



Dual Targeting of Multiple Myeloma Stem Cells and Myeloid-Derived Suppressor Cells for Treatment of Chemotherapy-Resistant Multiple Myeloma

Fatih M. Uckun^{1,2,3*}

¹ Department of Developmental Therapeutics, Immunology, and Integrative Medicine, Drug Discovery Institute, Ares Pharmaceuticals, St. Paul, MN, United States, ² Clinical Research Program, Aptevo Therapeutics, Seattle, WA, United States, ³ Translational Oncology Program, Reven Pharmaceuticals, Westminster, CO, United States

OPEN ACCESS

Edited by:

Giovanni Martinelli, University of Bologna, Italy

Reviewed by:

Amol Padgaonkar, Prescient Healthcare Group, United States Cirino Botta, University of Palermo, Italy

*Correspondence: Fatih M. Uckun fatih.uckun@aresmit.com

Specialty section:

This article was submitted to Hematologic Malignancies, a section of the journal Frontiers in Oncology

Received: 18 August 2021 Accepted: 21 October 2021 Published: 10 November 2021

Citation:

Uckun FM (2021) Dual Targeting of Multiple Myeloma Stem Cells and Myeloid-Derived Suppressor Cells for Treatment of Chemotherapy-Resistant Multiple Myeloma. Front. Oncol. 11:760382. doi: 10.3389/fonc.2021.760382 Here we review the insights and lessons learned from early clinical trials of T-cell engaging bispecific antibodies (BsABs) as a new class of biotherapeutic drug candidates with clinical impact potential for the treatment of multiple myeloma (MM). BsABs are capable of redirecting host T-cell cytotoxicity in an MHC-independent manner to malignant MM clones as well as immunosuppressive myeloid-derived suppressor cells (MDSC). T-cell engaging BsAB targeting the BCMA antigen may help delay disease progression in MM by destroying the MM cells. T-cell engaging BsAB targeting both the malignant MM clones and the MDSC in the bone marrow microenvironment (BMME). BsABs may facilitate the development of a new therapeutic paradigm for achieving improved survival in MM by altering the immunosuppressive BMME. T-cell engaging BsiABs targeting the CD123 antigen may help delay disease progression in MM by depleting both the MDSC in the BMME and destroying the MM stem cells that also carry the CD123 antigen on their surface.

Keywords: tumor microenvironment (TME), multiple myeloma (MM), bispecific T-cell engagers (BiTEs), bispecific antibodies (BsABs), bone marrow microenvironment (BMME), myeloid-derived suppressor cells (MDSC)

MULTIPLE MYELOMA AND DRUG RESISTANCE

MM is a heterogenous hematologic malignancy and relapses due to resistant disease are common (1-4). Resistance of the malignant clones to multiple drugs hampers a more successful treatment outcome after contemporary standard of care regimens in MM (1-4). Personalized therapy platforms have been designed to overcome the drug resistance, including precision medicines, kinase inhibitors, CAR-T cells, and antibody therapeutics (5-8). Effective treatment of patients with drug-resistant relapsed disease continues to be an unmet medical need (1-4).

1

IMMUNOSUPPRESSIVE BONE MARROW MICROENVIRONMENT IN MULTIPLE MYELOMA

The immunosuppressive bone marrow microenvironment (BMME) in MM contains cellular elements that facilitate the immune evasion of malignant MM clones (9–13). These immunosuppressive cells include MDSCs, an immature myeloid cell population capable of inhibiting effector cytotoxic T-cell (CTL) populations as well as natural killer (NK) cells and contribute to the T-cell exhaustion which is a hallmark of the BMME in MM patients (4, 14–20). In addition, regulatory T cells (Tregs), regulatory B-cells (Bregs), and tumor-associated macrophages (TAM) also contribute to BMME-associated immunosuppression (20). The immunosuppressive BMME in MM has been implicated in clonal evolution and immune evasion of MM cells accelerating disease progression (4, 20).

Expanded populations of MDSC, representing CD33⁺CD123⁺ immature myeloid cells within the bone marrow mononuclear cell fraction contribute to the immunosuppressive BMME by inhibiting both memory and cytotoxic effector T-cell populations as well as natural killer (NK) cells, thereby promoting the immune evasion of MM clones (4, 9–15, 20) (**Figure 1**). MDSCs along with MM cell

derived interleukin 10 (IL-10), TGF- β and IL-6 inhibit dendritic cell (DC) maturation and their antigen-presenting function, which further aggravates the immunosuppression (6). The abundance of MDSCs is associated with a higher risk of rapidly progressive disease and poor survival outcomes in MM (9–11). MDSCs are activated by exosomes and support the development of Tregs, promote angiogenesis and growth of MM cells besides inhibiting the immune effector cells (21–26).

Several strategies are being explored to overcome the immunosuppressive cellular elements of the BMME in MM patients, including the use autologous hematopoietic stem cell transplantation (AHSCT) (4, 20, 27–29) to remodel the BMME by establishing a more favorable ratio between effective MM-specific CTLs *versus* Tregs and other immunosuppressive cells. Treatment strategies aimed at further enhancing the anti-MM immunity can be employed as post-AHSCT interventions, including MM- or MDSC-directed monoclonal antibodies (MoAb) (20, 27, 28). New generation multi-parameter minimal residual leukemia (MRD) detection techniques provide a unique opportunity to evaluate the effect of new treatment modalities that are applied as part of AHSCT or post-AHSCT on the quality and length of complete remission in both newly diagnosed high-risk MM as well as RR MM (29, 30).



express CD38 and CD123 antigens on their surface.

