



# V $\gamma$ 9V $\delta$ 2 T Cells as Strategic Weapons to Improve the Potency of Immune Checkpoint Blockade and Immune Interventions in Human Myeloma

Barbara Castella<sup>1</sup>, Assunta Melaccio<sup>2</sup>, Myriam Foglietta<sup>1,3</sup>, Chiara Riganti<sup>4</sup> and Massimo Massaia<sup>1,3\*</sup>

<sup>1</sup> Laboratorio di Immunologia dei Tumori del Sangue, Centro Interdipartimentale di Ricerca in Biologia Molecolare, Università degli Studi di Torino, Turin, Italy, <sup>2</sup> Dipartimento di Scienze Biomediche ed Oncologia Umana, Sezione di Medicina Interna ed Oncologia, Università degli studi di Bari "A. Moro", Bari, Italy, <sup>3</sup> SC Ematologia, AO S.Croce e Carle, Cuneo, Italy, <sup>4</sup> Dipartimento di Oncologia, Università degli Studi di Torino, Turin, Italy

## OPEN ACCESS

### Edited by:

Nicola Giuliani,  
Università degli Studi di Parma, Italy

### Reviewed by:

Daniel Olive,  
Aix Marseille Université, France  
Wenwei Tu,  
University of Hong Kong, Hong Kong

### \*Correspondence:

Massimo Massaia  
massimo.massaia@unito.it

### Specialty section:

This article was submitted to  
Cancer Immunity and Immunotherapy,  
a section of the journal  
Frontiers in Oncology

**Received:** 02 August 2018

**Accepted:** 17 October 2018

**Published:** 06 November 2018

### Citation:

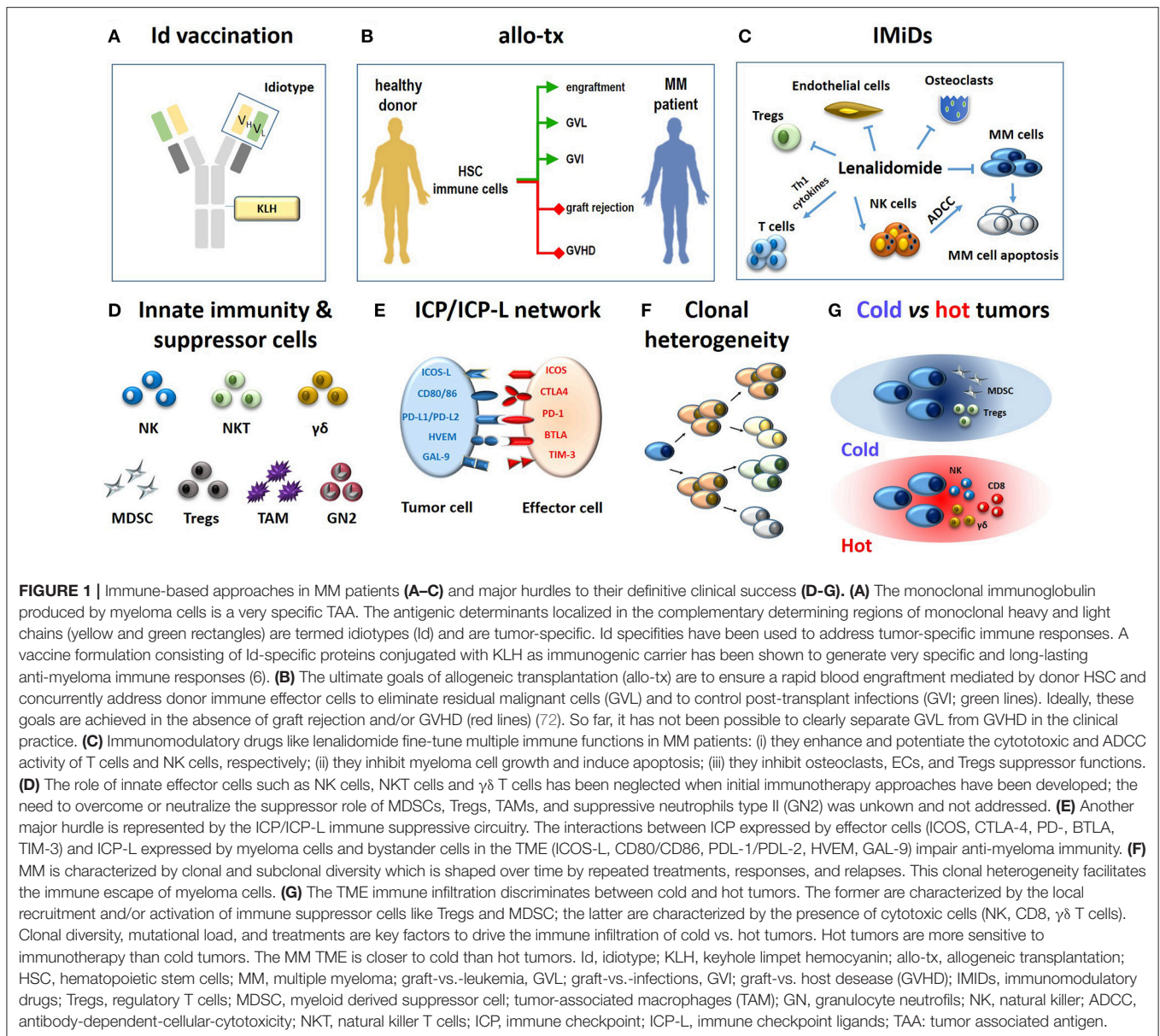
Castella B, Melaccio A, Foglietta M,  
Riganti C and Massaia M (2018)  
V $\gamma$ 9V $\delta$ 2 T Cells as Strategic Weapons  
to Improve the Potency of Immune  
Checkpoint Blockade and Immune  
Interventions in Human Myeloma.  
Front. Oncol. 8:508.  
doi: 10.3389/fonc.2018.00508

