



The Role of Macrophages in Oral Squamous Cell Carcinoma

Eleni Marina Kalogirou¹, Konstantinos I. Tosios¹ and Panagiotis F. Christopoulos^{2*}

¹ Department of Oral Medicine and Pathology, Faculty of Dentistry, National and Kapodistrian University of Athens, Athens, Greece, ² Department of Pathology, Oslo University Hospital and University of Oslo, Oslo, Norway

OPEN ACCESS

Edited by:

Amanda Psyrris,
University General Hospital
Attikon, Greece

Reviewed by:

Thorsten Fuereeder,
Medical University of Vienna, Austria
Sergios Doumas,
University Hospital of Larissa, Greece

*Correspondence:

Panagiotis F. Christopoulos
christopoulos.panagiotis
@rr-research.no

Specialty section:

This article was submitted to
Head and Neck Cancer,
a section of the journal
Frontiers in Oncology

Received: 08 October 2020

Accepted: 22 February 2021

Published: 18 March 2021

Citation:

Kalogirou EM, Tosios KI and
Christopoulos PF (2021) The Role of
Macrophages in Oral Squamous Cell
Carcinoma. *Front. Oncol.* 11:611115.
doi: 10.3389/fonc.2021.611115

Oral cancer is a common malignancy worldwide, with high disease-related death rates. Oral squamous cell carcinoma (OSCC) accounts for more than 90% of oral tumors, with surgical management remaining the treatment of choice. However, advanced and metastatic OSCC is still incurable. Thus, emphasis has been given lately in understanding the complex role of the oral tumor microenvironment (TME) in OSCC progression, in order to identify novel prognostic biomarkers and therapeutic targets. Tumor associated macrophages (TAMs) constitute a major population of the OSCC TME, with bipolar role in disease progression depending on their activation status (M1 vs. M2). Here, we provide an up to date review of the current literature on the role of macrophages during oral oncogenesis, as well as their prognostic significance in OSCC survival and response to standard treatment regimens. Finally, we discuss novel concepts regarding the potential use of macrophages as targets for OSCC immunotherapeutics and suggest future directions in the field.

Keywords: oral cancer, oral squamous cell carcinoma, tumor-associated macrophages (TAMs), M1-M2 phenotype, classical activation, macrophage polarization

INTRODUCTION

The global incidence of oral cancer was 377,713 new cases during 2020 (1), with a tendency for increasing occurrence even in countries where it is less common, such as Finland and Denmark (2, 3). More than 90% of oral malignancies are oral squamous cell carcinomas (OSCCs), mostly that of the mobile part of the tongue (4, 5). Surgical management is the treatment of choice for OSCC, occasionally supplemented by radiotherapy or chemotherapy. Despite the advances in treatment modalities for solid tumors, the 5-year survival rate is 50% (4, 5), establishing OSCC as the 7th and 10th cause of death for males and females, respectively, in Europe (6).

Prognosis of OSCC depends on various factors associated with patients' profile, i.e., ethnicity, gender, age, socioeconomic status, and lifestyle, mainly including smoking and alcohol consumption, as well as tumor's characteristics, i.e., site, size, regional or distant metastases, stage, depth of invasion and degree of differentiation (5). Most research efforts focus on the identification of novel biomarkers in blood, saliva, or tumor tissue samples that could facilitate early diagnosis and group patients into subpopulations with more predictable responses to certain treatment schemes (5, 7). Such biomarkers might be directly related to tumor cells *per se*, or concern cellular and/or other components of the surrounding tumor microenvironment (TME) (8). TME provides the essential requirements for cancer survival, growth and invasion (8) and macrophages are a key population of it (9).

We review the literature on the role of macrophages in OSCC, focusing on their pro-tumor and anti-tumor properties, as well as their prognostic significance for patients' survival and response to standard treatment regimens. We, also, discuss novel concepts regarding the potential use of macrophages as candidates for OSCC immunotherapy and suggest future directions in the field.

THE TUMOR MICROENVIRONMENT OF ORAL CANCER

The progression of OSCC depends on the interplay among the cancer cells, the surrounding host-derived stromal cells, e.g., fibroblasts, endothelial cells and pericytes, and the extracellular matrix non-cellular components composing the TME (10). Immune cells may have opposing functions in oral oncogenesis, i.e., a class of macrophages, myeloid-derived suppressor cells, regulatory T cells and CD4+ T helper type 2 (Th2) cells may share pro-tumor functions, whereas another class of macrophages, dendritic cells, natural killer cells, CD8+ T cells and CD4+ Th1 cells may have anti-tumor actions (11). Historically solid tumors are immunologically classified into "cold" and "hot," depending on the non-inflamed or inflamed TME milieu, respectively (12, 13). Inflamed tumors respond favorably to immune checkpoint blockade (ICB) therapy and are characterized by an abundance of tumor infiltrated lymphocytes (TILs) enriched for interferon- γ (IFN- γ)-expressing CD8+ T cells, expression of checkpoint markers including programmed death-1 ligand 1 (PD-L1) and high mutational burden. In contrast, non-inflamed tumors are poorly infiltrated by immune cells and rarely express PD-L1, while characterized by an immunosuppressive milieu, though they might also have high genomic instability. On the very end of the tumor immunity continuum are immunologically ignorant tumors, characterized by genomic stability, highly proliferative tumor cells and low infiltration of T cells, as well as low expression of antigen-presentation machinery markers including MHCI (14). Immunogenic tumors, characterized by the successful recognition of cancer antigens, vary (15, 16), but the presence of macrophages in the TME is independent of the tumor's immunogenicity status.

The interplay between the innate and the acquired immune system is responsible for the recognition of cancer

specific antigens as foreign antigens, a process termed *immunosurveillance* that eliminates cancer cells (17). Besides its natural protective role against tumors, the identification of a tumor promoting role of the immune system, mainly via clonal selection (those with low immunogenicity), has led to the refinement of the initial immunosurveillance hypothesis (18, 19) with that of cancer immunoediting (20, 21). This process consists of three phases, known as the 3Es of cancer immunoediting; elimination, equilibrium and escape. The elimination phase represents the original immunosurveillance concept and raises as an integrated and combined response of both the innate and adaptive immune system against developing tumors. Complete eradication of tumor cells at this stage means that there is not progression to the following two phases and thus homeostasis has achieved. If that's not the case then in the second phase, which is most probably the longest of the three (may last for years in humans) and the direct equivalent of a Darwinian selection process, the tumor cell variants that had survived the first phase enter a dynamic equilibrium with the immune system, which is able to contain but not completely vanish the tumor variants. The result of this sculpturing is the generation of heterogeneous and genetically unstable tumor clones with increased resistance to immune recognition. In the last phase, the selected tumor cell variants from the equilibrium phase can grow and expand to clinically detectable levels. Even though various genetic events and epigenetic modifications, as well as different direct and indirect mechanisms may be employed by cancer cells, the end result is the tumor escape from host's immune defenses (22, 23). In the first line of immune response are professional antigen presenting cells, e.g., dendritic cells, B cells and macrophages. Macrophages belong to the mononuclear phagocytic system and are the final differentiation stage of circulating monocytes that have been attracted by chemotactic factors to the tissue site, as a response to various microenvironmental stimuli (24, 25). Tissue resident macrophages originate from yolk sac during embryonic development, whereas monocyte-derived macrophages from bone marrow progenitor cells (26). *Tumor-associated macrophages* (TAMs) constitute the major leucocytic component of the OSCC TME (27).

