



Potential Molecular Biomarkers of Vestibular Schwannoma Growth: Progress and Prospects

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Vestibular schwannomas (VSs, also known as acoustic neuromas) are relatively rare benign brain tumors stem from the Schwann cells of the eighth cranial nerve. Tumor growth is the paramount factor for neurosurgeons to decide whether to choose aggressive treatment approach or careful follow-up with regular magnetic resonance imaging (MRI), as surgery and radiation can introduce significant trauma and affect neurological function, while tumor enlargement during long-term follow-up will compress the adjacent nerves and tissues, causing progressive hearing loss, tinnitus and vertigo. Recently, with the deepening research of VS biology, some proteins that regulate merlin conformation changes, inflammatory cytokines, miRNAs, tissue proteins and cerebrospinal fluid (CSF) components have been proposed to be closely related to tumor volume increase. In this review, we discuss advances in the study of biomarkers that associated with VS growth, providing a reference for exploring the growth course of VS and determining the optimal treatment strategy for each patient.

Keywords: vestibular schwannoma, growth, biomarker, target therapy, merlin

1 INTRODUCTION

VSs are histologically benign lesions deriving from the Schwann cells of the vestibulo-cochlear nerve commonly occur unilaterally, or bilaterally in the pathognomonic for the hereditary disorder neurofibromatosis type 2 (NF2) (1, 2). Generally, the mere presence of benign tumors is not a therapeutic indication. However, the slow but progressive growth of this intracranial tumor can confer compression to adjacent cranial nerve and brainstem, resulting in neurological deficits, balance difficulty and sensorineural hearing loss (SNHL) (3).

The goal of VS treatment has shifted from saving lives towards functional preservation, with multifaceted decision including watch-wait-rescan protocol, surgical resection and radiation (4, 5). However, surgery can be traumatic (6), while radiotherapy has a low hearing retention rate and can affect neurological function even after many years (7). We can see from the literature that two-thirds of VS did not grow during 3.6 years of follow-up, the average growth rate of sporadic VS was 1.1 mm/year diameter, and for NF2-related tumors 1.7 mm/year (8, 9), supporting the wait and watch policy in appropriate patients. A study showed that conservative management in small- to medium-sized VS (less than 2cm) can improve rates of facial nerve preservation and hearing protection in comparison to patients who undergo primary surgical treatment (10). Therefore, for stable or

involuting tumors with no mass effect, or elderly patients who will suffer higher risk of comorbidities, the continuing trend toward observation with regular follow-up imaging is reasonable (5, 11).

Nevertheless, observation policy carries the inherent risk of tumor progression and hearing deterioration in growing tumors, which emphasizing the importance of the identification of effective and convenient indicators to predict growth characteristics at the time of diagnosis to weigh the benefits and risks of active treatment against watchful observation. Previous studies have reported onset at an early age, increasing size in the first year, hearing loss, extrameatal tumor location, cerebellopontine angle extension, sway velocity with eyes open were risk factors for tumor growth (9, 12–15). Here, we provide a narrative review on the topic of biomarkers currently considered to be related to VS growth in order to assist surgeons to select the most accurate treatment approach and the development of innovative targeted therapies.

2 METHODS

The authors conducted a literature review using Web of Science and PubMed databases in order to identify important recent publications and synthesized them into a comprehensive review of potential biomarkers of VS growth. The search strategy included the search terms “growth”, “expression” and “vestibular schwannoma”. We have also scanned the reference lists of published articles for further potential hits of relevance to this review. Peer-reviewed full articles published in English till August 2021 were included to compile this review. Search results were judged for relevance by the research team using the title,

abstract and if necessary full text. Studies were included, if they 1) explored the relationship between biomarkers and the size or growth of sporadic VS or NF2-associated VS; 2) explored molecules that aberrantly expressed in VS compared with normal tissues; 3) drug research for VS treatment. Studies were excluded, if they 1) merely found some imaging manifestations or clinical features related to VS growth; 2) case reports.

3 RESULTS

We summarized the search results into five major types of biomarkers, which are summarized in **Table 1**.

3.1 Merlin Pathway Related Proteins

Neurofibromin 2 gene mutation and the function loss of its transcription protein merlin (an acronym for moesin-ezrin-radixin-like protein) are widely regarded to play a paramount role in the pathogenesis of both sporadic and bilateral VS (46, 47). Merlin belongs to the ERM (for ezrin, radixin, moesin) family of cytoskeleton linker proteins and shares a common structural organization with it: a relatively conserved N-terminal FERM domain, followed by a α -helical region and a charged hydrophilic-COOH terminal tail (48). The *neurofibromin 2* gene encodes for two major isoforms of merlin: exon 16 skipping production isoform 1 and exon 16 retention isoform 2, both of which carry full tumor suppressive function (49).

Merlin maintains the stability of the cell membrane by the binding of integral membrane proteins and spectrin actin cytoskeleton and mediates cell contact inhibition (50). Merlin exerts its growth suppressive function by modulating the activity of intracellular promitogenic signal cascades related to tumor

TABLE 1 | Summary table of potential biomarkers related to VS growth.