The advent of immune checkpoint (ICP) blockade has introduced an unprecedented paradigm shift in the treatment of cancer. Though very promising, there is still a substantial proportion of patients who do not respond or develop resistance to ICP blockade. *In vitro* and *in vivo* models are eagerly needed to identify mechanisms to maximize the immune potency of ICP blockade and overcome primary and acquired resistance to ICP blockade. V $\gamma$ 9V $\delta$ 2 T cells isolated from the bone marrow (BM) from multiple myeloma (MM) are excellent tools to investigate the mechanisms of resistance to PD-1 blockade and to decipher the network of mutual interactions between PD-1 and the immune suppressive tumor microenvironment (TME). V $\gamma$ 9V $\delta$ 2 T cells can easily be interrogated to dissect the progressive immune competence impairment generated in the TME by the long-lasting exposure to myeloma cells. BM MM V $\gamma$ 9V $\delta$ 2 T cells are PD-1<sup>+</sup> and anergic to phosphoantigen (pAg) stimulation; notably, single agent PD-1 blockade is insufficient to fully recover their anti-tumor activity *in vitro* indicating that additional players are involved in the anergy of V $\gamma$ 9V $\delta$ 2 T cells. In this mini-review we will discuss the value of V $\gamma$ 9V $\delta$ 2 T cells as investigational tools to improve the potency of ICP blockade and immune interventions in MM.

**Keywords:** V $\gamma$ 9V $\delta$ 2 T cells, immune checkpoint blockade, immunotherapy, tumor vaccination, multiple myeloma

## INTRODUCTION

Multiple myeloma (MM) is a disease characterized by the malignant growth of clonal plasma cells (hereafter referred to as myeloma cells) driven by intrinsic and extrinsic mechanisms. MM is uniformly preceded by a premalignant phase, termed monoclonal gammopathy of undetermined significance (MGUS). The risk of progression from MGUS to MM varies from 1 to 5% per year (1). Interestingly, myeloma cells isolated from the BM of MGUS already harbor many of the genetic and epigenetic abnormalities of myeloma cells isolated from patients with overt disease. Interestingly, long-term follow up has shown that almost 50% of high-risk MGUS never progresses to overt MM (2). These clinical data strongly support the concept that other factors, in addition to intrinsic myeloma cell features, are important to determine the fate and aggressiveness of myeloma cells.



The nature and relevance of the tumor microenvironment (TME) in MM have comprehensively been described elsewhere, including the role of immune cells (3, 4). We have anticipated these insights in the mid '80s, when we have shown a defective CD73 expression in CD8<sup>+</sup> cells which was correlated with the proliferative activity of BM PC in both MGUS and MM (5). These initial findings have been corroborated by many other preclinical studies leading to the pioneering development of active specific immunotherapy approaches. The unique expression of idiotype (Id) by clonal B cells encouraged the generation of a variety of Id-specific vaccines (from protein- to DNA-based vaccines) which were able to induce long-lasting and tumor-specific immune responses (6).

Clinical results in allo-transplanted MM patients have strengthened the perception that the only chance to permanently eliminate residual myeloma cells [including those surviving high dose melphalan and autologous stem cell transplantation (ASCT)] is the recognition and elimination by allogeneic immune effector cells (7). The development of immunomodulatory imide drugs (IMiDs) and the clinical results obtained with lenalidomide (including maintenance treatment after ASCT) have brought further evidences that immune cells in the TME are key targets to interrupt the myeloma cell prosurvival network (8).

These approaches have significantly impacted on the clinical outcome, but none of them has generated such an impressive cure rate to definitely change the natural history of the disease (Figure 1).

## RECONSIDERING THE IMMUNE COMPETENCE OF MGUS AND MM PATIENTS

The unsatisfactory results of immune-based approaches in MM should not generate a pessimistic view. The reasons are rooted in the increased knowledge about the pathogenesis of the disease, the pathophysiology of immune responses, and the innovative technologies available to monitor the disease, assess clinical responses, and develop novel strategies of immune interventions. Additional progresses have been made by shedding some misconceptions like the wisdom that MGUS are immunologically blessed conditions in which myeloma cells are held in check by very effective immune responses. This misconception was based on mouse models and preclinical results obtained in humans when much less was known about the mechanisms of immune surveillance and immune escape (9). Only recently, this misconception has been breached by us and others revealing that multiple immune dysfunctions are already present in MGUS (10–13).

Another misconception to be abandoned is that the remission state after ASCT represents a unique opportunity for immune interventions since it is possible to achieve a minimal residual disease (MRD) condition in this setting. We have shown more than 10 years ago that the T-cell receptor (TCR) repertoire is highly disrupted in patients in remission after ASCT (14). These results have been confirmed and consolidated (15) explaining why Id vaccination could not fulfil clinical expectations and why lenalidomide maintenance, even nowadays, significantly extends progression free survival (PFS), but does not definitely protect MM patients from late or very late relapse (8).

The time is ripe to apply more informative assays to investigate the immune competence of MGUS and MM. The aim of this minireview is to recapitulate how interrogating the immune competence of BM V $\gamma$ 9V $\delta$ 2 T cells has deepened our knowledge about the immune derangement occurring in MGUS and MM patients and how these informations can be applied to design more effective immune interventions in MM.

## V $\gamma$ 9V $\delta$ 2 T CELLS AS ULTRASENSITIVE TOOLS TO ASSESS THE IMMUNE SUPPRESSIVE TME COMMITMENT IN MGUS AND MM

V $\gamma$ 9V $\delta$ 2 T-cells are non-conventional T cells half-way between adaptive and innate immunity with a natural inclination to react against malignant B cells, including malignant myeloma cells (16). These cells are able to sense supra-physiological concentrations of phosphorylated metabolites (pAgs) generated in the mevalonate (Mev) pathway of mammalian cells. Isopentenyl pyrophosphate (IPP) is the prototypic pAg recognized by V $\gamma$ 9V $\delta$ 2 T cells. The pAgs-reactivity of V $\gamma$ 9V $\delta$ 2 T cells can be tested *in vivo* and *in vitro* by stimulating monocytes or dendritic cells (DC) with aminobisphosphonates like pamidronate or zoledronate (ZA). Both compounds inhibit farnesylpyrophosphate synthase in the Mev pathway (17, 18)

and induce intracellular IPP accumulation and extracellular IPP release that are detected by V $\gamma$ 9V $\delta$ 2 T cells. IPP recognition by V $\gamma$ 9V $\delta$ 2 T cells is mediated by the  $\gamma\delta$  TCR in association with the isoform A1 of the butyrophilin-3 (BTN3A1) protein family (19, 20).

