Check for updates

OPEN ACCESS

EDITED BY Ignacio Cabezudo, CONICET Rosario, Argentina

REVIEWED BY Ahmed Noah Badr, National Research Centre, Egypt Debomitra Dey, Anton Paar USA Inc, United States

*CORRESPONDENCE Han Lü Xiaohan1814@163.com Jian Chen Chenjian80@aliyun.com

[†]These authors have contributed equally to this work and share first authorship

RECEIVED 17 December 2024 ACCEPTED 16 April 2025 PUBLISHED 07 May 2025

CITATION

Chen H, Zhang F, Yang YX, Meng XH, Ding XQ, Jian TY, Niu GT, Tong B, Gai YN, Lü H and Chen J (2025) Effect of chitosan edible coating containing anthocyanins and tea polyphenols on cold storage of chilled pork. *Front. Nutr.* 12:1546618. doi: 10.3389/fnut.2025.1546618

COPYRIGHT

© 2025 Chen, Zhang, Yang, Meng, Ding, Jian, Niu, Tong, Gai, Lü and Chen. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Effect of chitosan edible coating containing anthocyanins and tea polyphenols on cold storage of chilled pork

Han Chen^{1†}, Fang Zhang^{2†}, Yi-xin Yang², Xiu-hua Meng¹, Xiao-qin Ding¹, Tun-yu Jian¹, Guan-ting Niu¹, Bei Tong¹, Ya-nan Gai¹, Han Lü^{2*} and Jian Chen^{1*}

¹Jiangsu Key Laboratory for the Research and Utilization of Plant Resources, Institute of Botany, Jiangsu Province and Chinese Academy of Sciences (Nanjing Botanical Garden Mem. Sun Yat-Sen), Nanjing, China, ²School of Pharmacy, Nanjing University of Chinese Medicine, Nanjing, China

Preserving fresh meat has been a long-standing challenge due to microbial growth and lipid oxidation. In this study, chitosan (CS) active coatings loaded with blackberry anthocyanins (BA), tea polyphenols (TP), and their combination, designated as CS/BA, CS/TP, and CS/BA/TP, were applied for chilled pork preservation. Compared to CS coating alone, CS/BA/TP coating exhibited a 9.3% lower total viable count (TVC), a 45.5% reduction in thiobarbituric acid reactive substances (TBARS), and a 26.6% decrease in total volatile basic nitrogen (TVB-N) after 12 days of storage at 4°C. Furthermore, the combined use of TP and BA exhibited a synergistic effect, with the most pronounced impact observed in oxidative stability, while also improving microbial inhibition and maintaining color stability. These results highlight CS/BA/TP coatings as a promising natural preservative for extending chilled pork shelf life.

KEYWORDS

chitosan, anthocyanins, tea polyphenols, edible coatings, pork preservation

Introduction

Preservation of fresh meat is a long-standing challenge due to its plentiful nutrients, which promote the growth of spoilage microorganisms and food-borne pathogens (19). Refrigeration storage is widely used to slow lipid oxidation and spoilage in meat. However, chilled meat remains susceptible to microbial contamination during storage and transport (20).

Synthetic petrochemical-based polymers have been widely used in meat packaging materials; however, due to their environmental pollution and the migration of toxic substances, biodegradable packaging materials have garnered significant interest in recent years (1). Edible films and coatings are thin layers used for food packaging and are made from edible components. Natural polysaccharides are excellent sources for edible coatings due to their biodegradable and inexpensive properties (2). Chitosan (CS) is obtained from chitin through deacetylation in an alkaline medium. It is a copolymer made up of β -(1–4)-2-acetamido-D-glucose and β -(1–4)-2-amino-D-glucose units, with the latter typically making up more than 60% of the composition. CS is an exceptional edible material due to its effective barrier properties against oxygen (O₂) and carbon dioxide (CO₂), along with excellent antimicrobial characteristics (21). However, its limited biological activity necessitates the incorporation of bioactive compounds, such as polyphenols, to enhance its functional properties (3).

