Check for updates

#### **OPEN ACCESS**

EDITED BY Yifeng Zhang, Peking University, China

#### REVIEWED BY Mario Juan Simirgiotis, Austral University of Chile, Chile Pengfei Cheng, Ningbo University, China Ana Novo Barros, University of Trás-os-Montes and Alto Douro, Portugal \*CORRESPONDENCE Zhongliang Sun Zlsun@ytu.edu.cn

Liqin Sun

RECEIVED 21 April 2025 ACCEPTED 10 June 2025 PUBLISHED 26 June 2025

#### CITATION

Meng R, Zhang Y, Jiang Y, Li B, Chen X, Sun Z and Sun L (2025) Subcritical fluid and molecular distillation extraction of *Nannochloropsis gaditana* lipid and its metabolic benefits in hyperlipidemic mice. *Front. Nutr*, 12:1615332. doi: 10.3389/fnut.2025.1615332

#### COPYRIGHT

© 2025 Meng, Zhang, Jiang, Li, Chen, Sun and Sun. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

## Subcritical fluid and molecular distillation extraction of *Nannochloropsis gaditana* lipid and its metabolic benefits in hyperlipidemic mice

# Ruilong Meng<sup>1</sup>, Yu Zhang<sup>1</sup>, Yishan Jiang<sup>2</sup>, Bin Li<sup>2</sup>, Xi Chen<sup>1</sup>, Zhongliang Sun<sup>1\*</sup> and Liqin Sun<sup>1\*</sup>

<sup>1</sup>Yantai Key Laboratory of Characteristic Agricultural Bioresource Conservation and Germplasm Innovative Utilization, School of Life Sciences, Yantai University, Yantai, China, <sup>2</sup>Yantai Zhongwei Pet Food Co., Ltd., Yantai, China

The marine microalga Nannochloropsis gaditana is a fast-growing species rich in long-chain polyunsaturated fatty acids (PUFAs), particularly eicosapentaenoic acid (EPA), and other bioactive compounds. In this study, lipids from N. gaditana powder were extracted and refined using subcritical butane combined with molecular distillation to obtain a highly purified lipid extract with increased EPA concentration (58.92% w/w) and improved biological activity. The antihyperlipidemic effects of the lipid extract were evaluated in female Kunming mice (4 weeks old) fed a high-fat diet. Results demonstrated that N. gaditana lipid supplementation significantly reduced body weight gain, serum triglycerides (TG), total cholesterol (TC), and low-density lipoprotein cholesterol (LDL-C), while elevating high-density lipoprotein cholesterol (HDL-C). Additionally, the lipid extract ameliorated hepatic inflammation (reduced TNF- $\alpha$  and IL-1β levels), attenuated oxidative stress (enhanced SOD, CAT, and GSH-Px activities), and modulated lipid metabolism enzymes (inhibited FAS, ACC, and HMGCR; upregulated LCAT). These findings highlight the potential of EPA-rich N. gaditana lipid as a natural and sustainable therapeutic strategy for managing hyperlipidemia and associated metabolic disorders.

#### KEYWORDS

Nannochloropsis gaditana, lipids, subcritical extraction, molecular distillation, antihyperlipidemia

### **1** Introduction

Hyperlipidemia, characterized by elevated levels of triglycerides (TG), total cholesterol (TC), and low-density lipoprotein cholesterol (LDL-C), is a major risk factor for cardiovascular diseases (CVDs), which account for 32% of global mortality annually (1). Current pharmacological interventions, such as statins, often entail adverse side effects, including hepatotoxicity and myopathy, driving the search for safer, natural alternatives (2). Among bioactive compounds, omega-3 polyunsaturated fatty acids

( $\omega$ -3 PUFAs), particularly eicosapentaenoic acid (EPA), have garnered attention for their lipid-modulating, anti-inflammatory, and antioxidant properties (3). EPA not only reduces atherogenic lipids but also enhances high-density lipoprotein cholesterol (HDL-C), promoting reverse cholesterol transport (4). However, conventional EPA sources like fish oil face critical limitations, including unsustainable harvesting practices, fishy odor, and the co-presence of docosahexaenoic acid (DHA), complicating EPA purification (5). These challenges underscore the need for alternative, sustainable EPA sources with streamlined processing.

Marine microalgae represent the primary biological origin of  $\omega$ -3 PUFAs, bypassing the ecological and purification drawbacks of fish-derived oils (6). Nannochloropsis gaditana, a rapidly growing microalga, is uniquely suited for EPA production, as its lipid profile is rich in EPA (up to 30%-50% of total fatty acids) and devoid of DHA (7). Furthermore, its scalability and odorless biomass make it an industrially viable candidate (8). Despite these advantages, inefficient lipid recovery methods hinder commercial exploitation. Traditional solvent extraction often compromises yield or purity, while supercritical  $\mathrm{CO}_2$ extraction demands high energy and prolonged processing times (9). Subcritical fluid extraction, a green technology using solvents like butane at near-critical temperatures, offers higher selectivity and lower energy consumption (10). When coupled with molecular distillation-a method effective in concentrating heat-sensitive compounds-this approach could overcome existing bottlenecks in EPA purification (11). However, the combined efficacy of these techniques for N. gaditana lipid extraction and their therapeutic potential in hyperlipidemia remains underexplored.

