Check for updates

OPEN ACCESS

EDITED BY Daniel Tomé, AgroParisTech Institut des Sciences et Industries du Vivant et de L'environnement, France

REVIEWED BY Jorn Trommelen, Maastricht University Medical Centre, Netherlands

*CORRESPONDENCE Glenda Courtney-Martin ⊠ glenda.courtney-martin@sickkids.ca

RECEIVED 14 March 2024 ACCEPTED 06 May 2024 PUBLISHED 21 May 2024

CITATION

Paoletti A, Elango R and Courtney-Martin G (2024) Metabolic availability of amino acids in humans. *Front. Nutr.* 11:1400676. doi: 10.3389/fnut.2024.1400676

COPYRIGHT

© 2024 Paoletti, Elango and Courtney-Martin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Metabolic availability of amino acids in humans

Alyssa Paoletti¹, Rajavel Elango^{2,3,4} and Glenda Courtney-Martin^{1,5,6}*

¹Research Institute, Hospital for Sick Children, Toronto, ON, Canada, ²Department of Pediatrics, University of British Columbia, Vancouver, BC, Canada, ³School of Population and Public Health, University of British Columbia, Vancouver, BC, Canada, ⁴BC Children's Hospital Research Institute, BC Children's Hospital, Vancouver, BC, Canada, ⁵Department of Nutritional Sciences, University of Toronto, Toronto, ON, Canada, ⁶Department of Kinesiology, University of Toronto, Toronto, ON, Canada

Knowledge of amino acid bioavailability and the effect of combining complementary protein sources are required to determine how to best meet an individual's protein and indispensable amino acid needs. Traditionally, protein quality of foods has been assessed using digestibility data. Digestibility may overestimate bioavailability of some amino acids particularly those more susceptible to heat and processing. The indicator amino acid oxidation (IAAO) method has been validated and applied to determine amino acid bioavailability termed metabolic availability of the first limiting amino acid of a proteinaceous food. The metabolic availability of the limiting amino acid in the test protein is determined as a ratio of the indicator amino acid oxidation response to graded intakes of the test protein compared to the indicator response to a reference protein (crystalline amino acid patterned after egg protein). The IAAO method has also been applied to assess the effect of protein complementation directly in humans on the overall protein quality of the diet. The results demonstrate that protein complementation augments the limiting amino acid supply and increases protein synthesis.

KEYWORDS

protein quality, metabolic availability, amino acids, indicator amino acid oxidation, slope ratio assay

1 Introduction

The nutritive value of proteins depends on their amino acid content and their bioavailability (1). Therefore, to understand the extent to which dietary proteins can meet the requirements for indispensable amino acids, the requirement for each indispensable amino acid and their bioavailability in food proteins must be known at each stage of the life cycle. Digestibility and bioavailability are often used interchangeably, based on the assumption that if an amino acid is digested and absorbed, it is available to the body for protein synthesis. *Digestibility* refers to the net absorption of amino acid across the intestine whereas *bioavailability* refers to the proportion of the total amino acids that are digested and absorbed in a form suitable for protein synthesis (2). Using growth response and amino acid retention in growing pigs Batterham et al. demonstrated that bioavailability was lower than ileal digestibility for lysine, threonine, methionine and tryptophan (2) due to their increased sensitivity to heat and processing. In the quantity normally consumed, plant protein sources may fail to fulfil the requirements for these indispensable amino acid which makes them the *limiting amino acid* in plant-based diets (3). For those consuming a strict plant-based diet, the

limiting amino acid will restrict the body's capacity to build proteins (4). Therefore, the limiting amino acid and its bioavailability are primary determinants of a food's protein quality.

The provision of complementary protein sources each containing a limiting amino acid in the same meal, helps to increase the content of that amino acid in the diet. This is known as *protein complementation*, a traditional and practical strategy used to improve the protein quality of plant-based diets (4). Thus, an ideal tool for protein quality assessment should possess the capacity to evaluate the effectiveness of complementation so that recommendations can be made regarding appropriate ratios required to optimize protein quality (5) of the diet. This review will discuss the application of the IAAO method in the determination of protein quality of foods for human consumption with a focus on its novel application to assess the effects of protein complementation directly in humans.

