



Clinical and Preclinical Outcomes of Combining Targeted Therapy With Radiotherapy

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In the era of precision medicine, radiation medicine is currently focused on the precise delivery of highly conformal radiation treatments. However, the tremendous developments in targeted therapy are yet to fulfill their full promise and arguably have the potential to dramatically enhance the radiation therapeutic ratio. The increased ability to molecularly profile tumors both at diagnosis and at relapse and the co-incident progress in the field of radiogenomics could potentially pave the way for a more personalized approach to radiation treatment in contrast to the current "one size fits all" paradigm. Few clinical trials to date have shown an improved clinical outcome when combining targeted agents with radiation therapy, however, most have failed to show benefit, which is arguably due to limited preclinical data. Several key molecular pathways could theoretically enhance therapeutic effect of radiation when rationally targeted either by directly enhancing tumor cell kill or indirectly through the abscopal effect of radiation when combined with novel immunotherapies. The timing of combining molecular targeted therapy with radiation is also important to determine and could greatly affect the outcome depending on which pathway is being inhibited.

Keywords: cancer, DNA damage, combination (combined) therapy, radiation therapy, radiosenisitizing agent, targeted therapy

INTRODUCTION

A plethora of factors are involved in the development and progression of cancer in individuals such as family history, age, sex, primary site of origin and driver mutations; thus, treatment depends upon the goal of therapy - curative or palliative. Treatment for cancer involves multiple approaches including surgery, chemotherapy, immunotherapy, small molecules that target certain cancer signaling pathways, and radiation depending on cancer type or status. The use of multiple treatments concurrently is referred to as multi-modality treatment. Radiation therapy plays a crucial role in the management of cancer. Also known as radiotherapy (RT), it is a method of impeding cancer cell division by using high-energy ionizing radiation to induce DNA damage and disrupt cell cycle progression. In the treatment of cancer, RT can be given alone or coupled with

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chemotherapy or surgery and is aimed at reducing local tumor burden. The primary advantage, however, that RT confers over chemotherapy is the ability to precisely target the tumor and reduce systemic side effects. Epidemiological studies have reported that almost 54% of breast cancer survivors were treated with radiation therapy in 2016 and this is projected to become 60% by 2030 (1). Treatment mode is usually determined by stage and type of cancer, genetic mutations, age, and overall health of patient.

RT can be delivered in several ways; the most commonly used modality is broadly defined as External Beam Radiation Therapy (EBRT), which includes Stereotactic Body Radiation therapy (SBRT) and Stereotactic Radiosurgery (SRS). EBRT most typically uses a linear accelerator to deliver radiation directly into the cancer site in the form of photons. Depending on the location of the tumor, this radiation can be of high or low energy. For instance, high energy EBRT is used in the treatment of head and neck cancer, breast, lung, and eye cancer (2-5) while lower energy photons are used for more superficial cancers such as melanoma (6). Another modality of delivery is brachytherapy, which utilizes a radioactive source placed as close to the tumor as possible and can be given in conjunction with EBRT (5, 7, 8). Some examples of cancers where brachytherapy is frequently administered are cervical, vaginal, and prostate cancer (8-11). Ideally, RT will preferentially or more frequently damage DNA of cancer cells, with less or reparable damage to surrounding healthy cells. Similar to the brachytherapy concept, IntraOperative Radiation Therapy (IORT) constitutes the precise delivery of radiation to the tumor/tumor bed during surgery while minimizing exposure to the surrounding healthy tissues. IORT can be done utilizing electrons, low-kV X-rays, and high dose rate (HDR) brachytherapy. TARGIT, an international randomized clinical trial designed to test the hypothesis that delivering a single dose of targeted IORT in patients eligible for breast conserving surgery (+ EBRT in patients at high risk for local recurrence) is equivalent to a conventional course of postoperative EBRT showed that there was no statistically significant difference between EBRT and the IORT approach with respect to local recurrence-free survival, invasive local recurrence-free survival, mastectomy-free survival, distant disease-free survival or breast cancer mortality (12). In a study looking at brain metastases, retrospective data suggests that IORT is a safe and effective tool in the adjuvant setting following surgical resection of brain metastases; an area that continues to be under debate (13). IORT is currently under investigation in the adjuvant setting following the maximal safe resection of recurrent glioblastoma multiforme (GBM) (NCT04763031, NCT04681677).

Conventional fractionated EBRT was traditionally based off the classical "four R's" of radiation biology: reassortment, repair, reoxygenation, and repopulation (14), to which radiosensitivity was later added (15). IORT on the other hand is generally performed with either low energy X-rays or electrons; both of which are considered low linear energy transfer (LET) radiation compared to high energy X-rays used in conventional EBRT. Unlike high LET radiation where the linear quadratic model (L-Q) model predicts that radiobiological effectiveness (RBE) should decrease as the dose per fraction increases (16), evidence suggests that this may not be true for low-LET radiation. With a predicted higher RBE, emerging evidence suggests that IORT can be effective by overwhelming the repair system leading to increased genomic instability and thus more cancer cell killing. Additionally, IORT performed during surgery eliminates repopulation of residual tumor cells in the tumor bed, which could theoretically happen during wound healing (17). The ability of IORT to eliminate repopulation could also be attributed to the radiation-induced bystander effect (RIBE) which is thought to be more common with high dose/fraction as is the case with IORT. Abscopal effect in normal nonirradiated cells in the vicinity of tumor could reduce tumor recurrence, modifying the wound microenvironment, and eradicating residual tumor cells when applied immediately after surgical procedure (18).

Additionally, SBRT or SRS is used to deliver very high doses of radiation to the primary sites or metastatic sites in few treatments (1-5), with extraordinary precision made possible by real-time monitoring of the patient under CT scan throughout the duration of therapy. Together they can be combined into a term Stereotactic Ablative Radiotherapy (SABR). Unlike IORT, which arguably does not fit the current L-Q model, current data suggests that this is not the case for SABR, which behaves biologically similar to conventionally fractionated EBRT. However, the higher tumor control that is achievable with SABR when compared to conventional EBRT is attributed to a more geometrically precise technique of dose delivery that allows for prescribing high biological effective doses (BED), which were simply unachievable with conventional dose delivery techniques (19). Additionally, ablative effect on the surrounding tumor endothelium provides additional mechanism of death that is not as prominent in conventionally fractionated EBRT. Emerging data suggest that better tumor control with SABR could also partly be attributable to the abscopal effect brought about by high dose radiation in nonirradiated cells such as enhanced endothelial cell damage and/or enhanced tumor immunity similar to what was suggested in the setting of IORT (20).

HOW DOES RADIATION WORK: THE BIOLOGIC EFFECTS OF RADIATION

Effects of Radiation Therapy: DNA Damage

Ionizing radiation introduces energy into molecular structures which then releases electrons creating ions that are capable of breaking covalent bonds. The breakdown of these covalent bonds within DNA produces DNA breaks, including double-stranded breaks. Radiation also leads to the generation of reactive oxygen species (ROS) which oxidize lipids and proteins and are capable of damaging DNA in many ways, including single-strand breaks. This damage leads to cell death and failure of mitosis.

Consequently, highly proliferating cells are most susceptible to damage due to radiation. DNA damage is not an uncommon phenomenon, with as many as 50,000 lesions, or instances of

DNA damage, in each cell, every day. Cellular mechanisms of DNA repair are able to fix this continuous damage and maintain functional DNA. Endogenously induced lesions are generally isolated and more evenly distributed throughout DNA. Damage resulting from radiation is far less dispersed. When two or more lesions are found within two helical turns, this is referred to as a clustered damage site, and these are far more difficult to repair than isolated lesions (21, 22). The most highly damaging effect of ionizing radiation is considered to be the double-stranded DNA breaks where both phosphodiester backbones of the two strands of DNA are broken within 10 base pairs (23-28). Doublestranded DNA breaks are likely particularly cytotoxic as they are not regularly induced endogenously (28-30). The linear energy transfer (LET) ratio of the radiation determines the type of damage it induces in the DNA. Particles with a higher LET (e.g., protons, neutrons, alpha particles) results in roughly 90% of the damage occurring in the form of clustered damage sites, while low LET radiation (e.g., gamma rays, x-rays, and electrons) produces roughly 70% of its damage as isolated lesions and the remaining 30% in the form of clustered damage sites (23, 31).

Radiation kills cancer cells either by damaging the DNA directly or generating excessive ROS which damages the DNA (**Figure 1**). However, cancer cells can become resistant to RT *via* several mechanisms which enhance their DNA repair capacity or suppress the functions of tumor suppressors (32). Therefore, strategies that disrupt the DNA repair machinery or the detection of DNA damage has largely been explored to enhance radiosensitization of tumors. Inhibitors of DNA repair proteins have widely been studied alone or in conjunction with radiotherapy to enhance tumor suppression. For instance, the inhibition of the DNA base excision repair (BER) protein apurinic/apyrimidinic endonuclease, APE1, has been shown to suppress growth of several cancers (33, 34). Similarly, overexpression of APE1 has been linked to radioresistance (35,

36), and suppression has been shown to enhance cancer cells to RT (37). Inhibition of several other DNA repair proteins such as Poly (ADP-ribose) polymerase (PARP) and ataxia telangiectasia mutated (ATM) have demonstrated similar effects (38, 39). The quantity and characteristics of DNA damage are also impacted by the tumor microenvironment, with the oxygen levels of the tumor being of particular importance. Hypoxic tumors do not respond as well to radiation therapy compared to tumors that are well oxygenated. This is because oxygen reacts very quickly with DNA radicals that result from radiation to produce DNA lesions when it is present. Molecules that will react with the DNA radicals can be introduced and function in a similar capacity to oxygen, such as nitroaromatic compounds (e.g., nimorazole, nitrotriazole or sanazole) (40, 41). Nitric oxide is another molecule that is of interest in this regard, though some of its effect may be due to increased oxygen tension of the tumor microenvironment (TME) (42). Due to the potential clinical impact, many preclinical studies have investigated the use of radiosensitizing agents to increase tumor cells' susceptibility to RT which will be discussed in the sections below (43-46).

Effects of Radiation Therapy: Cellular Damage

Traditionally, RT has been reported to arrest cancer cell proliferation by inducing DNA damage through stimulation of cell death mechanisms such as apoptosis, necrosis, and senescence. However, radiation can also inhibit cell proliferation by disrupting the neoplastic cells physically through damage to the cell membrane and organelles, and thereby interfering with signal transduction (47–49). Damage to several organelles including the endoplasmic reticulum, ribosome, lysosome, and mitochondria have been implicated in the effects of RT-induced tumor cell death (50–59).

The mitochondria, in particular, is an important target of RT as it regulates cellular respiration and metabolism, and altered



metabolism is considered a hallmark of cancer (Figure 1) (60). RT-induced damage within the mitochondrial DNA can induce programmed cell death in cancer cells (61). The mitochondrial respiratory chain generates ROS as a byproduct of cellular respiration in normal cells. On the other hand, excess ROS production can potentiate tumor growth. Together, this suggests that cellular response to ROS varies according to levels of ROS generated in the cells. For instance, tumorigenic events such as hypoxia or oncogene activation can induce tumor growth by generating abundant ROS to drive cell cycle progression, metastasis, angiogenesis, etc. However, RT can generate an ROS overload which can arrest the cell cycle and induce apoptosis through mitochondrial collapse in cancer cells (62, 63). For example, FLASH radiation is a novel radiotherapy technology, defined as a single ultra-high dose-rate (\geq 40 Gy/s) radiotherapy, which unlike conventional dose-rate radiation (described above) leads to strikingly differential responses between healthy and tumor tissues. This differential effect has been attributed to multiple theoretical mechanisms such as distinct mechanisms of DNA damage and the significantly higher ability of FLASH to produce ROS at a rate that can't be scavenged by tumor cells compared to healthy cells which have a lower oxidant load and higher catalase reduction reserve capacity. More future studies are needed to better understand the mechanism of FLASH and its clinical implications (64).

Several strategies targeting the mitochondria to sensitize cancer cells to RT have been investigated (65-67). The mitochondrial respiratory chain generates ROS as a byproduct of cellular respiration, and RT also generates an ROS overload which can induce apoptosis through mitochondrial collapse in cancer cells (62). LKB1 (also known as serine-threonine kinase 11, STK11) is a tumor suppressor and functions in the AMPK (adenosine monophosphate-activated protein kinase) pathway necessary for cell metabolism, homeostasis, and autophagy (68). In esophageal cancer, overexpressed LKB1 has been reported to confer resistance to radiation therapy, activate autophagy, and inhibit apoptosis (69). One of the metabolic changes that cancer cells initiate during low glucose conditions is the switch from glycolysis to oxidative phosphorylation (OXPHOS) to adjust to fluctuating microenvironmental conditions (70, 71). Irradiated human esophageal adenocarcinoma cells had a higher number of mitochondria with additional mitochondrial mutations compared to their non-irradiated counterparts. Analysis of patient tumors of esophageal adenocarcinoma showed an increase of ATP5B, a marker of OXPHOS, in patients who had poor response to neoadjuvant chemoradiation therapy, suggesting that changes in mitochondrial metabolism can potentially play a role in radioresistance (71).

RADIOGENOMICS AND RATIONAL DESIGN FOR RADIATION-TARGETED THERAPY COMBINATIONS

The combination of radiation therapy and traditional cytotoxic chemotherapy is a clinically well-established approach to

improve overall survival of cancer patients (72). However, to date, despite the significant advancements in developing molecularly targeted therapy, little progress has been made in identifying and defining optimal targeted therapy and radiotherapy combinations to improve the efficacy of cancer treatment (73). The rapidly growing arsenal of targeted therapies can be categorized according to their respective effects on one or more of the hallmarks of carcinogenesis which were coined by Hanahan and Weinberg (74, 75). Importantly, the clinical success of these agents was largely based on the identification of predictive biomarkers of response, which enabled the selection of patients and/or tumors that would benefit from these novel agents. This subsequently led to the rise of precision medicine and simultaneously sparked interest in the concept of 'precision radiation medicine', yet that concept remains in its infancy.

Precision radiation medicine proposes to leverage genomic information derived from human cancers or preclinical tumor models to identify subsets that are sensitive to specific radiation/ drug combinations, radiation alone at tailored doses or predict those at high risk for radiation-related normal tissue side effects (76, 77). As our knowledge of how radiation works evolved over time (as outlined above), several groups have attempted to characterize preclinical models, particularly cell lines to identify genomic signatures that are predictive of radiation sensitivity. The largest effort to date was done by Yard et al., who underwent large-scale profiling of cellular survival after exposure to radiation in a diverse collection of 533 genetically annotated human tumor cell lines and were able to demonstrate the wide range of radiation susceptibility and the novel genetic features driving that diversity (78). Currently, there are several genomic signatures that have been clinically validated for guiding radiation treatment. For example, OncotypeDX[®], a 21 gene classifier that was initially validated to predict the benefit of adjuvant chemotherapy in hormone receptor positive breast cancer, is currently used to estimate the risk of locoregional recurrence after radiation for invasive breast cancer and therefore guide addition or omission of radiation in the adjuvant setting (79). Similarly, for ductal carcinoma in situ (DCIS), DCISionRT[®] is a multigene assay (80) that has been prospectively validated in 327 patients with DCIS that participated in the E5194 trial (81) to help inform decisionmaking regarding the addition of radiation in the adjuvant setting in conjunction with clinic-pathologic criteria (82, 83). Decipher[®] is a 22 gene classifier that was developed as a prognostic tool for men with high-risk prostate cancer and was prospectively validated to guide that addition of postprostatectomy radiation in that risk group whether in the adjuvant or salvage setting (84, 85). In the 2019, the American Society of Clinical Oncology (ASCO) guideline on molecular markers in localized prostate cancer, only Decipher was recommended to guide the decision between salvage and adjuvant radiation and Decipher® PORTOS was the only predictive signature of radiation response (86). Nonetheless, salvage radiation is generally preferred based on randomized data (87) and so far genetic testing is not part of the standard of care to guide radiation timing until validated in the randomized

setting (NCT02783950) (88). In a collaborative novel effort to personalize radiation dose based on genetics and transcend the 'one size fits all' paradigm, a novel algorithm that uses genomic adjusted radiation dose (GARD) was proposed to independently quantify differences in clinical outcomes across different cancers that are not attributed to the physical radiation dose alone. This effort aims to guide the integration of genomics into radiation dose decisions (89–92).

While several genomic signatures have been studied in the preclinical setting and a few have been clinically validated to better tailor radiation therapy, limited clinical trials with RT were designed to prospectively test whether specific patient subpopulations with distinct genomic signatures would benefit from radiation or not. For example, HN002 is a phase II study that evaluated radiation dose de-escalation in patients with human papilloma virus (HPV) positive oropharyngeal cancers who are thought to have improved survival outcomes due to impaired DNA repair (93-95). In that study, radiation dose deescalation was found to be non-inferior to standard dose, which justifies hypothesis testing in the phase III setting. Another eloquent example is in pediatric medulloblastoma where several trials are investigating tailoring radiation dose and technique based on distinct molecular subgroups rather than clinic-pathologic characteristics per say (96). Recently, the ACNS0331 trial demonstrated that reduction of boost volume but not craniospinal radiation dose is safe in average risk medulloblastoma patients and this may occur in a genetic subgroup-dependent manner (97).

The equally important aspect of radiation therapy, which is crucial for an optimal therapeutic ratio, is better understanding and prediction of normal tissue toxicity, particularly late side effects, which are usually irreversible and can severely impact quality of life. While demographic and clinical factors are wellrecognized culprits of late tissue toxicity, the evolving field of radiogenomics proposed genetic factors as key players as well. Kerns et al. proposed two arching goals for the field: first, identifying key molecular pathways that can predict radiationinduced normal tissue toxicity and second, developing an assay to identify the patients who are more likely to develop late tissue toxicities and therefore require tailored treatment (98). Several genome-wide association studies have identified associations between specific single nucleotide polymorphisms (SNPs) and radiation toxicity (99-101). The REQUITE international prospective toxicity profiling effort, initiated by The Radiogenomics Consortium, represents the largest study to date in that regard and has led to the creation of a centralized database of relevant clinical information including treatment, dosimetry, toxicity, and genome-wide SNP genotyping data in an effort to prospectively validate these findings for clinical use (102, 103).

Despite the efforts outlined above, the radiation oncology field significantly lags behind in designing clinical trials that are poised to prospectively test whether specific combinations of radiation and targeted therapy can particularly benefit a genomically distinct patient population. To that end, several collaborative efforts aimed to outline guidelines to usher the field toward optimizing the clinical development of novel drugradiotherapy combinations. Two key points were proposed: 1) reconsidering novel endpoints in clinical trial design such as local control, organ preservation, and patient reported outcomes, and 2) prioritizing the development of promising therapeutics that target relevant pathways to radiation such as DNA repair inhibitors and immunotherapies (104–106).

Traditional radiosensitizing agents (such as cisplatin and 5fluorouracil) typically exert their effect by augmenting DNA damage (72). As large genomic studies continue to unravel the landscape of DNA repair pathway deficiencies across different tumor types, it will be critical to propose novel rationally designed combinations of radiation and targeted therapy that fit specific genomic contexts (77). PARP1, WEE1, DNA-PK, ATM, ATR, and CHK1 are among the most critical mediators of DNA damage response (DDR) (Figure 1). DDR inhibitors (such as PARP inhibitors) were initially developed as monotherapy to target DDR defects that are present in tumor cells, but not in normal cells. This selectivity gave rise to the concept of synthetic lethality (107). Theoretically radiation is an attractive DNAdamaging agent that can be combined with novel DDR inhibitors to promote cell-selective radio-sensitization by three mechanisms: firstly, by increasing the amount of DNA damage to levels that induce apoptosis or cell death mechanisms rather than DNA repair or cell cycle arrest, secondly by exploiting synthetic lethality, and thirdly, by augmenting DNA damage and thus increasing the tumor mutation burden which in turn enhances tumor antigenicity and thus T-cell mediated killing (108).