DUAL TARGETING OF MM CELLS AND MDSCS

Dual targeting of MM cells and MDSCs using biotherapeutic agents has emerged as a very promising new therapeutic platform with a particularly high clinical impact potential. For example CD38 antigen is present on MM cells as well as MDSCs (4, 20, 31-34). Daratumumab, a complement-activating anti-CD38 MoAb capable of causing antibody-dependent cellular cytotoxicity (ADCC) and apoptosis in MM cells, showed significant single agent activity in relapsed MM patients and improved the survival outcome when used in combination with other active anti-MM agents such as bortezomib and dexamethasone or lenalidomide and dexamethasone (4, 20, 31-34). Similar results were obtained using alternative anti-CD38 MoAbs, such as Isatuximab (4, 20). Daratumumab has been shown to expand the immunoreactive CTL populations via depletion of CD38⁺ immunosuppressive cellular elements of the BMME (33). The immunomodulatory effects of Daratumumab improved the clinical responses of previously resistant MM patients to standard combination therapy (4, 20). Unfortunately, increased expression levels of complement inhibitors CD55 and CD59 as well as decreased cell surface expression levels of CD38 on MM cells may decrease the clinical activity of anti-CD38 antibodies (4, 35).

CLINICAL IMPACT POTENTIAL OF BISPECIFIC T-CELL ENGAGERS

BsABs capable of redirecting host T-cell cytotoxicity in an MHCindependent manner to malignant clones as well as immunosuppressive MDSCs (14–20, 35–39) are being explored as a new class of drug candidates in various hematologic malignancies (40). Bispecific CD3xBCMA antibodies targeting the B-cell maturation antigen (BCMA; CD269/TNFRS17) on MM cells, such as EM801, REGN5458 (NCT03761108) and AMG 420 (NCT03836053) showed single agent activity in relapsed/refractory MM patients (41-46) (Figure 2). CD3xBCMA BsABs, Elranatamab (PF-06863135) and Teclistamab are being evaluated in R/R MM patients (NCT04649359 and NCT03145181/NCT04557098). In the MajesTEC-1 Phase 1 study of the BCMAxCD3 BsAB Teclistamab in R/R MM (NCT03145181), both intravenous and subcutaneous (s.c) administration schedules were evaluated, and the recommended phase 2 dose was identified as 1.5 mg/kg administered s.c. once a week. At this dose level, the overall response rate was 65%. Grade 3-4 neutropenia was observed in 40% and Grade 1-2 CRS was observed in 70% of the patients (47). The second-generation CD3xBCMA BsAB AMG701 that has an Fc domain to extend its half-life (48) is being evaluated in an Phase 1/2 clinical study (NCT03287908). TNB-383B has been designed to reduce the risk of the class-specific AE cytokine release syndrome (CRS) (49). TeneoBio has shown that TNB-383B causes significantly lower cytokine release from activated T-cells. A clinical proof of concept study (Clinicaltrials.gov identifier: NCT0302577) was designed to study the effects of reducing the levels of γ -secretase cleaved soluble BCMA in MM patients by using a y-secretase inhibitor because soluble BCMA interferes with the mechanism of action of BCMA-targeting bispecific antibodies.

Bispecific CD3xCD38 antibodies have also been developed (50, 51) and entered clinical trials in patients with relapsed or refractory MM, such as AMG424 (NCT0344566) (51) and GBR1342 (NCT0330911). A CD38-reactive tri-specific antibody targeting CD3 and CD28 co-receptors on T-cells has also been developed to achieve augmented and sustained T-cell activation *via* CD28 engagement (52). A bispecific T-cell engaging CD3xCD307 antibody, named BFCR4350A, was developed targeting the FcRH5/CD307 antigen (53) on MM cells and it is currently being evaluated in a Phase 1 clinical trial (NCT03275103) (54).



Talquetamab is a GPRC5DxCD3 BsAB targeting the orphan G protein-coupled receptor GPRC5D that is abundantly expressed on MM cells. In a Phase 1 study in R/R MM patients testing both IV and SC administration schedules (NCT03399799), the RP2D was identified as 405 mcg/kg administered SC on a weekly basis (55). The RP2D level was well tolerated and exhibited promising activity with an overall response rate of 63%. CRS (79%), neutropenia (64%), anemia (57%) and dysgeusia (57%) were the most common treatment-emergent AEs. Furthermore, 7% of patients developed neurotoxicity and 32% developed infections. The overall response rate at the RP2D was 63% (55).

TARGETING CD123 ON MDSC

The α -chain of the IL-3 receptor, also known as the CD123 antigen, is abundantly expressed on MDSC (20). Furthermore, CD123 is also expressed on plasmacytoid dendritic cells (PDCs) that contribute to the growth of MM cells as well as cancer stem-like cells and osteoclast progenitors (56). Several biotherapeutic agents targeting CD123 have been developed, including the CD123-directed recombinant human IL3 fusion toxin Tagraxofusb (SL-401), MoAbs, BsABs targeting CD123 antigen, such as bispecific T-cell engagers (BiTEs), dual-affinity retargeting antibodies (DARTs), bispecific killer cell engagers, and tri-specific killer cell engagers (40, 57–59).

Targeting the BMME in MM with SL-401 has been shown to reduce the viability of PDCs and inhibit PDC-induced MM cell growth, impair the viability of CD123⁺ MM stem cells, and prevent osteoclastogenesis in preclinical model systems (60). SL-401 is being assessed in combination with standard of care in a clinical study (NCT02661022) in relapsed/refractory MM patients with promising early evidence of clinical activity (61, 62). Seroproteomics analysis of MM patient serum samples reportedly showed a reduction of PDC-derived soluble proteins in SL-401 treated patients (63).