TAMs may have promoting or inhibitory effects on OSCC cancer cells proliferation, invasion and migration (10). They are directed to the tumor site by chemotactic cues produced by cancer cells or stromal cells, including vascular endothelial growth factor (VEGF), colony stimulating factor-1 (CSF-1), placental growth factor, and chemokines, such as the chemokine (C-C motif) ligand (CCL) 2/monocyte chemotactic protein-1, CCL3/macrophage inflammatory protein-1alpha, CCL4/macrophage inflammatory protein-1beta, and the CCL5/regulated on activation, normal T-cell expressed and secreted (RANTES) (28, 29). TAMs are usually more prevalent in tumor stroma than in OSCC cell nests (30).

Initially, TAMs recruited in the hypoxic TME environment may show an anti-tumor activity, e.g., via the tumor-antigen presentation to T cells and the induction of the T-cell mediated cancer cytotoxicity (31). TAMs have been shown to be effective in inducing the cytotoxic activation of naive (CD45RO) or memory T cell (CD45RA) against *Streptococcus salivarius*,

Abbreviations: Akt, Protein kinase B; CAR, chimeric antigen receptor; CAR-M, CAR engineered macrophages; CCL, chemokine (C-C motif) ligand; CD, cluster of differentiation; CSF-1, colony stimulating factor-1; CXCL, C-X-C Motif Chemokine Ligand; EGF, epidermal growth factor; GLI-1, glioma-associated oncogene homolog 1; ICB, immune checkpoint blockade; IFN- γ , interferon- γ ; IL, interleukin; iNOS, inducible nitric oxide synthase; MHCI, Major Histocompatibility Complex (MHC) Class I; MIF, migration inhibitor factor; MMP, matrix metalloproteinase; NF-kB, nuclear factor kappa beta; NO, nitric oxide; OSCC, oral squamous cell carcinoma; PD-L1, programmed death-1 ligand 1; PI3K, phosphatidylinositol 3-kinase; RACK1, receptor for activated C kinase 1; SIRP α , Signal regulatory protein α ; SOCS1, suppressor of cytokine signaling 1; STAT6, signal transduction and transcriptional activator 6; TAMs, tumor-associated macrophages; TGF- β , transforming growth factor beta; T β RII, type II TGF- β receptor; Th, T helper type; TILs, tumor infiltrated lymphocytes; TLR, toll-like receptor; TME, tumor microenvironment; TNF- α , tumor necrosis factor- α ; VEGF, vascular endothelial growth factor.

while their depletion caused decreased levels of granzyme B expressed by CD8+ T cells (32). On the other hand, TAMs may be a source of proangiogenic molecules and growth factors promoting cell proliferation, such as VEGF, epidermal growth factor (EGF), fibroblast growth factor, platelet-derived growth factor, transforming growth factor beta (TGF- β) and CCL2, as well as matrix metalloproteinases (MMPs), responsible for the degradation of the basement membrane and extracellular matrix, thus facilitating the progression of OSCC (28, 29, 33). The EGF-mediated pro-tumor role of TAMs in OSCC has been highlighted in a study showing increased cell proliferation and invasion of OSCC cells co-cultured with TAMs-derived conditioned medium, while the proliferation and invasion activity was hampered after cancer cell treatments with an anti-EGF receptor antibody (34). In another study, TAMs-induced progression of OSCC was associated with promotion of epithelial-to-mesenchymal transition of OSCC cells, as a fibroblast-like phenotype was observed in an OSCC and two head and neck cell lines following co-culture with TAMs conditioned medium (35).

A critical factor defining the pro-tumor or anti-tumor properties of TAMs is their activation state.

ACTIVATION STATES OF MACROPHAGES

Besides their antigen-presentation capabilities, macrophages possess various other properties necessary to maintain homeostasis, e.g., phagocytosis of pathogens and cell debris, destruction of antibody-coated cells or microbes, secretion of growth factors, cytokines and chemokines, expression of membranous co-stimulatory or co-inhibitory molecules, and wound healing via matrix destruction, regeneration and angiogenesis (36, 37). The wide and contradictory functions of macrophages are represented by the two-edged model of M1 or M2 macrophages, associated with Th1 or Th2 responses, respectively (25, 38). Metabolic events seem to drive the polarization fate of macrophages (39); the differential metabolism of L-arginine, either via inducible nitric oxide synthase (iNOS) to NO and citruline, or via arginase to ornithine and urea, defines the orientation toward M1 or M2 activation, respectively (40).

The M1 macrophage phenotype arises as the consequence of classical activation stimulated by bacterial components, e.g., lipopolysaccharides, or by the Th1-driven cytokine IFN- γ (10, 41). M1 macrophages possess enhanced antigen presentation properties and lysosome activity, favoring Th1 responses (42). Although there is evidence to the contrary (43), induction of the M1 macrophage activation phenotype is considered beneficial against cancer. M1 exert an anti-tumor effect via direct cancer cytotoxicity through production of reactive oxygen and nitrogen species and/or indirectly via the production of pro-inflammatory cytokines, e.g. interleukin (IL)-12, IL-23 and tumor necrosis factor- α (TNF α), and chemokines [e.g. CCL5, C-X-C Motif Chemokine Ligand (CXCL)5, CXCL9, and CXCL10] and activation of other effector cells (10). Of note, the macrophage-mediated NO-cytotoxicity of cancer cells is enhanced by close proximity and cell contact of macrophages to cancer cells

via induction of apoptotic mechanisms to the latter, but is independent of the cancer-antigen recognition (44), suggesting the immunotherapeutic potentials of macrophages in “cold” tumors. However, the role of classical activated macrophages in tumor cell-specific responses via cytotoxic T cells is equally important, as the levels of the CD8+ T cell-expressing granzyme B were significantly higher in the presence of M1- than M2-polarized TAMs in patients with OSCC (32).