Preclinical evidence suggests that DDR inhibitors can act as potent radiosensitizers and potentially have greater cytotoxic effects in cancer cells compared to normal cells (Figure 2). This also brought about the idea of synthetic lethality in which cancer cells, unlike their healthy counterparts, carry DNA repair defects, making them particularly vulnerable to DDR inhibitors, especially when simultaneously targeted with a DNA damaging agent such as radiation (109, 110). For example, PARP inhibitors have been shown to be potent radiosensitizers, irrespective of the tumor's homologous recombination (HR) status (111), albeit at lower doses in HR-deficient tumors (112). Similarly, Adavosertib, a WEE1 inhibitor is also an effective radiosensitizer (113, 114). Inhibition of WEE1 abrogates the G2/M checkpoint which is crucial for P53 mutant cancer cells, which also lack the G1 checkpoint. Therefore, WEE1 inhibition represents another form of tumor-selective radiosensitization (115). Induction of replication stress is another appealing mechanism that can selectively enhance radiation sensitivity in cancer cells particularly in the context of cMyc and KRAS mutations (116, 117). Several DDR inhibitors including PARP, WEE1, and ATR inhibitors have been implicated in the induction of replication stress either as monotherapy or in combination with other DDR inhibitors together with RT (118). Several clinical trials are currently testing the premise of combining radiation with DDR inhibitors in various disease sites.

In the era of immunotherapy, modulation of the host and the tumor microenvironment holds a lot of promise when combined with radiation as demonstrated in a plethora of eloquent



preclinical studies. Radiation and immunotherapy agents are thought to interact through five distinct mechanisms based of the modified Steel hypothesis (119): (1) spatial cooperation, (2) temporal modulation, (3) biological cooperation, (4) cytotoxic enhancement, and (5) normal tissue protection (120). Radiation has immunostimulatory and immunosuppressive effects. Radiation can induce immunogenic cell death and increase expression of tumor specific antigens and thus sensitize tumors to the effects of immunotherapy (121, 122). In the preclinical setting, Twyman-Saint Victor et al. demonstrated synergy between radiation therapy and combined anti-PD-1/PD-L1 and anti-CTLA4 blockade. In this study, the combination led to an increased response within the tumor as the radiation induced the diversification of the T-cell repertoire in tumorassociated lymphocytes and the immune checkpoint inhibitors inhibited T-regulatory cells (Tregs), which resulted in an increase in the CD8/Treg ratio and subsequently led to improved outcomes compared to either modality alone in a variety of tumor models (123). The abscopal effect of radiation refers to another form of RT-immunotherapy synergy where anecdotal studies (mostly in patients with melanoma) have shown tumor response in non-irradiated lesions presumably due to an incited systemic immune response resulting from local radiation treatment (20, 124-127). Conversely, radiation can promote tumor infiltration by suppressive regulatory T cells, inhibitory macrophage and myeloid-derived suppressor cell lineages (128, 129), therefore combination with immunotherapy in that context is crucial to maintain the anticipated cytotoxic effect of RT. The optimal dose, fractionation, volume, and sequencing of RT with immunotherapy remain to be elucidated to strike the balance

between the immunostimulatory and immunosuppressive effects of radiation and to fulfill the modified Steel criteria (76, 77, 120).

Thus far, the failure to predict treatment efficacy using genetic variables represents one of the most significant obstacles to the personalization of radiation-based treatment regimens. The potential success of radiosensitizing-targeted therapy is contingent upon our better understanding of radiogenomics, which pertain to defining biomarkers of response and genetic determinants of late tissue toxicity (106, 130, 131). Moving forward, two key concepts need to be considered in order to facilitate rational design of novel radiation-targeted therapy combinations that are effective: redefining end points of interest and efficacy and identifying and validating biomarkers that can enable the early identification of ineffective or toxic compounds. These two key concepts will require the optimization of preclinical models that can accurately recapitulate the complexity of human tumors and thus faithfully predict promising combinations and subsequently re-thinking clinical trial design in a way that is relevant to radiation and its paradigm.

BUILDING PREDICTIVE EXPERIMENTAL MODELS IN THE VALIDATION OF COMBINATION THERAPY THAT INCLUDES RADIATION

For a small molecule to be maximally effective as radiosensitizer, it must be highly specific and directly toxic to the tumor. Tumor cells depend more heavily on certain signaling pathways over normal tissues, therefore combination of RT with small molecule inhibitors of these pathways offers an alternative strategy to chemoradiation that is potentially less toxic to surrounding healthy tissues. A general limitation to this is the lack of preclinical models that mimic the human cancer to a molecular level which provides information regarding predictive biomarkers that differentiate between radioresistance and radiosensitivity.

Preclinical models for studying cancer radiogenomics as well as cancer efficacy studies require recapitulation of human cancer on an anatomical and histological level in a manner that closely mimics the human tumor characteristics. The driver or passenger mutations, microenvironment, hypoxia, angiogenesis, immune components, and therapeutic response are all important factors to consider. Therefore, several approaches are being used to build multi-cellular in vitro models as well as in vivo models with appropriate genetic manipulations to capture the aforementioned characteristics in response to RT. Methods include genetic knockdown, knock-in, activation, tissue-specific expression, inducible expression, and sequential expression in traditional cell culture, 3-dimensional (3D), organoid, and xenograft models. My laboratory has focused on generating 3D mono- and co-cultures using various cancers such as pancreatic, colon, and bladder (132-135). The use of both tumor cells and CAFs with distinct fluorescent markers allows us to monitor the effects of both cell populations following selective pathway inhibition. For example, we demonstrated the enhancement of tumor cell killing with dual inhibition of APE1/Ref-1 as well as CA9 (carbonic anhydrase 9), a HIF-1 α target. Through blocking the full activation of HIF-1 α through APE1/Ref-1 and the cells ability to respond to changes in pH through CA9, the spheroid growth was dramatically reduced (135). This model is now being interrogated to understand the effects of RT on growth of the spheroids and the impact on the cells of the TME as well as RT in combination with targeted agents that would impact hypoxia as well as metabolic signaling.

In vitro models often use a panel of radiosensitive and radioresistant cell lines and compare the effects of select small molecule inhibitors or the effects of knocking down potentially important signaling molecules. Other approaches include generation of radioresistant lines and determining which molecular factors play a role in their resistance. 3D models can aid in recapitulating the cell-cell interactions within tumor and stroma, cytokine signaling, hypoxia response, and combination therapy involving RT and allow us to quantitate the effects on the tumor as well as cells from the TME such as CAFs (136-138). A study comparing radiosensitive and radioresistant non-small cell lung cancer (NSCLC) demonstrated that pathways previously implicated including DNA repair, apoptosis, and NFKB activation in NSCLC were involved in the cellular response to RT (54). Prostate cancer cell lines and the transgenic mouse model TRAMP (Transgenic adenocarcinoma of mouse prostate) used natural product, Nexrutine (Nx), to sensitize the prostate cancer cells to RT both in vivo and in vitro. Downregulation of ribosomal and cell cycle proteins as well as HIF-1 α were implicated in the sensitization of the tumors to Nx (56). These are just two examples of preclinical studies that utilize various

models to test the radioresistance and sensitivity of various cancer types. The predictability of the model and the complexity of the 3D or monolayer system in response to RT will enable the preclinical studies to have a greater impact on the rationale design of combination therapy which will ultimately lead to translational impact.

RATIONAL COMBINATIONS OF RADIATION AND TARGETED THERAPY IN THE PRECLINICAL SETTING

PARP proteins are involved in DDR and inhibitors of PARP have been widely studied for radiosensitization both preclinically and in the clinic (discussed below and Figure 2). Currently, there are four PARP inhibitors in the clinic: Olaparib, Rucaparib, Niraparib, and Talazoparib (Table 1). The efficacy of this combination therapy has also been studied in preclinical models of human non-small cell lung cancer (NSCLC): Calu-3 and Calu-6 cell lines. Even though both cell lines exhibited increased radiosensitization following Olaparib treatment in vitro, only xenografts of Clau-6 showed increased response to combination RT in vivo. Difference in response between Clau-3 and Clau-6 were most likely due to microenvironmental factors that contributed to the sensitivity of cells, indicating that preclinical modeling must be approached unbiased and carefully with the appropriate TME (139). Talazoparib and Niraparib have also been studied for their sensitizing effects. Primary melanoma cultures treated with combination therapy of Talazoparib, Niraparib and radiation, demonstrate that both PARP inhibitors sensitize melanoma cells to IR (162). A shortterm phase 1 clinical trial looking at the efficacy of combination therapy of radiation and Olaparib has determined the safety of the combination regimen in doses up to 200 mg/day without any side effects (163).

Apurinic/apyrimidinic endonuclease 1/Redox factor-1 (APE1/Ref-1) possesses multiple functions that could affect the cellular response to RT (Figure 2). APE1/Ref-1 is key in the base excision repair (BER) pathway of DNA lesions, acting as the major AP endonuclease in both the nucleus and mitochondria and in eukaryotic transcriptional regulation of gene expression as a reduction-oxidation (redox) factor (164-166). APE1 contributes to the repair of ionizing radiation through its ability to repair a 3'-phosphoglycolate end within a DNA strand break that is generated following ionizing radiation (IR) (167). A decrease in expression of APE1/Ref-1 in cancer cells results in apoptosis, cell cycle arrest, a decrease in proliferative capacity, a blockade of mitochondrial metabolism, and sensitization to various anti-cancer agents including RT (166, 168-170). Biochemical studies using oligonucleotides with clustered damage sites as would be encountered in a cell following RT demonstrate that APE1/Ref-1 can repair these types of DNA lesions (171). An inhibitor of the DNA repair activity of APE1/Ref-1 has been difficult to identify and develop preclinically, therefore two recent studies in pediatric and adult

TABLE 1 List of radiosensitizers	, respective mechanism of a	actions and preclinical mo	dels used to study them.
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Radiosensitizer	Mechanism	Cell models studied	References
Olaparib	Blocks DNA repair by inhibiting PARP	Breast cancer: MCF-7, MDA-MB-231, MDA-MB-231, T47D, BT-549, HCC-1954	(139, 140)
Rucaparib	Blocks DNA repair by inhibiting PARP	NSCLC: Clau-3, Clau-6 Cervical cancer: HeLa Prostate cancer: PC3, LNCaP, DU145, VCaP Neuroblastoma SK-N-BE(2c), UVW	(141–143)
Cetuximab	Inhibits epidermal growth factor (EGF) from binding to its receptor	HNSCC: HN30, HPV-negative HTB-43, UM-SCC1, UM- SCC2, UM-SCC6, HPV-positive UM-SCC47, UPCI : SCC090 cells	(144, 145)
Telaglenastat	Interferes with mitochondrial metabolism by inhibiting the conversion of glutamine into glutamate	HNSCC: FaDu, HN5, CAL-27 Lung Cancer: H460, A427, A549	(146, 147)
Tirapazamine	Selective for hypoxic cells; Generates reactive oxygen species which cause DNA damage	Human Nasopharyngeal Carcinoma: HNE-1 Cervical cancer: HeLa	(148, 149)
Everolimus	Inhibits mTOR kinase	NSCLC: NCI-H460, NCI-H661 Glioblastoma: GS-2	(150, 151)
Nimorazole	Generates reactive oxygen species which cause DNA damage	HNSC: HPV-negative FaDu, UTSCC5, UTSCC33 and HPV positive: UMSCC47, UDSCC2 UPCISCC90	(152)
Trametinib	Inhibits MEK	NSCLC: A549, H460 Melanoma: A375, D04, WM1631, WM1791c	(153, 154)
Adavosertib Peposertib	Inhibits Wee1 and impairs the G2 DNA damage checkpoint Inhibits DNA-PK and impairs DNA repair	Esophageal Cancer: OE33, FLO1 Leukemia: Molm-13, Molt-4 HNSCC: FaDu Colon Cancer: HCT116	(155) (156, 157)
Silver NP	Deposit high levels of energy in cells when exposed to ionizing radiation; ROS generation and DNA damage	Glioma: C6 Colon cancer: HCT116, HT29	(158, 159)
Gold NP	Deposit high levels of energy in cells when exposed to ionizing radiation; ROS generation and DNA damage	Breast Cancer: SK-BR-3	(160)
Bismuth NP	Not fully understood; Possibly by depositing high levels of energy in cells when exposed to ionizing radiation; ROS generation and DNA damage	Breast cancer: MCF-7, 4T1	(161)

brain tumors utilized nanoparticle delivery of APE1/Ref-1 siRNA to achieve sensitivity to RT (168, 169). One of APE1/ Ref-1's interacting protein partners is nucleophosmin 1 (NPM1) and perturbation of the APE1/Ref-1 - NPM1 interaction can lead to decreased DNA repair activity of APE1/Ref-1 and increase in sensitivity to chemotherapeutic agents such as bleomycin (172, 173). Recently in NSCLC cells, radiosensitizing agent YTR107 was shown to bind to NPM1, disrupt RAD51 foci formation, and synergize with PARP inhibition (174). These findings highlight the complex interplay between radiation-induced DNA damage and repair and the potential proteins that can be exploited as drug targets to sensitize cancer cells to RT. Due to APE1/Ref-1's role in the repair of DNA lesions induced by RT, the blockade of APE1/Ref-1 DNA repair activity could be highly effective in combination with RT. The caveat of course would be toxicity to normal tissues, and therefore development of tumor targeting strategies would be of paramount importance.

In addition to DNA repair activity, APE1/Ref-1 also plays an important role in signaling within the tumor and TME through the transcription factors (TFs) it regulates, and many of these TFs also play a role in inflammation (166). Functioning as a redox factor, APE1/Ref-1 stimulates the DNA binding activity of TFs by reducing cysteine residues within the TF (175). APE1/ Ref-1 activates TFs including HIF1a, STAT3, p53, NF-kB and others that directly govern critical cellular functions, including hypoxia, DNA repair, inflammation, and angiogenesis (166). Cells, both tumor and normal, possess reduction-oxidation

systems such as NRF2, thioredoxin, peroxiredoxins, and glutathione. In contrast, APE1/Ref-1 functions as a signaling molecule rather than a general redox system (176, 177). Our team has extensively characterized APE1/Ref-1 redox signaling inhibitors in several indications including cancer as well as chemotherapy- or IR-induced neuropathy (165, 178, 179). Vasko et al. demonstrated that the DNA repair function of APE1/Ref-1 was protective against the neurotoxicity induced by IR and APE1/Ref-1 redox inhibitor, APX3330 could protect dorsal root ganglia against IR-induced cytotoxicity (179). Blockade of APE1/Ref-1's redox activity could also sensitize radioresistant cancer cells or remodel the TME to affect the tumor's response to RT as HIF, STAT3, NF-kB, and others have been strongly implicated in the cellular response to RT (180–183).

Finally, inhibition of DDR signals by enhancing p53 function has also proven to be effective for radiosensitization in preclinical models. Several strategies employed for this revolve around suppressing the functions of proteins that inhibit p53. For instance, mouse double minute 2 homolog (MDM2) inhibitors have widely been studied for combination radiotherapy in several different cancers which enhance anti-tumor effects *in vitro* and *in vivo* (184–188).

Moving on from DNA damage, the traditional culprit in radiation medicine, now significant interest exists in developing radiosensitizers that more selectively radiosensitize tumors, but not normal tissues, by targeting signal transduction pathways that are more commonly activated in tumors, such as the EGFR pathway (189, 190). Growth factors are essential for cancer cell proliferation and inhibition of apoptosis, and therefore, can contribute to radioresistance via several mechanisms, including activating proteins or pathways involved in repairing radiationinduced DNA damage (191, 192). Preclinical evidence has supported a radiosensitizing role for EGFR inhibition (193) and indeed, the addition of cetuximab to RT in patients with head and neck squamous cell cancer was shown to improve tumor control and overall survival compared with radiation alone (189). However, understanding the impact of the spectrum of EGFR alterations on radiosensitivity remains to be understood (194, 195). Similarly, the blockage of ERBB2 (human epidermal growth factor receptor2 [HER2]), which is commonly amplified in a subset of breast cancer (196), can reverse ERBB2mediated radioresistance (197). These findings were translatable into the clinic which was evident from a recent analysis of the HERA trial which demonstrated the potential of combining radiotherapy with trastuzumab in reducing loco-regional recurrence rates in breast cancer patients with 1 to 3 positive lymph nodes (198). In the prostate cancer field, the combination of androgen deprivation therapy (ADT) and radiation in patients with intermediate and high risk prostate cancer is a well-established approach to prolonging survival in that subset of patients (199). Despite being one of the earliest examples of combining radiation with targeted therapy, the mechanism of synergy between ADT and radiation remains controversial. Initially much of the benefit was thought to be derived by the orchestrated effect of radiation controlling disease locally in the prostate and ADT treating micrometastatic disease elsewhere (200). Newer preclinical data suggests that ADT has direct effects in the prostate that result in radiosensitization via several mechanisms including relieving hypoxia (201), suppressing DNA repair (202) and deactivating androgen receptor (AR). The blockade of AR signaling is thought to regulate the transcription of DNA repair genes and thus mediate radioresistance (203). The modulation of several other oncogenic pathways could provide another approach to enhance radiation sensitivity such as intracellular signaling (i.e. PI3K/AKT pathway) (204) and tumor-associated epigenetic changes (205).

As mentioned previously, the rationale for targeting tumor metabolism to sensitize cancer cells to RT is well-established. Mitochondrial metabolism is crucial to cancer cell survival and RT-induced mitochondrial DNA damage as well as excess ROS generation provides an attractive target to suppress cancer cell proliferation and induce apoptosis (146, 147, 206). Glutamine metabolism facilitates cancer cell survival, and breakdown of glutamine is mediated by glutaminases, making them the focus for development of small molecule inhibitors. Indeed, preclinical data supports the combination of the glutaminase inhibitor, Telaglenastat (CB-839), in radiosensitization of cancer cells (Figure 2). Telaglenastat suppresses cancer cell proliferation alone and in combination with 5-FU or EGFR in several colorectal and lung cell lines (207, 208). Combination therapy of radiation and Telaglenastat diminishes cancer progression in cell culture and mouse models of head and neck squamous cell

carcinoma. Clonogenic cell survival assays with FaDu (pharynx), HN5 (tongue), and CAL-27 (tongue) cell lines treated with radiation and Telaglenastat demonstrated significantly diminished proliferation compared to radiation or Telaglenastat treatment alone. These findings were confirmed using xenograft models in which combination therapy was superior to monotherapy (147). Similar results have been reported in lung cancer radiosensitization where treatment with Telaglenastat increased efficacy of RT by 30% in multiple cell lines and in H460-derived tumor xenografts (146).