Major types	Action mechanisms	Biomarkers and their characteristics	References
Merlin pathway proteins	RTK signal proteins	VEGF expression was increased in VS, and anti-VEGF therapy was effective for NF2-VS.	(16)
		Pharmaceutical inhibition of PDGFR could reduce VS growth rate.	(17, 18)
		bFGF promoted proliferation and invasion of VS cells, and also might be a hearing protector in VS.	(16, 19)
Inflammatory signal	Ras signal proteins	ErbB family proteins were aberrantly expressed in VS, and its inhibition has therapeutic effects on VS.	(20)
		Merlin inhibits Ras signal transduction by interfering with GRB2 expression and inhibiting Rac1 and PAK1 activation.	(21–23)
		CPI-17–MYPT1 switch was responsible for the conformational change of merlin. CPI-17 was over-expressed in VS.	(24–26)
Tumor miRNAs	Hippo signal proteins	Merlin activated LATS1/2 and inhibited the destabilization of LATS1/2 by CRL4 ^{DCAF1} . LATS1/2 promoted the degradation of YAP and inhibited the transcription of pro-proliferation and anti-apoptotic genes stimulated by YAP.	(27, 28)
		The NF- κ B signal activated in VS was considered to be the core of VS pathophysiology.	(29, 30)
		COX-2 was highly expressed in VS and was associated with high proliferation rate.	(31)
Tumor proteins	Local inflammation	Macrophages, especially M2-type macrophages, were thought to promote VS growth.	(32, 33)
		High NLR was associated with VS growth.	(34)
		The upregulation of miR-29abc, miR-19, miR-340-5p, miR-21, miR-221 and downregulation of miR-744, let-7b were related to VS growth velocity.	(35, 36)
CSF components	Systemic inflammation	CD105 can be used as a MVD marker, and was correlated with VS size and growth rate.	(37)
		BDNF expression was correlated with mitotic activity in VS.	(38)
		The expression of MMP-2 and MMP-9 was correlated with VS growth rate and was higher in cystic VS than solid VS. The proteolytic activity of MMP-14 was related to the degree of SNHL in VS patients.	(39–41)
Mucopolysaccharide	Proteases	The expression of ADAM9 was higher in VS and related to functional impairment.	(42, 43)
		HA levels were elevated in NF2-VS and correlated with the proliferation rate of schwannoma cells.	(44)
		ABCA3, SCG1, KLF11, CA2D1, BASP1 and PRDX2 were associated with VS growth.	(45)

formation, including Ras/Raf/MEK/ERK (51), PI3K/Akt (52), c-JNK (53), Hippo signaling pathway, and the overall activity of the E3 ubiquitin ligase CRL4^{DCAF1} in the nucleus (54). Moreover, merlin was reported to assume a possible role in the mediation of cell cycle progression (55).

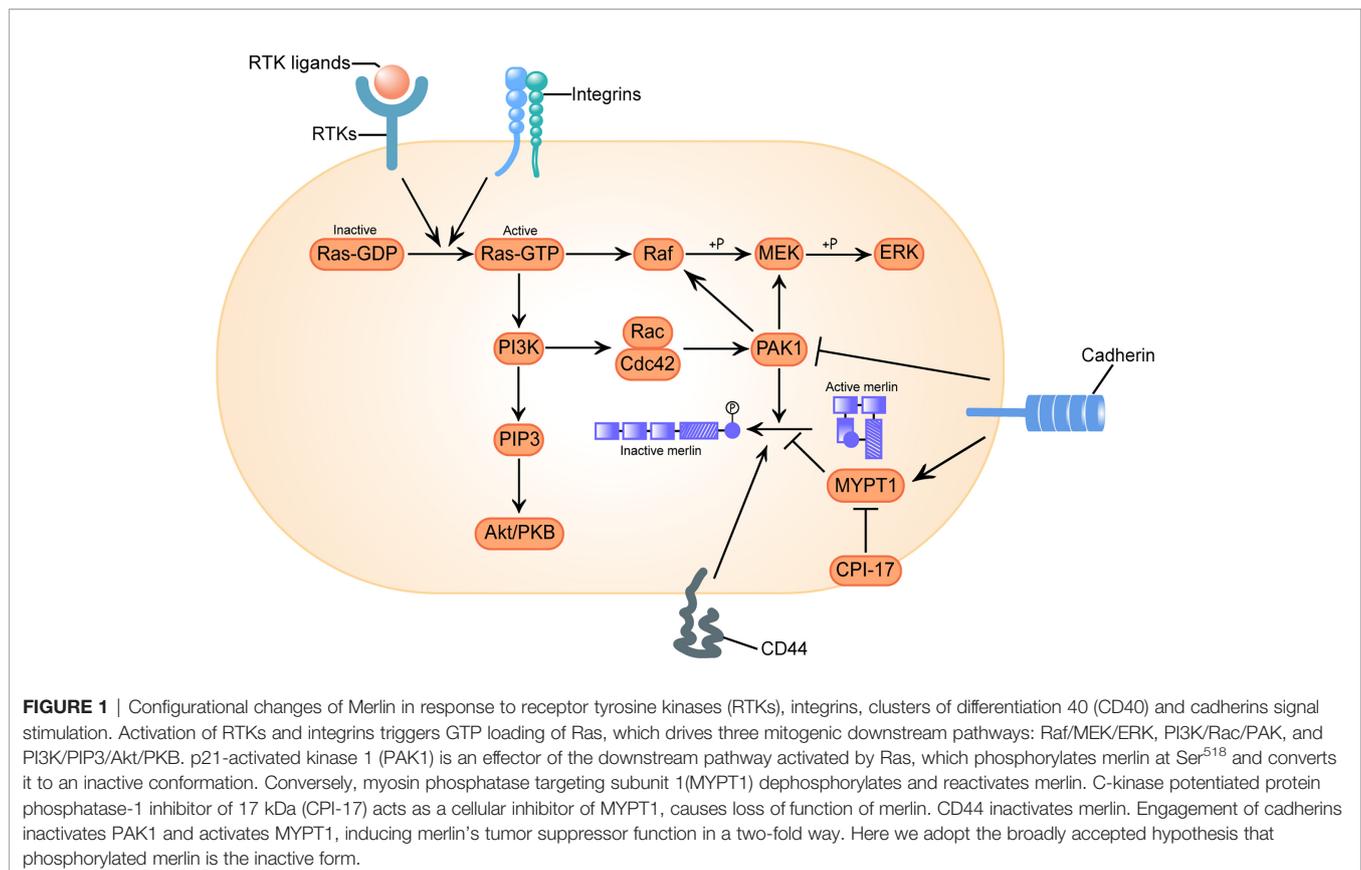
Classically, it was considered that the activity of merlin was regulated by phosphorylation on the main regulatory Ser⁵¹⁸: dephosphorylated merlin functions as a growth inhibitor in a closed conformation formed by intramolecular association of its N-terminal domain (NTD) and carboxy-terminal domain (CTD), while phosphorylated merlin cannot form a folded state and is functionally deficient (56). However, subsequent researches have shown that transcripts of the mutated *neurofibromin 2* gene or phosphorylated merlin have a more closed form, leading to impaired contact-dependent inhibition of proliferation (57, 58). There is also a controversial view holding that merlin's tumor suppressive function is independent of its conformational change (59). The activation of receptor tyrosine kinases (RTKs), integrins, CD44 and cadherin signal are responsible for the regulation of merlin's phosphorylation *via* the downstream effector p21-activated kinase 1 (PAK1) and myosin phosphatase targeting subunit 1 (MYPT1) (**Figure 1**) (60, 61).