In recent years, studies have confirmed that incorporating natural polyphenols into biodegradable food packaging films can boost their antioxidant and antibacterial properties, thereby enhancing their food preservation capabilities (22). Tea polyphenols (TP) are natural polyphenols derived from tea and belong to the flavanol group. In addition to being an effective food additive, TP functions as an antioxidant, antibacterial agent, and improving the performance of active food packaging films (4). Anthocyanins, a significant class of phenolic compounds found in nature, have been shown to enhance the physical and chemical properties of active films (23). Blackberry is a globally popular fruit rich in anthocyanins, with cyanidin-3-O-glucoside being the most prominent. Research has demonstrated that blackberry anthocyanins exhibit strong antioxidant and antibacterial properties (24). In our previous study, we found that combining blackberry anthocyanins (BA) and tea polyphenols (TP) effectively delayed lipid oxidation in edible oils during high-temperature processing (5). Recent studies have shown that chitosan coatings combined with TP and rosemary extract effectively extended the shelf life of large yellow croaker (6) and that chitosan-grape peel extract coatings reduced lipid oxidation and bacterial growth in beef during freeze-thaw cycles (7).

However, the effect of edible coatings containing TP and BA on meat preservation has not been thoroughly investigated. In this study, chitosan (CS) active coatings loaded with BA, TP, and a combination of both were applied for the preservation of pork. During storage, color parameters, pH value, total volatile basic nitrogen, total viable counts, and thiobarbituric acid reactive substances were measured to assess the effectiveness of the coatings in maintaining pork freshness. We hypothesize that integrating BA and TP into CS coatings will exhibit a synergistic effect, enhancing the antioxidant and antimicrobial properties of the coatings and extending the shelf life of chilled pork.

Materials and methods

Chemical and reagents

Chitosan powder (CS, ≥ 90 % degree of deacetylation) was obtained from Sangon Biotech (Shanghai) Co., Ltd. (Shanghai, China). Blackberry anthocyanins (BA) was prepared in our previous work. Tea polyphenol (TP, ≥ 98 % purity) was purchased from Yuanye Bio-Technology Co., Ltd. (Shanghai, China). The chemicals used in this study were analytical grade and obtained from Macklin Biochemical Co., Ltd. (Shanghai, China).

Preparation of chitosan coatings and pork samples

Based on our preliminary study, CS (2.25% w/v) was dispersed in a 1% (v/v) hydrochloric acid (HCl) solution under magnetic stirring at room temperature for 2 h to obtain CS solution, which appeared transparent and slightly yellowish. BA (0.44 wt%) and TP (6.67 wt%), dissolved in water, were individually added to the CS solution under magnetic stirring at room temperature for 1 h, based on the total weight of CS, to obtain the CS/BA and CS/TP solutions, respectively. The CS/BA solution exhibited a reddishpurple hue, while the CS/TP solution appeared light brown. The CS/BA/TP solution were prepared by adding mixed BA (0.44 wt%) and TP (6.67 wt%) solution to the CS solution under magnetic stirring at room temperature for 1 h, resulting in a dark purplishbrown solution. Subsequently, glycerol (1% v/v) was added under magnetic stirring to obtain the final coating solutions.

The devices and work surfaces were sterilized with 75% ethanol and exposed to ultraviolet (UV) light for 0.5 h before the experiment. The fresh pork tenderloin purchased from a local supermarket (Nanjing, China) was cut into steaks under sterile conditions and divided into five groups. The pork samples were individually dipped into the prepared coating solutions (CS, CS/BA, CS/TP, CS/BA/TP) for 60 s, then air-dried to form a coating. Uncoated pork samples were treated as control (Control). All samples were placed in sterile trays, sealed with plastic wrap and stored at $4 \pm 1^{\circ}$ C with a relative humidity of 80% in darkness for 12 days.

Color parameters

The color parameters [L* (lightness), a* (redness), and b* (yellowness)] of the pork samples were measured at three random locations on each surface of the sample at $4 \pm 1^{\circ}$ C using a portable spectrophotometer (NR10QC, Shenzhen ThreeNH Technology Co., Ltd., China). The total color difference (ΔE_t) was calculated according to Equation 1:

$$\Delta E_t = \sqrt{\left(L^* - L_0\right)^2 + \left(a^* - a_0\right)^2 + \left(b^* - b_0\right)^2} \tag{1}$$

where L_0 , a_0 and b_0 are the color parameters of day 0. L*, a* and b* are the color parameters of the samples at different storage times (3, 6, 9, and 12days) (8).

pH value

The pH was determined according to the Chinese national standard GB 5009.237-2016 (27). One gram of the sample was added to 10 g of 0.1 M potassium chloride solution. Then the mixture was homogenized, and the pH was measured using a digital pH meter (FiveEasy Plus, METTLER TOLEDO, USA). Three measurements were taken on each meat sample, and the average was recorded as the pH of the meat sample. Before use, the pH meter was calibrated with standard buffer solutions at pH 4.00, 6.86 and 9.18, respectively.

