



Amide Proton Transfer Weighted and Intravoxel Incoherent Motion Imaging in Evaluation of Prognostic Factors for Rectal Adenocarcinoma

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Objectives: To analyze the value of amide proton transfer (APT) weighted and intravoxel incoherent motion (IVIM) imaging in evaluation of prognostic factors for rectal adenocarcinoma, compared with diffusion weighted imaging (DWI).

Materials and Methods: Preoperative pelvic MRI data of 110 patients with surgical pathologically confirmed diagnosis of rectal adenocarcinoma were retrospectively evaluated. All patients underwent high-resolution T₂-weighted imaging (T₂WI), APT, IVIM, and DWI. Parameters including APT signal intensity (APT SI), pure diffusion coefficient (D), pseudo-diffusion coefficient (D*), perfusion fraction (f), and apparent diffusion coefficient (ADC) were measured in different histopathologic types, grades, stages, and structure invasion statuses. Receiver operating characteristic (ROC) curves were used to evaluate the diagnostic efficacy, and the corresponding area under the curves (AUCs) were calculated.

Results: APT SI, D and ADC values of rectal mucinous adenocarcinoma (MC) were significantly higher than those of rectal common adenocarcinoma (AC) ([3.192 ± 0.661%] vs. [2.333 ± 0.471%], [1.153 ± 0.238 × 10⁻³ mm²/s] vs. [0.792 ± 0.173 × 10⁻³ mm²/s], and [1.535 ± 0.203 × 10⁻³ mm²/s] vs. [0.986 ± 0.124 × 10⁻³ mm²/s], respectively; all *P* < 0.001). In AC group, the APT SI and D values showed significant differences between low- and high-grade tumors ([2.226 ± 0.347%] vs. [2.668 ± 0.638%], and [0.842 ± 0.148 × 10⁻³ mm²/s] vs. [0.777 ± 0.178 × 10⁻³ mm²/s], respectively, both *P* < 0.05). The D value had significant difference between positive and negative extramural vascular invasion (EMVI) tumors ([0.771 ± 0.175 × 10⁻³ mm²/s] vs. [0.858 ± 0.151 × 10⁻³ mm²/s], *P* < 0.05). No significant difference of APT SI, D, D*, f or ADC was observed in different T stages, N stages, perineural and lymphovascular invasions (all *P* > 0.05). The ROC curves showed that the AUCs of APT SI, D and ADC values for distinguishing MC from AC were 0.921, 0.893 and 0.995, respectively. The AUCs of APT SI and D values in distinguishing low- from high-grade AC were 0.737 and 0.663, respectively. The AUC of the D value for evaluating EMVI involvement was 0.646.

Conclusion: APT and IVIM were helpful to assess the prognostic factors related to rectal adenocarcinoma, including histopathological type, tumor grade and the EMVI status.

Keywords: APT, IVIM, rectal neoplasms, adenocarcinoma, magnetic resonance imaging

INTRODUCTION

Colorectal cancer is a common malignancy of the digestive system, 30-35% are occurred in the rectum, and 90% are classified as adenocarcinoma (1, 2). Many factors are associated with therapeutic schedule and prognosis of rectal cancer, including tumor location, histological type, tumor grade, T stage, N stage, and related imaging indicators based on MRI, such as circumferential resection margin (CRM), and extramural vascular invasion (EMVI) statuses (3, 4). Rectal mucinous adenocarcinoma (MC) is a common subtype of rectal adenocarcinomas, which has a poor prognosis, and it is not sensitive to neoadjuvant chemoradiotherapy (5, 6). The selection of individualized treatment options for rectal cancer is based on accurate imaging evaluation.

