doi: 10.11138/ccmbm/2015.12.1.022

56. Jo E, Lee S, Park B, Kim J. Potential mechanisms underlying the role of chronic inflammation in age-related muscle wasting. *Aging Clin Exp Res.* (2012) 24:412–22. doi: 10.3275/8464

57. Rom O, Reznick A. The role of E3 ubiquitin-ligases MuRF-1 and MAFbx in loss of skeletal muscle mass. *Free Radic Biol Med.* (2016) 98:218–30. doi: 10.1016/j. freeradbiomed.2015.12.031

58. Testa M, Cella P, Marinello P, Frajacomo F, Padilha C, Perandini P, et al. Resistance training attenuates activation of STAT3 and muscle atrophy in tumor-bearing mice. *Front Oncol.* (2022) 12:880787. doi: 10.3389/fonc.2022.880787

59. Wu H, Jang J, Dridi S, Ferrando A, Wolfe R, Kim I, et al. Net protein balance correlates with expression of autophagy, mitochondrial biogenesis, and fat metabolism-related genes in skeletal muscle from older adults. *Physiol Rep.* (2020) 8:e14575. doi: 10.14814/phy2.14575

60. Bodine S, Baehr L. Skeletal muscle atrophy and the E3 ubiquitin ligases MuRF1 and MAFbx/atrogin-1. *Am J Physiol Endocrinol Metab.* (2014) 307:E469–84. doi: 10.1152/ajpendo.00204.2014

61. Zhang Y, Wang J, Wang X, Gao T, Tian H, Zhou D, et al. The autophagic-lysosomal and ubiquitin proteasome systems are simultaneously activated in the skeletal muscle of gastric cancer patients with cachexia. *Am J Clin Nutr.* (2020) 111:570–9. doi: 10.1093/ajcn/nq2347

62. Sharma B, Dutt V, Kaur N, Mittal A, Dabur R. Tinospora cordifolia protects from skeletal muscle atrophy by alleviating oxidative stress and inflammation induced by sciatic denervation. *J Ethnopharmacol.* (2020) 254:112720. doi: 10.1016/j.jep.2020.112720

63. Spencer J, Eliazer S, Ilaria R Jr., Richardson J, Olson E. Regulation of microtubule dynamics and myogenic differentiation by MURF, a striated muscle RING-finger protein. *J Cell Biol.* (2000) 150:771–84. doi: 10.1083/jcb.150.4.771

64. Perera S, Mankoo B, Gautel M. Developmental regulation of MURF E3 ubiquitin ligases in skeletal muscle. *J Muscle Res Cell Motil.* (2012) 33:107–22. doi: 10.1007/s10974-012-9288-7

65. Lodka D, Pahuja A, Geers-Knörr C, Scheibe R, Nowak M, Hamati J, et al. Muscle RING-finger 2 and 3 maintain striated-muscle structure and function. *J Cachexia Sarcopenia Muscle*. (2016) 7:165–80. doi: 10.1002/jcsm.12057

66. Moriscot A, Baptista I, Bogomolovas J, Witt C, Hirner S, Granzier H, et al. MuRF1 is a muscle fiber-type II associated factor and together with MuRF2 regulates type-II fiber trophicity and maintenance. *J Struct Biol.* (2010) 170:344–53. doi: 10.1016/j.jsb.2010.02. 001

67. Min L, Choy E, Pollock R, Tu C, Hornicek F, Duan Z. Autophagy as a potential target for sarcoma treatment. *Biochim Biophys Acta Rev Cancer.* (2017) 1868:40–50. doi: 10.1016/j.bbcan.2017.02.004

68. Giordano F, Burattini S, Buontempo F, Canonico B, Martelli A, Papa S, et al. Diet modulation restores autophagic flux in damaged skeletal muscle cells. *J Nutr Health Aging.* (2019) 23:739–45. doi: 10.1007/s12603-019-1245-3

69. Pigna E, Berardi E, Aulino P, Rizzuto E, Zampieri S, Carraro U, et al. Aerobic exercise and pharmacological treatments counteract cachexia by modulating autophagy in colon cancer. *Sci Rep.* (2016) 6:26991. doi: 10.1038/srep26991

70. Russ D, Boyd I, McCoy K, McCorkle K. Muscle-specificity of age-related changes in markers of autophagy and sphingolipid metabolism. *Biogerontology*. (2015) 16:747–59. doi: 10.1007/s10522-015-9598-4