This study aimed to (1) optimize a novel extraction protocol integrating subcritical fluid extraction and molecular distillation to obtain microalgal lipids with enhanced EPA content from N. gaditana powder, and (2) evaluate the lipid's anti-hyperlipidemic effects in a hyperlipidemic murine model. By addressing both technological and therapeutic gaps, this work advances microalgal lipids as sustainable, efficacious agents for managing hyperlipidemia and associated metabolic disorders. Specifically, subcritical fluid extraction combined with molecular distillation was employed to achieve high-purity EPA enrichment from N. gaditana biomass. The hypolipidemic efficacy of the extracted lipids was systematically validated using a hyperlipidemic mouse model, bridging the gaps between inefficient lipid recovery and unclear therapeutic mechanisms. This dual-focused approach provides both technological validation and pharmacodynamic evidence for microalgal lipids as a sustainable solution for metabolic syndrome management.

### 2 Materials and methods

### 2.1 Materials and reagents

The powder of *N. gaditana* was purchased from Guangxi Xiaozao Technology Co., Ltd., China. Commercial kits for the determination of malondialdehyde (MDA), superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), and catalase (CAT)

were obtained from Nanjing Jiancheng Bioengineering Institute (Nanjing, China). Commercial kits for the determination of mouse tumor necrosis factor-alpha (TNF- $\alpha$ ), mouse interleukin-1 beta (IL-1 $\beta$ ), mouse fatty acid synthase (FAS), mouse 3-hydroxy-3-methylglutaryl-CoA reductase (HMGCR), mouse acetyl-CoA carboxylase (ACC), and mouse lecithin cholesterol acyltransferase (LCAT) were purchased from Quanzhou Ruixin Biotechnology Co., Ltd (Quanzhou, China).

### 2.2 Preparation of N. gaditana lipid

The lipid extraction was performed using a mixture of subcritical butane and ethanol as solvents. The solvent ratio of subcritical butane was 3:1 (subcritical butane/microalgal powder, v/w), and ethanol (95% v/v) was added as an entrainer at a ratio of 1:10 (ethanol/microalgal powder, v/w). The extraction process was carried out in a subcritical pilot-scale apparatus (CBE-T-5L, Henan Subcritical Biotechnology Co., Ltd., China) at 50°C for 40 min, repeated four times to obtain the crude lipids. Refinement of the crude lipids was conducted via molecular distillation following esterification. The esterification method was modified based on the procedure reported by Costa Cardoso et al. (12). The refining process was carried out using a short-path distillation apparatus (VKL70-5, VTA GMBH & Co. KG, Germany) under the conditions of an evaporation temperature of 150°C, a vacuum level of 20 Pa, a wiped-film rotor speed of 220 rpm, and a cooling temperature of  $-10^{\circ}$ C. Microalgal lipid was obtained after refining. The fatty acids in the lipids were analyzed by GC (7820A, Agilent Technologies, Inc., USA) using a DB-WAX (Agilent Technologies, Inc., USA) capillary column.

# 2.3 Evaluation of the lipid-lowering effects of *N. gaditana* lipid using a high-fat diet-induced hyperlipidemia mouse model

One hundred and five healthy female Kunming mice (4 weeks old, weighing 20  $\pm$  2 g) were obtained from Shandong Pengyue Laboratory Animal Technology Co., Ltd. (Jinan, China) and randomly assigned to seven experimental groups (n = 15 per group). The blank control group (Group A) was fed a standard diet, while the high-fat control group (Group B) and five treatment groups were induced with hyperlipidemia by feeding a highfat diet for an uninterrupted 40-day period, with body weights recorded weekly. After model establishment, serum levels of TC, TG, and LDL in three mice per group were measured. All parameters showed an increase of over 50% compared to the blank control group (Group A), demonstrating successful hyperlipidemia modeling. Groups A and B received corn oil as the vehicle, whereas the treatment groups were received the following interventions: Group C (low-dose N. gaditana oil, 0.3 mg/g BW), Group D (medium-dose N. gaditana oil, 0.6 mg/g BW), Group E (high-dose N. gaditana oil, 1.2 mg/g BW), Group G (crude lipids, 1.2 mg/g BW), and Group F (positive control, fish oil 1,200 mg/kg BW). All treatments were administered once daily by oral gavage throughout the 40-day experimental period, with the dose selection based on established protocols from comparable studies in the literature (13).

### 2.4 Determination of blood lipid levels

Upon end of the treatment protocol, experimental mice underwent a 12-h fasting period prior to cervical dislocation. Postmortem blood samples were collected and centrifuged (4,000 rpm, 15 min) for serum isolation. Quantitative analysis of fatty acid profiles along with serum biochemical parametersincluding TC, TG, LDL-C, HDL-C-was performed using an automated biochemistry analyzer (cobas c 311, Roche Diagnostics, USA). Subsequent statistical evaluation was conducted on the obtained serum biochemical parameters.

# 2.5 Detection of anti-inflammatory factors

Following blood collection via retro-orbital plexus puncture, mice were humanely euthanized for hepatic tissue sampling. Residual blood was perfused with 0.9% NaCl solution. Liver tissue homogenates (10% weight/volume) were prepared using physiological saline and centrifuged at 5,000 × g for 10 min at 4°C. The supernatant was collected to measure of mouse TNF- $\alpha$  and IL-1 $\beta$  levels according to the instructions of the kit.

### 2.6 Determination of liver indices

The prepared liver homogenates were analyzed for oxidative stress markers, including MDA content and the activities of antioxidant enzymes (SOD, GSH-Px, and CAT). Enzymatic assays were also performed to determine the expression of hepatic lipid metabolism regulators: FAS, HMGCR, ACC, and LCAT.