2 Evaluation of protein quality using the slope ratio-IAAO method

The IAAO method has been applied to the study of protein quality of foods for human consumption by measuring the bioavailability termed metabolic availability (6, 7) of the first limiting amino acid in foods. The food under study is first analyzed for its protein and amino acid composition to identify the first limiting amino acid. Below the limiting amino acid requirement, the decrease in IAAO will be linear with increasing intakes of the limiting amino acid (Figure 1). This patterned IAAO response can be used to measure metabolic availability of a proteinaceous food containing a limiting amino acid because decreased oxidation of the indicator amino acid is inversely related to protein synthesis (5, 8). In order to do so, the IAAO response is combined with *slope-ratio* principles, which can be considered the gold standard against which other bioavailability methods are judged (2).

Thus, the IAAO response to intakes of the test limiting amino acid from a food (protein-bound amino acid) is measured and compared to the IAAO response to the same intakes of the test limiting amino acid from a reference protein. The reference protein is provided as free, crystalline amino acid since it has been shown that the true digestibility of crystalline amino acids in pigs is essentially 100% (10). Therefore, the relative ratio of the IAAO response in the food compared to the IAAO response in the reference protein is proportional to whole-body bioavailability of the limiting amino acid (5–7). It captures both absorption, and availability of the amino acid for metabolic processes (i.e., protein synthesis). Hence, any dietary losses of amino acids during these biological processes are accounted for (5–7).

2.1 Validating the application of the IAAO method for protein quality evaluation

The application of the IAAO method to the study of protein quality was first validated in piglets (6) in which the bioavailability of lysine from raw and heated peas were assessed. The test diets (raw and heated peas) supplied lysine below its requirement (11) to ensure that the IAAO response was linear with increasing intakes of dietary lysine. The IAAO response to feeding the raw peas and heated peas were compared to the IAAO response to feeding free lysine (reference diet) separately to estimate the metabolic availability of lysine from each diet. Free lysine was then added back to the heated peas to assess whether the method was sensitive to the effects of lysine supplementation. The bioavailability of lysine from raw and heated peas were determined to be 88 and 55%, respectively (6). These estimates were comparable to estimates obtained by Van Barneveld and colleagues using a slope-ratio growth assay (12). When free lysine was added back to the heated peas, a decline in the IAAO response was observed demonstrating increased metabolic availability of lysine for protein synthesis (6). This was similarly demonstrated by Van Barneveld and colleagues in which the addition of free lysine to heated peas reversed the growth depression in piglets caused by the heated peas (12). Overall, the distinct IAAO response to either raw peas,



heated peas, or heated peas plus free lysine demonstrates that phenylalanine oxidation directly reflects the bioavailability of lysine and that the changes in oxidation were due to changes in metabolically available dietary lysine for protein synthesis. It also highlights the ability of the method to capture the effects of heat processing which is important to human nutrition in which many foods are processed (i.e., subject to the effects of heat or chemical treatment) before consumption.

2.2 Criteria required for measuring protein quality using the IAAO method

To quantify the bioavailability of a limiting amino acid in a food, important criteria must be met. First, the test amino acid must be the first limiting amino acid responsible for driving the changes in the IAAO response (5) and must be the only limiting amino acid in the diet. This is achieved by providing a diet patterned after the amino acid composition of egg protein. Thus, all amino acids are fed in excess of their requirements except for the test amino acid. The indicator amino acid (i.e., most commonly phenylalanine in the presence of excess tyrosine) is also provided above its requirement and is held constant throughout all diets to ensure indicator oxidation is due only to changes in the intakes of the test amino acid. The dispensable amino acid alanine is used to balance the nitrogen content of the diets as the intakes of the test amino acid is varied (5). For protein quality studies, the indicator amino acid is provided at intakes around $30\,\text{mg}\cdot\text{kg}^{-1}\cdot$ d⁻¹ of ¹³C-phenylalanine in the presence of excess tyrosine: $40 \text{ mg} \cdot$ $kg^{-1} \cdot d^{-1}$ (5).