V $\gamma$ 9V $\delta$ 2 T cells are endowed with peculiar functional properties which make them very good candidates for immunotherapy: they do not require MHC restriction and co-stimulation; they produce pro-inflammatory cytokines (IFN- $\gamma$  and TNF- $\alpha$ ); they recognize antigens shared by a variety of stressed and tumor cells; they behave as professional antigen-presenting cells (21); they can provide help to B cells to produce antibodies (22); and they can induce DC maturation boosting  $\alpha\beta$  T cell priming and MHC-restricted antigen-specific T-cell responses (23). We believe that this multifaceted array of immune functions gives a unique predisposition to V $\gamma$ 9V $\delta$ 2 T cells to behave as very sensitive biosensors of the immune suppressive TME commitment occurring in the BM of MGUS and MM patients (24).

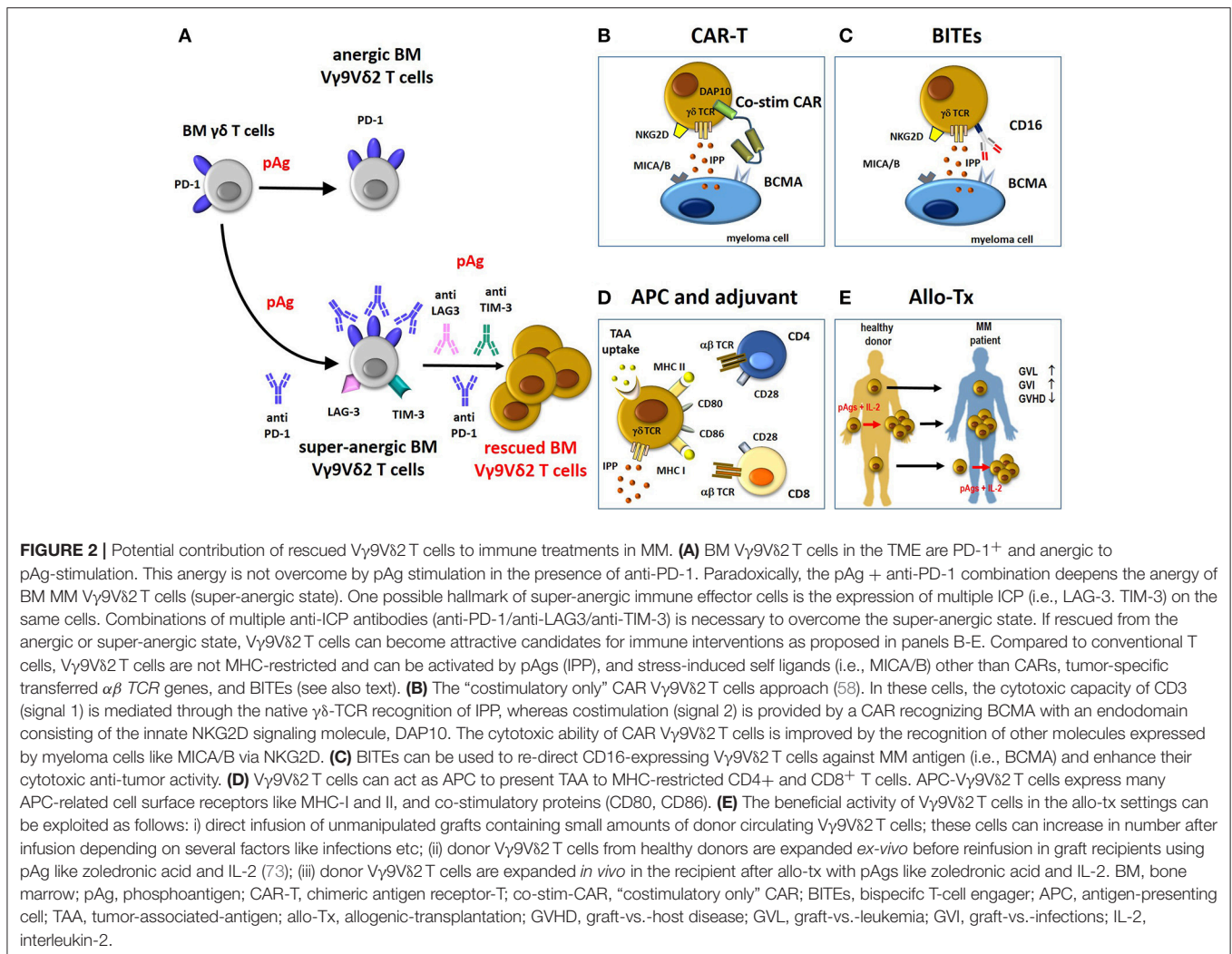
We have previously shown in a large series of patients (MGUS:  $n = 10$ ; MM at diagnosis:  $n = 70$ ; MM in remission:  $n = 52$ ; MM in relapse:  $n = 24$ ) that BM MM V $\gamma$ 9V $\delta$ 2 T cells are unable to properly react to pAgs stimulation in terms of proliferation, CD107 expression and IFN- $\gamma$  production. This is an early and long-lasting immune dysfunction, already detectable in MGUS individuals, largely anticipating that of CD8+ T cells and not disappearing even when most of tumor cells have been cleared by ASCT as in MM in remission. The investigation of pAgs reactivity of BM MM V $\gamma$ 9V $\delta$ 2 T cells has been instrumental to show that the frequency of immune suppressor cells in the TME [bone marrow stromal cells (BMSC), regulatory T cells (Tregs) and myeloid-derived suppressor cells (MDSC)] are similar in the BM of MGUS, MM at diagnosis and MM in remission.