CD123-targeting, CD3-engaging BsAB, such as Flotetuzumab (59) and APVO436 (64) bring cytotoxic T-cells (CTLs) within close vicinity of target CD123⁺ cells to create "cytolytic synapses" as a short bridge between target cells and CTLs, triggering CTL activation and destruction of targeted cells (Figures 2, 3). These dual-function anti-MM drug candidates are currently in clinical trials for treatment of CD123-expressing hematologic malignancies with early clinical proof of concept for their ability to destroy CD123 + malignant clones, including CRs in relapsed or refractory AML patients (NCT02152956, NCT03647800). However, the clinical potential of a CD123xCD3 bispecific antibody in MM therapy may be limited as the bulk population of MM cells lack CD123 and depletion of CD123⁺ MM stem cells alone is unlikely to be an effective strategy for monotherapy. Therefore, clinical feasibility and efficacy studies of combinations of CD123 targeting BsAB with active anti-MM drugs such as pomalidomide that appeared to have augmented activity in the presence of the anti-CD123 fusion toxin tagraxofusp (3, 4, 8, 60), biotherapeutic agents, such as CD3xBCMA BsABs, daratumumab, elotuzumab (4-7, 20), and CAR-T cells (4, 65) are needed to gain insights into the clinical impact potential of CD123xCD3 BsABs.

Notably, MDSCs have been shown to significantly suppress the CTL engaging activity of a BCMAxCD3 BsAB, but the MDSCsuppressed CTL activity could be restored by addition of a hypomethylating agent (HMA) capable of epigenetically altering the MDSC transcriptome *via* reversal of the aberrant DNA methylation (66). Therefore, MDSC-targeting BsAB could be potentiated by HMAs. It is noteworthy that a combination of the CD123xCD3 BsAB APVO436 with azacitidine is being evaluated in one of the cohorts in the ongoing expansion phase of a Phase 1B AML study (NCT03647800).

CYTOKINE RELEASE SYNDROME (CRS)

CD3-engaging BsABs act as agonists and activate T-cells in the presence of tumor cells expressing the target tumor-associated antigen, which can lead to excessive T-cell activation with release of inflammatory cytokines and development of the potentially lifethreatening systemic inflammation, known as cytokine release syndrome (CRS) (67-71). For example, BsAB AMG330 binds CD33 antigen on AML cells as well as MDSCs cells and CD3E on T-cells. In an open-label Phase 1 study (Clinicaltrial.gov identifier: NCT#02520427), AMG330 was given at doses ranging from 0.5- $720\,\mu$ g/d in the manner of continuous IV infusion among 55 patients with R/R AML (NCT02520427). AMG 330-related AEs included CRS (67%; Grade \geq 3 in 13%) as the most frequent AEs (72). Similarly, CRS was observed in 63% of AML patients treated with AMG673, a new version of AMG330 (Grade ≥3 in 18%; Clinicaltrial.gov identifier: NCT03224819) (72). Flotetuzumab (MGD006) is a bispecific, dual-affinity re-targeting (DART) antibody reactive with both CD3 antigen on T-cells and CD123 antigen on AML cells and MDSCs. This CD3 engaging bispecific antibody exhibited promising single agent activity in therapyrefractory AML patients with primary induction failure as well as patients with an early first relapse. CRS was observed in all AML patients treated with Flotetuzumab (73) and 58% of AML patients treated with Vibecotamab (XmAb14045), another CD3xCD123 BsAB (74). By comparison, only 10 of 46 patients (21.7%) treated with the CD3xCD123 BsAB APVO436 developed CRS (64).

IL-6 is one of the driving pro-inflammatory cytokines that contribute CRS and its pulmonary, cardiovascular, renal, and neurologic complications (60, 64-71). Cytokine profiling in patients who developed CRS after APVO436 infusion indicates that the predominant cytokine in this inflammatory cytokine response is IL-6, which agrees with our current knowledge regarding CRS that occurs in the context of BsAB therapy (20, 60, 64-71, 75). Within 1-2 days following the first dose of APVO436, the mean serum IL-6 concentration in these patients who developed CRS was elevated 145-fold over baseline (755 vs 5.2) and at the end of one week it was still elevated 83-fold over baseline. In most cases, CRS events were transient and medically manageable with standard of care including the use of dexamethasone and anti-IL-6:IL-6R antibody Tocilizumab or anti-IL-6 antibody Siltuximab (antibody against IL-6). However, CRS can be life-threatening even with the use of Tocilizumab or Siltuximab (60, 64-71). Therefore, development of consistently effective prevention and treatment regimens against CRS



remains an urgent and unmet medical need. Identification of such regimens would further advance the field of immunotherapy. We recently reported the robust anti-inflammatory activity of RJX in animal models of CRS (76). RJX has been shown to block the production of IL-6, TNF- α , as well as TGF- β and reverse inflammation-induced tissue injury and multi-organ damage in mouse models of sepsis and CRS (76). RJX is currently being evaluated for its ability to prevent COVID-19 associated CRS in a double-blind randomized clinical study (NCT04708340). Because of its safety and easy use, RJX may emerge as an attractive adjunct to BsAB platforms to mitigate the risk of severe CRS.

NEUROTOXICITY

Neurotoxicity is a treatment-emergent adverse event (AE) for BsABs, and it is often associated with CRS (77). The signs and symptoms vary from patient to patient and include headache, tremor, confusion, expressive and nominal dysphasia, impaired attention, apraxia, and lethargy occurring as early and common manifestations (77–79). Consensus grading criteria were developed by the ASTCT based on the use of the Immune Effector Cell-Associated Encephalopathy (ICE) screening tool (78). The CD19xCD3 BsAB blinatumumab has been reported to cause neurotoxicity in 70% of patients with B-lineage non-Hodgkin's lymphomas (NHL). By comparison, it is less common with CD20xCD3 or CD123xCD3 BsABs (80). In a recent Phase 1 dose escalation study of the CD123xCD3 BsAB APVO436, APVO436-related transient neurotoxicity occurred only in 5 of 46 patients (10.9%). It occurred during the first cycle in 4 of the 5 patients and in Cycle 8 in the remaining patient. It was mild with Grade 1 AEs including headache, tremor, dizziness, lethargy, insomnia, memory loss, and confusion (64). A single case of Grade 3 confusion was encountered on the first day of treatment and resolved within a day. Neurotoxicity did not show any dose-dependence. Gender, race, age, absolute lymphocyte count or percentage of lymphocytes in peripheral blood did not predict neurotoxicity. Neurotoxicity occurred in 3 patients who also experienced CRS and in 2 patients who did not develop CRS (75). Conversely, of 10 patients who developed CRS, 7 did not experience any neurotoxicity (75).