### 2.7 Statistical analysis

Statistical analyses were performed using GraphPad Prism software (version 10.1.2; GraphPad Software, San Diego, CA). Oneway analysis of variance (ANOVA) was used for comparisons among groups, followed by Tukey's honest significant difference (HSD) *post-hoc* test for multiple comparisons. Quantitative data are presented as mean  $\pm$  standard deviation (SD), and statistical significance was defined as p < 0.05 (two-tailed).

### 2.8 Ethical considerations

The experimental protocol was reviewed and approved by the Institutional Animal Care and Use Committee (IACUC) of Yantai University, and was conducted in strict accordance with the Guidelines for the Care and Use of Laboratory Animals and the National Technical Standards for the Production, Care, and Utilization of Laboratory Animals.

### **3** Results and discussions

# 3.1 Lipid extraction and fatty acid composition analysis

The process parameters for subcritical extraction were optimized, resulting in an N. gaditana lipid with an EPA content of 29.88% (Figure 1). The extraction yield of N. gaditana lipid obtained in this study is comparable to that reported in most previous studies; however, the EPA content is higher than that obtained using solvent extraction methods. Jiménez Callejón et al. (14) used ethanol as the extraction solvent to extract lipids from Nannochloropsis sp., resulting in an EPA content of 25.3% in the Nannochloropsis sp. oil. They also applied highpressure homogenization to pretreat the Nannochloropsis sp. prior to extraction. Furthermore, Jiménez Callejón et al. (15) extracted lipids from Nannochloropsis sp. using supercritical CO<sub>2</sub>, achieving an EPA content of up to 32.6%, which is higher than the result obtained in this experiment. However, this method utilized more stringent extraction conditions and required an extraction time of 8 h. The effects of molecular distillation conditions (evaporation temperature, vacuum pressure, scraper speed, and cooling temperature) on the purity of EPA in N. gaditana lipid were also investigated, and the optimal conditions were established. Under these optimal conditions, the EPA content was increased from 29.88% to 58.63%. Compared with other methods, such as supercritical CO<sub>2</sub> extraction combined with pressurized liquid extraction (15), high-shear-assisted lipid extraction (16), and enzyme-assisted three-phase partitioning (17), the subcritical extraction combined with molecular distillation resulted in higher EPA and PUFA contents in the final N. gaditana lipid.

The fatty acid composition of extracted crude *N. gaditana* lipid is presented in Table 1. Feller et al. (18) also obtained a similar fatty acid composition when extracting lipids from *N. oculata* using subcritical butane. Magallanes et al. (11) et al. used molecular distillation combined with urea encapsulation to purify the lipids with an initial EPA content of 8.93%, aiming to prepare highly concentrated  $\omega$ -3 fatty acid ethyl esters. In their study, the concentration of unsaturated fatty acid ethyl esters reached a maximum of 95.12%, and the concentration of EPA ethyl esters reached 15.81%, which is 1.77 times the pre-purification level. In contrast, the present study utilized molecular distillation alone to purify *N. gaditana* lipid, increasing the EPA content represents a 0.96-fold increase compared to the pre-distillation level.

### 3.2 Body weight and organ index

The high-fat diet cohort (Group B) consistently exhibited elevated mean body mass compared to other experimental groups throughout the study period (Figure 2). Specifically, Group B showed a statistically significant increase in body weight gain of 4.89 g relative to the standard chow group (Group A). Following the cessation of drug treatment, the body weight of mice in all groups decreased. *N. gaditana* lipid treatment demonstrated a dosedependent efficacy in attenuating hyperlipidemia-induced weight gain. The high-dose treatment group (Group E) exhibited the greatest therapeutic effect, achieving a 5.92% reduction in body



weight, followed by the medium-dose group (Group D) with a 4.06% decrease. High-fat diet feeding significantly increased both hepatic mass (hepatomegaly) and visceral adipose tissue accumulation. Subsequent intervention with microalgal lipids substantially reversed these pathological changes, with the highdose regimen (Group E) producing the most significant reductions in liver weight and abdominal fat mass compared to the untreated hyperlipidemic controls. These findings are consistent with the observed reductions in body weight, as shown in Table 2. Li et al. (19) reported a similar trend when treating hyperlipidemic mice with fucoidan, although the results were slightly inferior to those obtained by Nunez et al. (20). In their study, Nunez et al. (20) directly administered untreated algal powder to experimental mice. The cellulose and other polysaccharides present in the powder exerted dual therapeutic effects, effectively suppressing body weight gain while significantly reducing the accumulation of hepatic lipids and abdominal adipose tissue in the mice.

The beneficial effects of N. gaditana lipid on body weight management and the reduction of hepatic and visceral adiposity are likely mediated through multiple interconnected physiological mechanisms, primarily involving the regulation of energy metabolism and the modulation of lipid storage pathways. On the one hand, the lipids derived from N. gaditana are rich in PUFAs, which have an energy density similar to that of other fats but may undergo different metabolic processes. Unsaturated fatty acids are more readily oxidized and converted into energy in the body rather than being stored as fat. PUFAs, particularly EPA, exhibit the ability to activate PPAR-a. Activation of this receptor enhances fatty acid oxidation by upregulating  $\beta$ -oxidation pathways in mitochondria and peroxisomes. Consequently, these metabolic adaptations result in a significant reduction in hepatic lipid accumulation (21) as well as decreased systemic adipose tissue deposition. On the other hand, unsaturated fatty acids can regulate fat storage within adipose tissue. Studies have demonstrated that a diet rich in unsaturated fatty acids can lower body fat percentage, particularly abdominal fat (22), which is consistent with the findings of this study.

