



## OPEN ACCESS

## EDITED BY

Xiaolong Ji,  
Zhengzhou University of Light Industry, China

## REVIEWED BY

Hridayesh Anuragi,  
Indian Council of Agricultural Research  
(ICAR), India  
Xuefeng Guo,  
Tarim University, China  
Jing Zhang,  
Academy of National Food and Strategic  
Reserves Administration, China

## \*CORRESPONDENCE

Hosameldeen Mohamed Husien

✉ 0086@43yzu.edu.cn

Mengzhi Wang

✉ mengzhiwangyz@126.com

<sup>†</sup>These authors have contributed equally to  
this work

RECEIVED 21 April 2025

ACCEPTED 04 June 2025

PUBLISHED 20 June 2025

## CITATION

Mohai Ud Din R, Eman S, Zafar MH, Chong Z,  
Saleh AA, Husien HM and Wang M (2025)  
*Moringa oleifera* as a multifunctional feed  
additive: synergistic nutritional and  
immunomodulatory mechanisms in livestock  
production.  
*Front. Nutr.* 12:1615349.  
doi: 10.3389/fnut.2025.1615349

## COPYRIGHT

© 2025 Mohai Ud Din, Eman, Zafar, Chong,  
Saleh, Husien and Wang. This is an  
open-access article distributed under the  
terms of the [Creative Commons Attribution  
License \(CC BY\)](#). The use, distribution or  
reproduction in other forums is permitted,  
provided the original author(s) and the  
copyright owner(s) are credited and that the  
original publication in this journal is cited, in  
accordance with accepted academic  
practice. No use, distribution or reproduction  
is permitted which does not comply with  
these terms.

# *Moringa oleifera* as a multifunctional feed additive: synergistic nutritional and immunomodulatory mechanisms in livestock production

Raza Mohai Ud Din<sup>1†</sup>, Salwa Eman<sup>1†</sup>,  
Muhammad Hammad Zafar<sup>1</sup>, Zhang Chong<sup>1</sup>, Ahmed A. Saleh<sup>2,3</sup>,  
Hosameldeen Mohamed Husien<sup>1,4\*</sup> and Mengzhi Wang<sup>1,5\*</sup>

<sup>1</sup>Laboratory of Metabolic Manipulation of Herbivorous Animal Nutrition, College of Animal Science and Technology, Yangzhou University, Yangzhou, Jiangsu, China, <sup>2</sup>College of Animal Science and Technology, Yangzhou University, Yangzhou, Jiangsu, China, <sup>3</sup>Department of Animal and Fish Production, Faculty of Agriculture (Al-Shatby), Alexandria University, Alexandria, Egypt, <sup>4</sup>College of Veterinary Medicine, Albutana University, Rufaa, Sudan, <sup>5</sup>State Key Laboratory of Sheep Genetic Improvement and Healthy Production, Xinjiang Academy of Agricultural Reclamation Sciences, Shihezi, Xinjiang, China

Investigating *Moringa oleifera* (*M. oleifera*)' is potential as a livestock feed additive, this review explores its nutritional and phytochemical profiles and its mechanistic roles, specifically focusing on its immunomodulatory and antioxidant properties. *M. oleifera* is a rich source of diverse bioactive compounds, including polyphenols, alkaloids, terpenoids, flavonoids (e.g., quercetin, kaempferol), saponins, and tocopherols. These compounds exert significant immunomodulatory effects by modulating cytokine production and immune cell activity. Notably, *Moringa*-derived arabinogalactans (water-soluble polysaccharides comprising arabinose and galactose monomers) activate the gut-associated immune system through beneficial modulation of gut microbiota composition, increasing genera such as *Muribaculaceae* and *Lactobacillus*. The immunomodulatory activity is mediated via multiple pathways, including the promotion of anti-inflammatory cytokine secretion (e.g., IL-10) and the inhibition of pro-inflammatory enzymes [e.g., cyclooxygenase-2 (COX-2)]. Furthermore, *M. oleifera* exhibits potent antioxidant capabilities by enhancing endogenous defenses, neutralizing reactive oxygen species, and mitigating oxidative stress-induced tissue damage. These findings underscore *M. oleifera* is potential to enhance disease resistance and immune function in animals, advocating for its strategic incorporation into sustainable animal nutrition practices.

## KEYWORDS

*Moringa oleifera*, nutritional profile, immunomodulatory mechanism, animal, feed additive

## Introduction

The entire livestock sector is facing unprecedented threats due to rising global consumption of animal-derived products like meat, milk, and eggs. Looking at the current situation of animal global meat production, there is a 55 percent increase between 2000 and 2022, reaching a total of 361 million tons (1). A key development during this period was the rise of chicken, which accounted for the largest share of this growth and surpassed pork as the most produced

meat globally in 2022. Asia, home to nearly 60% of the world's population, is an unparalleled force in global livestock production and consumption. The sheer demographic weight and ongoing economic growth within the continent position it as the primary driver of global livestock trends (2). China stands as the world's largest livestock-producing country, a position that grants its internal dynamics significant influence over global markets and trends. Many publications have predicted the changes by 2050 regarding livestock production and consumption. By 2050, the demand for animal products globally is estimated to increase by 60 to 70%, and developing countries will have a bulk of this increase (3). Due to population growth, urbanization, and income growth, livestock, along with their products will change rapidly by 2050. While in animal health, welfare, and food security concerns, biotechnology and nanotechnology will play a key role (4). Poultry meat demand in sub-Saharan Africa is expected to rise by 214% by 2050, and for pork, by 161%, due to the main factor- urbanization, as well as demand for animal-sourced foods (5). China's livestock industry change has severe global implications, especially large changes and effects expected in the 2050s (6). These changes have created an alarming situation regarding food security. Climate change affects livestock production and emission of greenhouse gases, hence the need to develop appropriate technologies for sustainable production and contribute to the global food supply (7). This growth requires improvement in feed supply technologies to increase productivity efficiently and cost-effectively.

Now the major inputs in livestock feed production are the traditional crops such as maize and soybean meals, which are the energy and source of protein, respectively, in animal feed. Farm animals account for over 30% of global food consumption, primarily relying on grains, with soybeans making up 90% of that total. A minimal amount of these grains is utilized within factory farming operations (8). However, the cultivation of these crops has its economic and environmental consequences. Soybean, for example, is one of the causes of deforestation more particularly in South America, and they also significantly contribute to greenhouse gas emissions (9). Soybean trade impacts the environment and socio-economy of the world, and therefore, there is a need to find ways to increase sustainability in the trade (10). Moreover, competition drawn from feeding humans and animals using these crops continues to complicate food security issues around the world (11, 12). The development of the "maize/soybean system" has taken place and changed the structure of competition and interaction between human and animal consumption of vegetable proteins (12).

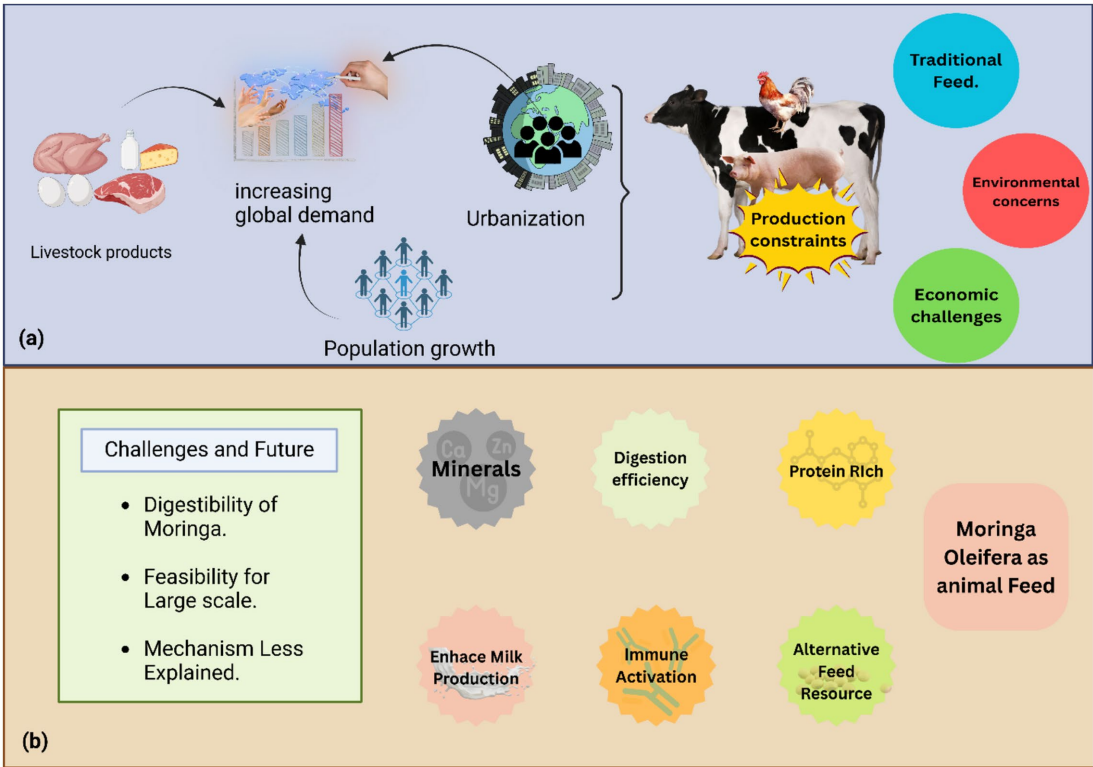
Environmental sustainability is another pressing concern. Feed production currently accounts for a significant proportion of global agricultural land use, water consumption, and nitrogen pollution. Food systems are dependent on livestock greenhouse gas emissions, which can only be tackled on a global scale while supporting food security (13). The cultivation of soybean and maize, and exclusive for the feeding of stock rations, has been linked with undesirable effects on lands and water resources, planetary nutrient imbalances (14). These environmental impacts highlight the urgent need to identify and integrate alternative feed resources.

In addition to environmental challenges, economic volatility in feed prices poses significant issues for farmers. Grain prices in particular have trended higher in 2005 and 2006, which has put pressure on livestock feed costs and has also resulted in high volatility

shocks (15). High and frequently changing feed costs, as well as high and rapidly changing output prices, are challenges to livestock farmers (16).

To meet the rising demand for livestock and poultry feed, researchers have sought alternatives to its traditional ingredients, which could be novel sources of protein and energy. Following recent years, innovations like the use of agricultural byproducts, insects, microalgae, and drought-resistant plants have increased. Tree leaves, along with traditional crops like *camelina* and oil seeds, offer promising alternative feed resources that can either replace or complement conventional crops in ruminant diets, leading to improved animal performance in a sustainable manner (17). Moreover, the livestock sector has identified insect-based products as a viable option to support sustainable development within the industry (18). Incorporating tree leaves as a feed ingredient presents a beneficial strategy, as they typically possess higher nutritional value than grasses, making them more appealing to herbivores (19).

A versatile plant, *Moringa oleifera* (*M. oleifera*), with a lot of nutrients, can be used as an alternative feed and forage to traditional animal feed and fodder with no negative effects on health, survival, and reproduction (20). *M. oleifera* holds significant promise for addressing the livestock feeding crisis due to its rich nutrient content, elevated protein biological value, and positive effects on animal nutrition (8). Using *M. oleifera* leaves in place of sunflower seed cake for goat feed promotes dry matter consumption and improves product breakdown capabilities without losing nitrogen content (21). It contains phenolic and flavonoid compounds that have been associated with enhanced health, improved feed conversion efficiency, and better growth performance in livestock (20, 22). Due to its abundant nutrients, high protein biological value, good feeding effect, and great potential make *M. oleifera* is suitable mean to deal with the feeding crisis for livestock (8). The phenolics in *M. oleifera* leaves include a wide variety of kaempferol derivatives, caffeoylquinic acid, and feruloylquinic acid, and are responsible for their antioxidant capacity (23). Antioxidant potential in *M. oleifera* leaves appears very strong when combined with flavonoids, flavanols, phenolics, and proanthocyanidins elements (24). The direct radical scavenging action and indirect enhancement of cellular antioxidant defenses are expressed by antioxidant compounds in *M. oleifera* leaves. These substances completely remove free radicals while boosting antioxidant enzyme function, including superoxide dismutase and catalase, to lower oxidative stress levels (23, 25). Besides having antioxidant ability, *M. oleifera* leaves are rich in protein, minerals, vitamins, and essential amino acids (26). Analyses show *M. oleifera* leaves contain 28.7% crude protein alongside 7.1% fat, while the protein content exists as insoluble compounds that display poor *in vitro* digestibility (27). Ruminant farmers can prepare concentrated mixtures at 20% concentration, which improves goat performance while reducing methane releases (28). The introduction is explained graphically in Figure 1. Considering the importance of *M. oleifera*, in this review, we will give detailed nutritional composition of *M. oleifera* and the mechanism of action of its bioactive components as immunomodulator. Application of *M. oleifera* in animal feed is discussed in detail, along with challenges of *M. oleifera* as animal feed and future directions. This review systematically evaluates *M. oleifera* nutritional-phytochemical synergy and its translational potential for sustainable livestock production.



**FIGURE 1**  
(a) This figure illustrates how increasing global demand for livestock products, driven by population growth and urbanization, creates operational difficulties for the worldwide livestock industry, including production constraints, reliance on traditional feed, environmental concerns from maize and soybean cultivation, and economic risks from variable feed costs. (b) This figure presents *M. oleifera* as a sustainable and alternative animal feed, highlighting its protein and mineral richness, and its potential to enhance digestion efficiency, activate immune responses, and increase milk production, thereby mitigating environmental damage and market fluctuations. The figure also identifies key research areas for *M. oleifera*'s effective implementation, including its digestibility, large-scale feasibility, and precise mechanisms of action.

