



Targeting the Stroma in the Management of Pancreatic Cancer

Penelope Edwards^{1†}, Byung Woog Kang^{2†} and Ian Chau^{1*}

¹ Department of Medicine, Royal Marsden Hospital, London, United Kingdom, ² Department of Oncology/Hematology, Kyungpook National University Hospital, School of Medicine, Kyungpook National University, Daegu, South Korea

Pancreatic cancer (PC) presents extremely aggressive tumours and is associated with poor survival. This is attributed to the unique features of the tumour microenvironment (TME), which is known to create a dense stromal formation and poorly immunogenic condition. In particular, the TME of PC, including the stromal cells and extracellular matrix, plays an essential role in the progression and chemoresistance of PC. Consequently, several promising agents that target key components of the stroma have already been developed and are currently in multiple stages of clinical trials. Therefore, the authors review the latest available evidence on novel stroma-targeting approaches, highlighting the potential impact of the stroma as a key component of the TME in PC.

OPEN ACCESS

Edited by:

Sumit Sahni, Royal North Shore Hospital, Australia

Reviewed by:

Jeremy Wilson, University of New South Wales, Australia Emily K Colvin, The University of Sydney, Australia

*Correspondence:

Ian Chau ian.chau@rmh.nhs.uk [†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Gastrointestinal Cancers, a section of the journal Frontiers in Oncology

Received: 05 April 2021 Accepted: 30 June 2021 Published: 14 July 2021

Citation:

Edwards P, Kang BW and Chau I (2021) Targeting the Stroma in the Management of Pancreatic Cancer. Front. Oncol. 11:691185. doi: 10.3389/fonc.2021.691185 Keywords: pancreatic cancer, stroma, immune checkpoint inhibition (ICI), hyaluronic acid, hedgehog (Hh), bruton kinase inhibitor, SPARC

INTRODUCTION

Pancreatic cancer (PC) has a 5-year survival rate of less than 8% (1). The treatment of operable PC involves surgery and adjuvant chemotherapy, yet only around 20% of patients present with an operable disease (2). Even for patients who achieve R0 surgical resection, the likelihood of local relapse is 20% and metastatic disease is 80%. For inoperable patients, chemotherapy is offered, yet the response rate (RR) is generally low with only a marginal improvement in efficacy. In a metastatic setting, triplet therapy includes folinic acid, 5-flurouracil, irinotecan and oxaliplatin (FOLFIRINOX), which has a median overall survival (OS) of 11.1 months (3). Comparatively, the OS for gemcitabine alone is 6.6 months, with a modest improvement to a 8.7 months when adding nanoparticle albumin-bound paclitaxel (nab-paclitaxel) (4, 5). Despite the recent development of new chemotherapeutic regimens, patients with PC still show a dismal prognosis, thus new treatment options are urgently required.

The dense stromal formation surrounding PC and tumour microenvironment (TME) contributes to the aggressiveness of PC (6). The stromal cells consist of cancer-associated fibroblasts (CAFs), immune cells, and endothelial cells, while the extracellular matrix (ECM) includes collagen, fibronectin, glycosaminoglycan, hyaluronic acid (HA), and proteinases (7). In particular, the fibrotic stromal reaction and desmoplastic TME are generally related with the recruitment and activation of CAFs, extensive infiltration of ECM components, and an altered tumour vasculature (8). Current evidence suggests that this stromal remodelling is directly involved in tumourigenesis and cancer progression (9). Moreover, the fibrotic reaction inhibits the immune response, which is impaired as a result of several immunosuppressive mechanisms (10). Therefore, new treatment strategies that target the tumour stroma may be effective for patients with PC.

1

Cancer research studied the factors that promote normal pancreatic cells to develop into preneoplastic lesions and eventually to invasive cancer (11). Activating mutations have been found in Kirsten Rat Sarcoma Viral Oncogene Homologue (KRAS), p16, and p53, plus around 10% of PC has been found to have a hereditary or genetic component, with germline alterations most commonly related to breast cancer gene 2 (BRCA2) and others (12, 13). Recently, following a comprehensive integrated genomic analysis of 456 pancreatic ductal adenocarcinomas using various platforms, the Cancer Genome Atlas Research Network (TCGA) proposed four distinct subtypes: squamous, aberrantly differentiated endocrine exocrine (ADEX), pancreatic progenitor, and immunogenic tumours (14). Interestingly, the squamous and immunogenic subtypes are especially related to the TME than the tumour cells (11). Another recent investigation also established a stromal classification with two subtypes ECM-rich and immunerich, where the ECM-rich subtype was associated with a shorter survival compared to immune-rich tumours (15). Similar to this classification, digital deconvolution of transcriptomic data identified two distinct stromal-specific gene expression signatures: normal and activated, where normal stroma was dominated by markers for pancreatic stellate cells (PSCs), whereas activated stroma was characterized by inflammatory signatures associated with a significantly worse prognosis (16). Consequently, based on a better molecular characterization of PC that includes the stroma, targeting the appropriate stromal components may lead to a better outcome and represent a promising approach for precision medicine in PC treatment.

ROLE OF DESMOPLASTIC STROMA IN TUMOUR MICROENVIRONMENT OF PANCREATIC CANCER

A desmoplastic stroma helps to define PC and is the basis of a complex TME (17). As mentioned above, a desmoplastic stroma is a very complex and heterogeneous network, and organized by interactions among various cell types and acellular components (9) (**Figure 1**).

CAFs are a major cellular component of PC stroma and derived from several origins, such as PSCs, fibroblasts, and epithelial, endothelial, and mesenchymal cells, through multiple activation pathways including epithelial-mesenchymal transition (EMT) (18, 19) (Table 1). PSCs are major sources of CAFs and found in the periacinar, perivascular, or periductal sites of the exocrine pancreas (26-28). In normal conditions, PSCs are quiescent with long cytoplasmic extensions and vitamin-A storing fat droplets (28). In the case of activation, PSCs transform into a myofibroblast-like phenotype that can support pancreatic fibrosis and tumour growth in PC (20). Moreover, adipocytes, pericytes, monocytes, endothelial cells, and bone marrow-derived or adipose-derived mesenchymal stem cells can differentiate into CAFs (21-25). These diverse origin of CAFs may explain their multiple roles and abilities in terms of tumour growth and progression in PC (29). These pathways are inactive in normal tissue, yet activated by secreted factors, such as tumour necrosis factor α (TNF α), transforming growth factor β (TGF- β), platelet derived growth factor (PDGF), interleukins, and sonic hedgehog (SHh), where these activated,



TABLE 1	Potential sources of	cancer-associated	fibroblasts (CAFs).
---------	----------------------	-------------------	---------------------

Origin	Description					
Pancreatic stellate cells (PSCs)	PSCs are the most studied CAF subtype and may participate in cancer pathogenesis after transforming from a quiescent state into an "activated" state	(20)				
Bone marrow-derived mesenchymal stem cells (BM-MSCs)	BM-MSCs can differentiate into a substantial proportion of CAFs in cancers.	(21)				
Adipocytes	Human adipose tissue-derived stem cells are a source of CAFs and exhibit the functional properties of CAFs.	(22)				
Pericytes	Perivascular cells have been identified as a major source of profibrotic cells in acute liver injury	(23)				
Epithelial cells	A substantial number of organ fibroblasts appear via a novel direction reversal of the epithelial cell fate.	(24)				
Endothelial cells	The endothelial to mesenchymal transition is associated with a phenotype conversion into fibroblast-like cells.	(25)				

specialized forms of fibroblasts express alpha-smooth muscle actin (aSMA), stromal cell-derived factor 1 alpha, fibroblast activation protein, and fibroblast specific protein-1 (7, 30). Once activated, CAFs are known to work as a key component of the TME with diverse functions, including matrix remodelling, metabolic effects, and immune crosstalk (29). CAFs are also a substantial source of growth factors, cytokines and exosomes that can induce the characteristic desmoplasia of the stroma. CAFs eventually promote various tumour-promoting processes, including cancer cell growth, proliferation, metastasis, and tumour angiogenesis and contribute to tumour therapy resistance (31) (Figure 2). Ohlund et al. recently identified two distinct populations of inflammatory fibroblasts (iCAFs) and myofibroblasts (myoCAFs) in PC (32). myoCAFs are located in close proximity to tumour cells and express high levels of α SMA, while iCAFs are localized at distal sites and exhibit low levels of α SMA and high levels of inflammatory mediators, contributing to an immunosuppressive environment (18, 29).

The ECM contains proteins, such as collagen, integrin, several types of glycans, and multiple soluble factors, with diverse physical and biochemical properties (33). These factors function as a dynamic molecular system that controls cells by altering proliferation, cytoskeletal organization, cellular differentiation, and receptor signalling (34). The predominant glycosaminoglycan is HA, which increases tumour tissue stiffness, promotes metastasis, and regulates tumour immunity (35). Under normal circumstances, the amount of HA is controlled by a balance between synthesis and degradation (36). However, this balance can be shifted towards a higher concentration in PC and this HA-rich microenvironment can then promote tumour proliferation by enhancing cell growth, migration, invasion, EMT, and metastasis (36, 37).



Several studies have reported increased expression of HA and its receptors in PC tissues, which is also associated with poor prognosis (38). *In vitro* studies have also revealed that HA can stimulate PC cell migration. Moreover, animal studies found accelerated tumour growth from the accumulation of extracellular HA due to the forced expression of synthesizing genes (39). In addition to its significant tumour-promoting effects, HA increases the interstitial fluid pressure, thereby compressing intratumoural blood vessels and resulting in hypoxia. This hypoxia may induce angiogenesis and, in an experimental model, the induction of HA overproduction was found to accelerate angiogenesis through stromal cell recruitment (40).

Along with HA, extracellular proteins, including connective tissue growth factor (CTGF) and secreted protein acidic and rich in cysteine (SPARC) are also essential for maintaining physiologic homeostasis and modulate multiple biologic processes, such as wound repair, tissue remodeling, angiogenesis, matrix cell adhesion, cell differentiation, proliferation, and migration (9, 41). The functions of these molecules might be in part mediated by interaction with PSCs/CAFs and several growth factors, like TGF- β and fibroblast growth factor (FGF) (16). Although the role of ECM components in carcinogenesis remains controversial in PC, they are highly expressed in various types of cancer (42). Plus, several studies have demonstrated that overexpression of ECM components generally indicates a worse prognosis in PC (43). Thus, when taken together, ECM components could represent a promising therapeutic target in the treatment of PC.

PC is also characterized by excessive dense ECM deposition associated with vasculature collapse and limited oxygen delivery, promoting the induction of an invasive and treatment-resistant phenotype (44, 45). In response to hypoxia, PC tumour cells produce several angiogenic molecules, including hypoxiainducible factors (HIFs), vascular endothelial growth factors (VEGFs), FGFs, PDGF, Matrix metallopeptidase 9 (MMP-9), and interleukin-8. Hypoxia-promoted changes lead to increased tumour proliferation, metabolic changes, and also contribute to an immunosuppressive phenotype in PC (46). Multiple studies have already explored the relationship between angiogenic mediators regulated by hypoxia and PC tumour cell survival (47-53). For example, the contribution of HIF-1 α and VEGF-A to the aggressiveness of PC is not only via angiogenesis but also via direct stimulation of tumour cell proliferation and metastatic capacity (45, 54, 55). Furthermore, the abnormal formation of the vasculature inhibits effective drug penetration and uptake, contributing to the lack of efficiency of conventional chemotherapies in PC treatment (56). This explains the attempts to target the hypoxic environment in PC either directly or indirectly

An immune defect in the TME is another interesting aspect for treating PC. Although not fully elucidated, the mechanism of immune suppression may be due to a low immunogenicity, the activation of stromal cells, recruitment of immunosuppressive cells, dense fibrotic stroma, or the secretion of immunosuppressing cytokines (8, 10, 57). PC is known to exhibit a low mutation burden and lack of significant neoantigens, leading to the dysfunction of immune effector cells, such as T cells and natural killer cells (58, 59). CAFs can also reduce activated T cell infiltration

of the tumour site and secrete a variety of cytokines, including TGF- β that is associated with EMT (16, 60). This can promote the migration of immunosuppressive inflammatory cells such as regulatory T cells (Treg), myeloid-derived suppressor cells (MDSCs), and tumour-associated macrophages (TAMs) through the CXC motif chemokine ligand (CXCL)-CXC chemokine receptor (CXCR) axis (9, 61). Furthermore, a TME-induced desmoplastic reaction can create a physical barrier for effective T cell migration, thereby evading the immune response. Recent studies have also implicated the role of the intratumoural vasculature which can interact with the immune system and suppress antitumour immunity (8). Therefore, these findings suggest that strategies that can alter the immune suppressive stroma and restore T-cell-mediated immune surveillance may potentially improve the outcome for PC patients.