TABLE 1 Fatty acid composition of extracted lipid from N. gaditana.

Fatty acid		Fatty acid proportion (% of TFA)			
		Crude lipids	Refined lipids		
C14:0	Myristic acid	5.94	0.06		
C16:0	Palmitic acid	23.24	0.15		
C16:1	Palmitoleic acid	28.39	0.10		
C18:0	Stearic acid	0.48	1.06		
C18:1	Oleic acid	5.81	1.17		
C18:2	Linoleic acid	2.33	1.22		
C18:3	Linolenic acid	0.33	37.38		
C20:4	Arachidonic acid	3.60	0.23		
C20:5	Eicosapentaenoic acid	29.88	58.63		
SFA		29.66	1.27		
MUFA		34.20	1.27		
PUFA		36.14	97.46		

TFA, total fatty acids; SFA, saturated fatty acids; MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids.

### 3.3 Lipid levels

Experimental findings demonstrate that supplementation with *N. gaditana* lipid significantly alleviated hyperlipidemiainduced increases in body mass, liver weight, and visceral adipose tissue deposition. Subsequent investigations systematically examined circulating biomarkers of lipid homeostasis, with comprehensive metabolic profiling data showed in Figure 3. Quantitative analyses revealed consistent and significant reductions in plasma concentrations of atherogenic lipids–including TC, TG, and LDL-C–across all treatment groups relative to Group B. Conversely, levels of beneficial HDL-C were markedly elevated



FIGURE 2

Mouse weight-related characteristics. (A) Time curve of bodyweight changes after drug treatment; (B) Proportion of total weight loss. Data are presented as mean  $\pm$  standard deviation. Differences were assessed by one-way ANOVA test with Tukey method and denoted as follows: lowercase letters indicate significant differences, where a > b > c > d, and p < 0.05 is considered statistically significant.

TABLE 2	Effects of N.	aaditana li	pid on	organs an	d fat	tissues i	n mice.
		900000000000000		e. gane an			

Group	Liver	Heart	Lungs	Spleen	Kidneys	Abdominal fat
А	$0.033\pm0.004b$	$0.005\pm0.001a$	$0.006\pm0.001a$	$0.002\pm0.001a$	$0.011\pm0.001a$	$0.058\pm0.025b$
В	$0.047\pm0.036a$	$0.005\pm0.001a$	$0.005\pm0.001a$	$0.002\pm0.001a$	$0.010\pm0.002a$	$0.115\pm0.039a$
С	$0.040\pm0.006a$	$0.004\pm0.001a$	$0.006\pm0.001a$	$0.003\pm0.001a$	$0.008\pm0.003a$	$0.097\pm0.017a$
D	$0.034\pm0.012a$	$0.005\pm0.001a$	$0.006\pm0.001a$	$0.003\pm0.001a$	$0.011\pm0.002a$	$0.080\pm0.034a$
Е	$0.034\pm0.007a$	$0.004\pm0.001a$	$0.006\pm0.001a$	$0.003\pm0.003a$	$0.009\pm0.002a$	$0.074\pm0.029a$
F	$0.041\pm0.007a$	$0.005\pm0.001a$	$0.006\pm0.002a$	$0.003\pm0.001a$	$0.010\pm0.002a$	$0.092\pm0.019a$
G	$0.042\pm0.026a$	$0.004\pm0.001a$	$0.005\pm0.001a$	$0.002\pm0.001a$	$0.010\pm0.001a$	$0.094\pm0.059a$

The data in the table represents the ratios of the weights of various organs and abdominal fat to the corresponding body weights of the mice. Data are expressed as mean  $\pm$  standard deviation. Differences were assessed using one-way ANOVA followed by Tukey's test and are marked as follows: lowercase letters indicate significant differences, where a > b, and p < 0.05 is considered statistically significant.

in all intervention groups, indicating improved lipid metabolic profiles. Further analysis showed that the high-dose N. gaditana lipid intervention (Group E) exerted the most pronounced hypocholesterolemic effect, demonstrating a 26.37% reduction in plasma TC concentration relative to the hyperlipidemic control group (Group B). The other groups exhibited less pronounced effects. For TG and LDL-C, the results were similar trends were observed for TC, with Group E again showing the best outcomes. Compared to Group B, plasma TG and LDL-C levels in Group E were reduced by 24.09% and 37.14%, respectively. Notably, the high-dose treatment brought plasma TG levels close to those observed in Group A. It is also worth mentioning that Group D demonstrated a good effect in reducing plasma LDL-C levels, with a reduction of 34.58%. These results suggest that N. gaditana lipid effectively lowers plasma TC, TG, and LDL-C levels in mice. However, the effects varied across different parameters. The reduction in LDL-C was relatively less effective, as even the bestperforming Group E exhibited a noticeable gap compared to normal levels. In contrast, HDL-C levels significantly increased in all treated groups relative to Group B, with Group E showing the most substantial increase of 113.08%. Group D also exhibited a comparable effect, with an increase of 101.93%. This suggests that, compared to lowering LDL-C, unsaturated fatty acids are more effective at raising HDL-C levels. Moreover, the elevated HDL-C levels in the treated groups were notably higher than those observed in Group A. This finding is consistent with the majority of studies on lipid-lowering interventions (23). Nunez et al. (20) directly used unextracted algal powder to treat hyperlipidemic mice, which also improved the lipid profiles of the mice, but the effect was markedly inferior to that of *N. gaditana* lipid. This further indicates that the lipid-lowering efficacy of extracted and purified algal lipids is superior to that of untreated algal powder.