Tumor hypoxia is another well-established mediator of radioresistance (209) and typically indicative of aggressive and treatment-resistant disease. Targeting tumor hypoxia by traditional cytotoxic chemotherapy has served as a cornerstone for concurrent chemoradiation regimens for decades. However, the validation of biomarkers of tumor hypoxia in patients that could guide the implementation of novel rationally designed combinations of radiation and hypoxia-targeting agents remains underexplored (105). Historically, several methods have been investigated in order to override hypoxia-mediated radioresistance. Such methods included: hyperbaric oxygen (210), oxygen mimetics which belong to the nitroimidazole class of agents (211), and hypoxia activated cytotoxic prodrugs such as tirapazamine (212). More recently, with the advent of the concept of normalizing tumor blood flow using anti-angiogenic therapy (AAT), several studies proposed RT-AAT combinations to alter oxygenation and improve therapeutic response. In xenograft mouse models, PI3K targeted inhibition led to improved tumor local control following radiation, which was associated with normalization of vasculature and increasing intrinsic radiosensitivity (213). In patients with NSCLC, PI3K inhibition led to reduction in tumor hypoxia as measured by FMISO PET in patients and was well tolerated in combination with palliative thoracic radiation (214). In GBM where angiogenesis is thought to be the hallmark of pathogenesis and VEGF its main driver (215), combining VEGF/EGFR with RT has been shown to halt the growth of glioma cells preclinically (216) and to have a significant synergistic anti-tumor effect with RT (217, 218).

The role of the tumor microenvironment on response to RT alone and in combination with chemotherapy or targeted agents is an important and understudied area. Stromal normalization is one approach to modulating the tumor microenvironment and reducing tumor hypoxia particularly with respect to radiation. Cancer-associated fibroblasts (CAFs) are naturally radioresistant, and data suggests that radiation can induce their pro-tumorigenic capabilities. However, the concept of combining RT with CAF targeting has not been investigated to date (219). Alternatively, another novel paradigm of targeting tumor hypoxia is the modulation of the tumor microenvironment by altering tumor metabolism through the inhibition of oxidative phosphorylation and thus decreasing tumor oxygen consumption rate and relieving hypoxia (220). Atovaquone, an FDA approved anti-malarial that functions through inhibition of mitochondrial complex III has been shown in pre-clinical models to alleviate tumor hypoxia and in turn results in tumor radiosensitization (221).

Finally, owing to rapid advances in nanotechnology, nanomaterials have attracted particular attention to enhance the anticancer efficacy of radiotherapy (158, 161, 222, 223). Nanoparticle delivery enhances tumor targeting while simultaneously improving effectiveness of radiotherapy by increasing local deposition of ionizing radiation dose or by augmenting production of ROS, DNA damage and cell cycle arrest (224). Silver nanoparticles were reported to sensitize both hypoxic and normoxic glioma U251 cells and C6 cells to radiotherapy (222). In additional studies, silver nanoparticles surface modified with polyethyleneglycol (PEG) and aptamer improved nanoparticle penetration and targeting in 3D glioma models, and conjugation with PEG/aptamer further enhanced radiosensitization in C6 xenograft models as well (158). The development of theragnostics further expand the scope of nanoparticles for multifunctional use (161). For instance, PEG conjugated bismuth gadolinium oxide nanoparticles (BiGdO3) not only sensitized breast cancer MCF-7 and 4T1 lines and 4T1 xenograft models to radiation, but the bismuth and gadolinium also allowed for MRI and CT imaging (161).

Even with the multitude of preclinical studies looking at combining RT with targeted therapy, chemotherapy, or immunotherapy, there are still very few examples of combinations that have translated into success clinically. We will now highlight some examples as well as future directions (**Table 1** and **Figure 2**).

RADIATION-TARGETED THERAPY COMBINATIONS IN THE CLINIC: STORIES OF SUCCESS AND FAILURE

A large body of preclinical evidence exists to support novel radiation-targeted therapy combinations. However, to date the EGFR inhibitor cetuximab remains to be the only molecular targeted agent approved by the U.S. Food and Drug Administration (FDA) for use with radiation therapy in head and neck cancer (189). Interestingly however the equivalence of cetuximab and cisplatin as radiosensitizers in head and neck cancer has been a crucial point of contention in the field. A small randomized trial by Margini et al. suggested that cetuximab was inferior to cisplatin when combined with radiation in patients with locoregionally advanced head and neck cancer (225). Two recent large, randomized trials have provided more conclusive evidence that cetuximab is indeed inferior. In the De-ESCALaTE Human Papilloma Virus (HPV trial), patients with low-risk HPV-positive oropharyngeal cancer had higher rates of local recurrence and lower overall survival when treated with cetuximab-RT compared to when treated with cisplatin-RT (226). That was also the case in the RTOG 1016 trial (227).

Although cetuximab was relatively successful as a radiosensitizer in the setting of head and neck cancer, it failed to show promising results in other cancers where EGFR signaling is relevant (**Figure 2**) (228–230). There is also a multitude of phase I/II data that demonstrated similarly disappointing results for other EGFR inhibitors. For example, EGFR is amplified in around 40% of GBM cases and its overexpression is associated with poor prognosis (231-233). Three phase II studies have examined the role of erlotinib, an oral tyrosine kinase inhibitor of the human EGF receptor that is FDA approved for the treatment of non-small cell lung and pancreatic cancers, given concurrently with RT plus temozolomide and have demonstrated widely contrasting results with respect to survival and toxicity. The overall trend however pointed towards increased toxicity with no substantial survival benefit. Phase I and II clinical trials have also been developed to study the combination of RT with erlotinib in pancreatic cancer in both the adjuvant and unresectable, locally advanced settings. Although toxicity profile was acceptable, only modest increases in efficacy have been observed (234-238). Alternative strategies for EGFR targeting have also been attempted in the early clinical settings. For instance, m-TOR targeting which is downstream of the EGFR/PI3K pathway have been trialed in the GBM setting. Two multi-institutional phase II studies have investigated the use of m-TOR inhibitor, Everolimus, in combination with standard RT plus TMZ, The North Central Cancer Treatment Group (NCCTG) N057K trial (239) and The Radiation Therapy Oncology Group (RTOG) 0913 trial (240). Despite having distinct designs, both trials showed no improvement in survival and increased toxicity. The rationale for the combination of EGFR inhibitors with RT is mainly based on the role of EGFR in driving the disease rather than on how the two modalities might work together to kill the tumor. Perhaps in future studies, combinations of RT with targeted agents need to be more rationally designed in order to see greater success clinically.

Another targeted radiosensitizer that has been relatively successful in the clinical setting is nimorazole. Nimorazole is a targeted radiosensitizer which selectively targets hypoxic tumor cells and has been shown in a phase III trial to significantly improve locoregional control by 16% in patients with cancer of the supraglottic larynx and pharynx when combined with radiation compared to radiation alone (241). However, nimorazole is currently only used in Denmark and has failed to become adopted as standard of care in the United States and elsewhere (242). In order to overcome hypoxia to sensitize tumors to radiation, Accelerated Radiation, Carbogen, and Nicotinamide, also known as the ARCON regimen, has demonstrated promising locoregional control rates and yet toxicity in a two large phase II studies in patients with head and neck cancer (243) and bladder cancer, respectively (244). This led to the phase III BCON trial which showed improved locoregional control and overall survival in bladder cancer patients who were treated using that regimen compared to patients treated with conventionally fractionated radiation alone (245). However, in a phase III study testing this regimen in laryngeal cancer patients, there was no significant improvement in either local control nor organ preservation rates in ARCON treated patients albeit with benefit in patients with hypoxic tumors (246). Taken together, this regimen has not been widely adopted due to practical difficulties in delivering this regimen, proper patient selection due difficulties in accurately determining highly hypoxic tumors, and inconclusive results from phase III data (247). Tirapazamine, the most clinically developed drug among hypoxia-activated cytotoxic prodrugs, which represent another class of hypoxia-targeted

radiosensitizers (212), have failed in phase III trials to demonstrate improved outcomes when combined with chemoradiation compared to conventional chemoradiation alone in both cervical (248) and head and neck cancers (249). Similarly, VEGF targeting which theoretically represents another attractive way of normalizing tumor vasculature and overcoming hypoxia, failed to improve OS in GBM patients where VEGF targeting was particularly alluring given its centrality to the disease pathogenesis (250-252). Interestingly however, another study showed that GBM patients that have increased tumor oxygenation following anti-angiogenic therapy when combined with conventional chemoradiation live significantly longer (253). Alternatively, targeting the stroma has been clinically attempted for radiosensitization with the goal of modulating RT-induced inflammatory responses (247). Recently, a phase II trial in patients with locally advanced pancreatic cancer has shown that addition of losartan to chemoradiation enhanced tumor shrinkage and enabled more margin negative resections likely due to interfering with TGF- β signaling in CAFs which are characteristic of the desmoplastic tumor microenvironment in pancreatic cancer (254).

Predictive biomarkers of response, which served as the premise of the systemic targeted therapy revolution, are needed in the radiation oncology field to improve trial design and success rates. To that goal, several early-stage clinical trials are currently underway; testing radiation resistance pathways that have been validated in the preclinical setting. For example, KRas, a proto-oncogene that is frequently mutated in a wide range of cancers (255) is a well-known driver of resistance to cancer therapy including radiation (256-258). Several exploratory clinical trials have demonstrated a link between KRas mutation status and decreased likelihood of locoregional control following radiation treatment (259-261). Midostaurin, a multikinase inhibitor that is FDA approved for treatment of FLT3 mutant acute myeloid leukemia (262) is currently being tested in phase Ib trial to be given concurrently with conventional chemoradiation in rectal cancer patients (263). This was based on an in vitro screen of 32 cell lines that represented lung, colorectal, head and neck, and genitourinary cell lines and identified Midostaurin as a potential radiosensitizer for KRas mutant cancers (264). Trametinib, a MEK inhibitor that is FDA approved for treatment of metastatic melanoma, is also being tested in a phase I trial in combination with chemoradiation for locally advanced KRas mutant NSCLC (265). Importantly, KRas has been so far inaccessible for direct inhibition until the recent FDA approval of sotorasib for the management of KRas mutated NSCLC based of the CodeBreaK 100 trial (266). It will be interesting to see how this could change the landscape of radiosensitization in the setting of KRas mutated cancer in the near future.

As discussed previously, DNA damage response is central to radiation response. However, so far there are many perceived challenges to clinically implementing this combination such as optimal sequencing, ideal genetic background, and importantly therapeutic window to avoid increased toxicity (267). There are numerous ongoing phase I/II trials combining radiation or conventional chemoradiation with novel targeted DDR inhibitors. Among DDR inhibitors, PARP inhibitors are the most clinically developed followed by WEE1 inhibitor, Adavosertib (Figure 2). In inflammatory or locally recurrent breast cancer, a phase I multicenter study evaluated veliparib, a PARP inhibitor, and concurrent RT for 30 patients. The study showed overall acceptable toxicity with only five (16.7%) patients experiencing a dose limiting toxicity (DLT) within 10 weeks from RT initiation. Although severe acute toxicity did not exceed 30% at even the highest dose, nearly half of the surviving patients demonstrated G3 adverse events at 3 years. Of the 30 patients, 15 experienced disease control failures during the 3 years of followup and 13 died which highlights the importance of long-term monitoring of toxicity in trials of radiosensitizing agents (268). A phase II trial comparing radiation with or without Olaparib in patients with inflammatory breast cancer, which is known to be particularly aggressive with dismal prognosis (269), is currently recruiting (NCT03598257). In pancreatic cancer, if the patient is homology recombination repair deficient (HRD), this may render the tumor particularly vulnerable to PARPi (270). Velaparib concurrent with chemo-RT was tested in a phase I study of 30 patients with locally advanced disease. Sixteen DLTs were detected in 12 patients (40%). Interestingly, median OS for DDR pathway gene-altered- and DDR-intact patients was 19 and 14 months, respectively. The most commonly mutated DDR gene was ARID1A (n = 4). Loss of ARID1A impairs both checkpoint activation and the repair of DSBs, which sensitizes cells to DSB-inducing treatments such as RT and PARP inhibitors (271). PARP inhibitors are also being tested in conjunction with other forms of targeted therapy such as EGFR inhibitors. A recent phase I study showed that Olaparib may be safely combined with concurrent cetuximab and radiation for patients with locally advanced head and neck squamous cell carcinoma who have a long smoking history. That combination has also demonstrated improved 2 year OS in that subset of patients compared to historical controls (72% vs 60% 2 year OS) (272). Other classes of DDR inhibitors such as WEE1 (Adavosertib), ATM, and DNA-PK inhibitors are currently being tested in phase I trials either in conjunction with radiation alone or chemoradiation in multiple disease sites. A recently completed phase I study evaluated Adavosertib in combination with RT and full-dose gemcitabine for 34 patients with locally advanced pancreatic cancer (273). In that study, median OS was 21.7 months which compares favorably with that of patients treated in the LAP07 trial (11.9-13.6 months), which had similar eligibility criteria and used gemcitabine (274). This sets Adavosertib as a promising drug in terms of clinical development compared to PARP inhibitors. The DNA-PKc inhibitor M3814 (Peposertib) has demonstrated promising anti-tumor activity in a recently published phase Ia study and is currently being tested concurrent with radiation in at least four phase I clinical trials covering different disease sites and different radiation fractionation regimens (275). ATM, ATR, and CHK1 inhibitors are also currently in several early phase clinical trials. Taken together, validating biomarkers of response for these novel agents to identify the subset of patients who will derive

the most benefit and the most acceptable toxicity in return remains to be a challenge (276).

Nanotechnology offers a new area of exciting research where nanoparticles can be used for targeted radiotherapy, either as sensitizers of external beams or as delivery vehicles for therapeutic radionuclides (277). In a phase II/III study, NBTXR3, a first-in-class radiosensitizer hafnium oxide nanoparticle, which is activated by radiation therapy, a significantly higher pathologic complete response was observed in the patients whose soft tissue sarcomas were injected with NBTXR3 prior to radiation compared to those who were not. There was no significant difference in toxicity between the two groups and no treatment-related death occurred (278). Although this is very promising data in the sarcoma field where very few patients achieve pathologic complete response with preoperative radiation and possibly in other cancers as well, a lot of challenges lie ahead for the clinical implementation of this technology and overcoming its limitations, particularly optimization of delivery (279).

The PACIFIC trial has revolutionized the management and therefore the outcomes of patients with locally advanced NSCLC. It has also set unprecedented clinical evidence supporting the interplay of chemoradiation and immunotherapy (280, 281). Importantly however it has posed many pressing questions regarding the optimal dosing, sequencing, and safety of combining radiation with immunotherapy. Currently, a plethora of clinical trials are attempting to answer those questions. Recently, the DETERRED trial demonstrated the safety and efficacy of adding Atezolizumab (anti-PD-L1) concurrently with chemoradiation (282) as well as the Phase 2 KEYNOTE-799 with concurrent delivery of Pembrolizumab (anti-PD-1) and radiation in locally advanced NSCLC (283). It will therefore be important to compare that regimen with the PACIFIC regimen where Durvalumab (anti-PD-L1) was given after chemoradiation in the consolidation setting. In head and neck cancer, a number of phase I/II clinical trials are testing the feasibility of combining chemoradiation with immunotherapy in the definitive setting. Collectively, those early studies have demonstrated the safety of the combination (284-287). A recent report by Weiss et al. showed that concurrent definitive immunoradiotherapy for patients with stage III-IV head and neck cancer who are ineligible for cisplatin had 24-month PFS and overall survival rates were 71% which exceeded their primary hypothesis (288). However, a substantial clinical benefit is yet to be proven in the phase III setting.

In the metastatic setting, several prospective trials have been conducted to test the abscopal effect of radiation, which stems from many anecdotal reports and arguably stimulated much of the hype regarding the combination of radiation and immunotherapy (289). The abscopal effect of radiation refers to the shrinkage or disappearance of sites of metastasis that were not directly treated with radiation. Although the mechanisms of this observation are still being elucidated, it is believed that the addition of immunotherapy to radiation regimens allows the immune system to mount a more systemic response against the tumor. PEMBRO-RT is a phase II study which asked the question whether stereotactic body radiotherapy (SBRT) enhances the effect of immune checkpoint inhibition in nonirradiated lung cancer lesions in metastatic NSCLC. In that study, patients with metastatic NSCLC were randomized to receiving pembrolizumab either alone or after SBRT, which was delivered to a single tumor site. There was a trend towards better overall response (ORR) and improved PFS in the combination arm but did not reach statistical significance. Interestingly, the benefit was more evident in patients with PD-L1 negative tumors and in subgroup analysis, improved ORR and PFS reached statistical significance in that group of patients (290). This again highlights the importance of discovering and understanding what molecular markers are important in the response to RT alone and in combination with targeted agents. In metastatic head and neck cancer, a similar phase II study randomized patients to either Nivolumab (anti-PD-1) alone or after SBRT to one metastatic site. Unfortunately the study did not find improvement in response, PFS, or OS between the two arms and there was no evidence of an abscopal effect with the addition of SBRT to Nivolumab in unselected patients with metastatic HNSCC (291). Interestingly however, in the neoadjuvant setting in early stage resectable NSCLC, concurrent SBRT and Durvalumab was safe and associated with significantly better pathological response compared to neoadjuvant Durvalumab alone demonstrating a robust evidence of abscopal immune-modulatory effect of radiation (292). These contrasting results could probably be attributed to the hypothesis that immunotherapy is generally more effective with less disease burden and therefore the abscopal effect could be captured in that setting (293). Taken together, phase III data is needed to validate the combinatorial benefit of radiation and immunotherapy in the metastatic setting and also better defining correlates of response based on biomarkers.

As outlined above there are many clinical trials testing different radiosensitization paradigms. That is not meant to be a comprehensive list but rather to paint a picture for the diverse nature of signaling mechanisms that could potentially be targeted to improve the therapeutic ratio of radiation. Importantly, while there are examples of successful radiationtargeted therapy combination in clinic, failures certainly outweigh those few successes. Therefore, a lot remains to be done in to decrease attrition rates of novel radiosensitizers in the clinic.

THE CHALLENGES AHEAD FOR CLINICAL IMPLEMENTATION

Oncology drug development has witnessed a significant growth over the last decade that was coupled with improved cancer outcomes and unprecedented drop in cancer related death rates (294). However, the development of novel radiosensitizers lagged behind reflecting lack of incentive by pharmaceutical industry to invest in this pipeline. This huge gap led to holding a collaborative workshop by the FDA-AACR-ASTRO in 2018 to bring together various stakeholders including representatives of academia, industry, patient advocacy groups and the FDA to identify key challenges and design a roadmap for bridging this gap (104). This effort was also preceded by similar efforts in the UK highlighting the



importance of this issue in the overall goal of improving cancer control rates where radiation therapy plays a central role as a curative and palliative treatment (105). As highlighted in Figure 3, the main challenges identified were: (1) lack of regulatory guidance by the FDA detailing the approval pathway for drug-radiotherapy combination particularly with regard to the extent of required preclinical data, (2) choice of adequate model systems that can reflect tumor complexity and heterogeneity and enable testing various radiation techniques and schedules, (3) complexity of the definition of 'safety' in the radiation setting as it should take into account normal tissue toxicity and long term toxicity which are not traditionally considered in drug only studies, (4) perceived impracticality of traditional clinical trial regulatory endpoints (such as OS and PFS) when testing novel drug-radiotherapy combinations particularly in the curative setting and finally (5) historically limited collaboration among medical and radiation oncologists particularly in the United states which is crucial for aligning research perspectives and goals. Moving forward, overcoming these hurdles and prioritizing communication among key stakeholders in the field will be crucial to propel the radiosensitizer pipeline. The year 2020 was arguably a landmark year for drug-radiotherapy combinations, with two novel radiosensitizers getting fast track and breakthrough designations:

REFERENCES

- Bryant AK, Banegas MP, Martinez ME, Mell LK, Murphy JD. Trends in Radiation Therapy Among Cancer Survivors in the United States, 2000– 2030. *Cancer Epidemiol Prev Biomarkers* (2017) 26(6):963–70. doi: 10.1158/ 1055-9965.EPI-16-1023
- Dupin C, Lang P, Dessard-Diana B, Simon J-M, Cuenca X, Mazeron J-J, et al. Treatment of Head and Neck Paragangliomas With External Beam Radiation Therapy. *Int J Radiat Oncol Biol Phys* (2014) 89(2):353–9. doi: 10.1016/j.ijrobp.2014.02.010
- Mondal D, Sharma DN. External Beam Radiation Techniques for Breast Cancer in the New Millennium: New Challenging Perspectives. J Egyptian Natl Cancer Institute (2016) 28(4):211–8. doi: 10.1016/j.jnci.2016.08.001

NBTXR3 and Debio 1143 respectively (294). However, the field is yet to witness new market approvals as we strive to overcome challenges and improve patient outcomes.