Total volatile basic nitrogen (TVB-N) value

TVB-N value of the pork was measured according to the Chinese national standard GB 5009.228-2016 (26). Ten gram of sample was homogenized with 50 mL of distilled water and filtered after impregnating for 30 min. One milliliter of boric acid solution and 1 drop of mixed indicator were added into the central chamber of the diffusion dish. Simultaneously, the 1 mL of filtrate

and 1 mL of saturated sodium carbonate solution were added into the outer chamber of the diffusion dish. All solution were mixed evenly and stored in the sealed diffusion dish at $37 \pm 1^{\circ}$ C for 2 h. The solution in the central chamber was then titrated with hydrochloric acid. The TVB-N value of each sample was determined based on triplicate measurements, and volatile nitrogen bases were calculated in terms of milligrams of nitrogen per 100 g of the sample using the following equation:

 $TVB - N (mg/100g) = C \times (V - V_0) \times 14 \times 5000/W$

Where V is the volume of hydrochloric acid consumed, V_0 is the volume of hydrochloric acid consumed in the titration process of blank (ml), W is the weight of meat sample (g), and C is the concentration of hydrochloric acid (g/mol).

Thiobarbituric acid reactive substances value

The lipid oxidation of pork was evaluated by measuring TBARS according to the Chinese national standard GB 5009.181-2016 (25). Briefly, 5 g of ground sample was blended and homogenized with 50 mL of 7.5% trichloroacetic acid containing 0.1% EDTA, and stirred at 50°C for 30 min using a magnetic stirrer. After cooling to room temperature, the homogenate was filtered with doublelayer quantitative slow filter paper. The thiobarbituric acid (TBA) reagent was prepared by dissolving 0.2883 g of TBA in 100 mL of distilled water to obtain a 0.02 M solution. Then, 5 mL of filtrate was transferred to a screw cap test tube, and after adding with 5 mL of 0.02 M TBA solution, the solution was reacted in a water bath at $90^\circ\mathrm{C}$ for 30 min and analyzed optically at 532 nm using a spectrophotometer. The content of malondialdehyde in the sample was calculated according to the standard curve of malondialdehyde, referred to as the TBARS value, and expressed as milligrams of malondialdehyde per kilogram of sample (mg MDA/kg).

Microbial analysis

Total viable count (TVC) was measured by the plate count agar (PCA) method to assess microbial growth in samples at 0, 3, 6, 9 and 12 days of refrigerated storage duration. Meat samples (5 g) were homogenized with 45 mL of 0.1% peptone water. Ten-fold serial dilutions of each sample homogenate were made and applied to agar plates, which were then incubated at 37 $^{\circ}$ C for 48 h. The results were recorded as log CFU (colony-forming units) per gram of meat sample.

Statistical analysis

Statistical analysis was performed using GraphPad Prism version 8 (GraphPad Software, Inc., CA, USA) and Origin version 2019b (OriginLab Corporation, USA). Each independent analysis was performed in triplicate (n = 3) for which results were expressed as the mean \pm standard deviation. Date collected were analyzed

using two-way-analysis of variance (ANOVA; treatment × storage) followed by Duncan's test to compare differences between means of parameters at the 5% significance level. Differences were considered significant at p < 0.05.

In this study, the independent variables were coating treatment (CS, CS/BA, CS/TP, and CS/BA/TP) and storage time (0, 3, 6, 9, and 12 days). The dependent variables included color parameters (L, a, b*), pH, total volatile basic nitrogen (TVB-N), total viable counts (TVC), and thiobarbituric acid reactive substances (TBARS), which were used to assess the effectiveness of the coatings in preserving pork quality.

Results and discussion

Color parameters

Meat color is an important indicator for consumer choice. As shown in Figure 1, with increased storage time, the color of control pork samples darkened, and water loss and dry shrinkage occurred. Bacterial colonies appeared on day 9, with more colonies appearing on day 12. In comparison, the changes in the different coating groups were less significant, and no obvious colonies appeared.