71. García-Prat L, Martínez-Vicente M, Perdiguero E, Ortet L, Rodriguez-Ubreva J, Rebollo E, et al. Autophagy maintains stemness by preventing senescence. *Nature*. (2016) 529:37–42. doi: 10.1038/nature16187

72. Revuelta M, Matheu A. Autophagy in stem cell aging. *Aging Cell*. (2017) 16:912–5. doi: 10.1111/acel.12655

73. Carter H, Kim Y, Erlich A, Zarrin-Khat D, Hood D. Autophagy and mitophagy flux in young and aged skeletal muscle following chronic contractile activity. *J Physiol.* (2018) 596:3567–84. doi: 10.1113/JP275998

74. Urbina-Varela R, Castillo N, Videla L, Del Campo A. Impact of mitophagy and mitochondrial unfolded protein response as new adaptive mechanisms underlying old pathologies: sarcopenia and non-alcoholic fatty liver disease. *Int J Mol Sci.* (2020) 21:7704. doi: 10.3390/ijms21207704

75. Zhang Y, Oliveira A, Hood D. The intersection of exercise and aging on mitochondrial protein quality control. *Exp Gerontol.* (2020) 131:110824. doi: 10.1016/j.exger.2019.110824

76. Davuluri G, Welch N, Sekar J, Gangadhariah M, Alchirazi KA, Mohan ML, et al. Activated protein phosphatase 2A disrupts nutrient sensing balance between mechanistic target of rapamycin complex 1 and adenosine monophosphate-activated protein kinase, causing sarcopenia in alcohol-associated liver disease. *Hepatology.* (2021) 73:1892–908. doi: 10.1002/hep.31524

77. Kny M, Fielitz J. Hidden agenda - the involvement of endoplasmic reticulum stress and unfolded protein response in inflammation-induced muscle wasting. *Front Immunol.* (2022) 13:878755. doi: 10.3389/fimmu.2022.878755

78. Hulmi J, Hentilä J, DeRuisseau K, Oliveira B, Papaioannou K, Autio R, et al. Effects of muscular dystrophy, exercise and blocking activin receptor IIB ligands on the unfolded protein response and oxidative stress. *Free Radic Biol Med.* (2016) 99:308–22. doi: 10.1016/j.freeradbiomed.2016.08.017

79. Gallot Y, Bohnert K. Confounding roles of ER stress and the unfolded protein response in skeletal muscle atrophy. *Int J Mol Sci.* (2021) 22:2567. doi: 10.3390/ ijms22052567

80. Timmerman K, Volpi E. Amino acid metabolism and regulatory effects in aging. *Curr Opin Clin Nutr Metab Care.* (2008) 11:45–9. doi: 10.1097/MCO.0b013e3282f2 a592

81. Fujita S, Volpi E. Amino acids and muscle loss with aging. J Nutr. (2006) 136(1 Suppl):277S-80S. doi: 10.1093/jn/136.1.277S

82. Drummond M, Dreyer H, Pennings B, Fry C, Dhanani S, Dillon E, et al. Skeletal muscle protein anabolic response to resistance exercise and essential amino acids is delayed with aging. *J Appl Physiol (1985)*. (2008) 104:1452–61. doi: 10.1152/japplphysiol. 00021.2008

83. Aragon A, Tipton K, Schoenfeld B. Age-related muscle anabolic resistance: inevitable or preventable? *Nutr Rev.* (2022): Epub ahead of print. doi: 10.1093/nutrit/nuac062

84. Brook M, Wilkinson D, Mitchell W, Lund J, Phillips B, Szewczyk N, et al. Synchronous deficits in cumulative muscle protein synthesis and ribosomal biogenesis underlie age-related anabolic resistance to exercise in humans. *J Physiol.* (2016) 594:7399–417. doi: 10.1113/JP272857

85. Argilés J, Busquets S, Stemmler B, López-Soriano F. Cachexia and sarcopenia: mechanisms and potential targets for intervention. *Curr Opin Pharmacol.* (2015) 22:100–6. doi: 10.1016/j.coph.2015.04.003

86. Xu Z, Tan Z, Zhang Q, Gui Q, Yang Y. The effectiveness of leucine on muscle protein synthesis, lean body mass and leg lean mass accretion in older people: a systematic review and meta-analysis. *Br J Nutr.* (2015) 113:25–34. doi: 10.1017/S000711451400 2475