Magnetic resonance imaging (MRI) is the most accurate test for preoperative assessment of rectal cancer. Conventional high-resolution MRI imaging, especially the small-field-of-view and thin-layer T₂ weighted imaging (T₂WI), not only clearly distinguishes the various layers of the rectal wall, but also displays the mesorectal fascia and EMVI (7, 8). Functional MRI has become increasingly widespread in recent years. Diffusion weighted imaging (DWI) is an example of functional MRI that reflects changes in tissue microenvironments by measuring the diffusion of water molecules in tissues. It has been applied in tumor TN stage, grading, and prognosis of rectal cancer in previous studies. However, the results were lack of consistency (9). Zhu et al. found the ADC values of low-grade adenocarcinoma were higher than those of high-grade adenocarcinoma, but the difference was not statistically significant (10). Several new MRI techniques have been used to evaluate the pathological features of rectal cancer, including intravoxel incoherent motion (IVIM), diffusion kurtosis imaging (DKI), and dynamic contrast-enhanced (DCE) imaging (11, 12). IVIM provides diffusion and perfusion information within tissue through the biexponential modelling of images acquired by multiple b values (13). Previous studies showed the ability of IVIM for the differential diagnosis of malignant and benign tumors, as well as reflect the biological behavior and predict prognosis (14–16). Amide proton transfer (APT) weighted imaging is a noninvasive molecular imaging technique based on chemical exchange saturation transfer (CEST). It measures the endogenous moving proteins and peptides by detecting the reduction in bulk water intensity, which indirectly reflects changes of the internal metabolism (17, 18). APT weighted imaging has been applied in studies of various cancers, it exhibits an excellent ability in tumor differentiation, grading, and discrimination of treatment related necrosis from recurrence (19–21). Li et al. suggested the utility of APT and IVIM may be a useful technique in the

diagnosis and predicting the differentiation of squamous cell carcinoma (22). Jia et al. found a prediction model incorporating APT and IVIM in the tumor may be useful for predicting the response of hepatocellular carcinoma (HCC) to transarterial chemoembolization (TACE) pretreatment (23). There are few studies using APT on research of rectal cancer. Nishie et al. observed APT weighted imaging can predict the tumor response to neoadjuvant chemotherapy in patients with locally advanced rectal cancer (24). Previous studies have reported that tumors with high-grade, more advanced T stage, and lymph node metastasis had higher APT signal intensity (APT SI) (25–27). However, the previous studies were commonly with limited sample sizes, without involvement of the histopathologic type, perineural invasion and lymphovascular invasion, and without comparison to IVIM.

This study aims to investigate the ability of APT and IVIM in evaluation of prognostic factors for rectal adenocarcinoma, thereby to evaluate its reference value for assessing the malignant degree and predicting tumor aggressiveness, compared with results by conventional DWI.