The alternative polarization, induced by Th2-derived cytokines (e.g., IL-4, IL-10, IL-13) that hamper the production of pro-inflammatory cytokines such as TNF- α and IFN- γ , gives rise to the pro-tumor M2 macrophage phenotype (10, 25, 45, 46). M2 macrophages produce anti-inflammatory cytokines, such as IL-1 receptor antagonist, IL-10 and TGF- β , promote the immunosuppressive functions of Foxp3+ regulatory T cells, and enhance the expression of proangiogenic factors (e.g., VEGF) and proteolytic enzymes (e.g., MMPs). Those effects result in the inhibition of anti-tumor immunity mediated by Th1 cells and cytotoxic T cells and, finally, to the escape from immune surveillance (10, 41, 47). The immune escape mechanisms of OSCC might be promoted by the infiltrating TAMs, as a positive correlation has been observed between the levels of CD68+ and CD163+ TAMs and the OSCC cells that expressed the checkpoint PD-L1 protein (48). The simultaneous secretion of IL-10 and expression of the PD-L1 by TAMs has been suggested as a potential mechanism for CD3+ T cell negative regulation by TAMs, identified by the expression of the M2-related markers CD163 and CD204 in OSCC patients (49). M2-polarized TAMs in close proximity of OSCC cells can induce the migration and invasion of the latter, via activation of NF- κ B and favor the production of growth factors (e.g., EGF and TGF- β) that promote tumor progression (50). In addition, the expression of the proteins Sonic Hedgehog, Indian Hedgehog and glioma-associated oncogene homolog 1 (GLI-1) in tumor cells and endothelial cells in the OSCC TME, along with the co-expression of Indian Hedgehog ligand in CD163+ TAMs, may be indicative of the role of Hedgehog pathway in promoting neovascularization in OSCC (51).

TAMs are characterized by striking plasticity and except for the M1 and M2 end-stage polarization they may acquire various intermediate activation states, by simultaneously expressing markers related to both M1 (e.g., elevated levels of TNF- α , MMP9, CCL2, CCL5, CXCL9, CXCL10, and CXCL16), and M2 (e.g., increased levels of IL-10, arginase-1, and peroxisome proliferator-activated receptor γ) phenotype (10). As within most solid tumors, the macrophage balance in OSCC tends toward the M2 phenotype (41, 47, 52–54). Co-culture of a monocyte/macrophage like cell line (RAW264.7) with conditioned medium derived from OSCC cell lines stimulated the expression of pro-tumor cytokines and chemokines associated with the M2 phenotype, e.g., IL-10, CCL22, and VEGF-A (54). The receptor for activated C kinase 1 (RACK1) has also been shown to induce OSCC progression, by favoring the polarization toward the M2 phenotype and reducing the levels of M1-phenotype related molecules, such as IL-6, CCL5, and CSF, in an NF- κ B axis-dependent manner (55). TAMs-induced VEGF expression in OSCC has also been associated

with activation of the TGF- β 1/T β RII/Smad3 signaling pathway (56). Okubo et al. (57) developed a xenograft mouse model using a human tongue squamous cell carcinoma (SCC) cell line and reported that the irradiation-induced tumor infiltration by CD11b+ bone marrow-derived cells that acquired an M2-like phenotype, promoted tumor re-vascularization, progression and recurrence after radiotherapy. The potential mechanisms of M2-polarization of TAMs in OSCC also include the OSCC-derived exosomes enclosing micro-RNAs (miR-29a-3p) regulating the activity of SOCS1/STAT6 signals (58), as well as activation of the Axl/PI3K/Akt/NF- κ B signaling pathway (52).

The predominance of M1 or M2 phenotype in different subpopulations of OSCC patients has been significantly correlated with the disease outcome (31).

PROGNOSTIC SIGNIFICANCE OF MACROPHAGES IN DISEASE OUTCOME

The prognostic significance of TAMs in OSCC has predominantly been evaluated by immunohistochemistry, where the presence of TAMs was correlated with survival parameters, either directly with overall survival and progression/disease/recurrence free survival, or indirectly with clinicopathological factors associated with disease outcome (31, 59). Several TAMs markers have been used, such as the pan-macrophage marker CD68 (both M1 and M2) (30, 35, 47, 54, 56, 60–66), CD204, expressed by dendritic cells and macrophages (54), and markers highly expressed -but not restricted to- M1 (CD11c) (55) or M2 [CD163 (7, 35, 41, 42, 54, 60, 61, 67), CD206 (34, 55)] phenotype.

A growing body of evidence supports the adverse prognostic role of CD68+ TAMs for OSCC patients overall survival (30, 35, 56, 62, 64) or disease-free survival (30, 64, 66). Moreover, elevated numbers of CD68+ TAMs have significantly been associated with high grade (54, 56), increased size (63, 64) and advanced stage (30, 56, 60, 63, 64), lymph node (30, 35, 54, 61, 63, 64) or distant metastasis (47), or high recurrence rate of OSCC (64). In contrast, Wei et al. (65) found that lower mean CD68+ TAMs density was significantly associated with lower differentiation grade and higher clinical stages of OSCC, as well as shorter 5-year survival rate. The latter contrasting findings might be attributed to CD68 staining of both M1 and M2 TAMs, and emphasized the need to identify more specific markers for M1 and M2 subpopulations (65).

Increased levels of CD163+ TAMs, currently considered as an M2-preferable marker, have been correlated with worst overall survival (7, 35, 42, 60, 61, 67) or disease-free survival (7, 67), as well as more frequent recurrence (35) of OSCC. A higher number of CD163+ TAMs has, also, been associated with OSCC poor differentiation (41) and positive lymph nodes (7, 42, 54, 61). A mechanistic explanation for the latter finding was suggested by Yagamoto et al. (54), who reported a significant correlation between the number of CD163+ M2 TAMs and the lymphatic vessel density in the TME of OSCC tissue samples, while immunofluorescence analysis revealed the co-expression of VEGF-C in CD163+ TAMs. These results indicated that M2

polarized TAMs may induce lymphangiogenesis through VEGF-C, eventually promoting lymph node metastasis in OSCC (54). Low expression of CD163+ TAMs has also been reported as a negative prognostic factor in OSCC cases presenting high IL-10 and low IFN- γ expression (42). High levels of CD206+ (M2-associated) TAMs have also been correlated with advanced stage, increased tumor size, lymph node metastasis, as well as short disease-free survival (34). Finally, the high M2/M1 ratio, represented by CD206+/CD11c TAMs, showed a significant association with poor disease outcome (55).

The prognostic effects of TAMs in OSCC have, also, been assessed indirectly, through the expression levels of molecules usually secreted by or associated with macrophages. A higher number of CCL2+ cells in the TME, which were found co-expressed in CD163+ TAMs, has been associated with lower 5-year overall survival rates (68). The levels of macrophage migration inhibitor factor (MIF) in OSCC tissues have been adversely correlated with overall and recurrence-free survival, while increased MIF serum concentration is considered indicative for early recurrence (69, 70).