Taxonomy, botanical characters, and cultivation

The initial description of *M. oleifera* was made in 1785 by the French naturalist Jean Baptiste Lamarck. The name “*M. oleifera*” is thought to originate from the Tamil word “murungai,” which translates to “twisted hand length structure of the young *M. oleifera* fruit.” In Latin, “oleum” signifies “oil,” while “ferre” means “to bear” (29). *M. oleifera* from the Moringaceae family represents a widely grown plant species that demonstrates significant medical characteristics as well as valuable nutritional benefits (30). Which has 13 species of different trees and shrubs, having the potential to be used for medicinal and nutritional purposes (31). All species have their native origin (Table 1).

Among these, *M. oleifera* stands out as the most economically important variety that grows throughout Asia and is spreading across Africa and America (31, 32). It is a widely planted tree because it possesses high nutritional value while offering prospects to fight malnutrition (33).

*M. oleifera* is considered to have high phytonutrient content, with the ability of drought, is used to deal with malnutrition, and also has nutraceutical properties (34). It is a perennial tree with a height range from 5 to 12 cm (35).

*M. oleifera* thrives in tropical and subtropical regions, especially in areas with average annual rainfall between 1,000 and 2,000 mm and

TABLE 1 Species of the Moringaceae family with their native origin.

Species	Native region
<i>M. rivae</i>	Indigenous to Ethiopia and Kenya
<i>M. pygmaea</i>	Native to Somalia
<i>M. arborea</i>	Indigenous to Kenya
<i>M. borziana</i>	Native to Kenya and Somalia
<i>M. stenopetala</i>	Indigenous to Ethiopia and Kenya
<i>M. ovalifolia</i>	Indigenous to Angola and Namibia
<i>M. longituba</i>	Indigenous to Somalia, Kenya, and Ethiopia
<i>M. ruspoliana</i>	Native to Ethiopia
<i>M. peregrine</i>	Indigenous to the Horn of Africa and the Red Sea
<i>M. drouhardii</i> , <i>M. hildebrandi</i>	Indigenous to Madagascar
<i>M. concanensis</i>	Indigenous to the sub-Himalayan tracts of Northern
<i>M. oleifera lam</i>	Native to northwestern India and northeastern Pakistan

high levels of solar radiation (36). It has various climatic adaptations, including temperate, subtropical, and tropical regions, which makes it suitable for the semi-arid regions as a contributor to nutritional security (37). High vigor and resistance to salinity stress are promoted

more effectively when seeds of *M. oleifera* are pre-soaked for 24 h, making them suitable for planting in areas that are subjected to salinity (38). *M. oleifera* is a high-yielder in terms of biomass production. The annual biomass yield of *M. oleifera* is reported as 43 to 115 t/h (39). The leaf production is 1–5 kg per tree annually, which is equal to 10,000–50,000 kg/h if plants are cultivated at 1 m × 1 m spacing (40). Due to its distinctive cultivation attributes, *M. oleifera* is currently grown in India, southern China, and certain regions of Africa (41).

## Nutritional and phytochemical profile

*M. oleifera* is a superior nutritional source because of its extensive nutritional benefits, which make it an important supplement for livestock feed. *M. oleifera* demonstrates remarkable potential to solve livestock feeding problems through its supply of rich nutrients and excellent protein value, alongside positive nutritional advantages (8). Extensive research analysis has reported that *M. oleifera* leaves have great amounts of protein, vitamins, and amino acids, along with bioactive components that are beneficial to livestock. It offers valuable nutrient content as animal feed because it contains significant amounts of protein, along with carotenoids and minerals together with vitamins, and phytochemicals (42).

## Macronutrients and minerals

*M. oleifera* leaves are a rich source of essential macronutrients, providing building blocks for animal growth, development, and productivity. These macronutrients, including protein, lipids, and carbohydrates, significantly influence the overall nutritional value of *M. oleifera* as a feed resource. The leaves of *M. oleifera* contains 19.34–28.7%, providing a rich source of essential amino acids required for muscle growth and maintenance (27). Due to its high protein, *M. oleifera* is comparable with other conventional feed resources like maize in terms of percentage and with soybean, having a comparatively similar profile of amino acids (Table 2). The analysis of dietary fiber becomes essential for understanding plant cell wall nutritional value and its effects on animal digestive and absorption processes (43). *M. oleifera* leaves contain approximately 8.07% crude fiber, contributing to their overall nutritional profile (44). In general, the feed source with

low fiber content is considered good due to better digestibility. Another appealing feature of *M. oleifera* is its content. In recent studies, the mineral content in the form of ash is reported to be 11.65% in *M. oleifera* leaves (45), which is significantly higher than soybean meal, which is 5.6–7.2 (46). Analysis of *M. oleifera* leaves cultivated in Gaborone, Botswana, revealed a variable ash content ranging from 5.6 to 9.1%, with a mean value of 7.34%. This variation underscores the influence of environmental factors and sample origin on the mineral composition of *M. oleifera* leaves (47). Another experiment testified to an ash content of 6.00% for *M. oleifera* leaf protein concentrate (48). The mineral composition of *M. oleifera* leaves exhibits variability, influenced by factors such as edaphic conditions and environmental parameters during cultivation. For instance, research has demonstrated comparable calcium concentrations in *M. oleifera* leaves (11,153 mg/kg) and roots (12,834 mg/kg), while seed calcium levels (565 mg/kg) were significantly lower. This observation highlights the division and difference accumulation of minerals within different plant tissues (49). There is a considerable amount of lipids in the leaves of *M. oleifera*, contributing to their nutritional value. On the dry meter basis, analysis of *M. oleifera* leaves samples have a range of lipid concentration from 1.7 to 10.42%. This variation is likely attributable to different factors, including the specific source of leaf sample, the analytical methods employed for lipid determination, and environmental conditions during cultivation (47). A mean lipid content of  $7.8 \pm 0.13\%$  is shown by analysis of *M. oleifera* leaves from Botswana and Gaborone. This value exceeds previously reported values of 2.3, 5.2, and 3.0%, pointing out potential regional changes in *M. oleifera* lipid composition (50). The lipid fraction of *M. oleifera* leaves are characterized by the predominance of unsaturated fatty acids, notably palmitic, oleic, and linoleic acids (51). These purchases fulfill essential physiological roles and contribute to the energetic value of the leaves. A monounsaturated fatty acid, Oleic acid, has been implicated in the modulation of inflammatory responses and positive cardiovascular health outcomes (47).

## Amino acids

Serving as the fundamental building blocks of proteins, amino acids are essential components in animal feed. Although they have a structural role, they also participate in cellular processes, including gene expression, cell signaling, and metabolic regulation. Therefore, that's sufficient supply of amino acids is necessary for supporting overall well-being, optimal growth, development, reproduction, and lactation of animals (52). The metabolic pathways that are crucial for sporting growth, reproductive, and lactation functions are organized by amino acids, so in that way contributing to enhanced animal health (53). *M. oleifera*, due to its amino acid profile, is presented as a primary amino acid supplement in animal feed formulations, specifically when integrated with conventional forages (8). Phytochemical analyses have revealed the presence of 16–19 amino acids in *M. oleifera*, encompassing all ten essential amino acids: threonine, tyrosine, methionine, valine, phenylalanine, isoleucine, leucine, histidine, lysine, and tryptophan (54). *M. oleifera* exhibits comparatively elevated levels of lysine, leucine, histidine, glutamic acid, valine, isoleucine, alanine, phenylalanine, and arginine relative to other woody plant species (55). Some recent studies have reported *M. oleifera* is a notable source of essential amino acids, including

TABLE 2 Proximate analysis: comparison of *M. oleifera* leaf powder with soybean meals and maize on a dry matter basis.

Nutrient	<i>M. oleifera</i> leaves (dry matter basis) g/100 g	Soybean meal (dry matter basis) g/100 g	Corn (maize) (dry matter basis) g/100 g
Crude protein	25–30	43.8–49.9	8–10
Crude fiber	8.07	5–7	2–3
Crude fat	2–5	18–20	3–4
Carbohydrate	40–45	35–40	70–75
ash	11.65	5–7	2–3
References	(163)	(8, 46)	(8, 164)



threonine, valine, methionine, leucine, isoleucine, phenylalanine, histidine, lysine, and arginine (56, 57). Amino acids that cannot be synthesized *de novo* by animals are classified as essential. Efficient protein synthesis depends on the availability of both essential and non-essential amino acids at the ribosomal site, in proportions commensurate with the animal's physiological needs. A deficit in any single amino acid can constrain the utilization of other amino acids within the dietary protein. The amino acid that initially restricts the rate of protein synthesis is defined as the first limiting amino acid. The second limiting amino acid, representing the next most deficient amino acid, can also negatively impact growth even when the first limiting amino acid is supplemented (58). For instance, in weaned calves consuming a corn and soybean meal-based diet, methionine has been identified as the first limiting amino acid, with lysine subsequently becoming limiting (59). Researchers have found that Supplementation with synthetic methionine or methionine-rich feed ingredients can improve dietary amino acid balance and consequently enhance animal performance (60, 61). Maintaining an appropriate balance between essential and non-essential amino acids in animal feed formulations is vital for ensuring optimal nutrition. As indicated in Table 3, *M. oleifera* leaves provide a variety of amino acids, with essential amino acids making up over 50% of the total amino acid content. Although the methionine content in *M. oleifera* leaves exceeds that of corn meal, it remains about two-thirds of the amount found in soybean meal (Table 3). Sulfur-containing amino acids are crucial for preserving cellular integrity and may also contribute to the detoxification of heavy metals through chelation (62). Dietary

supplementation of cystine, a sulfur-containing amino acid, may be necessary in *M. oleifera* leaf meal formulations to ensure that animal requirements for sulfur-containing amino acids are met.

## Mineral content

Ash content is often regarded as a measure of total mineral content. *M. oleifera* leaves serve as an important source of essential minerals, such as calcium, iron, potassium, and sodium (63). The levels of these minerals in *M. oleifera* leaves are generally higher compared to those found in other tree leaf species. Calcium ions are crucial for various cellular functions, including the regulation of cell motility, gene transcription, muscle contraction, and exocytosis (64, 65). *M. oleifera* leaves exhibit higher calcium content and bio accessibility compared to spinach and sweet potato leaves, suggesting their potential to enhance calcium intake, particularly in tropical and warm temperate regions (66). Notably, iron deficiency is frequently observed in many plant-based foods, except those derived from *M. oleifera* leaves. *M. oleifera* leaves provide substantially higher iron levels compared to other plant sources; for example, the iron content of *M. oleifera* leaves is reportedly 25 times greater than that of spinach (67). While *M. oleifera* leaves are a source of magnesium, which can positively influence milk yield and composition, for example, Magnesium supplementation in cattle diets has been reported to increase milk fat concentration and yield, with one study noting a 12% increase within 4 days (68). The recommendation to add excess magnesium salt to cattle feed is not universally supported. Although magnesium is essential for various physiological functions, including milk production, excessive intake can have detrimental effects. Cows do possess homeostatic mechanisms to regulate mineral balance, but their capacity to handle excess magnesium is limited. Hypermagnesemia, a condition characterized by elevated magnesium levels in the blood, can result from over-supplementation and lead to serious health problems, including muscle weakness, cardiac arrhythmias, and even death (69). Therefore, determining the appropriate magnesium level in cattle feed should be based on the animal's growth stage, production status, and dietary context, rather than simply adding excess magnesium (Table 4).

## Bioactive compounds and their distribution

*M. oleifera*, contributing to its recognized medicinal, nutritional, and therapeutic properties, presents a notable abundance of bioactive compounds (70). Bioactive compounds in *M. oleifera* include polyphenols, flavonoids, carotenoids, terpenoids, alkaloids, glucosinolates, tocopherols, and saponins. These compounds play important roles in enhancing animal health and optimizing livestock productivity (71). *M. oleifera* tree with its seeds, leaves, and bark packed with health boosting compounds like polyphenols, vitamins, and essential amino acids is considered a nutritional powerhouse (72). For example, glucosinolates and flavonoids play a crucial role as natural immune regulators, helping animals to combat infections and stress (73). Moreover, its saponins and tannins improve metabolic efficiency and nutrient absorption in animals, leading to healthier herds (74).

TABLE 3 Comparison: amino acid profile (g/100 g dry weight) of *M. oleifera* with conventional feed resources.

Amino acids	<i>M. oleifera</i> leaves (g/100 g)	Soybean meals (g/100 g)	Corn meal (g/100 g)
<b>Essential</b>			
Leucine	1.96	2.75	–
Lysine	1.637	2.43	0.22
Valine	1.413	1.70	0.26
Isoleucine	1.177	1.57	0.26
Phenylalanine	1.64	1.79	0.31
Methionine	0.297	0.60	0.43
Cystine	0.01	0.62	0.34
Tryptophan	0.486	0.64	1.03
Threonine	1.357	1.44	0.40
<b>Non-essential</b>			
Alanine	3.033	3.033	1.25
Aspartic Acid	1.43	1.43	1.97
Glutamic Acid	2.53	–	–
Glycine	1.533	2.048	–
Proline	1.203	–	–
Histidine	0.72	1.148	0.23
Serine	1.087	2.378	–
Tyrosine	2.65	1.53	0.08
Reference	(54)	(8, 165)	(8, 166)

TABLE 4 Comparison of mineral profile between *M. oleifera*, soybean, and maize.