## ROLE OF IMMUNE CHECKPOINTS (ICP) AND ICP-LIGANDS (ICP-L) IN THE IMMUNE SUPPRESSIVE TME COMMITMENT OF MGUS AND MM PATIENTS

Immune checkpoints (ICP) are key regulators of immune activation, immune homeostasis, and autoimmunity driven by interactions with the corresponding ligands (ICP-L) expressed by surrounding cells (25). In cancer, the ICP/ICP-L network is often hijacked by tumor cells to suppress anti-tumor immune responses. This has led to the development of anti-ICP/ICP-L monoclonal antibodies (mAbs) to treat a variety of cancers with heterogeneous results.

Among the ICP/ICP-L pairs identified so far, the PD-1/PD-L1 axis plays a major role in the generation of the immune suppressive TME in MM. PD-L1 expression in myeloma cells is higher in MM and SMM than in MGUS and predicts an increased risk of disease progression (26, 27). Paiva et al. have shown a significant upregulation of PD-L1 expression in residual myeloma cells of MM patients who are in first complete





peptides (44, 45). Cancer testis antigens are under investigation as potential TAA in MM patients (50, 52).

Despite a growing enthusiasm, immunotherapy progresses are still facing many hurdles. The majority of MM treated with anti-CD38 mAbs (daratumumab) eventually progress and the mechanisms involved in resistance to daratumumab are largely unknown. CAR T cells also are not free from handicaps like reduced expression of BCMA on myeloma cells, short persistence or loss *in vivo* of functional CAR T cells (44–49). Bispecific CAR T cells targeting simultaneously two myeloma associated antigens may compensate the decreased BCMA expression, but it may also increase on-target off-tumor toxicity. MHC down-regulation on tumor cells may compromise the therapeutic efficacy of  $\alpha\beta$ TCR gene transferred T cells, whereas the eventual recognition of cross reactive epitopes from alternative target antigens may account for considerable on-target off-tumor toxicity. Autoimmune fatal complications have occurred with MAGE-A3 enhanced affinity  $\alpha\beta$ TCR gene transferred T cells (51). Another drawback of  $\alpha\beta$ TCR gene transfer to conventional CD3<sup>+</sup>  $\alpha\beta$  T cells is the formation of mixed TCR dimers with unknown specificities

due to pairing of endogenous and introduced  $\alpha$  and  $\beta$  TCR chains (53).

BM V $\gamma$ 9V $\delta$ 2 T cells can be very attractive candidates to deliver antitumor responses in MM, provided that they are rescued from the immune dysfunction they are afflicted. These cells recognize a broader range of targets (including metabolic targets like IPP and self-induced stress ligands) and possess a more favorable safety profile than conventional T cells (16). This unique feature has been exploited to reduce the potential “off target” toxicity of CAR V $\gamma$ 9V $\delta$ 2 T cells (54–57). Fisher et al (58) have designed “costimulatory only” CAR V $\gamma$ 9V $\delta$ 2 T cells in which activation signals 1 and 2 are provided by separate receptors. In these dual-receptor CAR V $\gamma$ 9V $\delta$ 2 T cells, the cytotoxic capacity of CD3 (signal 1) is mediated via the native  $\gamma\delta$ -TCR recognizing IPP, whereas costimulation (signal 2) is provided by a CAR-mediated recognition of TAA mediated by DAP10, the endodomain consisting of the NKG2D receptor (**Figure 2B**). Normal healthy tissues which do not express IPP do not activate V $\gamma$ 9V $\delta$ 2 TCR and are spared from V $\gamma$ 9V $\delta$ 2 T cell cytotoxicity. Interestingly, these “costimulation only” CAR V $\gamma$ 9V $\delta$ 2 T cells express lower

levels of PD1 and TIM3 than traditional CAR V $\gamma$ 9V $\delta$ 2 T cells after long term culture (58).

V $\gamma$ 9V $\delta$ 2 T cells are excellent candidates for  $\alpha\beta$ TCR gene transfer without the risk of expression of undesired mixed TCR dimers (59). Another interesting approach is to engineer  $\alpha\beta$  T cells to express tumor-specific V $\gamma$ 9V $\delta$ 2 TCRs (TEGs) to redirect  $\alpha\beta$  T cells against cancer cells (60). V $\gamma$ 9V $\delta$ 2 TCR-redirection  $\alpha\beta$  T cells very efficiently kill cancer cell lines *in vitro* and primary acute myeloid leukemia blasts in a humanized mouse model. Very recently, TEGs have also been generated in MM patients and shown to be able to recognize and kill myeloma cells in a 3D model (61). V $\gamma$ 9V $\delta$ 2 T cells can also be redirected against myeloma cells with BITEs (Figure 2C). The bispecific antibody [(HER2) $\times$ CD16] has been used to re-direct CD16<sup>+</sup> V $\gamma$ 9V $\delta$ 2 T cells against Her2<sup>+</sup> tumor cells that were killed with very high efficiency (62). HLA-independent recognition of TAA by tumor-redirection CAR V $\gamma$ 9V $\delta$ 2 T cells or BITEs-activated V $\gamma$ 9V $\delta$ 2 T cells may prelude to the development of allogeneic “off the shelf” CAR products.