3.1.1 RTKs

The over-expression and over-activation of at least four types of RTKs and their ligands involved in the progression or invasion of

sporadic and NF2-associated VS: ErbB (62), platelet-derived growth factor (PDGF) (63), basic fibroblast growth factor (bFGF) (19) and vascular endothelial growth factor (VEGF) (64).

VEGF is one of the most prominent angiogenesis stimulator that can regulate irregular blood vessel sprouting and growth except for simple remodeling of the capillary basement membrane (65), and lead to development of an immuno suppressive tumor microenvironment (66). Compared with normal vestibular nerve, the expression of VEGF, VEGFR-1/Flt and VEGFR-2/Flk as well as the coreceptor NP1 were considerable increased in VS tissues, suggesting at least a part of neoplastic growth was induced *via* the promotion of angiogenesis (39, 64, 67). Tissue microarray analysis of 182 sporadic VSs found significantly higher VEGF levels in the groups of recurrent and preoperatively irradiated tumors when compared to primary VS patients (67). However, there are contradictory views as to whether VEGF expression is correlated with sporadic VS growth characteristics. Koutsimpelas et al. initially found a positive correlation between VEGF expression and tumor volume, tumor growth index (calculated by dividing the maximal tumor diameter by the patient's age) and microvessel density (MVD, defined by CD31 staining) (16), but further investigation with expanded sample size demonstrated that although tumors with high proliferation index and high levels of VEGF and its receptor were more common than those with low proliferation activity expressing low levels of VEGF and its receptor, the expression of neither VEGF nor its receptors correlated with the proliferation indexes (Ki-67) or the growth characteristics of the tumors (67).



Multiple anti-VEGF therapy studies in patients with progressive NF2-VS have shown that bevacizumab can not only ameliorate hearing loss and reduce tumor size (68), but also normalize the tumor vasculature and reduce vasogenic edema, which improved the delivery of oxygenation, a potent radiosensitizer, and therefore reduced radiation dose and minimized radiation-related neurotoxicity (69).

As a regulator of cell growth and division, PDGF can regulate stemness in schwannoma cell lines and have a role in tumorigenesis (70). The PDGF family consists of five different disulphide-linked dimers built up of four different polypeptide chains encoded by four different genes. These isoforms, PDGF-AA, PDGF-AB, PDGF-BB, PDGF-CC and PDGF-DD, act *via* two RTKs, PDGF receptors α and β (71). Nilotinib (17) and Gleevec (18, 63, 72) pharmacologically inhibit these two receptors and their main downstream signaling pathways that have been proved to be over-expressed and activated in VS, and could also potentially reduce the angiogenic activity and growth rate of VS.

bFGF (also known as FGF-2 or FGF- β) was not only an identified angiogenic cytokine (73), but also implicated in tumor maintenance and metastasis (74). bFGF was identified as a mediator to protect auditory neurons from acoustic trauma and aminoglycoside ototoxicity, it was 3.5-fold higher in good hearing VS *versus* poor hearing VS (75), and its plasma concentration increased while patient's hearing improved after bevacizumab regimen (76). bFGF is also a known mitogen that promotes the proliferation of cell cultures derived from sporadic VS and increased the invasive phenotype mediated by Akt and ERK in HEI-193 cells (19, 77). In sporadic VS, increased levels of bFGF were positively correlated with tumor volume, tumor growth index and MVD (16). The mechanism by which bFGF stimulates mitosis might divergent from the way it modulates hearing. This notion, together with the results of cytokine array analysis where levels of ErbB, a well-established growth modulator, are not correlate with hearing status, may partly explain the irrelevance of tumor growth rate or tumor volume with VS patients' hearing outcome (75).

The ErbB family consists of four members: ErbB-1/EGFR/Her1, ErbB-2/Neu/Her2/p185, ErbB-3/Her3, and ErbB-4/Her4. ErbB family is upregulated in many malignant tumors, and also aberrantly expressed in VS. It has been reported that 68% of EGFR (62% sporadic and 75% NF2-associated VS), 84% of ErbB2 (76% sporadic and 94% NF2-related VS) and 34% of ErbB3 were upregulated in VS (20). Of EGFR ligands, EGF was up-regulated in all NF2-related VS, but none of the sporadic VS; Neuregulin was up-regulated in 86% of sporadic VS and 19% of NF2-related VS (20). In HEI-193 cells, the addition of EGF increased cellular invasion by 10-fold, which could be reduced by the inhibition of PI3K/Akt (19). Both dual small molecule inhibitor of EGFR and ErbB2 Lapatinib and the EGFR inhibitor Erlotinib demonstrated their therapeutic effects in VS (78–80).

3.1.2 Ras and Related Raf/MEK/ERK, PI3K/Akt and Rac/PAK

The small G-protein Ras, the downstream oncoprotein of RTKs that cycles between an inactive GDP-bound and an active GTP-

bound state, play a role in the cascade of cell proliferation and division. In its active state, Ras can interact with several different effectors, thereby triggering an array of downstream signaling networks that responsible for promoting cellular transformation and driving tumorigenesis, such as Ras/Raf/MEK/ERK and Ras/PI3K/Akt pathways (81–83). It is well known that activation of Ras leads to subsequent activation of Rac/Cdc42 and its downstream effector PAK1, whose phosphorylation of Raf on Ser³³⁸ and MEK on Ser²⁹⁸ is required for effective signal transfer of Ras (**Figure 1**) (84, 85).

An increasing body of literature suggests that merlin counteract Ras-induced transformation at multiple levels. Merlin interferes with the expression of endogenous growth factor receptor binding 2 (GRB2) protein (21), directly reduces the GTP-loading of Ras and Rac to restrain their activation (86), and directly binds to the p21 binding domain (PBD) of PAK1 to interrupt PAK1 activation and its recruitment to focal adhesions (22, 87). Merlin competitively binds to angiominin and releases the Rac1 negative regulator Rich1, which ultimately attenuates Rac1 signaling (23). Dysfunction of merlin would allow for enhanced Ras signal transduction and accelerated tumor growth rate (24).