CONCLUSION

Recombinant T-cell engaging humanized BsABs redirect host T-cell cytotoxicity in an target antigen-expressing cells in patients with hematologic malignancies. They can be used both for targeting drug-resistant MM clones as well as the immune-suppressive cell populations in the BMME (**Figure 3**). Dual targeting of drug-resistant MM clones and immunosuppressive MDSC has the potential to change the therapeutic landscape for MM and improve the survival outcomes of high-risk as well as relapsed/refractory MM patients. The definition of optimal strategies for overcoming the

immunosuppressive BMME in MM may require randomized clinical studies with parallel cohorts and adaptive trial designs. CD3xCD123 BsAB have clinical impact potential in MM as they may help treatment outcomes by blocking immune evasion *via* depletion of CD123⁺ MDSC and by reducing the drug-resistant tumor load *via* CTL-mediated MHC-independent destruction of MM stem cells.

REFERENCES

- Schürch CM, Rasche L, Frauenfeld L, Weinhold N, Fend F. A Review on Tumor Heterogeneity and Evolution in Multiple Myeloma: Pathological, Radiological, Molecular Genetics, and Clinical Integration. *Virchows Arch* (2020) 476:337–51. doi: 10.1007/s00428-019-02725-3
- Holthof LC, Mutis T. Challenges for Immunotherapy in Multiple Myeloma: Bone Marrow Microenvironment-Mediated Immune Suppression and Immune Resistance. *Cancers* (2020) 12(4):988. doi: 10.3390/cancers12040988
- Lee JH, Kim SH. Treatment of Relapsed and Refractory Multiple Myeloma. Blood Res (2020) 55(S1):S43–53. doi: 10.5045/br.2020.S008
- Uckun FM, Qazi S, Demirer T, Champlin RE. Contemporary Patient-Tailored Treatment Strategies Against High Risk and Relapsed or Refractory Multiple Myeloma. *EBioMedicine* (2019) 39:612–20. doi: 10.1016/j.ebiom.2018.12.004
- Soekojo CY, Ooi M, de Mel S, Chang WJ. Immunotherapy in Multiple Myeloma. *Cells* (2020) 9(3):601. doi: 10.3390/cells9030601
- Yamamoto L, Amodio N, Gulla A, Anderson KC. Harnessing the Immune System Against Multiple Myeloma: Challenges and Opportunities. Front Oncol (2021) 10:606368. doi: 10.3389/fonc.2020.606368
- Minnie SA, Hill GR. Immunotherapy of Multiple Myeloma. J Clin Invest (2020) 130(4):1565–75. doi: 10.1172/JCI129205
- Gulla A, Anderson KC. Multiple Myeloma: The (R)Evolution of Current Therapy and a Glance Into Future. *Haematologica* (2020) 105(10):2358–67. doi: 10.3324/haematol.2020.247015
- Kawano Y, Moschetta M, ManierS, Glaveys, Gorgun GT, Roccaro AM, Anderson KC, Ghobrial IM. Targeting the Bone Marrow Microenvironment in Multiple Myeloma. *Immunol Rev* (2015) 263:160–72. doi: 10.1111/imr.12233
- García-Ortiz A, Rodríguez-García Y, Encinas J, Maroto-Martín E, Castellano E, Teixidó J, et al. The Role of Tumor Microenvironment in Multiple Myeloma Development and Progression. *Cancers* (2021) 13:217. doi: 10. 3390/cancers13020217
- Ostrand-Rosenberg S, Sinha P, Beury DW, Clements VK. Cross-Talk Between Myeloid-Derived Suppressor Cells (MDSC), Macrophages, and Dendritic Cells Enhances Tumor-Induced Immune Suppression. *Semin Cancer Biol* (2012) 22(4):275–81. doi: 10.1016/j.semcancer.2012.01.011
- Urashima M, Ogata A, Chauhan D, Hatziyanni M, Vidriales MB, Dedera DA, et al. Transforming Growth Factor-Beta1: Differential Effects on Multiple Myeloma Versus Normal B Cells. Blood (1996) 87:1928–38. doi: 10.1182/ blood.V87.5.1928.1928
- Nakamura K, Kassem S, Cleynen A, Chretien ML, Guillerey C, Putz EM, et al. Dysregulated IL-18 Is a Key Driver of Immunosuppression and a Possible Therapeutic Target in the Multiple Myeloma Microenvironment. *Cancer Cell* (2018) 33:634–48.e635. doi: 10.1016/j.ccell.2018.02.007
- Domagala M, Laplagne C, Leveque E, Laurent C, Fournié J-J, Espinosa E, et al. Cancer Cells Resistance Shaping by Tumor Infiltrating Myeloid Cells. *Cancers* (2021) 13(2):165. doi: 10.3390/cancers13020165
- Uhel F, Azzaoui I, Le Gallou S, Fest T, Tarte K, Roussel M. Early-Stage Myeloid-Derived Suppressor Cell Count: Basophil Exclusion Matters. J Allergy Clin Immunol (2019) 144(4):1125–7. doi: 10.1016/j.jaci.2019.06.027
- Khan ANH, Emmons TR, Wong JT, Alqassim E, Singel KL, Mark J, et al. Quantification of Early-Stage Myeloid-Derived Suppressor Cells in Cancer Requires Excluding Basophils. *Cancer Immunol Res* (2020) 8(6):819–28. doi: 10.1158/2326-6066.CIR-19-0556
- Dysthe M, Parihar R. Myeloid-Derived Suppressor Cells in the Tumor Microenvironment. Adv Exp Med Biol (2020) 1224:117–40. doi: 10.1007/ 978-3-030-35723-8_8
- Gustafson MP, Lin Y, Maas ML, Van Keulen VP, Johnston PB, Peikert T, et al. A Method for Identification and Analysis of Non-Overlapping Myeloid

