Check for updates

OPEN ACCESS

EDITED BY Michael Gänzle, University of Alberta, Canada

REVIEWED BY Per Erik Joakim Saris, University of Helsinki, Finland Hongzhi Wu, Chinese Academy of Tropical Agricultural Sciences, China

*CORRESPONDENCE Yifei Lu ⊠ luyf8893@enzemed.com Xiaoshu Chen ⊠ 13506662820@139.com

[†]These authors have contributed equally to this work

RECEIVED 06 April 2025 ACCEPTED 15 May 2025 PUBLISHED 29 May 2025

CITATION

Zou H, Zhu T, Chen X and Lu Y (2025) Dietary live microbe intake and its association with Parkinson's disease in American adults: an NHANES analysis (1999–2018). *Front. Nutr.* 12:1606922. doi: 10.3389/fnut.2025.1606922

COPYRIGHT

© 2025 Zou, Zhu, Chen and Lu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Dietary live microbe intake and its association with Parkinson's disease in American adults: an NHANES analysis (1999–2018)

He Zou^{1†}, Tao Zhu^{2†}, Xiaoshu Chen^{1*} and Yifei Lu^{2*}

¹Department of Cardiology, Wenzhou Third Clinical Institute Affiliated to Wenzhou Medical University, Wenzhou People's Hospital, Wenzhou, China, ²Department of Cardiology, Taizhou Hospital of Zhejiang Province Affiliated to Wenzhou Medical University, Taizhou, China

Background: Diet plays a crucial role in sustaining a healthy body, and microbes have attracted significant scholarly interest in recent years as an essential component of diet. This study aims to explore the association between dietary live microbe intake and the incidence of Parkinson's Disease (PD) among adults over 40 years old.

Methods: A total of 26,033 subjects in NHANES 1999–2018 were included in this study, comprising 314 patients with PD, which were divided into three groups based on their intake of foods with different levels of microbial content: high, medium, and low. The assessment on subjects' dietary live microbe intake was conducted through the self-reported questionnaire.

Results: The distribution of subjects based on dietary live microbe intake was as follows: 34.5% had a low intake, 45.4% had a medium intake, and 20.1% had a high intake. A multivariable logistic regression analysis revealed that subjects with high (OR = 0.52, 95% CI: 0.37–0.74) and medium (OR = 0.73, 95% CI: 0.57–0.93) dietary live microbe intake exhibits a reduced prevalence of PD compared to those with low intake. Restricted cubic spline (RCS) analysis indicated a significant linear association between dietary live microbe intake and the prevalence of PD. Furthermore, stratified analyses demonstrate that the association among research variables was more pronounced in subjects without obesity.

Conclusion: This study demonstrates an inverse linear association between dietary live microbe intake and the prevalence of PD.

KEYWORDS

NHANES, Parkinson's disease, dietary live microbe, MedHi, obesity

Introduction

Parkinson's disease (PD) manifests as a clinical syndrome typified by symptoms, such as cogwheel rigidity, bradykinesia, postural instability, slow shuffling gait, and resting tremors (1). As the second most prevalent neurodegenerative disorder followed by Alzheimer's disease, PD constitutes a significant societal concern and poses a critical global health challenge. Projections indicate that the incidence of PD will double over the next 30 years (2–5), with the increasing affected subjects from 6.9 million in 2015 to 14.2 million by the end of 2040 (6). Understanding the risk factors and pathogenesis of PD is crucial for improving the prognosis of those impacted by the disease and their quality of life.

The etiology of PD has been widely recognized as multifactorial, involving an interplay of environmental and genetic influences (7, 8). Among these environmental determinants, diet has emerged as a significant factor. It is well established that microorganisms are prevalent in various foods consumed, such as natural foods and fermented products (9). These microorganisms influence both the preservation and flavor profile of food and are intricately associated with health conditions (10). Growing evidence has shown that gut microbiota may be the important contributor to central nervous system (CNS) dysfunction (11, 12). Gut microbiota alterations can increase both intestinal and blood-brain barrier permeabilities, leading to a continuous accumulation of gut microbiota-derived molecules and metabolites in brain, thereby promoting neuroinflammation in CNS (13). Gut dysbiosis has been found to be strongly associated with the pathology of PD in all clinical and preclinical studies (14–17). Alterations in microbiome induced microglial activation and neuroinflammation in mice overexpressing α -Syn, in turn promoting motor dysfunction of mice (16).

Dietary live microbes see those microorganisms capable of surviving in the gastrointestinal tract and exerting beneficial effects, including probiotic bacteria and other advantageous microorganisms (probiotic candidates) (18). These microbes may prevent the colonization of exogenous pathogenic bacteria within the gastrointestinal tract by promoting the homeostasis of intestinal microbiota and inhibiting harmful bacteria. And metabolites produced by the colonization of microbes in the gastrointestinal tract will also neutralize and inhibit the production of harmful bacterial toxins, reducing local and systemic inflammatory reactions in human body (19). Substantial research has established probiotics contributing to keep healthy by modulating immune function and gut microbiota (20, 21), with observed benefits in various conditions, including digestive disorders, chronic liver disease, chronic kidney disease, Alzheimer's disease, and PD (22-25). Furthermore, an increased intake of synbiotics and probiotics has been demonstrated to alter gut microbial composition, thereby supporting healthy aging (26-29).

In summary, PD has been considered to be associated with the dysbiosis of systemic inflammation and gut microbiota, suggesting that dietary live microbes may serve as a potential target for the management and prevention of PD. While previous studies have predominantly centered on the advantages of probiotic supplements, there is a significant gap in the research on the exploration of the impact of total intake derived from natural foods. Dietary live microbes exist in various foods, including unpeeled fruits and vegetables, as well as fermented dairy products (30). These natural foods are easy to obtain, inexpensive, and can be consumed regularly. Given that consuming dietary live microbes through food sources might provide a comprehensive and sustainable means to enhancing immune function and gut health, it is crucial to explore the association between dietary live microbes and PD. Based on previous studies that estimated dietary live microbe intake based on the viable bacteria per gram of food (31, 32), this study examined the dietary live microbe intake in American adults from NHANES 1999-2018 and explored its association with PD in American adults. We hypothesized that higher intakes of dietary live microbes would be associated with a lower risk of PD (see Figure 1).

Methods

Research subjects and design

NHANES, a comprehensive research initiative conducted biennially by NCHS, aims to assess the association among disease prevention, nutrition, and health promotion (33), by taking a series of interviews and physical examinations, encompassing a wide range of sections including dietary, demographic, and laboratory data, as well as various health assessments.