Mineral	<i>M. oleifera</i> (mg/100 g)	Maize (mg/100 g)	Soybean (mg/100 g)
Ca	2016.5	10	310–1,593
K	1845	286	1,548–2,190
Na	8.13	15.9	–
Fe	19.37	2.3	58.22–172
Mg	322.5	139	280–580
Mn	1.6	–	41–193
Cr	0.02	–	–
Al	1.7	–	–
References	(163)	(164)	(167)

The leaves of *M. oleifera* are a rich source of nutrients, loaded with bioactive substances like phytosterols, polyphenols, and essential vitamins that actively reinforce livestock health. Flavonoids, such as kaempferol and quercetin, are among the most important immunomodulatory compounds found in leaves of *M. oleifera* (75). These compounds have been shown to influence the activity of immune cells and production of cytokines to modulate the cellular immune responses (76). *M. oleifera* is renowned for its diverse phytochemical composition. This includes a variety of polysaccharides contributing to their reported health benefits (77). Polysaccharides are complex carbohydrates made up of monosaccharide units, such as galactose, arabinose, rhamnose, glucose, and xylose (45). Among these, Arabinogalactan is identified as a significant polysaccharide found in *M. oleifera* leaves (78). These leaves are abundant in essential nutrients, including minerals, protein, and vitamins, as well as a diverse array of bioactive compounds such as saponins, tannins, and flavonoids (79). These elements take part in observed, antioxidant, anti-inflammatory, and immunomodulatory effects of plants (80, 81). *M. oleifera* pods are also a rich nutrient source and have bioactive components. They have protein, fiber, and immunoglobulin data, which are important components of immune function. Studies have shown that pods of *M. oleifera* can boost cell-mediated immunity in broiler chicken (82). Similarly, the seeds of *M. oleifera* are also a good source of oil and contain bioactive components like tannins and saponins. These cells can modulate immune response and have shown influence on immune cell activity (83). There is an enzyme in *M. oleifera* seeds called myrosinase, which can catalyze the production of isothiocyanates on plant damage or processing (84). The seeds of *M. oleifera* have shown anti-cancer activity by preventing cancer cell proliferation (85). They also have antimicrobial and anti-inflammatory activities (86). The detailed bioactive components in different parts of *M. oleifera* in graphically shown in Figure 2.

## Immunomodulatory mechanisms

### Gut microbiota modulation

Immunomodulation in livestock involves the targeted manipulation of the immune system to strengthen immune responses, ultimately leading to improved disease resistance and overall health

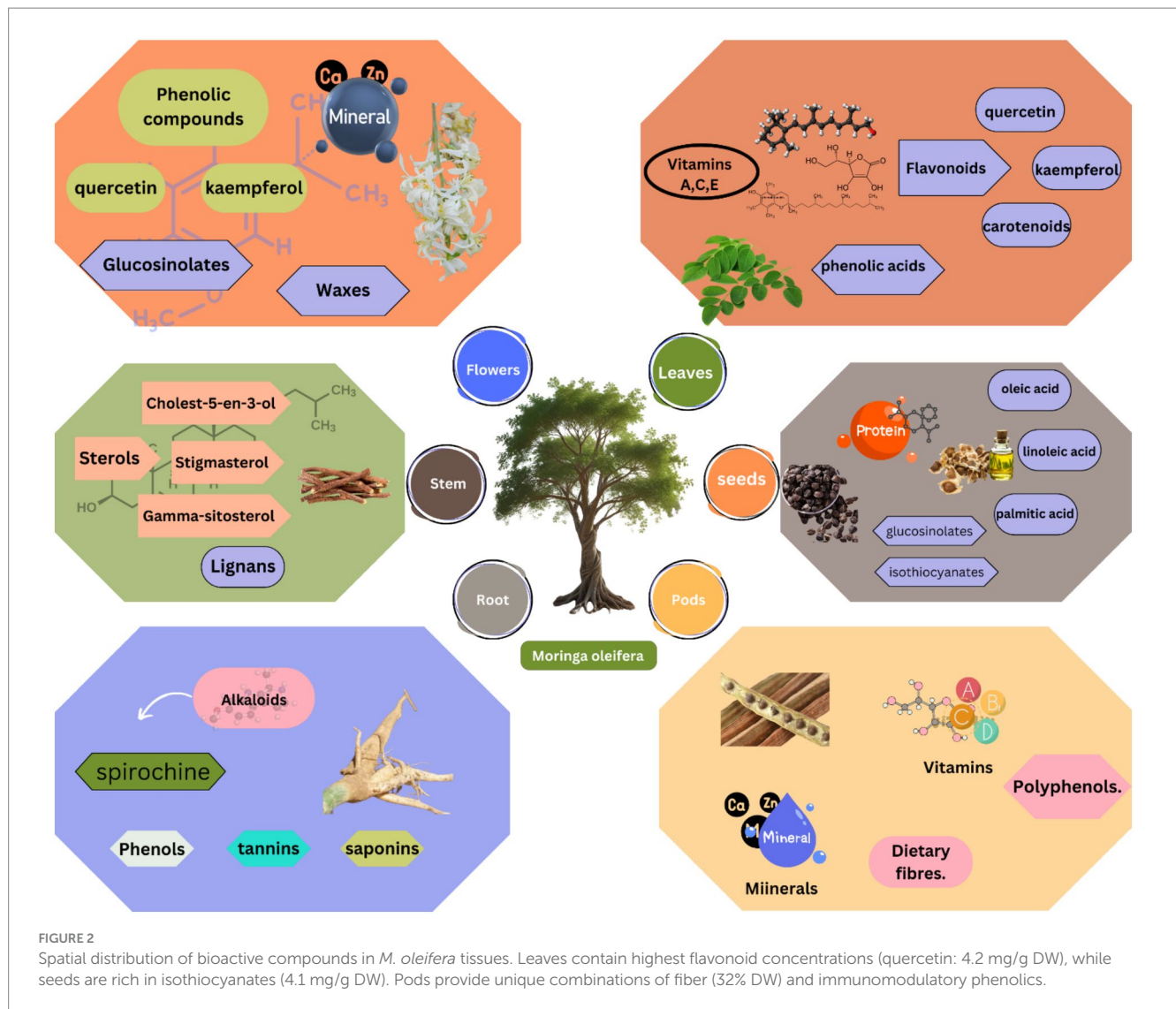
(87). Dietary immunomodulation involves the incorporation of specific nutrients and bioactive compounds into animal feed to optimize immune function (88). For example, probiotics have demonstrated efficacy in improving livestock health by modulating gut microbiota composition and stimulating host immune responses through the secretion of specific factors and competitive exclusion of pathogenic bacteria (89). Likewise, the administration of immunomodulatory feed additives in cattle has been employed to modulate physiological parameters and enhance performance under stressful conditions, such as transportation (90). *M. oleifera*, commonly known as the “miracle tree,” is recognized for its immunomodulatory effects (91). These properties are attributed to its diverse array of bioactive compounds, including essential amino acids, oleic acid, vitamins, flavonoids, polyphenols, and minerals (92). *M. oleifera* contains flavonoids, like kaempferol and quercetin, that express significant anti-inflammatory activity (91). The activity of pro-inflammatory enzymes, such as cyclooxygenase and lipoxygenase is inhibited by these compounds to reduce the production of inflammatory mediators (93). This mechanism takes part in the reduction of inflammation and modulation of immune response in animals. There are different pathways for immunomodulation in animals, including Gut-associated immune system activation, antioxidant defense, and anti-inflammatory action. Comparison of bioactive compounds and immunomodulatory properties of *M. oleifera* with conventional feed crops (Supplementary Table S1).

### Gut-associated immune system activation

The gut, being the largest immunological organ, plays an important role both in digestion and nutrient absorption (94). The intestine of animals contains a vast and complex population of microorganisms, comprising billions of bacteria (95). These microbes play a critical role in nutrient absorption and digestion, taking part significantly in the body's immune function and participating in a range of other biochemical and physiological processes (96). The composition of the intestinal microbiota of a healthy animal is predominantly composed of the phyla *Bacteroidetes*, *Firmicutes*, *Proteobacteria*, and *Actinobacteria*. *Bacteroidetes* and *Firmicutes* are particularly represented by the most abundant phyla, often covering 90% of the total intestinal microbial community (97). Any changes in intestinal flora can induce pathological changes within the intestinal tissue. Moreover, such disruption can contribute to the formation of carcinogenic compounds and chronic inflammation, thereby causing a significant risk to animal health (98).

Polysaccharides present in *M. oleifera* has been linked with different biological activities, including antioxidant properties, immunomodulatory effects, and potential antimicrobial actions (78). The diverse and significant biological activities of polysaccharides extracted from *M. oleifera* have been highlighted by recent researchers, leading to their increased prominence. Research teams have isolated and characterized MOP-1 as a newly discovered arabinogalactan that shows efficient *in vitro* antioxidant effects from *M. oleifera* leaves (99). Dong et al. (100) extracted MOP-2 from *M. oleifera* leaves and then examined the *in vitro* immunomodulatory activity.

The immunomodulatory mechanism of *M. oleifera* leaf polysaccharides has been shown in recent studies. For instance, a study by Mohamed Husien et al. (78) has shown that high doses of



*M. oleifera* polysaccharides promote intestinal health in UC mice by modulating gut microbiome compositions. The mechanism of action of *M. oleifera* in gut-associated immunomodulation is shown in Figure 3. Inflammatory Bowel Disease (IBD) is an inflammatory conditions that affect the gastrointestinal tract and can cause an increase in *Bacteroides* as reported previously (101). Treatment with MOLF-H significantly reduces *Bacteroides* abundance (29% decrease,  $p < 0.05$ ) while increasing *Firmicutes* (40% rise) in DSS-induced colitis models (101). This observation aligns with a prior study that reported a 40% higher prevalence of *Firmicutes* compared to *Bacteroides* in mice subjected to a high-fat diet (102). *Bacteroides* and a few *Firmicutes* species, notably *Bacteroides* and *Lactobacillus*, have been implicated in modulating physiological conditions in mice subjected to dextran sodium sulfate (DSS) treatment (103). A decrease in *Lactobacillus* abundance has been correlated with an increase in ulcerative colitis (UC) induced by DSS. This suggests that *Lactobacillus* plays a beneficial role in immunomodulation (104). The study demonstrated that treatment with MOLF-H resulted in increased *Lactobacillus* levels (105). Research studies show that supplementation with *M. oleifera* leads

to elevated *Lactobacillus* levels when obesity occurs through high-fat diet intake (104). Recent research exploring the interplay between gut microbiota and host immune responses has revealed that members of the Muribaculaceae family, a dominant component of the murine gut microbiota, can modulate host immunity. Natural killer (NK) cell activity is influenced by sucrose, while the nuclear factor-kappa B-alpha (NF- $\kappa$ B) signaling pathways experience impairment because of its presence (106). Treatment with MOLF induces changes in the gut microbiota composition, specifically increasing the abundance of families such as Muribaculaceae (107). These bacterial families have been identified to support greater activity of NK cells. The innate immune system relies on NK cells to perform recognition and elimination of abnormal infected cells (108). The pathogenic bacterium *Helicobacter* exists as an established cause of different gastric abnormalities. High *Helicobacter* counts in the body tend to worsen the outcomes of IBD (109). Research using mice established that *M. oleifera* polysaccharide administration minimized *Helicobacter* growth levels, while DSS treatment usually increases *Helicobacter* levels (105, 107).