AUTHOR CONTRIBUTIONS

FMU conceived the review, analyzed the contents of relevant publications, wrote the original draft of the manuscript revised the manuscript, provided final approval for submission of the final version. No medical writer or editor was involved.

Immunophenotypes in Humans. PloS One (2015) 10(3):e0121546. doi: 10.1371/journal.pone.0121546

- Edwards JV, Zhang H, Mackall CL. Abstract 3818: CD11c+CD123+ Human Myeloid Dendritic Cells Generated Using IL4, IL13 and IL3 Show Characteristics of Myeloid Derived Suppressor Cells. In: Proceedings of the 101st Annual Meeting of the American Association for Cancer Research; 2010 Apr 17-21; Washington, Dc. *Philadelphia (PA): AACR; Cancer Res* (2010) 70 (8 Suppl):Abstract nr 3818. doi: 10.1158/1538-7445.AM10-3818
- Uckun FM. Overcoming the Immunosuppressive Tumor Microenvironment in Multiple Myeloma. *Cancers (Basel)* (2021) 13(9):2018. doi: 10.3390/ cancers13092018
- Chen T, Moscvin M, Bianchi G. Exosomes in the Pathogenesis and Treatment of Multiple Myeloma in the Context of the Bone Marrow Microenvironment. *Front Oncol* (2020) 10:608815. doi: 10.3389/fonc.2020.608815
- Moloudizargari M, Abdollahi M, Asghari MH, Zimta AA, Neagoe IB, Nabavi SM. The Emerging Role of Exosomes in Multiple Myeloma. *Blood Rev* (2019) 38:100595. doi: 10.1016/j.blre.2019.100595
- Li M, Xia B, Wang Y, James You M, Zhang Y. Potential Therapeutic Roles of Exosomes in Multiple Myeloma: A Systematic Review. J Cancer (2019) 10 (24):6154–60. doi: 10.7150/jca.31752
- Raimondo S, Urz O, Conigliaro A, Bosco L, Parisi S, Carlisi M, et al. Extracellular Vesicle microRNAs Contribute to the OsteogenicInhibition of Mesenchymal Stem Cells in Multiple Myeloma. *Cancers (Basel)* (2020) 12 (2):449. doi: 10.3390/cancers12020449
- Raimondi L, De Luca A, Fontana S, Amodio N, Costa V, Carina V, et al. Multiple Myeloma-Derived Extracellular Vesicles Induce Osteoclastogenesis Through the Activation of the XBP1/IRE1α Axis. *Cancers (Basel)* (2020) 12 (8):2167. doi: 10.3390/cancers12082167
- 26. Wang J, De Veirman K, Faict S, Frassanito MA, Ribatti D, Vacca A, et al. Multiple Myeloma Exosomes Establish a Favourable Bone Marrow Microenvironment With Enhanced Angiogenesis and Immunosuppression. *J Pathol* (2016) 239:162–73. doi: 10.1002/path.4712
- Al Hamed R, Bazarbachi AH, Malard F, Harousseau JL, Mohty M. Current Status of Autologous Stem Cell Transplantation for Multiple Myeloma. *Blood Cancer J* (2019) 9(4):44. doi: 10.1038/s41408-019-0205-9
- Jagannath S, Abonour R, Durie BGM, Narang M, Terebelo HR, Gasparetto CJ, et al. Impact of Post-ASCT Maintenance Therapy on Outcomes in Patients With Newly Diagnosed Multiple Myeloma in Connect Mm. *Blood Adv* (2018) 2(13):1608–15. doi: 10.1182/bloodadvances.2018017186
- Hu B, Thall P, Milton DR, Sasaki K, Bashir Q, Shah N, et al. High-Risk Myeloma and Minimal Residual Disease Postautologous-HSCT Predict Worse Outcomes. *Leuk Lymphoma* (2018) 60(2):442–52. doi: 10.1080/ 10428194.2018.1485908
- Caers J, Garderet L, Kortüm KM, O'Dwyer ME, van de Donk NWCJ, Binder M, et al. An European Myeloma Network Recommendation on Tools for Diagnosis and Monitoring of Multiple Myeloma: What to Use and When. *Haematologica* (2018) 103(11):1772–84. doi: 10.3324/haematol.2018.189159. pii: haematol.2018.189159.
- 31. Giri S, Grimshaw A, Bal S, Godby K, Kharel P, Djulbegovic B, et al. Evaluation of Daratumumab for the Treatment of Multiple Myeloma in Patients With High-Risk Cytogenetic Factors: A Systematic Review and Meta-Analysis. *JAMA Oncol* (2020) 6(11):1759–65. doi: 10.1001/jamaoncol.2020.4338
- 32. Voorhees PM, Kaufman JL, Laubach JP, Sborov DW, Reeves B, Rodriguez C, et al. Daratumumab, Lenalidomide, Bortezomib, and Dexamethasone for Transplant-Eligible Newly Diagnosed Multiple Myeloma: GRIFFIN. *Blood* (2020) 136(8):936–45. doi: 10.1182/blood.2020005288
- Krejcik J, Casneuf T, Nijhof IS, Verbist B, Bald J, Plesner T, et al. Daratumumab Depletes CD38+ Immune Regulatory Cells, Promotes T-Cell