THERAPEUTIC STRATEGIES TARGETING STROMAL CELLS IN PANCREATIC CANCER

Several promising agents that target key components of the stroma have already been developed and are currently in multiple stages of clinical trials in PC (**Table 2**).

Hyaluronic Acid

Targeting HA deposition can be divided into 3 approaches: depleting stromal HA, inhibiting HA synthesis, and blocking HA signalling (79). In a mouse model, pegvorhyaluronidase alfa (PEGPH20), the PEGylated form of a recombinant human hyaluronidase, has been shown to deplete or reduce HA within the stroma, thereby improving tumour perfusion and drug delivery (80). Moreover, when combined with chemotherapy, PEGPH20 exhibited a synergistic effect with substantially tumour inhibition (81). Such preclinical studies led to a phase Ib study by Hingorani et al. to evaluate the safety and efficacy of escalating doses of intravenous PEGPH20 combined with gemcitabine in patients with metastatic PC (62). The combination was well tolerated and demonstrated therapeutic benefit in 28 patients. Plus, patients with high levels of HA expression showed an improved objective response and median survival. These results then prompted an investigation of PEGPH20 with gemcitabine plus nab-paclitaxel in the phase II HALO 202 trial (82). However, stage one produced an imbalance of thromboembolic (TE) events for patients receiving PEGPH20, therefore, the study protocol was revised to exclude high-TE-risk patients and include prophylactic enoxaparin. In stage two using the revised protocol, the primary endpoints were progressionfree survival (PFS) and the incidence of TEs. The randomized study of 279 patients with untreated metastatic PC administered PEGPH20 plus chemotherapy or chemotherapy alone. The patients that received PEGPH20 plus gemcitabine and nabpaclitaxel benefited from an improved median PFS (6.0 versus 5.3 months, P=0.049) and the TE events were comparable after including enoxaparin. The benefit was even more pronounced in

TABLE 2 | Selected phase II and III studies evaluating stroma-targeting agents in pancreatic cancer.

Target	Agents	Phase	Setting	Treatment arms	Ν	PE	RR (%)	PFS (months)	OS (months)	Reference
Hyaluronic acid	PEGPH20	II	First-line	PEGPH20/Gemcitabine/Nab- paclitaxel	116	PFS/ TE	40	6.0	9.6	(62)
				Gemcitabine/Nab-paclitaxel	113		33	5.3	9.2	
	PEGPH20	III	First-line/HA-high	PEGPH20/Gemcitabine/Nab- paclitaxel	327	OS	34	7.1	11.2	(63)
				Gemcitabine/Nab-paclitaxel	165		27	7,1	11.5	
	PEGPH20	lb/ll	First-line	PEGPH20/mFOLFIRINOX	55	OS	33	4.3	7.7	(64)
				mFOLFIRINOX	56		45	6.2	14.4	
CTGF	Pamrevlumab (FG-3019)	1/11	LAPC	Pamrevlumab/Gemcitabine/Nab- paclitaxel	37	-	30	-	-	(65)
	Pamrevlumab	III	LAPC	Pamrevlumab/Gemcitabine/Nab- paclitaxel	256			Ongoing		NCT03941093
Hh pathway	Vismodegib	II	First-line	Vismodegib/Gemcitabine/Nab- paclitaxel	71	-	40	5.4	9.8	(66)
FAK	Defactinib	l (exp.)	Refractory solid tumour/PC	Defactinib/Gemcitabine/ Pembrolizumb	43			Ongoing		NCT02546531
	Defactinib	П	Resectable	Defactinib/Pembrolizumb	36			Ongoing		NCT03727880
HGF pathway	Cabzantinib	Ι	First- or Second- line	Cabozantinib/Gemcitabine	12	-	-	4.7	10.1	(67)
	Cabozantinib	II	Second-line or more	Cabozantinib/Atezolizumab	29			Ongoing		NCT04820179
SPARC	Nab-paclitaxel	1/11	First-line	Gemcitabine/Nab-paclitaxel	67	-	46	7.1	10.3	(68)
	Nab-paclitaxel	III	First-line	Gemcitabine/Nab-paclitaxel	431	OS	23	6.7	8.5	(5)
				Gemcitabine	430		7	3.7	5.5	(-)
BTK	Ibrutinib	III	First-line	Ibrutinib/Gemcitabine/Nab- palictaxel	211	OS/ PFS	29	5.3	9.7	(69)
				Gemcitabine/Nab-palictaxel	213		42	6.0	10.8	
Antiangiogenic										
agents										
VEGF	Bevacizumab	III	First-line	Bevacizumab/Gemcitabine/Erlotinib	306	OS	13.5	4.6	7.1	(70)
				Gemcitabine/Erlotinib	301		8.6	3.6	6.0	
	Bevacizumab	III	First-line	Bevacizumab/Gemcitabine	302	OS	13	3.8	5.8	(71)
				Gemcitabine	300		10	2.9	5.9	
VEGFR	Axitinib	III	First-line	Axitinib/Gemcitabine	314	OS	12	4.4	8.5	(72)
				Gemcitabine	316		4	4.4	8.3	
VEGFR	Sorafenib	III	First-line	Sorafenib/Gemcitabine	52	PFS	23	3.8	8.0	(73)
				Gemcitabine	52		19	5.7	9.2	
VEGF	Aflibercept	III	First-line	Aflibercept/Gemcitabine	271	OS	-	3.7	6.5	(74)
Immunotherapy				Gemcitabine	275		-	3.7	7.8	
Vaccine	GVAX/CRS-207	П	Second-line or	$GVAX/Cy \rightarrow CRS-207$	61	OS	0	-	6.0	(75)
			more	GVAX/Cy	29		0	-	3.4	
	GVAX/CRS-207	llb	Second-line or	$\text{GVAX/Cy} \rightarrow \text{CRS-207}$	29	OS	-	2.4	4.3	(!38)
			more	CRS-207	31		-	-	4.1	
				Single-agent chemotherapy	30		9.1	2.4	9.1	
ICIs CTLA4	Ipilimumab	Ш	LAPC or	Ipilimumab	27		0		4.5	(76)
	ipiinnunao	П	metastatic	ipiinnanao	21	-	0	-	4.0	(70)
Dual ICIs CTLA4/PD-1	lpilimumab/Nivolumab	1/11	Solid tumours	Inilimumah /Nival-mah				Oncoina		NCT01928394
CTLA4/PD-1 CTLA4/PD-L1	Tremelimumab/	1/11	Second-line	lpilimumab/Nivolumab Tremelimumab/Durvalumab	-	סס	0.1	Ongoing	0.1	
UILA4/MD-LI	Durvalumab	11	Second-III le	Durvalumab	32 33	RR	3.1 0	1.5 1.5	3.1 3.6	(77)
ICls/CTx					00		0	1.0	0.0	
PD-1/CTx	Pembrolizumab	lb/ll	First-line	Pembrolizumab/Gemcitabine/Nab- paclitaxel	11	CR	27	9.1	15.0	(78)
CTLA4/PD-L1/ CTx ICls/Others	Tremelimumab/ Durvalumab	II	First-line	Tremelimumab/Durvalumab/ Gemcitabine/Nab-paclitaxel	190			Ongoing		NCT02879318
PD-1/Vaccine	Pembrolizumab/GVAX/ CRS-207	II	metastatic	Pembrolizumab/CRS-207/ Epacadostat ± GVAX/Cy	70			Ongoing		NCT03006302
PD-1/Vaccine	Nivolumab/GVAX/CRS- 207	II	metastatic	Nivolumab/CRS-207/GVAX/Cy	93			Ongoing		NCT02243371

(Continued)

TABLE 2 | Continued

Target	Agents	Phase	Setting	Treatment arms	N	PE	RR (%)	PFS (months)	OS (months)	Reference
CTLA4/PD-1/ Vaccine	lpilimumab/Nivolumab/ GVAX/CRS-207	Ш	metastatic	Ipilimumab/Nivolumab/GVAX/CRS- 207	63			Ongoing		NCT03190265
PD-L1/ Hyaluronic acid	Atezolizumab/PEGPH20	Ш	Resectable	Atezolizumab/PEGPH20	40			Ongoing		NCT03979066

N, patient number; PE, Primary endpoint; RR, response rate; PFS, progression-free survival; OS, overall survival; PEGPH20, pegvorhyaluronidase alfa; Nab-paclitaxel, nanoparticle albumin-bound paclitaxel; TE, thromboembolic events; mFOLFIRINOX, leucovorin, 5-fluorouracil, irinotecan, and oxaliplatin; CTGF, connective tissue growth factor; LAPC, locally advanced pancreatic cancer; Hh, hedgehog; FAK, focal adhesion kinase; exp., expansion; PC, pancreatic cancer; HGF, hepatocyte growth factor; SPARC, secreted protein acidic and rich in cysteine; BTK, bruton tyrosine kinase; VEGF, vascular endothelial growth factor; VEGFR, vascular endothelial growth factor receptor; Cy, cyclophosphamide; ICIs, immune checkpoint inhibitors; CTx, chemotherapy; CR, complete response.

patients with HA-high tumours, showing a median PFS of 9.2 months for the PEGPH20 arm *versus* 5.2 months for the control arm (P=0.048). The proportion of patients with HA-high tumours was 34% as defined as extracellular matrix HA staining \geq 50% of tumour surface at any intensity. Notwithstanding, the PEGPH20 treatment was also associated with an increased incidence and severity of other manageable adverse events (AEs), such as fatigue, muscle spasms, arthralgia, edema, and neutropenia.

Due to encouraging phase I and II studies, PEGPH20 was investigated in the phase III Halo 301 trial (NCT02715804), which used a combination of PEGPH20 and gemcitabine plus nab-paclitaxel in previously untreated patients with stage IV HA-high PC (63, 83). In this study, the HA status was prospectively determined using a VENTANA HA assay, with HA-high defined as ≥50% staining of a tumour sample. The primary endpoint was OS, with PFS, the objective RR, and safety included as secondary study endpoints. According to predefined criteria, 494 patients with HA-high PC were randomly assigned in a 2:1 ratio to receive first-line chemotherapy with gemcitabine and nab-paclitaxel in combination with either PEGPH20 or a placebo, and prophylactic enoxaparin was given for the TE risk. However, HALO 301 did not meet its primary endpoint of OS (11.2 months versus 11.5 months, P=0.97) (63, 84). Moreover, the combination arm did not improve the PFS or duration of response. This negative finding could imply that targeting the stromal component can affect resistance to chemotherapies or be attributed to compensatory mechanisms overcoming tumour stroma inhibition, suggesting that targeting desmoplasia alone is not sufficient and a combination of stromal modifying agents is needed. The results from the HALO 301 trial are also consistent with the results from a recently presented randomized phase I/II study (SWOG S1313) evaluating the efficacy of PEGPH20 and modified FOLFIRINOX in patients with metastatic PC. This trial closed early due to an inferior survival and significantly higher AEs in the combination arm (64). Consequently, new studies using PEGPE20 include the addition of immune checkpoint inhibitors (ICIs) (NCT03634332, NCT03979066). The therapeutic potential of PEGPH20 combined with an immune checkpoint inhibitor was studied in a murine synergic breast tumour model (85). The combination of PEGPH20 with oncolytic reovirus therapy and anti-programmed cell death protein ligand-1 (PD-L1)-targeting antibody (anti-PD-L1) resulted in enhanced anti-tumour activity and also extended survival. In addition, PEGPH20 facilitated the infiltration of cytotoxic T lymphocytes and improved the delivery of chemotherapy and programmed cell death protein-1 (PD-1)/ PD-L1 antibodies (86). Therefore, these data suggest that the removal of HA by PEGPH20 can increase the efficacy of immune checkpoint therapy and also enhance the accumulation of immunotherapeutic antibodies in HA-rich tumours. Plus, PEGPE20 is also being tested as a main drug in biomarkerdriven trials (NCT03193190).