In this study, a retrospective analysis was conducted on the data from 10 cycles of the NHANES dataset spanning the period from 1999 to 2018. Subjects aged 40 years old and older (n = 36,252) were included. Based on predefined exclusion criteria, subjects with incomplete data for dietary information required to assess dietary live microbe intake (n = 4,255) were excluded. Specifically, these subjects lacked the information on the types or quantities of foods consumed, which is essential for estimating microbial content accurately. Additionally, subjects without data on PD status (n = 18,610) were excluded, as PD diagnosis was a critical variable for this study. Pregnant females (n = 28)were excluded due to the unique physiological and dietary changes during pregnancy, which could confound the association between dietary live microbe intake and PD. Lastly, subjects with missed covariate data (n = 5,900), including demographic information, health conditions, or lifestyle factors, were excluded to ensure the completeness and reliability of the data for multivariable analysis. As a result, the final sample comprised 26,033 subjects, as illustrated in Figure 2.

Assessment on dietary live microbe intake

In this study, the data on dietary intake of live microbes were obtained through the estimation of live bacterial quantities (per gram) conducted by four domain experts (MES, MLM, CH, and RH). These experts evaluated 9,388 food codes across 48 subgroups within the NHANES database (30). The estimation process employed 24-h dietary recall data from the United States Department of Agriculture (USDA) and was cross-referenced with corresponding food codes in the NHANES database, ensuring alignment with the food and nutrient database (31, 32). The microbial content was categorized into three levels: High (>107 CFU/g), medium (104-107 CFU/g), and low (<10⁴ CFU/g). Foods characterized by low microbial content predominantly consist of heat-treated or pasteurized items, including milk, gravies, seafood, prepared meats, and sauces. In contrast, foods with medium microbial content were primarily unpeeled fresh vegetables and fruits, which inherently harbored moderate levels of microorganisms. Additionally, foods with high microbial content encompass unpasteurized fermented products, such as kefir, yogurt, and kimchi, in addition to probiotic supplements.

Subjects were stratified into three distinct groups according to microbial content in the diet: low intake (all consumed foods demonstrated low microbial content), medium intake (any consumed food demonstrated medium but not high microbial content), and high intake (any consumed food demonstrated high microbial content). Moreover, subjects were divided into three categories based on the intake of MedHi foods, which quantified the dietary live microbe intake: Group 1 including those who did not consume any MedHi foods; Group 2 including those whose MedHi food intake was above



zero but below the median level; and Group 3 including those whose MedHi food intake was above the median level.

Diagnosis of PD

In this study, subjects with PD were ascertained by pinpointing those prescribed "Anti-Parkinson Agents" based on the subjects' selfreported prescription medication usage. Given the limitations of the NHANES dataset concerning available medication codes, the diagnosis of PD was attributed solely to those currently receiving treatment. Consequently, those who did not report anti-Parkinsonian medication were deemed as non-PDs, defining PD aligned with the diagnostic criteria, which has been employed in previous studies (34, 35).

Covariates

Based on previous studies (36) and variables provided by NHANES, covariates of interest including self-reported health conditions, physical examination results, and sociodemographic variables, were identified as potential confounding factors. Trained researchers collected self-reported health information and sociodemographic variables, including smoking status, total energy intake, past medical history, and alcohol abuse during MEC and family interviews. Sociodemographic variables included education level, age, marital status, gender, PIR, and race. Physical examination results, including blood pressure and BMI, were collected by trained health technicians at MEC. Additionally, the definitions for diabetes, hypertension, and hyperlipidemia were consistent with those previously reported (37). The incidence of stroke, coronary heart disease (CHD), and cancers was determined through self-reported diagnoses. Alcohol abuse was characterized by taking at least 12 alcoholic beverages within 1 year. Smoking was defined by taking a minimum of 100 cigarettes over lifetime. The assessment on probiotic supplement intake was carried out through the dietary supplement questionnaire (30).

Statistical analysis

The normality of continuous variables was assessed by representing them as either the median with interquartile range or mean \pm SD. Through multivariable logistic regression analyses, the association between dietary live microbe intake and PD was explored across three progressively adjusted models: Model 1 (unadjusted), Model 2 (accounted for gender, race, and age) and Model 3 (extensively adjusted for an array of factors, including gender, age, race, BMI, physical activities, diabetes, education level, hypertension, smoking status, CHD, stroke, PIR, total energy intake, cancers, hyperlipidemia, marital status, and alcohol abuse). The heterogeneity between dietary live microbe



intake and PD was explored through interactions and subgroup analyses on various variables. In addition, RCS regression analysis was conducted to assess the linearity and explore the dose–response association between dietary live microbe intake and PD, adjusting for covariates included in Model 3. During the sensitivity analyses, multiple logistic regression analyses were performed to further explore the association between dietary live microbe intake and PD, including extra adjustments for the intake of non-food prebiotics and probiotics. The statistical assessments were carried out with Free Statistics software (Version 2.0, China) and R software (Version 4.1.1, Vienna, Austria).

Results

Demographic characteristics

Table 1 delineates the baseline characteristics of all subjects, categorized based on the absence or presence of PD. Among the

cohort, 314 patients (1.2%) were diagnosed with PD, with the average age of 60 years old, including 13,192 (50.7%) females. In terms of dietary live microbe intake classified by varying microbial content, 34.5% of adults had a low intake, 45.4% had a medium intake, and 20.1% had a high intake. According to the MedHi classification, 34.5, 32.5, and 32.9% of the adults were categorized into groups as G1, G2, and G3, respectively. Subjects with PD were more likely to be older, non-Hispanic White, living alone, with a higher BMI, a lower household income, and a combination of hypertension, CHD, and stroke (p < 0.05). The proportion of subjects with a high dietary live microbe intake was significantly lower among PD subjects compared to non-PD subjects (p < 0.05). The basic attributes of the subjects, divided into high, medium, and low intake groups, as well as G3, G2, and G1 groups, are shown in Supplementary Tables S1, S2. Notably, the high intake group had a lower prevalence of PD (p < 0.05).

Association between dietary live microbe intake and PD

Table 2 presents the findings from logistic regression analyses examining the association between varying levels of dietary live microbe intake and the prevalence of PD. Upon accounting for all potential confounders, the analysis reveals a substantial decrease in the prevalence of PD among subjects with high (OR = 0.52, 95% CI: 0.37–0.74) and medium (OR = 0.73, 95% CI: 0.57–0.93) intake, compared to those with a low intake. Additionally, subjects were stratified into three groups based on MedHi intake to quantify the dietary live microbe intake. In the totally adjusted model, the prevalence of PD was found to be 31% lower in the G2 group (OR = 0.64, 95% CI: 0.52–0.90) and 36% lower in the G3 group (OR = 0.64, 95% CI: 0.48–0.85), compared to the G1 group. RCS analyses further illustrated a significant negative linear association between dietary live microbe intake and the prevalence of PD (Figure 3).