Another unique feature of V $\gamma$ 9V $\delta$ 2 T cells is their capacity to act as antigen-presenting cells (APC) to boost antigen-specific immune responses mediated by CD8<sup>+</sup> cells (21, 63) (Figure 2D). Combination therapy of V $\gamma$ 9V $\delta$ 2 T-APC-based vaccines with ICP blockade may have synergistic activity leading to enhanced anti-tumor immune responses and long-lived immuno-surveillance (64, 65). These adjuvant properties are not lost even after chimerization of V $\gamma$ 9V $\delta$ 2 T cells as demonstrated by Capsomidis A. (57)

Lastly, the multifunctional properties of V $\gamma$ 9V $\delta$ 2 T cells may also be beneficial in the allo-tx setting (allo-tx) (Figure 2E) (66). V $\gamma$ 9V $\delta$ 2 T cells have been reported to cause less graft-vs.-host disease (GVHD) than  $\alpha\beta$  T cells while retaining graft-vs.-leukemia activity (GVL) (67, 68). A protective effect of V $\gamma$ 9V $\delta$ 2 T cells against both leukemia cell regrowth and infections has been reported in haploidentical HSCT depleted of TCR- $\alpha\beta$ /CD19

lymphocytes (69). Lastly, recent studies suggest an overall favorable effect of high V $\gamma$ 9V $\delta$ 2 T cells immune reconstitution after HSCT; patients with elevated numbers of V $\gamma$ 9V $\delta$ 2 T cells had a significantly higher overall survival rate and a decreased rate of acute GVHD compared to patients with low V $\gamma$ 9V $\delta$ 2 T cell counts (70).

## CONCLUSIONS

Investigation of BM MM V $\gamma$ 9V $\delta$ 2 T cells has been useful to gather a faithful picture of the immune suppressive TME in MGUS and MM. Understanding the mechanisms that are responsible for BM V $\gamma$ 9V $\delta$ 2 T-cell dysfunction, with special regard to resistance to PD-1 blockade, can help to overcome ICP resistance and safely integrate ICP/ICP-L blockade in the immune treatments of MGUS and MM patients. The use of nanotechnologies may improve delivery of antagonistic antibodies to block ICP inhibitory receptors compared to free antibodies and improve T cell activation (71).

Finally, the functional rescue of BM V $\gamma$ 9V $\delta$ 2 T cells is an attractive opportunity to exploit their multifaceted immune functions to carry on *ex-vivo* and *in vivo* adoptive immunotherapy interventions.

## AUTHOR CONTRIBUTIONS

All authors have made a substantial contributions to text and figures and have approved the manuscript for submission

## FUNDING

This study was supported by the Italian Association for Cancer Research (AIRC) (IG15232 to CR and IG 16985 to MM). B.C. is a post-doc research fellow supported by AIRC (IG 16985).

## REFERENCES

1. Van Nieuwenhuijzen N, Spaan I, Raymakers R, Peperzak V. From MGUS to multiple myeloma, a paradigm for clonal evolution of premalignant cells. *Cancer Res.* (2018) 78:2449–56. doi: 10.1158/0008-5472.CAN-17-3115
2. Brendan M Weiss. Advances in understanding monoclonal gammopathy of undetermined significance as a precursor of multiple myeloma. *Expert Rev Hematol.* (2010) 3:165–74. doi: 10.1586/ehm.10.13
3. Bianchi G, Munshi NC. Pathogenesis beyond the cancer clone(s) in multiple myeloma. *Blood* (2015) 125:3049–58. doi: 10.1182/blood-2014-11-568881
4. Ghobrial IM, Detappe A, Anderson KC, Steensma DP. The bone-marrow niche in MDS and MGUS: implications for AML and MM. *Nat Rev Clin Oncol.* (2018) 15:219–33. doi: 10.1038/nrclinonc.2017.197
5. Massaia M, Ma DD, Boccadoro M, Golzio F, Gavarotti P, Dianzani U, et al. Decreased ecto-5'-nucleotidase activity of peripheral blood lymphocytes in human monoclonal gammopathies: correlation with tumor cell kinetics. *Blood* (1985) 65:530–4.
6. Massaia M, Borrione P, Battaglio S, Mariani S, Beggiano, Napoli P, et al. Idiotype Vaccination in human myeloma: generation of tumor-specific immune responses after high-dose chemotherapy. *Blood* (1999) 94:673–83.
7. Rosenblatt J, Avigan D. Role of immune therapies for myeloma. *JNCCN* (2015) 13:1440–7 doi: 10.6004/jnccn.2015.0168
8. Fostier K, Caers J, Meuleman N, Broos K, Corthals J, Thielemans K, et al. Impact of lenalidomide maintenance on the immune environment of multiple myeloma patients with low tumor burden after autologous stem cell transplantation. *Oncotarget* (2018) 9:20476–89. doi: 10.18632/oncotarget.24944
9. Calcinotto A, Ponzoni M, Ria R, Grioni M, Cattaneo E, Villa I, et al. Modifications of the mouse bone marrow microenvironment favor angiogenesis and correlate with disease progression from asymptomatic to symptomatic multiple myeloma. *Oncoimmunology* (2015) 4:e1008850. doi: 10.1080/2162402X.2015.1008850
10. Das R, Strowig T, Verma R, Koduru S, Hafemann A, Hopf S, et al. Microenvironment-dependent growth of preneoplastic and malignant plasma cells in humanized mice. *Nat Med.* (2016) 22:1351–7. doi: 10.1038/nm.4202
11. Zheng MM, Zhang Z, Bemis K, Belch AR, Pilarski LM, Shively JE, et al. The systemic cytokine environment is permanently altered in multiple myeloma. *PLoS ONE* (2013) 8:e58504. doi: 10.1371/journal.pone.0058504
12. Foglietta M, Castella B, Mariani S, Coscia M, Godio L, Ferracini R, et al. The bone marrow of myeloma patients is steadily inhabited by a normal-sized pool of functional regulatory T cells irrespective of the disease status. *Haematologica* (2014) 99:1605–10. doi: 10.3324/haematol.2014.105866
13. Feng X, Acharya C, An G, Wen, Li Zhang K, Kalbasi A, et al. Targeting CD38 suppresses induction and function of T regulatory cells to