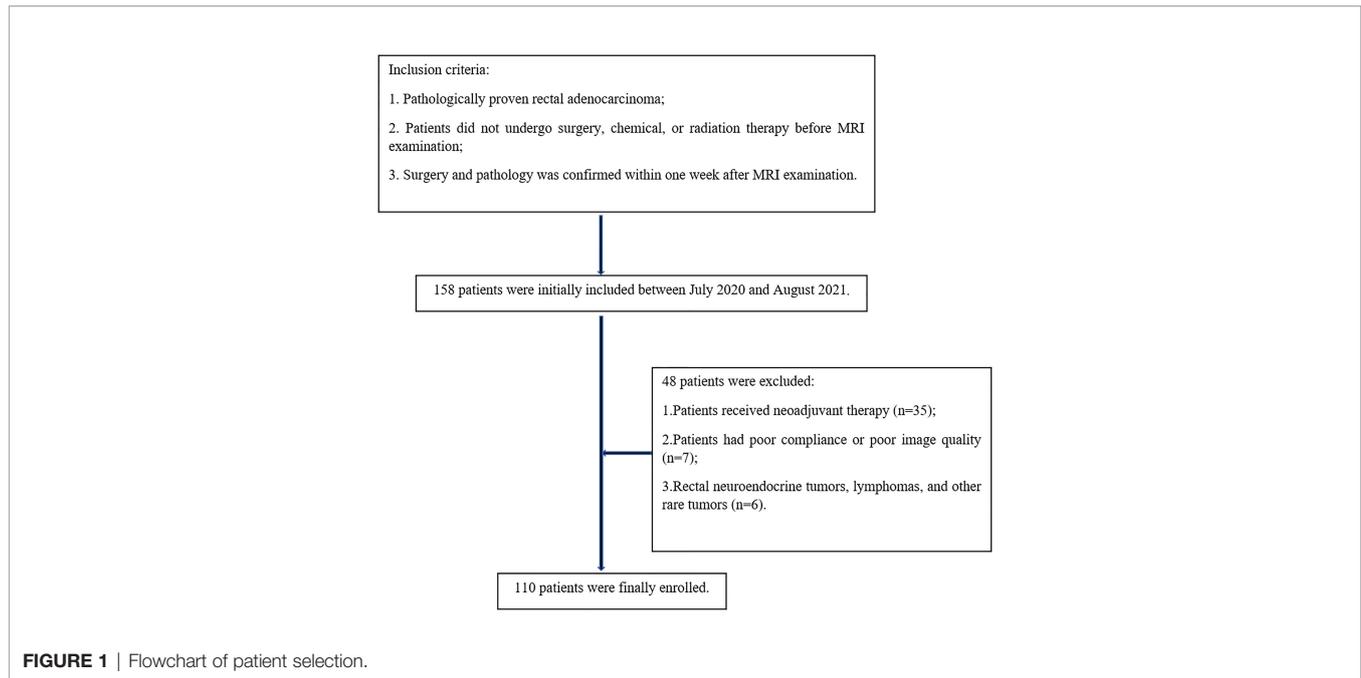
MATERIALS AND METHODS

Participants

Preoperative pelvic MRI data of 158 patients with pathologically confirmed rectal cancer at our hospital were collected between July 2020 and August 2021. The inclusion criteria were as follows: pathologically proven rectal adenocarcinoma; patients did not undergo surgery, chemical, or radiation therapy before MRI examination; surgery and pathology was confirmed within one week after MRI examination. The exclusion criteria were as follows: patients received neoadjuvant therapy (n=35); patients had poor compliance or poor image quality (n=7); rectal neuroendocrine tumor, lymphomas, and other rare tumors (n=6). Finally, 110 patients were enrolled in this study (Figure 1).

MR Imaging Protocols

MRI were performed on a 3T scanner (Ingenia CX, Philips Healthcare, Best, the Netherlands) with a 32-channel phase array coil. Patients were instructed to empty the rectum before examination. To suppress intestinal movement artifacts, 20 mg raceanisodamine hydrochloride injection (Suicheng Pharmaceutical Co, Ltd.) was given intramuscularly 5-10 min before examination. The scanning sequences included T₂WI, T₁-weighted imaging (T₁WI), DWI, APT, IVIM, and dynamic contrast-enhanced T₁WI. The oblique axial was positioned perpendicular to the long axis of the lesion. Detailed parameters for the sequences were listed in Table 1.

**TABLE 1** | MRI acquisition parameters.

| Parameters | APT | IVIM | DWI | T ₂ WI |
|---------------------------------------|----------|---|------------|-------------------|
| Sequence | TSE | EPI | EPI | TSE |
| TR/TE (ms) | 6540/8.3 | 4888/90 | 4421/77 | 3000/100 |
| Field of view (mm ²) | 230×181 | 240×240 | 200×129 | 180×180 |
| Slice thickness (mm) | 5 | 5 | 4 | 4 |
| No. of slices | 9 | 24 | 24 | 24 |
| Matrix | 116×90 | 72×67 | 80×52 | 300×235 |
| Spatial resolution (mm ³) | 2×2×5 | 3.3×3.58×5 | 2.5×2.46×4 | 0.6×0.7×4 |
| b-values (s/mm ²) | N/A | 0, 10, 20, 50, 100, 200, 400, 800, 1200 | 0, 800 | N/A |
| Bandwidth (Hz/pixel) | 647.2 | 2977.8 | 2365.9 | 254.1 |
| TSE factor | 174 | N/A | N/A | 15 |
| Fat suppression | Yes | No | No | NO |
| Acquisition time | 6min0s | 4min53s | 2min26s | 2min54s |

T₂WI, T₂-weighted imaging; TSE, turbo spin echo; EPI, echo-planar imaging; TR, repetition time; TE, echo time; N/A, not applicable.