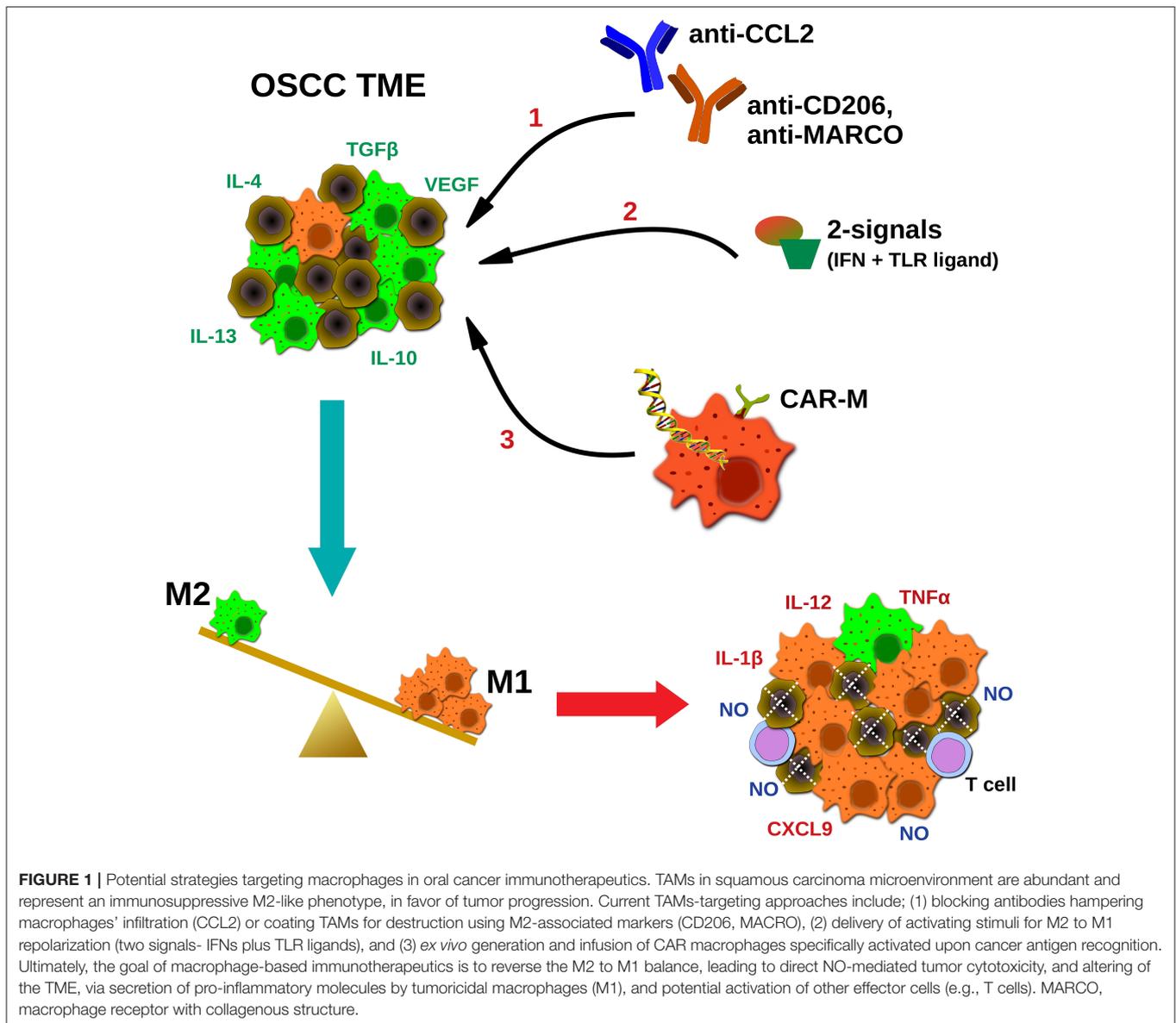
Other immunohistochemical studies have reported a significantly higher expression of CD68+ TAMs (62, 64) or CD163+ M2-polarized macrophages (42, 71) in OSCC compared to oral potentially malignant lesions, e.g., epithelial dysplasia/hyperplasia. TAM markers' expression showed a progressive increase from hyperplasia to low and high degree of dysplasia (64, 72), indicating a possible role of macrophages in promoting the evolution of oral premalignant lesions to invasive OSCC. Finally, significantly increased levels of CD68+ TAMs have been reported in OSCC compared to verrucous carcinoma (73), which is considered less aggressive (74).

In conclusion, an increased immunohistochemical expression of TAMs, predominantly those of M2 polarization, has been associated with adverse OSCC prognosis.

TARGETING MACROPHAGES IN ORAL CANCER IMMUNOTHERAPEUTICS; LESSONS LEARNED FROM OTHER SOLID TUMOR MODELS

Given the bipolar role of macrophages in tumor development and their tendency to acquire an immunosuppressive M2-like phenotype during disease progression, current macrophage-based immunotherapeutic approaches aim to alter the M1 to M2 balance in the TME, in favor of the M1. For this purpose several strategies have been developed, including; hampering the monocyte infiltration and macrophage differentiation in the tumor sites (e.g., CCL2 or CSF-1 blocking antibodies), deletion of M2 macrophages in the TME (e.g., antibodies against CD206, scavenger receptors or other M2-associated molecules), or repolarization toward the M1 phenotype and enhancement of their anti-tumor properties (e.g., delivery of activating stimuli and/or antibodies inducing macrophage phagocytosis, or *ex vivo* macrophage manipulation) (75–77) (**Figure 1**).

Even though numerous studies point to the strong potentials of macrophage-based immunotherapeutics in various solid



tumors, via regulation of the tumor immunity in different levels [reviewed in DeNardo and Ruffell (39)], the direct evidence in OSCC models is yet very limited. In a study using THP-1 human cell line as monocyte-derived macrophage model, pre-activated toward the M1 (IFN- γ + lipopolysaccharide) or the M2 (IL-4+IL-13) phenotype, it was found that M2 induced the migration and invasion of human tongue SCC cell line HSC-3, when co-cultured with the latter (50). In contrast, using various *in vitro* migration assays, co-cultures with M1 macrophages reduced the invasion of HSC-3 cells, alluding to the anti-tumor properties of this phenotype (50).