Connective Tissue Growth Factor

CTGF is a profibrotic extracellular protein with an abundantly elevated expression in PC (16). CTGF is mediated by chemokine signalling, which promotes fibrosis and collagen deposition, which in turn causes cancer progression and metastasis (87, 88). Pamrevlumab (FG-3019), a fully human, recombinant DNA-derived IgG1 kappa monoclonal antibody against CTGF, attenuates the malignant properties of different human cancers and is currently under clinical trial for the treatment of PC. In a preclinical setting using a mouse model, enhanced antitumour activity was observed when using pamrevlumab as a single agent or in combined with gemcitabine (50, 89). In a phase I study, the safety and efficacy of increasing doses of pamrevlumab were evaluated in combination with two chemotherapy agents, gemcitabine and erlotinib, in 75 patients with previously untreated Stage III/IV PC (65). The results showed that pamrevlumab was well tolerated with no dose-limiting toxicity or dose-related trends in the type or incidence of serious AEs. In a subsequent phase I/II study using gemcitabine/nab-paclitaxel with pamrevlumab or gemcitabine/nab-paclitaxel alone for patients with locally advanced PC, pamrevlumab combination arm showed a higher percentage of surgical resection (33.3% versus 7.7%) and improved median survival rate, plus the combination was feasible and well tolerated with no incremental safety signals (90). Accordingly, a phase III (LAPIS) trial is currently ongoing to investigate the efficacy of pamrevlumab plus gemcitabine with nabpaclitaxel in locally advanced, unresectable PC (NCT03941093).

Hedgehog Pathway

The Hh signalling pathway is one of twelve core signalling pathways and processes in PC and plays a critical role in regulating the dense stroma of pancreatic cancer *in vivo* and *in vitro* (91, 92). Saridegib (IPI-926) is a potent and specific inhibitor of smoothened (Smo), a key signalling transmembrane protein in the Hh pathway (93). Preclinical studies using a mouse model showed that saridegib depleted tumour-associated stromal tissue and increased the intratumoural mean vessel density, leading to a decreased tumour burden and prolonged survival (94). However, subsequent human clinical trials failed to show a sustained benefit for this agent (95, 96). Vismodegib (GDC-0449) is another Hh pathway inhibitor that blocks Hh signalling by binding to Smo (93). Similarly, in contrast to preclinical results, the addition of vismodegib to chemotherapy did not improve the overall outcomes in phase I and II studies (66, 97). Therefore, these disappointing results suggest that inhibiting the Hh pathway ironically stimulates aggressive PC clones and invigorates the metastatic capacity (59). Plus, the inhibition of CAFs via suppressing the Hh signalling pathway may induce a decreased immune response and vascular dysfunction, revealing the complex relationship between Hh signalling and desmoplastic stroma (91). Therefore, there are currently no ongoing studies on Hh signalling inhibitors in PC, except for the biomarker-driven trial (NCT02465060).

Focal Adhesion Kinase

Focal adhesion kinase (FAK) is a nonreceptor protein tyrosine kinase that plays a central role in the collagen pathway and regulates integrin-mediated ECM signalling (98). FAK is frequently overexpressed and dysregulated in a variety of cancers, including PC, and correlated with poor prognosis (98, 99). Interestingly, in preclinical studies, FAK activation induced a fibrotic and immunosuppressive TME, while inhibition of the FAK pathway was found to interrupt the aggressive effects of collagens on PC biology (100, 101). Therefore, these observations led to an investigation of a combination therapy of FAK inhibitors with gemcitabine and pembrolizumab, a humanized monoclonal antibody directed against PD-1. Defactinib (VS-6063) is a selective, orally active, competitive adenosine triphosphate (ATP) inhibitor of FAK. A recent phase I/IIa clinical trial evaluated a dose escalation of defactinib in combination with pembrolizumab and gemcitabine for patients with advanced solid tumours, and subsequently at an optimal dose for an expanded patient cohort with PC (102). Preliminary data have showed that the combination therapy was welltolerated with no dose-limiting toxicity (DLT), and the common treatment-related AEs included fatigue, nausea, myalgia, vomiting, anorexia, pruritus, and fever. The expanded patient cohort is currently being investigating with pending correlative and efficacy data (103). Another phase II study of defactinib plus pembrolizumab is also ongoing for resectable PC patients (NCT03727880).

Hepatocyte Growth Factor Pathway

Hepatocyte growth factor (HGF), also known as scatter factor, is the ligand of c-Met (mesenchymal-epithelial transition tyrosine kinase receptor or HGF receptor) that is often secreted by stromal cells and regulates the stromal-tumour interactions in PC (104). The binding of HGF to c-Met leads to the dimerization and phosphorylation of c-Met and triggers several downstream signallings (105). This axis also plays an important role in the pathogenesis of pancreatic cancer and its progression. C-Met overexpression has already been identified in PC and correlated with aggressive disease (106), while elevated serum HGF levels have been associated with disease progression (107). Plus, c-Met activation has been linked with resistance to gemcitabine (108). Interestingly, in vitro and in vivo models have shown that crizotinib, an ATP competitive multi-target protein kinase inhibitor, increases sensitivity to gemcitabine and inhibits peritoneal dissemination (109, 110). A recent preclinical study demonstrated that the inhibition of both HGF and c-Met combined with gemcitabine improved the reduction of tumour volume and metastasis (105). Another report also indicated that long-term treatments involving cabozantinib induced less resistance and even improved the efficacy of gemcitabine (108). Therefore, these preclinical studies led to a phase I trial of cabozantinib, an orally bioavailable c-Met inhibitor, and gemcitabine in advanced PC (67). This trial involved the treatment of 12 patients and no maximum tolerated dose was determined, plus the DLT was relatively high at all dose levels. Another phase II clinical trial investigated the safety and antitumor activity of cabozantinib combined with atezolizumab in patients with PC (NCT04820179). Thus, several preclinical and early clinical studies have tested the inclusion of c-Met inhibitors, such as rilotumumab, crizotinib, capmatinib, and tivantinib, in combination treatments for solid tumours (104). However, the remaining major challenge is to identify potential predictive biomarkers to facilitate the appropriate selection of specific patient populations most likely to benefit from combination therapies.

Secreted Protein Acidic and Rich in Cysteine

SPARC is a multifunctional calcium-binding glycoprotein which modulates interactions between cells and the extracellular environment, thereby participating in cell development, matrix cell adhesion, wound repair, tissue remodeling, and angiogenesis (41, 111). SPARC may also influence PC cell proliferation, migration, metastatic, and escape mechanisms (112). Thus, SPARC has been investigated as a potential therapeutic target, where the presence of SPARC in the tumour stroma could be used to deliver albumin-conjugated molecules into the tumour and TME. Meanwhile, nab-paclitaxel, a nanoparticle form of paclitaxel, is known to deplete tumour stroma through interaction between albumin and SPARC (5). Plus, nabpaclitaxel has exhibited clinical antitumour activity in several cancer types that overexpressed SPARC (43, 113). In metastatic PC, nab-paclitaxel was used in multiple-stage clinical trials, where the addition of nab-paclitaxel to gemcitabine showed activity and efficacy in first-line treatment, improving survival and the overall RR (112, 114). Interestingly, patients with high stromal-SPARC expression exhibited a significant increase in OS compared to patients with low stromal-SPARC expression (17.8 versus 8.1 months), representing a significant predictor of OS in a multivariate analysis (68). Based on these results, a large, openlabel, international, randomized, phase III trial (Metastatic Pancreatic Adenocarcinoma Clinical Trial, MPACT) followed with 861 metastatic PC patients as a first-line setting (5). The patients were randomly assigned 1:1 to receive either nabpaclitaxel plus gemcitabine or gemcitabine alone. As mentioned above, since the trial met its primary endpoint of OS, this combination is now a standard regimen in the treatment

of metastatic PC. However, the stromal and tumour levels of SPARC, measured by immunohistochemistry, showed no association with survival (115, 116). These conflicting results on SPARC as a prognostic and predictive biomarker in PC may have been impacted by different methodologies of protein detection and staining evaluation, the origin of the tumour samples, and the level of the cutoff value. Moreover, recent data has suggested that specific tumour delivery of nabpaclitaxel is not directly related to SPARC expression, and nab-paclitaxel does not usually deplete tumour stroma (117). Furthermore, the efficacy of nab-paclitaxel may be dependent on the drug internalization by TAMs (118). Consequently, for PC patients, the potential link between SPARC expression and treatment efficacy remains unclear, and more precise methods are needed to analyse SPARC expression. Plus, discovering a novel targeted nanoparticle and enhanced drug delivery system using SPARC could improve the pharmacologic and therapeutic properties of conventional cancer treatment for patients with PC.

Bruton Tyrosine Kinase

Ibrutinib is a first-generation Bruton tyrosine kinase (BTK) inhibitor that irreversibly binds to cysteine at position 481 in the kinase domain and blocks enzyme activity (119). Protein tyrosine kinase BTK, which is essential for B cell maturation and expressed by other immune cell populations, has been implicated in the immune regulation and function of myeloid cells (120, 121). BTK has also been shown to exhibit anti-fibrotic effects in PC by effectively inhibiting the infiltration of mast cells in both transgenic mice and patient-derived xenograft models. Furthermore, ibrutinib reduces stromal fibrosis and inhibits tumour progression (122). Preclinical data for ibrutinib plus gemcitabine showed enhanced antitumour activities versus gemcitabine alone in both transgenic mouse and patientderived xenograft PC models (122). Early developmental clinical trials also showed preliminary efficacy in solid tumours, including PC (NCT02562898) (123). Thus, based on these encouraging results, ibrutinib was included in a recent randomized phase III RESOLVE trial (NCT02436668) that assessed ibrutinib in combination with gemcitabine and nabpaclitaxel or a placebo plus chemotherapy as the first-line treatment for patients with metastatic PC (124). Although the full results have yet to be published, it has been revealed that the study failed to meet its primary endpoint of prolonged PFS and no statistically significant PFS or OS benefit was shown when adding ibrutinib to chemotherapy (69). Although both treatment arms showed a similar proportion of patients with grade \geq 3 AEs, the patients in the ibrutinib arm were treated for a shorter time period and received lower cumulative doses of all agents compared with the placebo arm. Therefore, this emphasizes the need for clinical biomarkers to predict the sensitivity and resistance to BTK inhibitors and identify the best combination partners for synergistic effects and lower toxicity. Despite these disappointing results, more recent studies are investigating the combination of BTK inhibitors with ICIs. Indeed, multiple cell types, such as MDSCs, macrophages, dendritic cells, and endothelial cells, in the TME are regulated by BTK (119). In a preclinical study, ibrutinib treatment successfully reprogrammed

macrophages for increasing CD8+ T cells to assist with tumour control in PC (125). Moreover, the synergistic activity of ibrutinib with ICIs has also been described in preclinical models, including regulating tumour-induced immune tolerance by enhancing the activity of tumour-infiltrating cells and reducing the secretion of immunosuppressive cytokines (126). A recent phase Ib/II study evaluated ibrutinib plus durvalumab, a PD-L1-targeting antibody, in patients with relapsed or refractory solid tumours (127). Although ibrutinib plus durvalumab had an acceptable safety profile, the combination showed limited activity with an overall RR of 2% and median OS of 4.2 months for 49 PC patients. Another randomized phase II study evaluated the effect of a secondgeneration BTK inhibitor in patients with advanced PC using acalabrutinib alone or in conjunction with pembrolizumab, an anti-PD-1 antibody (128). The monotherapy and combination treatments both showed minimal clinical benefit. Therefore, additional trials are needed to refine the dual inhibition of BTK and the PD-1/PD-L1 pathway in PC.