Merlin itself is regulated by PAK reciprocally, it was phosphorylated at Ser⁵¹⁸ by active PAK and therefore lose the inhibition of cell transformation. Activated Rac expression induces merlin phosphorylation and decreases the association between merlin and cytoskeleton (60). Based on the study of the role of Ras pathway signal transduction in the growth of VS, preclinical assessment of PAK inhibitors (88), MEK1/2 inhibitors (81) and Ailanthone, the down regulator of Ras and Raf, all exhibited certain antitumor properties (89).

3.1.3 MYPT1

MYPT1 is a phosphatase that forms the C-kinase potentiated protein phosphatase-1 inhibitor of 17 kDa (CPI-17)–MYPT1 switch along with its most specific and potent inhibitor CPI-17, regulating the phosphorylation of both merlin and other ERM family proteins (24, 90). Although highly homologous, the activity changes of merlin and other ERM proteins after C-terminal phosphorylation are opposite, and fulfils an antagonistic role in Ras activity control: the phosphorylation inactivates merlin, which counteracts Ras-induced transformation (91), but activates ERM proteins, which are essential for proper Ras activation (92). Thus, the oncogenic protein CPI-17 may activate Ras signaling in a two-fold way.

A previous systematically investigation on schwannomas showed that CPI-17 stained negative in non-tumor pathologies, but specifically up-regulated in over 90% of schwannomas, primarily in sporadic schwannomas (25). Moreover, high CPI-17 levels were found to correlate with higher Ki-67 proliferation indices, indicating a putative role of CPI-17 in schwannoma progression (25). Xu et al. found the over-expression of CPI-17 was a prominent feature of sporadic VS tissues, and there was a significantly positive correlation between CPI-17 expression and merlin phosphorylation (26), confirming the rationality of the causal chain CPI-17–MYPT1–merlin dysfunction in VS.

3.1.4 Yes-Associated Protein (YAP)

The deregulation of the Hippo pathway and the accompanying activation of Yes-associated protein (YAP) has been implicated in VS cell proliferation (27). As described in **Figure 2**, merlin initiates Hippo pathway to promote the phosphorylation and degradation of YAP and its homologous protein transcriptional coactivator with PDZ-binding motif (TAZ) (93, 94). In the inactivated state of merlin, YAP/TAZ migrates into the nucleus and binds to TEA domain family members (TEAD), stimulating the transcription of pro-proliferation and anti-apoptotic genes (27, 95).

The E3 ubiquitin ligase CRL4^{DCAF1} can directly ubiquitinate and destabilize LATS1/2 to activate YAP (54). Merlin can translocate into the nucleus, binds with high affinity to CRL4^{DCAF1} and blocks its function (28). It is postulated that this merlin-CRL4^{DCAF1}-LATS1/2-YAP signaling axis might be the key mechanism of merlin's tumor suppressive effect. Although no studies have directly confirmed the correlation between VS tumorigenesis and CRL4^{DCAF1} activity, targeted inhibition of the upstream activator of CRL4^{DCAF1}, NEDD8-activating enzyme (NAE), can induce inhibitory YAP phosphorylation and reduce cell proliferation in mouse *neurofibromin 2*-mutant schwannoma cells and human *neurofibromin 2*-mutant mesothelioma cell line (96), indicating a growth-promoting role of CRL4^{DCAF1} in *neurofibromin 2*-inactivated tumors.

3.2 Inflammatory Signal

At present, there is increasing evidence that a variety of solid tumors contain a certain degree of tumor-associated inflammation that play a role in the cancer formation, progression and metastasis (97). Inflammatory response in cancer patients comprises both the local inflammatory response and the systemic response. The local inflammatory response is mediated by chemokines, cytokines, growth factors and matrix metalloproteinases (MMPs) secreted from *in situ* tumor cells, stromal cells, and infiltrating immune cells, and can become accomplices to cancer development by enhancing cell survival, invasion, neovascularization, and adaptive immunity suppression (98). Meanwhile, inflammation in the tumor microenvironment could be reflected in peripheral circulation, i.e., systemic inflammatory response that characterized by white cell components in peripheral blood, such as the platelet-to-lymphocyte ratio (PLR) and neutrophil-to-lymphocyte ratio (NLR), which have been highlighted as probable markers of pathologic responses for numerous types of solid tumors (99, 100).

It has been demonstrated that the volume increase of VS is not merely based on cell proliferation, but is also implicated with (neo)vascularization, intratumoral hemorrhage, cyst formation and inflammatory reaction (32, 101, 102).