Stratified analyses

To assess the stability of the association between dietary live microbe intake and PD, subgroup analyses and interaction tests were performed across various factors, including gender, age, and a history of diabetes, CHD, hypertension, hyperlipidemia, and stroke. These analyses did not reveal any significant association among these variables. Notably, the inverse association between a high dietary live microbe intake and the prevalence of PD was more pronounced in subjects without obesity, as illustrated in Figures 4, 5.

Sensitivity analyses

To mitigate the potential confounding effects of non-dietary prebiotic and probiotic use, adjustments were incorporated into models (see Table 3). The analysis indicated a substantially lower prevalence of PD in the high intake group compared to the low intake group (OR = 0.85, 95% CI: 0.76–0.94). Moreover, the prevalence of PD was 47% lower in the G3 group than in the G1 group (OR = 0.53, 95% CI: 0.37–0.75).

TABLE 1 Baseline characteristics of the study participants.

Characteristics	Total (<i>n</i> = 26,033)	Non-PD (<i>N</i> = 25,719)	PD (<i>n</i> = 314)	<i>p</i> -value
Age, year	60.0 ± 12.6	60.0 ± 12.6	64.1 ± 13.2	<0.001
Gender, <i>n</i> (%)				0.504
Male	12,841 (49.3)	12,692 (49.3)	149 (47.5)	
Female	13,192 (50.7)	13,027 (50.7)	165 (52.5)	
PIR	2.68 ± 1.62	2.68 ± 1.62	2.29 ± 1.55	<0.001
BMI, kg/m ²	29.3 ± 6.5	29.3 ± 6.5	30.0 ± 6.9	0.044
Race, <i>n</i> (%)				<0.001
Mexican American	4,245 (16.3)	4,213 (16.4)	32 (10.2)	
Other Hispanic	2045 (7.9)	2021 (7.9)	24 (7.6)	
Non-Hispanic White	12,816 (49.2)	12,618 (49.1)	198 (63.1)	
Non-Hispanic Black	5,328 (20.5)	5,278 (20.5)	50 (15.9)	
Other Race	1,599 (6.1)	1,589 (6.2)	10 (3.2)	
Physical activity, <i>n</i> (%)				0.101
Low	22,049 (84.7)	21,783 (84.7)	266 (90.5)	
High	3,984 (15.3)	3,936 (15.3)	48 (9.5)	
Marital group, <i>n</i> (%)				0.002
Married or living with partner	17,337 (66.6)	17,154 (66.7)	183 (58.3)	
Widowed or divorced	6,780 (26.0)	6,685 (26)	95 (30.3)	
Never married	1916 (7.4)	1880 (7.3)	36 (11.5)	
Education level, <i>n</i> (%)				0.154
Less than high school	7,673 (29.5)	7,569 (29.4)	104 (33.1)	
High school or above	18,360 (70.5)	18,150 (70.6)	210 (66.9)	
Smoking status, n (%)				0.516
Current or ever	13,123 (50.4)	12,959 (50.4)	164 (52.2)	
Never	12,910 (49.6)	12,760 (49.6)	150 (47.8)	
Drinking status, n (%)				0.430
Current or ever	17,781 (68.3)	17,573 (68.3)	208 (66.2)	
Never	8,252 (31.7)	8,146 (31.7)	106 (33.8)	
Diabetes, n (%)	5,021 (19.3)	4,948 (19.2)	73 (23.2)	0.073
Hypertension, <i>n</i> (%)	12,148 (46.7)	11,972 (46.5)	176 (56.1)	<0.001
Hyperlipidemia, n (%)	20,017 (76.9)	19,780 (76.9)	237 (75.5)	0.550
CHD, n (%)	1,633 (6.3)	1,605 (6.2)	28 (8.9)	0.049
Stroke, <i>n</i> (%)	1,317 (5.1)	1,285 (5)	32 (10.2)	<0.001
Cancer, <i>n</i> (%)	3,486 (13.4)	3,434 (13.4)	52 (16.6)	0.097
Total energy intake, kcal	1976.6 ± 910.2	1977.7 ± 911.0	1882.7 ± 843.8	0.066
Category of MedHi ^a				<0.001
Low	8,990 (34.5)	8,853 (34.4)	137 (43.6)	
Medium	11,808 (45.4)	11,674 (45.4)	134 (42.7)	
High	5,235 (20.1)	5,192 (20.2)	43 (13.7)	
Category of MedHi ^b				<0.001
G1	8,990 (34.5)	8,853 (34.4)	137 (43.6)	
G2	8,472 (32.5)	8,381 (32.6)	91 (29)	
G3	8,571 (32.9)	8,485 (33)	86 (27.4)	

Values are mean ± SD, median (IQR) or number (%). BMI, body mass index; PIR, poverty-income ratio; CHD, coronary heart disease.

*Participants were also classified into three different groups considering the general intake of foods with varying contents of microbe: low (all foods consumed were Low); moderate (any foods consumed were Medium but not High); and high (any foods consumed were High).

^bParticipants were categorized into three groups based on the MedHi consumption to quantify the ingestion of live microbes: G1, consumers without intakes of any MedHi food; G2, those with intakes of MedHi food above zero but below the median level for consumers; G3, those with intakes of MedHi food above the median level for consumers.

TABLE 2 Association between dietary live microbe intake and prevalence of PD.

Subgroups	Mode	el 1	Mode	l 2	Model 3					
	OR (95%CI)	P-value	OR (95%CI)	P-value	OR (95%CI)	P-value				
Per100 unit increase	0.92 (0.85 ~ 0.99)	0.032	0.90 (0.83 ~ 0.98)	0.012	0.92 (0.84 ~ 0.99)	0.033				
Category of MedHi ^a										
Low	1(Ref)		1(Ref)		1(Ref)					
Medium	0.74 (0.58 ~ 0.94)	0.015	0.69 (0.54 ~ 0.88)	0.003	0.73 (0.57 ~ 0.93)	0.010				
High	0.54 (0.38 ~ 0.76)	<0.001	0.49 (0.34 ~ 0.69)	< 0.001	0.52 (0.37 ~ 0.74)	< 0.001				
P for trend	0.73 (0.63 ~ 0.86)	<0.001	0.69 (0.59 ~ 0.82)	< 0.001	0.72 (0.61 ~ 0.85)	< 0.001				
Category of MedHi ^b										
G1	1(Ref)		1(Ref)		1(Ref)					
G2	0.70 (0.54 ~ 0.92)		0.65 (0.50 ~ 0.86)	0.002	0.69 (0.52 ~ 0.90)	0.007				
G3	0.65 (0.50 ~ 0.86)	< 0.001	0.60 (0.46 ~ 0.79)	< 0.001	0.64 (0.48 ~ 0.85)	0.002				
<i>P</i> for trend	0.80 (0.70 ~ 0.92)	<0.001	0.77 (0.67 ~ 0.88)	<0.001	0.79 (0.69 ~ 0.91)	0.001				

Model 1: None covariates were adjusted.