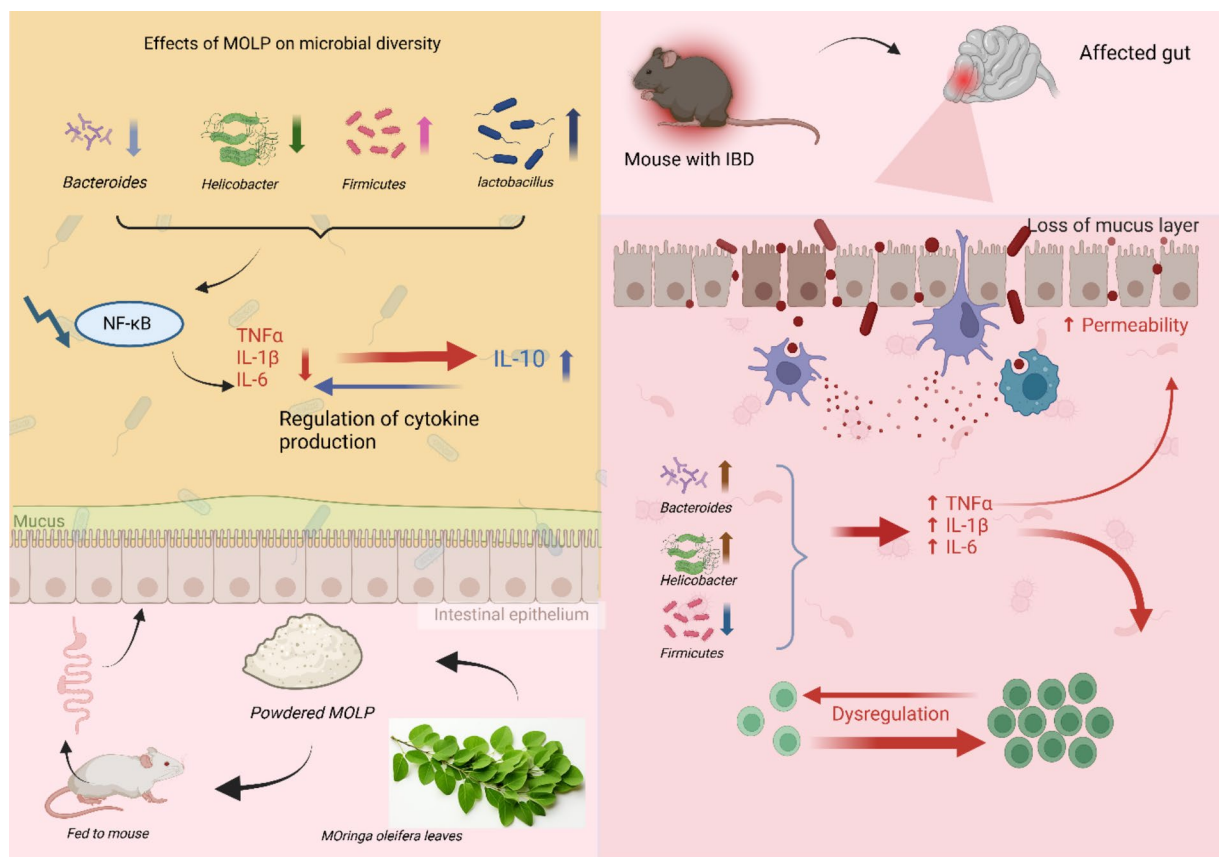


FIGURE 3

Explain the mechanism by which *M. oleifera* polysaccharides (MOLP) mitigate inflammatory bowel disease (IBD) involves modulation of gut microbiota composition and subsequent dampening of pro-inflammatory signaling. In IBD, dysbiosis is characterized by an increased abundance of pro-inflammatory bacteria (e.g., *Bacteroidetes* and *Helicobacter*) and a reduction in anti-inflammatory taxa (e.g., *Firmicutes*) contributes to mucosal barrier disruption and increased intestinal permeability. Oral administration of powdered *M. oleifera* leaves, rich in MOLP, appears to shift the gut microbial profile towards a more favorable composition, evidenced by increased beneficial genera such as *Lactobacillus* and the phylum *Firmicutes*. This modulation of the gut microbiota is associated with the downregulation of NF- $\kappa$ B signaling pathway and a consequent regulation of cytokine production, leading to a reduction in intestinal inflammation.

## Anti-inflammatory pathways

The natural protective mechanism against stimuli is inflammation (110). The initial inflammatory response leads to acute inflammation, yet chronic inflammation occurs when the response endures for multiple weeks up to several years (111). Inflammation is essential for tissue regeneration and repair, necessitating optimal activation of both the innate and adaptive immune systems to mount an effective response to injury (112). Activated macrophages, key players in the inflammatory response, release a suite of pro-inflammatory cytokines, including tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 beta (IL-1 $\beta$ ), interferon- $\gamma$  (IFN- $\gamma$ ), and interleukin-6 (IL-6) (113). These cells also generate reactive oxygen and nitrogen species, such as nitric oxide (NO), synthesized by inducible nitric oxide synthase (iNOS), contributing to the oxidative stress environment characteristic of inflammation (114).

Anti-inflammation, a process involving active suppression of pro-inflammatory signaling and restoration of tissue homeostasis, is crucial for limiting immunopathology following pathogen clearance or sterile injury. Chronic inflammation and subsequent tissue damage can be the result of failure to adequately control the inflammation cascade

(115). The production of immunomodulatory molecules, like IL-10, and the regulation of specific signaling pathways to suppress the activity of pro-inflammatory responses can resolve the problem of inflammation, culminating in the repair of tissue homeostasis and preservation of immune equilibrium (116). This complex regulatory mechanism is crucial for maintaining physiological health (117). *M. oleifera* delivers anti-inflammatory benefits through its mix of health-enhancing chemical substances, including isothiocyanates, flavonoids, and phenolic acids. The anti-inflammatory effects of *M. oleifera* compounds are believed to be mediated through multiple mechanisms (93).

## Inhibition of pro-inflammatory enzymes

iNOS is an enzyme expressed in various immune cells, such as macrophages, in response to pro-inflammatory stimuli like lipopolysaccharide (LPS) and cytokines (118). Upon induction, iNOS catalyzes the conversion of L-arginine to NO. In contrast to the constitutive isoforms, endothelial NOS (eNOS) and neuronal NOS (nNOS), iNOS produces substantial quantities of NO over prolonged periods. Excessive NO production, especially in the presence of



superoxide, can lead to the formation of peroxynitrite, a highly reactive species capable of damaging proteins, lipids, and DNA, thus contributing to the exacerbation of inflammation and tissue injury (119). The isothiocyanates, flavonoids, and phenolic acids present in *M. oleifera* can attenuate iNOS expression, thereby modulating NO production. This downregulation is often mediated through the suppression of upstream signaling pathways, particularly the NF- $\kappa$ B pathway (74). These bioactive compounds can stabilize the NF- $\kappa$ B inhibitor, I $\kappa$ B, preventing NF- $\kappa$ B translocation to the nucleus. Consequently, the transcriptional activity of the iNOS gene is diminished, resulting in reduced iNOS protein levels and a subsequent decrease in NO synthesis.

Similarly, Cyclooxygenase-2 (COX-2), an inducible enzyme, catalyzes the conversion of arachidonic acid to prostaglandins, such as prostaglandin E2 (PGE2) (120). These prostaglandins are pivotal inflammatory mediators implicated in the pathogenesis of pain, fever, and edema. In contrast to COX-1, which is constitutively expressed, COX-2 expression is markedly upregulated in response to inflammatory stimuli, positioning it as a key driver of inflammatory processes (121). The bioactive constituents of *M. oleifera* attenuates COX-2 expression, primarily through suppression of the NF- $\kappa$ B signaling pathway. Inhibition of NF- $\kappa$ B diminishes the transcriptional activity of the COX-2 gene, resulting in decreased COX-2 protein levels and a subsequent reduction in the synthesis of pro-inflammatory prostaglandins (93, 122). Furthermore, certain flavonoids present in *M. oleifera* may also exert a direct inhibitory effect on COX-2 enzyme activity.

## Regulation of cytokine production

Cytokines are a diverse group of signaling molecules that play a crucial role in the complex process of inflammation (123). Following tissue injury or pathogen invasion, immune cells, including macrophages, dendritic cells, and T lymphocytes, rapidly synthesize and secrete a variety of cytokines to initiate and regulate the inflammatory cascade (124). Pro-inflammatory cytokines, including TNF- $\alpha$ , IL-1 $\beta$ , and IL-6, are rapidly released at the site of tissue damage or infection (125). These cytokines initiate a cascade of events, activating resident cells and recruiting additional immune cells, such as neutrophils and monocytes, to the affected area, thereby amplifying and propagating the inflammatory response. It is very important to maintain a delicate equilibrium between pro-inflammatory and anti-inflammatory cytokines and prostaglandins. This equilibrium is essential for the development of targeted therapeutic strategies aimed at mitigating the detrimental effects of chronic inflammation and sepsis (126).

Administration of *M. oleifera* bio actives, such as *M. oleifera* isothiocyanate-1 (MIC-1) or MOLP, have demonstrated a significant decrease in tissue concentration and serum of pro-inflammatory cytokines, including TNF- $\alpha$ , IL-6, and IL-1 $\beta$ , in experimental models of acute inflammation or sepsis such as LPS-induced sepsis in mice (127). Moreover, along with suppression of pro-inflammatory signaling, certain *M. oleifera* extracts have shown the capacity to enhance the production of anti-inflammatory cytokines, such as IL-10, in animal models. This increase in IL-10 contributes to counterbalancing the inflammatory cascade and promoting resolution. For example, in a murine model of DSS-induced colitis,

administration of MOLP not only reduced colonic levels of TNF- $\alpha$  and IL-1 $\beta$  but also concurrently elevated IL-10 expression, thereby facilitating the resolution of inflammation (78).

## Mechanism of action as an antioxidant

Oxidative stress occurs when the generation of reactive oxygen species (ROS) overwhelms the body's endogenous antioxidant defense mechanisms, leading to an imbalance that favors ROS accumulation and subsequent cellular damage (128). This imbalance can arise from either increased ROS production, decreased antioxidant capacity, or a combination of both (129). During oxidative phosphorylation, the process by which ATP is generated in mitochondria, electrons can escape from the electron transport chain, resulting in the formation of superoxide radicals ( $O_2^-$ ) (130). These superoxide radicals can subsequently be converted into other ROS, including hydrogen peroxide ( $H_2O_2$ ) and highly reactive hydroxyl radicals ( $OH\cdot$ ). While ROS plays essential physiological roles in cellular signaling and host defense, excessive ROS production overwhelms endogenous antioxidant systems, leading to a state of oxidative stress (131, 132). Activated immune cells, such as neutrophils and macrophages, generate ROS as a crucial component of the innate immune response against invading pathogens. While this ROS production is a physiological process essential for microbial killing, chronic inflammation can result in sustained and excessive ROS generation, which contributes to oxidative damage of host tissues and the progression of various diseases. Excessive production of ROS can lead to significant oxidative damage to critical cellular components, including lipids, proteins, and DNA (133). Such damage can impair cellular function, trigger cell death pathways (apoptosis or necrosis), and contribute to the aging process and the development of a wide range of pathological conditions (134). Oxidative stress can negatively impact immune function, rendering animals more susceptible to infections and disease (135). The mechanism of oxidative damage through phosphorylation is shown in Figure 4. In ruminants, for instance, oxidative stress has been linked to compromised immune responses, particularly during periods of physiological stress, such as the periparturient period or heat stress (136).

*M. oleifera* leaves are recognized for their significant antioxidant capacity, a property attributed to their rich composition of bioactive compounds (137, 138). These antioxidants play a critical role in scavenging free radicals and mitigating oxidative stress within biological systems (139). The abundance of bioactive compounds, including flavonoids, phenolic acids, and vitamin C, in *M. oleifera* leaves contributes significantly to their ability to mitigate oxidative damage and inflammation (140). Specific antioxidant constituents, such as quercetin, chlorogenic acid, and beta-carotene, present in *M. oleifera* contribute significantly to its protective effects against oxidative damage and inflammation (141, 142). *M. oleifera* has a series of events in its mechanism as an antioxidant, which are shown in Figure 4. These are scavenging free radicals, increasing endogenous antioxidant defenses, reducing lipid peroxidation, and modulating cellular signaling pathways that are related to oxidative stress (79). The 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay is a widely used method to assess the activity of an antioxidant. Antioxidants reacted with that free radical DPPH by donating either an electron or a hydrogen atom (143). After the reaction, DPPH reduces to a non-radical form,

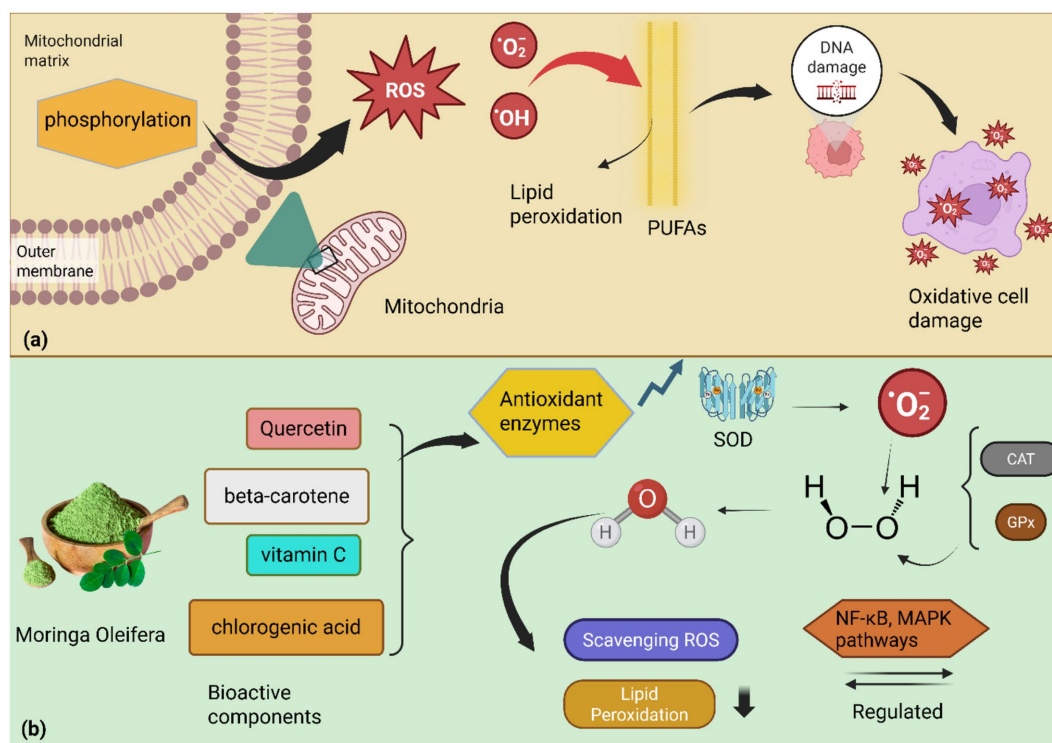


FIGURE 4

(a) Oxidative stress mechanism: mitochondrial oxidative phosphorylation generates superoxide radicals ( $O_2^{\bullet-}$ ) due to electron leakage from the electron transport chain. These radicals further transform into reactive oxygen species (ROS), including hydrogen peroxide ( $H_2O_2$ ) and highly reactive hydroxyl radicals ( $OH^\bullet$ ). When ROS production overwhelms cellular antioxidant defenses, it leads to an imbalance that attacks polyunsaturated fatty acids in cell membranes, initiating lipid peroxidation. Byproducts of lipid peroxidation subsequently cause DNA damage and overall oxidative cell damage. (b) *M. oleifera*'s antioxidant action: *M. oleifera* combats oxidative stress through its bioactive components, including quercetin, beta-carotene, vitamin C, and chlorogenic acid, which provide potent antioxidant protection. These antioxidants directly scavenge ROS and neutralize free radicals like superoxide ( $O_2^{\bullet-}$ ). *M. oleifera* also enhances the activity of key antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx), enhancing cellular defense against oxidative damage. By scavenging ROS and boosting antioxidant enzyme activities, *M. oleifera* reduces lipid damage in membranes. Furthermore, *M. oleifera* modulates cellular signaling pathways, including NF- $\kappa$ B and MAPK, which regulate inflammatory responses triggered by oxidative stress, thereby ensuring cell health through its rigorous antioxidant properties.