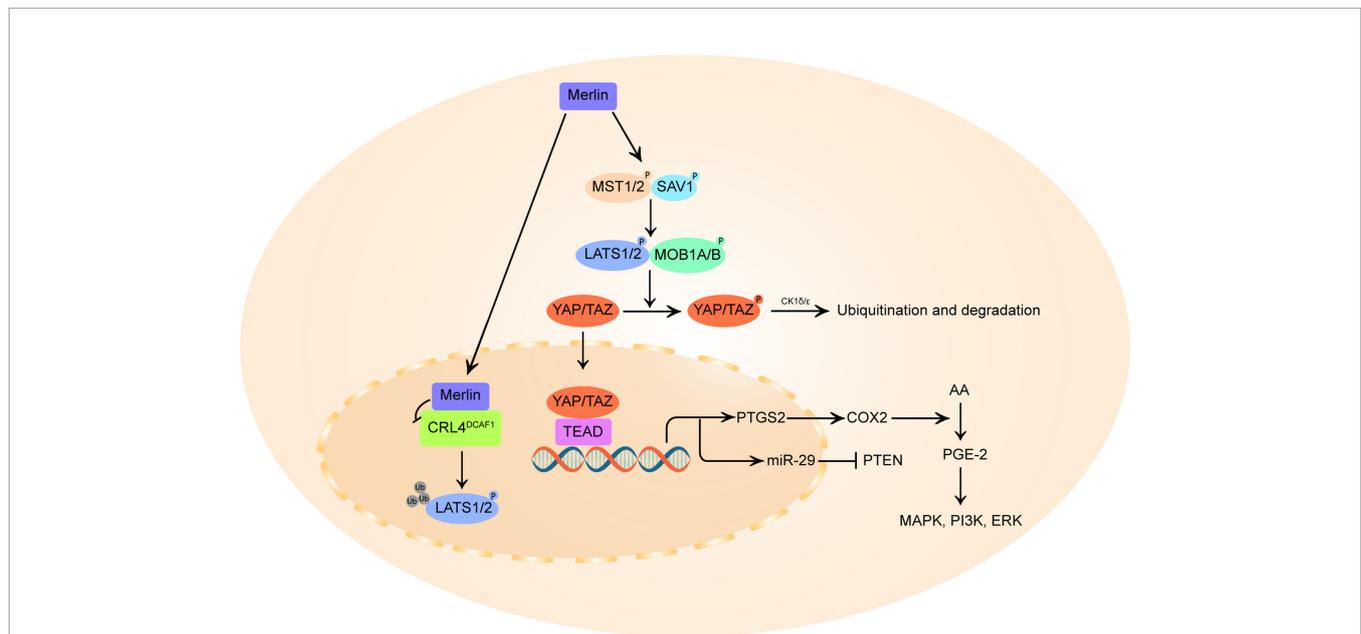


FIGURE 2 | Schematic diagram of merlin and Hippo pathway signaling. On the one hand, merlin initiates the Hippo pathway by directly activating mammalian STE20-like protein (MST1/2), which in turn phosphorylates large tumor suppressor homolog 1/2 (LATS1/2), or by recruiting LATS 1/2 to the plasma membrane for phosphorylation by MST1/2 kinases. The activated LATS1/2 directly phosphorylates Yes-associated protein (YAP) at Ser³⁹⁷ and transcriptional coactivator with PDZ-binding motif (TAZ) at Ser³¹¹, priming the neighboring phosphodegron motif for phosphorylation by CK1δ/ε kinases and multi-level control of YAP/TAZ levels. In parallel, merlin also translocates to the nucleus and blocks the activity of the nuclear E3 ubiquitin ligase CRL4^{DCAF1}, which promotes the ubiquitylation of LATS1/2 to activate YAP. In merlin-deficient cells, YAP/TAZ accumulates in the nucleus and forms hybrid transcription factors with TEA domain (TEAD) family proteins to promote the transcription of pro-proliferative genes, including PTGS2 and miR-29. PTGS2 encodes cyclooxygenase-2 (COX-2) that catalyzes the conversion of arachidonic acid (AA) to prostaglandin E-2 (PGE-2), thus promoting PGE-2 mediated MAPK, ERK, PI3K pathway. By inducing miR-29, YAP can also inhibit the translation of phosphatase and tensin homolog (PTEN), a broadly downregulated tumor suppressor in vestibular schwannoma.

3.2.1 Nuclear Factor-Kappa B (NF- κ B)

NF- κ B is a ubiquitous, evolutionary conserved transcription factor central to cell growth, apoptosis, inflammation and various malignant diseases (103). As a group of homo- and hetero-dimeric proteins composed of members of the Rel family, NF- κ B complexes includes p65/RelA, RelB, c-Rel, p50/p105 (NF- κ B1), and p52/p100 (NF- κ B2) five subunits (104), and resides in the cytoplasm of non-stimulating cells in the form of complexes with inhibitor of κ B (I κ B). The NF- κ B signaling pathway is activated by inflammatory cytokines or growth factors, and induces gene transcription of these factors in a feedback-loop way (Figure 3) (105–107).

Studies have shown that merlin negatively regulates NF- κ B signaling, and the elevated NF- κ B signal was considered as the hub of the interaction network of aberrant expressed molecules in VS pathobiology (29, 30, 108). In the absence of merlin, overexpressed NIK and IKK α induced high NF- κ B dependent transcription (29), while merlin expression blocks NF- κ B activation by the inhibition of p65, NIK, IKK α , tumor necrosis factor- α (TNF- α)-induced I κ B degradation, NF- κ B–DNA binding and endogenous NF- κ B signaling (30). Besides, activated NF- κ B is thought to be the mechanism by which elevated p75^{NTR} expression promotes cell survival in VS (109). Hyper NF- κ B activity in VS potentiated hepatocyte growth factor (HGF) to c-Met autocrine feed-forward loop to promote

tumor cell proliferation (108), and increased the expression of factors that reported to be increased in VS or correlated with the prognosis or absolute tumor growth rate of VS, such as matrix metalloproteinase 2 (MMP-2), MMP-9 (40), MMP-14 (41), COX-2 (110), interleukin-1 (IL-1), IL-6, TNF- α (64) and signal transducers and activators of transcription 1 (STAT1) (111).

3.2.2 Cyclooxygenase-2 (COX-2)

COX-2 is an isozyme of the COX family, catalyzes the conversion of arachidonic acid to prostaglandins, including the biosynthesis of prostaglandin E-2 (PGE-2). In a healthy state, COX-2 is involved in maintaining cell homeostasis, while its response to homeostatic dysregulation might lead to the development of cancer (112). Studies have demonstrated that COX-2 is upregulated in various solid tumors and involved in cancer inflammation (113). COX-2 is released to the cancer microenvironment by type II macrophages, cancer-associated fibroblasts and tumor cells, and can suppress tumor cell apoptosis, enhance cell adhesion to promote tumor-induced angiogenesis and achieve an aggressive phenotype (114, 115).

COX-2 presented at high levels in majority (96.67%) of VS tissue samples, and those VSs with higher COX-2 expression showed higher proliferation rate (31). Transcription of COX-2 is thought to be promoted by NF- κ B. Aspirin modifies both NF- κ B signaling and COX-2 expression (116), and has been shown to