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ME and NC contributed equally to the writing and planning of the content of the review. RR provided clinical input and expertise to the review and MF edited and directed the writing and content within the review. All authors contributed to the article and approved the submitted version.

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- Higgins KA, O'Connell K, Liu Y, Gillespie TW, McDonald MW, Pillai RN, et al. National Cancer Database Analysis of Proton *Versus* Photon Radiation Therapy in Non-Small Cell Lung Cancer. *Int J Radiat Oncol Biol Phys* (2017) 97(1):128–37. doi: 10.1016/j.ijrobp.2016.10.001
- Kim J-Y, Park Y. Treatment of Retinoblastoma: The Role of External Beam Radiotherapy. *Yonsei Med J* (2015) 56(6):1478. doi: 10.3349/ ymj.2015.56.6.1478
- Magnuson WJ, Halligan JB. Successful Treatment of Melanoma Metastatic to the Left Atrium Using External Beam Radiation Therapy. *Oncology* (2010) 24(7):650.
- Wang T-H, Huang P-I, Hu Y-W, Lin K-H, Liu C-S, Lin Y-Y, et al. Combined Yttrium-90 Microsphere Selective Internal Radiation Therapy and External Beam Radiotherapy in Patients With Hepatocellular

Carcinoma: From Clinical Aspects to Dosimetry. *PloS One* (2018) 13(1): e0190098. doi: 10.1371/journal.pone.0190098

- Yang J, Cai H, Xiao ZX, Wang H, Yang P. Effect of Radiotherapy on the Survival of Cervical Cancer Patients: An Analysis Based on SEER Database. *Medicine* (2019) 98(30):e16421. doi: 10.1097/MD.000000000016421
- Glaser SM, Beriwal S. Brachytherapy for Malignancies of the Vagina in the 3D Era. J Contemp Brachytherapy (2015) 7(4):312. doi: 10.5114/ jcb.2015.54053
- Falk AT, Claren A, Benezery K, François E, Gautier M, Gerard J-P, et al. Interstitial High-Dose Rate Brachytherapy as Boost for Anal Canal Cancer. *Radiat Oncol* (2014) 9(1):1–9. doi: 10.1186/s13014-014-0240-4
- Zaorsky NG, Davis BJ, Nguyen PL, Showalter TN, Hoskin PJ, Yoshioka Y, et al. The Evolution of Brachytherapy for Prostate Cancer. *Nat Rev Urol* (2017) 14(7):415. doi: 10.1038/nrurol.2017.76
- Vaidya JS, Bulsara M, Baum M, Wenz F, Massarut S, Pigorsch S, et al. New Clinical and Biological Insights From the International TARGIT-A Randomised Trial of Targeted Intraoperative Radiotherapy During Lumpectomy for Breast Cancer. Br J Cancer (2021) 125(3):380–9. doi: 10.14324/000.wp.10121050
- Cifarelli CP, Brehmer S, Vargo JA, Hack JD, Kahl KH, Sarria-Vargas G, et al. Intraoperative Radiotherapy (IORT) for Surgically Resected Brain Metastases: Outcome Analysis of an International Cooperative Study. *J Neuro-Oncology* (2019) 145(2):391–7. doi: 10.1007/s11060-019-03309-6
- 14. Withers HR. The Four R's of Radiotherapy. Adv Radiat Biol 5: Elsevier; (1975) 241-71. doi: 10.1016/B978-0-12-035405-4.50012-8
- Steel GG, McMillan TJ, Peacock J. The 5Rs of Radiobiology. Int J Radiat Biol (1989) 56(6):1045–8. doi: 10.1080/09553008914552491
- Joiner MC, Burmeister JW, Dörr W. Linear Energy Transfer and Relative Biological Effectiveness. In: *Basic Clinical Radiobiology*. CRC Press (2018). p. 54–60. doi: 10.1201/9780429490606-6
- Herskind C, Ma L, Liu Q, Zhang B, Schneider F, Veldwijk MR, et al. Biology of High Single Doses of IORT: RBE, 5 R's, and Other Biological Aspects. *Radiat Oncol* (2017) 12(1):1–14. doi: 10.1186/s13014-016-0750-3
- Sologuren I, Rodríguez-Gallego C, Lara PC. Immune Effects of High Dose Radiation Treatment: Implications of Ionizing Radiation on the Development of Bystander and Abscopal Effects. *Trans Cancer Res* (2014) 3:18–31. doi: 10.3390/ijms15010927
- Brown JM, Carlson DJ, Brenner DJ. The Tumor Radiobiology of SRS and SBRT: Are More Than the 5 Rs Involved? *Int J Radiat Oncol Biol Phys* (2014) 88(2):254–62. doi: 10.1016/j.ijrobp.2013.07.022
- Marconi R, Strolin S, Bossi G, Strigari L. A Meta-Analysis of the Abscopal Effect in Preclinical Models: Is the Biologically Effective Dose a Relevant Physical Trigger? *PloS One* (2017) 12(2):e0171559. doi: 10.1371/ journal.pone.0171559
- Riballo E, Kühne M, Rief N, Doherty A, Smith GC, Recio M-J, et al. A Pathway of Double-Strand Break Rejoining Dependent Upon ATM, Artemis, and Proteins Locating to γ-H2AX Foci. *Mol Cell* (2004) 16 (5):715–24. doi: 10.1016/j.molcel.2004.10.029
- Eccles LJ, O'Neill P, Lomax ME. Delayed Repair of Radiation Induced Clustered DNA Damage: Friend or Foe? *Mutat Research/Fundamental Mol Mech Mutagenesis* (2011) 711(1-2):134–41. doi: 10.1016/j.mrfmmm. 2010.11.003
- Gulston M, Fulford J, Jenner T, de Lara C, O'Neill P. Clustered DNA Damage Induced by γ Radiation in Human Fibroblasts (HF19), Hamster (V79-4) Cells and Plasmid DNA Is Revealed as Fpg and Nth Sensitive Sites. *Nucleic Acids Res* (2002) 30(15):3464–72. doi: 10.1093/nar/gkf467
- Asaithamby A, Chen DJ. Mechanism of Cluster DNA Damage Repair in Response to High-Atomic Number and Energy Particles Radiation. *Mutat Research/Fundamental Mol Mech Mutagenesis* (2011) 711(1-2):87–99. doi: 10.1016/j.mrfmmm.2010.11.002
- Nickoloff JA, Sharma N, Taylor L. Clustered DNA Double-Strand Breaks: Biological Effects and Relevance to Cancer Radiotherapy. *Genes* (2020) 11 (1):99. doi: 10.3390/genes11010099
- Hanai M, Yazu K, Hieda R. On the Experimental Distinction Between Ssbs and Dsbs in Circular DNA. *Int J Radiat Biol* (1998) 73(5):475–9. doi: 10.1080/095530098142013
- 27. Van Der Schans G. Gamma-Ray Induced Double-Strand Breaks in DNA Resulting From Randomly-Inflicted Single-Strand Breaks: Temporal Local

Denaturation, a New Radiation Phenomenon? Int J Radiat Biol Related Stud Physics Chem Med (1978) 33(2):105–20. doi: 10.1080/09553007814550011

- Zheng Y, Sanche L. Clustered DNA Damages Induced by 0.5 to 30 eV Electrons. Int J Mol Sci (2019) 20(15):3749.
- Sutherland BM, Bennett PV, Cintron NS, Guida P, Laval J. Low Levels of Endogenous Oxidative Damage Cluster Levels in Unirradiated Viral and Human DNAs. Free Radical Biol Med (2003) 35(5):495–503. doi: 10.1016/ S0891-5849(03)00327-7
- Bennett PV, Cintron NS, Gros L, Laval J, Sutherland BM. Are Endogenous Clustered DNA Damages Induced in Human Cells? *Free Radical Biol Med* (2004) 37(4):488–99. doi: 10.1016/j.freeradbiomed.2004.05.004
- Nikjoo H, O'Neill P, Terrissol M, Goodhead D. Quantitative Modelling of DNA Damage Using Monte Carlo Track Structure Method. *Radiat Environ biophysics* (1999) 38(1):31–8. doi: 10.1007/s004110050135
- Pavlopoulou A, Bagos PG, Koutsandrea V, Georgakilas AG. Molecular Determinants of Radiosensitivity in Normal and Tumor Tissue: A Bioinformatic Approach. *Cancer Lett* (2017) 403:37–47. doi: 10.1016/ j.canlet.2017.05.023
- Fishel ML, Jiang Y, Rajeshkumar N, Scandura G, Sinn AL, He Y, et al. Impact of APE1/Ref-1 Redox Inhibition on Pancreatic Tumor Growth. *Mol Cancer Ther* (2011) 10(9):1698–708. doi: 10.1158/1535-7163.MCT-11-0107
- 34. Gampala S, Shah F, Zhang C, Rhodes SD, Babb O, Grimard M, et al. Exploring Transcriptional Regulators Ref-1 and STAT3 as Therapeutic Targets in Malignant Peripheral Nerve Sheath Tumours. Br J Cancer (2021) 124(9):1566–80. doi: 10.1038/s41416-021-01270-8
- Robertson KA, Bullock HA, Xu Y, Tritt R, Zimmerman E, Ulbright TM, et al. Altered Expression of Ape1/ref-1 in Germ Cell Tumors and Overexpression in NT2 Cells Confers Resistance to Bleomycin and Radiation. *Cancer Res* (2001) 61(5):2220–5.
- Naidu MD, Mason JM, Pica RV, Fung H, Peña LA. Radiation Resistance in Glioma Cells Determined by DNA Damage Repair Activity of Ape1/Ref-1. *J Radiat Res* (2010) 51(4):393–404. doi: 10.1269/jrr.09077
- 37. Cun Y, Dai N, Xiong C, Li M, Sui J, Qian C, et al. Silencing of APE1 Enhances Sensitivity of Human Hepatocellular Carcinoma Cells to Radiotherapy *In Vitro* and in a Xenograft Model. *PloS One* (2013) 8(2): e55313. doi: 10.1371/journal.pone.0055313
- Zhao W, Hu H, Mo Q, Guan Y, Li Y, Du Y, et al. Function and Mechanism of Combined PARP-1 and BRCA Genes in Regulating the Radiosensitivity of Breast Cancer Cells. *Int J Clin Exp Pathol* (2019) 12(10):3915.
- 39. Teng P-N, Bateman NW, Darcy KM, Hamilton CA, Maxwell GL, Bakkenist CJ, et al. Pharmacologic Inhibition of ATR and ATM Offers Clinically Important Distinctions to Enhancing Platinum or Radiation Response in Ovarian, Endometrial, and Cervical Cancer Cells. *Gynecologic Oncol* (2015) 136(3):554–61. doi: 10.1016/j.ygyno.2014.12.035
- Metwally MAH, Frederiksen KD, Overgaard J. Compliance and Toxicity of the Hypoxic Radiosensitizer Nimorazole in the Treatment of Patients With Head and Neck Squamous Cell Carcinoma (HNSCC). *Acta Oncol* (2014) 53 (5):654–61. doi: 10.3109/0284186X.2013.864050
- Wardman P. Nitroimidazoles as Hypoxic Cell Radiosensitizers and Hypoxia Probes: Misonidazole, Myths and Mistakes. *Br J Radiol* (2018) 92 (1093):20170915. doi: 10.1259/bjr.20170915
- Somasundaram V, Basudhar D, Bharadwaj G, No JH, Ridnour LA, Cheng RY, et al. Molecular Mechanisms of Nitric Oxide in Cancer Progression, Signal Transduction, and Metabolism. *Antioxidants Redox Signaling* (2019) 30(8):1124–43. doi: 10.1089/ars.2018.7527
- 43. Fokas E, Prevo R, Pollard J, Reaper P, Charlton P, Cornelissen B, et al. Targeting ATR *In Vivo* Using the Novel Inhibitor VE-822 Results in Selective Sensitization of Pancreatic Tumors to Radiation. *Cell Death Dis* (2012) 3(12):e441–e. doi: 10.1038/cddis.2012.181
- 44. Riaz MA, Sak A, Erol YB, Groneberg M, Thomale J, Stuschke M. Metformin Enhances the Radiosensitizing Effect of Cisplatin in Non-Small Cell Lung Cancer Cell Lines With Different Cisplatin Sensitivities. *Sci Rep* (2019) 9 (1):1–16. doi: 10.1038/s41598-018-38004-5
- Wu C-H, Chen C-Y, Yeh C-T, Lin K-H. Radiosensitization of Hepatocellular Carcinoma Through Targeting Radio-Associated Microrna. Int J Mol Sci (2020) 21(5):1859. doi: 10.3390/ijms21051859
- 46. Masoudi-Khoram N, Abdolmaleki P, Hosseinkhan N, Nikoofar A, Mowla SJ, Monfared H, et al. Differential miRNAs Expression Pattern of Irradiated

Breast Cancer Cell Lines is Correlated With Radiation Sensitivity. *Sci Rep* (2020) 10(1):1–12. doi: 10.1038/s41598-020-65680-z

- Shimura T, Sasatani M, Kawai H, Kamiya K, Kobayashi J, Komatsu K, et al. A Comparison of Radiation-Induced Mitochondrial Damage Between Neural Progenitor Stem Cells and Differentiated Cells. *Cell Cycle* (2017) 16 (6):565–73. doi: 10.1080/15384101.2017.1284716
- Lei G, Zhang Y, Koppula P, Liu X, Zhang J, Lin SH, et al. The Role of Ferroptosis in Ionizing Radiation-Induced Cell Death and Tumor Suppression. *Cell Res* (2020) 30(2):146–62. doi: 10.1038/s41422-019-0263-3
- Corre I, Niaudet C, Paris F. Plasma Membrane Signaling Induced by Ionizing Radiation. *Mutat Research/Reviews Mutat Res* (2010) 704(1-3):61–7. doi: 10.1016/j.mrrev.2010.01.014
- 50. Zhang S-R, Zhang X-C, Liang J-F, Fang H-M, Huang H-X, Zhao Y-Y, et al. Chalcomoracin Inhibits Cell Proliferation and Increases Sensitivity to Radiotherapy in Human Non-Small Cell Lung Cancer Cells via Inducing Endoplasmic Reticulum Stress-Mediated Paraptosis. Acta Pharmacologica Sin (2020) 41(6):825–34. doi: 10.1038/s41401-019-0351-4
- 51. Qiao Q, Sun C, Han C, Han N, Zhang M, Li G. Endoplasmic Reticulum Stress Pathway PERK-eIF 2α Confers Radioresistance in Oropharyngeal Carcinoma by Activating NF-κb. *Cancer Sci* (2017) 108(7):1421–31. doi: 10.1111/cas.13260
- 52. Gong C, Yang Z, Zhang L, Wang Y, Gong W, Liu Y. Quercetin Suppresses DNA Double-Strand Break Repair and Enhances the Radiosensitivity of Human Ovarian Cancer Cells via P53-Dependent Endoplasmic Reticulum Stress Pathway. OncoTargets Ther (2018) 11:17. doi: 10.2147/OTT.S147316
- 53. Yasui H, Takeuchi R, Nagane M, Meike S, Nakamura Y, Yamamori T, et al. Radiosensitization of Tumor Cells Through Endoplasmic Reticulum Stress Induced by PEGylated Nanogel Containing Gold Nanoparticles. *Cancer Lett* (2014) 347(1):151–8. doi: 10.1016/j.canlet.2014.02.005
- Yang HJ, Youn H, Seong KM, Jin Y-W, Kim J, Youn B. Phosphorylation of Ribosomal Protein S3 and Antiapoptotic TRAF2 Protein Mediates Radioresistance in Non-Small Cell Lung Cancer Cells. J Biol Chem (2013) 288(5):2965–75. doi: 10.1074/jbc.M112.385989
- 55. Zhao Y, Tan M, Liu X, Xiong X, Sun Y. Inactivation of Ribosomal Protein S27-Like Confers Radiosensitivity *via* the Mdm2-P53 and Mdm2-MRN-ATM Axes. *Cell Death Dis* (2018) 9(2):1–11. doi: 10.1038/s41419-017-0192-3
- Hussain SS, Huang S-B, Bedolla RG, Rivas P, Basler JW, Swanson GP, et al. Suppression of Ribosomal Protein RPS6KB1 by Nexrutine Increases Sensitivity of Prostate Tumors to Radiation. *Cancer Lett* (2018) 433:232– 41. doi: 10.1016/j.canlet.2018.07.009
- Wu P-H, Onodera Y, Giaccia AJ, Le Q-T, Shimizu S, Shirato H, et al. Lysosomal Trafficking Mediated by Arl8b and BORC Promotes Invasion of Cancer Cells That Survive Radiation. *Commun Biol* (2020) 3(1):1–15. doi: 10.1038/s42003-020-01339-9
- Bagshaw RD, Callahan JW, Mahuran DJ. The Arf-Family Protein, Arl8b, is Involved in the Spatial Distribution of Lysosomes. *Biochem Biophys Res Commun* (2006) 344(4):1186–91. doi: 10.1016/j.bbrc.2006.03.221
- Cho YH, Lee S-H, Lee S-J, Kim HN, Koh J-Y. A Role of Metallothionein-3 in Radiation-Induced Autophagy in Glioma Cells. *Sci Rep* (2020) 10(1):1–11. doi: 10.1038/s41598-020-58237-7
- 60. Tang L, Wei F, Wu Y, He Y, Shi L, Xiong F, et al. Role of Metabolism in Cancer Cell Radioresistance and Radiosensitization Methods. J Exp Clin Cancer Res (2018) 37(1):1–15. doi: 10.1186/s13046-018-0758-7
- 61. Tann AW, Boldogh I, Meiss G, Qian W, Van Houten B, Mitra S, et al. Apoptosis Induced by Persistent Single-Strand Breaks in Mitochondrial Genome: Critical Role of EXOG (5'-EXO/endonuclease) in Their Repair. *J Biol Chem* (2011) 286(37):31975–83. doi: 10.1074/jbc.M110.215715
- 62. Li N, Yu L, Wang J, Gao X, Chen Y, Pan W, et al. A Mitochondria-Targeted Nanoradiosensitizer Activating Reactive Oxygen Species Burst for Enhanced Radiation Therapy. *Chem Sci* (2018) 9(12):3159–64. doi: 10.1039/ C7SC04458E
- 63. Liou G-Y, Storz P. Reactive Oxygen Species in Cancer. Free Radical Res (2010) 44(5):479–96. doi: 10.3109/10715761003667554
- Lin B, Gao F, Yang Y, Wu D, Zhang Y, Feng G, et al. FLASH Radiotherapy: History and Future. Front Oncol (2021) 11:1890. doi: 10.3389/ fonc.2021.644400
- 65. Ni K, Lan G, Veroneau SS, Duan X, Song Y, Lin W. Nanoscale Metal-Organic Frameworks for Mitochondria-Targeted Radiotherapy-

Radiodynamic Therapy. Nat Commun (2018) 9(1):1-13. doi: 10.1038/ s41467-018-06655-7