The IAAO response to changes in the dietary intake of the test amino acid (in $mg \cdot kg^{-1} \cdot d^{-1}$) must be linear (statistical validity) to permit the calculation of bioavailability according to the principles of the slope-ratio assay (13). This is accomplished by providing intakes of the limiting amino acid that fall below the lower confidence limit of its dietary requirement (6, 14–16). Therefore, it is necessary to know the dietary requirement of the test amino acid determined by the IAAO method *a priori* to appropriately select intakes located on the linear portion of the IAAO slope (Figure 1). Additionally, the bioavailability of the limiting amino acid in the food is measured relative to a reference protein which is fed as free crystalline amino acids. This reference protein is assumed to be 100% bioavailable (10) which means that the IAAO reference slope signifies the maximal unit increase in protein synthesis (5).

To compare the IAAO response of the test protein to the IAAO response of the reference protein, the slopes require a common intercept (fundamental validity) (13). Thus, the design of the study diets must be carefully considered. For example, it is possible to test $0 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$ of the limiting amino acid from the reference diet (free crystalline amino acid) and observe a biologically meaningful IAAO response. However, since amino acid containing foods are the test source of the limiting amino acid from the test food. Therefore, the IAAO response from the reference diet at $0 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$ could not be compared to the IAAO response to feeding $0 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$ from the test food. To reconcile this statistical issue, a base amount of the crystalline form of the limiting amino acid is added to all the diets, both reference and test. This basal amount of the test assumes that

the IAAO response is the same at this basal intake which enables the two regression lines (reference and test) to intersect at a common intercept (13).

The IAAO response must demonstrate good repeatability to permit accurate quantification of metabolic availability. This is achieved with a repeated measures design which reduces intra individual variation. The repeated measures design is made possible by the use of the minimally invasive IAAO protocol which combines a short 2 day adaptation (17) period to a 9h fed-state oxidation day (14-16, 18, 19). Despite this, a major drawback of the IAAO method is that only one amino acid is evaluated at a time, rendering the procedure time consuming and expensive compared to the other available digestibility-based methods. However, dietary intakes are tightly controlled, thus, low variation (i.e., CV = <10%) in the IAAO derived bioavailability estimates have been obtained (14, 18, 19). The IAAO combined slope-ratio method has been successfully applied in humans to assess the protein quality of a variety of commonly consumed grains and pulses (14, 18, 19). Most recently, IAAO method has been adapted and applied to study the effectiveness of protein complementation directly in humans.

3 Novel application of IAAO to assess protein complementation in humans

Knowledge of the limiting amino acid content and its bioavailability from plant proteins determined using the IAAO combined slope-ratio method permits the direct quantification of the individual amounts of each individual plant foods required to fulfil an individual's protein and amino acid needs. In this way, the premise on which protein complementation is based is met by ensuring adequate quantities of all amino acids are present at the site of protein synthesis (3). Thus, in the context of a meal, the goal is to augment the limiting amino acid intake by combining two complementary plant proteins; one limiting and the other sufficient in that amino acid. For example, pulses like chickpeas are limiting in methionine (20) whereas rice contains sufficient methionine. To effectively complement a chickpeabased diet with a grain such as rice, knowledge on the bioavailability of methionine in both protein sources is required.

3.1 Rice and chickpea mixed meal

The IAAO method was first applied to the study of protein complementation in humans, by combining chickpeas and rice in a mixed meal (14). In that study, the separately determined bioavailability of methionine from steamed rice (bioavailability = 100%) and cooked chickpeas (bioavailability = 63%) were used to design a mixed-meal of cooked rice plus chickpea (18). The aim was to enhance the methionine intake from chickpea protein by adding methionine from rice protein. The impact of complementation was assessed by comparing the IAAO responses at each methionine intake from the cooked chickpeas alone to that of the IAAO responses at the same methionine intakes from the cooked chickpea plus rice after adjusting for the bioavailability of methionine in chickpeas. When this was done, a decline in IAAO (6–14%) at each methionine intake was observed compared to the IAAO responses of methionine intakes from the cooked chickpeas alone (18). This demonstrates an increase