Angiogenesis

Various stromal components, such as collagen, fibronectin, HA, VEGF, TGF-B, and CTGF are closely associated with a fibrotic and hypoxic tumour condition, which forms a hard mass and releases several proangiogenic factors, including VEGF, MMP-9, interleukin-8, and fibroblast growth factor-2 (129). These stromal cells and ECM play a crucial role in stimulating or inhibiting angiogenesis via multiple pathways and numerous genetic alterations in PC (56). Several studies have already demonstrated that overexpression of VEGF is related with tumour progression and a poor prognosis, and antiangiogenic treatment reduces tumour cell growth (130). Consequently, anticipating that antiangiogenic therapy could be effective for PC, a single-arm phase II trial with a combination of bevacizumab and gemcitabine did show clinical benefits (131). However, in two large phase III studies, the addition of bevacizumab to chemotherapy failed to reach the primary endpoint of OS (70, 71). Despite several subsequent clinical trials using bevacizumab and other chemotherapy backbones, no treatment benefits were reported (130).

Moreover, subsequent phase III clinical trials of antiangiogenic agents targeting vascular endothelial growth factor receptor (VEGFR) or VEGF, including axitinib, sorafenib, and aflibercept, showed no significant prolongation of OS (56, 72-74). Such findings are also consistent with two meta-analyses, that indicated no increase in OS with any VEGF or VEGFR-targeting treatment (130, 132). There are various explanations for the limited clinical outcomes of antiangiogenic agents in PC (133). Antiangiogenic therapy may inhibit the blood supply to tumours and inevitably diminish the drug delivery, which would partially explain the combined need for inhibition of angiogenesis and alternative mechanisms in the treatment of PC (134). Antiangiogenic therapy per se may support a conversion to a more aggressive phenotype, promoting the induction of invasive and treatment-resistant tumours (129). In addition, the angiogenesis pathway comprises a complex network of crosstalk with many parallel

cascades, so its inhibition may induce compensatory upregulation of proangiogenic factors, thereby paradoxically increasing tumour growth (56). Moreover, an adaptive response to hypoxia primarily mediated by HIFs confers more aggressive phenotypes in PC cells (129). Finally, continuous hypoxic microenvironment remodelling can stimulate the regulation of autophagy and generation of reactive oxygen species and alternative pathways, including the metabolism of glucose or glutamine, while also contributing to therapeutically resistant behaviour of PC (135). Therefore, considering its complex and multifaceted functionality, further studies of antiangiogenic therapy in PC are needed to identify the best combination partners for synergistic effects, especially with stromal depletion strategies that target specific stromal features. Exploring potential biomarkers will also be important to select the appropriate PC patient populations.

Immune Cells

The ability to evade immune surveillance is a recognized hall mark in PC biology (136). Since PC is also well characterized by immune-quiescent desmoplastic stroma and the dominance of immunosuppressive cells, some recent research has been focusing on the development of immune-based therapies for PC (57). One promising immunotherapeutic strategy is to stimulate T-cell priming and dendritic cell activation, as the resulting activation of tumour-specific T cells and their migration into the TME may be the key to surmounting innate immune suppression and correcting the lack of effector T cells (17, 58). One of the most widely evaluated PC vaccines is GVAX, which is an irradiated allogeneic whole tumour cell vaccine engineered to express a granulocyte-macrophage colonystimulating factor (137). In early phase studies, GVAX has been found to be tolerable and effective in promoting the development of antitumour immunity (138, 139). These results then prompted a randomized phase 2 trial evaluating GVAX pancreas in combination with cyclophosphamide followed by CRS-207, a live-attenuated listeria vaccine expressing a PCassociated antigen mesothelin in previously treated metastatic PC. This triple combination therapy demonstrated a longer OS with minimal toxicity and enhanced mesothelin-specific CD8 cytotoxic cells correlated with the improved OS (75). However, a subsequent phase IIb trial (ECLIPSE) failed to show any superiority of this combination over chemotherapy (140). This may have been due to the failure of sufficient T cell induction or quick exhaustion of effective immune cells (58). Plus, major contributors, including CAFs, MDSCs, or TAMs may regulate multiple immunosuppressive mechanisms, indicating the dynamic nature and heterogeneity of the immune response in PC (18, 57). Thus, the addition of a new combination approach seems to be the ideal strategy to achieve a more impressive response in PC, and ICIs could be such a treatment.

ICIs can lead to T cell activation, thereby generating an effective immune response. Nevertheless, a broad spectrum of clinical trials for each ICI monotherapy has shown limited clinical success. For example, a phase II study with ipilimumab, a monoclonal antibody targeting cytotoxic T-lymphocyte-associated protein 4 (CTLA-4), showed no significant improvement in survival (76). In early clinical trials, other single-agent anti-PD-1 agents, including pembrolizumab and nivolumab have also been ineffective in the treatment of PC (141, 142). Consequently, combination approaches have been explored to enhance the immune responses for better therapeutic effects when treating PC. For a dual inhibition setting, a recent randomized phase II study evaluated 65 previously treated metastatic PC patients for the efficacy of durvalumab alone or in combination with the anti-CTLA-4 antibody, tremelimumab (77). Although the combination therapy produced a modest improvement with a 3-month disease control rate (DCR) of 9.4%, as the desired threshold efficacy (10%) was not achieved, the study was closed. Another ongoing clinical study is evaluating ipilimumab in combination with nivolumab (NCT01928394) (137).

ICIs in combination with chemotherapeutic agents are an increasing focus as an alternative option for the treatment of PC. A recent phase Ib trial reported that an ipilimumab and gemcitabine combination was safe, including a delayed response in one patient (143). Tremelimumab plus gemcitabine has also demonstrated a tolerable toxicity with median survival of 7.4 months (144). Moreover, the preliminary results from a randomized phase 2 trial (CCTG PA.7) evaluating the efficacy of gemcitabine and nabpaclitaxel in combination with durvalumab and tremelimumab showed a median PFS of 2.5 months and median OS of 8.5 months (NCT02879318) (145). Plus, pembrolizumb was combined with gemcitabine and nab-paclitaxel in a phase Ib/II study including 17 patients with metastatic PC. Of the 11 evaluable chemotherapy naïve patients, this combination showed an interesting outcome with a DCR of 100% and median PFS of 9.1 months, which seems to be much better than typically reported for first-line chemotherapy (78).

Combining of ICIs with a vaccine or other immunomodulatory molecules is another potentially effective regimen, and is currently under clinical investigation to determine the efficacy in PC. Le et al. evaluated combining ipilimumab and the GVAX vaccine in a phase Ib trial, where 30 patients were randomly assigned to single-agent ipilimumab versus ipilimumab combined with GVAX. As a result, the combination patients showed a better 1-year OS benefit versus ipilimumab alone (27% versus 7%), prompting additional GAVX clinical trials with various modalities (146). Numerous similar clinical trials are also underway, including agonist CD40 antibodies, indoleamine 2,3-dioxygenase inhibitors, BTK inhibitors, anti-lymphocyte activating gene-3 monoclonal antibodies, CXCL/CXCR inhibitors, FAK inhibitors, T cell immunoglobulin-related agents, and adoptive cell transfer therapy (so-called 'CAR T-cell therapy'). Although most of these trials are still ongoing, combined strategies using different therapeutic approaches with ICIs may provide supporting evidence to optimize PC treatment using ICIs (137, 147).

DISCUSSION AND PERSPECTIVE

Despite substantial advances in the development of new agents and an improved understanding of PC genetics, cytotoxicbackboned chemotherapy remains the key treatment with only a few novel agents being translated into clinical practice. Current research is focused on targeting the specific component of the PC tumour stroma, yet the results so far are disappointing, and there have been few significant advanced in antiangiogenic therapy or immunotherapy. The PC microenvironment is characterized by increased desmoplasia and energy metabolic disorders (28). Plus, accumulating evidence has revealed that the PC stroma is enriched with CAFs or PSCs that produce excessive amounts stromal elements and a matrix, leading to a desmoplastic process (59). These components are also responsible for the generation of a rigid barrier that results in elevated tumour pressure, a hypovascular microenvironment, and attenuated drug delivery (17). A rich stroma and severe fibrotic reaction have also been shown to play a role in remodelling the TME (28). In addition to the mechanical factors, TME alterations in PC can also support the metabolism of cancer cells, even in a nutrient-depleted and hypoxic TME. In particular, an amplified desmoplastic reaction can significantly impair the immune response and augment the immune tolerance (8). Thus, recent data also emphasizes the importance of multiple targeted approaches, including stromal depletion and immune modulation. Therefore, proactively designed combination strategies are needed for more successful development of stroma-targeting agents. Plus, an improved understanding of the interaction complexity between tumour cells and the heterogeneous TME components will help to identify stromal-based biomarkers for categorizing patients who can receive clinical benefit in PC.

Molecular classification could also help to improve patient selection and enable better differentiation of responders and non-responders. For example, as an immunogenic subtype exhibits an upregulation of immune-related genes, this subtype

REFERENCES

- 1. Ryan DP, Hong TS, Bardeesy N. Pancreatic Adenocarcinoma. N Engl J Med (2014) 371:1039–49. doi: 10.1056/NEJMra1404198
- Cid-Arregui A, Juarez V. Perspectives in the Treatment of Pancreatic Adenocarcinoma. World J Gastroenterol (2015) 21:9297–316. doi: 10.3748/ wjgv21.i31.9297
- Conroy T, Desseigne F, Ychou M, Bouché O, Guimbaud R, Bécouarn Y, et al. FOLFIRINOX Versus Gemcitabine for Metastatic Pancreatic Cancer. *N Engl J Med* (2011) 364:1817–25. doi: 10.1056/NEJMoa1011923
- Braiteh F, Patel MB, Parisi M, Ni Q, Park S, Faria C. Comparative Effectiveness and Resource Utilization of Nab-Paclitaxel Plus Gemcitabine vs FOLFIRINOX or Gemcitabine for the First-Line Treatment of Metastatic Pancreatic Adenocarcinoma in a US Community Setting. *Cancer Manag Res* (2017) 9:141–8. doi: 10.2147/CMAR.S126073
- Von Hoff DD, Ervin T, Arena FP, Chiorean EG, Infante J, Moore M, et al. Increased Survival in Pancreatic Cancer With Nab-Paclitaxel Plus Gemcitabine. *N Engl J Med* (2013) 369:1691–703. doi: 10.1056/NEJMoa1304369
- Veenstra VL, Garcia-Garijo A, van Laarhoven HW, Bijlsma MF. Extracellular Influences: Molecular Subclasses and the Microenvironment in Pancreatic Cancer. *Cancers (Basel)* (2018) 10:34. doi: 10.3390/cancers10020034
- Murakami T, Hiroshima Y, Matsuyama R, Homma Y, Hoffman RM, Endo I. Role of the Tumor Microenvironment in Pancreatic Cancer. Ann Gastroenterol Surg (2019) 3:130–7. doi: 10.1002/ags3.12225
- Vennin C, Murphy KJ, Morton JP, Cox TR, Pajic M, Timpson P. Reshaping the Tumor Stroma for Treatment of Pancreatic Cancer. *Gastroenterology* (2018) 154:820–38. doi: 10.1053/j.gastro.2017.11.280
- Lafaro KJ, Melstrom LG. The Paradoxical Web of Pancreatic Cancer Tumor Microenvironment. Am J Pathol (2019) 189:44–57. doi: 10.1016/j.ajpath.2018.09.009
- Brunner M, Wu Z, Krautz C, Pilarsky C, Grützmann R, Weber GF. Current Clinical Strategies of Pancreatic Cancer Treatment and Open Molecular Questions. Int J Mol Sci (2019) 20:4543. doi: 10.3390/ijms20184543

could feasibly be treated with ICIs (148). Meanwhile, a basal-like subtype may benefit more from gemcitabine-based regimens than from FOLFIRINOX (149). Plus, the presence of pancreatic stellate cells in an ADEX subtype could also imply resistance to gemcitabine or radiotherapy (150). More recently, Tiriac et al. performed drug sensitivity profiling of PC organoid models and identified novel functional subtypes for defining gene expression signatures to predict chemotherapy sensitivity (151). Although further studies are required to validate these results in PC, designing and testing novel specific targeted therapies for each subtype may help to avoid unnecessary treatments and expedite the application of an effective drug combination.