$\alpha,\alpha$ -diphenyl- $\beta$ -picryl hydrazine, that results in loss of its purple color (144). Studies have shown that extracts of *M. oleifera* have a significant effect in reducing DPPH free radicals (145, 146). The radical scavenging and the antioxidant properties of different extracts of *M. oleifera* leaves from various agro-climatic regions were examined by Siddhuraju and Becker (147). The results found that the aqueous and aqueous ethanol extracts of freeze-dried leaves of *M. oleifera* inhibit 89.7–92.0% of peroxidation of linoleic acid and possess scavenging activities on superoxide radicals in the  $\beta$ -carotene-linoleic acid system. *M. oleifera* enhance the activity of endogenous antioxidant enzymes, in which glutathione peroxidase (GPx), catalase (CAT), and superoxide dismutase (SOD). This enhancement of these enzymes boosts the immune function against oxidative stress (148).

Free radicals interact with lipids in cell membranes, initiating a chain reaction of lipid peroxidation that can lead to cell damage and contribute to various diseases impacting the immune system (149, 150). Lipid peroxidation is recognized as a well-established biomarker of oxidative stress (151). This process primarily affects polyunsaturated fatty acids (PUFAs) such as linoleic acid, linolenic acid, and arachidonic acid, which are essential components of cell membranes (150, 152). The byproducts of this reaction include malondialdehyde

(MDA) and 4-hydroxy-2-hexenal (153). These compounds possess mutagenic, cytotoxic, and neurotoxic properties, allowing them to alter DNA, damage cells, and harm nerve tissues (154).

*M. oleifera* has proven effective in inhibiting lipid peroxidation by lowering MDA levels. Research indicates that the percentage of lipid peroxidation observed in *M. oleifera* leaves and stems was 85.88 and 77.63%, respectively (155). In a study involving Swiss albino mice experiencing oxidative stress, pre-treatment with *M. oleifera* leaf extract successfully restored glutathione (GSH) levels, effectively reducing lipid peroxidation (156, 157). Similarly, a daily administration of *M. oleifera* extract for 60 days to rats with carbon tetrachloride (CCl<sub>4</sub>)-induced hepatic lipid peroxidation demonstrated a reduction in hepatotoxicity, attributed to phenolic compounds and flavonoids, including  $\beta$ -sitosterol, quercetin, and kaempferol found in the extract (158). In broiler chickens, supplementing *M. oleifera* leaf meal up to 5% of dry matter intake has shown improvements in fatty acid profiles and a reduction in lipid peroxidation (159). Recent studies on Nile tilapia (*Oreochromis niloticus*) have indicated that *M. oleifera* leaf extracts enhance feed utilization and growth while also improving the innate immune response, evidenced by increased lysosome levels and phagocytic activity (160, 161). A recent study involving crayfish

(*Procambarus clarkii*) found that incorporating 1% of fermented *M. oleifera* leaves into the diet significantly improved growth performance and antioxidant capacity (162). Overall, due to its rich phenolic content, *M. oleifera* exhibits strong antioxidant properties.

## Future directions

Multiple essential investigations need completion before *M. oleifera* can achieve its complete potential as an environmentally friendly animal feed source for nutrition and health benefits. Research needs to focus on developing better processing methods that will boost the bioactive compound availability in *M. oleifera*. Scientific research is necessary to develop drying techniques and fermentation processes with enzymatic treatments, which will make *M. oleifera* nutrients more accessible after processing. Research needs to progress further to create well-balanced animal feed compositions that effectively integrate *M. oleifera* into animal dietary plans. There is a need to evaluate *M. oleifera* connection with dietary elements to discover optimal combinations that maximize nutrient uptake and promote animal development with improved wellness.

The ongoing research must focus on extended investigations to determine how *M. oleifera* supplementation affects livestock throughout multiple periods. More research about *M. oleifera* long-term influence on livestock performance must investigate its effects on reproductive outcomes and disease resistance, and animal welfare, regardless of demonstrated short-term enhancements in growth and feed conversion. The inconsistent bioactive component levels in *M. oleifera* require scientists to investigate the effects of growing conditions, together with the differences observed across cultivars. Extraction of *M. oleifera* cultivars alongside optimal cultivation methods that maximize beneficial compound concentrations will optimize the effectiveness of *M. oleifera* as a feed resource.

Research must determine the molecular processes that explain *M. oleifera* ability to modulate immune functions. The evaluation of *M. oleifera* bioactive compound-substance interactions with immune pathways and antioxidant systems, and gut microbiota needs comprehensive research to determine its potential as a natural animal feed immunostimulant. Research into *M. oleifera* biological mechanisms will optimize its value as a health promoter for livestock during times of stress.

Environmental assessment methods and economic modeling practices need to be implemented to determine the complete advantages *M. oleifera* has over conventional feed crops. Studies using life-cycle assessments need to evaluate the environmental effects of *M. oleifera* farming in addition to its capacity to minimize greenhouse gas emissions, water usage, and land degradation relative to maize and soybean as conventional feed crops. The cost-effectiveness assessment of *M. oleifera* in extended-scale livestock production requires economic studies to determine expense-to-worth ratios and the resulting lower feed costs while enhancing livestock health.

The rising acceptance of *M. oleifera* as an essential feed ingredient needs proper regulations to guarantee its secure usage within the livestock industry. Research about *M. oleifera* safety profiles alongside compliance with animal feed regulations will enable its integration into international commercial feeding systems. Driving market demand for sustainable livestock production with *M. oleifera* will

be facilitated by educating consumers about its advantages. The successful adoption of *M. oleifera* requires research-based partnerships between researchers and both agricultural stakeholders and policymakers who will develop strategies for global livestock adoption.

*M. oleifera* shows extensive value because it represents a sustainable source of nourishing feed material for agricultural purposes. Research efforts into *M. oleifera* must advance through optimization of processing techniques, along with formulation research and studies of molecular effects and environmental and economic assessments. The implementation of these tactics ensures *M. oleifera* significantly contributes to solving current international issues regarding food supply stability and livestock health as well as environmental preservation.

## Conclusion

*M. oleifera* is increasingly recognized as a promising solution for sustainable livestock production due to its rich nutritional content and therapeutic properties. Packed with protein, essential amino acids, vitamins, and minerals, it enhances animal growth and overall health. Its immunomodulatory effects help regulate gut immunity and reduce oxidative stress, boosting disease resistance in livestock. Additionally, *M. oleifera* serves as a viable alternative to traditional feed sources like soybeans and maize, addressing concerns of feed supply, environmental impact, and price stability. Cultivating *M. oleifera* in arid regions can also contribute to food security in developing countries. While its potential is significant, the effective use of *M. oleifera* requires further research on diet optimization and its long-term effects on livestock performance. Overall, *M. oleifera* offers substantial benefits for enhancing livestock nutrition and promoting sustainable agricultural practices. *M. oleifera* triple-action benefits—nutritional (complete EAA profile), environmental (3.5 × lower carbon footprint than soybean), and therapeutic (40–60% cytokine reduction)—position it as a transformative feed additive. Critical findings include:

- 1 Optimal inclusion: 15% for ruminants (↑ milk yield 12%), 5% for poultry (↑ weight gain 8%).
- 2 Processing protocols: Freeze-drying retains 92% flavonoids vs. 67% in sun-drying.
- 3 Economic viability: 0.18/kg production cost vs. 0.31/kg for soybean meal.

Priority research areas:

- Long-term toxicity (>6 months consumption).
- Breed-specific formulation optimization.
- Policy frameworks for smallholder adoption.

## Author contributions

RM: Software, Methodology, Writing – original draft, Data curation, Investigation, Conceptualization. SE: Writing – review & editing. MZ: Writing – review & editing. ZC: Writing – review & editing. AS: Writing – review & editing. HH: Writing – review &



editing, Supervision. MW: Funding acquisition, Project administration, Supervision, Writing – review & editing.

## Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was supported by the National 14th Five-Year Plan Key Research and Development Program (2024YFD1300204, 2023YFD1301705), high-end foreign experts from the Ministry of Science and Technology of China (G2023014066L), and the Priority Academic Program Development of Jiangsu Higher Education Institution (PAPD), P.R. China.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## References

- World Food. World food and agriculture – statistical yearbook 2023. Rome. (2023).
- Yang C, Li Q, Wang X, Cui A, Chen J, Liu H, et al. Human expansion-induced biodiversity crisis over Asia from 2000 to 2020. *Research*. (2023) 6:14–7. doi: 10.34133/research.0226
- Makkar H. Review: feed demand landscape and implications of food-not feed strategy for food security and climate change. *Animal*. (2017) 12:1744–54. doi: 10.1017/S175173111700324X
- Yitbarek MB. Livestock and livestock product trends by 2050. *International Journal of Animal Research (IJAR)* (2019). 4:30.
- Erdaw M. Contribution, prospects and trends of livestock production in sub-Saharan Africa: a review. *Int J Agric Sustain*. (2023) 21:2247776. doi: 10.1080/14735903.2023.2247776
- Bai Z, Wenqi L, Velthof G, Wei Z, Havlik P, Oenema O, et al. China's livestock transition: driving forces, impacts, and consequences. *Sci Adv*. (2018) 4:eaar8534. doi: 10.1126/sciadv.aar8534
- Rojas-Downing M, Nejadhashemi A, Harrigan T, Woznicki S. Climate change and livestock: impacts, adaptation, and mitigation. *Clim Risk Manag*. (2017) 16:145–63. doi: 10.1016/J.CRM.2017.02.001
- Su B, Chen X. Current status and potential of *Moringa oleifera* leaf as an alternative protein source for animal feeds. *Front Vet Sci*. (2020) 7:53. doi: 10.3389/fvets.2020.00053
- Parrini S, Aquilani C, Pugliese C, Bozzi R, Sirtori F. Soybean replacement by alternative protein sources in pig nutrition and its effect on meat quality. *Animals*. (2023) 13:494. doi: 10.3390/ani13030494
- Boerema A, Peeters A, Swolfs S, Vandevenne F, Jacobs S, Staes J, et al. Soybean trade: balancing environmental and socio-economic impacts of an intercontinental market. *PLoS One*. (2016) 11:e0155222. doi: 10.1371/journal.pone.0155222
- Khan NM, Qadeer A, Khan A, Nasir A, Sikandar A, Adil M, et al. Alternative sources of proteins in FARM animal feeding. *J Microbiol Biotechnol Food Sci*. (2024) 13:e10605. doi: 10.55251/jmbfs.10605
- Dronne Y. Agricultural raw materials for food and feed: the world. *INRA Prod Anim*. (2018) 31:165–80. doi: 10.20870/PRODUCTIONS-ANIMALES.2018.31.3.2345
- Uwizeye A, de I, Opio C, Schulte R, Falcucci A, Tempio G, et al. Nitrogen emissions along global livestock supply chains. *Nat Food*. (2020) 1:437–46. doi: 10.1038/s43016-020-0113-y
- Wang J, Liu Q, Hou Y, Qin W, Lesschen J, Zhang F, et al. International trade of animal feed: its relationships with livestock density and N and P balances at country level. *Nutr Cycl Agroecosyst*. (2017) 110:197–211. doi: 10.1007/s10705-017-9885-3
- Tejeda H, Goodwin B. Price volatility, nonlinearity, and asymmetric adjustments in corn, soybean, and cattle markets: implications of ethanol-driven (market) shocks. (2009). Available online at: <https://consensus.app/papers/price-volatility-nonlinearity-and-asymmetric-tejeda-goodwin/32015c95e5915f5be8537186d29158186/> (Accessed December 5, 2024).
- Minh CLT, Lebaillay P, Nguyen T. Cost, return analysis and constraints in livestock production and marketing in Hai Duong, Vietnam. (2013) 1194–1199. Available online at: <https://consensus.app/papers/cost-return-analysis-and-constraints-in-livestock-minh-lebaillay/c68848f9a21c52fba73135ce9a78848b/> (Accessed December 5, 2024).
- Halmemies-Beauchet-Filleau A, Rinne M, Lamminen M, Mapato C, Ampapon T, Wanapat M, et al. Review: alternative and novel feeds for ruminants: nutritive value, product quality and environmental aspects. *Animal*. (2018) 12:s295–309. doi: 10.1017/S1751731118002252
- Gasco L, Biasato I, Dabbou S, Schiavone A, Gai F. Animals fed insect-based diets: state-of-the-art on digestibility, performance and product quality. *Animals*. (2019) 9:170. doi: 10.3390/ani9040170
- Bryant JP, Joly K, Chapin FS, DeAngelis DL, Kielland K. Can antibrowsing defense regulate the spread of woody vegetation in arctic tundra? *Ecography Cop*. (2014) 37:204–11. doi: 10.1111/j.1600-0587.2013.00436.x
- Rizwan N, Rizwan D, Banday M. *Moringa oleifera*: the miracle tree and its potential as non-conventional animal feed: a review. *Agric Rev*. (2022) 45:369–379. doi: 10.18805/ag.r-2405
- Sarwatt S, Kapange S, Kakengi A. Substituting sunflower seed-cake with *Moringa oleifera* leaves as a supplemental goat feed in Tanzania. *Agrofor Syst*. (2002) 56:241–7. doi: 10.1023/A:1021396629613
- Sonkar N, Singh N, Santra A, Mishra S, Verma L, Soni A. Application of munga (*Moringa oleifera*) in livestock feed: a review. *Int J Chem Stud*. (2020) 8:1729–35. doi: 10.22271/chemi.2020.v8.i1y.8513
- Nouman W, Anwar F, Gull T, Newton A, Rosa E, Domínguez-Perles R. Profiling of polyphenolics, nutrients and antioxidant potential of germplasm's leaves from seven cultivars of *Moringa oleifera* lam. *Ind Crop Prod*. (2016) 83:166–76. doi: 10.1016/J.INDCROP.2015.12.032
- Moyo B, Oyedemi S, Masika P, Muchenje V. Polyphenolic content and antioxidant properties of *Moringa oleifera* leaf extracts and enzymatic activity of liver from goats supplemented with *Moringa oleifera* leaves/sunflower seed cake. *Meat Sci*. (2012) 91:441–7. doi: 10.1016/j.meatsci.2012.02.029
- Tumer T, Rojas-Silva P, Poulev A, Raskin I, Waterman C. Direct and indirect antioxidant activity of polyphenol- and isothiocyanate-enriched fractions from *Moringa oleifera*. *J Agric Food Chem*. (2015) 63:1505–13. doi: 10.1021/jf505014n
- Falowo A, Mukumbo F, Idamokoro E, Lorenzo J, Afolayan A, Muchenje V. Multi-functional application of *Moringa oleifera* lam. In nutrition and animal food products: a review. *Food Res Int*. (2018) 106:317–34. doi: 10.1016/j.foodres.2017.12.079
- Teixeira E, Carvalho M, Neves V, Silva M, Arantes-Pereira L. Chemical characteristics and fractionation of proteins from *Moringa oleifera* lam. Leaves. *Food Chem*. (2014) 147:51–4. doi: 10.1016/j.foodchem.2013.09.135
- Leitanthem VK, Chaudhary P, Maiti S, Mohini M, Mondal G. Impact of *Moringa oleifera* leaves on nutrient utilization, enteric methane emissions, and performance of goat kids. *Animals*. (2023) 13:1–15. doi: 10.3390/ani13010097