Model 2: gender, race and age were adjusted.

Model 3, gender, age, race, BMI, physical activities, diabetes, education level, hypertension, smoking status, CHD, stroke, cancer, PIR, hyperlipidemia, marital status, drinking status, total energy intake were adjusted.

^aParticipants were also classified into three different groups considering the general intake of foods with varying contents of microbe: low (all foods consumed were Low); moderate (any foods consumed were Medium but not High); and high (any foods consumed were High).

^bParticipants were categorized into three groups based on the MedHi consumption to quantify the ingestion of live microbes: G1, consumers without intakes of any MedHi food; G2, those with intakes of MedHi food above zero but below the median level for consumers; G3, those with intakes of MedHi food above the median level for consumers.

with intakes of MedHi food above zero but below the median level for consumers; G3, those with intakes of MedHi food above the median level for consume

Discussion

In this study, the data were gathered from a sample of 26,033 adults. After comprehensive analysis of the data, it was revealed that within the model adjusted for all confounding variables, the prevalence of PD decreased by 48 and 27% in the high and medium dietary live microbe intake groups, respectively, compared to the low intake group. RCS analyses demonstrated a linear and inverse association between dietary live microbe intake and the prevalence of PD, which was particularly pronounced among subjects without obesity. Sensitivity analyses further confirmed the reliability and consistency of these findings.

Numerous scholars have explored the impact of dietary live microbe intake on human health. Hill et al. (38) established an association between diet abundant in dietary live microbes and reductions in blood glucose levels, systolic blood pressure, inflammatory markers, and TG levels. Similarly, Han and Wang (39) reported that a higher dietary live microbe intake could potentially lower the risk of cardiovascular diseases. In addition, Wang et al. (32) found an inverse association between dietary live microbe intake and the incidence of depression. Moreover, those regularly consuming foods rich in dietary live microbes tended to display the improved cognitive performance (40). Numerous studies also have explored the potential benefits of probiotic supplements. Tamtaji et al. (41) proposed that an increased intake of prebiotics and probiotics may enhance gut flora homeostasis and reduce motor severity ratings in subjects with PD. Jin et al. (42) supported the hypothesis that prebiotics and probiotics may alleviate constipation symptoms in this population, through an analysis on 11 randomized controlled trials involving 765 PD patients. A recent meta-analysis conducted in 2023 suggested that probiotics may alleviate the depression and both motor and non-motor symptoms in subjects with PD (43). Similarly, it was the first time to find that subjects with a higher dietary live microbe intake may exhibit a reduced prevalence of PD.

The negative association between dietary live microbe intake and PD is more pronounced in non-obese individuals, which may be attributed to several key factors. Firstly, research indicates that obesity is associated with reduced microbiota diversity, characterized by an increased abundance of Firmicutes and a decreased abundance of Bacteroidetes (44). This imbalance may hinder the colonization and beneficial effects of dietary live microbes (45), whereas the diverse microbiota in non-obese individuals offers a more favorable environment for these microbes. Secondly, non-obese individuals experience lower levels of chronic inflammation compared to obese individuals, who often have elevated pro-inflammatory cytokines (46). The anti-inflammatory properties of dietary live microbes can be diminished in the presence of high baseline inflammation, which may reduce their protective effects against PD. Additionally, non-obese individuals typically demonstrate a more favorable metabolic profile, including better insulin sensitivity (47). In contrast, obesity is associated with metabolic disturbances that can increase oxidative stress and inflammation (48), potentially weakening the protective effects of dietary live microbes. Lastly, dietary patterns exhibit significant differences between the two groups. Obese individuals typically consume high-fat, high-sugar diets that are low in fiber and dietary live microbes, which exacerbate gut dysbiosis and diminish the benefits of dietary live microbes. On the contrary, non-obese individuals tend to have more balanced diets that are rich in fermented foods, thereby enhancing their overall health benefits (49, 50).

In this study, a significant correlation between the intake of dietary live microbes and a reduced risk of PD was identified. This correlation may be influenced by various metabolic and biological mechanisms. Firstly, dietary live microbes can modulate gut microbiota, thereby promoting the proliferation of beneficial bacterial genera such as *Bifidobacterium and lactobacilli* (51). These beneficial bacteria are known to produce short-chain fatty acids (SCFAs), including propionate, acetate, and butyrate, possessing neuroprotective properties and anti-inflammatory (52), which can strengthen the intestinal barrier, reducing



microbial translocation and systemic inflammation which are key factors in the progression of PD (53). Additionally, dietary live microbe intakes can influence the immune system by stimulating the production of antiinflammatory cytokines and enhancing the function of regulatory T cells (54), helping to mitigate the chronic low-grade inflammation often observed in PD patients. Furthermore, these microbes may also exert antioxidant effects, reducing oxidative stress, a significant contributor to neuronal damage in PD (55, 56). Dietary live microbe intakes can offer a promising non-pharmacological approach by improving gut health, modulating inflammation, and reducing oxidative stress, to potentially lower the risk of PD.

To the best of our knowledge, this is the first study to explore the correlation between dietary microbe intake and the prevalence of PD. This study possesses several notable strengths. Firstly, the data adopted in this study were sourced from NHANES, a nationally representative database, with data collection and compilation conducted by professionals. Secondly, the study assessed subjects' dietary live microbe intake through their self-reported food intake. Microbe intake was quantified by categorizing subjects into high, medium, and low intake groups based on microbial content, as well as

into G3, G2, and G1 groups according to MedHi intake. Thirdly, the overall microbial content of all diets has been estimated by analyzing the data from 24-h dietary recall assessments, rather than concentrating exclusively on the intake of probiotics and the supplements.