AUTHOR CONTRIBUTIONS

PE and BK drafted the manuscript. IC managed and reviewed the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

PE and IC would like to acknowledge National Health Service funding to the National Institute for Health Research Biomedical Research Centre at the Royal Marsden NHS Foundation Trust and The Institute of Cancer Research.

- Lai E, Puzzoni M, Ziranu P, Pretta A, Impera V, Mariani S, et al. New Therapeutic Targets in Pancreatic Cancer. *Cancer Treat Rev* (2019) 81:101926. doi: 10.1016/j.ctrv.2019.101926
- Hilgers W, Kern SE. Molecular Genetic Basis of Pancreatic Adenocarcinoma. Genes Chromosomes Cancer (1999) 26:1–12. doi: 10.1002/(SICI)1098-2264 (199909)26:1<1::AID-GCC1>3.0.CO;2-X
- Canto MI, Hruban RH, Fishman EK, Kamel IR, Schulick R, Zhang Z, et al. Frequent Detection of Pancreatic Lesions in Asymptomatic High-Risk Individuals. *Gastroenterology* (2012) 142:796–e15. doi: 10.1053/j.gastro. 2012.01.005
- Bailey P, Chang DK, Nones K, Johns AL, Patch A-M, Gingras M-C, et al. Genomic Analyses Identify Molecular Subtypes of Pancreatic Cancer. *Nature* (2016) 531:47–52. doi: 10.1038/nature16965
- Maurer C, Holmstrom SR, He J, Laise P, Su T, Ahmed A, et al. Experimental Microdissection Enables Functional Harmonisation of Pancreatic Cancer Subtypes. *Gut* (2019) 68:1034–43. doi: 10.1136/gutjnl-2018-317706
- Abbassi R, Schmid RM. Evolving Treatment Paradigms for Pancreatic Cancer. Visc Med (2019) 35:362–72. doi: 10.1159/000503797
- Sarantis P, Koustas E, Papadimitropoulou A, Papavassiliou AG, Karamouzis MV. Pancreatic Ductal Adenocarcinoma: Treatment Hurdles, Tumor Microenvironment and Immunotherapy. World J Gastrointest Oncol (2020) 12:173–81. doi: 10.4251/wjgo.v12.i2.173
- Storz P, Crawford HC. Carcinogenesis of Pancreatic Ductal Adenocarcinoma. Gastroenterology (2020) 158:2072–81. doi: 10.1053/j.gastro.2020.02.059
- von Ahrens D, Bhagat TD, Nagrath D, Maitra A, Verma A. The Role of Stromal Cancer-Associated Fibroblasts in Pancreatic Cancer. J Hematol Oncol (2017) 10:76. doi: 10.1186/s13045-017-0448-5
- Omary MB, Lugea A, Lowe AW, Pandol SJ. The Pancreatic Stellate Cell: A Star on the Rise in Pancreatic Diseases. J Clin Invest (2007) 117:50–9. doi: 10.1172/JCI30082
- Chen X, Song E. Turning Foes to Friends: Targeting Cancer-Associated Fibroblasts. Nat Rev Drug Discov (2019) 18:99–115. doi: 10.1038/s41573-018-0004-1

- Jotzu C, Alt E, Welte G, Li J, Hennessy BT, Devarajan E, et al. Adipose Tissue Derived Stem Cells Differentiate Into Carcinoma-Associated Fibroblast-Like Cells Under the Influence of Tumor Derived Factors. *Cell Oncol* (2011) 34:55–67. doi: 10.1007/s13402-011-0012-1
- Dulauroy S, Di Carlo SE, Langa F, Eberl G, Peduto L. Lineage Tracing and Genetic Ablation of ADAM12(+) Perivascular Cells Identify a Major Source of Profibrotic Cells During Acute Tissue Injury. *Nat Med* (2012) 18:1262–70. doi: 10.1038/nm.2848
- Iwano M, Plieth D, Danoff TM, Xue C, Okada H, Neilson EG. Evidence That Fibroblasts Derive From Epithelium During Tissue Fibrosis. J Clin Invest (2002) 110:341–50. doi: 10.1172/JCI0215518
- Zeisberg EM, Potenta S, Xie L, Zeisberg M, Kalluri R. Discovery of Endothelial to Mesenchymal Transition as a Source for Carcinoma-Associated Fibroblasts. *Cancer Res* (2007) 67:10123–8. doi: 10.1158/0008-5472.CAN-07-3127
- Awaji M, Singh RK. Cancer-Associated Fibroblasts' Functional Heterogeneity in Pancreatic Ductal Adenocarcinoma. *Cancers (Basel)* (2019) 11:29. doi: 10.3390/cancers11030290
- Sunami Y, Böker V, Kleeff J. Targeting and Reprograming Cancer-Associated Fibroblasts and the Tumor Microenvironment in Pancreatic Cancer. *Cancers (Basel)* (2021) 13:697. doi: 10.3390/cancers13040697
- Wu Y, Zhang C, Jiang K, Werner J, Bazhin AV, D'Haese JG. The Role of Stellate Cells in Pancreatic Ductal Adenocarcinoma: Targeting Perspectives. *Front Oncol* (2020) 10:621937. doi: 10.3389/fonc.2020.621937
- Sahai E, Astsaturov I, Cukierman E, DeNardo DG, Egeblad M, Evans RM, et al. A Framework for Advancing Our Understanding of Cancer-Associated Fibroblasts. *Nat Rev Cancer* (2020) 20:174–86. doi: 10.1038/s41568-019-0238-1
- Zeng S, Pöttler M, Lan B, Grützmann R, Pilarsky C, Yang H. Chemoresistance in Pancreatic Cancer. Int J Mol Sci (2019) 20:4504. doi: 10.3390/ijms20184504
- Musa M. Single-Cell Analysis on Stromal Fibroblasts in the Microenvironment of Solid Tumours. Adv Med Sci (2020) 65:163–9. doi: 10.1016/j.advms.2019.12.001
- Öhlund D, Handly-Santana A, Biffi G, Elyada E, Almeida AS, Ponz-Sarvise M, et al. Distinct Populations of Inflammatory Fibroblasts and Myofibroblasts in Pancreatic Cancer. J Exp Med (2017) 214:579–96. doi: 10.1084/jem.20162024
- Thomas D, Radhakrishnan P. Tumor-Stromal Crosstalk in Pancreatic Cancer and Tissue Fibrosis. *Mol Cancer* (2019) 18:14. doi: 10.1186/ s12943-018-0927-5
- Sonbol HS. Extracellular Matrix Remodeling in Human Disease. J Microsc Ultrastruct (2018) 6:123–8. doi: 10.4103/JMAU.JMAU_4_18
- Xu S, Xu H, Wang W, Li S, Li H, Li T, et al. The Role of Collagen in Cancer: From Bench to Bedside. J Transl Med (2019) 17:309–. doi: 10.1186/s12967-019-2058-1
- Sato N, Cheng X-B, Kohi S, Koga A, Hirata K. Targeting Hyaluronan for the Treatment of Pancreatic Ductal Adenocarcinoma. *Acta Pharm Sin B* (2016) 6:101–5. doi: 10.1016/j.apsb.2016.01.002
- Sato N, Kohi S, Hirata K, Goggins M. Role of Hyaluronan in Pancreatic Cancer Biology and Therapy: Once Again in the Spotlight. *Cancer Sci* (2016) 107:569–75. doi: 10.1111/cas.12913
- Kanat O, Ertas H. Shattering the Castle Walls: Anti-Stromal Therapy for Pancreatic Cancer. World J Gastrointest Oncol (2018) 10:202–10. doi: 10.4251/wjgo.v10.i8.202
- 39. Kultti A, Zhao C, Singha NC, Zimmerman S, Osgood RJ, Symons R, et al. Accumulation of Extracellular Hyaluronan by Hyaluronan Synthase 3 Promotes Tumor Growth and Modulates the Pancreatic Cancer Microenvironment. *BioMed Res Int* (2014) 2014:817613. doi: 10.1155/ 2014/817613
- 40. Koyama H, Hibi T, Isogai Z, Yoneda M, Fujimori M, Amano J, et al. Hyperproduction of Hyaluronan in Neu-Induced Mammary Tumor Accelerates Angiogenesis Through Stromal Cell Recruitment: Possible Involvement of Versican/PG-M. Am J Pathol (2007) 170:1086–99. doi: 10.2353/ajpath.2007.060793
- Vaz J, Ansari D, Sasor A, Andersson R. SPARC: A Potential Prognostic and Therapeutic Target in Pancreatic Cancer. *Pancreas* (2015) 44:1024–35. doi: 10.1097/MPA.000000000000409