At day 0, after coating with different films, the brightness (L* value) of chilled pork samples increased (Figure 2A). The L* value of the control group increased on day 3 and then decreased rapidly, dropping from 57.22 \pm 1.61 to 42.30 \pm 1.38 by day 12. The alteration in L* value in the CS coating samples were similar to that in the control group, but the extent was lower. The changes in L* values for the CS/BA, CS/TP, and CS/BA/TP coating samples were smaller, with the L* values in the CS/BA/TP samples remaining relatively stable (p > 0.05) over 12 days. This indicates that the CS/BA/TP coating can help maintain the light value of pork.

Compared with the control group, coating with CS increased the redness index (a* values) of pork (Figure 2B), while the a* values of the CS/BA, CS/TP, and CS/BA/TP coating samples were lower than those of the control sample on day 0, indicating that the addition of BA and TP may influence the redness index of the meat. The a* values of all pork samples decreased significantly with storage time, suggesting that these coatings have no obvious effect on maintaining the red color of pork.

With the increase in storage time, the b* value of the control group samples increased gradually (Figure 2C), which is consistent with the meat color changing from red to brown. Lipid oxidation in meat may transform white fat into a yellow color. The formation of yellow pigments affected by lipid oxidation in meat is related to the non-enzymatic browning reactions of lipid oxidation products (28). All coatings decreased the yellowness b* values of pork compared to control samples on day 0. In comparison, the increase in b* values of the coating samples ware smaller than that of the control group, with the CS/BA/TP group showing the least increase. The CS coating could significantly inhibit the increase of yellowness value, which may be related to its oxygen barrier ability, thereby retarding lipid oxidation (21). The addition of BA or TP increased the effectiveness of CS in inhibiting the increase in b* value, with the simultaneous addition of both BA and TP showing the best effect. This improvement may be due to the antioxidant properties



of BA and TP, which may enhance the anti-lipid oxidation effects of the CS coating.

The total color difference (ΔE_t) between days (3 vs. 1, 6 vs. 1, 9 vs. 1 and 12 vs.1) for five batches (Control, CS, CS/BA, CS/TP, and CS/BA/TP) was calculated (Figure 2D). Most remarkable differences were observed on day 9 vs. 1 (CS > CS/TP> Control > CS/BA > CS/BA/TP). The ΔE_t value of the CS/BA/TP group was always the lowest among all groups, which indicated that the CS/BA/TP coating promoted the color stability of pork during storage.

Microbiological analyses

Microbiological growth is the most important factor influencing the changes that cause spoilage in meat. The total viable count (TVC) is an important microbial index to evaluate the hygienic quality, safety and spoilage of meat. As shown in Figure 3A, the TVC values of all coating groups were significantly lower than those of the control sample at 0 days. Throughout the storage period, the TVC values of all samples gradually increased, but the coating groups consistently exhibited significantly lower TVC values compared to the control group (p < 0.05). After the 9-day storage period, the TVC of control group samples were 7.09 \pm 0.04 log CFU/g, up to the threshold defined for the quality of pork (29). While all of the coating samples were below this threshold after the 12-day storage period, indicating that coating may contributed to extend the meat shelf-life by limiting the oxygen availability for microbial growth (9). At the end of the storage period, the TVC of the CS/BA/TP coating was 6.21 \pm 0.15 log CFU/g, significantly lower than that of CS, CS/BA, and CS/TP (p < 0.05). Similar to other composite chitosan coatings for preservation, the incorporation of one or more active substances demonstrated superior antibacterial performance (4, 10).

The observed antimicrobial effects of the coatings align with previous reports on chitosan (30), and both TP and BA also demonstrate antibacterial properties (24, 31). The peptidoglycan layer and teichoic acid are key components of Gram-positive bacterial cell walls, and TP has been reported to disrupt the peptidoglycan structure, leading to bacterial inhibition (11). Furthermore, studies suggest that polyphenol-based compounds



Changes of color parameters including L* (A), a*(B), b* values (C) and ΔE_t (D) of pork samples with different packaging groups during cold storage (4°C).



can enhance bacterial damage when combined with other bioactive agents, demonstrating a synergistic antibacterial effect (9). This experiment further confirmed the antibacterial effects of TP and BA in meat preservation. This result is consistent with findings from other studies on pork preservation (4, 10, 12). The active coating loaded with both BA and TP was more effective in reducing the TVC of pork during storage than other coatings, suggesting a synergistic effect in preventing microbial spoilage.