- 66. Yamazaki T, Kirchmair A, Sato A, Buqué A, Rybstein M, Petroni G, et al. Mitochondrial DNA Drives Abscopal Responses to Radiation That are Inhibited by Autophagy. *Nat Immunol* (2020) 21(10):1160–71. doi: 10.1038/s41590-020-0751-0
- Atkinson J, Kapralov AA, Yanamala N, Tyurina YY, Amoscato AA, Pearce L, et al. A Mitochondria-Targeted Inhibitor of Cytochrome C Peroxidase Mitigates Radiation-Induced Death. *Nat Commun* (2011) 2(1):1–9. doi: 10.1038/ncomms1499
- Momcilovic M, Shackelford D. Targeting LKB1 in Cancer–Exposing and Exploiting Vulnerabilities. Br J Cancer (2015) 113(4):574–84. doi: 10.1038/ bjc.2015.261
- 69. He Q, Li J, Dong F, Cai C, Zou X. LKB1 Promotes Radioresistance in Esophageal Cancer Cells Exposed to Radiation, by Suppression of Apoptosis and Activation of Autophagy *via* the AMPK Pathway. *Mol Med Rep* (2017) 16(2):2205–10. doi: 10.3892/mmr.2017.6852
- Porporato PE, Filigheddu N, Bravo-San Pedro JM, Kroemer G, Galluzzi L. Mitochondrial Metabolism and Cancer. *Cell Res* (2018) 28(3):265–80. doi: 10.1038/cr.2017.155
- 71. Lynam-Lennon N, Maher SG, Maguire A, Phelan J, Muldoon C, Reynolds JV, et al. Altered Mitochondrial Function and Energy Metabolism is Associated With a Radioresistant Phenotype in Oesophageal Adenocarcinoma. *PloS One* (2014) 9(6):e100738. doi: 10.1371/journal.pone.0100738
- Lawrence YR, Vikram B, Dignam JJ, Chakravarti A, Machtay M, Freidlin B, et al. NCI–RTOG Translational Program Strategic Guidelines for the Early-Stage Development of Radiosensitizers. J Natl Cancer Institute (2013) 105 (1):11–24. doi: 10.1093/jnci/djs472
- Ataman OU, Sambrook SJ, Wilks C, Lloyd A, Taylor AE, Wedge SR. The Clinical Development of Molecularly Targeted Agents in Combination With Radiation Therapy: A Pharmaceutical Perspective. Int J Radiat Oncol Biol Phys (2012) 84(4):e447–e54. doi: 10.1016/j.ijrobp.2012.05.019
- 74. Hanahan D, Weinberg RA. The Hallmarks of Cancer. *Cell* (2000) 100(1):57– 70. doi: 10.1016/S0092-8674(00)81683-9
- Hanahan D, Weinberg RA. Hallmarks of Cancer: The Next Generation. *cell* (2011) 144(5):646–74. doi: 10.1016/j.cell.2011.02.013
- Willers H, Keane FK, Kamran SC. Toward a New Framework for Clinical Radiation Biology. *Hematology/Oncology Clinics* (2019) 33(6):929–45. doi: 10.1016/j.hoc.2019.07.001
- Kamran SC, Mouw KW. Applying Precision Oncology Principles in Radiation Oncology. JCO Precis Oncol (2018) 2:1–23. doi: 10.1200/PO.18.00034
- Yard BD, Adams DJ, Chie EK, Tamayo P, Battaglia JS, Gopal P, et al. A Genetic Basis for the Variation in the Vulnerability of Cancer to DNA Damage. *Nat Commun* (2016) 7(1):1–14. doi: 10.1038/ncomms11428
- Mamounas EP, Tang G, Fisher B, Paik S, Shak S, Costantino JP, et al. Association Between the 21-Gene Recurrence Score Assay and Risk of Locoregional Recurrence in Node-Negative, Estrogen Receptor–Positive Breast Cancer: Results From NSABP B-14 and NSABP B-20. J Clin Oncol (2010) 28(10):1677. doi: 10.1200/JCO.2009.23.7610
- Solin LJ, Gray R, Baehner FL, Butler SM, Hughes LL, Yoshizawa C, et al. A Multigene Expression Assay to Predict Local Recurrence Risk for Ductal Carcinoma *in Situ* of the Breast. *J Natl Cancer Institute* (2013) 105(10):701– 10. doi: 10.1093/jnci/djt067
- Solin LJ, Gray R, Hughes LL, Wood WC, Lowen MA, Badve SS, et al. Surgical Excision Without Radiation for Ductal Carcinoma *in Situ* of the Breast: 12-Year Results From the ECOG-ACRIN E5194 Study. *J Clin Oncol* (2015) 33 (33):3938. doi: 10.1200/JCO.2015.60.8588
- Rakovitch E, Baehner R, Shak S, Miller D, Cherbavaz D, Anderson J, et al. A Large Prospectively Designed Study of the DCIS Score: Recurrence Risk After Local Excision for Ductal Carcinoma *in Situ* Patients With and Without Irradiation. *Int J Radiat Oncology*• *Biology*• *Phys* (2015) 93(3): S135–S6. doi: 10.1016/j.ijrobp.2015.07.323
- Rakovitch E, Sutradhar R, Nofech-Mozes S, Gu S, Fong C, Hanna W, et al. 21-Gene Assay and Breast Cancer Mortality in Ductal Carcinoma *in Situ.* JNCI: J Natl Cancer Institute (2021) 113(5):572–9. doi: 10.1093/jnci/djaa179
- 84. Den RB, Yousefi K, Trabulsi EJ, Abdollah F, Choeurng V, Feng FY, et al. Genomic Classifier Identifies Men With Adverse Pathology After Radical

Prostatectomy Who Benefit From Adjuvant Radiation Therapy. J Clin Oncol (2015) 33(8):944. doi: 10.1200/JCO.2014.59.0026

- 85. Gore JL, du Plessis M, Santiago-Jiménez M, Yousefi K, Thompson DJ, Karsh L, et al. Decipher Test Impacts Decision Making Among Patients Considering Adjuvant and Salvage Treatment After Radical Prostatectomy: Interim Results From the Multicenter Prospective PRO-IMPACT Study. *Cancer* (2017) 123(15):2850–9. doi: 10.1002/cncr.30665
- Eggener SE, Rumble RB, Armstrong AJ, Morgan TM, Crispino T, Cornford P, et al. Molecular Biomarkers in Localized Prostate Cancer: ASCO Guideline. J Clin Oncol (2020) 38(13):1474–94. doi: 10.1200/JCO.19.02768
- Vale CL, Fisher D, Kneebone A, Parker C, Pearse M, Richaud P, et al. Adjuvant or Early Salvage Radiotherapy for the Treatment of Localised and Locally Advanced Prostate Cancer: A Prospectively Planned Systematic Review and Meta-Analysis of Aggregate Data. *Lancet* (2020) 396 (10260):1422-31. doi: 10.1016/S0140-6736(20)31952-8
- Morgan TM, Miller DC, Dunn R, Susan L, Okoth L, Johnson A, et al. Prospective Randomized Trial of Genomic Classifier Impact on Treatment Decisions in Patients at High Risk of Recurrence Following Radical Prostatectomy (G-MINOR). Am Soc Clin Oncol (2018). doi: 10.1200/ JCO.2018.36.6_suppl.TPS154
- Scott JG, Berglund A, Schell MJ, Mihaylov I, Fulp WJ, Yue B, et al. A Genome-Based Model for Adjusting Radiotherapy Dose (GARD): A Retrospective, Cohort-Based Study. *Lancet Oncol* (2017) 18(2):202–11. doi: 10.1016/S1470-2045(16)30648-9
- Scott JG, Sedor G, Ellsworth P, Scarborough JA, Ahmed K, Eschrich SA, et al. Pan-Cancer Prediction of Radiotherapy Benefit using Genomic-Adjusted Radiation Dose (GARD): A Cohort-based Pooled Analysis. *Lancet Oncol* (2021) 22(9):1221–9. doi: 10.1016/S1470-2045(21)00347-8
- Ahmed KA, Liveringhouse CL, Mills MN, Figura NB, Grass GD, Washington IR, et al. Utilizing the Genomically Adjusted Radiation Dose (GARD) to Personalize Adjuvant Radiotherapy in Triple Negative Breast Cancer Management. *EBioMedicine* (2019) 47:163–9. doi: 10.1016/ j.ebiom.2019.08.019
- Ahmed KA, Scott JG, Arrington JA, Naghavi AO, Grass GD, Perez BA, et al. Radiosensitivity of Lung Metastases by Primary Histology and Implications for Stereotactic Body Radiation Therapy Using the Genomically Adjusted Radiation Dose. J Thorac Oncol (2018) 13(8):1121–7. doi: 10.1016/ j.jtho.2018.04.027
- 93. Rieckmann T, Tribius S, Grob TJ, Meyer F, Busch C-J, Petersen C, et al. HNSCC Cell Lines Positive for HPV and P16 Possess Higher Cellular Radiosensitivity Due to an Impaired DSB Repair Capacity. *Radiotherapy* Oncol (2013) 107(2):242–6. doi: 10.1016/j.radonc.2013.03.013
- Kimple RJ, Smith MA, Blitzer GC, Torres AD, Martin JA, Yang RZ, et al. Enhanced Radiation Sensitivity in HPV-Positive Head and Neck Cancer. *Cancer Res* (2013) 73(15):4791–800. doi: 10.1158/0008-5472.CAN-13-0587
- 95. Tinhofer I, Stenzinger A, Eder T, Konschak R, Niehr F, Endris V, et al. Targeted Next-Generation Sequencing Identifies Molecular Subgroups in Squamous Cell Carcinoma of the Head and Neck With Distinct Outcome After Concurrent Chemoradiation. Ann Oncol (2016) 27(12):2262–8. doi: 10.1093/annonc/mdw426
- Northcott PA, Korshunov A, Witt H, Hielscher T, Eberhart CG, Mack S, et al. Medulloblastoma Comprises Four Distinct Molecular Variants. J Clin Oncol (2011) 29(11):1408. doi: 10.1200/JCO.2009.27.4324
- Michalski JM, Janss AJ, Vezina LG, Smith KS, Billups CA, Burger PC, et al. Children's Oncology Group Phase III Trial of Reduced-Dose and Reduced-Volume Radiotherapy With Chemotherapy for Newly Diagnosed Average-Risk Medulloblastoma. J Clin Oncol (2021) 39(24):2685–97. doi: 10.1200/ JCO.20.02730
- Kerns SL, Ostrer H, Rosenstein BS. Radiogenomics: Using Genetics to Identify Cancer Patients at Risk for Development of Adverse Effects Following Radiotherapy. *Cancer Discov* (2014) 4(2):155–65. doi: 10.1158/ 2159-8290.CD-13-0197
- 99. Kerns SL, Stock R, Stone N, Buckstein M, Shao Y, Campbell C, et al. A 2-Stage Genome-Wide Association Study to Identify Single Nucleotide Polymorphisms Associated With Development of Erectile Dysfunction Following Radiation Therapy for Prostate Cancer. Int J Radiat Oncol Biol Phys (2013) 85(1):e21–e8. doi: 10.1016/j.ijrobp.2012.08.003

- 100. Kerns SL, Stone NN, Stock RG, Rath L, Ostrer H, Rosenstein BS. A 2-Stage Genome-Wide Association Study to Identify Single Nucleotide Polymorphisms Associated With Development of Urinary Symptoms After Radiotherapy for Prostate Cancer. J Urol (2013) 190(1):102–8. doi: 10.1016/j.juro.2013.01.096
- 101. Barnett GC, Thompson D, Fachal L, Kerns S, Talbot C, Elliott RM, et al. A Genome Wide Association Study (GWAS) Providing Evidence of an Association Between Common Genetic Variants and Late Radiotherapy Toxicity. *Radiotherapy Oncol* (2014) 111(2):178–85. doi: 10.1016/j.radonc.2014.02.012
- 102. West C, Azria D, Chang-Claude J, Davidson S, Lambin P, Rosenstein B, et al. The REQUITE Project: Validating Predictive Models and Biomarkers of Radiotherapy Toxicity to Reduce Side-Effects and Improve Quality of Life in Cancer Survivors. *Clin Oncol* (2014) 26(12):739–42. doi: 10.1016/ j.clon.2014.09.008
- 103. Kerns SL, Fachal L, Dorling L, Barnett GC, Baran A, Peterson DR, et al. Radiogenomics Consortium Genome-Wide Association Study Meta-Analysis of Late Toxicity After Prostate Cancer Radiotherapy. JNCI: J Natl Cancer Institute (2020) 112(2):179–90. doi: 10.1093/jnci/djz075
- 104. Ahmad SS, Crittenden MR, Tran PT, Kluetz PG, Blumenthal GM, Bulbeck H, et al. Clinical Development of Novel Drug–Radiotherapy Combinations. *Clin Cancer Res* (2019) 25(5):1455–61. doi: 10.1158/1078-0432.CCR-18-2466
- 105. Sharma RA, Plummer R, Stock JK, Greenhalgh TA, Ataman O, Kelly S, et al. Clinical Development of New Drug–Radiotherapy Combinations. *Nat Rev Clin Oncol* (2016) 13(10):627. doi: 10.1038/nrclinonc.2016.79
- 106. Hall WA, Bergom C, Thompson RF, Baschnagel AM, Vijayakumar S, Willers H, et al. Precision Oncology and Genomically Guided Radiation Therapy: A Report From the American Society for Radiation Oncology/American Association of Physicists in Medicine/National Cancer Institute Precision Medicine Conference. *Int J Radiat Oncol Biol Phys* (2018) 101(2):274–84. doi: 10.1016/j.ijrobp.2017.05.044
- 107. Pilié PG, Tang C, Mills GB, Yap TA. State-Of-the-Art Strategies for Targeting the DNA Damage Response in Cancer. Nat Rev Clin Oncol (2019) 16(2):81–104. doi: 10.1038/s41571-018-0114-z
- 108. Samstein RM, Riaz N. The DNA Damage Response in Immunotherapy and Radiation. Adv Radiat Oncol (2018) 3(4):527–33. doi: 10.1016/ j.adro.2018.08.017
- 109. Topatana W, Juengpanich S, Li S, Cao J, Hu J, Lee J, et al. Advances in Synthetic Lethality for Cancer Therapy: Cellular Mechanism and Clinical Translation. J Hematol Oncol (2020) 13(1):1–22. doi: 10.1186/s13045-020-00956-5
- 110. Morgan MA, Lawrence TS. Molecular Pathways: Overcoming Radiation Resistance by Targeting DNA Damage Response Pathways. *Clin Cancer Res* (2015) 21(13):2898–904. doi: 10.1158/1078-0432.CCR-13-3229
- 111. Wéra A-C, Lobbens A, Stoyanov M, Lucas S, Michiels C. Radiation-Induced Synthetic Lethality: Combination of Poly (ADP-Ribose) Polymerase and RAD51 Inhibitors to Sensitize Cells to Proton Irradiation. *Cell Cycle* (2019) 18(15):1770–83. doi: 10.1080/15384101.2019.1632640
- 112. Lesueur P, Chevalier F, Austry J-B, Waissi W, Burckel H, Noël G, et al. Poly-(ADP-Ribose)-Polymerase Inhibitors as Radiosensitizers: A Systematic Review of Pre-Clinical and Clinical Human Studies. *Oncotarget* (2017) 8 (40):69105. doi: 10.18632/oncotarget.19079
- 113. Karnak D, Engelke CG, Parsels LA, Kausar T, Wei D, Robertson JR, et al. Combined Inhibition of Wee1 and PARP1/2 for Radiosensitization in Pancreatic Cancer. *Clin Cancer Res* (2014) 20(19):5085–96. doi: 10.1158/ 1078-0432.CCR-14-1038
- 114. Kausar T, Schreiber JS, Karnak D, Parsels LA, Parsels JD, Davis MA, et al. Sensitization of Pancreatic Cancers to Gemcitabine Chemoradiation by WEE1 Kinase Inhibition Depends on Homologous Recombination Repair. *Neoplasia* (2015) 17(10):757–66. doi: 10.1016/j.neo.2015.09.006
- 115. Cuneo KC, Morgan MA, Davis MA, Parcels LA, Parcels J, Karnak D, et al. Weel Kinase Inhibitor AZD1775 Radiosensitizes Hepatocellular Carcinoma Regardless of TP53 Mutational Status Through Induction of Replication Stress. *Int J Radiat Oncol Biol Phys* (2016) 95(2):782–90. doi: 10.1016/ j.ijrobp.2016.01.028
- 116. Al Zubaidi T, Gehrisch OF, Genois M-M, Liu Q, Lu S, Kung J, et al. Targeting the DNA Replication Stress Phenotype of KRAS Mutant Cancer Cells. *Sci Rep* (2021) 11(1):1–12. doi: 10.1038/s41598-021-83142-y