High-resolution T₂WI was helpful for estimating tumor location, the relationship with peritoneal reflection, tumor size, bowel circumferential involvement, CRM, and EMVI statuses. APT weighted images were acquired by using a 3D turbo spin echo (TSE) sequence for optimized signal-to-noise ratio. The continuous RF saturation for a duration of 2 seconds (each RF coil was turned on and off for 500 msec to generate four block RF pulses at 2 μT amplitude) (20). For convenience, the water frequency (around 4.75 ppm in the proton MR spectrum) is placed at 0 ppm of the Z-spectrum, in which the water signal saturation is measured as a function of saturation frequency. Data were acquired with seven different saturation frequency offsets with respect to the water resonance (± 3.5 , ± 3.42 , ± 3.58 , -1560 ppm). A B₀ map was derived from three echo acquisitions at +3.5 ppm for B₀ correction (28). IVIM (with b values of 0, 10, 20, 50, 100, 200, 400, 800, 1200 s/mm²) were performed in the oblique axial plane

using a single-shot echo planar imaging (ss-EPI) sequence with comparable parameters. The diffusion gradients were applied simultaneously along with three orthogonal directions. DWI (with b values of 0, 800 s/mm²) was also performed using the ss-EPI sequence.

Data Processing and Analysis

APT weighted images were automatically generated on the console at the time of scan completion. After MR scans, all images were uploaded to the IntelliSpace Portal (ISP v10, Philips Healthcare) workstation for post processing or quantitative measurements. The MTR_{asym} (magnetization transfer ratio asymmetry) value at the frequency offset of +3.5 ppm was displayed as percent level (relative to S₀) in the final APT images, and referred as APT SI:

$$\text{APT SI} = \text{MTR}_{\text{asym}} [\Delta\omega = +3.5\text{ppm}] (\%)$$

The IVIM data were processed by the application of advanced diffusion analysis (ADA) on the workstation with maps of the pure diffusion coefficient (D), pseudo-diffusion coefficient (D*) and perfusion fraction (f) generated. The linear fitting equation is as follows:

$$S_b/S_0 = (1 - f) \exp(-b \times D) + f \exp(-b \times D^*)$$

where S_b is the MR signal intensity with diffusion gradient; S_0 is the MR signal intensity without diffusion gradient. The ADC maps were generated immediately after DWI data acquisition.

MRI images were analyzed by two radiologists experienced in gastrointestinal diseases diagnosis. Regions of interest (ROIs) on APT SI, D, D*, f and ADC images were manually selected for analysis, according to T₂WI and pathology results. The ROIs of APT SI were drawn on APT-T2 merged images, then the same ROIs were copied to the ADC images for measuring values. The ROIs of D were drawn on D images, then the same ROIs were showed on the D* and f images for quantitative measurements. The ROIs were drawn at the level of the maximum extent of the tumor and the levels above and below it, and the averaged values were taken. Necrotic, cystic, and hemorrhagic regions were avoided.