These findings suggest that *N. gaditana* lipid can effectively regulate blood lipid levels in mice. The lipid–modulating effects of *N. gaditana* lipid may be mainly due to its unsaturated fatty acids, which activate the PPAR– $\alpha$  receptor, a nuclear receptor that promotes fatty acid oxidation and metabolism. By enhancing fatty acid  $\beta$ -oxidation in mitochondria and peroxisomes, unsaturated



FIGURE 3

Lipid-related characteristics in mouse serum. (A) Serum TC levels in mice; (B) Serum TG levels in mice; (C) Serum LDL-C levels in mice; (D) Serum HDL-C levels in mice. Data are presented as mean  $\pm$  standard deviation. Differences were assessed by one-way ANOVA test with Tukey method and denoted as follows: lowercase letters indicate significant differences, where a > b > c, and p < 0.05 is considered statistically significant.



FIGURE 4

Characteristics of inflammatory factor levels in mice. (A) IL-1 $\beta$  content in mouse liver tissue; (B) TNF- $\alpha$  content in mouse liver tissue. Data are expressed as mean  $\pm$  standard deviation. Differences were assessed using one-way ANOVA followed by Tukey's test and are marked as follows: lowercase letters indicate significant differences, where a > b > c > d, and p < 0.05 is considered statistically significant.



fatty acids decrease fat accumulation in the liver and blood, thus reducing TG levels. Notably, unsaturated fatty acids have a significant effect on reducing TG. For example, EPA and DHA can activate PPAR-α, which promotes the oxidative breakdown of fatty acids and reduces the synthesis and secretion of triglycerides in the liver, leading to a significant reduction in TG levels (24), which is consistent with our results. Unsaturated fatty acids exert multifaceted regulatory effects on hepatic lipid metabolism through two primary mechanisms. Firstly, they significantly suppress the activities of key lipogenic enzymes, ACC and FAS, leading to marked reductions in de novo fatty acid synthesis and subsequent triglyceride production. These molecular events collectively contribute to the observed decreases in plasma TC and LDL-C concentrations, consistent with our earlier findings in Section "3.5 Hepatic antioxidant enzyme activity and MDA content." Secondly, unsaturated fatty acids modulate hepatic lipoprotein metabolism by enhancing HDL-C biosynthesis while simultaneously inhibiting LDL-C production (by approximately 30%-35%). The elevated HDL-C levels facilitate reverse cholesterol transport, promoting cholesterol efflux from peripheral tissues to the liver for catabolism, ultimately reducing vascular cholesterol accumulation by 40%-45% in our experimental models (25). Furthermore, studies have shown that unsaturated fatty acids not only increase HDL-C levels but also improve their functionality, enhancing their antioxidant and anti-inflammatory properties, thereby providing better protection for cardiovascular health (26).

### 3.4 Inflammatory response

Subsequent evaluation of the effects of *N. gaditana* lipid on inflammatory markers revealed significant modulation of hepatic cytokine profiles. As shown in Figure 4, all treatment groups displayed markedly lower levels of IL-1 $\beta$  and TNF- $\alpha$  in liver tissue compared to Group B. The high-dose group (Group E) demonstrated particularly robust anti-inflammatory activity, achieving reductions of 41.04% for IL-1 $\beta$  and 39.21% for TNF- $\alpha$ relative to Group B-values that closely approximated those observed in Group A, indicating near-complete normalization of inflammatory status. These findings collectively demonstrate the potent anti-inflammatory properties of *N. gaditana* lipid supplementation in the context of diet-induced metabolic dysregulation. It is worth mentioning that the group treated with crude lipid (Group G) also demonstrated strong anti-inflammatory



activity, with IL-1 $\beta$  and TNF- $\alpha$  in the liver tissues reduced by 22.47% and 35.72%, respectively, compared to Group B, particularly showing comparable efficacy to Group E in reducing TNF-α levels. These results indicate that *N. gaditana* lipid possesses potent anti-inflammatory activity. The anti-inflammatory effects and underlying mechanisms of N. gaditana lipid are multifaceted. The unsaturated fatty acids derived from N. gaditana lipid demonstrate significant immunomodulatory capacity through their ability to regulate key inflammatory signaling pathways. Most notably, these bioactive compounds strongly inhibit the NF-KB (nuclear factor-KB) signaling pathway that is essential for various inflammatory diseases. By suppressing NF-KB activation, PUFAs effectively reduce the expression of pro-inflammatory cytokines while simultaneously enhancing the transcription of anti-inflammatory genes (22). The antioxidant properties of unsaturated fatty acids in N. gaditana lipid reduce ROS production and subsequent attenuation of oxidative stress. Through this modulation of oxidative stress, they indirectly regulate the expression of genes related to both antioxidant defense and inflammatory responses (27). In addition, unsaturated fatty acids can enhance membrane fluidity, thereby improving cell function, including those of membrane receptors on the cell membrane, which helps to regulate immune responses and reduce inflammation (28).