However, it is essential to recognize several limitations inherent in this study. Firstly, the database employed in this study defined PD based on the anti-Parkinsonian medication rather than a clinical diagnosis by healthcare professionals, leading to potential biases. Specifically, subjects with mild PD symptoms who do not require medication might have been included in the control group, while subjects with other neurological disorders prescribed drugs on PD might have been misclassified into the PD group. Secondly, the approach endorsed by the International Society for the Science of Probiotics and Prebiotics for classifying the dietary live microbe intake was adopted in order to gauge the microbial content. Although these classification principles are primarily based on statistical values (9, 57–59) and authoritative reviews from previous literature (30), they predominantly rely on indirect assessments rather than direct microbial testing. Furthermore, these methods do not take into account potential variations in microbial content resulting from factors such as

Subgroup Age, years	Microbe.level	adj.OR_95CI	P value			0.954
<60	Low	1(Ref)			1	
	Medium	0.80 (0.54~1.19)	0.268			
	High	0.65 (0.38~1.12)	0.118	-		
>60						
	Low	1(Ref)			· · · · ·	
	Medium	$0.69 (0.50 \sim 0.94)$	0.020			
Gender	Figh	0.47 (0.29~0.75)	0.001	•		0.541
Male						0.011
	Low	1(Ref)			+	
	Medium	0.77 (0.54~1.09)	0.134		•	
Fomalo	High	0.44 (0.25~0.78)	0.005			
remale	Low	1(Ref)			↓	
	Medium	0.70 (0.50~0.99)	0.043			
	High	0.58 (0.37~0.91)	0.018	F		
BMI, kg/m2						0.017
<30	Low	1(Ref)				
	Medium	0.67 (0.48~0.93)	0.016			
	High	0.35 (0.21~0.59)	<0.001		_	
>30						
	Low	1(Ref)	0.252		_	
	Hiah	0.79 (0.49~1.18)	0.262			
Diabetes	i ngin	0.10 (0.10 1.20)	0.010			0.534
No						
	Low	1(Ref)			•	
	Medium	0.77 (0.58~1.02)	0.073	_		
Yes	riigii	0.00 (0.00-0.79)	0.002		•••	
	Low	1(Ref)			•	
	Medium	0.58 (0.35~0.96)	0.034	-		
Unartonaian	High	0.50 (0.23~1.05)	0.067		• •	0 767
No						0.767
	Low	1(Ref)			•	
	Medium	0.80 (0.55~1.16)	0.239			
Ma a	High	0.52 (0.31~0.89)	0.018		••	
res	Low	1(Ref)				
	Medium	0.68 (0.49~0.95)	0.022		—	
	High	0.53 (0.33~0.86)	0.009			
Hyperlipidemia						0.177
No	Low	1(Pof)				
	Medium	$0.55(0.34 \sim 0.91)$	0.021	-	_ •	
	High	0.34 (0.15~0.74)	0.006	←		
Yes						
	Low	1(Ref)	0 104		.	
	High	0.60 (0.60~1.06)	0.124	-		
CHD						0.054
No						
	Low	1(Ref)	0.001		•	
	wealum High	0.55 (0.51~0.85)	0.001	-		
Yes	ingn	0.01 (0.00~0.74)	~0.001	-		
	Low	1(Ref)			+	
	Medium	2.07 (0.83~5.21)	0.120		•	•
01-1-1	High	0.64 (0.12~3.27)	0.591	•	•	+
No						0.570
	Low	1(Ref)			•	
	Medium	0.76 (0.58~0.98)	0.036		—	
	High	0.54 (0.37~0.78)	0.001	-		
Yes	Law	4/D-6)				
	LOW	1(Ret) 0.49 (0.22~1.10)	0.082		<u> </u>	
	High	0.40 (0.22~1.10)	0.062			4
		(
				0.20	1.0 1	1.5
				OR	(95%CI)	

Age, years	Microbellever	auj.ort_5561	i value		0.980
<60				r	
	G1	1(Ref)		•	
	G2	0.74 (0.48~1.14)	0.174		
> 00	G3	0.77 (0.49~1.20)	0.250	• • ••	
>60	C1	1(Pof)			
	G	1(Rel)	0.020		
	G2 G3	0.00(0.47 - 0.94) 0.59(0.41~0.85)	0.020	· · · · ·	
Gender	00	0.00 (0.41 0.00)	0.004		0 887
Male					0.007
Male	G1	1(Ref)		•	
	G2	0.68 (0.45~1.01)	0.057	→	
	G3	0.67 (0.45~1.01)	0.053		
Female		,			
	G1	1(Ref)		•	
	G2	0.70 (0.48~1.01)	0.056	→ →→	
	G3	0.63 (0.42~0.92)	0.018		
BMI, kg/m2					0.061
<30					
	G1	1(Ref)		+	
	G2	0.65 (0.46~0.94)	0.020	—	
	G3	0.49 (0.33~0.73)	<0.001		
>30					
	G1	1(Ref)		•	
	G2	0.74 (0.49~1.13)	0.165	• • •	
	G3	0.87 (0.58~1.31)	0.515	••	
Diabetes					0.533
No		1/D-0			
	G1	1(Ref)	0.057	†	
	G2	0.74 (0.54~1.01)	0.057		
	G3	0.66 (0.47~0.91)	0.010	—	
res	04	1(D-6)			
	GI	1(Rel)	0.020		
	G2 C2	$0.52(0.29 \sim 0.94)$	0.030		
Hyportonsion	63	0.59 (0.55~1.04)	0.000		0 883
No					0.005
	G1	1(Ref)		•	
	G2	$0.76(0.51 \sim 1.14)$	0 186		
	G3	0.67 (0.44~1.03)	0.066	▶	
Yes		· · · ·			
	G1	1(Ref)		+	
	G2	0.64 (0.45~0.93)	0.019		
	G3	0.63 (0.44~0.92)	0.017		
Hyperlipidemia					0.060
No					
	G1	1(Ref)		+	
	G2	0.39 (0.21~0.72)	0.003		
	G3	0.59 (0.34~1.02)	0.061	•+	
Yes					
	G1	1(Ref)		•	
	G2	0.81 (0.60~1.10)	0.179	•+•	
	G3	0.67 (0.48~0.93)	0.017	► ••••	
CHD					0.099
NO	O 4	100			
	G1	1(Ref)	0.000	· · · · · · · · •	
	G2	0.63 (0.48~0.84)	0.002		
	G3	0.59 (0.44~0.80)	<0.001	→→→	
Yes	01	1(D-5)			
	G1	1(Ref)		†	
	G2	1.72 (0.63~4.70)	0.289		•
04	G3	1.77 (0.63~4.95)	0.276	• • •	
Stroke					0.182
No	<i></i>				
	G1	1(Ref)	0.045	• • • • •	
	G2	0.70 (0.52~0.93)	0.015		
	G3	0.69 (0.51~0.92)	0.012	••••	
Yes	<u>.</u>				
	G1	1(Ref)	0.070	· . · · · · ·	
	62	0.63 (0.28~1.43)	0.270		
	63	0.25 (0.07~0.87)	0.029	•	
					-
			0.2		.5
				UR (95%CI)	

TABLE 3 Association between dietary live microbe intake and prevalence of PD.