## Generative AI statement

The authors declare that no Gen AI was used in the creation of this manuscript.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## Supplementary material

The Supplementary material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fnut.2025.1615349/full#supplementary-material>



29. Boopathi NM, Abubakar BY. Botanical Descriptions of *Moringa* spp., In: Boopathi, N.M., Raveendran, M., Kole, C. (eds). *The Moringa Genome. Compendium of Plant Genomes*. Springer, Cham (2021).
30. Tshabalala T, Ncube B, Madala N, Nyakudya T, Moyo H, Sibanda M, et al. Scribbling the cat: A case of the “miracle” plant, *Moringa oleifera*. *Plan Theory*. (2019) 8:510. doi: 10.3390/plants8110510
31. Dhiman J. A review on medicinal uses of *Moringa oleifera*. *J Drug Deliv Ther*. (2023) 13:197–201. doi: 10.22270/jddt.v13i11.6042
32. Alshoaibi A. Seed germination, seedling growth and photosynthetic responses to temperature in the tropical tree *Moringa oleifera* and its relative desert, *Moringa peregrina*. *Egypt J Bot*. (2021) 61:541–51. doi: 10.21608/EJBO.2021.63271.1631
33. Rajalakshmi R, Rajalakshmi S, Parida A. Evaluation of the genetic diversity and population structure in drumstick (*Moringa oleifera* L.) using SSR markers. *Curr Sci*. (2017) 112:1250–6. doi: 10.18520/CS/V112/I06/1250-1256
34. Anzano A, Ammar M, Papaiani M, Grauso L, Sabbah M, Capparelli R, et al. *Moringa oleifera* lam.: a phytochemical and pharmacological overview. *Horticulturae*. (2021) 7:409. doi: 10.3390/horticulturae7100409
35. Farooq F, Rai M, Tiwari A, Khan A, Farooq S. Medicinal properties of *Moringa oleifera*: an overview of promising healer. *J Med Plants Res*. (2012) 6:4368–74. doi: 10.5897/JMPR12.279
36. Dania SO, Akpansubi P, Eghagara OO. Comparative effects of different fertilizer sources on the growth and nutrient content of *Moringa (Moringa oleifera)* seedling in a greenhouse trial. *Adv Agric*. (2014) 2014:726313. doi: 10.1155/2014/726313
37. Mashamaite CV, Ramatsitsi MN, Manyevere A. *Moringa oleifera* lam.: a versatile climate-smart plant for nutritional security and therapeutic usage in semi-arid regions. *J Agric Food Res*. (2024) 16:101217. doi: 10.1016/j.jafr.2024.101217
38. Santos A, Silva-Mann R, Ferreira R, Brito A. Water pre-hydration as priming for *Moringa oleifera* lam. Seeds under salt stress. *Trop Subtrop Agroecosyst* (2011) 14 201–207. Available online at: <https://consensus.app/papers/water-prehydration-as-priming-for-moringa-oleifera-lam-santos-silva-mann/9c085dead6cd57f9bf4a2aec797e6ca6/> (Accessed January 2, 2024).
39. Kholif A, Morsy T, Gouda G, Anale U, Galyean M. Effect of feeding diets with processed *Moringa oleifera* meal as protein source in lactating Anglo-Nubian goats. *Anim Feed Sci Technol*. (2016) 217:45–55. doi: 10.1016/j.ANIFEEDSCI.2016.04.012
40. Sánchez NR, Ledin S, Ledin I. Biomass production and chemical composition of *Moringa oleifera* under different management regimes in Nicaragua. *Agrofor Syst*. (2006) 66:231–42. doi: 10.1007/s10457-005-8847-y
41. Oduro I, Ellis WO, Owusu D. Nutritional potential of two leafy vegetables: *Moringa oleifera* and *Ipomoea batatas* leaves. *Sci Res Essays*. (2008) 3:057–60. doi: 10.5897/SRE.9000686
42. El-Hack M, Alagawany M, Elrys A, Desoky E, Tolba H, Elnahal A, et al. Effect of forage *Moringa oleifera* L. (moringa) on animal health and nutrition and its beneficial applications in soil, plants and water purification. *Agriculture*. (2018) 8:145. doi: 10.3390/AGRICULTURE8090145
43. Knudsen K. The nutritional significance of “dietary fibre” analysis. *Anim Feed Sci Technol*. (2001) 90:3–20. doi: 10.1016/S0377-8401(01)00193-6
44. Lesten ECC, Emmanuel CM. Proximate, physical and chemical composition of leaves and seeds of *Moringa (Moringa oleifera)* from Central Malawi: a potential for increasing animal food supply in the 21st century. *Afr J Agric Res*. (2018) 13:2872–80. doi: 10.5897/ajar2018.13535
45. Ahmed M, Marrez DA, Abdelmoeen NM, Mahmoud EA, Abdel-Shakur Ali M, Decsi K, et al. Proximate analysis of *Moringa oleifera* leaves and the antimicrobial activities of successive leaf Ethanolic and aqueous extracts compared with green chemically synthesized ag-NPs and crude aqueous extract against some pathogens. *Int J Mol Sci*. (2023) 24:3529. doi: 10.3390/ijms24043529
46. Kumar V, Rani A, Chauhan GS. Nutritional value of soybean. In: *The soybean: botany, production and uses*. (ed.) Guriqbal S. Wallingford UK: CABI (2010). 375–403.
47. Masitha EP, Seifu E, Teketay D. Nutritional composition and mineral profile of leaves of *Moringa oleifera* provenances grown in Gaborone, Botswana. *Food Prod Process Nutr*. (2024) 6:3. doi: 10.1186/s43014-023-00183-8
48. Laura R. Nutritional and mineral composition of leaves, roots and seeds of *Moringa oleifera* lam. Tree from Tenerife, Spain. *J Soc Trop Plant Res*. (2021) 8:1–5. doi: 10.22271/tpr.2021.v8.i1.001
49. Sodamade A, Bolaji OS, Adeboye OO. Proximate analysis, mineral contents and functional properties of *Moringa oleifera* leaf protein concentrate. *IOSR J Appl Chem*. (2013) 4:47–51. doi: 10.9790/5736-0464751
50. Waterman C, Rojas-Silva P, Tumer TB, Kuhn P, Richard AJ, Wicks S, et al. Isothiocyanate-rich *Moringa oleifera* extract reduces weight gain, insulin resistance, and hepatic gluconeogenesis in mice. *Mol Nutr Food Res*. (2015) 59:1013–24. doi: 10.1002/mnfr.201400679
51. Sánchez-Machado D, Núñez-Gastélum J, Reyes-Moreno C, Ramírez-Wong B, López-Cervantes J. Nutritional quality of edible parts of *Moringa oleifera*. *Food Anal Methods*. (2010) 3:175–80. doi: 10.1007/S12161-009-9106-Z
52. Wu G, Bazer F, Dai Z, Li D, Wang J, Wu Z. Amino acid nutrition in animals: protein synthesis and beyond. *Annu Rev Anim Biosci*. (2014) 2:387–417. doi: 10.1146/annurev-animal-022513-114113
53. Sefer M, Petronijevic RB, Trbovic D, Ciric J, Baltic T, Parunovic N, et al. Amino acids in animal feed: significance and determination techniques. *IOP Conf Ser Earth Environ Sci*. (2021) 854:012082. doi: 10.1088/1755-1315/854/1/012082
54. Moyo B, Masika PJ, Hugo A, Muchenje V. Nutritional characterization of *Moringa (Moringa oleifera* lam.) leaves. *Afr J Biotechnol*. (2011) 10:12925–33. doi: 10.5897/ajb10.1599
55. Kong C, Adeola O. Evaluation of amino acid and energy utilization in feedstuff for swine and poultry diets. *Asian Australas J Anim Sci*. (2014) 27:917–25. doi: 10.5713/ajas.2014.r.02
56. Abbas R, Elsharbas F, Fadlelmula AA. Nutritional values of *Moringa oleifera*, total protein, amino acid, vitamins, minerals, carbohydrates, total fat and crude fiber, under the semi-arid conditions of Sudan. *J Microb Biochem Technol*. (2018) 10:56–8. doi: 10.4172/1948-5948.1000396
57. Aderinola TA, Fagbemi TN, Enujiugha VN, Alashi AM, Aluko RE. Amino acid composition and antioxidant properties of *Moringa oleifera* seed protein isolate and enzymatic hydrolysates. *Heliyon*. (2018) 4:e00877. doi: 10.1016/j.heliyon.2018.e00877
58. Park BC. Amino acid imbalance-biochemical mechanism and nutritional aspects. *Asian Australas J Anim Sci*. (2006) 19:1361–8. doi: 10.5713/ajas.2006.1361
59. Abe M, Iriki T, Funaba M, Onda S. Limiting amino acids for a corn and soybean meal diet in weaned calves less than three months of age. *J Anim Sci*. (1998) 76:628–36. doi: 10.2527/1998.762628x
60. Monte Singer W, Zhang B, Rouf Mian MA, Huang H. Soybean amino acids in health, genetics, and evaluation. *Soybean Hum Consum Anim Feed*. (2020) 19:1361–1368. doi: 10.5772/intechopen.89497
61. Bunchasak C. Role of dietary methionine in poultry production. *J Poult Sci*. (2009) 46:169–79. doi: 10.2141/jpsa.46.169
62. Čolović M, Vasić V, Djurić D, Krstić D. Sulphur-containing amino acids: protective role against free radicals and heavy metals. *Curr Med Chem*. (2018) 25:324–35. doi: 10.2174/0929867324666170609075434
63. Valdez-Solana M, Mejía-García V, Téllez-Valencia A, García-Arenas G, Salas-Pacheco J, Alba-Romero J, et al. Nutritional content and elemental and phytochemical analyses of *Moringa oleifera* grown in Mexico. *J Chem*. (2015) 20:1–9. doi: 10.1155/2015/860381
64. Bootman M. Calcium signaling. *Current Medicinal Chemistry*. (2018) 25:1–12.
65. Luan S, Wang C. Calcium signaling mechanisms across kingdoms. *Annu Rev Cell Dev Biol*. (2021) 37:311–40. doi: 10.1146/annurev-cellbio-120219-035210
66. Allen J, Issa J, Cai W. Calcium content, *in vitro* digestibility, and bioaccessibility in leaves of spinach (*Spinacia oleracea*), sweet potato (*Ipomoea batatas*), and drumstick tree (*Moringa oleifera*). *F1000Res*. (2014) 3:65. doi: 10.12688/F1000RESEARCH.3287.1
67. Rajbhar Y, Rajbhar G, Rawat P, Shukla S, Kumar M. Grow *Moringa (Moringa oleifera)*, the miracle tree on the earth. *Hortic Int J*. (2018) 2:166–72. doi: 10.15406/hij.2018.02.00047
68. Razzaghi A, Vakili A, Khorrami B, Ghaffari M, Rico D. Effect of dietary supplementation or cessation of magnesium-based alkalizers on milk fat output in dairy cows under milk fat depression conditions. *J Dairy Sci*. (2022) 105:2275–87. doi: 10.3168/jds.2021-20457
69. Pinotti L, Manoni M, Ferrari L, Tretola M, Cazzola R, Givens I. The contribution of dietary magnesium in farm animals and human nutrition. *Nutrients*. (2021) 13:1–15. doi: 10.3390/nu13020509
70. Ferreira PPM, Farias DF, Oliveira JTDA, Carvalho ADF. *Moringa oleifera*: bioactive compounds and nutritional potential *Moringa oleifera*: compostos bioativos e potencialidade nutricional. *Rev Nutr*. (2008) 21:431–7. doi: 10.1590/S1415-52732008000400007
71. Lata M, Mondal BC. *Moringa oleifera* leaf meal: a sustainable approach for poultry production: a review. *Arch Curr Res Int*. (2024) 24:176–85. doi: 10.9734/acri/2024/v24i11959
72. Gopalakrishnan L, Doriya K, Kumar DS. *Moringa oleifera*: a review on nutritive importance and its medicinal application. *Food Sci Human Wellness*. (2016) 5:49–56. doi: 10.1016/j.fshw.2016.04.001
73. Leone A, Spada A, Battezzati A, Schiraldi A, Aristil J, Bertoli S. Cultivation, genetic, ethnopharmacology, phytochemistry and pharmacology of *Moringa oleifera* leaves: an overview. *Int J Mol Sci*. (2015) 16:12791–835. doi: 10.3390/ijms160612791
74. Kou X, Li B, Olayanju JB, Drake JM, Chen N. Nutraceutical or pharmacological potential of *Moringa oleifera* lam. *Nutrients*. (2018) 10:343. doi: 10.3390/nu10030343
75. Poluan JC, Zubair MS, Ramadan AP, Hayati F. Narrative review: potential of flavonoids from *Moringa (Moringa oleifera* lam.) leaves as immunomodulators. *Jurnal Farmasi Galenika*. (2023) 9:270–83. doi: 10.22487/j24428744.2023.v9.i2.16265
76. Nfambi J, Bbosa GS, Sembajwe LE, Gakunga J, Kasolo JN. Immunomodulatory activity of methanolic leaf extract of *Moringa oleifera* in Wistar albino rats. *J Basic Clin Physiol Pharmacol*. (2015) 26:603–11. doi: 10.1515/jbcp-2014-0104
77. Li L, Ma L, Wen Y, Xie J, Yan L, Ji A, et al. Crude polysaccharide extracted from *Moringa oleifera* leaves prevents obesity in association with modulating gut microbiota in high-fat diet-fed mice. *Front Nutr*. (2022) 9:1–17. doi: 10.3389/fnut.2022.861588
78. Mohamed Husien H, Peng WL, Su H, Zhou RG, Tao Y, Huang JJ, et al. *Moringa oleifera* leaf polysaccharide alleviates experimental colitis by inhibiting inflammation