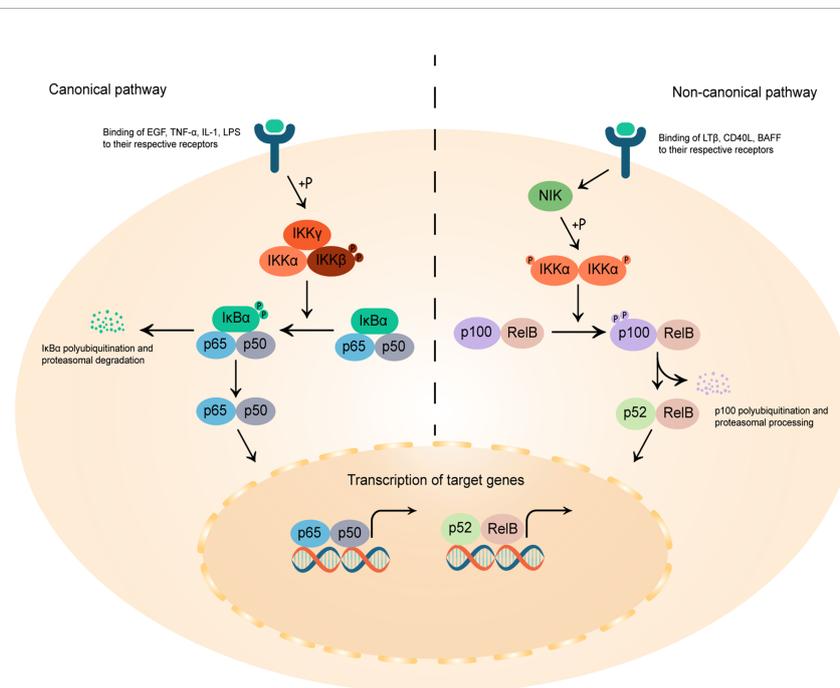


FIGURE 3 | Schematic diagram of the canonical and non-canonical NF- κ B signaling pathway. In the canonical pathway, on the left, the binding of epidermal growth factor (EGF), tumor necrosis factor- α (TNF- α), interleukin-1 (IL-1) and lipopolysaccharides (LPS) to their respective receptors lead to the phosphorylation of inhibitor of κ B (I κ B)-kinase β (IKK β) in the IKK complex, which in turn phosphorylates I κ B α , culminating in its polyubiquitination and proteasomal degradation. The free NF- κ B homo- or heterodimers, in this case p65/p50, then translocate into the nucleus to bind to κ B motif and promote target gene transcription. The non-canonical pathway, on the right, is activated by the binding of lymphotoxin β (LT β), CD40 ligand (CD40L), and B-cell activating factor (BAFF) to their respective receptors, causing IKK α dimers phosphorylation by the NF- κ B-inducing kinase (NIK). Phosphorylation of p100 by activated IKK α initiates proteasomal processing of p100 N-terminal to form p52. The p52/RelB heterodimers can then undergo nuclear translocation and modulates the gene transcription of pro-inflammatory cytokines in a feedback-loop way.

have therapeutic effects in several tumors with high COX-2 levels (117, 118). Although some studies have shown that aspirin may halt VS growth (119, 120), the latest meta-analysis disproved this viewpoint (121).

3.2.3 Macrophages

Tumor associated macrophages (TAMs) can be broadly classified into two categories, both of which are differentiated from monocytes in response to stimulus signals (122). The first type is M1-type inflammatory macrophages, also known as classically activated macrophages. M1-type macrophages are critically important in host defense and killing of tumor cells by the production of pro-inflammatory cytokines such as interferon- γ (IFN- γ), TNF- α and IL-18, which have potent microbiome-killing properties and hence are considered as 'good' macrophages (123). In order to avoid collateral damage to healthy cells/tissues by M1-type macrophages, M2-type macrophages, or alternatively activated macrophages, usually presented at the later stage of inflammatory response to suppressing destructive immunity, promoting wound repair and fibrosis. The effects of M2-type macrophages are manifested in the tumor microenvironment as inhibiting anti-tumor immunity, promoting tumor matrix remodeling and inducing angiogenesis (123). Collective data have shown that the high densities of M2-type macrophages in the tumor microenvironment are associated with the secretion of VEGF and MMP-9 (124), and worse prognosis in numerous cancer types (125, 126).

In a previous study, it was demonstrated that macrophages, rather than tumor cells, accounted for the majority of proliferating cells in the growing sporadic VS (32). Using Iba-1 as a pan-macrophage marker, the study of 24 sporadic VSs and 20 NF2-related VSs showed that the microvessel surface area was positively correlated with TAM density, and that Iba-1+ macrophages contributed significantly to VEGF production (127). This supports the idea that targeting macrophages along with vascular supply should be viewed as promising therapeutic options in both VS groups.

de Vries et al. determined the role of M2-type macrophages in promoting angiogenesis and volumetric tumor growth in sporadic VS tissue sections. They compared the expression of CD163, a specific marker for M2-type macrophages, in 10 rapidly growing VSs and 10 slow-growing VSs, and found that CD163 expression was significantly higher in fast-growing VS, and tumors with higher CD163 expression had more microvessels (as assessed by CD31). Consistent with these findings, the levels of macrophage colony-stimulating factor (M-CSF), an important cytokine that stimulates macrophage polarization into an M2-type macrophage, and its synergistic cytokine interleukin-34 (IL-34), are increased in fast-growing VS, suggesting their potential function as promoters of tumor progression (33). Recently, Bi et al. demonstrated variable expression of immune regulatory markers as well as immune infiltrates in VS (128). They found both tumor volume and volumetric growth were positively correlated with CD163 expression, and the relationship between PD-L1 and growth

strengthened with increasing CD163 infiltration, suggesting a prominent role of immunotherapy in VS treatment (128).

3.2.4 Neutrophil-to-Lymphocyte Ratio (NLR)

As a representative index of systemic inflammation, preoperative peripheral blood NLR is related to the pathological characteristics of many tumors (129). The pretreatment NLR is calculated according to the absolute neutrophil count (ANC) and the absolute lymphocyte count (ALC) of routine blood tests obtained prior to any intervention. A high NLR indicated greater systemic inflammation and elevated circulating concentrations of pro-inflammatory and angiogenic cytokines, which can enhance tumor progression, immunosuppression and peritumoral stroma formation, and therefore is considered as a promising negative prognostic biomarker in solid tumors (130).

The utility of NLR has been pointed for predicting oncological growth in patients with VS. Kontorinis et al. found a significant difference of NLR between 79 growing VSs and 82 non-growing VSs, with high NLR was observed predominantly in the growing VS group (34). However, there is no such discrepancy between regressing VS and growing VS (131). A probable speculation is that NLR-associated inflammation functions not only in the pathomechanism of tumor growth, but also in the decrease of tumor size. Currently, the critical limitation needs to be obviated for NLR's clinical application and further investigation is that there is no consensus on the cutoff value of NLR (34).