Expansion, and Skews T-Cell Repertoire in Multiple Myeloma. *Blood* (2016) 128:384–94. doi: 10.1182/blood-2015-12-687749

- 34. Adams HC 3rd, Stevenaert F, Krejcik J, Van der Borght K, Smets T, Bald J, et al. High-Parameter Mass Cytometry Evaluation of Relapsed/Refractory Multiple Myeloma Patients Treated With Daratumumab Demonstrates Immune Modulation as a Novel Mechanism of Action. *Cytometry A* (2019) 95(3):279–89. doi: 10.1002/cyto.a.23693
- van de Donk NWCJ, Usmani SZ. CD38 Antibodies in Multiple Myeloma: Mechanisms of Action and Modes of Resistance. *Front Immunol* (2018) 9:2134. doi: 10.3389/fimmu.2018.02134
- Parker KH, Beury DW, Ostrand-Rosenberg S. Myeloid-Derived Suppressor Cells: Critical Cells Driving Immune Suppression in the Tumor Microenvironment. Adv Cancer Res (2015) 128:95–139. doi: 10.1016/ bs.acr.2015.04.002
- Movahedi K, Guilliams M, Van den Bossche J, Van den Bergh R, Gysemans C, Beschin A, et al. Identification of Discrete Tumor-Induced Myeloid-Derived Suppressor Cell Subpopulations With Distinct T Cell-Suppressive Activity. *Blood* (2008) 111(8):4233–44. doi: 10.1182/blood-2007-07-099226
- Marvel D, Gabrilovich DI. Myeloid-Derived Suppressor Cells in the Tumor Microenvironment: Expect the Unexpected. J Clin Invest (2015) 125(9):3356– 64. doi: 10.1172/JCI80005
- Younos IH, Abe F, . Talmadge JE. Myeloid-Derived Suppressor Cells: Their Role in the Pathophysiology of Hematologic Malignancies and Potential as Therapeutic Targets. *Leukemia Lymphoma* (2015) 56(8):2251-63. doi: 10.3109/10428194.2014.987141
- Einsele H, Borghaei H, Orlowski RZ, Subklewe M, Roboz GJ, Zugmaier G, et al. The BiTE (Bispecific T-Cell Engager) Platform: Development and Future Potential of a Targeted Immuno-Oncology Therapy Across Tumor Types. *Cancer* (2020) 126:3192–201. doi: 10.1002/cncr.32909
- Topp MS, Duell J, Zugmaier G, Attal M, Moreau P, Langer C, et al. Anti-B-Cell Maturation Antigen BiTE Molecule AMG 420 Induces Responses in Multiple Myeloma. J Clin Oncol (2020) 38(8):775–83. doi: 10.1200/ JCO.19.02657
- 42. Topp MS, Duell J, Zugmaier G, Attal M, Moreau P, Langer C, et al. Treatment With AMG 420, an Anti-B-Cell Maturation Antigen (BCMA) Bispecific T-Cell Engager (BiTE[®]) Antibody Construct, Induces Minimal Residual Disease (MRD) Negative Complete Responses in Relapsed and/or Refractory (R/R) Multiple Myeloma (MM) Patients: Results of a First-in-Human (FIH) Phase I Dose Escalation Study. *Blood* (2018) 132(Suppl 1):1010. doi: 10.1182/blood-2018-99-109769
- Gavriatopoulou M, Ntanasis-Stathopoulos I, Dimopoulos MA, Terpos E. Anti-BCMA Antibodies in the Future Management of Multiple Myeloma. Expert Rev Anticancer Ther (2019) 19:319-26. doi: 10.1080/ 14737140.2019.1586539
- 44. Shah N, Chari A, Scott E, Mezzi K, Usmani SZ. B-Cell Maturation Antigen (BCMA) in Multiple Myeloma: Rationale for Targeting and Current Therapeutic Approaches. *Leukemia* (2020) 34:985–1005. doi: 10.1038/ s41375-020-0734-z
- Madduri D, Roske A, Brayer J, Zonder J, Bensinger WI, Li J, et al. REGN5458, a BCMA X CD3 Bispecific Monoclonal Antibody, Induces Deep and Durable Responses in Patients With Relapsed/Refractory Multiple Myeloma (RRMM). Presented at: 2020 ASH Annual Meeting & Exposition. *Blood* (2020) 136 (Suppl 1):41–2. doi: 10.1182/blood-2020-139192
- 46. Seckinger A, Delgado JA, Moser S, Moreno L, Neuber B, Grab A, et al. Target Expression, Generation, Preclinical Activity, and Pharmacokinetics of the BCMA-T Cell Bispecific Antibody EM801 for Multiple Myeloma Treatment. *Cancer Cell* (2017) 31(3):396–410. doi: 10.1016/j.ccell.2017.02.002
- Usmani SZ, Garfall AL, van de Donk NWCJ, Nahi H, San-Miguel JF, Oriol A, et al. Teclistamab, a B-Cell Maturation Antigen × CD3 Bispecific Antibody, in Patients With Relapsed or Refractory Multiple Myeloma (MajesTEC-1): A Multicentre, Open-Label, Single-Arm, Phase 1 Study. *Lancet* (2021) 398(10301):665–74. doi: 10.1016/S0140-6736(21)01338-6
- Goldstein RL, Goyos A, Li CM, Deegen P, Bogner P, Sternjak A, et al. AMG 701 Induces Cytotoxicity of Multiple Myeloma Cells and Depletes Plasma Cells in Cynomolgus Monkeys. *Blood Adv* (2020) 4:4180–94. doi: 10.1182/ bloodadvances.2020002565
- 49. Suurs FV, Lub-de Hooge MN, de Vries EGE, de Groot DJA. A Review of Bispecific Antibodies and Antibody Constructs in Oncology and Clinical