Our previous work using primary mouse bone marrow derived macrophages showed that macrophages pre-activated with two simultaneous signals [IFNs plus toll-like receptors (TLRs) ligands] completely blocked the proliferation of Lewis lung carcinoma cell line (LLC), when co-cultured with the

latter. Similar results were shown using a macrophage-like cell line (J774.A1) and the mineral-oil induced plasmacytoma cell line (MOPC315), showing the immunotherapeutic potentials of macrophages in different tumor models (78, 79). This macrophage-mediated growth inhibition was shown to be dependent on NO production, since blocking of the latter rescued cancer cell proliferation. Of note, these anti-tumor macrophages secreted significant amounts of pro-inflammatory factors including IL-12p70, TNF- α and the T cell chemoattractant CXCL9 (78, 79), further enhancing the notion for altering the TME and promoting the activation of other effector cells. In this context, recent elegant studies stratifying cutting edge technologies, point to the tumoricidal potentials of macrophages. The CD47-Signal regulatory protein α (SIRP α) axis has gained great interest in immunotherapeutic approaches lately. The interaction of CD47 (also known as “don’t eat me” signal)

expressed by all cells in the body, with its receptor SIRP α found mainly on macrophages and other phagocytes, hampers the engulfment of the ligand expressing cells (CD47+) by the phagocytes. The upregulation of CD47 is now considered an evade mechanism (innate immune checkpoint) adopted by cancer cells (80, 81). Alvey et al. (82) showed that *ex vivo* manipulated, SIRP α -inhibited macrophages when systemically injected to mice were able to accumulate into the tumor sites (human lung cancer model), leading to tumor cells engorgement and cancer regression. However, this anti-tumor effect lasted for 1-2 weeks, followed by differentiation of the donor macrophages toward non-phagocytic, high SIRP α TAMs, alluding to the high plasticity of macrophages in response to microenvironmental factors and the need for more permanent approaches. A very recent study, stratifying the chimeric antigen receptor (CAR) technology from T cells, showed that human monocyte-derived CAR engineered macrophages (CAR-M), demonstrated antigen-specific phagocytosis *in vitro*, while a single systemic injection of CAR-M in mice models, significantly reduced tumor burden and improved the overall survival (83). Furthermore, CAR-M demonstrated tumor specific localization in various tumors models and longevity in mice. Intriguingly, they also showed outstanding indirect anti-tumor properties including; secretion of pro-inflammatory molecules, resistance to immunosuppressive cytokines (M1 phenotype persistence), conversion of M2 macrophages to M1, as well as recruitment of and presentation of antigens to T cells (83).

Thus, as shown with various other solid tumor models, macrophage-based approaches may hold strong therapeutic potentials. Whether this is also true for OSCC remains to be elucidated.

FUTURE PERSPECTIVES AND CONCLUSIONS

Advanced OSCC remains practically incurable, as this is reflected on the poor 5-year survival and increased death rates. Although

surgical treatment might lead to complete disease management in most of the cases of stage I and II OSCC, cancer recurrence is not rare, while adjuvant chemo-radiotherapy is usually required in advanced stages (4, 5). Given the recent advances in cancer research, the need for implementation of novel diagnostic and treatment tools in its management, has emerged.

TAMs are abundant in the OSCC TME, displaying a bipolar role in disease progression depending on their activation status. A growing body of literature suggests that macrophages may serve as a valid prognostic and therapeutic tool in the arsenal of OSCC treatment modalities. Even though macrophage-based immunotherapeutic research in OSCC is yet inaugural, evidence from various other solid tumor models points to the strong therapeutic potentials of macrophages via inducing direct tumor cytotoxicity and indirectly via altering the immunosuppressive TME and induction of a systemic anti-tumor immunity and activation of other effector cells. Identification of novel M1- and M2- specific markers and further experimentation implying macrophages' tumoricidal activation in OSCC models, might close the gap from bed-to bedside, leading to the clinical implementation of novel macrophage-based strategies, in prognosis and immuno-treatment of this cancer type.

AUTHOR CONTRIBUTIONS

EMK performed the literature research and wrote the manuscript. KT contributed to manuscript writing and formatting. PC conceived the idea, designed the lay out, wrote and reviewed the manuscript. All authors have approved the final draft.

FUNDING

This research was supported by funds provided by the South-Eastern Norway Regional Health Authority, Project Number 2019059 to PC.

REFERENCES

- Global Cancer Observatory: Cancer Today. International Agency for Research (2021). Available online at: <https://gco.iarc.fr/today/fact-sheets-cancers> (accessed February 19, 2021).
- Chi AC, Day TA, Neville BW. Oral cavity and oropharyngeal squamous cell carcinoma—an update. *CA Cancer J Clin.* (2015) 65:401–21. doi: 10.3322/caac.21293
- Warnakulasuriya S. Global epidemiology of oral and oropharyngeal cancer. *Oral Oncol.* (2009) 45:309–16. doi: 10.1016/j.oraloncology.2008.06.002
- García-Martín JM, Varela-Centelles P, González M, Seoane-Romero JM, Seoane J, García-Pola MJ. Epidemiology of oral cancer. In: Panta P, editor. *Oral Cancer Detection*. Cham: Springer. (2019). p. 81–93. doi: 10.1007/978-3-319-61255-3_3
- Warnakulasuriya S, Greenspan JS. Epidemiology of oral and oropharyngeal cancers. In: Warnakulasuriya S, Greenspan JS, editors. *Textbook of Oral Cancer Textbooks in Contemporary Dentistry*. Cham: Springer. (2020). p. 5–21. doi: 10.1007/978-3-030-32316-5_2
- Ferlay J, Steliarova-Foucher E, Lortet-Tieulent J, Rosso S, Coebergh JW, Comber H, et al. Cancer incidence and mortality patterns in Europe: estimates for 40 countries in (2012). *Eur J Cancer.* (2013) 49:1374–403. doi: 10.1016/j.ejca.2012.12.027
- Fujita Y, Okamoto M, Goda H, Tano T, Nakashiro K, Sugita A, et al. Prognostic significance of interleukin-8 and CD163-positive cell-infiltration in tumor tissues in patients with oral squamous cell carcinoma. *PLoS ONE.* (2014) 9:e110378. doi: 10.1371/journal.pone.0110378
- Salo T, Vered M, Bello IO, Nyberg P, Bitu CC, Zlotogorski Hurvitz A, et al. Insights into the role of components of the tumor microenvironment in oral carcinoma call for new therapeutic approaches. *Exp Cell Res.* (2014) 325:58–64. doi: 10.1016/j.yexcr.2013.12.029
- Li Z, Liu FY, Kirkwood KL. The p38/MKP-1 signaling axis in oral cancer: impact of tumor-associated macrophages. *Oral Oncol.* (2020) 103:104591. doi: 10.1016/j.oraloncology.2020.104591
- Peltanova B, Raudenska M, Masarik M. Effect of tumor microenvironment on pathogenesis of the head and neck squamous cell carcinoma: a systematic review. *Mol Cancer.* (2019) 18:63. doi: 10.1186/s12943-019-0983-5
- Mohan SP, Bhaskaran MK, George AL, Thirutheri A, Somasundaran M, Pavithran A. Immunotherapy in oral cancer. *J Pharm Bioallied Sci.* (2019) 11(Suppl 2):S107–11. doi: 10.4103/JPBS.JPBS_31_19