Pathologic Analyses

Pathological reports of rectal cancer were referred to standardized templates, including surgical procedures, gross and histological types, tumor grade, pathological stage, perineural invasion, lymphovascular invasion, cut edge infringement, and immunohistochemistry. According to world health organization (WHO) grading criteria, rectal common adenocarcinoma (AC) was classified as grade 1 (G1, well differentiated, >95% gland forming), grade 2 (G2, moderately differentiated, 50-95% gland forming), or grade 3 (G3, poorly differentiated, 0-49% gland forming). According to two-tiered grading system of WHO criteria, G1 and G2 tumors were classified as low-grade tumors, G3 tumors were classified as high-grade tumors. The staging criteria were evaluated according to the American Joint Committee on Cancer (AJCC) 8th edition. T staging was classified as pT1-2 and pT3-4 stage based on depth of tumor invasion. pT1-2 stage cancer was defined as disease confined to the muscularis propria, including pT1 and pT2 stage, and pT3-4 stage cancer was defined as disease extending beyond the muscularis propria, including pT3 and pT4 stage. Lymph node staging was performed based on results of postoperative pathology including pN0 stage: lack of regional lymph node metastasis, pN1 stage: less than 3 regional lymph node metastasis, and pN2 stage: 4 or more regional lymph node metastasis. Perineural invasion, lymphovascular invasion, and EMVI statuses were classified into positive and negative groups.

Statistical Analysis

SPSS 22.0 software (IBM, Armonk, NY) was used for statistical analysis. The Kolmogorov-Smirnov test was performed for analyzing normality. Data conforming to the normal distribution were expressed as mean \pm standard deviation (SD). The intraclass correlation coefficient (ICC) was used to evaluate the interobserver consistency of the measured

parameters. ICC values of less than 0.40, 0.41–0.75, and greater than 0.75 were considered to indicate poor, fair, and good agreement, respectively. The t-test for independent samples was used to compare APT SI, D, D*, f and ADC parameters between pathological types (MC vs. AC), WHO grades (low- vs. high-grade), pT stages (pT1-2 vs. pT3-4), pN stages (pN1-2 vs. pN0), perineural invasion (positive vs. negative), lymphovascular invasion (positive vs. negative), and EMVI statuses (positive vs. negative). For parameters with significant differences between groups, the receiver operating characteristic (ROC) curve was used to analyze their diagnostic efficacy using the software of MedCalc v. 20.0 (MedCalc Software, Ostend, Belgium). DeLong test was used to compare the differences of area under ROC curves (AUCs). The forward model of binary logistic regression was applied for parameter fusion. Differences with $P < 0.05$ were considered statistically significant.

RESULTS

The Pathological Results of Rectal Adenocarcinomas

Among 110 rectal adenocarcinomas, 17 cases were MC and 93 cases were AC. The mean age was 60.31 ± 10.84 years (age range 33–83 years). Within the AC group, 69 and 24 cases were low-grade and high-grade adenocarcinomas, respectively (**Figures 2–4**); 38 and 55 cases were pT1-2 stage and pT3-4 stage, respectively; 64 and 29 cases were pN0 stage and pN1-2 stage, respectively; 23 and 70 cases were positive and negative perineural invasion, respectively; 26 and 67 cases were positive and negative lymphovascular invasion, respectively; 25 and 68 cases were positive and negative EMVI, respectively; 90 and 3 cases were positive and negative CRM, respectively. Clinical features, histopathologic characteristics were summarized in **Table 2**.

Interobserver Agreement

The intraclass correlation coefficient were 0.942 (95% CI 0.831–0.967) for APT SI; 0.862 (95% CI, 0.714–0.913) for D; 0.762 (95% CI, 0.632–0.825) for D*; 0.859 (95% CI, 0.697–0.912) for f; and 0.916 (95% CI, 0.850–0.933) for ADC, respectively. There were good agreements between two observers for measurements of APT SI, D, D*, f, and ADC values.

Comparison of the Parameters in Different Groups of Rectal Adenocarcinomas

Detailed results were showed in **Tables 3, 4** and **Figure 5**. The APT SI, D and ADC values of MC were significantly higher than those of AC (all $P < 0.001$). Within the AC group, the APT SIs were significantly lower, and the D values were higher in low-grade adenocarcinomas than in high-grade ones ($P = 0.001$ and 0.025 ; respectively). The D values were significantly lower in positive than in negative EMVI tumors ($P = 0.045$). No significant difference of APT SI, D, D*, f or ADC observed in other groups (all $P > 0.05$).