- 117. Ubhi T, Brown GW. Exploiting DNA Replication Stress for Cancer Treatment. Cancer Res (2019) 79(8):1730–9. doi: 10.1158/0008-5472.CAN-18-3631
- 118. Parsels LA, Karnak D, Parsels JD, Zhang Q, Vélez-Padilla J, Reichert ZR, et al. PARP1 Trapping and DNA Replication Stress Enhance Radiosensitization With Combined WEE1 and PARP Inhibitors. *Mol Cancer Res* (2018) 16(2):222–32. doi: 10.1158/1541-7786.MCR-17-0455
- Bentzen SM, Harari PM, Bernier J. Exploitable Mechanisms for Combining Drugs With Radiation: Concepts, Achievements and Future Directions. *Nat Clin Pract Oncol* (2007) 4(3):172–80. doi: 10.1038/ncponc0744
- 120. Jagodinsky JC, Harari PM, Morris ZS. The Promise of Combining Radiation Therapy With Immunotherapy. *Int J Radiat Oncol Biol Phys* (2020) 108 (1):6–16. doi: 10.1016/j.ijrobp.2020.04.023
- 121. Golden EB, Frances D, Pellicciotta I, Demaria S, Helen Barcellos-Hoff M, Formenti SC. Radiation Fosters Dose-Dependent and Chemotherapy-Induced Immunogenic Cell Death. *Oncoimmunology* (2014) 3(4):e28518. doi: 10.4161/onci.28518
- 122. Ngwa W, Irabor OC, Schoenfeld JD, Hesser J, Demaria S, Formenti SC. Using Immunotherapy to Boost the Abscopal Effect. *Nat Rev Cancer* (2018) 18(5):313–22. doi: 10.1038/nrc.2018.6
- 123. Twyman-Saint Victor C, Rech AJ, Maity A, Rengan R, Pauken KE, Stelekati E, et al. Radiation and Dual Checkpoint Blockade Activate Non-Redundant Immune Mechanisms in Cancer. *Nature* (2015) 520(7547):373–7. doi: 10.1038/nature14292
- 124. Stamell EF, Wolchok JD, Gnjatic S, Lee NY, Brownell I. The Abscopal Effect Associated With a Systemic Anti-Melanoma Immune Response. *Int J Radiat* Oncol Biol Phys (2013) 85(2):293–5. doi: 10.1016/j.ijrobp.2012.03.017
- 125. Postow MA, Callahan MK, Barker CA, Yamada Y, Yuan J, Kitano S, et al. Immunologic Correlates of the Abscopal Effect in a Patient With Melanoma. *New Engl J Med* (2012) 366(10):925–31. doi: 10.1056/NEJMoa1112824
- 126. Golden EB, Demaria S, Schiff PB, Chachoua A, Formenti SC. An Abscopal Response to Radiation and Ipilimumab in a Patient With Metastatic Non– Small Cell Lung Cancer. *Cancer Immunol Res* (2013) 1(6):365–72. doi: 10.1158/2326-6066.CIR-13-0115
- 127. Golden EB, Chhabra A, Chachoua A, Adams S, Donach M, Fenton-Kerimian M, et al. Local Radiotherapy and Granulocyte-Macrophage Colony-Stimulating Factor to Generate Abscopal Responses in Patients With Metastatic Solid Tumours: A Proof-of-Principle Trial. *Lancet Oncol* (2015) 16(7):795–803. doi: 10.1016/S1470-2045(15)00054-6
- 128. Merrick A, Errington F, Milward K, O'Donnell D, Harrington K, Bateman A, et al. Immunosuppressive Effects of Radiation on Human Dendritic Cells: Reduced IL-12 Production on Activation and Impairment of Naive T-Cell Priming. *Br J Cancer* (2005) 92(8):1450–8. doi: 10.1038/sj.bjc.6602518
- 129. Chiang C-S, Fu S-Y, Wang S-C, Yu C-F, Chen F-H, Lin C-M, et al. Irradiation Promotes an M2 Macrophage Phenotype in Tumor Hypoxia. *Front Oncol* (2012) 2:89. doi: 10.3389/fonc.2012.00089
- 130. Quon H, McNutt T, Lee J, Bowers M, Jiang W, Lakshminarayanan P, et al. Needs and Challenges for Radiation Oncology in the Era of Precision Medicine. Int J Radiat Oncol Biol Phys (2019) 103(4):809–17. doi: 10.1016/ j.ijrobp.2018.11.017
- 131. Baumann M, Krause M, Overgaard J, Debus J, Bentzen SM, Daartz J, et al. Radiation Oncology in the Era of Precision Medicine. *Nat Rev Cancer* (2016) 16(4):234. doi: 10.1038/nrc.2016.18
- 132. Caston RA, Shah F, Starcher CL, Wireman R, Babb O, Grimard M, et al. Combined Inhibition of Ref-1 and STAT3 Leads to Synergistic Tumour Inhibition in Multiple Cancers Using 3D and *In Vivo* Tumour Co-Culture Models. *J Cell Mol Med* (2020) 25(2):784–800. doi: 10.1111/jcmm.16132
- 133. Fishel ML, Xia H, McGeown J, McIlwain DW, Elbanna M, Craft AA, et al. Antitumor Activity and Mechanistic Characterization of APE1/Ref-1 Inhibitors in Bladder Cancer. *Mol Cancer Ther* (2019) 18(11):1947–60. doi: 10.1158/1535-7163.MCT-18-1166
- 134. Prabhu L, Wei H, Chen L, Demir O, Sandusky G, Sun E, et al. Adapting AlphaLISA High Throughput Screen to Discover a Novel Small-Molecule Inhibitor Targeting Protein Arginine Methyltransferase 5 in Pancreatic and Colorectal Cancers. *Oncotarget* (2017) 8(25):39963–77. doi: 10.18632/ oncotarget.18102
- 135. Logsdon DP, Shah F, Carta F, Supuran CT, Kamocka M, Jacobsen MH, et al. Blocking HIF Signaling via Novel Inhibitors of CA9 and APE1/Ref-1

Dramatically Affects Pancreatic Cancer Cell Survival. Sci Rep (2018) 8 (1):13759. doi: 10.1038/s41598-018-32034-9

- 136. Logsdon DP, Grimard M, Luo M, Shahda S, Jiang Y, Tong Y, et al. Regulation of HIF1alpha Under Hypoxia by APE1/Ref-1 Impacts CA9 Expression: Dual Targeting in Patient-Derived 3d Pancreatic Cancer Models. *Mol Cancer Ther* (2016) 15(11):2722–32. doi: 10.1158/1535-7163.MCT-16-0253
- 137. Selby M, Delosh R, Laudeman J, Ogle C, Reinhart R, Silvers T, et al. 3d Models of the NCI60 Cell Lines for Screening Oncology Compounds. SLAS Discovery (2017) 22(5):473–83. doi: 10.1177/2472555217697434
- 138. Chen O, Michlíková S, Eckhardt L, Wondrak M, De Mendoza AM, Krause M, et al. Efficient Heat Shock Response Affects Hyperthermia-Induced Radiosensitization in a Tumor Spheroid Control Probability Assay. *Cancers (Basel)* (2021) 13(13):3168. doi: 10.3390/cancers13133168
- 139. Jiang Y, Verbiest T, Devery AM, Bokobza SM, Weber AM, Leszczynska KB, et al. Hypoxia Potentiates the Radiation-Sensitizing Effect of Olaparib in Human Non-Small Cell Lung Cancer Xenografts by Contextual Synthetic Lethality. Int J Radiat Oncol Biol Phys (2016) 95(2):772–81. doi: 10.1016/ j.ijrobp.2016.01.035
- 140. Lafontaine J, Boisvert J-S, Glory A, Coulombe S, Wong P. Synergy Between Non-Thermal Plasma With Radiation Therapy and Olaparib in a Panel of Breast Cancer Cell Lines. *Cancers* (2020) 12(2):348. doi: 10.3390/cancers12020348
- 141. Saha S, Howarth R, Pappworth I, Marchbank K, Curtin N. Potential Use of the PARP Inhibitor Rucaparib to Enhance Cervical Cancer Treatment. *Eur J Cancer* (2020) 138:S37. doi: 10.1016/S0959-8049(20)31169-2
- 142. Chatterjee P, Choudhary GS, Sharma A, Singh K, Heston WD, Ciezki J, et al. PARP Inhibition Sensitizes to Low Dose-Rate Radiation TMPRSS2-ERG Fusion Gene-Expressing and PTEN-Deficient Prostate Cancer Cells. *PloS One* (2013) 8(4):e60408. doi: 10.1371/journal.pone.0060408
- 143. Nile DL, Rae C, Hyndman IJ, Gaze MN, Mairs RJ. An Evaluation In Vitro of PARP-1 Inhibitors, Rucaparib and Olaparib, as Radiosensitisers for the Treatment of Neuroblastoma. BMC Cancer (2016) 16(1):1–13. doi: 10.1186/s12885-016-2656-8
- 144. McDaniel NK, Iida M, Nickel KP, Longhurst CA, Fischbach SR, Rodems TS, et al. AXL Mediates Cetuximab and Radiation Resistance Through Tyrosine 821 and the C-ABL Kinase Pathway in Head and Neck Cancer. *Clin Cancer Res* (2020) 26(16):4349–59. doi: 10.1158/1078-0432.CCR-19-3142
- 145. Zeng L, Beggs RR, Cooper TS, Weaver AN, Yang ES. Combining Chk1/2 Inhibition With Cetuximab and Radiation Enhances *In Vitro* and *In Vivo* Cytotoxicity in Head and Neck Squamous Cell Carcinoma. *Mol Cancer Ther* (2017) 16(4):591–600. doi: 10.1158/1535-7163.MCT-16-0352
- 146. Boysen G, Jamshidi-Parsian A, Davis MA, Siegel ER, Simecka CM, Kore RA, et al. Glutaminase Inhibitor CB-839 Increases Radiation Sensitivity of Lung Tumor Cells and Human Lung Tumor Xenografts in Mice. *Int J Radiat Biol* (2019) 95(4):436–42. doi: 10.1080/09553002.2018.1558299
- 147. Wicker CA, Hunt BG, Krishnan S, Aziz K, Parajuli S, Palackdharry S, et al. Glutaminase Inhibition With Telaglenastat (CB-839) Improves Treatment Response in Combination With Ionizing Radiation in Head and Neck Squamous Cell Carcinoma Models. *Cancer Lett* (2021) 502:180–8. doi: 10.1016/j.canlet.2020.12.038
- 148. Xu P, Huang J-M, Ren Y, Zha X, Deng B-F, Wu J-H, et al. Regulation of Hypoxia-Induced mRNA Expressions of HIF-1alpha and Osteopontin and *In Vitro* Radiosensitization by Tirapazamine in Human Nasopharyngeal Carcinoma HNE-1 and CNE-1 Cells. *Chin J Cancer* (2010) 29(2):126–30. doi: 10.5732/cjc.009.10500
- 149. Liu Y, Liu Y, Bu W, Xiao Q, Sun Y, Zhao K, et al. Radiation-/Hypoxia-Induced Solid Tumor Metastasis and Regrowth Inhibited by Hypoxia-Specific Upconversion Nanoradiosensitizer. *Biomaterials* (2015) 49:1–8. doi: 10.1016/j.biomaterials.2015.01.028
- 150. Chen Y, Li W-W, Peng P, Zhao W-H, Tian Y-J, Huang Y, et al. Mtorc1 Inhibitor RAD001 (Everolimus) Enhances Non-Small Cell Lung Cancer Cell Radiosensitivity *In Vitro via* Suppressing Epithelial–Mesenchymal Transition. *Acta Pharmacologica Sin* (2019) 40(8):1085–94. doi: 10.1038/ s41401-019-0215-y
- 151. Chaumeil MM, Ozawa T, Park I, Scott K, James CD, Nelson SJ, et al. Hyperpolarized 13c MR Spectroscopic Imaging can be Used to Monitor Everolimus Treatment *In Vivo* in an Orthotopic Rodent Model of Glioblastoma. *Neuroimage* (2012) 59(1):193–201. doi: 10.1016/ j.neuroimage.2011.07.034

- 152. Sørensen BS, Busk M, Olthof N, Speel E-J, Horsman MR, Alsner J, et al. Radiosensitivity and Effect of Hypoxia in HPV Positive Head and Neck Cancer Cells. *Radiotherapy Oncol* (2013) 108(3):500–5. doi: 10.1016/ j.radonc.2013.06.011
- 153. Tao Z, Le Blanc JM, Wang C, Zhan T, Zhuang H, Wang P, et al. Coadministration of Trametinib and Palbociclib Radiosensitizes KRAS-Mutant Non–Small Cell Lung Cancers In Vitro and In Vivo. Clin Cancer Res (2016) 22(1):122–33. doi: 10.1158/1078-0432.CCR-15-0589
- 154. Schick U, Kyula J, Barker H, Patel R, Zaidi S, Gregory C, et al. Trametinib Radiosensitises RAS-And BRAF-Mutated Melanoma by Perturbing Cell Cycle and Inducing Senescence. *Radiotherapy Oncol* (2015) 117(2):364–75. doi: 10.1016/j.radonc.2015.06.026
- 155. Yang L, Shen C, Pettit CJ, Li T, Hu AJ, Miller ED, et al. Weel Kinase Inhibitor AZD1775 Effectively Sensitizes Esophageal Cancer to Radiotherapy. *Clin Cancer Res* (2020) 26(14):3740–50. doi: 10.1158/1078-0432.CCR-19-3373
- 156. Haines E, Nishida Y, Carr MI, Montoya RH, Ostermann LB, Zhang W, et al. DNA-PK Inhibitor Peposertib Enhances P53-Dependent Cytotoxicity of DNA Double-Strand Break Inducing Therapy in Acute Leukemia. *Sci Rep* (2021) 11(1):1–14. doi: 10.1038/s41598-021-90500-3
- 157. Zenke FT, Zimmermann A, Sirrenberg C, Dahmen H, Kirkin V, Pehl U, et al. Pharmacologic Inhibitor of DNA-PK, M3814, Potentiates Radiotherapy and Regresses Human Tumors in Mouse Models. *Mol Cancer Ther* (2020) 19 (5):1091–101. doi: 10.1158/1535-7163.MCT-19-0734
- 158. Zhao J, Liu P, Ma J, Li D, Yang H, Chen W, et al. Enhancement of Radiosensitization by Silver Nanoparticles Functionalized With Polyethylene Glycol and Aptamer As1411 for Glioma Irradiation Therapy. *Int J nanomedicine* (2019) 14:9483. doi: 10.2147/IJN.S224160
- 159. Habiba K, Aziz K, Sanders K, Santiago CM, Mahadevan LSK, Makarov V, et al. Enhancing Colorectal Cancer Radiation Therapy Efficacy Using Silver Nanoprisms Decorated With Graphene as Radiosensitizers. *Sci Rep* (2019) 9 (1):1–9. doi: 10.1038/s41598-019-53706-0
- 160. Cala PG, Chambrier I, Cook MJ, Haines AH, Field RA, Russell DA. Targeted Photodynamic Therapy of Breast Cancer Cells Using Lactose-Phthalocyanine Functionalized Gold Nanoparticles. J colloid Interface Sci (2018) 512:249–59. doi: 10.1016/j.jcis.2017.10.030
- 161. Rajaee A, Wang S, Zhao L, Wang D, Liu Y, Wang J, et al. Multifunction Bismuth Gadolinium Oxide Nanoparticles as Radiosensitizer in Radiation Therapy and Imaging. *Phys Med Biol* (2019) 64(19):195007. doi: 10.1088/ 1361-6560/ab2154
- 162. Jonuscheit S, Jost T, Gajdošová F, Wrobel M, Hecht M, Fietkau R, et al. PARP Inhibitors Talazoparib and Niraparib Sensitize Melanoma Cells to Ionizing Radiation. *Genes* (2021) 12(6):849. doi: 10.3390/genes12060849
- 163. Loap P, Loirat D, Berger F, Ricci F, Vincent-Salomon A, Ezzili C, et al. Combination of Olaparib and Radiation Therapy for Triple Negative Breast Cancer: Preliminary Results of the RADIOPARP Phase 1 Trial. *Int J Radiat* Oncol Biol Phys (2021) 109(2):436–40. doi: 10.1016/j.ijrobp.2020.09.032
- 164. Malfatti MC, Antoniali G, Codrich M, Burra S, Mangiapane G, Dalla E, et al. New Perspectives in Cancer Biology From a Study of Canonical and Non-Canonical Functions of Base Excision Repair Proteins With a Focus on Early Steps. *Mutagenesis* (2019) 35(1):129–49. doi: 10.1093/mutage/gez051
- 165. Shah F, Logsdon D, Messmann RA, Fehrenbacher JC, Fishel ML, Kelley MR. Exploiting the Ref-1-APE1 Node in Cancer Signaling and Other Diseases: From Bench to Clinic. NPJ Precis Oncol (2017) 1:19. doi: 10.1038/s41698-017-0023-0
- 166. Caston RA, Gampala S, Armstrong L, Messmann RA, Fishel ML, Kelley MR. The Multifunctional APE1 DNA Repair-Redox Signaling Protein as a Drug Target in Human Disease. *Drug Discov Today* (2021) 26(1):218–28. doi: 10.1016/j.drudis.2020.10.015
- 167. Parsons JL, Dianova II, Dianov GL. APE1 Is the Major 3'-Phosphoglycolate Activity in Human Cell Extracts. *Nucleic Acids Res* (2004) 32(12):3531–6. doi: 10.1093/nar/gkh676
- 168. Kievit FM, Wang K, Ozawa T, Tarudji AW, Silber JR, Holland EC, et al. Nanoparticle-Mediated Knockdown of DNA Repair Sensitizes Cells to Radiotherapy and Extends Survival in a Genetic Mouse Model of Glioblastoma. *Nanomedicine* (2017) 13(7):2131–9. doi: 10.1016/ j.nano.2017.06.004

- 169. Liu Z, Yan H, Li H. Silencing of DNA Repair Sensitizes Pediatric Brain Tumor Cells to Gamma-Irradiation Using Gold Nanoparticles. *Environ Toxicol Pharmacol* (2017) 53:40–5. doi: 10.1016/j.etap.2017.04.017
- 170. Gampala S, Shah F, Lu X, Moon H, Sandusky G, Hulsey E, et al. Ref-1 Redox Activity Alters Cancer Cell Metabolism in Pancreatic Cancer: Exploiting This Novel Finding as a Potential Target. J Exp Clin Can Res (2021) 40 (1):251. doi: 10.21203/rs.3.rs-125304/v1
- 171. Bignon E, Gattuso H, Morell C, Dehez F, Georgakilas AG, Monari A, et al. Correlation of Bistranded Clustered Abasic DNA Lesion Processing With Structural and Dynamic DNA Helix Distortion. *Nucleic Acids Res* (2016) 44 (18):8588–99. doi: 10.1093/nar/gkw773
- 172. Vascotto C, Fantini D, Romanello M, Cesaratto L, Deganuto M, Leonardi A, et al. APE1/Ref-1 Interacts With NPM1 Within Nucleoli and Plays a Role in the rRNA Quality Control Process. *Mol Cell Biol* (2009) 29(7):1834–54. doi: 10.1128/MCB.01337-08
- 173. Poletto M, Malfatti MC, Dorjsuren D, Scognamiglio PL, Marasco D, Vascotto C, et al. Inhibitors of the Apurinic/Apyrimidinic Endonuclease 1 (APE1)/nucleophosmin (NPM1) Interaction That Display Anti-Tumor Properties. *Mol Carcinog* (2016) 55(5):688–704. doi: 10.1002/mc.22313
- 174. Traver G, Sekhar KR, Crooks PA, Keeney DS, Freeman ML. Targeting NPM1 in Irradiated Cells Inhibits NPM1 Binding to RAD51, RAD51 Foci Formation and Radiosensitizes NSCLC. *Cancer Lett* (2021) 500:220–7. doi: 10.1016/j.canlet.2020.12.023
- 175. Kelley MR, Georgiadis MM, Fishel ML. APE1/Ref-1Role in Redox Signaling: Translational Applications of Targeting the Redox Function of the DNA Repair/Redox Protein APE1/Ref-1. Curr Mol Pharmacol (2012) 5(1):36–53. doi: 10.2174/1874467211205010036
- 176. Hawkes HJ, Karlenius TC, Tonissen KF. Regulation of the Human Thioredoxin Gene Promoter and Its Key Substrates: A Study of Functional and Putative Regulatory Elements. *Biochim Biophys Acta* (2014) 1840 (1):303–14. doi: 10.1016/j.bbagen.2013.09.013
- 177. Tell G, Damante G, Caldwell D, Kelley MR. The Intracellular Localization of APE1/Ref-1: More Than a Passive Phenomenon? *Antioxid Redox Signal* (2005) 7(3-4):367–84. doi: 10.1089/ars.2005.7.367
- 178. Kelley MR, Wikel JH, Guo C, Pollok KE, Bailey BJ, Wireman R, et al. Identification and Characterization of New Chemical Entities Targeting Apurinic/Apyrimidinic Endonuclease 1 for the Prevention of Chemotherapy-Induced Peripheral Neuropathy. J Pharmacol Exp Ther (2016) 359(2):300–9. doi: 10.1124/jpet.116.235283
- 179. Vasko MR, Guo C, Thompson EL, Kelley MR. The Repair Function of the Multifunctional DNA Repair/Redox Protein APE1 Is Neuroprotective After Ionizing Radiation. DNA Repair (Amst) (2011) 10(9):942–52. doi: 10.1016/ j.dnarep.2011.06.004
- 180. Galeaz C, Totis C, Bisio A. Radiation Resistance: A Matter of Transcription Factors. Front Oncol (2021) 662840. doi: 10.3389/fonc.2021.662840
- 181. Wang X, Zhang X, Qiu C, Yang N. STAT3 Contributes to Radioresistance in Cancer. Front Oncol (2020) 10:1120. doi: 10.3389/fonc.2020.01120
- 182. Mortezaee K, Najafi M, Farhood B, Ahmadi A, Shabeeb D, Musa AE. NFkappaB Targeting for Overcoming Tumor Resistance and Normal Tissues Toxicity. J Cell Physiol (2019) 234(10):17187–204. doi: 10.1002/jcp.28504
- 183. Pordanjani SM, Hosseinimehr SJ. The Role of NF-kB Inhibitors in Cell Response to Radiation. Curr Med Chem (2016) 23(34):3951-63. doi: 10.2174/0929867323666160824162718
- 184. Yi H, Yan X, Luo Q, Yuan L, Li B, Pan W, et al. A Novel Small Molecule Inhibitor of MDM2-P53 (APG-115) Enhances Radiosensitivity of Gastric Adenocarcinoma. J Exp Clin Cancer Res (2018) 37(1):1–10. doi: 10.1186/ s13046-018-0765-8
- 185. Feng FY, Zhang Y, Kothari V, Evans JR, Jackson WC, Chen W, et al. MDM2 Inhibition Sensitizes Prostate Cancer Cells to Androgen Ablation and Radiotherapy in a P53-Dependent Manner. *Neoplasia* (2016) 18(4):213–22. doi: 10.1016/j.neo.2016.01.006
- 186. Werner LR, Huang S, Francis DM, Armstrong EA, Ma F, Li C, et al. Small Molecule Inhibition of MDM2-p53 Interaction Augments Radiation Response in Human Tumors. *Mol Cancer Ther* (2015) 14(9):1994–2003. doi: 10.1158/1535-7163.MCT-14-1056-T
- 187. Phelps D, Bondra K, Seum S, Chronowski C, Leasure J, Kurmasheva RT, et al. Inhibition of MDM2 by RG7388 Confers Hypersensitivity to X-