- reverse immunosuppression in multiple myeloma. *Blood* (2016) 128:2106. doi: 10.1158/1078-0432.CCR-16-3192
14. Toubai T, Hirate D, Shono Y, Ota S, Ibata M, Mashiko S, et al. Chimerism and T-cell receptor repertoire analysis after unrelated cord blood transplantation with a reduced-intensity conditioning regimen following autologous stem cell transplantation for multiple myeloma. *Int J Lab Hematol.* (2008) 30:75–81. doi: 10.1111/j.1751-553X.2007.00903
  15. Chung D J, Katherine B. Pronschinske, Justin A. Shyer, Sneha Sharma, et al. Young T cell exhaustion in multiple myeloma relapse after autotransplant: optimal timing of immunotherapy. *Cancer Immunol Res.* (2016) 4:61–71. doi: 10.1158/2326-6066
  16. Castella B, Vitale C, Coscia M, Massaia M. Vc9Vd2 T cell-based immunotherapy in hematological malignancies: from bench to bedside. *Cell. Mol. Life Sci.* (2011) 68:2419–2432. doi: 10.1007/s00018-011-0704-8
  17. Kunzmann V, Bauer E, Feurle J, Weissinger F, Tony HP, Wilhelm M. Stimulation of gammadelta T cells by aminobisphosphonates and induction of antiplasmacell activity in multiple myeloma. *Blood* (2000) 96:384–92.
  18. Thompson K, Rogers MJ. Statins prevent bisphosphonate-induced gammadelta-T-cell proliferation and activation in vitro. *J Bone Miner Res.* (2004) 19:278–88. doi: 10.1359/JBMR.0301230
  19. Riganti C, Castella B, Massaia M. ABCA1, apoA-I, and BTN3A1: a legitimate ménage à trois in dendritic cells. *Front Immunol.* (2018) 9:1246. doi: 10.3389/fimmu.2018.01246
  20. Castella B, Kopecka J, Sciancalepore P, Mandili G, Foglietta M, Mitro N, et al. The ATP-binding cassette transporter A1 regulates phosphoantigen release and Vy9V82 T cell activation by dendritic cells. *Nat Commun.* (2017) 8:15663. doi: 10.1038/ncomms15663
  21. Brandes M, Willmann K, Bioley G, Lévy N, Eberl M, Luo M, et al. Cross-presenting human gammadelta T cells induce robust CD8+ alphabeta T cell responses. *Proc Natl Acad Sci USA* (2009) 106:2307–12. doi: 10.1073/pnas.0810059106
  22. Born WK, Huang Y, Reinhardt RL, Huang H, Sun D, O'Brien RL.  $\gamma\delta$  T cells and B cells. *Adv Immunol.* (2017) 134:1–45. doi: 10.1016/bs.ai.2017.01.002
  23. Petrasca A, Doherty DG. Human V82(+)  $\gamma\delta$  T cells differentially induce maturation, cytokine production, and alloreactive T cell stimulation by dendritic cells and B cells. *Front Immunol.* (2014) 5:650. doi: 10.3389/fimmu.2014.00650
  24. Castella B, Foglietta M, Sciancalepore P, Rigoni M, Coscia M, Griggio V, et al. Anergic bone marrow V9V2 T cells as early and long-lasting markers of PD-1-targetable microenvironment-induced immune suppression in human myeloma. *Oncoimmunology* (2015) 4:e1047580. doi: 10.1080/2162402X.2015.1047580
  25. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer* (2012) 12:252–64. doi: 10.1038/nrc3239
  26. Liu J, Hamrouni A, Wolowicz D, Coiteux V, Kuliczowski K, Hetuin D, et al. Plasma cells from multiple myeloma patients express B7-H1 (PD-L1) and increase expression after stimulation with IFN- $\gamma$  and TLR ligands via a MyD88-, TRAF6-, and MEK-dependent pathway. *Blood* (2007) 110:296–304. doi: 10.1182/blood-2006-10-051482
  27. Paiva B, Azpilikueta A, Puig N, Ocio EM, Sharma R, Oyajobi BO, et al. PD-L1/PD-1 presence in the tumor microenvironment and activity of PD-1 blockade in multiple myeloma. *Leukemia* (2015) 29:2110–3. doi: 10.1038/leu.2015.79
  28. Görgün G, Samur MK, Cowens KB, Paula S, Bianchi G, Anderson JE, et al. Lenalidomide enhances immune checkpoint blockade-induced immune response in multiple myeloma. *Clin Cancer Res.* (2015) 21:4607–18. doi: 10.1158/1078-0432.CCR-15-0200
  29. Rosenblatt J, Glotzbecker B, Mills H, Vasir B, Tzachanis D, Levine JD, et al. PD-1 blockade by CT-011, anti-PD-1 antibody, enhances ex vivo T-cell responses to autologous dendritic cell/myeloma fusion vaccine. *J Immunother.* (2011) 34:409–18. doi: 10.1097/CJI.0b013e31821ca6ce
  30. Benson DM Jr, Bakan CE, Mishra A, Hofmeister CC, Efebera Y, Becknell B, et al. The PD-1/PD-L1 axis modulates the natural killer cell versus multiple myeloma effect: a therapeutic target for CT-011, a novel monoclonal anti-PD-1 antibody. *Blood* (2010) 116:2286–94. doi: 10.1182/blood-2010-02-271874
  31. Koyama S, Akbay EA, Li YY, Herter-Sprie GS, Buczkowski KA, Richards WG, et al. Adaptive resistance to therapeutic PD-1 blockade is associated with upregulation of alternative immune checkpoints. *Nat Commun* (2016) 7:10501. doi: 10.1038/ncomms10501