Challenges. Pharm Ther (2019) 201:103-19. doi: 10.1016/j.pharmthera. 2019.04.006

- Fayon M, Martinez-Cingolani C, Abecassis A, Roders N, Nelson E, Choisy C, et al. Bi38-3 Is a Novel CD38/CD3 Bispecific T-Cell Engager With Low Toxicity for the Treatment of Multiple Myeloma. *Haematologica* (2021) 106 (4):1193–7. doi: 10.3324/haematol.2019.242453
- Zuch de Zafra CL, Fajardo F, Zhong W, Bernett MJ, Muchhal US, Moore GL, et al. Targeting Multiple Myeloma With AMG 424, a Novel Anti-CD38/CD3 Bispecific T-Cellrecruiting Antibody Optimized for Cytotoxicity and Cytokine Release. *Clin Cancer Res* (2019) 25(13):3921–33. doi: 10.1158/1078-0432.CCR-18-2752
- Wu L, Seung E, Xu L, Rao E, Lord DM, Wei RR, et al. Trispecific Antibodies Enhance the Therapeutic Efficacy of Tumor-Directed T Cells Through T Cell Receptor Co-Stimulation. *Nat Cancer* (2020) 1:86–98. doi: 10.1038/s43018-019-0004-z
- 53. Ise T, Nagata S, Kreitman RJ, Wilson WH, Wayne AS, Stetler-Stevenson M, et al. Elevation of Soluble CD307 (IRTA2/FcRH5) Protein in the Blood and Expression on Malignant Cells of Patients With Multiple Myeloma, Chronic Lymphocytic Leukemia, and Mantle Cell Lymphoma. *Leukemia* (2007) 21 (1):169–74. doi: 10.1038/sj.leu.2404445
- Alhallak K, Sun J, Jeske A, Park C, Yavner J, Bash H, et al. Bispecific T Cell Engagers for the Treatment of Multiple Myeloma: Achievements and Challenges. *Cancers* (2021) 13:2853. doi: 10.3390/cancers13122853
- 55. Berdeja JG, Krishnan AY, Oriol A, Van deDonk N, Rodríguez-Otero P, Askari E, et al. Updated Results of a Phase 1, First-in-Human Study of Talquetamab, a G Protein-Coupled Receptor Family C Group 5 Member D (GPRC5D) × CD3 Bispecific Antibody, in Relapsed/Refractory Multiple Myeloma (MM). J Clin Oncol (2021) 39(15_suppl):8008–8. doi: 10.1200/JCO.2021.39. 15_suppl.8008
- Chauhan D, Singh AV, Brahmandam M, Carrasco R, Bandi M, Hideshima T, et al. Functional Interaction of Plasmacytoid Dendritic Cells With Multiple Myeloma Cells: A Therapeutic Target. *Cancer Cell* (2009) 16:309–23. doi: 10.1016/j.ccr.2009.08.019
- Testa U, Pelosi E, Frankel A. CD 123 Is a Membrane Biomarker and a Therapeutic Target in Hematologic Malignancies. *Biomark Res* (2014) 2(1):4. doi: 10.1186/2050-7771-2-4
- Kovtun Y, Jones GE, Adams S, Harvey L, Audette CA, Wilhelm A, et al. A CD123-Targeting Antibody-Drug Conjugate, IMGN632, Designed to Eradicate AML While Sparing Normal Bone Marrow Cells. *Blood Adv* (2018) 2:848–58. doi: 10.1182/bloodadvances.2018017517
- Uy GL, Aldoss I, Foster MC, Sayre PH, Wieduwilt MJ, Advani AS, et al. Flotetuzumab as Salvage Immunotherapy for Refractory Acute Myeloid Leukemia. Blood (2021) 137(6):751–62. doi: 10.1182/blood.2020007732
- Ray A, Das DS, Song Y, Macri V, Richardson P, Brooks CL, et al. A Novel Agent SL-401 Induces Anti-Myeloma Activity by Targeting Plasmocytoid Dendritic Cells, Osteoclastogenesis and Cancer Stem-Like Cells. *Leukemia* (2017) 31:2652–60. doi: 10.1038/leu.2017.135
- Htut M, Gasparetto C, Zonder J, Matin T.G. III, Scott EC, Chen J, et al. Results From Ongoing Phase 1/2 Trial of SL-401 in Combination With Pomalidomide and Dexamethasone in Relapsed or Refractory Multiple Myeloma. *Blood* (2016) 128:5696. doi: 10.1182/blood.V128.22.5696.5696
- Alkharabsheh O, Frankel AE. Clinical Activity and Tolerability of SL-401 (Tagraxofusp): Recombinant Diphtheria Toxin and Interleukin-3 in Hematologic Malignancies. *Biomed* (2019) 7(1):6. doi: 10.3390/ biomedicines7010006
- 63. Chauhan D, Ray A, Song Y, Olguin A, Chen J, Brooks C, et al. Biological Correlates to Support a Clinical Role for Tagraxofusp, a Novel Targeted Therapy Directed to CD123, in Combination With Pomalidomide and Dexamethasone, to Target Plasmacytoid Dendritic Cells in Poor-Risk Patients With Multiple Myeloma. J Clin Oncol (2020) 38(15_suppl):e20537– 7. doi: 10.1200/JCO.2020.38.15_suppl.e20537
- 64. Uckun FM, Lin TL, Mims AS, Patel P, Lee C, Shahidzadeh A, et al. A Clinical Phase 1b Study of the CD3xCD123 Bispecific Anti-Body APVO436 in Patients With Relapsed/Refractory Acute Myeloid Leukemia or Myelodysplastic Syndrome. *Cancers* (2021) 13:4113. doi: 10.3390/cancers
- 65. Berdeja JG, Madduri D, Usmani SZ, Jakubowiak A, Agha M, Cohen AD, et al. Ciltacabtagene Autoleucel, a B-Cell Maturation Antigen-Directed Chimeric Antigen Receptor T-Cell Therapy in Patients With Relapsed or Refractory