12. Galon J, Bruni D. Approaches to treat immune hot, altered and cold tumours with combination immunotherapies. *Nat Rev Drug Discov.* (2019) 18:197–218. doi: 10.1038/s41573-018-0007-y
13. Gajewski TF. The next hurdle in cancer immunotherapy: overcoming the non-T-cell-inflamed tumor microenvironment. *Semin Oncol.* (2015) 42:663–71. doi: 10.1053/j.seminoncol.2015.05.011
14. Hegde PS, Karanikas V, Evers S. The where, the when, and the how of immune monitoring for cancer immunotherapies in the era of checkpoint inhibition. *Clin Cancer Res.* (2016) 22:1865–74. doi: 10.1158/1078-0432.CCR-15-1507
15. Blankenstein T, Coulie PG, Gilboa E, Jaffee EM. The determinants of tumour immunogenicity. *Nat Rev Cancer.* (2012) 12:307–13. doi: 10.1038/nrc3246
16. Lechner MG, Karimi SS, Barry-Holson K, Angell TE, Murphy KA, Church CH, et al. Immunogenicity of murine solid tumor models as a defining feature of *in vivo* behavior and response to immunotherapy. *J Immunother.* (2013) 36:477–89. doi: 10.1097/01.cji.0000436722.46675.4a
17. Outh-Gauer S, Morini A, Tartour E, Lepine C, Jung AC, Badoual C. The microenvironment of head and neck cancers: papillomavirus involvement and potential impact of immunomodulatory treatments. *Head Neck Pathol.* (2020) 14:330–40. doi: 10.1007/s12105-020-01147-x
18. Burnet FM. The concept of immunological surveillance. *Prog Exp Tumor Res.* (1970) 13:1–27. doi: 10.1159/000386035
19. Thomas L. "Discussion" in *Cellular Humoral Aspects of the Hypersensitive States*, ed. H.S. Lawrence. New York, NY: Hoeber-Harper. (1959). p. 529–532.
20. Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoeediting: from immunosurveillance to tumor escape. *Nat Immunol.* (2002) 3:991–8. doi: 10.1038/nri102-991
21. Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ, et al. IFN γ and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature.* (2001) 410:1107–11. doi: 10.1038/35074122
22. Dunn GP, Old LJ, Schreiber RD. The immunobiology of cancer immunosurveillance and immunoeediting. *Immunity.* (2004) 21:137–48. doi: 10.1016/j.immuni.2004.07.017
23. Dunn GP, Old LJ, Schreiber RD. The three Es of cancer immunoeediting. *Annu Rev Immunol.* (2004) 22:329–60. doi: 10.1146/annurev.immunol.22.012703.104803
24. Mantovani A, Bottazzi B, Colotta F, Sozzani S, Ruco L. The origin and function of tumor-associated macrophages. *Immunol Today.* (1992) 13:265–70. doi: 10.1016/0167-5699(92)90008-U
25. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol.* (2002) 23:549–55. doi: 10.1016/S1471-4906(02)02302-5
26. Shi C, Pamer EG. Monocyte recruitment during infection and inflammation. *Nat Rev Immunol.* (2011) 11:762–74. doi: 10.1038/nri3070
27. Essa AA, Yamazaki M, Maruyama S, Abe T, Babkair H, Raghieb AM, et al. Tumour-associated macrophages are recruited and differentiated in the neoplastic stroma of oral squamous cell carcinoma. *Pathology.* (2016) 48:219–27. doi: 10.1016/j.pathol.2016.02.006
28. Coussens LM, Werb Z. Inflammation and cancer. *Nature.* (2002) 420:860–7. doi: 10.1038/nature01322
29. Mantovani A, Sica A. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr Opin Immunol.* (2010) 22:231–7. doi: 10.1016/j.coi.2010.01.009
30. Ni YH, Ding L, Huang XF, Dong YC, Hu QG, Hou YY. Microlocalization of CD68+ tumor-associated macrophages in tumor stroma correlated with poor clinical outcomes in oral squamous cell carcinoma patients. *Tumour Biol.* (2015) 36:5291–8. doi: 10.1007/s13277-015-3189-5
31. Alves AM, Diel LF, Lamers ML. Macrophages and prognosis of oral squamous cell carcinoma: a systematic review. *J Oral Pathol Med.* (2018) 47:460–7. doi: 10.1111/jop.12643
32. Wang J, Yang L, Mao X, Li Z, Lin X, Jiang C. Streptococcus salivarius-mediated CD8(+) T cell stimulation required antigen presentation by macrophages in oral squamous cell carcinoma. *Exp Cell Res.* (2018) 366:121–6. doi: 10.1016/j.yexcr.2018.03.007
33. Tampa M, Mitran MI, Mitran CI, Sarbu MI, Matei C, Nicolae I, et al. Mediators of inflammation - a potential source of biomarkers in oral squamous cell carcinoma. *J Immunol Res.* (2018) 2018:1061780. doi: 10.1155/2018/1061780