Food	Amino acid	Population	Metabolic availability (%)	Complementary food	% Decrease in indicator oxidation	Reference
Soy protein isolate	Methionine	Young adult males	71.8			(21)
Casein	Methionine	Young adult males	87.4			(21)
Cooked white rice	Lysine	Young adult males	97			(14)
Oven browned white rice	Lysine	Young adult males	70			(14)
Cornmeal	Lysine	Young adult males	71			(15)
Cornmeal	Tryptophan	Young adult males	80			(15)
Millet	Lysine	Young adult males	97	Lentils	27	(19)
Lentils	Lysine	Young adult males	80			(19)
Sorghum	Lysine	Young adult males	94	Lentils	19	(16)
Chickpeas	Methionine	Young adult males	63	Rice	14	(18)
Rice	Methionine	Young adult males	100			(18)
Lentils	Methionine	Young adult males	69	Rice	16	(22)
Warm rice	Lysine	School-aged children	97.5			(23)

TABLE 1 Metabolic availability values and the effect of protein complementation derived using the IAAO method in humans.

in net protein synthesis due to the increased intake of methionine from the complementation of the two protein sources. Thus, more methionine is provided in the mixed diet that is available from chickpea alone reflecting superior protein quality of the mixed meal. The effect of protein complementation by combining millet (limiting in lysine but sufficient in methionine) with lentils (sufficient in lysine but limiting in methionine) have also been assessed (19).

The metabolic availability values obtained using the IAAO method and the results of the protein complementation studies are outlined in Table 1.

4 Conclusion

The IAAO method was validated in pigs and successfully applied in humans to determine the metabolic availability of the limiting amino acid in plant foods and to assess the effect of protein complementation. Nonetheless, the IAAO method has some limitations. As mentioned above, only one indispensable amino acid is evaluated at a time, rendering the procedure time consuming and expensive to study several foods. Another limitation concerns protein sources with more than one limiting amino acid in which case the bioavailability of each limiting amino acid will need to be determined separately. However, the method is non-invasive which makes it suitable for application in vulnerable populations like children, pregnant women and older adults. A key advantage is IAAO measures metabolic availability rather than digestibility alone, and hence can be useful for assessing the effect of processing and in developing reference values for major food protein sources and in the validation of new techniques for estimating amino acid bioavailability.

Author contributions

AP: Writing – original draft. RE: Writing – review & editing. GC-M: Writing – review & editing, Conceptualization, Funding acquisition, Project administration, Resources, Supervision.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by Canadian Institutes for Health Research (RE & GCM) (Grant no. 201603PJT).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Tome D. Criteria and markers for protein quality assessment – a review. *Br J Nutr.* (2012) 108:S222–9. doi: 10.1017/S0007114512002565

2. Batterham ES. Availability and utilization of amino acids for growing pigs. *Nutr Res Rev.* (1992) 5:1–18. doi: 10.1079/NRR19920004

3. Block RJ, Mitchell HH. The correlation of the amino-acid composition of proteins with their nutritive value. *Nutr Abstr Rev.* (1946) 16:249–78.

4. Institute of Medicine. Dietary reference intakes: the essential guide to nutrient requirements. In: Otten JJ, Pitzi Hellwig J and Meyers LD, editors. Washington, D.C.: The National Academies Press (2006).

5. FAO. Research approaches and methods for evaluating the protein quality of human foods. Rome: Report of an FAO Expert Working Group (2014).

6. Moehn S, Bertolo RFP, Pencharz PB, Ball RO. Development of the Indicator amino acid oxidation technique to determine the availability of amino acids from dietary protein in pigs. J Nutr. (2005) 135:2866–70. doi: 10.1093/jn/135.12.2866

7. Elango R, Levesque C, Ball RO, Pencharz PB. Available versus digestible amino acids – new stable isotope methods. *Br J Nutr.* (2012) 108:S306–14. doi: 10.1017/S0007114512002498

8. Ball RO, Bayley HS. Influence of dietary protein concentration on the oxidation of phenylalanine by the Young pig. *Br J Nutr.* (1986) 55:651–8. doi: 10.1079/BJN19860071

9. Levesque CL, Moehn S, Pencharz PB, Ball RO. Review of advances in metabolic bioavailability of amino acids. *Livest Sci.* (2010) 133:4–9. doi: 10.1016/j. livsci.2010.06.013

10. Chung TK, Baker DH. Apparent and true amino acid digestibility of a crystalline amino acid mixture and of casein: comparison of values obtained with Ileal-cannulated pigs and Cecectomized cockerels. *J Anim Sci.* (1992) 70:3781–90. doi: 10.2527/1992.70123781x