Multiple Myeloma (CARTITUDE-1): A Phase 1b/2 Open-Label Study. Lancet (2021) 398(10297):314–24. doi: 10.1016/S0140-6736(21)00933-8

- 66. Perez C, Botta C, Zabaleta A, Puig N, Cedena MT, Goicoechea I, et al. Immunogenomic Identification and Characterization of Granulocytic Myeloid-Derived Suppressor Cells in Multiple Myeloma. *Blood* (2020) 136 (2):199–209. doi: 10.1182/blood.2019004537
- Hay K, Hanafi L, Li D, Gust J, Liles WC, Wurfel MM, et al. Kinetics and Biomarkers of Severe Cytokine Release Syndrome After CD19 Chimeric Antigen Receptor-Modified T-Cell Therapy. *Blood* (2017) 130(21):2295– 306. doi: 10.1182/blood-2017-06-793141
- Aldoss I, Khaled SK, Budde E, Stein AS. Cytokine Release Syndrome With the Novel Treatments of Acute Lymphoblastic Leukemia: Pathophysiology, Prevention, and Treatment. *Curr Oncol Rep* (2019) 21(1):4. doi: 10.1007/ s11912-019-0753-y
- Fajgenbaum DC, June CH. Cytokine Storm. N Engl J Med (2020) 383 (23):2255–73. doi: 10.1056/NEJMra2026131
- Lee D, Gardner R, Porter D, Louis CU, Ahmed N, Jensen M, et al. Current Concepts in the Diagnosis and Management of Cytokine Release Syndrome. *Blood* (2014) 124(2):188–95. doi: 10.1182/blood-2014-05-552729
- Morris EC, Neelapu SS, Giavridis T, Sadelain M. Cytokine Release Syndrome and Associated Neurotoxicity in Cancer Immunotherapy. *Nat Rev Immunol* (2021) 17:1–12. doi: 10.1038/s41577-021-00547-6
- Daver N, Alotaibi AS, Bücklein V, Subklewe M. T-Cell-Based Immunotherapy of Acute Myeloid Leukemia: Current Concepts and Future Developments. *Leukemia* (2021) 35(7):1843–63. doi: 10.1038/s41375-021-01253-x
- Rettig MP, Muth J, Curtis T, Fehr E, Guo K, Zhao J, et al. Flotetuzumab as Salvage Immunotherapy for Refractory Acute Myeloid Leukemia. *Blood* (2021) 137(6):751–62. doi: 10.1182/blood.2020007732
- 74. Ravandi F, Bashey A, Stock W, Foran JM, Mawad R, Egan D, et al. Complete Responses in Relapsed/Refractory Acute Myeloid Leukemia (Aml) Patients on a Weekly Dosing Schedule of Vibecotamab (XmAb14045), a CD123 X CD3 T Cell-Engaging Bispecific Antibody; Initial Results of a Phase 1 Study. *Blood* (2020) 136:4–5. doi: 10.1182/blood-2020-134746
- 75. Uckun FM, Watts J, Mims AS, Patel P, Wang E, Shami PJ, et al. Risk, Characteristics and Biomarkers of Cytokine Release Syndrome in Patients with Relapsed/Refractory AML or MDS Treated with CD3xCD123 Bispecific Antibody APVO436. *Cancers* (2021) 13(21):5287. doi: 10.3390/ cancers13215287

- 76. Uckun FM, Carlson J, Orhan C, Powell J, Pizzimenti NM, Van Wyk H, et al. Rejuveinix Shows a Favorable Clinical Safety Profile in Human Subjects and Exhibits Potent Preclinical Protective Activity in the Lipopolysaccharide-Galactosamine Mouse Model of Acute Respiratory Distress Syndrome and Multi-Organ Failure. *Front Pharmacol* (2020) 11:594321. doi: 10.3389/ fphar.2020.594321
- Salvaris R, Ong J, Gregory GP. Bispecific Antibodies: A Review of Development, Clinical Efficacy and Toxicity in B-Cell Lymphomas. J Pers Med (2021) 11:355. doi: 10.3390/jpm11050355
- Lee DW, Santomasso BD, Locke FL, Ghobadi A, Turtle CJ, Brudno JN, et al. ASTCT Consensus Grading for Cytokine Release Syndrome and Neurologic Toxicity Associated With Immune Effector Cells. Biol. Blood Marrow Transplant (2019) 25:625–38. doi: 10.1016/j.bbmt.2018.12.758
- Santomasso BD, Park JH, Salloum D, Riviere I, Flynn J, Mead E, et al. Clinical and Biological Correlates of Neurotoxicity Associated With CAR T-Cell Therapy in Patients With B-Cell Acute Lymphoblastic Leukemia. *Cancer Discov* (2018) 8:958–71. doi: 10.1158/2159-8290.CD-17-1319
- Phillips TJ, Olszewski AJ, Munoz J, Kim TM, Yoon DH, Greil R, et al. Mosunetuzumab, a Novel CD20/CD3 Bispecific Antibody, in Combination With CHOP Confers High Response Rates in Patients With Diffuse Large B-Cell Lymphoma. *Blood* (2020) 136:37–8. doi: 10.1182/blood-2020-136295

Conflict of Interest: FU was employed by Ares Pharmaceuticals, and he was a consultant for Aptevo Therapeutics and for Reven Pharmaceuticals.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Uckun. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.