87. Ma W, Zhang W, Zhai J, Li H, Zhao T, Han L, et al. Branched chain and other amino acid intakes are inversely associated with sarcopenia among community elders in Qingdao. *China. Asia Pac J Clin Nutr.* (2022) 31:97–107. doi: 10.6133/apjcn.202203_31(1).0011

88. Pascual-Fernández J, Fernández-Montero A, Córdova-Martínez A, Pastor D, Martínez-Rodríguez A, Roche E. Sarcopenia: molecular pathways and potential targets for intervention. *Int J Mol Sci.* (2020) 21:8844. doi: 10.3390/ijms21228844

89. Cava E, Fontana L. Will calorie restriction work in humans? Aging (Albany NY). (2013) 5:507–14. doi: 10.18632/aging.100581

90. van Norren K, Rusli F, van Dijk M, Lute C, Nagel J, Dijk F, et al. Behavioural changes are a major contributing factor in the reduction of sarcopenia in caloric-restricted ageing mice. *J Cachexia Sarcopenia Muscle.* (2015) 6:253–68. doi: 10.1002/jcsm.12024

91. Ruan G, Ge Y, Xie H, Hu C, Zhang Q, Zhang X, et al. Association between systemic inflammation and malnutrition with survival in patients with cancer sarcopenia-a prospective multicenter study. *Front Nutr.* (2022) 8:811288. doi: 10.3389/fnut.2021. 811288

92. Peng H, Tan X. The prognostic significance of sarcopenia and the neutrophil-tolymphocyte ratio in elderly patients with esophageal squamous cell carcinoma. *Cancer Manag Res.* (2021) 13:3209–18. doi: 10.2147/CMAR.S302274

93. Patel H, Al-Shanti N, Davies L, Barton S, Grounds M, Tellam R, et al. Lean mass, muscle strength and gene expression in community dwelling older men: findings from the hertfordshire sarcopenia study (HSS). *Calcif Tissue Int.* (2014) 95:308–16. doi: 10. 1007/s00223-014-9894-z

94. Shokri-Mashhadi N, Moradi S, Heidari Z, Saadat S. Association of circulating C-reactive protein and high-sensitivity C-reactive protein with components of sarcopenia: a systematic review and meta-analysis of observational studies. *Exp Gerontol.* (2021) 150:111330. doi: 10.1016/j.exger.2021.111330

95. Carter C, Justice J, Thompson L. Lipotoxicity, aging, and muscle contractility: does fiber type matter? *Geroscience*. (2019) 41:297–308. doi: 10.1007/s11357-019-00 077-z

96. Kob R, Bollheimer L, Bertsch T, Fellner C, Djukic M, Sieber C, et al. Sarcopenic obesity: molecular clues to a better understanding of its pathogenesis? *Biogerontology*. (2015) 16:15–29. doi: 10.1007/s10522-014-9539-7

97. Li C, Yu K, Shyh-Chang N, Jiang Z, Liu T, Ma S, et al. Pathogenesis of sarcopenia and the relationship with fat mass: descriptive review. *J Cachexia Sarcopenia Muscle*. (2022) 13:781–94. doi: 10.1002/jcsm.12901

98. Livshits G, Kalinkovich A. Inflammaging as a common ground for the development and maintenance of sarcopenia, obesity, cardiomyopathy and dysbiosis. *Ageing Res Rev.* (2019) 56:100980. doi: 10.1016/j.arr.2019.100980

99. Haran P, Rivas D, Fielding R. Role and potential mechanisms of anabolic resistance in sarcopenia. *J Cachexia Sarcopenia Muscle*. (2012) 3:157–62. doi: 10.1007/s13539-012-0068-4

100. Nishikawa H, Asai A, Fukunishi S, Nishiguchi S, Higuchi K. Metabolic syndrome and sarcopenia. *Nutrients.* (2021) 13:3519. doi: 10.3390/nu1310 3519

101. Kim J, Kwon E, Han Y. Allulose attenuated age-associated sarcopenia via regulating IGF-1 and myostatin in aged mice. *Mol Nutr Food Res.* (2022) 66: e2100549.

102. Liu P, Hu Y, Wang M, Kang L. Nutrient weight against sarcopenia: regulation of the IGF-1/PI3K/Akt/FOXO pathway in quinoa metabolites. *Curr Opin Pharmacol.* (2021) 61:136–41. doi: 10.1016/j.coph.2021.10.001