Subgroups	Mode	el 1	Mode	l 2	Model 3					
	OR (95%CI)	P-value	OR (95%CI)	P-value	OR (95%CI)	P-value				
Per100 unit increase	0.92 (0.85 ~ 0.99)	0.032	0.92 (0.84 ~ 0.99)	0.033	0.85 (0.76 ~ 0.94)	0.002				
Category of MedHi ^a										
Low	1(Ref)		1(Ref)		1(Ref)					
Medium	0.74 (0.58 ~ 0.94)	0.015	0.73 (0.57 ~ 0.93)	0.010	0.69 (0.51 ~ 0.93)	0.016				
High	0.54 (0.38 ~ 0.76)	<0.001	0.52 (0.37 ~ 0.74)	< 0.001	0.41 (0.26 ~ 0.63)	< 0.001				
<i>P</i> for trend	0.73 (0.63 ~ 0.86)	<0.001	0.72 (0.61 ~ 0.85)	<0.001	0.65 (0.53 ~ 0.8)	< 0.001				
Category of MedHi ^b										
G1	1(Ref)		1(Ref)		1(Ref)					
G2	0.70 (0.54 ~ 0.92)		0.69 (0.52 ~ 0.90)	0.007	0.68 (0.49 ~ 0.95)	0.022				
G3	0.65 (0.50 ~ 0.86)	< 0.001	0.64 (0.48 ~ 0.85)	0.002	0.53 (0.37 ~ 0.75)	< 0.001				
<i>P</i> for trend	0.80 (0.70 ~ 0.92)	< 0.001	0.79 (0.69 ~ 0.91)	0.001	0.72 (0.61 ~ 0.86)	< 0.001				

Model 1: none covariates were adjusted.

Model 2: gender, age, race, BMI, physical activities, diabetes, education level, hypertension, smoking status, CHD, stroke, cancer, PIR, hyperlipidemia, marital status, drinking status, total energy intake, non-food prebiotic use (yes or no), and non-food probiotic use (yes or no) were adjusted.

Model 3: model 2+ non-food prebiotic use (yes or no), and non-food probiotic use (yes or no) were adjusted.

*Participants were also classified into three different groups considering the general intake of foods with varying contents of microbe: low (all foods consumed were Low); moderate (any foods consumed were High).

^bParticipants were categorized into three groups based on the MedHi consumption to quantify the ingestion of live microbes: G1, consumers without intakes of any MedHi food; G2, those with intakes of MedHi food above zero but below the median level for consumers; G3, those with intakes of MedHi food above the median level for consume.

food preparation, storage, and processing. This oversight may contribute to misclassifications in the assessment of dietary live microbe intake levels. However, direct assessment requires a long time and huge expenditure, which limits its application. Thirdly, due to the varied dietary habits among individuals with different dietary patterns, further research is needed to explore the association between dietary live microbe intake from different food sources and the prevalence of PD. Fourthly, given the cross-sectional design of this study, it is crucial to acknowledge that causal inferences cannot be conducted. Fifthly, the potential impact of unidentified confounding variables may limit the interpretability of these findings, particularly concerning the consumption of other specific food groups that are rich in live microorganisms. Finally, the study was unable to determine whether the observed health benefits were attributable to the food itself or to the presence of live bacteria within the food, thereby limiting the ability to ascertain the true impact on the prevalence of PD.

Conclusion

To sum up, this study indicates that a higher dietary live microbe intake associates with a lower prevalence of PD. Given the increasing prevalence of PD and the possible neuroprotective effects of dietary live microbes, the results of this study have important ramifications for public health initiatives and clinical approaches.

Data availability statement

Publicly available datasets were analyzed in this study. This data can be found here: NHANES, https://wwwn.cdc.gov/nchs/nhanes.

Ethics statement

The studies involving humans were approved by National Center for Health Statistics Ethics Review Board. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

HZ: Conceptualization, Investigation, Writing – original draft. TZ: Conceptualization, Investigation, Writing – review & editing. XC: Data curation, Methodology, Writing – review & editing. YL: Conceptualization, Data curation, Investigation, Methodology, Software, Supervision, Writing – review & editing.

Funding

The author(s) declare that no financial support was received for the research and/or publication of this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The authors declare that no Gen AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations,

References

1. Reich SG, Savitt JM. Parkinson's disease. *Med Clin North Am.* (2019) 103:337–50. doi: 10.1016/j.mcna.2018.10.014

2. Ben-Shlomo Y, Darweesh S, Llibre-Guerra J, Marras C, San Luciano M, Tanner C. The epidemiology of Parkinson's disease. *Lancet.* (2024) 403:283–92. doi: 10.1016/S0140-6736(23)01419-8

3. Xu Q, Ou X, Li J. The risk of falls among the aging population: a systematic review and Meta-analysis. *Front Public Health*. (2022) 10:902599. doi: 10.3389/fpubh.2022.902599

4. Dorsey ER, Constantinescu R, Thompson JP, Biglan KM, Holloway RG, Kieburtz K, et al. Projected number of people with Parkinson disease in the Most populous nations, 2005 through 2030. *Neurology*. (2007) 68:384–6. doi: 10.1212/01.wnl.0000247740.47667.03

5. Tolosa E, Garrido A, Scholz SW, Poewe W. Challenges in the diagnosis of Parkinson's disease. *Lancet Neurol.* (2021) 20:385–97. doi: 10.1016/S1474-4422(21)00030-2

6. Dorsey ER, Bloem BR. The Parkinson pandemic-a call to action. *JAMA Neurol.* (2018) 75:9–10. doi: 10.1001/jamaneurol.2017.3299

7. Wirdefeldt K, Adami HO, Cole P, Trichopoulos D, Mandel J. Epidemiology and etiology of Parkinson's disease: a review of the evidence. *Eur J Epidemiol*. (2011) 26:1–58. doi: 10.1007/s10654-011-9581-6

8. Tansey MG, Wallings RL, Houser MC, Herrick MK, Keating CE, Joers V. Inflammation and immune dysfunction in Parkinson disease. *Nat Rev Immunol.* (2022) 22:657–73. doi: 10.1038/s41577-022-00684-6

9. Rezac S, Kok CR, Heermann M, Hutkins R. Fermented foods as a dietary source of live organisms. *Front Microbiol.* (2018) 9:1785. doi: 10.3389/fmicb.2018.01785

10. Wastyk HC, Fragiadakis GK, Perelman D, Dahan D, Merrill BD, Yu FB, et al. Gutmicrobiota-targeted diets modulate human immune status. *Cell.* (2021) 184:4137–4153.e14. doi: 10.1016/j.cell.2021.06.019