Radiation in Xenograft Models of Childhood Sarcoma. *Pediatr Blood Cancer* (2015) 62(8):1345–52. doi: 10.1002/pbc.25465

- Rew Y, Sun D. Discovery of a Small Molecule MDM2 Inhibitor (AMG 232) for Treating Cancer. J medicinal Chem (2014) 57(15):6332–41. doi: 10.1021/ jm500627s
- 189. Bonner JA, Harari PM, Giralt J, Azarnia N, Shin DM, Cohen RB, et al. Radiotherapy Plus Cetuximab for Squamous-Cell Carcinoma of the Head and Neck. New Engl J Med (2006) 354(6):567–78. doi: 10.1056/ NEJMoa053422
- 190. Nieder C, Pawinski A, Dalhaug A, Andratschke N. A Review of Clinical Trials of Cetuximab Combined With Radiotherapy for Non-Small Cell Lung Cancer. *Radiat Oncol* (2012) 7(1):1–7. doi: 10.1186/1748-717X-7-3
- 191. Toulany M. Targeting DNA Double-Strand Break Repair Pathways to Improve Radiotherapy Response. *Genes* (2019) 10(1):25. doi: 10.3390/ genes10010025
- 192. Das AK, Chen BP, Story MD, Sato M, Minna JD, Chen DJ, et al. Somatic Mutations in the Tyrosine Kinase Domain of Epidermal Growth Factor Receptor (EGFR) Abrogate EGFR-Mediated Radioprotection in Non–Small Cell Lung Carcinoma. *Cancer Res* (2007) 67(11):5267–74. doi: 10.1158/0008-5472.CAN-07-0242
- 193. Huang S-M, Bock JM, Harari PM. Epidermal Growth Factor Receptor Blockade With C225 Modulates Proliferation, Apoptosis, and Radiosensitivity in Squamous Cell Carcinomas of the Head and Neck. *Cancer Res* (1999) 59(8):1935–40.
- 194. Pfäffle HN, Wang M, Gheorghiu L, Ferraiolo N, Greninger P, Borgmann K, et al. EGFR-Activating Mutations Correlate With a Fanconi Anemia–like Cellular Phenotype That Includes PARP Inhibitor Sensitivity. *Cancer Res* (2013) 73(20):6254–63. doi: 10.1158/0008-5472.CAN-13-0044
- 195. Johung KL, Yao X, Li F, James BY, Gettinger SN, Goldberg S, et al. A Clinical Model for Identifying Radiosensitive Tumor Genotypes in Non–Small Cell Lung Cancer. *Clin Cancer Res* (2013) 19(19):5523–32. doi: 10.1158/1078-0432.CCR-13-0836
- 196. Braunstein LZ, Taghian AG, Niemierko A, Salama L, Capuco A, Bellon JR, et al. Breast-Cancer Subtype, Age, and Lymph Node Status as Predictors of Local Recurrence Following Breast-Conserving Therapy. *Breast Cancer Res Treat* (2017) 161(1):173–9. doi: 10.1007/s10549-016-4031-5
- 197. Liang K, Lu Y, Jin W, Ang KK, Milas L, Fan Z. Sensitization of Breast Cancer Cells to Radiation by Trastuzumab. *Mol Cancer Ther* (2003) 2(11):1113–20.
- 198. Abi Jaoude J, de Azambuja E, Makki M, Tamim H, Tfayli A, Geara F, et al. Post-Mastectomy Radiation Therapy in Human Epidermal Growth Factor Receptor 2 Positive Breast Cancer Patients: Analysis of the HERA Trial. *Int J Radiat Oncol Biol Phys* (2020) 106(3):503–10. doi: 10.1016/ j.ijrobp.2019.10.022
- 199. Wolff RF, Ryder S, Bossi A, Briganti A, Crook J, Henry A, et al. A Systematic Review of Randomised Controlled Trials of Radiotherapy for Localised Prostate Cancer. *Eur J Cancer* (2015) 51(16):2345–67. doi: 10.1016/ j.ejca.2015.07.019
- 200. Messing EM, Manola J, Yao J, Kiernan M, Crawford D, Wilding G, et al. Immediate Versus Deferred Androgen Deprivation Treatment in Patients With Node-Positive Prostate Cancer After Radical Prostatectomy and Pelvic Lymphadenectomy. Lancet Oncol (2006) 7(6):472–9. doi: 10.1016/S1470-2045(06)70700-8
- 201. Milosevic M, Chung P, Parker C, Bristow R, Toi A, Panzarella T, et al. Androgen Withdrawal in Patients Reduces Prostate Cancer Hypoxia: Implications for Disease Progression and Radiation Response. *Cancer Res* (2007) 67(13):6022–5. doi: 10.1158/0008-5472.CAN-07-0561
- 202. Wo JY, Zietman AL. Why does androgen deprivation enhance the results of radiation therapy? Urol Oncol (2008) 26(5):522–9. doi: 10.1016/ j.urolonc.2008.03.008
- 203. Polkinghorn WR, Parker JS, Lee MX, Kass EM, Spratt DE, Iaquinta PJ, et al. Androgen Receptor Signaling Regulates DNA Repair in Prostate Cancers. *Cancer Discov* (2013) 3(11):1245–53. doi: 10.1158/2159-8290.CD-13-0172
- 204. Zumsteg ZS, Morse N, Krigsfeld G, Gupta G, Higginson DS, Lee NY, et al. Taselisib (GDC-0032), a Potent β-Sparing Small Molecule Inhibitor of PI3K, Radiosensitizes Head and Neck Squamous Carcinomas Containing Activating PIK3CA Alterations. *Clin Cancer Res* (2016) 22(8):2009–19. doi: 10.1158/1078-0432.CCR-15-2245

- Chinnaiyan P, Vallabhaneni G, Armstrong E, Huang S-M, Harari PM. Modulation of Radiation Response by Histone Deacetylase Inhibition. *Int J Radiat Oncol Biol Phys* (2005) 62(1):223–9. doi: 10.1016/j.ijrobp.2004.12.088
- 206. Gampala S, Shah F, Lu X, Moon H-R, Sandusky G, Hulsey E, et al. Ref-1 Redox Activity Alters Cancer Cell Metabolism in Pancreatic Cancer: Exploiting This Novel Finding as a Potential Target. (2020) 40(1):251. doi: 10.21203/rs.3.rs-125304/v1
- 207. Zhao Y, Feng X, Chen Y, Selfridge JE, Gorityala S, Du Z, et al. 5-Fluorouracil Enhances the Antitumor Activity of the Glutaminase Inhibitor CB-839 Against PIK3CA-Mutant Colorectal Cancers. *Cancer Res* (2020) 80 (21):4815–27. doi: 10.1158/0008-5472.CAN-20-0600
- 208. Momcilovic M, Bailey ST, Lee JT, Fishbein MC, Magyar C, Braas D, et al. Targeted Inhibition of EGFR and Glutaminase Induces Metabolic Crisis in EGFR Mutant Lung Cancer. *Cell Rep* (2017) 18(3):601–10. doi: 10.1016/ j.celrep.2016.12.061
- 209. Gray LH, Conger AD, Ebert M, Hornsey S, Scott O. The Concentration of Oxygen Dissolved in Tissues at the Time of Irradiation as a Factor in Radiotherapy. Br J Radiol (1953) 26(312):638–48. doi: 10.1259/0007-1285-26-312-638
- 210. Overgaard J. Hypoxic Modification of Radiotherapy in Squamous Cell Carcinoma of the Head and Neck-A Systematic Review and Meta-Analysis. *Radiotherapy Oncol* (2011) 100(1):22–32. doi: 10.1016/ j.radonc.2011.03.004
- 211. Bentzen J, Toustrup K, Eriksen JG, Primdahl H, Andersen LJ, Overgaard J. Locally Advanced Head and Neck Cancer Treated With Accelerated Radiotherapy, the Hypoxic Modifier Nimorazole and Weekly Cisplatin. Results From the DAHANCA 18 Phase II Study. *Acta Oncol* (2015) 54 (7):1001–7. doi: 10.3109/0284186X.2014.992547
- 212. Mistry IN, Thomas M, Calder ED, Conway SJ, Hammond EM. Clinical Advances of Hypoxia-Activated Prodrugs in Combination With Radiation Therapy. Int J Radiat Oncol Biol Phys (2017) 98(5):1183–96. doi: 10.1016/ j.ijrobp.2017.03.024
- 213. Fokas E, Im JH, Hill S, Yameen S, Stratford M, Beech J, et al. Dual Inhibition of the PI3K/mTOR Pathway Increases Tumor Radiosensitivity by Normalizing Tumor Vasculature. *Cancer Res* (2012) 72(1):239–48. doi: 10.1158/0008-5472.CAN-11-2263
- 214. McGowan DR, Skwarski M, Bradley KM, Campo L, Fenwick JD, Gleeson FV, et al. Buparlisib With Thoracic Radiotherapy and Its Effect on Tumour Hypoxia: A Phase I Study in Patients With Advanced Non-Small Cell Lung Carcinoma. *Eur J Cancer* (2019) 113:87–95. doi: 10.1016/j.ejca.2019.03.015
- Jain RK, Di Tomaso E, Duda DG, Loeffler JS, Sorensen AG, Batchelor TT. Angiogenesis in Brain Tumours. Nat Rev Neurosci (2007) 8(8):610–22. doi: 10.1038/nrn2175
- 216. Rich JN, Sathornsumetee S, Keir ST, Kieran MW, Laforme A, Kaipainen A, et al. ZD6474, a Novel Tyrosine Kinase Inhibitor of Vascular Endothelial Growth Factor Receptor and Epidermal Growth Factor Receptor, Inhibits Tumor Growth of Multiple Nervous System Tumors. *Clin Cancer Res* (2005) 11(22):8145–57. doi: 10.1158/1078-0432.CCR-05-0319
- 217. Sandström M, Johansson M, Bergström P, Bergenheim AT, Henriksson R. Effects of the VEGFR Inhibitor ZD6474 in Combination With Radiotherapy and Temozolomide in an Orthotopic Glioma Model. J Neuro-Oncology (2008) 88(1):1–9. doi: 10.1007/s11060-008-9527-3
- Damiano V, Melisi D, Bianco C, Raben D, Caputo R, Fontanini G, et al. Cooperative Antitumor Effect of Multitargeted Kinase Inhibitor ZD6474 and Ionizing Radiation in Glioblastoma. *Clin Cancer Res* (2005) 11(15):5639–44. doi: 10.1158/1078-0432.CCR-05-0174
- 219. Wang Z, Tang Y, Tan Y, Wei Q, Yu W. Cancer-Associated Fibroblasts in Radiotherapy: Challenges and New Opportunities. *Cell Communication Signaling* (2019) 17(1):1–12. doi: 10.1186/s12964-019-0362-2
- 220. Ashton TM, McKenna WG, Kunz-Schughart LA, Higgins GS. Oxidative Phosphorylation as an Emerging Target in Cancer Therapy. *Clin Cancer Res* (2018) 24(11):2482–90. doi: 10.1158/1078-0432.CCR-17-3070
- 221. Ashton TM, Fokas E, Kunz-Schughart LA, Folkes LK, Anbalagan S, Huether M, et al. The Anti-Malarial Atovaquone Increases Radiosensitivity by Alleviating Tumour Hypoxia. Nat Commun (2016) 7(1):1–13. doi: 10.1038/ncomms12308
- 222. Liu Z, Tan H, Zhang X, Chen F, Zhou Z, Hu X, et al. Enhancement of Radiotherapy Efficacy by Silver Nanoparticles in Hypoxic Glioma Cells. *Artif*

Cells Nanomedicine Biotechnol (2018) 46(sup3):\$922-30. doi: 10.1080/ 21691401.2018.1518912

- 223. Song G, Chao Y, Chen Y, Liang C, Yi X, Yang G, et al. All-In-One Theranostic Nanoplatform Based on Hollow TaOx for Chelator-Free Labeling Imaging, Drug Delivery, and Synergistically Enhanced Radiotherapy. Advanced Funct Materials (2016) 26(45):8243-54. doi: 10.1002/adfm.201603845
- 224. Kempson I. Mechanisms of Nanoparticle Radiosensitization. Wiley Interdiscip Reviews: Nanomedicine Nanobiotechnology (2021) 13(1):e1656. doi: 10.1002/wnan.1656
- 225. Magrini SM, Buglione M, Corvò R, Pirtoli L, Paiar F, Ponticelli P, et al. Cetuximab and Radiotherapy *Versus* Cisplatin and Radiotherapy for Locally Advanced Head and Neck Cancer: A Randomized Phase II Trial. *J Clin Oncol* (2016) 34(5):427–35. doi: 10.1200/JCO.2015.63.1671
- 226. Mehanna H, Robinson M, Hartley A, Kong A, Foran B, Fulton-Lieuw T, et al. Radiotherapy Plus Cisplatin or Cetuximab in Low-Risk Human Papillomavirus-Positive Oropharyngeal Cancer (De-ESCALaTE HPV): An Open-Label Randomised Controlled Phase 3 Trial. *Lancet* (2019) 393 (10166):51–60. doi: 10.1016/S0140-6736(18)32752-1
- 227. Gillison ML, Trotti AM, Harris J, Eisbruch A, Harari PM, Adelstein DJ, et al. Radiotherapy Plus Cetuximab or Cisplatin in Human Papillomavirus-Positive Oropharyngeal Cancer (NRG Oncology RTOG 1016): A Randomised, Multicentre, Non-Inferiority Trial. *Lancet* (2019) 393 (10166):40–50. doi: 10.1016/S0140-6736(18)32779-X
- 228. Bradley JD, Paulus R, Komaki R, Masters G, Blumenschein G, Schild S, et al. Standard-Dose Versus High-Dose Conformal Radiotherapy With Concurrent and Consolidation Carboplatin Plus Paclitaxel With or Without Cetuximab for Patients With Stage IIIA or IIIB Non-Small-Cell Lung Cancer (RTOG 0617): A Randomised, Two-by-Two Factorial Phase 3 Study. Lancet Oncol (2015) 16 (2):187–99. doi: 10.1016/S1470-2045(14)71207-0
- 229. Garg MK, Zhao F, Sparano JA, Palefsky J, Whittington R, Mitchell EP, et al. Cetuximab Plus Chemoradiotherapy in Immunocompetent Patients With Anal Carcinoma: A Phase II Eastern Cooperative Oncology Group– American College of Radiology Imaging Network Cancer Research Group Trial (E3205). J Clin Oncol (2017) 35(7):718. doi: 10.1200/JCO.2016.69.1667
- 230. Sparano JA, Lee JY, Palefsky J, Henry DH, Wachsman W, Rajdev L, et al. Cetuximab Plus Chemoradiotherapy for HIV-Associated Anal Carcinoma: A Phase II AIDS Malignancy Consortium Trial. J Clin Oncol (2017) 35(7):727. doi: 10.1200/JCO.2016.69.1642
- 231. Libermann TA, Nusbaum HR, Razon N, Kris R, Lax I, Soreq H, et al. Amplification, Enhanced Expression and Possible Rearrangement of EGF Receptor Gene in Primary Human Brain Tumours of Glial Origin. *Nature* (1985) 313(5998):144–7. doi: 10.1038/313144a0
- 232. Chakravarti A, Chakladar A, Delaney MA, Latham DE, Loeffler JS. The Epidermal Growth Factor Receptor Pathway Mediates Resistance to Sequential Administration of Radiation and Chemotherapy in Primary Human Glioblastoma Cells in a RAS-Dependent Manner. *Cancer Res* (2002) 62(15):4307–15.
- 233. Barker FGII, Simmons ML, Chang SM, Prados MD, Larson DA, Sneed PK, et al. EGFR Overexpression and Radiation Response in Glioblastoma Multiforme. Int J Radiat Oncol Biol Phys (2001) 51(2):410–8. doi: 10.1016/ S0360-3016(01)01609-1
- 234. Iannitti D, Dipetrillo T, Akerman P, Barnett JM, Maia-Acuna C, Cruff D, et al. Erlotinib and Chemoradiation Followed by Maintenance Erlotinib for Locally Advanced Pancreatic Cancer: A Phase I Study. Am J Clin Oncol (2005) 28(6):570–5. doi: 10.1097/01.coc.0000184682.51193.00
- 235. Duffy A, Kortmansky J, Schwartz G, Capanu M, Puleio S, Minsky B, et al. A Phase I Study of Erlotinib in Combination With Gemcitabine and Radiation in Locally Advanced, Non-Operable Pancreatic Adenocarcinoma. *Ann Oncol* (2008) 19(1):86–91. doi: 10.1093/annonc/mdm441
- 236. Ma WW, Herman JM, Jimeno A, Laheru D, Messersmith WA, Wolfgang CL, et al. A Tolerability and Pharmacokinetic Study of Adjuvant Erlotinib and Capecitabine With Concurrent Radiation in Resected Pancreatic Cancer. *Trans Oncol* (2010) 3(6):373–9. doi: 10.1593/tlo.10196
- 237. Bao PQ, Ramanathan RK, Krasinkas A, Bahary N, Lembersky BC, Bartlett DL, et al. Phase II Study of Gemcitabine and Erlotinib as Adjuvant Therapy for Patients With Resected Pancreatic Cancer. *Ann Surg Oncol* (2011) 18 (4):1122–9. doi: 10.1245/s10434-010-1401-9