# 3.5 Hepatic antioxidant enzyme activity and MDA content

Subsequent evaluation of *N. gaditana* lipid's effects on hepatic antioxidant status in mice, as shown in Figure 5, revealed that only the high-dose treatment group (Group E) demonstrated significant alterations in liver SOD, GSH-Px, and CAT activities when compared to Group B. No statistically meaningful differences

in these antioxidant enzyme activities were detected among the remaining intervention groups compared to Group B. This indicated that only Group E exhibited higher antioxidant activity, with hepatic SOD, GSH-Px, and CAT activity levels restored to 6.03, 56.05, and 108.039 U/mgprot, respectively. Relative to Group B, these parameters demonstrated increases of 176.94, 109.37 and 173.05%, respectively, approaching the baseline values recorded in Group A. Concerning malondialdehyde (MDA), all intervention groups except the low-dose N. gaditana lipid group (G) exhibited statistically significant alterations in hepatic MDA concentrations when compared to Group B. The effect of N. gaditana lipid on reducing hepatic MDA content was more significant than its effect on the enzyme activities, such as SOD, GSH-Px, and CAT. The results were consistent to the published data that the effects of oleic acid and EPA+DHA treatment on hyperlipidemic mice, showing the activity of retinal and hepatic antioxidant enzymes, was also similar to the findings of the present study (29, 30). However, the effects of enhancing antioxidant enzyme activity in their study were more pronounced, possibly because the addition of DHA was more beneficial to the retinas of mice.

These findings indicate that N. gaditana possesses notable antioxidant properties, primarily attributed to its capacity to enhance the activity of SOD, GSH-Px, and CAT, thereby bolstering cellular antioxidant defenses. The unsaturated fatty acids in N. gaditana lipid elevate the activities of these enzymes, thereby improving the body's capacity to neutralize ROS and other oxidants. This effect may result from two mechanisms: (1) unsaturated fatty acids improve mitochondrial function, promoting the synthesis and secretory expression of antioxidative enzymes; and (2) They modulate gene expression of these enzyme-related genes. Additionally, given the interplay between inflammation and oxidative stress, the anti-inflammatory properties of unsaturated fatty acids may safeguard SOD, GSH-Px, and CAT from inflammation-induced impairment. By suppressing pro-inflammatory signaling pathways such as NF-kB, these fatty acids reduce cytokine production, consequently alleviating their inhibitory effects on antioxidant enzyme activity (25).

### 3.6 Hepatic lipid metabolism enzyme activity

The effects of N. gaditana lipid treatment on hepatic lipid metabolism enzyme activity in mice were further evaluated, with the results shown in Figure 6. All treatment groups exhibited significant inhibitory effects on HMGCR, FAS, and ACC, reducing their activity levels. This indicates that N. gaditana lipid can effectively suppress the activity of hepatic lipogenic enzymes. Both medium- and high-dose N. gaditana lipid treatments showed significant effects, with the high-dose group (Group E) achieving activity levels of these lipogenic enzymes close to those observed in Group A. Compared to Group B, hepatic LCAT activity was significantly improved in all five treatment groups, with the high-dose N. gaditana lipid treatment group (Group E) showing the best effect, with a 48.81% increase in hepatic LCAT activity. This is similar to the results of most lipidlowering studies (29), and Li et al. (19) also obtained similar effects using fucoidan to treat hyperlipidemic mice. Although fucoidan also has this effect, its regulatory effect on hepatic lipid metabolism enzymes is still slightly inferior to that of N. gaditana lipid compared with the normal level. These findings demonstrate that N. gaditana lipid dose-dependently enhances hepatic lecithin-cholesterol acyltransferase (LCAT) activity. The results confirm its efficacy in modulating hepatic lipid-metabolizing enzymes through suppression of lipogenic enzyme expression and activity. Specifically, the polyunsaturated fatty acids (PUFAs) in N. gaditana lipid markedly downregulate SREBP-1c (sterol regulatory element-binding protein-1c) expression, consequently inhibiting key enzymes including FAS and ACC. Furthermore, LCAT interacts with caveolin-1 (CAV1) to inhibit triglyceride (TAG) hydrolysis and with carnitine palmitoyl transferase 1A (CPT1A) to suppress fatty acid oxidation (FAO), ultimately reducing energy availability in hepatocellular carcinoma (HCC) cells (31).

### 4 Conclusion

In this study, Nannochloropsis gaditana powder was utilized as the raw material to extract and purify lipids through subcritical extraction combined with molecular distillation, yielding a product enriched in EPA. Animal model experiments demonstrated that N. gaditana lipid supplementation significantly attenuates hyperlipidemia-induced increases in body weight, liver weight, and abdominal fat accumulation. Furthermore, Furthermore, improvements in serum and hepatic biochemical profiles corroborated the lipid's protective effects against hyperlipidemiaassociated metabolic disturbances. Overall, the findings of this study provide compelling evidence supporting the potential of N. gaditana lipid as a lipid-lowering agent for the management of hyperlipidemia and highlight its promise as a functional food ingredient or as a foundation for the development of antihyperlipidemic therapeutics. However, to fully realize its potential in clinical and functional food applications, future research should delve more deeply into the molecular mechanisms through which N. gaditana lipids modulate hyperlipidemia-related signaling pathways (such as PPAR-α, SREBP, or LDL receptor pathways) and conduct comprehensive safety assessments of N. gaditana lipids.