TVB-N value

During meat spoilage, the degradation of proteins and other nitrogen (N)-containing compounds causes the accumulation of organic amines, commonly known as total volatile basic nitrogen (TVB-N), which are toxic and can lead to considerable changes in color and flavor, seriously affecting the quality of the meat. Therefore, TVB-N content is an important factor in evaluating meat quality (32).

As shown in Figure 3B, all groups exhibited a gradual increase in TVB-N with extended storage time, which may be related to the proliferation of microorganisms that accelerate the decomposition of pork proteins and produce amines (33). Throughout the entire storage period, the TVB-N value of the control showed a significant increasing trend. In contrast, all coatings significantly delayed the increase in TVB-N value (p < 0.05) compared to the control samples. According to the results of the microbiological analyses above, CS coating may suppress microbial growth, and the addition of BA and TP can enhance this effect. All of these factors may lead to lower production of TVB-N in meat. It was suggested that a TVB-N value of 15 mg/100 g may serve as the upper threshold for acceptable freshness of pork. The TVB-N value of the control sample reached this threshold on day 9, indicating that its shelf life is no more than 9 days. TVB-N values of the CS and CS/BA coating samples did not reach the threshold until day 12. However, the TVB-N values of the CS/TP and CS/BA/TP coatings were still below the threshold value on day 12, indicating that their shelf life was extended to 12 days.

pH values

As shown in Figure 3C, the pH value of the control group significantly increased throughout the entire storage period. Compared to the control group, there was no significant change in pH in any of the coating groups within the first 6 days. The pH of the CS group significantly increased on day 9, while there was no significant change in the CS/BA group, and the CS/TP and CS/BA/TP coating groups showed a slight decrease in pH. All coating groups exhibited an increase in pH by the 12th day, but the pH of the CS/BA and CS/TP or CS/BA/TP coating groups was significantly lower than that of the CS coating group. The pH increase in the control group can be attributed to protein denaturation and the buildup of organic amines, such as ammonia, amines, and trimethylamine, which are generated during the degradation of amino acids through autolytic or microbial processes (34). Studies have shown that chitosan-gelatin coating

can create a protective barrier on the surface of pork, reduce the exchange of substances between the meat and the surrounding environment, and slow down the propagation of microorganisms and the decomposition rate of proteins to some extent (4). The differences in pH trends among the coating groups may be attributed to the distinct mechanisms of BA and TP. BA contains organic acids, which may contribute to a direct pH-lowering effect (13), whereas TP exhibits stronger antimicrobial properties, effectively inhibiting microbial metabolism and acid production, leading to a more gradual pH decline (14).

The pH of all coating samples in this experiment is significantly lower than that of the control group, which supports the previous researches. Moreover, the pH growth rates of the CS/BA, CS/TP, and CS/BA/TP groups were significantly lower than that of the CS group (p < 0.05). This indicates that the addition of BA, TP, and BA/TP effectively inhibited the increase in pH of chilled pork, likely due to the further inhibition of protein decomposition and microbial reproduction in the meat by the addition of BA and TP had a synergistic effect in inhibiting the increase in pH of chilled pork.