34. Haque A, Moriyama M, Kubota K, Ishiguro N, Sakamoto M, Chinju A, et al. CD206(+) tumor-associated macrophages promote proliferation and invasion in oral squamous cell carcinoma via EGF production. *Sci Rep.* (2019) 9:14611. doi: 10.1038/s41598-019-51149-1
35. Hu Y, He MY, Zhu LF, Yang CC, Zhou ML, Wang Q, et al. Tumor-associated macrophages correlate with the clinicopathological features and poor outcomes via inducing epithelial to mesenchymal transition in oral squamous cell carcinoma. *J Exp Clin Cancer Res.* (2016) 35:12. doi: 10.1186/s13046-015-0281-z
36. Chawla A. Control of macrophage activation and function by PPARs. *Circ Res.* (2010) 106:1559–69. doi: 10.1161/CIRCRESAHA.110.216523
37. Franken L, Schiwon M, Kurts C. Macrophages: sentinels and regulators of the immune system. *Cell Microbiol.* (2016) 18:475–87. doi: 10.1111/cmi.12580
38. Mills CD, Kincaid K, Alt JM, Heilman MJ, Hill AM. M-1/M-2 macrophages and the Th1/Th2 paradigm. *J Immunol.* (2000) 164:6166–73. doi: 10.4049/jimmunol.164.12.6166
39. DeNardo DG, Ruffell B. Macrophages as regulators of tumour immunity and immunotherapy. *Nat Rev Immunol.* (2019) 19:369–82. doi: 10.1038/s41577-019-0127-6
40. Mills CD. Anatomy of a discovery: m1 and m2 macrophages. *Front Immunol.* (2015) 6:212. doi: 10.3389/fimmu.2015.00212
41. Mori K, Hiroi M, Shimada J, Ohmori Y. Infiltration of m2 tumor-associated macrophages in oral squamous cell carcinoma correlates with tumor malignancy. *Cancers.* (2011) 3:3726–39. doi: 10.3390/cancers3043726
42. Wang S, Sun M, Gu C, Wang X, Chen D, Zhao E, et al. Expression of CD163, interleukin-10, and interferon-gamma in oral squamous cell carcinoma: mutual relationships and prognostic implications. *Eur J Oral Sci.* (2014) 122:202–9. doi: 10.1111/eos.12131
43. Xiao M, Zhang J, Chen W, Chen W. M1-like tumor-associated macrophages activated by exosome-transferred THBS1 promote malignant migration in oral squamous cell carcinoma. *J Exp Clin Cancer Res.* (2018) 37:143. doi: 10.1186/s13046-018-0815-2
44. Fauskanger M, Haabeth OAW, Skjeldal FM, Bogen B, Tveita AA. Tumor killing by CD4(+) T cells is mediated via induction of inducible nitric oxide synthase-dependent macrophage cytotoxicity. *Front Immunol.* (2018) 9:1684. doi: 10.3389/fimmu.2018.01684
45. Allavena P, Garlanda C, Borrello MG, Sica A, Mantovani A. Pathways connecting inflammation and cancer. *Curr Opin Genet Dev.* (2008) 18:3–10. doi: 10.1016/j.gde.2008.01.003
46. Gordon S. Alternative activation of macrophages. *Nat Rev Immunol.* (2003) 3:23–35. doi: 10.1038/nri978
47. Costa NL, Valadares MC, Souza PP, Mendonca EF, Oliveira JC, Silva TA, et al. Tumor-associated macrophages and the profile of inflammatory cytokines in oral squamous cell carcinoma. *Oral Oncol.* (2013) 49:216–23. doi: 10.1016/j.oraloncology.2012.09.012
48. Suarez-Sanchez FJ, Lequerica-Fernandez P, Suarez-Canto J, Rodrigo JP, Rodriguez-Santamarta T, Dominguez-Iglesias F, et al. Macrophages in oral carcinomas: relationship with cancer stem cell markers and PD-L1 expression. *Cancers.* (2020) 12(7). doi: 10.3390/cancers12071764
49. Kubota K, Moriyama M, Furukawa S, Rafiul H, Maruse Y, Jinno T, et al. CD163(+)CD204(+) tumor-associated macrophages contribute to T cell regulation via interleukin-10 and PD-L1 production in oral squamous cell carcinoma. *Sci Rep.* (2017) 7:1755. doi: 10.1038/s41598-017-01661-z
50. Pirila E, Vayrynen O, Sundquist E, Pakkila K, Nyberg P, Nurmenniemi S, et al. Macrophages modulate migration and invasion of human tongue squamous cell carcinoma. *PLoS ONE.* (2015) 10:e0120895. doi: 10.1371/journal.pone.0120895
51. Valverde Lde F, Pereira Tde A, Dias RB, Guimaraes VS, Ramos EA, Santos JN, et al. Macrophages and endothelial cells orchestrate tumor-associated angiogenesis in oral cancer via hedgehog pathway activation. *Tumour Biol.* (2016) 37:9233–41. doi: 10.1007/s13277-015-4763-6
52. Chiu KC, Lee CH, Liu SY, Chou YT, Huang RY, Huang SM, et al. Polarization of tumor-associated macrophages and Gas6/Axl signaling in oral squamous cell carcinoma. *Oral Oncol.* (2015) 51:683–9. doi: 10.1016/j.oraloncology.2015.04.004
53. Weber M, Iliopoulos C, Moebius P, Buttner-Herold M, Amann K, Ries J, et al. Prognostic significance of macrophage polarization in