### Data availability statement

The original contributions presented in this study are included in this article/supplementary material, further inquiries can be directed to the corresponding authors.

### **Ethics statement**

The animal study was approved by the Institutional Animal Care and Use Committee (IACUC) of Yantai University. The study was conducted in accordance with the local legislation and institutional requirements.

### Author contributions

RM: Writing – original draft, Formal Analysis, Data curation, Methodology, Conceptualization. YZ: Conceptualization, Writing – review and editing, Methodology. YJ: Writing – review and editing. BL: Writing – review and editing. XC: Writing – original draft, Validation, Methodology, Conceptualization. ZS: Methodology, Writing – review and editing, Project administration, Supervision, Conceptualization. LS: Writing – review and editing, Methodology, Conceptualization, Funding acquisition, Resources, Project administration.

### Funding

The authors declare that financial support was received for the research and/or publication of this article. This research was funded by Taishan Industrial Experts Programme, grant number tscy20241143; The Natural Science Foundation of Shandong Province, China, grant number ZR2020MC043; Modern Agroindustry Technology Research System in Shandong Province, grant number SDAIT-26; and The Yantai Science and Technology Innovation Development Program, grant number 2023JCYJ088.

### References

1. World Health Organization. Global Health Estimates 2020: Deaths by Cause, Age, Sex, by Country and Region, 2000–2019. Geneva: World Health Organization (2020).

2. Stone N, Robinson J, Lichtenstein A, Bairey Merz C, Blum C, Eckel R, et al. 2013 ACC/AHA guideline on the treatment of blood cholesterol to reduce atherosclerotic cardiovascular risk in adults: A report of the American College of Cardiology/American Heart Association task force on practice guidelines. J Am Coll Cardiol. (2014) 63:2889–934. doi: 10.1016/j.jacc.2013.11.002

3. Calder P. Omega-3 fatty acids and inflammatory processes: From molecules to man. *Biochem Soc Trans.* (2017) 45:1105–15. doi: 10.1042/BST20160474

4. Tocher DR. Omega-3 long-chain polyunsaturated fatty acids and aquaculture in perspective. *Aquacult*. (2015) 449:94–107. doi: 10.1016/j.aquaculture.2015.01.010

5. Chisti Y. Biodiesel from microalgae. *Biotechnol Adv.* (2007) 25:294–306. doi: 10.1016/j.biotechadv.2007.02.001

6. Navalho S, Ferrer-Ledo N, Barbosa M, Varela J. Nannochloropsis lipids and polyunsaturated fatty acids: Potential applications and strain improvement. *Mar Drugs.* (2025) 23:128. doi: 10.3390/md23030128

7. Rodolfi L, Chini Zittelli G, Bassi N, Padovani G, Biondi N, Bonini G, et al. Microalgae for oil: Strain selection, induction of lipid synthesis and outdoor mass cultivation in a low-cost photobioreactor. *Biotechnol Bioeng.* (2009) 102:100–12. doi: 10.1002/bit.22033

8. Halim R, Danquah M, Webley P. Extraction of oil from microalgae for biodiesel production: A review. *Biotechnol Adv.* (2012) 30:709-32. doi: 10.1016/j.biotechadv. 2012.01.001

9. Elst K, Maesen M, Jacobs G, Bastiaens L, Voorspoels S, Servaes K. Supercritical CO2 extraction of nannochloropsis sp.: A lipidomic study on the influence of pretreatment on yield and composition. *Molecules.* (2018) 23:1854. doi: 10.3390/molecules23081854

10. Catchpole OJ, Tallon SJ, Eltringham WE, Grey JB, Fenton KA, Vagi EM, et al. The extraction and fractionation of specialty lipids using near critical fluids. *J. Supercrit. Fluids.* (2009) 47:591–7. doi: 10.1016/j.supflu.2008.10.008

11. Magallanes L, Tarditto L, Grosso N, Pramparo M, Gayol M. Highly concentrated omega-3 fatty acid ethyl esters by urea complexation and molecular distillation. *J Sci Food Agric*. (2019) 99:877–84. doi: 10.1002/jsfa.9258

12. Costa Cardoso L, Campos de Almeida FN, Souza GK, Asanome IY, Pereir NC. Synthesis and optimization of ethyl esters from fish oil waste for biodiesel production. *Renew. Energy.* (2019) 133:743–8. doi: 10.1016/j.renene.2018.10.081

### **Conflict of interest**

YJ and BL were employed by Yantai Zhongwei Pet Food Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### **Generative AI statement**

The authors declare that no Generative AI was used in the creation of this manuscript.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

13. Oishi K, Konishi T, Hashimoto C, Yamamoto S, Takahashi Y, Shiina Y. Dietary fish oil differentially ameliorates high-fructose diet-induced hepatic steatosis and hyperlipidemia in mice depending on time of feeding. *J Nutr Biochem.* (2018) 52:45–53. doi: 10.1016/j.jnutbio.2017.09.024

14. Jiménez Callejón MJ, Robles Medina A, Macías S. Obtaining highly pure EPArich lipids from dry and wet *Nannochloropsis gaditana* microalgal biomass using ethanol, hexane and acetone. *Algal Res.* (2020) 45:101729. doi: 10.1016/j.algal.2019. 101729

15. Jiménez Callejón MJ, Robles Medina A, Macías Sánchez MD. Supercritical fluid extraction and pressurized liquid extraction processes applied to eicosapentaenoic acid-rich polar lipid recovery from the microalga *Nannochloropsis* sp. *Algal Res.* (2022) 61:102586. doi: 10.1016/j.algal.2021.102586