Lipid oxidation

Pork has a high fat content and is highly susceptible to lipid peroxidation. The resulting oxidative changes, including alterations in flavor, color, texture, and nutritional value are significant factors that contribute to quality deterioration during storage. Our previous research has demonstrated that BA and TP exhibit strong scavenging activity against ABTS and DPPH free radicals (5). TBARS assay was used to evaluate the lipid oxidation levels in different samples, with higher TBARS values indicating elevated levels of lipid oxidation. Studies have shown that the TBARS value of fresh meat should be controlled within 1.00 mg MDA/kg, as exceeding this threshold leads to noticeable oxidative deterioration and negatively impacts sensory quality (15). As shown in Figure 3D, the TBARS value of the control group increased rapidly with storage time. The TBARS values of the CS, CS/BA, and CS/TP coating groups showed a slow increase during the first 3 days and slightly decreased by the 6th day. This may be because CS, BA, and TP had not yet fully exerted their effects in the early stage of coating and were unable to fully inhibit the acceleration of the lipid oxidation process. After 3 days, the CS, CS/BA, and CS/TP coating solutions tightly bound to the pork samples, reducing oxygen permeability, and the antioxidant components began to take effect, slowing down the lipid oxidation process and leading to a decrease in TBARS values. After day 6, all coating groups exhibited a rapid increase in TBARS values, but the CS/BA/TP group maintained a significantly lower oxidation rate than the other groups. By the 9th day, both the control and CS groups had exceeded the limit of 1.00 mg MDA/kg, while the CS/TP, CS/BA, and CS/BA/TP groups remained below this limit. However, by the 12th day, all groups except the CS/BA/TP group had exceeded the limit of 1.00 mg MDA/kg, indicating that the coating with the combined addition of BA and TP had a synergistic effect in slowing lipid oxidation. This synergy may be attributed to the complementary antioxidant mechanisms of BA and TP, where BA donates hydrogen atoms to scavenge free radicals, while TP enhances oxidative stability through metal ion chelation and radical regeneration (16). Consequently, the CS/BA/TP coating effectively inhibited TBARS accumulation, significantly extending the shelf life of chilled pork to 12 days.

Conclusion

In this study, chitosan (CS) active coatings loaded with blackberry anthocyanins (BA) and tea polyphenols (TP) and a combination of both were tested for their effects on pork preservation. Compared to the CS coating alone, the CS/BA/TP coating exhibited a 9.3% lower total viable count (TVC), a 45.5% reduction in thiobarbituric acid reactive substances (TBARS), and a 26.6% decrease in total volatile basic nitrogen (TVB-N) after 12 days of storage at 4°C. The combined addition of TP and BA demonstrated a synergistic effect, further enhancing antimicrobial and antioxidant properties. These results suggest that CS coatings incorporating TP and BA effectively extend the shelf life of chilled pork to 12 days, making them a promising natural preservation strategy.

While synthetic preservatives and modified atmosphere packaging (MAP) are widely used in the meat industry, natural biobased coatings like CS/BA/TP provide a clean-label alternative with strong antimicrobial and antioxidant properties (17).

Although the use of BA and TP may slightly increase production costs, their ability to extend shelf life, reduce spoilage, and minimize reliance on synthetic additives could offset these costs. Furthermore, the ease of application and biodegradability of CS-based coatings align with the growing demand for sustainable and natural food preservation solutions, supporting their potential for commercial adoption (18).

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

Author contributions

HC: Data curation, Investigation, Writing - original draft, Writing - review & editing. FZ: Data curation, Investigation, Writing – original draft, Writing – review & editing. YY: Formal analysis, Writing – original draft. XM: Methodology, Writing – original draft. XD: Visualization, Writing – original draft. TJ: Methodology, Writing – original draft. GN: Resources, Writing – original draft. BT: Methodology, Writing – original draft. YG: Resources, Writing – original draft. HL: Funding acquisition, Project administration, Supervision, Writing – review & editing, Writing – original draft. JC: Project administration, Supervision, Writing – review & editing, Writing – original draft.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This research was financially supported by grants from the Primary Research and Development Plan of Jiangsu Province (Nos. BE2022371 and BE2020344) and the National Natural Science Foundation of China (No. 32170377). Support was also received from Jiangsu Agricultural Science and Technology Innovation Fund (No. CX(23)3138) and Jiangsu Key Laboratory for Conservation and Utilization of Plant Resources (No. JSPKLB202315). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative Al statement

The author(s) declare that no Gen AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

^{1.} Yin Y, Woo MW. Transitioning of petroleum-based plastic food packaging to sustainable bio-based alternatives. *Sustain Food Technol.* (2024) 2:548–66. doi: 10.1039/D4FB00028E

^{2.} Kocira A, Kozłowicz K, Panasiewicz K, Staniak M, Szpunar-Krok E, Hortyńska P. Polysaccharides as edible films and coatings: Characteristics and influence on fruit and vegetable quality—a review. *Agronomy*. (2021) 11:813. doi: 10.3390/agronomy110 50813

^{3.} Wang Z, Zhang M, Liang S, Li Y. Enhanced antioxidant and antibacterial activities of chitosan/zein nanoparticle Pickering emulsion-incorporated chitosan coatings in the presence of cinnamaldehyde and tea polyphenol. *Int J Biol Macromol.* (2024) 266:131181. doi: 10.1016/j.ijbiomac.2024.131181