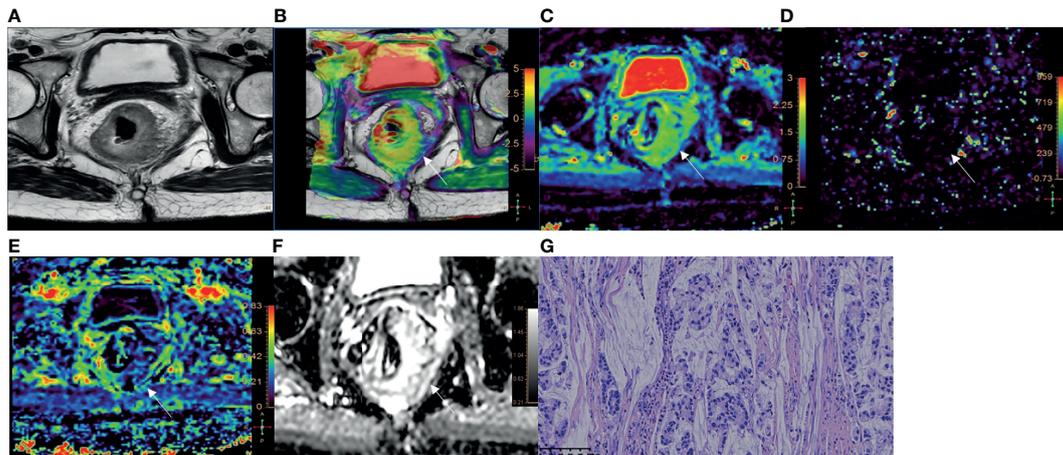


FIGURE 2 | A 51-year-old female with MC. (A) Oblique axial T_2 -weighted image showed a mass with high intensity in the rectum. (B) APT- T_2 merged image showed the mass with a mean APT SI of 3.4%. (C–E) D , D^* and f maps showed the mass with values of $1.42 \times 10^{-3} \text{ mm}^2/\text{s}$, $5.00 \times 10^{-3} \text{ mm}^2/\text{s}$ and 0.22, respectively. (F) The mass showed high intensity ($1.87 \times 10^{-3} \text{ mm}^2/\text{s}$) on the ADC map. (G) HE staining revealed mucinous adenocarcinoma. ($\times 200$).