11. Agus A, Planchais J, Sokol H. Gut microbiota regulation of tryptophan metabolism in health and disease. *Cell Host Microbe*. (2018) 23:716–24. doi: 10.1016/j.chom.2018.05.003

12. Zhu S, Jiang Y, Xu K, Cui M, Ye W, Zhao G, et al. The Progress of gut microbiome research related to brain disorders. *J Neuroinflammation*. (2020) 17:25. doi: 10.1186/s12974-020-1705-z

13. Roy Sarkar S, Banerjee S. Gut Microbiota in Neurodegenerative Disorders. J Neuroimmunol. (2019) 328:98–104. doi: 10.1016/j.jneuroim.2019.01.004

14. Keshavarzian A, Green SJ, Engen PA, Voigt RM, Naqib A, Forsyth CB, et al. Colonic bacterial composition in Parkinson's disease. *Mov Disord*. (2015) 30:1351–60. doi: 10.1002/mds.26307

15. Lai F, Jiang R, Xie W, Liu X, Tang Y, Xiao H, et al. Intestinal pathology and gut microbiota alterations in a Methyl-4-Phenyl-1,2,3,6-Tetrahydropyridine (Mptp) mouse model of Parkinson's disease. *Neurochem Res.* (2018) 43:1986–99. doi: 10.1007/s11064-018-2620-x

16. Sampson TR, Debelius JW, Thron T, Janssen S, Shastri GG, Ilhan ZE, et al. Gut microbiota regulate motor deficits and Neuroinflammation in a model of Parkinson's disease. *Cell.* (2016) 167:1469–1480.e12. doi: 10.1016/j.cell.2016.11.018

17. Pant A, Bisht KS, Aggarwal S, Maiti TK. Human gut microbiota and Parkinson's disease. Prog Mol Biol Transl Sci. (2022) 192:281–307. doi: 10.1016/bs.pmbts.2022.08.004

18. Hill C, Guarner F, Reid G, Gibson GR, Merenstein DJ, Pot B, et al. Expert consensus document. The international scientific Association for Probiotics and Prebiotics Consensus Statement on the scope and appropriate use of the term probiotic. *Nat Rev Gastroenterol Hepatol.* (2014) 11:506–14. doi: 10.1038/nrgastro.2014.66

19. Reid G, Younes JA, Van der Mei HC, Gloor GB, Knight R, Busscher HJ. Microbiota restoration: natural and supplemented recovery of human microbial communities. *Nat Rev Microbiol.* (2011) 9:27–38. doi: 10.1038/nrmicro2473

20. Kamada N, Chen GY, Inohara N, Nunez G. Control of pathogens and Pathobionts by the gut microbiota. *Nat Immunol.* (2013) 14:685–90. doi: 10.1038/ni.2608

or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnut.2025.1606922/ full#supplementary-material

21. Leclercq S, Matamoros S, Cani PD, Neyrinck AM, Jamar F, Starkel P, et al. Intestinal permeability, gut-bacterial Dysbiosis, and behavioral markers of alcoholdependence severity. *Proc Natl Acad Sci USA*. (2014) 111:E4485–93. doi: 10.1073/pnas.1415174111

22. Cooper TE, Khalid R, Chan S, Craig JC, Hawley CM, Howell M, et al. Synbiotics, prebiotics and probiotics for people with chronic kidney disease. *Cochrane Database Syst Rev.* (2023) 10:CD013631. doi: 10.1002/14651858.CD013631.pub2

23. Kaufmann B, Seyfried N, Hartmann D, Hartmann P. Probiotics, prebiotics, and Synbiotics in nonalcoholic fatty liver disease and alcohol-associated liver disease. *Am J Physiol Gastrointest Liver Physiol.* (2023) 325:G42–61. doi: 10.1152/ajpgi.00017.2023

24. Quigley EMM. Prebiotics and probiotics in digestive health. *Clin Gastroenterol Hepatol.* (2019) 17:333–44. doi: 10.1016/j.cgh.2018.09.028

25. Atak ES, Yildiz D, Kocaturk RR, Temizyurek A, Ozcan OO, Erguzel TT, et al. Therapeutic targets of probiotics in Parkinson disease: a systematic review of randomized controlled trials. *Basic Clin Neurosci.* (2024) 15:165–74. doi: 10.32598/bcn.2022.3665.1

26. Lim MY, Nam YD. Gut microbiome in healthy aging versus those associated with frailty. *Gut Microbes.* (2023) 15:2278225. doi: 10.1080/19490976.2023.2278225

27. Ghosh TS, Das M, Jeffery IB, O'Toole PW. Adjusting for age improves identification of gut microbiome alterations in multiple diseases. *eLife*. (2020) 9:9. doi: 10.7554/eLife.50240

28. Shanahan F, van Sinderen D, O'Toole PW, Stanton C. Feeding the microbiota: transducer of nutrient signals for the host. *Gut.* (2017) 66:1709–17. doi: 10.1136/gutjnl-2017-313872

29. Perez Martinez G, Bauerl C, Collado MC. Understanding gut microbiota in Elderly's health will enable intervention through probiotics. *Benef Microbes.* (2014) 5:235–46. doi: 10.3920/BM2013.0079

30. Marco ML, Hutkins R, Hill C, Fulgoni VL, Cifelli CJ, Gahche J, et al. A classification system for defining and estimating dietary intake of live microbes in us adults and children. *J Nutr.* (2022) 152:1729–36. doi: 10.1093/jn/nxac074

31. Huo X, Jia S, Zhang X, Sun L, Liu X, Liu L, et al. Association of Dietary Live Microbe Intake with abdominal aortic calcification in us adults: a cross-sectional study of Nhanes 2013-2014. *Front Nutr.* (2023) 10:1267607. doi: 10.3389/fnut.2023.1267607

32. Wang X, Wang H, Yu Q, Fu S, Yang Z, Ye Q, et al. High dietary live microbe intake is correlated with reduced risk of depressive symptoms: a cross-sectional study of Nhanes 2007-2016. *J Affect Disord*. (2024) 344:198–206. doi: 10.1016/j.jad.2023.10.015

33. Lin X, Lin H, Xu J, Yang S, Miao L. Relative fat mass as a predictor of gallstones: Insights from National Health and nutrition examination survey data. *Lipids Health Dis.* (2025) 24:78. doi: 10.1186/s12944-025-02480-2

34. Fan J, Wang Y, Guo X, Cao S, Zhan S, Li R. Association between Life's essential 8 and Parkinson's disease: a case-control study. *BMC Public Health*. (2025) 25:411. doi: 10.1186/s12889-025-21648-0