- Melstrom LG, Salazar MD, Diamond DJ. The Pancreatic Cancer Microenvironment: A True Double Agent. J Surg Oncol (2017) 116:7–15. doi: 10.1002/jso.24643
- Whatcott CJ, Han H, Von Hoff DD. Orchestrating the Tumor Microenvironment to Improve Survival for Patients With Pancreatic Cancer: Normalization, Not Destruction. *Cancer J* (2015) 21:299–306. doi: 10.1097/PPO.00000000000140
- Erkan M, Kurtoglu M, Kleeff J. The Role of Hypoxia in Pancreatic Cancer: A Potential Therapeutic Target? *Expert Rev Gastroenterol Hepatol* (2016) 10:301–16. doi: 10.1586/17474124.2016.1117386
- Longo V, Brunetti O, Gnoni A, Cascinu S, Gasparini G, Lorusso V, et al. Angiogenesis in Pancreatic Ductal Adenocarcinoma: A Controversial Issue. Oncotarget (2016) 7:58649–58. doi: 10.18632/oncotarget.10765
- Daniel SK, Sullivan KM, Labadie KP, Pillarisetty VG. Hypoxia as a Barrier to Immunotherapy in Pancreatic Adenocarcinoma. *Clin Transl Med* (2019) 8:10. doi: 10.1186/s40169-019-0226-9
- Provenzano PP, Cuevas C, Chang AE, Goel VK, Von Hoff DD, Hingorani SR. Enzymatic Targeting of the Stroma Ablates Physical Barriers to Treatment of Pancreatic Ductal Adenocarcinoma. *Cancer Cell* (2012) 21:418–29. doi: 10.1016/j.ccr.2012.01.007
- Berchtold S, Grünwald B, Krüger A, Reithmeier A, Hähl T, Cheng T, et al. Collagen Type V Promotes the Malignant Phenotype of Pancreatic Ductal Adenocarcinoma. *Cancer Lett* (2015) 356:721–32. doi: 10.1016/j.canlet. 2014.10.020
- Erkan M, Adler G, Apte MV, Bachem MG, Buchholz M, Detlefsen S, et al. StellaTUM: Current Consensus and Discussion on Pancreatic Stellate Cell Research. *Gut* (2012) 61:172–8. doi: 10.1136/gutjnl-2011-301220
- Ijichi H, Chytil A, Gorska AE, Aakre ME, Bierie B, Tada M, et al. Inhibiting Cxcr2 Disrupts Tumor-Stromal Interactions and Improves Survival in a Mouse Model of Pancreatic Ductal Adenocarcinoma. J Clin Invest (2011) 121:4106–17. doi: 10.1172/JCI42754
- Kalbasi A, Komar C, Tooker GM, Liu M, Lee JW, Gladney WL, et al. Tumor-Derived CCL2 Mediates Resistance to Radiotherapy in Pancreatic Ductal Adenocarcinoma. *Clin Cancer Res* (2017) 23:137–48. doi: 10.1158/1078-0432.CCR-16-0870
- Komar G, Kauhanen S, Liukko K, Seppänen M, Kajander S, Ovaska J, et al. Decreased Blood Flow With Increased Metabolic Activity: A Novel Sign of Pancreatic Tumor Aggressiveness. *Clin Cancer Res* (2009) 15:5511–7. doi: 10.1158/1078-0432.CCR-09-0414
- 53. McCarty MF, Somcio RJ, Stoeltzing O, Wey J, Fan F, Liu W, et al. Overexpression of PDGF-BB Decreases Colorectal and Pancreatic Cancer Growth by Increasing Tumor Pericyte Content. J Clin Invest (2007) 117:2114–22. doi: 10.1172/JCI31334
- 54. Büchler P, Reber HA, Büchler MW, Friess H, Lavey RS, Hines OJ. Antiangiogenic Activity of Genistein in Pancreatic Carcinoma Cells is Mediated by the Inhibition of Hypoxia-Inducible Factor-1 and the Down-Regulation of VEGF Gene Expression. *Cancer* (2004) 100:201–10. doi: 10.1002/cncr.11873
- 55. Shibaji T, Nagao M, Ikeda N, Kanehiro H, Hisanaga M, Ko S, et al. Prognostic Significance of HIF-1 Alpha Overexpression in Human Pancreatic Cancer. Anticancer Res (2003) 23:4721–7.
- Li S, Xu H-X, Wu C-T, Wang W-Q, Jin W, Gao H-L, et al. Angiogenesis in Pancreatic Cancer: Current Research Status and Clinical Implications. *Angiogenesis* (2019) 22:15–36. doi: 10.1007/s10456-018-9645-2
- Yao W, Maitra A, Ying H. Recent Insights Into the Biology of Pancreatic Cancer. *EBioMedicine* (2020) 53:102655. doi: 10.1016/j.ebiom.2020.102655
- Upadhrasta S, Zheng L. Strategies in Developing Immunotherapy for Pancreatic Cancer: Recognizing and Correcting Multiple Immune "Defects" in the Tumor Microenvironment. J Clin Med (2019) 8:1472. doi: 10.3390/jcm8091472
- Uzunparmak B, Sahin IH. Pancreatic Cancer Microenvironment: A Current Dilemma. Clin Transl Med (2019) 8:2–. doi: 10.1186/s40169-019-0221-1
- Ene-Obong A, Clear AJ, Watt J, Wang J, Fatah R, Riches JC, et al. Activated Pancreatic Stellate Cells Sequester CD8+ T Cells to Reduce Their Infiltration of the Juxtatumoral Compartment of Pancreatic Ductal Adenocarcinoma. *Gastroenterology* (2013) 145:1121–32. doi: 10.1053/j.gastro.2013.07.025
- Li J, Byrne KT, Yan F, Yamazoe T, Chen Z, Baslan T, et al. Tumor Cell-Intrinsic Factors Underlie Heterogeneity of Immune Cell Infiltration and

Response to Immunotherapy. *Immunity* (2018) 49:178–93.e7. doi: 10.1016/j.immuni.2018.06.006

- 62. Hingorani SR, Harris WP, Beck JT, Berdov BA, Wagner SA, Pshevlotsky EM, et al. Phase Ib Study of PEGylated Recombinant Human Hyaluronidase and Gemcitabine in Patients With Advanced Pancreatic Cancer. *Clin Cancer Res* (2016) 22:2848–54. doi: 10.1158/1078-0432.CCR-15-2010
- 63. Tempero MA, Cutsem EV, Sigal D, Oh D-Y, Fazio N, Macarulla T, et al. HALO 109-301: A Randomized, Double-Blind, Placebo-Controlled, Phase 3 Study of Pegvorhyaluronidase Alfa (PEGPH20) + Nab-Paclitaxel/Gemcitabine (AG) in Patients (Pts) With Previously Untreated Hyaluronan (HA)-High Metastatic Pancreatic Ductal Adenocarcinoma (mPDA). J Clin Oncol (2020) 38:638. doi: 10.1200/JCO.2020.38.4_suppl.638
- 64. Ramanathan RK, McDonough SL, Philip PA, Hingorani SR, Lacy J, Kortmansky JS, et al. Phase IB/II Randomized Study of FOLFIRINOX Plus Pegylated Recombinant Human Hyaluronidase Versus FOLFIRINOX Alone in Patients With Metastatic Pancreatic Adenocarcinoma: SWOG S1313. J Clin Oncol (2019) 37:1062–9. doi: 10.1200/JCO.18.01295
- 65. Picozzi VJ, Pipas JM, Koong A, Giaccia A, Bahary N, Krishnamurthi SS, et al. FG-3019, a Human Monoclonal Antibody to Connective Tissue Growth Factor (CTGF), With Gemcitabine/Erlotinib (G/E) in Patients With Locally Advanced or Metastatic Pancreatic Ductal Adenocarcinoma (PDAC). J Clin Oncol (2014) 32:4138. doi: 10.1200/jco.2014.32.15_suppl.4138
- 66. De Jesus-Acosta A, Sugar EA, O'Dwyer PJ, Ramanathan RK, Von Hoff DD, Rasheed Z, et al. Phase 2 Study of Vismodegib, a Hedgehog Inhibitor, Combined With Gemcitabine and Nab-Paclitaxel in Patients With Untreated Metastatic Pancreatic Adenocarcinoma. Br J Cancer (2020) 122:498–505. doi: 10.1038/s41416-019-0683-3
- 67. Zhen DB, Griffith KA, Ruch JM, Camphausen K, Savage JE, Kim EJ, et al. A Phase I Trial of Cabozantinib and Gemcitabine in Advanced Pancreatic Cancer. *Invest New Drugs* (2016) 34:733–9. doi: 10.1007/s10637-016-0376-1
- Von Hoff DD, Ramanathan RK, Borad MJ, Laheru DA, Smith LS, Wood TE, et al. Gemcitabine Plus Nab-Paclitaxel is an Active Regimen in Patients With Advanced Pancreatic Cancer: A Phase I/II Trial. J Clin Oncol (2011) 29:4548–54. doi: 10.1200/JCO.2011.36.5742
- 69. Tempero M, Oh D, Macarulla T, Reni M, Van Cutsem E, Hendifar A, et al. Ibrutinib in Combination With Nab-Paclitaxel and Gemcitabine as First-Line Treatment for Patients With Metastatic Pancreatic Adenocarcinoma: Results From the Phase 3 RESOLVE Study. Ann Oncol (2019) 30:iv126. doi: 10.1093/annonc/mdz154.001
- Cutsem EV, Vervenne WL, Bennouna J, Humblet Y, Gill S, Laethem J-LV, et al. Phase III Trial of Bevacizumab in Combination With Gemcitabine and Erlotinib in Patients With Metastatic Pancreatic Cancer. J Clin Oncol (2009) 27:2231–7. doi: 10.1200/JCO.2008.20.0238
- 71. Kindler HL, Niedzwiecki D, Hollis D, Sutherland S, Schrag D, Hurwitz H, et al. Gemcitabine Plus Bevacizumab Compared With Gemcitabine Plus Placebo in Patients With Advanced Pancreatic Cancer: Phase III Trial of the Cancer and Leukemia Group B (CALGB 80303). J Clin Oncol (2010) 28:3617–22. doi: 10.1200/JCO.2010.28.1386
- 72. Kindler HL, Ioka T, Richel DJ, Bennouna J, Létourneau R, Okusaka T, et al. Axitinib Plus Gemcitabine Versus Placebo Plus Gemcitabine in Patients With Advanced Pancreatic Adenocarcinoma: A Double-Blind Randomised Phase 3 Study. *Lancet Oncol* (2011) 12:256–62. doi: 10.1016/S1470-2045(11)70004-3
- 73. Gonçalves A, Gilabert M, François E, Dahan L, Perrier H, Lamy R, et al. BAYPAN Study: A Double-Blind Phase III Randomized Trial Comparing Gemcitabine Plus Sorafenib and Gemcitabine Plus Placebo in Patients With Advanced Pancreatic Cancer. Ann Oncol (2012) 23:2799–805. doi: 10.1093/ annonc/mds135
- 74. Rougier P, Riess H, Manges R, Karasek P, Humblet Y, Barone C, et al. Randomised, Placebo-Controlled, Double-Blind, Parallel-Group Phase III Study Evaluating Aflibercept in Patients Receiving First-Line Treatment With Gemcitabine for Metastatic Pancreatic Cancer. *Eur J Cancer* (2013) 49:2633–42. doi: 10.1016/j.ejca.2013.04.002
- 75. Le DT, Wang-Gillam A, Picozzi V, Greten TF, Crocenzi T, Springett G, et al. Safety and Survival With GVAX Pancreas Prime and Listeria Monocytogenes-Expressing Mesothelin (CRS-207) Boost Vaccines for Metastatic Pancreatic Cancer. J Clin Oncol (2015) 33:1325–33. doi: 10.1200/JCO.2014.57.4244
- 76. Royal RE, Levy C, Turner K, Mathur A, Hughes M, Kammula US, et al. Phase 2 Trial of Single Agent Ipilimumab (Anti-CTLA-4) for Locally

Advanced or Metastatic Pancreatic Adenocarcinoma. J Immunother (2010) 33:828–33. doi: 10.1097/CJI.0b013e3181eec14c