- and maintaining intestinal barrier. *Front Nutr.* (2022) 9:1–14. doi: 10.3389/fnut.2022.1055791
79. Vergara-Jimenez M, Almatrafi MM, Fernandez ML. Bioactive components in *Moringa oleifera* leaves protect against chronic disease. *Antioxidants.* (2017) 6:91. doi: 10.3390/antiox6040091
80. Kusmiyati K, Rahmawati E, Waangsir FWF, Selasa P. Alkaloids, flavonoids, tannins and saponins contents in *Moringa oleifera* leaves. *Indones J Glob Health Res.* (2022) 4:139–44. doi: 10.37287/ijghr.v4i1.832
81. Goswami R, Arya D, Siddiqui R, Kumar S, Sarma O, Arora N. The multifaceted benefits and applications of *Moringa oleifera*: a comprehensive review. *Eur J Nutr Food Saf.* (2024) 16:1–13. doi: 10.9734/ejnf/2024/v16i81489
82. Eladia R, Ampode KM. Moringa (*Moringa oleifera* lam.) pod meal: nutrient analysis and its effect on the growth performance and cell-mediated immunity of broiler chickens. *J Anim Health Prod.* (2021) 9:170–7. doi: 10.17582/journal.jahp/2021/9.2.170.177
83. Dzuvoor CKO, Pan S, Amanze C, Amuzu P, Asakiya C, Kubi F. Bioactive components from *Moringa oleifera* seeds: production, functionalities and applications—a critical review. *Crit Rev Biotechnol.* (2022) 42:271–93. doi: 10.1080/07388551.2021.1931804
84. Pop OL, Kerezi AD, Ciont C. A comprehensive review of *Moringa oleifera* bioactive compounds—cytotoxicity evaluation and their encapsulation. *Food Secur.* (2022) 11:1–18. doi: 10.3390/foods11233787
85. Mitsogianni M, Koutsidis G, Mavroudis N, Trafalis DT, Botaitis S, Franco R, et al. The role of isothiocyanates as cancer anti-melanoma agents. *Antioxidants.* (2019) 8:1–35. doi: 10.3390/antiox8040106
86. Habtemariam S. Anti-inflammatory therapeutic mechanisms of Isothiocyanates: insights from Sulforaphane. *Biomedicines.* (2024) 12:1169. doi: 10.3390/biomedicines12061169
87. Byrne KA, Loving CL, McGill JL. Innate immunomodulation in food animals: evidence for trained immunity? *Front Immunol.* (2020) 11:1099. doi: 10.3389/fimmu.2020.01099
88. Bobeck EA. Nutrition and health: companion animal applications: functional nutrition in livestock and companion animals to modulate the immune response. *J Anim Sci.* (2020) 98:1–8. doi: 10.1093/JAS/SKAA035
89. Kober H AKM, Rajoka MSR, Mehwish HM, Villena J, Kitazawa H. Immunomodulation potential of probiotics: a novel strategy for improving livestock health, immunity, and productivity. *Microorganisms.* (2022) 10:388. doi: 10.3390/microorganisms10020388
90. Batista LHC, Oliveira IM, Prados LF, Araújo LC, Ferreira IM, de Abreu MJ, et al. The strategic use of an immunomodulatory feed additive in supplements for grazing young Nellore bulls transported after weaning: performance, physiological, and stress parameters. *Agriculture.* (2023) 13:1027. doi: 10.3390/agriculture13051027
91. Pareek A, Pant M, Gupta MM, Kashania P, Ratan Y, Jain V, et al. *Moringa oleifera*: an updated comprehensive review of its pharmacological activities, Ethnomedicinal, Phytopharmaceutical formulation, clinical, phytochemical, and toxicological aspects. *Int J Mol Sci.* (2023) 24:2098. doi: 10.3390/ijms24032098
92. Xiao X, Wang J, Meng C, Liang W, Wang T, Zhou B, et al. *Moringa oleifera* lam and its therapeutic effects in immune disorders. *Front Pharmacol.* (2020) 11:1–9. doi: 10.3389/fphar.2020.566783
93. Chiş A, Noubissi PA, Pop OL, Mureşan CI, Fokam Tagne MA, Kamgang R, et al. Bioactive compounds in *Moringa oleifera*: mechanisms of action, focus on their anti-inflammatory properties. *Plan Theory.* (2024) 13:20. doi: 10.3390/plants13010020
94. Pamuru RR, Sucharitha K V, Vadde R. Immuno-Oncology of Colorectal Cancer. In: Vadde R, Nagaraju, G.P. (eds) Immunotherapy for Gastrointestinal Malignancies. Diagnostics and Therapeutic Advances in GI Malignancies. Springer, Singapore.
95. Min L, Chi Y, Dong S. Gut microbiota health closely associates with PCB153-derived risk of host diseases. *Ecotoxicol Environ Saf.* (2020) 203:111041. doi: 10.1016/j.ecoenv.2020.111041
96. Lu D, Huang Y, Kong Y, Tao T, Zhu X. Gut microecology: why our microbes could be key to our health. *Biomed Pharmacother.* (2020) 131:110784. doi: 10.1016/j.biopha.2020.110784
97. Magne F, Gotteland M, Gauthier L, Zazueta A, Pessoa S, Navarrete P, et al. The firmicutes/bacteroidetes ratio: A relevant marker of gut dysbiosis in obese patients? *Nutrients.* (2020) 12:1474. doi: 10.3390/nu12051474
98. Singh S, Sharma P, Sarma DK, Kumawat M, Tiwari R, Verma V, et al. Implication of obesity and gut microbiome dysbiosis in the etiology of colorectal cancer. *Cancers (Basel).* (2023) 15:1–28. doi: 10.3390/cancers15061913
99. He T-B, Huang Y-P, Huang Y, Wang X-J, Hu J-M, Sheng J. Structural elucidation and antioxidant activity of an arabinogalactan from the leaves of *Moringa oleifera*. *Int J Biol Macromol.* (2018) 112:126–33. doi: 10.1016/j.ijbiomac.2018.01.110
100. Liu X, Xu Q-X, Wang D-B, Zhao J, Wu Y, Liu Y, et al. Improved methane production from waste activated sludge by combining free ammonia with heat pretreatment: performance, mechanisms and applications. *Bioresour Technol.* (2018) 268:230–6. doi: 10.1016/j.biortech.2018.07.109
101. Guo C, Wang Y, Zhang S, Zhang X, Du Z, Li M, et al. *Crataegus pinnatifida* polysaccharide alleviates colitis via modulation of gut microbiota and SCFAs metabolism. *Int J Biol Macromol.* (2021) 181:357–68. doi: 10.1016/j.ijbiomac.2021.03.137
102. Do MH, Lee H-B, Oh M-J, Jhun H, Choi SY, Park H-Y. Polysaccharide fraction from greens of *Raphanus sativus* alleviates high fat diet-induced obesity. *Food Chem.* (2021) 343:128395. doi: 10.1016/j.foodchem.2020.128395
103. Miranda PM, De Palma G, Serkis V, Lu J, Louis-Auguste MP, McCarville JL, et al. High salt diet exacerbates colitis in mice by decreasing *Lactobacillus* levels and butyrate production. *Microbiome.* (2018) 6:57. doi: 10.1186/s40168-018-0433-4
104. Elabd EMY, Morsy SM, Elmal HA. Investigating of *moringa oleifera* role on gut microbiota composition and inflammation associated with obesity following high fat diet feeding. *Open Access Maced J Med Sci.* (2018) 6:1359–64. doi: 10.3889/oamjms.2018.313
105. Husien HM, Rehman SU, Duan Z, Wang M. Effect of *Moringa oleifera* leaf polysaccharide on the composition of intestinal microbiota in mice with dextran sulfate sodium-induced ulcerative colitis. *Front Nutr.* (2024) 11:1–10. doi: 10.3389/fnut.2024.1409026
106. Zeng X, Cao Y, Huang K, Yan Y, Chen D, Zhao Y, et al. Ascorbic acid derivative 2-o- $\beta$ -d-glucopyranosyl-L-ascorbic acid from the fruit of *lycium barbarum* modulates microbiota in the small intestine and colon and exerts an immunomodulatory effect on cyclophosphamide-treated BALB/c mice. *J Agric Food Chem.* (2020) 68:11128–43. doi: 10.1021/acs.jafc.0c04253
107. Wen Z, Tian H, Liang Y, Guo Y, Deng M, Liu G, et al. *Moringa oleifera* polysaccharide regulates colonic microbiota and immune repertoire in C57BL/6 mice. *Int J Biol Macromol.* (2022) 198:135–46. doi: 10.1016/j.ijbiomac.2021.12.085
108. Cooper MA, Colonna M, Yokoyama WM. Hidden talents of natural killers: NK cells in innate and adaptive immunity. *EMBO Rep.* (2009) 10:1103–10. doi: 10.1038/embor.2009.203
109. Bretto E, Frara S, Armandi A, Caviglia GP, Saracco GM, Bugianesi E, et al. *Helicobacter pylori* in inflammatory bowel diseases: active protagonist or innocent bystander? *Antibiotics.* (2024) 13:267. doi: 10.3390/antibiotics13030267
110. Korniluk A, Koper O, Kemona H, Dymicka-Piekarska V. From inflammation to cancer. *Ir J Med Sci.* (2017) 186:57–62. doi: 10.1007/s11845-016-1464-0
111. Sindhu R, Sood N, Puri V, Arora S. Various animal models for preclinical testing of anti-inflammatory agents. (2017). Available online at: <https://consensus.app/papers/various-animal-models-for-preclinical-testing-of-sindhu-sood/316ec8d26de85b9cb62f360a58a58b98/> (Accessed February 10, 2024).
112. Cooke J. Inflammation and its role in regeneration and repair. *Circ Res.* (2019) 124:1166–8. doi: 10.1161/CIRCRESAHA.118.314669
113. Rajan TS, Giacompo S, Iori R, De Nicola GR, Grassi G, Pollastro F, et al. Anti-inflammatory and antioxidant effects of a combination of cannabidiol and moringin in LPS-stimulated macrophages. *Fitoterapia.* (2016) 112:104–15. doi: 10.1016/j.fitote.2016.05.008
114. Marrocco A, Ortiz LA. Role of metabolic reprogramming in pro-inflammatory cytokine secretion from LPS or silica-activated macrophages. *Front Immunol.* (2022) 13:1–13. doi: 10.3389/fimmu.2022.936167
115. Bordon Y. Inflammasome blockade keeps the thymus young. *Nat Rev Immunol.* (2012) 12:154. doi: 10.1038/nri3188
116. Iyer SS, Cheng G. Role of interleukin 10 transcriptional regulation in inflammation and autoimmune disease. *Crit Rev Immunol.* (2012) 32:23–63. doi: 10.1615/critrevimmunol.v32.i1.30
117. Fioranelli M, Rocca MG, Flavin D, Cota L. Regulation of inflammatory reaction in health and disease. *Int J Mol Sci.* (2021) 22:1–13. doi: 10.3390/ijms22105277
118. Saini R, Singh S. Inducible nitric oxide synthase: an asset to neutrophils. *J Leukoc Biol.* (2018) 105:49–61. doi: 10.1002/JLB.4RU0418-161R
119. Packer M, Porteous C, Murphy M. Superoxide production by mitochondria in the presence of nitric oxide forms peroxynitrite. *IUBMB Life.* (1996) 40:527–34. doi: 10.1080/15216549600201103
120. Frejborg E, Salo T, Salem A. Role of cyclooxygenase-2 in head and neck tumorigenesis. *Int J Mol Sci.* (2020) 21:9246. doi: 10.3390/ijms21239246
121. Simmons DL, Botting RM, Hla T. Cyclooxygenase isozymes: the biology of prostaglandin synthesis and inhibition. *Pharmacol Rev.* (2004) 56:387–437. doi: 10.1124/pr.56.3.3
122. Luetragoon T, Sranujit RP, Noysang C, Thongsri Y, Potup P, Suphrom N, et al. Bioactive compounds in *moringa oleifera* lam. Leaves inhibit the pro-inflammatory mediators in lipopolysaccharide-induced human monocyte-derived macrophages. *Molecules.* (2020) 25:1–16. doi: 10.3390/molecules25010191
123. Leonard WJ, Lin JX. Cytokine receptor signaling pathways. *J Allergy Clin Immunol.* (2000) 105:877–88. doi: 10.1067/mai.2000.106899
124. Chang D, Dela Cruz C, Sharma L. Beneficial and detrimental effects of cytokines during influenza and COVID-19. *Viruses.* (2024) 16:308. doi: 10.3390/v16020308
125. Megha K, Joseph X, Akhil V, Mohanan P. Cascade of immune mechanism and consequences of inflammatory disorders. *Phytomedicine.* (2021) 91:153712. doi: 10.1016/j.phymed.2021.153712
126. Wautier J, Wautier M. Pro- and anti-inflammatory prostaglandins and cytokines in humans: A Mini review. *Int J Mol Sci.* (2023) 24:24. doi: 10.3390/ijms24119647