- early stage oral squamous cell carcinomas. *Oral Oncol.* (2016) 52:75–84. doi: 10.1016/j.oraloncology.2015.11.001
54. Yamagata Y, Tomioka H, Sakamoto K, Sato K, Harada H, Ikeda T, et al. CD163-positive macrophages within the tumor stroma are associated with lymphangiogenesis and lymph node metastasis in oral squamous cell carcinoma. *J Oral Maxillofac Surg.* (2017) 75:2144–53. doi: 10.1016/j.joms.2017.03.009
 55. Dan H, Liu S, Liu J, Liu D, Yin F, Wei Z, et al. RACK1 promotes cancer progression by increasing the M2/M1 macrophage ratio via the NF-kappaB pathway in oral squamous cell carcinoma. *Mol Oncol.* (2020) 14:795–807. doi: 10.1002/1878-0261.12644
 56. Sun H, Miao C, Liu W, Qiao X, Yang W, Li L, et al. TGF-beta1/TbetaRII/Smad3 signaling pathway promotes VEGF expression in oral squamous cell carcinoma tumor-associated macrophages. *Biochem Biophys Res Commun.* (2018) 497:583–90. doi: 10.1016/j.bbrc.2018.02.104
 57. Okubo M, Kioi M, Nakashima H, Sugiura K, Mitsudo K, Aoki I, et al. M2-polarized macrophages contribute to neovasculogenesis, leading to relapse of oral cancer following radiation. *Sci Rep.* (2016) 6:27548. doi: 10.1038/srep27548
 58. Cai J, Qiao B, Gao N, Lin N, He W. Oral squamous cell carcinoma-derived exosomes promote M2 subtype macrophage polarization mediated by exosome-enclosed miR-29a-3p. *Am J Physiol Cell Physiol.* (2019) 316:C731–40. doi: 10.1152/ajpcell.00366.2018
 59. Kumar AT, Knops A, Swendseid B, Martinez-Outschoom U, Harshyne L, Philp N, et al. Prognostic significance of tumor-associated macrophage content in head and neck squamous cell carcinoma: a meta-analysis. *Front Oncol.* (2019) 9:656. doi: 10.3389/fonc.2019.00656
 60. Fujii N, Shomori K, Shiomi T, Nakabayashi M, Takeda C, Ryoke K, et al. Cancer-associated fibroblasts and CD163-positive macrophages in oral squamous cell carcinoma: their clinicopathological and prognostic significance. *J Oral Pathol Med.* (2012) 41:444–51. doi: 10.1111/j.1600-0714.2012.01127.x
 61. He KF, Zhang L, Huang CF, Ma SR, Wang YF, Wang WM, et al. CD163+ tumor-associated macrophages correlated with poor prognosis and cancer stem cells in oral squamous cell carcinoma. *Biomed Res Int.* (2014) 2014:838632. doi: 10.1155/2014/838632
 62. Li C, Shintani S, Terakado N, Nakashiro K, Hamakawa H. Infiltration of tumor-associated macrophages in human oral squamous cell carcinoma. *Oncol Rep.* (2002) 9:1219–23. doi: 10.3892/or.9.6.1219
 63. Liu SY, Chang LC, Pan LF, Hung YJ, Lee CH, Shieh YS. Clinicopathologic significance of tumor cell-lined vessel and microenvironment in oral squamous cell carcinoma. *Oral Oncol.* (2008) 44:277–85. doi: 10.1016/j.oraloncology.2007.02.007
 64. Lu CF, Huang CS, Tjui JW, Chiang CP. Infiltrating macrophage count: a significant predictor for the progression and prognosis of oral squamous cell carcinomas in Taiwan. *Head Neck.* (2010) 32:18–25. doi: 10.1002/hed.21138
 65. Wei W, Hu-Jie C. Association of the infiltration of tumor-associated macrophages, expression of Smad7 protein and prognosis in oral squamous cell carcinoma. *Arch Oral Biol.* (2018) 95:22–9. doi: 10.1016/j.archoralbio.2018.07.002
 66. Fang WY, Chen YW, Hsiao JR, Liu CS, Kuo YZ, Wang YC, et al. Elevated S100A9 expression in tumor stroma functions as an early recurrence marker for early-stage oral cancer patients through increased tumor cell invasion, angiogenesis, macrophage recruitment and interleukin-6 production. *Oncotarget.* (2015) 6:28401–24. doi: 10.18632/oncotarget.4951
 67. Matsuoka Y, Yoshida R, Nakayama H, Nagata M, Hirotsue A, Tanaka T, et al. The tumour stromal features are associated with resistance to 5-FU-based chemoradiotherapy and a poor prognosis in patients with oral squamous cell carcinoma. *APMIS.* (2015) 123:205–14. doi: 10.1111/apm.12344
 68. Kimura S, Nanbu U, Noguchi H, Harada Y, Kumamoto K, Sasaguri Y, et al. Macrophage CCL22 expression in the tumor microenvironment and implications for survival in patients with squamous cell carcinoma of the tongue. *J Oral Pathol Med.* (2019) 48:677–85. doi: 10.1111/jop.12885
 69. Kang Y, Zhang Y, Sun Y. Macrophage migration inhibitory factor is a novel prognostic marker for human oral squamous cell carcinoma. *Pathol Res Pract.* (2018) 214:1192–8. doi: 10.1016/j.prp.2018.06.020
 70. MB DES, Curioni OA, Kanda JL, MB DEC. Serum and salivary macrophage migration inhibitory factor in patients with oral squamous cell carcinoma. *Oncol Lett.* (2014) 8:2267–75. doi: 10.3892/ol.2014.2513
 71. Stasikowska-Kanicka O, Wagrowska-Danilewicz M, Danilewicz M. CD8+ and CD163+ infiltrating cells and PD-L1 immunoeexpression in oral leukoplakia and oral carcinoma. *APMIS.* (2018) 126:732–8. doi: 10.1111/apm.12881
 72. Ye X, Zhang J, Lu R, Zhou G. Signal regulatory protein alpha associated with the progression of oral leukoplakia and oral squamous cell carcinoma regulates phenotype switch of macrophages. *Oncotarget.* (2016) 7:81305–21. doi: 10.18632/oncotarget.12874
 73. Chawla H, Urs AB, Augustine J. Association of macrophages with angiogenesis in oral epithelial dysplasia, oral verrucous carcinoma, and oral squamous cell carcinoma: an immunohistochemical study. *Appl Immunohistochem Mol Morphol.* (2017) 25:203–8. doi: 10.1097/PAI.0000000000000284
 74. Vallonthael AG, Singh MK, Dinda AK, Kakkar A, Thakar A, Das SN. Expression of cell cycle-associated proteins p53, pRb, p16, p27, and correlation with survival: a comparative study on oral squamous cell carcinoma and verrucous carcinoma. *Appl Immunohistochem Mol Morphol.* (2016) 24:193–200. doi: 10.1097/PAI.0000000000000179
 75. Mantovani A, Marchesi F, Malesci A, Laghi L, Allavena P. Tumour-associated macrophages as treatment targets in oncology. *Nat Rev Clin Oncol.* (2017) 14:399–416. doi: 10.1038/nrclinonc.2016.217
 76. Tang X, Mo C, Wang Y, Wei D, Xiao H. Anti-tumour strategies aiming to target tumour-associated macrophages. *Immunology.* (2013) 138:93–104. doi: 10.1111/imm.12023
 77. Georgoudaki AM, Prokopec KE, Boura VF, Hellqvist E, Sohn S, Ostling J, et al. Reprogramming tumor-associated macrophages by antibody targeting inhibits cancer progression and metastasis. *Cell Rep.* (2016) 15:2000–11. doi: 10.1016/j.celrep.2016.04.084
 78. Muller E, Christopoulos PF, Halder S, Lunde A, Beraki K, Speth M, et al. Toll-like receptor ligands and interferon-gamma synergize for induction of antitumor M1 macrophages. *Front Immunol.* (2017) 8:1383. doi: 10.3389/fimmu.2017.01383
 79. Muller E, Speth M, Christopoulos PF, Lunde A, Avdagic A, Oynebraten I, et al. Both Type I and Type II interferons can activate antitumor M1 macrophages when combined with TLR stimulation. *Front Immunol.* (2018) 9:2520. doi: 10.3389/fimmu.2018.02520
 80. Jaiswal S, Jamieson CH, Pang WW, Park CY, Chao MP, Majeti R, et al. CD47 is upregulated on circulating hematopoietic stem cells and leukemia cells to avoid phagocytosis. *Cell.* (2009) 138:271–85. doi: 10.1016/j.cell.2009.05.046
 81. Oldenborg PA, Zheleznyak A, Fang YF, Lagenaur CF, Gresham HD, Lindberg FP. Role of CD47 as a marker of self on red blood cells. *Science.* (2000) 288:2051–4. doi: 10.1126/science.288.5473.2051
 82. Alvey CM, Spinler KR, Irianto J, Pfeifer CR, Hayes B, Xia Y, et al. SIRPA-inhibited, marrow-derived macrophages engorge, accumulate, and differentiate in antibody-targeted regression of solid tumors. *Curr Biol.* (2017) 27:2065–77 e6. doi: 10.1016/j.cub.2017.06.005
 83. Klichinsky M, Ruella M, Shestova O, Lu XM, Best A, Zeeman M, et al. Human chimeric antigen receptor macrophages for cancer immunotherapy. *Nat Biotechnol.* (2020) 38:947–53. doi: 10.1038/s41587-020-0462-y

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Kalogirou, Tosios and Christopoulos. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.