- 77. O'Reilly EM, Oh D-Y, Dhani N, Renouf DJ, Lee MA, Sun W, et al. Durvalumab With or Without Tremelimumab for Patients With Metastatic Pancreatic Ductal Adenocarcinoma: A Phase 2 Randomized Clinical Trial. JAMA Oncol (2019) 5:1431–8. doi: 10.1001/jamaoncol.2019.1588
- Weiss GJ, Blaydorn L, Beck J, Bornemann-Kolatzki K, Urnovitz H, Schütz E, et al. Phase Ib/II Study of Gemcitabine, Nab-Paclitaxel, and Pembrolizumab in Metastatic Pancreatic Adenocarcinoma. *Invest New Drugs* (2018) 36:96– 102. doi: 10.1007/s10637-017-0525-1
- Seufferlein T, Ducreux M, Hidalgo M, Prager G, Cutsem E. More Than a Gel & Hyaluronic Acid, a Central Component in the Microenvironment of Pancreatic Cancer. *Eur Oncol Haematol* (2018) 14:40. doi: 10.17925/ EOH.2018.14.1.40
- Thompson CB, Shepard HM, O'Connor PM, Kadhim S, Jiang P, Osgood RJ, et al. Enzymatic Depletion of Tumor Hyaluronan Induces Antitumor Responses in Preclinical Animal Models. *Mol Cancer Ther* (2010) 9:3052– 64. doi: 10.1158/1535-7163.MCT-10-0470
- Jacobetz MA, Chan DS, Neesse A, Bapiro TE, Cook N, Frese KK, et al. Hyaluronan Impairs Vascular Function and Drug Delivery in a Mouse Model of Pancreatic Cancer. *Gut* (2013) 62:112–20. doi: 10.1136/gutjnl-2012-302529
- 82. Hingorani SR, Zheng L, Bullock AJ, Seery TE, Harris WP, Sigal DS, et al. HALO 202: Randomized Phase II Study of PEGPH20 Plus Nab-Paclitaxel/ Gemcitabine Versus Nab-Paclitaxel/Gemcitabine in Patients With Untreated, Metastatic Pancreatic Ductal Adenocarcinoma. *J Clin Oncol* (2017) 36:359–66. doi: 10.1200/JCO.2017.74.9564
- Doherty GJ, Tempero M, Corrie PG. HALO-109–301: A Phase III Trial of PEGPH20 (With Gemcitabine and Nab-Paclitaxel) in Hyaluronic Acid-High Stage IV Pancreatic Cancer. *Future Oncol* (2018) 14:13–22. doi: 10.2217/fon-2017-0338
- Hakim N, Patel R, Devoe C, Saif MW. Why HALO 301 Failed and Implications for Treatment of Pancreatic Cancer. *Pancreas (Fairfax)* (2019) 3:e1-4. doi: 10.17140/POJ-3-e010
- 85. Lee J, Wilkinson GA, Kimbler T, Blouw B, Thompson CB. Abstract 2206: Degradation of Hyaluronan in the Tumor Microenvironment Enhances Oncolytic Reovirus and Anti-PD-L1 Therapy in a Murine Syngeneic Breast Tumor Model. *Cancer Res* (2020) 80:2206. doi: 10.1158/1538-7445.AM2020-2206
- Singha NC, Nekoroski T, Zhao C, Symons R, Jiang P, Frost GI, et al. Tumor-Associated Hyaluronan Limits Efficacy of Monoclonal Antibody Therapy. *Mol Cancer Ther* (2015) 14:523–32. doi: 10.1158/1535-7163.MCT-14-0580
- Abreu JG, Ketpura NI, Reversade B, De Robertis EM. Connective-Tissue Growth Factor (CTGF) Modulates Cell Signalling by BMP and TGF-Beta. *Nat Cell Biol* (2002) 4:599–604. doi: 10.1038/ncb826
- Heng ECK, Huang Y, Black SA Jr, Trackman PC. CCN2, Connective Tissue Growth Factor, Stimulates Collagen Deposition by Gingival Fibroblasts *via* Module 3 and Alpha6- and Beta1 Integrins. *J Cell Biochem* (2006) 98:409–20. doi: 10.1002/jcb.20810
- Neesse A, Frese KK, Bapiro TE, Nakagawa T, Sternlicht MD, Seeley TW, et al. CTGF Antagonism With mAb FG-3019 Enhances Chemotherapy Response Without Increasing Drug Delivery in Murine Ductal Pancreas Cancer. Proc Natl Acad Sci U.S.A. (2013) 110:12325–30. doi: 10.1073/pnas.1300415110
- 90. Picozzi VJ, Pishvaian MJ, Mody K, Winter JM, Glaspy JA, Larson T, et al. Effect of Anti-CTGF Human Recombinant Monoclonal Antibody Pamrevlumab on Resectability and Resection Rate When Combined With Gemcitabine/Nab-Paclitaxel in Phase 1/2 Clinical Study for the Treatment of Locally Advanced Pancreatic Cancer Patients. J Clin Oncol (2018) 36:4016-. doi: 10.1200/JCO.2018.36.15_suppl.4016
- Gu J, Saiyin H, Fu D, Li J. Stroma A Double-Edged Sword in Pancreatic Cancer: A Lesson From Targeting Stroma in Pancreatic Cancer With Hedgehog Signaling Inhibitors. *Pancreas* (2018) 47:382–9. doi: 10.1097/ MPA.000000000001023
- Jones S, Zhang X, Parsons DW, Lin JC-H, Leary RJ, Angenendt P, et al. Core Signaling Pathways in Human Pancreatic Cancers Revealed by Global Genomic Analyses. *Science* (2008) 321:1801–6. doi: 10.1126/science.1164368
- Cortes JE, Gutzmer R, Kieran MW, Solomon JA. Hedgehog Signaling Inhibitors in Solid and Hematological Cancers. *Cancer Treat Rev* (2019) 76:41–50. doi: 10.1016/j.ctrv.2019.04.005

- Olive KP, Jacobetz MA, Davidson CJ, Gopinathan A, McIntyre D, Honess D, et al. Inhibition of Hedgehog Signaling Enhances Delivery of Chemotherapy in a Mouse Model of Pancreatic Cancer. *Science* (2009) 324:1457–61. doi: 10.1126/science.1171362
- 95. Richards DA, Stephenson J, Wolpin BM, Becerra C, Hamm JT, Messersmith WA, et al. A Phase Ib Trial of IPI-926, a Hedgehog Pathway Inhibitor, Plus Gemcitabine in Patients With Metastatic Pancreatic Cancer. J Clin Oncol (2012) 30:213. doi: 10.1200/jco.2012.30.4_suppl.213
- 96. Ko AH, LoConte NK, Kantoff E, Ross RW, Trehu EG, Tempero MA, et al. A Phase Ib Trial of FOLFIRINOX Plus Saridegib, an Oral Hedgehog (Hh) Inhibitor, in Pts With Advanced Pancreatic Cancer (PDAC). *Pancreas* (2012) 30:3105–. doi: 10.1200/jco.2012.30.15_suppl.3105
- McCleary-Wheeler AL, Carr RM, Palmer SR, Smyrk TC, Allred JB, Almada LL, et al. Phase 1 Trial of Vismodegib and Erlotinib Combination in Metastatic Pancreatic Cancer. *Pancreatology* (2020) 20:101–9. doi: 10. 1016/j.pan.2019.11.011
- Parkin A, Man J, Timpson P, Pajic M. Targeting the Complexity of Src Signalling in the Tumour Microenvironment of Pancreatic Cancer: From Mechanism to Therapy. *FEBS J* (2019) 286:3510–39. doi: 10.1111/febs.15011
- 99. Kanteti R, Mirzapoiazova T, Riehm JJ, Dhanasingh I, Mambetsariev B, Wang J, et al. Focal Adhesion Kinase a Potential Therapeutic Target for Pancreatic Cancer and Malignant Pleural Mesothelioma. *Cancer Biol Ther* (2018) 19:316–27. doi: 10.1080/15384047.2017.1416937
- 100. Jiang H, Hegde S, Knolhoff BL, Zhu Y, Herndon JM, Meyer MA, et al. Targeting Focal Adhesion Kinase Renders Pancreatic Cancers Responsive to Checkpoint Immunotherapy. Nat Med (2016) 22:851–60. doi: 10.1038/nm.4123
- 101. Weniger M, Honselmann KC, Liss AS. The Extracellular Matrix and Pancreatic Cancer: A Complex Relationship. *Cancers (Basel)* (2018) 10:316. doi: 10.3390/cancers10090316
- 102. Wang-Gillam A, Lockhart AC, Tan BR, Suresh R, Lim K-H, Ratner L, et al. Phase I Study of Defactinib Combined With Pembrolizumab and Gemcitabine in Patients With Advanced Cancer. J Clin Oncol (2018) 36:380. doi: 10.1200/JCO.2018.36.4_suppl.380
- 103. Osipov A, Saung MT, Zheng L, Murphy AG. Small Molecule Immunomodulation: The Tumor Microenvironment and Overcoming Immune Escape. J Immunother Cancer (2019) 7:224. doi: 10.1186/s40425-019-0667-0
- 104. Moosavi F, Giovannetti E, Peters GJ, Firuzi O. Combination of HGF/MET-Targeting Agents and Other Therapeutic Strategies in Cancer. Crit Rev Oncol/Hematol (2021) 160:103234. doi: 10.1016/j.critrevonc.2021.103234
- Pothula SP, Xu Z, Goldstein D, Pirola RC, Wilson JS, Apte MV. Targeting HGF/c-MET Axis in Pancreatic Cancer. Int J Mol Sci (2020) 21:9170. doi: 10.3390/ijms21239170
- 106. Yu J, Ohuchida K, Mizumoto K, Ishikawa N, Ogura Y, Yamada D, et al. Overexpression of C-Met in the Early Stage of Pancreatic Carcinogenesis; Altered Expression Is Not Sufficient for Progression From Chronic Pancreatitis to Pancreatic Cancer. World J Gastroenterol (2006) 12:3878– 82. doi: 10.3748/wjg.v12.i24.3878
- 107. Ueda T, Takeyama Y, Hori Y, Nishikawa J, Yamamoto M, Saitoh Y. Hepatocyte Growth Factor in Assessment of Acute Pancreatitis: Comparison With C-Reactive Protein and Interleukin-6. J Gastroenterol (1997) 32:63-70. doi: 10.1007/BF01213298
- 108. Hage C, Rausch V, Giese N, Giese T, Schönsiegel F, Labsch S, et al. The Novel C-Met Inhibitor Cabozantinib Overcomes Gemcitabine Resistance and Stem Cell Signaling in Pancreatic Cancer. *Cell Death Dis* (2013) 4:e627. doi: 10.1038/cddis.2013.158
- 109. Avan A, Caretti V, Funel N, Galvani E, Maftouh M, Honeywell RJ, et al. Crizotinib Inhibits Metabolic Inactivation of Gemcitabine in C-Met-Driven Pancreatic Carcinoma. *Cancer Res* (2013) 73:6745–56. doi: 10.1158/0008-5472.CAN-13-0837
- 110. Takiguchi S, Inoue K, Matsusue K, Furukawa M, Teramoto N, Iguchi H. Crizotinib, a MET Inhibitor, Prevents Peritoneal Dissemination in Pancreatic Cancer. Int J Oncol (2017) 51:184–92. doi: 10.3892/ijo.2017.3992
- Rossi MK, Gnanamony M, Gondi CS. The 'Sparc' of Life: Analysis of the Role of Osteonectin/SPARC in Pancreatic Cancer (Review). *Int J Oncol* (2016) 48:1765–71. doi: 10.3892/ijo.2016.3417
- 112. Giordano G, Pancione M, Olivieri N, Parcesepe P, Velocci M, Di Raimo T, et al. Nano Albumin Bound-Paclitaxel in Pancreatic Cancer: Current

Evidences and Future Directions. World J Gastroenterol (2017) 23:5875-86. doi: 10.3748/wjg.v23.i32.5875