^{4.} Zhang H, Li X, Kang H, Peng X. Effect of tannic acid-grafted chitosan coating on the quality of fresh pork slices during cold storage. *Meat Sci.* (2022) 188:108779. doi: 10.1016/j.meatsci.2022.108779

5. Chen H, Zhao H-f, Meng X-h, Chen J, Wu W-l, Li W-l, et al. Effect of blackberry anthocyanins and its combination with tea polyphenols on the oxidative stability of lard and olive oil. *Front Nutr.* (2023) 10:1286209. doi: 10.3389/fnut.2023.1286209

6. Li T, Hu W, Li J, Zhang X, Zhu J, Li X. Coating effects of tea polyphenol and rosemary extract combined with chitosan on the storage quality of large yellow croaker (*Pseudosciaena crocea*). *Food Control.* (2012) 25:101-6. doi: 10.1016/j.foodcont.2011.10.029

7. Cao Y, Song Z, Dong C, Yu Q, Han L. Chitosan coating with grape peel extract: a promising coating to enhance the freeze-thaw stability of beef. *Meat Sci.* (2023) 204:109262. doi: 10.1016/j.meatsci.2023.109262

8. Vargas-Ramella M, Lorenzo JM, Zamuz S, Valdés ME, Moreno D, Balcázar MCG, et al. The antioxidant effect of colombian berry (*Vaccinium meridionale* Sw.) extracts to prevent lipid oxidation during pork patties shelf-life. *Antioxidants* (2021) 10:1290. doi: 10.3390/antiox10081290

9. Wang J, Chen J, Sun Y, He J, Zhou C, Xia Q, et al. Ultraviolet-radiation technology for preservation of meat and meat products: recent advances and future trends. *Food Control.* (2023) 148:109684. doi: 10.1016/j.foodcont.2023.109684

10. Fang Z, Lin D, Warner RD, Ha M. Effect of gallic acid/chitosan coating on fresh pork quality in modified atmosphere packaging. *Food Chem.* (2018) 260:90–6. doi: 10.1016/j.foodchem.2018.04.005

11. Karinou E, Schuster CF, Pazos M, Vollmer W, Gründling A. Inactivation of the monofunctional peptidoglycan glycosyltransferase SgtB allows *Staphylococcus aureus* to survive in the absence of lipoteichoic acid. *J Bacteriol.* (2019) 201:e00574-18. doi: 10.1128/JB.00574-18

12. Zhang H, He P, Kang H, Li X. Antioxidant and antimicrobial effects of edible coating based on chitosan and bamboo vinegar in ready to cook pork chops. *LWT*. (2018) 93:470–6. doi: 10.1016/j.lwt.2018.04.005

13. Veberic R, Stampar F, Schmitzer V, Cunja V, Zupan A, Koron D, et al. Changes in the contents of anthocyanins and other compounds in blackberry fruits due to freezing and long-term frozen storage. *J Agric Food Chem.* (2014) 62:6926–35. doi: 10.1021/jf405143w

14. Yang Y, Zhang T. Antimicrobial activities of tea polyphenol on phytopathogens: a review. *Molecules.* (2019) 24:816. doi: 10.3390/molecules24040816

15. Ali Ghoflgar Ghasemi M, Hamishehkar H, Javadi A, Homayouni-Rad A, Jafarizadeh-Malmiri H. Natural-based edible nanocomposite coating for beef meat packaging. *Food Chem.* (2024) 435:137582. doi: 10.1016/j.foodchem.2023.137582

16. Xu X, Li F, Zhang X, Li P, Zhang X, Wu Z, et al. *In vitro* synergistic antioxidant activity and identification of antioxidant components from *Astragalus membranaceus* and *Paeonia lactiflora*. *PLoS ONE*. (2014) 9:e96780. doi: 10.1371/journal.pone.0096780

17. Czerwiński K, Rydzkowski T, Wróblewska-Krepsztul J, Thakur VK. Towards impact of modified atmosphere packaging (MAP) on shelf-life of polymer-film-packed food products: challenges and sustainable developments. *Coatings*. (2021) 11:1504. doi: 10.3390/coatings11121504