127. Sailaja BS, Aita R, Maledatu S, Ribnický D, Verzi MP, Raskin I. Moringa isothiocyanate-1 regulates Nrf2 and NF- $\kappa$ B pathway in response to LPS-driven sepsis and inflammation. *PLoS One*. (2021) 16:e0248691. doi: 10.1371/journal.pone.0248691
128. Jomova K, Raptova R, Alomar SY, Alwasel SH, Nepovimova E, Kuca K, et al. Reactive oxygen species, toxicity, oxidative stress, and antioxidants: chronic diseases and aging. *Arch Toxicol*. (2023) 97:2499–574. doi: 10.1007/s00204-023-03562-9
129. Afzal S, Abdul Manap AS, Attiq A, Albokhadaim I, Kandeel M, Alhojaily SM. From imbalance to impairment: the central role of reactive oxygen species in oxidative stress-induced disorders and therapeutic exploration. *Front Pharmacol*. (2023) 14:1–22. doi: 10.3389/fphar.2023.1269581
130. Brand M. Mitochondrial generation of superoxide and hydrogen peroxide as the source of mitochondrial redox signaling. *Free Radic Biol Med*. (2016) 100:14–31. doi: 10.1016/j.freeradbiomed.2016.04.001
131. Jain A, Shakkarpude J. Oxidative stress: a biomarker for animal health and production: a review. *Indian J Anim Res*. (2024) 58:1–12. doi: 10.18805/IJAR.B-5300
132. Zhang L, Wang X, Cueto R, Effi C, Zhang Y, Tan H, et al. Biochemical basis and metabolic interplay of redox regulation. *Redox Biol*. (2019) 26:101284. doi: 10.1016/j.redox.2019.101284
133. Andrés Juan C, Pérez M, de la Lastra J, Plou FJ, Pérez-Lebeña E, Reinbothe S. Molecular sciences the chemistry of reactive oxygen species (ROS) revisited: outlining their role in biological macromolecules (DNA, lipids and proteins) and induced pathologies. *Int J Mol Sci*. (2021) 22:4642. doi: 10.3390/ijms
134. Redza-Dutordoir M, Averill-Bates DA. Activation of apoptosis signalling pathways by reactive oxygen species. *Biochim Biophys Acta, Mol Cell Res*. (2016) 1863:2977–92. doi: 10.1016/j.bbamcr.2016.09.012
135. Adamo SA. The effects of stress hormones on immune function may be vital for the adaptive reconfiguration of the immune system during fight-or-flight behavior. *Integr Comp Biol*. (2014) 54:419–26. doi: 10.1093/icb/icu005
136. Khan MZ, Huang B, Kou X, Chen Y, Liang H, Ullah Q, et al. Enhancing bovine immune, antioxidant and anti-inflammatory responses with vitamins, rumen-protected amino acids, and trace minerals to prevent periparturient mastitis. *Front Immunol*. (2024) 14:1290044. doi: 10.3389/fimmu.2023.1290044
137. Younis N, Khan MI, Zahoor T, Faisal MN. Phytochemical and antioxidant screening of *Moringa oleifera* for its utilization in the management of hepatic injury. *Front Nutr*. (2022) 9:1–11. doi: 10.3389/fnut.2022.1078896
138. Singh VK. A review of *Moringa oleifera* as animal fodder: nutritive value and health aspects. *Int J Vet Sci Anim Husband*. (2024) 9:81–4.
139. Olvera-Aguirre G, Mendoza-Taco MM, Moo-Huchin VM, Lee-Rangel HA, Roque-Jiménez JA, Gómez-Vázquez A, et al. Effect of extraction type on bioactive compounds and antioxidant activity of *Moringa oleifera* lam. Leaves. *Agriculture*. (2022) 12:1–9. doi: 10.3390/agriculture12091462
140. Shalaby EA, Shanab SMM, El-Raheem WMA, Hanafy EA. Biological activities and antioxidant potential of different biosynthesized nanoparticles of *Moringa oleifera*. *Sci Rep*. (2022) 12:18400–14. doi: 10.1038/s41598-022-23164-2
141. Paikra BK, Dhongade HKJ, Gidwani B. Phytochemistry and pharmacology of *Moringa oleifera* lam. *J Pharmacopuncture*. (2017) 20:194–200. doi: 10.3831/kpi.2017.20.022
142. Modisaojang-Mojanaga MM, Ogbuwu IP, Oguttu JW, Mbajiorgu CA. Moringa leaf meal improves haemato-biochemical and production indices in broiler chickens: a review. *Comp Clin Path*. (2019) 28:621–32. doi: 10.1007/s00580-019-02900-7
143. Baliyan S, Mukherjee R, Priyadarshini A, Vibhuti A, Gupta A, Pandey RP, et al. Determination of antioxidants by DPPH radical scavenging activity and quantitative phytochemical analysis of *Ficus religiosa*. *Molecules*. (2022) 27:1326. doi: 10.3390/molecules27041326
144. Gulcin İ, Alwasel SH. DPPH radical scavenging assay. *Processes*. (2023) 11:2248. doi: 10.3390/pr11082248
145. Sreelatha S, Padma PR. Antioxidant activity and total phenolic content of *Moringa oleifera* leaves in two stages of maturity. *Plant Foods Hum Nutr*. (2009) 64:303–11. doi: 10.1007/s11130-009-0141-0
146. Ojha PS, Tripathi V, Dwivedi P. Antioxidant capacity and radical scavenging activities of *Moringa oleifera*. *Acta Sci Agric*. (2024) 8:35–7.
147. Siddhuraju P, Becker K. Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera* lam.) leaves. *J Agric Food Chem*. (2003) 51:2144–55. doi: 10.1021/jf020444+
148. Duranti G, Maldini M, Crognale D, Horner K, Dimauro I, Sabatini S, et al. *Moringa oleifera* leaf extract upregulates nrf2/ho-1 expression and ameliorates redox status in c2c12 skeletal muscle cells. *Molecules*. (2021) 26:26. doi: 10.3390/molecules26165041
149. Gaschler M, Stockwell B. Lipid peroxidation in cell death. *Biochem Biophys Res Commun*. (2017) 482:419–25. doi: 10.1016/j.bbrc.2016.10.086
150. Mortensen MS, Ruiz J, Watts JL. Polyunsaturated fatty acids drive lipid peroxidation during Ferroptosis. *Cells*. (2023) 12:804. doi: 10.3390/cells12050804
151. Wood LG, Gibson PG, Garg ML. Biomarkers of lipid peroxidation, airway inflammation and asthma. *Eur Respir J*. (2003) 21:177–86. doi: 10.1183/09031936.03.00017003a
152. Yuji T, Naito Y. What is oxidative stress? *Jpn Med Assoc J*. (2002) 124:271–6.
153. Larsson K, Harrysson H, Havenaar R, Alminger M, Undeland I. Formation of malondialdehyde (MDA), 4-hydroxy-2-hexenal (HHE) and 4-hydroxy-2-nonenal (HNE) in fish and fish oil during dynamic gastrointestinal in vitro digestion. *Food Funct*. (2016) 7:1176–87. doi: 10.1039/c5fo01401h
154. Papastergiadis A, Fatouh A, Jaxsens L, Lachat C, Shrestha K, Daelman J, et al. Exposure assessment of malondialdehyde, 4-Hydroxy-2-(E)-Nonenal and 4-Hydroxy-2-(E)-Hexenal through specific foods available in Belgium. *Food Chem Toxicol*. (2014) 73:51–8. doi: 10.1016/j.fct.2014.06.030
155. Fitri A, Toharmat T, Astuti DA, Tamura H. The potential use of secondary metabolites in *moringa oleifera* as an antioxidant source. *Media Peternak*. (2015) 38:169–75. doi: 10.3398/medpet.2015.38.3.169
156. Sinha M, Das DK, Bhattacharjee S, Majumdar S, Dey S. Leaf extract of *moringa oleifera* prevents ionizing radiation-induced oxidative stress in mice. *J Med Food*. (2011) 14:1167–72. doi: 10.1089/jmf.2010.1506
157. Sinha M, Das DK, Datta S, Ghosh S, Dey S. Amelioration of ionizing radiation induced lipid peroxidation in mouse liver by *Moringa oleifera* lam. Leaf extract. *Indian J Exp Biol*. (2012) 50:209–15.
158. Singh D, Arya PV, Aggarwal VP, Gupta RS. Evaluation of antioxidant and hepatoprotective activities of *Moringa oleifera* lam. Leaves in carbon tetrachloride-intoxicated rats. *Antioxidants*. (2014) 3:569–91. doi: 10.3390/antiox3030569
159. Nkukwana T, Muchenje V, Masika PJ, Dzama K, Descalzo A. Fatty acid composition and oxidative stability of breast meat from broiler chickens supplemented with *Moringa oleifera* leaf meal over a period of refrigeration. *Food Chem*. (2014) 142:255–61. doi: 10.1016/j.foodchem.2013.07.059
160. Kamble MT, Gallardo W, Salin KR, Pumpuang S, Chavan BR, Bhujel RC, et al. Effect of *Moringa oleifera* leaf extract on the growth performance, hematology, innate immunity, and disease resistance of Nile tilapia (*Oreochromis niloticus*) against *Streptococcus agalactiae* biotype 2. *Animals*. (2024) 14:953. doi: 10.3390/ani14060953
161. El-Kassas S, Aljahdali N, Abdo SE, Alaryani FS, Moustafa EM, Mohamed R, et al. *Moringa oleifera* leaf powder dietary inclusion differentially modulates the antioxidant, inflammatory, and histopathological responses of Normal and *Aeromonas hydrophila*-infected mono-sex Nile Tilapia (*Oreochromis niloticus*). *Front Vet Sci*. (2022) 9:918933. doi: 10.3389/fvets.2022.918933
162. Li Z, Luo W, Zhou Q, Sun C, Zheng X, Liu B, et al. Investigation of the fermentation process of *Moringa oleifera* leaves and its effects on the growth performance, antioxidant capacity, and intestinal microbiome of *Procambarus clarkii*. *Antioxidants*. (2024) 13:1355. doi: 10.3390/antiox13111355
163. Peñalver R, Martínez-zamora L, Lorenzo JM, Ros G, Nieto G. Nutritional and antioxidant properties of *Moringa oleifera* leaves in functional foods. *Food Secur*. (2022) 11:1–13. doi: 10.3390/foods11081107
164. Sanodiya P, Prasad K, Neel B, Mishra K. Maize nutritional quality and value addition: a brief overview. *Maize Journal* (2024) 11:75–82.
165. A W III, B J. Soybean meal quality and analytical techniques. In: Soybean and nutrition. *Intech Open*. (2011)
166. Li J, Zhang S, Gu X, Xie J, Zhu X, Wang Y, et al. Effects of alfalfa levels on carcass traits, meat quality, fatty acid composition, amino acid profile, and gut microflora composition of Heigai pigs. *Front Nutr*. (2022) 9:975455. doi: 10.3389/fnut.2022.975455
167. Moringa Job D. D. As a source of minerals? A comparison with corn and soybean. *Technical Report* (2018)1–22.