  32. Hallett WH, Jing W, Drobyski WR, Johnson BD. Immunosuppressive effects of multiple myeloma are overcome by PD-L1 blockade. *Biol Blood Marrow Transplant.* (2011) 17:1133–45. doi: 10.1016/j.bbmt.2011.03.011
  33. Ray A, Das DS, Song Y, Richardson P, Munshi NC, Chauhan D, et al. Targeting PD1-PD-L1 immune checkpoint in plasmacytoid dendritic cell interactions with T cells, natural killer cells and multiple myeloma cells. *Leukemia* (2015) 29:1441–4. doi: 10.1038/leu.2015.11
  34. Lesokhin AM, Ansell SM, Armand P, Scott EC, Halwani A, Gutierrez M, et al. Nivolumab in patients with relapsed or refractory hematologic malignancy: preliminary results of a phase Ib Study. *J Clin Oncol.* (2016) 34:2698–704. doi: 10.1200/JCO.2015.65.9789
  35. Suen H, Brown R, Yang S, Ho PJ, Gibson J, Joshua D. The failure of immune checkpoint blockade in multiple myeloma with PD-1 inhibitors in a phase I study. *Leukemia* (2015) 29:1621–2. doi: 10.1038/leu.2015.104
  36. Rosenblatt J, Avigan D. Targeting the PD-1/PD-L1 axis in multiple myeloma: a dream or a reality? *Blood* (2017) 129:275–79. doi: 10.1182/blood-2016-08-731885
  37. Giuliani M, Janji B, Berchem G. Activation of NK cells and disruption of PD-L1/PD-1 axis: two different ways for lenalidomide to block myeloma progression. *Oncotarget* (2017) 8:24031–44. doi: 10.18632/oncotarget.15234
  38. Bezman NA, Jhatakia A, Kearney AY, Brender T, Maurer M, Henning K et al. PD-1 blockade enhances elotuzumab efficacy in mouse tumor models. *Blood Adv.* (2017) 1:753–65. doi: 10.1182/bloodadvances.2017004382
  39. Tremblay-LeMay R, Rastgoo N, Chang H. Modulating PD-L1 expression in multiple myeloma: an alternative strategy to target the PD-1/PD-L1 pathway. *J Hematol Oncol.* (2018) 11:46. doi: 10.1186/s13045-018-0589-1
  40. Binder DC, Fu YX, Weichselbaum RR. Radiotherapy and immune checkpoint blockade: potential interactions and future directions. *Trends Mol Med.* (2015) 463–5. doi: 10.1016/j.molmed.2015.05.007
  41. Kumar S. Emerging options in multiple myeloma: targeted, immune, and epigenetic therapies. *Hematology Am Soc Hematol Educ Program.* (2017) 2017:518–24. doi: 10.1182/asheducation-2017.1.518
  42. Köhler M, Greil C, Hudecek M, Lonial S, Raje N, Wäsch R, et al. Current developments in immunotherapy in the treatment of multiple myeloma. *Cancer* (2018) 124:2075–85. doi: 10.1002/cncr.31243
  43. Lokhorst HM, Plesner T, Laubach JP, Nahi H, Gimsing P, Hansson M, et al. Targeting CD38 with Daratumumab Monotherapy in Multiple Myeloma. *N Engl J Med.* (2015) 373:1207–19. doi: 10.1056/NEJMoa1506348
  44. Vallet S, Pecherstorfer M, Podar K. Adoptive cell therapy in multiple Myeloma. *Expert Opin Biol Ther.* (2017) 17:1511–22. doi: 10.1080/14712598.2017.1375095
  45. Danhof S, Hudecek M, Smith EL. CARs and other T cell therapies for MM: The clinical experience. *Best Pract Res Clin Haematol.* (2018) 31:147–57. doi: 10.1016/j.beha.2018.03.002
  46. Ali SA, Shi V, Maric I, Wang M, Stroncek DF, Rose JJ, et al. T cells expressing an anti-B-cell 460 maturation antigen chimeric antigen receptor cause remissions of multiple myeloma. *Blood* (2016) 128:1688–700. doi: 10.1182/blood-2016-04-711903
  47. Cohen AD, Garfall AL, Stadtmauer EA, Lacey SE, Lancaster E, Vogl DT et al. B-Cell Maturation Antigen (BCMA)-Specific Chimeric Antigen Receptor T Cells (CART-BCMA) for Multiple Myeloma (MM): initial safety and efficacy from a phase I study. *Blood* (2016) 128:1147.
  48. Fan F (X), Zhao W, Liu J, He A, Chen Y, Cao X, et al. Durable remissions with BCMA-specific chimeric antigen receptor (CAR)-modified T cells in patients with refractory/relapsed multiple myeloma. *J Clin Oncol.* (2017) 35:abstr LBA3001. doi: 10.1200/JCO.2017.35.18\_suppl.LBA3001
  49. Berdeja JG, Yi Lin, Raje NS, DiCapua Siegel DS, Munshi NC, Liedtke M, et al. First-in-human multicenter study of bb2121 anti-BCMA CAR T-cell therapy for relapsed/refractory multiple myeloma: updated results. *J Clin Oncol.* (2017) 15:3010. doi: 10.1200/JCO.2017.35.15\_suppl.3010
  50. Rapoport AP, Stadtmauer EA, Binder-Scholl GK, Goloubeva O, Vogl DT, Lacey SE, et al. NY-ESO-1-engineered T cells mediate sustained antigen-specific chimeric antigen receptor engineered T cells for the treatment of antitumor effects in myeloma. *Nat Med.* (2015) 21:914–21. doi: 10.1038/nm.3910