35. Su J, Liu L, Wang R, Li C, Wang Z, Xu Q, et al. Association between dietary Betacarotene intake with Parkinson's disease and all-cause mortality among American adults aged 40 and older (Nhanes 2001-2018). *Front Nutr.* (2024) 11:1430605. doi: 10.3389/fnut.2024.1430605

36. Xu J, Li YC. Negative correlation between Cardiometabolic index and testosterone in male adults. *Front Endocrinol.* (2024) 15:1447230. doi: 10.3389/fendo.2024.1447230

37. Chen Y, Lin H, Xu J, Zhou X. Estimated glucose disposal rate is correlated with increased depression: a population-based study. *BMC Psychiatry*. (2024) 24:786. doi: 10.1186/s12888-024-06257-2

38. Hill C, Tancredi DJ, Cifelli CJ, Slavin JL, Gahche J, Marco ML, et al. Positive health outcomes associated with live microbe intake from foods, including fermented foods, assessed using the Nhanes database. *J Nutr.* (2023) 153:1143–9. doi: 10.1016/j.tjnut.2023.02.019

39. Han L, Wang Q. Association of Dietary Live Microbe Intake with cardiovascular disease in us adults: a cross-sectional study of Nhanes 2007-2018. *Nutrients*. (2022) 14:4908. doi: 10.3390/nu14224908

40. Tang H, Zhang X, Luo N, Huang J, Zhu Y. Association of dietary live microbes and non-dietary prebiotic/probiotic intake with cognitive function in older adults: evidence from Nhanes. J Gerontol A Biol Sci Med Sci. (2024) 79:175. doi: 10.1093/gerona/glad175

41. Tamtaji OR, Taghizadeh M, Daneshvar Kakhaki R, Kouchaki E, Bahmani F, Borzabadi S, et al. Clinical and metabolic response to probiotic Administration in People with Parkinson's disease: a randomized, double-blind, placebo-controlled trial. *Clin Nutr.* (2019) 38:1031–5. doi: 10.1016/j.clnu.2018.05.018

42. Jin X, Dong W, Chang K, Yan Y, Liu X. Efficacy of probiotic supplements on Parkinson's disease: a systematic review and Meta-analysis. *Complement Ther Med.* (2024) 82:103045. doi: 10.1016/j.ctim.2024.103045

43. Park JM, Lee SC, Ham C, Kim YW. Effect of probiotic supplementation on gastrointestinal motility, inflammation, motor, non-motor symptoms and mental health in Parkinson's disease: a meta-analysis of randomized controlled trials. *Gut Pathog.* (2023) 15:9. doi: 10.1186/s13099-023-00536-1

44. Gomes AC, Hoffmann C, Mota JF. The human gut microbiota: metabolism and perspective in obesity. *Gut Microbes*. (2018) 9:308–25. doi: 10.1080/19490976.2018.1465157

45. Chen Y, Ni H, Zhang H. Exploring the relationship between live microbe intake and obesity prevalence in adults. *Sci Rep.* (2024) 14:21724. doi: 10.1038/s41598-024-72961-4

46. Xu X, Xu J, Zhang M. Association between metabolic score for visceral fat and obstructive sleep apnea: a cross-sectional study. *Front Med.* (2024) 11:1480717. doi: 10.3389/fmed.2024.1480717

47. Zhang M, Weng X, Xu J, Xu X. Correlation between obstructive sleep apnea and weight-adjusted-waist index: a cross-sectional study. *Front Med.* (2024) 11:1463184. doi: 10.3389/fmed.2024.1463184

48. Karam BS, Chavez-Moreno A, Koh W, Akar JG, Akar FG. Oxidative stress and inflammation as central mediators of atrial fibrillation in obesity and diabetes. *Cardiovasc Diabetol.* (2017) 16:120. doi: 10.1186/s12933-017-0604-9

49. Wang H, Li W, Lai Q, Huang Q, Ding H, Deng Z. Inflammatory markers and risk of Parkinson's disease: a population-based analysis. *Parkinsons Dis.* (2024) 2024:4192853. doi: 10.1155/padi/4192853

50. Gao A, Lv J, Su Y. The inflammatory mechanism of Parkinson's disease: gut microbiota metabolites affect the development of the disease through the gut-brain Axis. *Brain Sci.* (2025) 15:159. doi: 10.3390/brainsci15020159

51. Kumar D, Bishnoi M, Kondepudi KK, Sharma SS. Gut microbiota-based interventions for Parkinson's disease: neuroprotective mechanisms and current perspective. *Probiot Antimicrob Proteins*. (2025). doi: 10.1007/s12602-024-10433-x

52. Roager HM, Licht TR. Microbial tryptophan catabolites in health and disease. *Nat Commun.* (2018) 9:3294. doi: 10.1038/s41467-018-05470-4

53. Koh A, De Vadder F, Kovatcheva-Datchary P, Backhed F. From dietary Fiber to host physiology: short-chain fatty acids as key bacterial metabolites. *Cell.* (2016) 165:1332–45. doi: 10.1016/j.cell.2016.05.041

54. Shibata N, Kunisawa J, Kiyono H. Dietary and microbial metabolites in the regulation of host immunity. *Front Microbiol.* (2017) 8:2171. doi: 10.3389/fmicb.2017.02171

55. Liu F, Ran Q, Zhang H, Chen J. The systemic immune-inflammation index and the risk of Parkinson's disease in the U.S.: a cross-sectional study. *J Clin Med.* (2025) 14:403. doi: 10.3390/jcm14020403

56. Jiao F, Zhou L, Wu Z. The microbiota-gut-brain Axis: a potential target in the small-molecule compounds and gene therapeutic strategies for Parkinson's disease. *Neurol Sci.* (2025) 46:561–78. doi: 10.1007/s10072-024-07878-x

57. Feroz F, Shimizu H, Nishioka T, Mori M, Sakagami Y. Bacterial and fungal counts of dried and semi-dried foods collected from Dhaka, Bangladesh, and their reduction methods. *Biocontrol Sci.* (2016) 21:243–51. doi: 10.4265/bio.21.243

58. Jeddi MZ, Yunesian M, Gorji ME, Noori N, Pourmand MR, Khaniki GR. Microbial evaluation of fresh, minimally-processed vegetables and bagged sprouts from chain supermarkets. *J Health Popul Nutr.* (2014) 32:391–9.

59. Sonnenburg ED, Sonnenburg JL. The ancestral and industrialized gut microbiota and implications for human health. *Nat Rev Microbiol.* (2019) 17:383–90. doi: 10.1038/s41579-019-0191-8