- 238. Herman JM, Fan KY, Wild AT, Hacker-Prietz A, Wood LD, Blackford AL, et al. Phase 2 Study of Erlotinib Combined With Adjuvant Chemoradiation and Chemotherapy in Patients With Resectable Pancreatic Cancer. Int J Radiat Oncol Biol Phys (2013) 86(4):678–85. doi: 10.1016/ j.ijrobp.2013.03.032
- 239. Ma DJ, Galanis E, Anderson SK, Schiff D, Kaufmann TJ, Peller PJ, et al. A Phase II Trial of Everolimus, Temozolomide, and Radiotherapy in Patients With Newly Diagnosed Glioblastoma: NCCTG N057K. *Neuro-Oncology* (2015) 17(9):1261–9. doi: 10.1093/neuonc/nou328
- 240. Chinnaiyan P, Won M, Wen PY, Rojiani AM, Werner-Wasik M, Shih HA, et al. A Randomized Phase II Study of Everolimus in Combination With Chemoradiation in Newly Diagnosed Glioblastoma: Results of NRG Oncology RTOG 0913. *Neuro-Oncology* (2018) 20(5):666–73. doi: 10.1093/ neuonc/nox209
- 241. Overgaard J, Hansen HS, Overgaard M, Bastholt L, Berthelsen A, Specht L, et al. A Randomized Double-Blind Phase III Study of Nimorazole as a Hypoxic Radiosensitizer of Primary Radiotherapy in Supraglottic Larynx and Pharynx Carcinoma. Results of the Danish Head and Neck Cancer Study (DAHANCA) Protocol 5-85. *Radiotherapy Oncol* (1998) 46(2):135–46. doi: 10.1016/s0167-8140(97)00220-x
- 242. Overgaard J. Hypoxic Radiosensitization: Adored and Ignored. J Clin Oncol (2007) 25(26):4066–74. doi: 10.1200/JCO.2007.12.7878
- 243. Kaanders JH, Pop LA, Marres HA, Bruaset I, van den Hoogen FJ, Merkx MA, et al. ARCON: Experience in 215 Patients With Advanced Head-and-Neck Cancer. Int J Radiat Oncol Biol Phys (2002) 52(3):769–78. doi: 10.1016/ S0360-3016(01)02678-5
- 244. Hoskin P, Rojas A, Saunders M. Accelerated Radiotherapy, Carbogen, and Nicotinamide (ARCON) in the Treatment of Advanced Bladder Cancer: Mature Results of a Phase II Nonrandomized Study. *Int J Radiat Oncol Biol Phys* (2009) 73(5):1425–31. doi: 10.1016/j.ijrobp.2008.06.1950
- 245. Hoskin PJ, Rojas AM, Bentzen SM, Saunders MI. Radiotherapy With Concurrent Carbogen and Nicotinamide in Bladder Carcinoma. J Clin Oncol (2010) 28(33):4912–8. doi: 10.1200/JCO.2010.28.4950
- 246. Janssens GO, Rademakers SE, Terhaard CH, Doornaert PA, Bijl HP, van den Ende P, et al. Accelerated Radiotherapy With Carbogen and Nicotinamide for Laryngeal Cancer: Results of a Phase III Randomized Trial. *J Clin Oncol* (2012) 30(15):1777–83. doi: 10.1200/JCO.2011.35.9315
- 247. Barker HE, Paget JT, Khan AA, Harrington KJ. The Tumour Microenvironment After Radiotherapy: Mechanisms of Resistance and Recurrence. *Nat Rev Cancer* (2015) 15(7):409–25. doi: 10.1038/nrc3958
- 248. DiSilvestro PA, Ali S, Craighead PS, Lucci JA, Lee Y-C, Cohn DE, et al. Phase III Randomized Trial of Weekly Cisplatin and Irradiation Versus Cisplatin and Tirapazamine and Irradiation in Stages IB2, IIA, IIB, IIIB, and IVA Cervical Carcinoma Limited to the Pelvis: A Gynecologic Oncology Group Study. J Clin Oncol (2014) 32(5):458. doi: 10.1200/JCO.2013.51.4265
- 249. Rischin D, Peters LJ, O'Sullivan B, Giralt J, Fisher R, Yuen K, et al. Tirapazamine, Cisplatin, and Radiation Versus Cisplatin and Radiation for Advanced Squamous Cell Carcinoma of the Head and Neck (TROG 02.02, HeadSTART): A Phase III Trial of the Trans-Tasman Radiation Oncology Group. Oncology (2008) 28(18):2989-95. doi: 10.1200/ jco.2008.26.15_suppl.lba6008
- 250. Gilbert MR, Dignam JJ, Armstrong TS, Wefel JS, Blumenthal DT, Vogelbaum MA, et al. A Randomized Trial of Bevacizumab for Newly Diagnosed Glioblastoma. *New Engl J Med* (2014) 370(8):699–708. doi: 10.1056/NEJMoa1308573
- 251. Chinot OL, Wick W, Mason W, Henriksson R, Saran F, Nishikawa R, et al. Bevacizumab Plus Radiotherapy–Temozolomide for Newly Diagnosed Glioblastoma. New Engl J Med (2014) 370(8):709–22. doi: 10.1056/ NEJMoa1308345
- 252. Lai A, Tran A, Nghiemphu PL, Pope WB, Solis OE, Selch M, et al. Phase II Study of Bevacizumab Plus Temozolomide During and After Radiation Therapy for Patients With Newly Diagnosed Glioblastoma Multiforme. *J Clin Oncol* (2011) 29(2):142. doi: 10.1200/JCO.2010.30.2729
- 253. Batchelor TT, Gerstner ER, Emblem KE, Duda DG, Kalpathy-Cramer J, Snuderl M, et al. Improved Tumor Oxygenation and Survival in Glioblastoma Patients Who Show Increased Blood Perfusion After Cediranib and Chemoradiation. *Proc Natl Acad Sci* (2013) 110(47):19059– 64. doi: 10.1073/pnas.1318022110

- 254. Murphy JE, Wo JY, Ryan DP, Clark JW, Jiang W, Yeap BY, et al. Total Neoadjuvant Therapy With FOLFIRINOX in Combination With Losartan Followed by Chemoradiotherapy for Locally Advanced Pancreatic Cancer: A Phase 2 Clinical Trial. JAMA Oncol (2019) 5(7):1020–7. doi: 10.1001/ jamaoncol.2019.0892
- Prior IA, Lewis PD, Mattos C. A Comprehensive Survey of Ras Mutations in Cancer. Cancer Res (2012) 72(10):2457–67. doi: 10.1158/0008-5472.CAN-11-2612
- 256. Bernhard EJ, Stanbridge EJ, Gupta S, Gupta AK, Soto D, Bakanauskas VJ, et al. Direct Evidence for the Contribution of Activated N-Ras and K-Ras Oncogenes to Increased Intrinsic Radiation Resistance in Human Tumor Cell Lines. *Cancer Res* (2000) 60(23):6597–600.
- 257. Cengel KA, Voong KR, Chandrasekaran S, Maggiorella L, Brunner TB, Stanbridge E, et al. Oncogenic K-Ras Signals Through Epidermal Growth Factor Receptor and Wild-Type H-Ras to Promote Radiation Survival in Pancreatic and Colorectal Carcinoma Cells. *Neoplasia* (2007) 9(4):341–8. doi: 10.1593/neo.06823
- 258. Kim I-A, Bae S-S, Fernandes A, Wu J, Muschel RJ, McKenna WG, et al. Selective Inhibition of Ras, Phosphoinositide 3 Kinase, and Akt Isoforms Increases the Radiosensitivity of Human Carcinoma Cell Lines. *Cancer Res* (2005) 65(17):7902–10. doi: 10.1158/0008-5472.CAN-05-0513
- 259. Mak RH, Hermann G, Lewis JH, Aerts HJ, Baldini EH, Chen AB, et al. Outcomes by Tumor Histology and KRAS Mutation Status After Lung Stereotactic Body Radiation Therapy for Early-Stage Non–Small-Cell Lung Cancer. *Clin Lung Cancer* (2015) 16(1):24–32. doi: 10.1016/j.cllc.2014.09.005
- 260. Hong TS, Wo JY, Borger DR, Yeap BY, McDonnell EI, Willers H, et al. Phase II Study of Proton-Based Stereotactic Body Radiation Therapy for Liver Metastases: Importance of Tumor Genotype. *JNCI: J Natl Cancer Institute* (2017) 109(9):djx031. doi: 10.1093/jnci/djx031
- 261. Jethwa KR, Jang S, Mullikin TC, Harmsen WS, Petersen MM, Olivier KR, et al. Association of Tumor Genomic Factors and Efficacy for Metastasis-Directed Stereotactic Body Radiotherapy for Oligometastatic Colorectal Cancer. *Radiotherapy Oncol* (2020) 146:29–36. doi: 10.1016/j.radonc.2020.02.008
- 262. Gallogly MM, Lazarus HM, Cooper BW. Midostaurin: A Novel Therapeutic Agent for Patients With FLT3-Mutated Acute Myeloid Leukemia and Systemic Mastocytosis. *Ther Adv Hematol* (2017) 8(9):245–61. doi: 10.1177/2040620717721459
- 263. Hong TS, Wo JY-L, Ryan DP, Zheng H, Borger DR, Kwak EL, et al. Phase Ib Study of Neoadjuvant Chemoradiation (CRT) With Midostaurin, 5-Fluorouracil (5-FU) and Radiation (XRT) for Locally Advanced Rectal Cancer: Sensitization of RAS Mutant Tumors. Am Soc Clin Oncol (2018) 36(15_suppl):e15674–e15674. doi: 10.1200/JCO.2018.36.15_suppl.e15674
- 264. Liu Q, Wang M, Kern AM, Khaled S, Han J, Yeap BY, et al. Adapting a Drug Screening Platform to Discover Associations of Molecular Targeted Radiosensitizers With Genomic Biomarkers. *Mol Cancer Res* (2015) 13 (4):713–20. doi: 10.1158/1541-7786.MCR-14-0570
- 265. Lin SH, Mok I, Leos D, Pasia MG, Thall PF, Lin HY, et al. NCI 9448: Phase I Study of Trametinib in Combination With Chemoradiation for KRAS-Mutant Non-Small Cell Lung Cancer. Am Soc Clin Oncol (2015) 33 (15_suppl):TPS7585-TPS7585. doi: 10.1200/jco.2015.33.15_suppl.tps7585
- 266. Hong DS, Fakih MG, Strickler JH, Desai J, Durm GA, Shapiro GI, et al. KRASG12C Inhibition With Sotorasib in Advanced Solid Tumors. *New Engl J Med* (2020) 383(13):1207–17. doi: 10.1056/NEJMoa1917239
- 267. O'Connor MJ. Targeting the DNA Damage Response in Cancer. Mol Cell (2015) 60(4):547–60. doi: 10.1016/j.molcel.2015.10.040
- 268. Jagsi R, Griffith KA, Bellon JR, Woodward WA, Horton JK, Ho A, et al. Concurrent Veliparib With Chest Wall and Nodal Radiotherapy in Patients With Inflammatory or Locoregionally Recurrent Breast Cancer: The TBCRC 024 Phase I Multicenter Study. J Clin Oncol (2018) 36(13):1317. doi: 10.1200/ JCO.2017.77.2665
- 269. Connolly E, Silvera D, Badura M, Braunstein S, Formenti S, Schneider R. Inflammatory Breast Cancer Radio-Resistance and Its Cancer Stem Cell Population are Oppositely Controlled by Translation Factor Eif4g. *Int J Radiat Oncol Biol Phys* (2010) 78(3):S221. doi: 10.1016/j.ijrobp.2010.07.531
- 270. Hu Y, Guo M. Synthetic Lethality Strategies: Beyond BRCA1/2 Mutations in Pancreatic Cancer. *Cancer Sci* (2020) 111(9):3111–21. doi: 10.1111/cas.14565
- 271. Tuli R, Shiao SL, Nissen N, Tighiouart M, Kim S, Osipov A, et al. A Phase 1 Study of Veliparib, a PARP-1/2 Inhibitor, With Gemcitabine and

Radiotherapy in Locally Advanced Pancreatic Cancer. *EBioMedicine* (2019) 40:375–81. doi: 10.1016/j.ebiom.2018.12.060

- 272. Karam SD, Reddy K, Blatchford PJ, Waxweiler T, DeLouize AM, Oweida A, et al. Final Report of a Phase I Trial of Olaparib With Cetuximab and Radiation for Heavy Smoker Patients With Locally Advanced Head and Neck Cancer. *Clin Cancer Res* (2018) 24(20):4949–59. doi: 10.1158/1078-0432.CCR-18-0467
- 273. Cuneo KC, Morgan MA, Sahai V, Schipper MJ, Parsels LA, Parsels JD, et al. Dose Escalation Trial of the Wee1 Inhibitor Adavosertib (AZD1775) in Combination With Gemcitabine and Radiation for Patients With Locally Advanced Pancreatic Cancer. J Clin Oncol (2019) 37(29):2643. doi: 10.1200/JCO.19.00730
- 274. Hammel P, Huguet F, van Laethem J-L, Goldstein D, Glimelius B, Artru P, et al. Effect of Chemoradiotherapy vs Chemotherapy on Survival in Patients With Locally Advanced Pancreatic Cancer Controlled After 4 Months of Gemcitabine With or Without Erlotinib: The LAP07 Randomized Clinical Trial. Jama (2016) 315(17):1844–53. doi: 10.1001/jama.2016.4324
- 275. Van Triest B, Damstrup L, Falkenius J, Budach V, Troost E, Samuels M, et al. A Phase Ia/Ib Trial of the DNA-PK Inhibitor M3814 in Combination With Radiotherapy (RT) in Patients (Pts) With Advanced Solid Tumors: Dose-Escalation Results. Am Soc Clin Oncol (2018) 36(15_suppl):2518. doi: 10.1200/JCO.2018.36.15_suppl.2518
- 276. Huang R-X, Zhou P-K. DNA Damage Response Signaling Pathways and Targets for Radiotherapy Sensitization in Cancer. Signal Transduction Targeted Ther (2020) 5(1):1–27. doi: 10.1038/s41392-020-0150-x
- 277. Kwatra D, Venugopal A, Anant S. Nanoparticles in Radiation Therapy: A Summary of Various Approaches to Enhance Radiosensitization in Cancer. *Transl Cancer Res* (2013) 2(4):330–42. doi: 10.3978/j.issn.2218-676X.2013.08.06
- 278. Bonvalot S, Rutkowski PL, Thariat J, Carrère S, Ducassou A, Sunyach M-P, et al. NBTXR3, a First-in-Class Radioenhancer Hafnium Oxide Nanoparticle, Plus Radiotherapy Versus Radiotherapy Alone in Patients With Locally Advanced Soft-Tissue Sarcoma (Act. In. Sarc): A Multicentre, Phase 2–3, Randomised, Controlled Trial. Lancet Oncol (2019) 20(8):1148–59. doi: 10.1016/S1470-2045(19)30326-2
- Pallares RM, Abergel RJ. Nanoparticles for Targeted Cancer Radiotherapy. Nano Res (2020) 1–11. doi: 10.1007/s12274-020-2957-8
- Antonia SJ, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, et al. Durvalumab After Chemoradiotherapy in Stage III Non–Small-Cell Lung Cancer. New Engl J Med (2017) 377(20):1919–29. doi: 10.1056/NEJMoa1709937
- Antonia SJ, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, et al. Overall Survival With Durvalumab After Chemoradiotherapy in Stage III NSCLC. New Engl J Med (2018) 379(24):2342–50. doi: 10.1056/NEJMoa1809697
- 282. Lin SH, Lin Y, Yao L, Kalhor N, Carter BW, Altan M, et al. Phase II Trial of Concurrent Atezolizumab With Chemoradiation for Unresectable NSCLC. J Thorac Oncol (2020) 15(2):248–57. doi: 10.1016/j.jtho.2019.10.024
- 283. Jabbour SK, Berman AT, Decker RH, Lin Y, Feigenberg SJ, Gettinger SN, et al. Phase 1 Trial of Pembrolizumab Administered Concurrently With Chemoradiotherapy for Locally Advanced Non–Small Cell Lung Cancer: A Nonrandomized Controlled Trial. JAMA Oncol (2020) 6(6):848–55. doi: 10.1001/jamaoncol.2019.6731
- 284. Weiss J, Bauman JR, Deal AM, Sheth S, Chera BS, Shen C, et al. Preliminary Toxicity Data From the Combination of Pembrolizumab and Definitive-Dose Radiotherapy for Locally Advanced Head and Neck Cancer With Contraindication to Cisplatin Therapy. Am Soc Clin Oncol (2018) 36 (15_suppl):6069. doi: 10.1200/JCO.2018.36.15_suppl.6069
- 285. Sun XS, Sire C, Tao Y, Martin L, Alfonsi M, Prevost JB, et al. A Phase II Randomized Trial of Pembrolizumab Versus Cetuximab, Concomitant With Radiotherapy (RT) in Locally Advanced (LA) Squamous Cell Carcinoma of the Head and Neck (SCCHN): First Results of the GORTEC 2015-01 "PembroRad" Trial. J Clin Oncol (2018) 36(15_suppl):6018. doi: 10.1200/ JCO.2018.36.15_suppl.6018
- 286. Powell SF, Gitau MM, Sumey CJ, Reynolds JT, Lohr M, McGraw S, et al. Safety of Pembrolizumab With Chemoradiation (CRT) in Locally Advanced Squamous Cell Carcinoma of the Head and Neck (LA-SCCHN). Am Soc Clin Oncol (2017) 35(15_suppl):6011. doi: 10.1200/JCO.2017.35.15_suppl.6011
- 287. Tao Y, Aupérin A, Sun X, Sire C, Martin L, Coutte A, et al. Avelumabcetuximab-radiotherapy Versus Standards of Care in Locally Advanced Squamous-Cell Carcinoma of the Head and Neck: The Safety Phase of a

Randomised Phase III Trial GORTEC 2017-01 (REACH). Eur J Cancer (2020) 141:21-9. doi: 10.1016/j.ejca.2020.09.008

- 288. Weiss J, Sheth S, Deal AM, Olson JEG, Patel S, Hackman TG, et al. Concurrent Definitive Immunoradiotherapy for Patients With Stage III–IV Head and Neck Cancer and Cisplatin Contraindication. *Clin Cancer Res* (2020) 26(16):4260–7. doi: 10.1158/1078-0432.CCR-20-0230
- Rodríguez-Ruiz ME, Vanpouille-Box C, Melero I, Formenti SC, Demaria S. Immunological Mechanisms Responsible for Radiation-Induced Abscopal Effect. *Trends Immunol* (2018) 39(8):644–55. doi: 10.1016/j.it.2018.06.001
- 290. Theelen WS, Peulen HM, Lalezari F, van der Noort V, De Vries JF, Aerts JG, et al. Effect of Pembrolizumab After Stereotactic Body Radiotherapy vs Pembrolizumab Alone on Tumor Response in Patients With Advanced Non–Small Cell Lung Cancer: Results of the PEMBRO-RT Phase 2 Randomized Clinical Trial. JAMA Oncol (2019) 5(9):1276–82. doi: 10.1001/jamaoncol.2019.1478
- 291. McBride S, Sherman E, Tsai CJ, Baxi S, Aghalar J, Eng J, et al. Randomized Phase II Trial of Nivolumab With Stereotactic Body Radiotherapy Versus Nivolumab Alone in Metastatic Head and Neck Squamous Cell Carcinoma. J Clin Oncol (2021) 39(1):30–7. doi: 10.1200/JCO.20.00290
- 292. Altorki NK, McGraw TE, Borczuk AC, Saxena A, Port JL, Stiles BM, et al. Neoadjuvant Durvalumab With or Without Stereotactic Body Radiotherapy in Patients With Early-Stage Non-Small-Cell Lung Cancer: A Single-Centre, Randomised Phase 2 Trial. *Lancet Oncol* (2021) 22(6):824–35. doi: 10.1016/ S1470-2045(21)00149-2

- 293. Alexander BM, Schoenfeld JD, Trippa L. Hazards of Hazard Ratios-Deviations From Model Assumptions in Immunotherapy. New Engl J Med (2018) 378(12):1158–9. doi: 10.1056/NEJMc1716612
- 294. Walker AJ, DeWeese TL, Viswanathan AN. Drug-Radiotherapy Combinations in 2020—A Landmark Year? *JAMA Oncol* (2021) 7(3):349– 50. doi: 10.1001/jamaoncol.2020.6139

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