51. Amos SM, Duong CP, Westwood JA, Ritchie DS, Junghans RP, Darcy PK, et al. Autoimmunity associated with immunotherapy of cancer. *Blood* (2011) 118:499–509 doi: 10.1182/blood-2011-01-325266
52. Caballero OL, Chen YT. Cancer-testis (CT) antigens: potential targets for immunotherapy. *Cancer Sci.* (2009) 100:2014–21. doi: 10.1111/j.1349-7006.2009.01303.x
53. van der Veken LT, Hagedoorn RS, van Loenen MM, Willemze R, Falkenburg JH, Heemskerk MH.  $\alpha\beta$  T-cell receptor engineered  $\gamma\delta$  T cells mediate effective antileukemic reactivity. *Cancer Res.* (2006) 66:3331–7. doi: 10.1158/0008-5472.CAN-05-4190
54. Rischer M, Pscherer S, Duwe S, Vormoor J, Jurgens H, Rossig C. Human gammadelta T cells as mediators of chimaeric-receptor redirected anti-tumour immunity. *Br J Haematol.* (2004) 126:583–92. doi: 10.1111/j.1365-2141.2004.05077.x
55. Deniger DC, Switzer K, Mi T, Maiti S, Hurton L, Singh H, et al. Bispecific T-cells expressing polyclonal repertoire of endogenous  $\gamma\delta$  T-cell receptors and introduced CD19-specific chimeric antigen receptor. *Mol Ther.* (2013) 21:638–47. doi: 10.1038/mt.2012.267
56. Du SH, Li Z, Chen C, Tan WK, Chi Z, Kwang TW, et al. Co-expansion of cytokine-induced killer cells and V $\gamma$ 9V $\delta$ 2 T cells for CAR T-cell therapy. *PLoS ONE* (2016) 11:e0161820. doi: 10.1371/journal.pone.0161820
57. Capsomidis A, Benthall G, Van Acker HH, Fisher J, Kramer AM, Abeln Z, et al. Chimeric antigen receptor-engineered human gamma delta T cells: enhanced cytotoxicity with retention of cross presentation. *Mol Ther.* (2018) 26:354–65. doi: 10.1016/j.ymthe.2017.12.001
58. Fisher J, Wisidagamage Don ND, Flutter B, Capsomidis A, Cheung GW, Gustafsson K, Anderson J. Avoidance of on-target off-tumor activation using a co-stimulation-only chimeric antigen receptor. *Mol Ther.* (2017) 25:1234–47. doi: 10.1016/j.ymthe.2017.03.002
59. van der Veken LT, Coccoris M, Swart E, Falkenburg JH, Schumacher TN, Heemskerk MH.  $\alpha\beta$  T cell receptor transfer to  $\gamma\delta$  T cells generates functional effector cells without mixed TCR dimers *in vivo*. *J Immunol.* (2009) 182:164–70. doi: 10.4049/jimmunol.182.1.164
60. Marcu-Malina V, Heijhuurs S, van Buuren M, Hartkamp L, Strand S, Sebastyen Z, et al. Redirecting  $\alpha\beta$  T cells against cancer cells by transfer of a broadly tumor-reactive  $\gamma\delta$ T-cell receptor. *Blood* (2011) 118:50–9. doi: 10.1182/blood-2010-12-325993
61. Braham MVJ, Minnema MC, Aarts T, Sebastyen Z, Straetmans T, Vyborova A, et al. Cellular immunotherapy on primary multiple myeloma expanded in a 3D bone marrow niche model. *Oncoimmunology* (2018) 7:e1434465. doi: 10.1080/2162402X.2018.1434465
62. Oberg HH, Kellner C, Gonnermann D, Sebens S, Bauerschlag D, Gramatzki M, et al. Tribody [(her2)2xcD16] is More effective Than Trastuzumab in enhancing  $\gamma\delta$  T cell and natural Killer cell cytotoxicity against her2-expressing cancer cells. *Front Immunol.* (2018) 9:814. doi: 10.3389/fimmu.2018.00814
63. Meuter S, Eberl M, Moser B. Prolonged antigen survival and cytosolic export in cross-presenting human gammadelta T cells. *Proc Natl Acad Sci USA.* (2010) 107:8730–5. doi: 10.1073/pnas.1002769107
64. Khan MWA, Eberl M, Moser B. Potential use of V9V2 T cells-based vaccines in cancer immunotherapy. *Front Immunol.* (2014) 5 :512. doi: 10.3389/fimmu.2014.00512
65. van Beek JJ, Wimmers F, Hato SV, de Vries IJ, Sködl AE. Dendritic cell cross talk with innate and innate-like effector cells in antitumor immunity: implications for DC vaccination. *Crit Rev Immunol.* (2014) 34:517–36.
66. de Witte MA, Sarhan D, Davis Z, Felices M, Vallera DA, Hinderlie P, et al. Early Reconstitution of NK and  $\gamma\delta$  T Cells and Its Implication for the Design of Post-Transplant Immunotherapy. *Biol Blood Marrow Transplant.* (2018) 24:1152–62. doi: 10.1016/j.bbmt.2018.02.023
67. Minculescu L, Sengeløv H. The role of gamma delta T cells in haematopoietic stem cell transplantation. *Scand J Immunol.* (2015) 81:459–68. doi: 10.1111/sji.12289
68. Scheper WJ, Gründer C, Straetmans T, Sebastyen ZI, Kuball JI. Hunting for clinical translation with innate-like immune cells and their receptors. *Leukemia* (2014) 28:1181–90. doi: 10.1038/leu.2013.37
69. Irma Airolidi, Alice Bertaina, Ignazia Prigione, Alessia Zorzoli, Daria Pagliara, Claudia Cocco, et al. gd T-cell reconstitution after HLA-haploidentical hematopoietic transplantation depleted of TCR-ab1/CD191 lymphocytes. *Blood* (2015) 125:2349–58 doi: 10.1182/blood-2014-09-599423
70. Lamb LS, Pillai S, Langford S, Bowersock J, Stasi AD, Saad A. Clinical-scale manufacturing of  $\gamma\delta$  T cells for protection against infection and disease recurrence following haploidentical peripheral blood stem cell transplantation and cyclophosphamide gvhd prophylaxis. *Bone Marrow Transplant.* (2018) 53:766–9. doi: 10.1038/s41409-018-0130-8
71. Hagan CT, Medik YB, Wang AZ. Nanotechnology approaches to improving cancer immunotherapy. *Adv Cancer Res.* (2018) 139:35–56. doi: 10.1016/bs.acr.2018.05.003
72. Chabannon C, Kuball J, Bondanza A, Dazzi F, Pedrazzoli P, Toubert A, et al. Hematopoietic stem cell transplantation in its 60s: a platform for cellular therapies. *Sci Transl Med.* (2018) 10:eap9630. doi: 10.1126/scitranslmed.aap9630
73. M Smetak, B Kimmel, J Birkmann, K Schaefer-Eckart, H Einsele, M Wilhelm, Kunzmann V. Clinical-scale single-step CD4+ and CD8+ cell depletion for donor innate lymphocyte infusion (DILI). *Bone Marrow Transplant.* (2008) 41:643–50. doi: 10.1038/sj.bmt.1705942

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Castella, Melaccio, Foglietta, Riganti and Massaia. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.