18. Bhalani DV, Nutan B, Singh Chandel AK. Sustainable Development of Membranes Encyclopedia of Green Materials. Springer: New York. (2024). p.1745-54. doi: 10.1007/978-981-97-4618-7 223

19. Aymerich T, Picouet PA, Monfort JM. Decontamination technologies for meat products. *Meat Sci.* (2008) 78:114–29. doi: 10.1016/j.meatsci.2007.07.007

20. Shao L, Chen S, Wang H, Zhang J, Xu X, Wang H, et al. Advances in understanding the predominance, phenotypes, and mechanisms of bacteria related to meat spoilage. *Trends Food Sci Technol.* (2021) 118:822–32. doi: 10.1016/j.tifs.2021.11.007

21. Elsabee MZ, and Abdou ES. Chitosan based edible films and coatings: a review. *Mater Sci Eng C.* (2013) 33:1819–41. doi: 10.1016/j.msec.2013.01.010

22. Biao Y, Yuxuan C, Qi T, Ziq, Y, Yourong Z, McClements DJ, et al. Enhanced performance and functionality of active edible films by incorporating tea polyphenols into thin calcium alginate hydrogels. *Food Hydrocoll*. (2019) 97:105197. doi: 10.1016/j.foodhyd.2019.105197

23. Qin Y, Xu F, Yuan L, Hu H, Yao X, Liu J, et al. Comparison of the physical and functional properties of starch/polyvinyl alcohol films containing anthocyanins and/or betacyanins. *Int J Biol Macromol.* (2020) 163:898–909. doi: 10.1016/j.ijbiomac.2020.07.065

24. Wu H, Di QR, Zhong L, Zhou JZ, Shan CJ, Liu XL, et al. Enhancement on antioxidant, anti-hyperglycemic and antibacterial activities of blackberry anthocyanins by processes optimization involving extraction and purification. *Front Nutr.* (2022) 9:1007691. doi: 10.3389/fnut.2022.10 07691

25. PRC-National-Standard. Determination of Malondialdehyde in Food of National Standard for Food Safety. GB 5009.181-2016 (2016).

26. PRC-National-Standard. Determination of Volatile Base Nitrogen in Food of National Standard for Food Safety. GB 5009.228-2016 (2016).

27. PRC-National-Standard Determination of pH Value in Food of National Standard for Food Safety. GB 5009.237-2016 (2017).

28. Rodríguez-Carpena JG, Morcuende D, Estévez M. Avocado by-products as inhibitors of color deterioration and lipid and protein oxidation in raw porcine patties subjected to chilled storage. *Meat Sci.* (2011) 89:166–73. doi: 10.1016/j.meatsci.2011.04.013

29. Huang L, Zhao J, Chen Q, Zhang Y. Rapid detection of total viable count (TVC) in pork meat by hyperspectral imaging. *J Food Res Int.* (2013) 54:821–8. doi: 10.1016/j.foodres.2013.08.011

30. Leceta I, Guerrero P, Ibarburu I, Dueñas MT, DE La Caba K. Characterization and antimicrobial analysis of chitosan-based films. *J Food Eng.* (2013) 116:889–99. doi: 10.1016/j.jfoodeng.2013.01.022

31. Amarowicz R, Pegg RB, Dykes GA, Troszynska A, Shahidi F. Antioxidant and Antibacterial Properties of Extracts of Green Tea Polyphenols. Washington, DC: ACS Publications (2005). doi: 10.1021/bk-2005-0909.ch009

32. Bekhit AE-DA, Holman BW, Giteru SG, Hopkins DL. Total volatile basic nitrogen (TVB-N) and its role in meat spoilage: a review. *Trends Food Sci Technol.* (2021) 109:280-302. doi: 10.1016/j.tifs.2021.01.006

33. Hussain MA, Sumon TA, Mazumder SK, Ali MM, Jang WJ, Abualreesh MH, et al. Essential oils and chitosan as alternatives to chemical preservatives for fish and fisheries products: a review. *Food Control* (2021) 129:108244. doi:10.1016/j.foodcont.2021.108244

34. Lorenzo JM, Batlle R, Gómez M. Extension of the shelf-life of foal meat with two antioxidant active packaging systems. *LWT-Food Sci.* (2014) 59:181–8. doi: 10.1016/j.lwt.2014.04.061

80