3.3 miRNAs

miRNAs are small non-coding RNAs that responsible for the post-transcriptional regulation of genes by partially or perfectly complementary to the target mRNAs, leading to protein synthesis abortion or transcription inhibition (132). miRNAs play an important role in tumor initiation and development, and are potential targets for therapeutic interventions in diverse cancers.

Several differentially expressed miRNAs have been identified in VS tissues (133, 134). Using principal component analysis, affected gene ontology analysis, and analysis of miRNA expression fold changes, Sass et al. determined the relationship between rapid tumor growth and upregulation of miR-29abc, miR-19, miR-340-5p, miR-21, miR-221 and downregulation of miR-744 and let-7b (35). miR-29 fulfills controversial function in tumor progression and invasion in different tumors (135–137), and was reported to be upregulated in previous VS studies (134). Concordant with this research, Cioffi et al. demonstrated that miR-21 was over-expressed in VS compared with normal vestibular nerve tissue (36). Yang et al. found that downregulation of miR-21 by aianthone reduced the proliferative potential of VS cells and induced apoptosis and autophagy (89). Other deregulated miRNAs also serve as modulators of tumor-related signaling pathways, such as miR-340 in Ras/Raf/MAPK (138) and let-7b in SOCS1/STAT (139). Notably, the contributions of miR-19, miR-21 and miR-221 to tumor growth are all related to their down-regulation of tumor suppressor PTEN (36, 136, 140), suggesting the role of PTEN and

its downstream PI3K/Akt/mTOR signaling pathway in VS cancerigenic process.

3.4 Proteins in Tumor Tissue

CD105, better known as endoglin, is a homodimeric transmembrane glycoprotein that highly expressed on activated angiogenic endothelial cells (141). Used as a marker of MVD, CD105 has proven itself as a potent prognostic indicator and a potential target for antiangiogenic therapy and chimeric antigen receptor-based T-cell (CAR-T) immunotherapy in several tumors (142–144). Calculated by immunohistochemically assessed CD105 expression, Gino et al. correlated the vessel cross-sectional area (VA) and vessel density (VD) with tumor dimensions and tumor growth rate in both NF2-associated VS and sporadic VS, and found a positive correlation between VD and tumor growth rate in NF2-VS, and VA and VD with tumor size in sporadic VS (37). Considering the great achievements of anti-angiogenesis therapy in VS and the specificity of CD105 to indicate the vascular front where sprouting takes place, it seems reasonable to use circulating CD105 to monitor tumor growth and to treat VS with anti-endoglin antibodies.

With several over-expressed neurotrophic factors in VS drawn attention as putative key mediators of tumor growth, Kramer et al. explored the relationship between gene expression profiles of neurotrophic factors and proliferation-associated Ki-67 labelling index, and observed significantly elevated brain derived neurotrophic factor (BDNF) expression that correlated with mitotic activity in VS (38). BDNF plays a pivotal role in myelination processing and supports the survival and synaptic integrity of the auditory nerve (145). This mitosis-promoting neurotrophic factor, together with the previously mentioned bFGF, which has the dual effects of hearing protection and growth stimulation of VS cells (16, 75), may partially explain the seemingly paradoxical phenomenon that VS patients with tumor growth do not necessarily experience hearing loss.

MMPs comprise a large family of zinc- and calcium-dependent proteolytic enzymes. Apart from their essential role in promoting cell differentiation and the reconstruction of the extracellular matrix in a regenerative milieu, MMPs play a fundamental role in tumor progression, invasion, and angiogenesis (146). Expression characteristics of several MMPs family members in VS have been explored. MMP-2 and its endogenous inhibitors tissue inhibitors of metalloproteinase (TIMP)-1 were found interstitially in sporadic solid VS, while the majority of MMP-9 was localized in tumor cells, and its concentration in tumor sample homogenates was positively correlated with the absolute tumor growth rate (40). Consistent with previous studies showing that the proteolytic activity of MMPs might degrade collagen, fibronectin, laminin and contribute to cyst formation and expansion, the expression of MMP-2 and MMP-9 were upregulated in cystic VS samples compared with solid VS (147). In cystic VS tissues, MMP-2 was localized to tumor cells on the cyst cavity inner surface and its levels were higher in the cystic fluid than in other samples (148).

Histologically, VS can be classified into two tissue types, where Antoni type A has a compact structure with interwoven bundles of long bipolar spindle cells and Antoni type B characterized by a

loose texture and small, uniform satellite cells. It is known that cystic VS consist of a large mass of Antoni type B area, where the expression of MMP-14 was significantly higher than that in Antoni type A (39). The abundance and proteolytic activity of MMP-14 in VS patients was correlated with the degree of SNHL and surgical outcomes (41). Besides, MMP-2, MMP-9 and MMP-14 are all principal proteins of the MMPs family in the vasculature, implying degeneration of tumor tissues by MMPs and the accompanying increased tumor vessel permeability and adhesion to the facial nerve are critical in VS tumorigenesis, cyst formation and preoperative hearing impairment.

As a membrane-anchored protein of the A-Disintegrin and Metalloproteinase (ADAM) protein family, the overexpression of ADAM9 in solid tumors has been correlated with aggressive tumor phenotypes and unfavorable clinical prognosis (149). ADAM9 may implicated in tumor progression and invasion either *via* non-proteolytic mechanisms that include interactions between tumor cells and endothelial or peritumoral stromal cells (150), or proteolytic mechanisms that involve an enzymatic modification called “shedding” or processing of cell-surface proteins (151). The value of ADAM9 inhibition in reducing migration and invasion of different solid tumors has been established (152, 153). The earliest expression assessment of ADAM9 in VS by Breun et al. found an 8.8-fold higher ADAM9 mRNA level in VS compared with in healthy peripheral nerves, and a strong correlation between ADAM9 mRNA expression and the degree of functional impairment (42). Further, in VS primary cell cultures, ADAM9 knock down caused a 58% reduction in cell numbers, which might be a hint that ADAM9 plays a role in inducing VS progression (43).