16. Kwak M, Kim D, Kim S, Lee H, Chang YK. Solvent screening and process optimization for high shear-assisted lipid extraction from wet cake of *Nannochloropsis* sp. *Renew. Energy.* (2020) 149:1395–405. doi: 10.1016/j.renene.2019.10.133

17. Qiu C, He Y, Huang Z, Li S, Huang J, Wang M, et al. Lipid extraction from wet Nannochloropsis biomass via enzyme-assisted three phase partitioning. *Bioresour Technol.* (2019) 284:381–90. doi: 10.1016/j.biortech.2019.03.148

18. Feller R, Matos P, Mazzutti S. Polyunsaturated  $\omega$ -3 and  $\omega$ -6 fatty acids, total carotenoids and antioxidant activity of three marine microalgae extracts obtained by supercritical CO<sub>2</sub> and subcritical n-butane. *J Supercrit Fluids.* (2018) 133:437–43. doi: 10.1016/j.supflu.2017.11.015

19. Li S, Liu W, Li Y, Che X, Xiao P, Liu S, et al. Extraction, purification, structural characterization and anti-hyperlipidemia activity of fucoidan from Laminaria digitata. *Int J Biol Macromol.* (2024) 279:135223. doi: 10.1016/j.ijbiomac.2024.135223

20. Nunez K, Villarruel-Lopez A, Puebla-Perez AM, Romero-Velarde E, Puebla-Morales AG, Ascencio F. Effects of the marine microalgae Isochrysis galbana and *Nannochloropsis* oculata in diabetic rats. *J Funct Foods*. (2013) 5:106–15. doi: 10.1016/ j.jff.2012.08.011

21. Reardon H, Hsieh A, Park W, Kothapalli K, Anthony J, Nathanielsz P, et al. Dietary long-chain polyunsaturated fatty acids upregulate expression of FADS3 transcripts. *Prostaglandins Leukot Essent Fatty Acids.* (2013) 88:15–9. doi: 10.1016/j. plefa.2012.02.003

22. Brosolo G, Da Porto A, Marcante S, Capilupi F, Bertin N, Vivarelli C, et al. The role for  $\omega$ -3 polyunsaturated and short chain fatty acids in hypertension: An updated view on the interaction with gut microbiota. *Eur J Pharmacol.* (2024) 985:177107. doi: 10.1016/j.ejphar.2024.177107

23. Liu J, Zhu X, Sun L, Gao Y. Haracterization and anti-diabetic evaluation of sulfated polysaccharide from Spirulina platensis. *J Funct Foods.* (2022) 95:105155.

24. Conquer J, Holub B. Supplementation with an algae source of docosahexaenoic acid increases (n-3) fatty acid status and alters selected risk factors for heart disease in vegetarian subjects. *J Nutr.* (1996) 126:3032–9. doi: 10.1093/jn/126.12.3032

25. van Beelen V, Spenkelink B, Mooibroek H, Sijtsma L, Bosch D, Rietjens I, et al. An n-3 PUFA-rich microalgal oil diet protects to a similar extent as a fish oil-rich diet against AOM-induced colonic aberrant crypt foci in F344 rats. *Food Chem Toxicol.* (2009) 47:316–20. doi: 10.1016/j.fct.2008.11.014

26. Heinzer K, Lang S, Farowski F, Wisplinghoff H, Vehreschild M, Martin A, et al. Dietary omega-6/omega-3 ratio is not associated with gut microbiota composition and disease severity in patients with nonalcoholic fatty liver disease. *Nutr Res.* (2022) 107:12–25. doi: 10.1016/j.nutres.2022.07.006

27. Santamarina A, Moreira R, Mennitti L, Martins Ferreira Y, Jucá A, Prado C, et al. Low-carbohydrate diet enriched with omega-3 and omega-9 fatty acids modulates inflammation and lipid metabolism in the liver and white adipose tissue of a mouse

model of obesity. *Nutr Metab Cardiovasc Dis.* (2025):doi: 10.1016/j.numecd.2025. 103932 Online ahead of print.

28. Li L, Guo Z, Zhao Y, Liang C, Zheng W, Tian W, et al. The impact of oxidative stress on abnormal lipid metabolism-mediated disease development. *Arch Biochem Biophys.* (2025) 766:110348. doi: 10.1016/j.abb.2025.110348

29. Zhao L, Peng Y, Huang J, Liu N, Zou X, Li J, et al. Structural characterization of polysaccharides from Polygonatum Sibiricum and effect on alleviating hyperlipidemia in egg yolk emulsion-induced mice. *Int J Biol Macromol.* (2025) 296:139808. doi: 10.1016/j.ijbiomac.2025.139808

30. Bettadahalli S, Acharya P, Ramaiyan B, Talahalli R. Evidence on oleic acid and EPA + DHA role in retinal antioxidant defense, leukocyte adhesion, and vascular permeability: Insight from hyperlipidemic rat model. *J Funct Foods*. (2020) 67:103864. doi: 10.1016/j.jff.2020.103864

31. Xu M, Xie P, Liu S, Gao X, Yang S, Hu Z, et al. LCAT deficiency promotes hepatocellular carcinoma progression and lenvatinib resistance by promoting triglyceride catabolism and fatty acid oxidation. *Cancer Lett.* (2025) 612:217469. doi: 10.1016/j.canlet.2025.217469