11. Bertolo RF, Moehn S, Pencharz PB, Ball RO. Estimate of the variability of the lysine requirement of growing pigs using the Indicator amino acid oxidation technique. *J Anim Sci.* (2005) 83:2535–42. doi: 10.2527/2005.83112535x

12. Van Barneveld RJ, Batterham ES, Skingle DC, Norton BW. The effect of heat on amino acids for growing pigs. 3. The availability of lysine from heat-treated field peas (*Pisum Sativum* cultivar Dundale) determined using the slope-ratio assay. *Br J Nutr.* (1995) 73:259–73. doi: 10.1079/bjn19950028

13. Littell RC, Lewis AJ, Henry PR. Statistical evaluation of bioavailability assays. In: *Bioavailability of nutrients for animals: amino acids, minerals, and vitamins*. Ammerman CB, Baker DH and Lewis AJ, editors. San Diego, California: Academic Press, Inc. (1995). 5–33.

14. Prolla IRD, Rafii M, Courtney-Martin G, Elango R, da Silva LP, Ball RO, et al. Lysine from cooked white Rice consumed by healthy Young men is highly metabolically available when assessed using the Indicator amino acid oxidation technique. *J Nutr.* (2013) 143:302–6. doi: 10.3945/jn.112.166728

15. Rafii M, Elango R, Ball RO, Pencharz PB, Courtney-Martin G. Metabolic availability of the limiting amino acids lysine and tryptophan in cooked white African cornmeal assessed in healthy Young men using the Indicator amino acid oxidation technique. *J Nutr.* (2018) 148:917–24. doi: 10.1093/jn/nxy039

16. Paoletti A, Fakiha A, Tul-Noor Z, Pencharz PB, Levesque CL, Ball RO, et al. Bioavailable lysine assessed using the Indicator amino acid oxidation method in healthy Young males is high when Sorghum is cooked by a moist cooking method. *J Nutr.* (2022) 152:770–8. doi: 10.1093/jn/nxab410

17. Szwiega S, Pencharz PB, Ball RO, Xu L, Kong D, Elango R, et al. Length of adaptation has no effect on the threonine requirement determined in healthy Young adult males using the Indicator amino acid oxidation method. *J Nutr.* (2023) 153:2016–26. doi: 10.1016/j.tjnut.2023.03.033

18. Rafii M, Pencharz PB, Ball RO, Tomlinson C, Elango R, Courtney-Martin G. Bioavailable methionine assessed using the Indicator amino acid oxidation method is greater when cooked chickpeas and steamed Rice are combined in healthy Young men. *J Nutr.* (2020) 150:1834–44. doi: 10.1093/jn/nxaa086

19. Fakiha A, Tul-Noor Z, Paoletti A, Pencharz PB, Ball RO, Levesque CL, et al. Bioavailable lysine, assessed in healthy Young men using Indicator amino acid oxidation, is greater when cooked millet and stewed Canadian lentils are combined. *J Nutr.* (2020) 150:2729–37. doi: 10.1093/jn/nxaa227

20. Nosworthy MG, Neufeld J, Frohlich P, Young G, Malcolmson L, House JD. Determination of the protein quality of cooked Canadian pulses. *Food Sci Nutr.* (2017) 5:896–903. doi: 10.1002/fsn3.473

21. Humayun MA, Elango R, Moehn S, Ball RO, Pencharz PB. Application of the Indicator amino acid oxidation technique for the determination of metabolic availability of sulfur amino acids from casein versus soy protein isolate in adult men. *J Nutr.* (2007) 137:1874–9. doi: 10.1093/jn/137.8.1874

22. Rafii M, Pencharz PB, Boileau K, Ball RO, Tomlinson C, Elango R, et al. Metabolic availability of methionine assessed using Indicator amino acid oxidation method, is greater when cooked lentils and steamed Rice are combined in the diet of healthy Young men. *J Nutr.* (2022) 152:1467–75. doi: 10.1093/jn/nxac049

23. Caballero K, Mandal R, Pratap-Singh A, Kitts DD, Ball RO, Pencharz PB, et al. Lysine Bioavailability in School-Age Children Consuming Rice Is Reduced by Starch Retrogradation. *J Nutr.* (2020) 150:3208–3215. doi: 10.1093/jn/nxaa276