3.5 Components in Cerebrospinal Fluid

VSs usually originate from Schwann cells of the cisternal portion of nerve rootlets exposed to CSF and may alter CSF composition by secreting proteins or altering CSF metabolism (154). CSF-based liquid biopsy approaches have been used to characterize protein expression during the pathogenesis of several brain tumors (155, 156), and differential components analyses of CSF are also expected to yielded diagnostic and therapeutic targets in VS.

Two studies have been done on CSF composition to identify biomarkers that predict VS growth. Ariannur et al. found the levels of hyaluronan (HA) were 17-fold higher in NF2-related VS cases compared to the controls, and the rate of HA synthesis and secretion by primary schwannoma cells was commensurate with their proliferation rate (44). HA is a cell-surrounding mucopolysaccharide that binds to the Schwann cell surface CD44 receptor to trigger an uninterrupted cell proliferation cascade. Deranged HA-CD44 interaction has been identified as one of the central causative factors for schwannoma and a tumor suppressor target of merlin (157). Schwannoma cells increase their self-reproductive potential by secreting HA under the innate condition, suggesting that elevated HA in the CSF of patients with NF2-VS may be used as an indicator of tumor growth.

By characterizing the CSF proteome among VS patients with different grades, Huang et al. found that ATP-binding cassette subfamily A member 3 (ABCA3), secretogranin-1 (SCG1),

Kruppel-like factor 11 (KLF11), voltage-dependent calcium channel subunit alpha-2/delta-1 (CA2D1), brain acid soluble protein 1 (BASP1), and peroxiredoxin-2 (PRDX2) in CSF were associated with VS growth (45). It is notable that some of them did not simply increase or decrease as the tumor grows, but reached their maximum or minimum values at certain phases, suggesting that volumetric growth of VS may be driven by different signaling pathways at different tumor stages.

4 DISCUSSION

VS is a benign but potentially devastating tumor whose aggressively growth can be associated with significant morbidity including deafness and facial neuropathy. Usually, treatment of VS relies on surgery, radiation and regular follow-up. Better knowledge of the pathogenesis of VS has led to several targeted therapies with the effect of reducing tumor volume or restoring patients' hearing, of which bevacizumab has achieved the most promising results (68).

The majority of VVs may not enlarge after initial diagnosis, with an average annual growth rate of 1.11 mm (9). In order to preserve hearing function and enhance the quality of life, a substantial proportion of newly diagnosed patients will choose wait-and-scan policy, who were put at the risk of sudden tumor growth due to extremely variable growth patterns of VS. Hence, it would be helpful to identify predictive factors for VS growth at time of diagnosis. Several epidemiological, clinical, and radiological characteristics were thought to be related to the increased risk of subsequent tumor growth, such as patients' age, tumor location, hearing loss, and grow at first follow-up (9, 12–14). But there were some inconsistencies among reports of predicting growth-related factors (9), making the accuracy of this approach debatable.

From the perspective of VS growth mechanisms, numerous achievements had been obtained in exploring the relationship between tumor growth and biomarkers. The exact mechanism of VS onset and progression has remained elusive, with only a broad association with genetic and epigenetic aberrations of merlin, inflammation factors, proteolytic enzymes and nutrient supply pathways. Notably, there are mutual interactions between abnormally expressed molecules in VS biology. Merlin, whose conformational changes are regulated by RTKs and integrin signals, is also an inhibitor of the pro-inflammatory transcription factor NF- κ B (30). On the other hand, high numbers of tumor infiltrating leukocytes in VS tissues can enhance cell survival, promote tumor growth and degenerative changes through the production of growth factors, cytokines, and MMPs (33, 128, 158). These evidences indicate that the progression of VS appears to be the result of interactions of these dysregulated pathways, drugs that target multiple signaling pathways simultaneously merit further investigation.

Despite the many recent advances in the identification of growth-related biomarkers, their translational researches have encountered several problems. First, current available VS animal models have some limitations. The commonly utilized animal models for the study of VS including patient-derived xenograft mice model (159), merlin-deficient Schwann cell or SC4 cells mouse

model (160, 161). Nevertheless, merlin-deficient Schwann cells may not accurately reproduce VS (162), and it takes more than 2 weeks for the construction of mouse model. Recently proposed zebrafish xenograft model with shorter period of model establishment (163), as well as the application of 3D *in vitro* cell culture system (164) and *ex vivo* organ cultures (165) that fully mirror the growth patterns of VS, promise to further elucidate the role of these biomolecules in the pathogenesis of VS. Second, clinical validation of these hypotheses should take tumor diameter and clinical course into consideration. Current studies demonstrate non-linear changes of protein expression as VS size increases (45), suggesting that tumor progression at different grades might be stimulated by different signaling pathways and can therefore be predicted by different biomarkers. Along similar lines, different stages of the natural course of VS might be driven by different mechanisms either. During the enlarge of VS, accelerated growth, regression, or quiescence of the tumor can occur at any stage (2). Classifying patients according to the growth characteristics of VS and analyzing biomarker levels in different groups would shed light on which pathway induces VS initiation and which involves in VS shrinkage or growth cessation. Third, research is urgently needed to address how to obtain tissue expression data in a non-traumatic way. MRI texture analysis have been previously associated with VEGF expression in head and neck squamous cell carcinoma (166), and might also reflected VEGF levels in VS. A more precise non-invasive *in vivo* marker expression evaluation technique is targeted molecular imaging [MRI, computed tomography (CT), positron emission computed tomography/single-photon emission computed tomography (PET/SPECT) and fluorescence] (161, 167–169). Recently, Morrison et al. injected the covalently linked compounds formed by anti-VEGFR2 or anti-Her2/Neu monoclonal antibodies and near-infrared probe into Schwann cell xenograft models in mice, and observed strong correlations between day 1 tumor fluorescence and eventual maximum tumor volume (170). Apart from advanced imaging techniques, the circulating biomarkers level could also serve as convenient and reliable indicators for the prediction of subsequent tumor growth (37, 44).

We highlight some of the known and novel indicators with potential to predict VS growth enlightened therapeutic targets of VS and are critical for clinicians in stratifying patient's risk preoperatively, formulating individualized therapeutic tactics, and guiding clinical trial enrollment.

AUTHOR CONTRIBUTIONS

YZ and JL wrote the first draft of the paper. JR and XH revised the draft. PZ and BW provided approval for publication of this review. All authors contributed to the article and approved the submitted version.

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