- 113. Han W, Cao F, Chen M-b, Lu R-z, Wang H-b, Yu M, et al. Prognostic Value of SPARC in Patients With Pancreatic Cancer: A Systematic Review and Meta-Analysis. *PloS One* (2016) 11:e0145803–e. doi: 10.1371/journal.pone.0145803
- 114. Zhang DS, Wang DS, Wang ZQ, Wang FH, Luo HY, Qiu MZ, et al. Phase I/II Study of Albumin-Bound Nab-Paclitaxel Plus Gemcitabine Administered to Chinese Patients With Advanced Pancreatic Cancer. *Cancer Chemother Pharmacol* (2013) 71:1065–72. doi: 10.1007/s00280-013-2102-4
- 115. Hidalgo M, Plaza C, Musteanu M, Illei P, Brachmann CB, Heise C, et al. SPARC Expression Did Not Predict Efficacy of Nab-Paclitaxel Plus Gemcitabine or Gemcitabine Alone for Metastatic Pancreatic Cancer in an Exploratory Analysis of the Phase III MPACT Trial. *Clin Cancer Res* (2015) 21:4811–8. doi: 10.1158/1078-0432.CCR-14-3222
- 116. Tabernero J, Chiorean EG, Infante JR, Hingorani SR, Ganju V, Weekes C, et al. Prognostic Factors of Survival in a Randomized Phase III Trial (MPACT) of Weekly Nab-Paclitaxel Plus Gemcitabine Versus Gemcitabine Alone in Patients With Metastatic Pancreatic Cancer. Oncologist (2015) 20:143–50. doi: 10.1634/theoncologist.2014-0394
- 117. Kim H, Samuel S, Lopez-Casas P, Grizzle W, Hidalgo M, Kovar J, et al. SPARC-Independent Delivery of Nab-Paclitaxel Without Depleting Tumor Stroma in Patient-Derived Pancreatic Cancer Xenografts. *Mol Cancer Ther* (2016) 15:680–8. doi: 10.1158/1535-7163.MCT-15-0764
- Cullis J, Siolas D, Avanzi A, Barui S, Maitra A, Bar-Sagi D. Macropinocytosis of Nab-Paclitaxel Drives Macrophage Activation in Pancreatic Cancer. *Cancer Immunol Res* (2017) 5:182–90. doi: 10.1158/2326-6066.CIR-16-0125
- Pal Singh S, Dammeijer F, Hendriks RW. Role of Bruton's Tyrosine Kinase in B Cells and Malignancies. *Mol Cancer* (2018) 17:57. doi: 10.1186/s12943-018-0779-z
- 120. Good L, Benner B, Carson WE. Bruton's Tyrosine Kinase: An Emerging Targeted Therapy in Myeloid Cells Within the Tumor Microenvironment. *Cancer Immunol Immunother: CII* (2021) 1–13. doi: 10.1007/s00262-021-02908-5
- 121. Messex JK, Liou GY. Targeting BTK Signaling in the Microenvironment of Solid Tumors as a Feasible Cancer Therapy Option. *Cancers (Basel)* (2021) 13:2198. doi: 10.3390/cancers13092198
- 122. Massó-Vallés D, Jauset T, Serrano E, Sodir NM, Pedersen K, Affara NI, et al. Ibrutinib Exerts Potent Antifibrotic and Antitumor Activities in Mouse Models of Pancreatic Adenocarcinoma. *Cancer Res* (2015) 75:1675–81. doi: 10.1158/0008-5472.CAN-14-2852
- 123. Molina-Cerrillo J, Alonso-Gordoa T, Gajate P, Grande E. Bruton's Tyrosine Kinase (BTK) as a Promising Target in Solid Tumors. *Cancer Treat Rev* (2017) 58:41–50. doi: 10.1016/j.ctrv.2017.06.001
- 124. Tempero MA, Coussens LM, Fong L, Manges R, Singh P, Li Y, et al. A Randomized, Double-Blind, Placebo-Controlled Study of Ibrutinib, a Bruton Tyrosine Kinase Inhibitor, With Nab-Paclitaxel and Gemcitabine in the First-Line Treatment of Patients With Metastatic Pancreatic Adenocarcinoma (RESOLVE). J Clin Oncol (2016) 34:TPS2601–TPS. doi: 10.1200/ JCO.2016.34.15_suppl.TPS2601
- 125. Gunderson AJ, Kaneda MM, Tsujikawa T, Nguyen AV, Affara NI, Ruffell B, et al. Bruton Tyrosine Kinase-Dependent Immune Cell Cross-Talk Drives Pancreas Cancer. *Cancer Discov* (2016) 6:270–85. doi: 10.1158/2159-8290.CD-15-0827
- 126. da Cunha-Bang C, Niemann CU. Targeting Bruton's Tyrosine Kinase Across B-Cell Malignancies. Drugs (2018) 78:1653–63. doi: 10.1007/s40265-018-1003-6
- 127. Hong D, Rasco D, Veeder M, Luke JJ, Chandler J, Balmanoukian A, et al. A Phase 1b/2 Study of the Bruton Tyrosine Kinase Inhibitor Ibrutinib and the PD-L1 Inhibitor Durvalumab in Patients With Pretreated Solid Tumors. Oncology (2019) 97:102–11. doi: 10.1159/000500571
- 128. Overman M, Javle M, Davis RE, Vats P, Kumar-Sinha C, Xiao L, et al. Randomized Phase II Study of the Bruton Tyrosine Kinase Inhibitor Acalabrutinib, Alone or With Pembrolizumab in Patients With Advanced Pancreatic Cancer. *J Immunother Cancer* (2020) 8:e000587. doi: 10.1136/jitc-2020-000587
- 129. Zhang Z, Ji S, Zhang B, Liu J, Qin Y, Xu J, et al. Role of Angiogenesis in Pancreatic Cancer Biology and Therapy. *BioMed Pharmacother* (2018) 108:1135–40. doi: 10.1016/j.biopha.2018.09.136

- 130. van Mackelenbergh MG, Stroes CI, Spijker R, van Eijck CHJ, Wilmink JW, Bijlsma MF, et al. Clinical Trials Targeting the Stroma in Pancreatic Cancer: A Systematic Review and Meta-Analysis. *Cancers (Basel)* (2019) 11:588. doi: 10.3390/cancers11050588
- 131. Kindler HL, Friberg G, Singh DA, Locker G, Nattam S, Kozloff M, et al. Phase II Trial of Bevacizumab Plus Gemcitabine in Patients With Advanced Pancreatic Cancer. J Clin Oncol (2005) 23:8033–40. doi: 10.1200/ JCO.2005.01.9661
- 132. Lu Z, Weniger M, Jiang K, Boeck S, Zhang K, Bazhin A, et al. Therapies Targeting the Tumor Stroma and the VEGF/VEGFR Axis in Pancreatic Ductal Adenocarcinoma: A Systematic Review and Meta-Analysis. *Target Oncol* (2018) 13:447–59. doi: 10.1007/s11523-018-0578-x
- Chandana S, Babiker HM, Mahadevan D. Therapeutic Trends in Pancreatic Ductal Adenocarcinoma (PDAC). *Expert Opin Investig Drugs* (2019) 28:161– 77. doi: 10.1080/13543784.2019.1557145
- 134. Annese T, Tamma R, Ruggieri S, Ribatti D. Angiogenesis in Pancreatic Cancer: Pre-Clinical and Clinical Studies. *Cancers (Basel)* (2019) 11:381. doi: 10.3390/cancers11030381
- 135. Tao J, Yang G, Zhou W, Qiu J, Chen G, Luo W, et al. Targeting Hypoxic Tumor Microenvironment in Pancreatic Cancer. J Hematol Oncol (2021) 14:14. doi: 10.1186/s13045-020-01030-w
- 136. Hanahan D, Weinberg Robert A. Hallmarks of Cancer: The Next Generation. *Cell* (2011) 144:646–74. doi: 10.1016/j.cell.2011.02.013
- 137. Schizas D, Charalampakis N, Kole C, Economopoulou P, Koustas E, Gkotsis E, et al. Immunotherapy for Pancreatic Cancer: A 2020 Update. *Cancer Treat Rev* (2020) 86:102016. doi: 10.1016/j.ctrv.2020.102016
- 138. Laheru D, Lutz E, Burke J, Biedrzycki B, Solt S, Onners B, et al. Allogeneic Granulocyte Macrophage Colony-Stimulating Factor-Secreting Tumor Immunotherapy Alone or in Sequence With Cyclophosphamide for Metastatic Pancreatic Cancer: A Pilot Study of Safety, Feasibility, and Immune Activation. *Clin Cancer Res* (2008) 14:1455–63. doi: 10.1158/ 1078-0432.CCR-07-0371
- 139. Lutz E, Yeo CJ, Lillemoe KD, Biedrzycki B, Kobrin B, Herman J, et al. A Lethally Irradiated Allogeneic Granulocyte-Macrophage Colony Stimulating Factor-Secreting Tumor Vaccine for Pancreatic Adenocarcinoma. A Phase II Trial of Safety, Efficacy, and Immune Activation. *Ann Surg* (2011) 253:328– 35. doi: 10.1097/SLA.0b013e3181fd271c
- 140. Le DT, Picozzi VJ, Ko AH, Wainberg ZA, Kindler H, Wang-Gillam A, et al. Results From a Phase IIb, Randomized, Multicenter Study of GVAX Pancreas and CRS-207 Compared With Chemotherapy in Adults With Previously Treated Metastatic Pancreatic Adenocarcinoma (ECLIPSE Study). *Clin Cancer Res* (2019) 25:5493–502. doi: 10.1158/1078-0432.CCR-18-2992
- 141. Patnaik A, Kang SP, Rasco D, Papadopoulos KP, Elassaiss-Schaap J, Beeram M, et al. Phase I Study of Pembrolizumab (MK-3475; Anti-PD-1 Monoclonal Antibody) in Patients With Advanced Solid Tumors. *Clin Cancer Res* (2015) 21:4286–93. doi: 10.1158/1078-0432.CCR-14-2607
- 142. Brahmer JR, Tykodi SS, Chow LQM, Hwu W-J, Topalian SL, Hwu P, et al. Safety and Activity of Anti-PD-L1 Antibody in Patients With Advanced Cancer. N Engl J Med (2012) 366:2455–65. doi: 10.1056/NEJMoa1200694
- 143. Kamath SD, Kalyan A, Kircher S, Nimeiri H, Fought AJ, Benson IIIA, et al. Ipilimumab and Gemcitabine for Advanced Pancreatic Cancer: A Phase Ib Study. Oncologist n/a (2020) 25(5):e808–15. doi: 10.1634/theoncologist. 2019-0473

- 144. Aglietta M, Barone C, Sawyer MB, Moore MJ, Miller WHJr., Bagalà C, et al. A Phase I Dose Escalation Trial of Tremelimumab (CP-675,206) in Combination With Gemcitabine in Chemotherapy-Naive Patients With Metastatic Pancreatic Cancer. Ann Oncol (2014) 25:1750–5. doi: 10.1093/ annonc/mdu205
- 145. Renouf DJ, Dhani NC, Kavan P, Jonker DJ, Wei A-c, Hsu T, et al. The Canadian Cancer Trials Group PA.7 Trial: Results From the Safety Run in of a Randomized Phase II Study of Gemcitabine (GEM) and Nab-Paclitaxel (Nab-P) Versus GEM, Nab-P, Durvalumab (D), and Tremelimumab (T) as First-Line Therapy in Metastatic Pancreatic Ductal Adenocarcinoma (mPDAC). J Clin Oncol (2018) 36:349–. doi: 10.1200/JCO.2018.36. 4_suppl.349
- 146. Le DT, Lutz E, Uram JN, Sugar EA, Onners B, Solt S, et al. Evaluation of Ipilimumab in Combination With Allogeneic Pancreatic Tumor Cells Transfected With a GM-CSF Gene in Previously Treated Pancreatic Cancer. J Immunother (2013) 36:382–9. doi: 10.1097/CJI.0b013e31829fb7a2
- 147. Li K-Y, Yuan J-L, Trafton D, Wang J-X, Niu N, Yuan C-H, et al. Pancreatic Ductal Adenocarcinoma Immune Microenvironment and Immunotherapy Prospects. *Chronic Dis Transl Med* (2020) 6:6–17. doi: 10.1016/j. cdtm.2020.01.002
- 148. Martinez-Useros J, Martin-Galan M, Garcia-Foncillas J. The Match Between Molecular Subtypes, Histology and Microenvironment of Pancreatic Cancer and Its Relevance for Chemoresistance. *Cancers (Basel)* (2021) 13:322. doi: 10.3390/cancers13020322
- 149. O'Kane GM, Grünwald BT, Jang GH, Masoomian M, Picardo S, Grant RC, et al. GATA6 Expression Distinguishes Classical and Basal-Like Subtypes in Advanced Pancreatic Cancer. *Clin Cancer Res* (2020) 26:4901–10. doi: 10.1158/1078-0432.CCR-19-3724
- 150. Al-Assar O, Demiciorglu F, Lunardi S, Gaspar-Carvalho MM, McKenna WG, Muschel RM, et al. Contextual Regulation of Pancreatic Cancer Stem Cell Phenotype and Radioresistance by Pancreatic Stellate Cells. *Radiother Oncol: J Eur Soc Ther Radiol Oncol* (2014) 111:243–51. doi: 10.1016/j.radonc.2014.03.014
- 151. Tiriac H, Belleau P, Engle DD, Plenker D, Deschênes A, Somerville TDD, et al. Organoid Profiling Identifies Common Responders to Chemotherapy in Pancreatic Cancer. *Cancer Discov* (2018) 8:1112–29. doi: 10.1158/1538-7445.PANCA19-C57

Conflict of Interest: IC: Advisory Board: Eli-Lilly, Bristol Meyers Squibb, MSD, Bayer, Roche, Merck-Serono, Five Prime Therapeutics, Astra-Zeneca, Oncologie International, Pierre Fabre, Boehringer Ingelheim. Research funding: Eli-Lilly, Janssen-Cilag, Sanofi Oncology, Merck-Serono. Honorarium: Eli-Lilly.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Edwards, Kang and Chau. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.