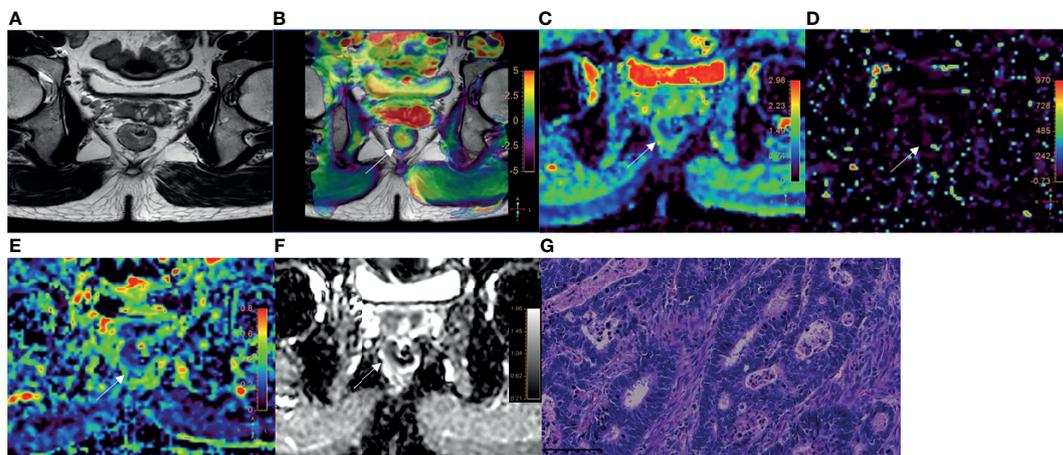


FIGURE 3 | A 52-year-old male with AC of grade 2. (A) Oblique axial T_2 -weighted image showed a mass with slightly high intensity in the rectum. (B) APT- T_2 merged image showed the mass with a mean APT SI of 2.1%. (C–E) D , D^* and f maps showed the mass with values of $0.81 \times 10^{-3} \text{ mm}^2/\text{s}$, $7.74 \times 10^{-3} \text{ mm}^2/\text{s}$ and 0.19, respectively. (F) The mass showed low intensity ($1.03 \times 10^{-3} \text{ mm}^2/\text{s}$) on the ADC map. (G) HE staining revealed moderately differentiated adenocarcinoma. ($\times 200$).

Comparison of ROC Curves for Distinguishing MC From AC, Low- From High-Grade AC, and Distinguishing EMVI Status

ROC curves of APT SI, D , and ADC values between MC and AC, low- and high-grade AC, positive and negative EMVI were listed in **Figure 6**. The ROC curves for distinguishing MC from AC were shown in **Figure 6A** using the APT SI, D and ADC values with the AUCs of 0.921, 0.893, 0.995, respectively. The comparison among these AUCs showed no significant difference (APT SI vs. D : $Z=0.352$, $P=0.725$; APT SI vs. ADC:

$Z=2.457$, $P=0.140$; and D vs. ADC: $Z=1.607$, $P=0.108$; respectively). The AUCs for distinguishing low- from high-grade AC using the APT SI and D values were 0.737 and 0.663, respectively (**Figure 6B**), without significant difference ($Z=0.748$, $P=0.455$). The AUC was increased to 0.806 through the combination of APT SI and D values (**Figure 6B**). The comparison of AUCs showed significant differences between the combined parameter and APT SI ($Z=1.962$, $P=0.049$) or D values ($Z=2.040$, $P=0.041$). The AUC for distinguishing positive EMVI from negative EMVI using the D value was 0.646 (**Figure 6C**). The diagnostic performance and optimal diagnostic threshold of parameters were listed in **Table 5**.

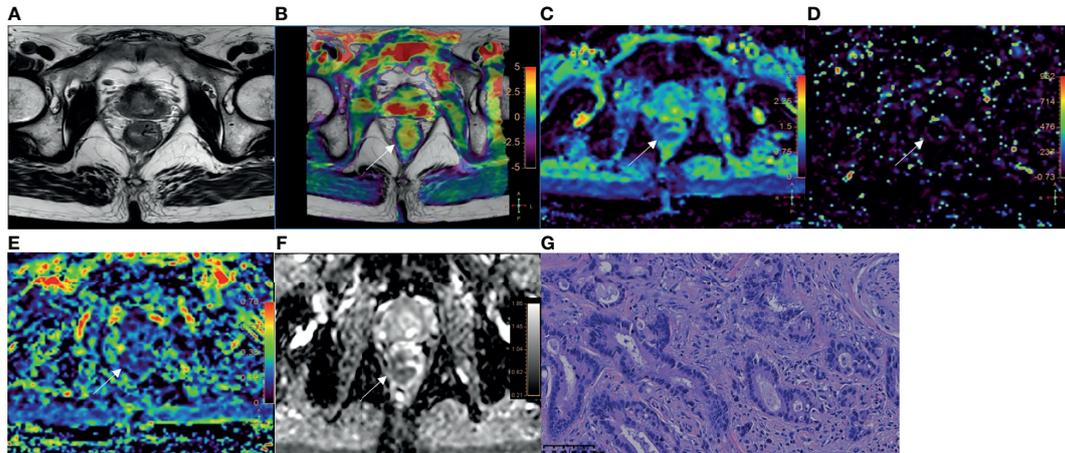


FIGURE 4 | A 78-year-old male with AC of grade 3. **(A)** Oblique axial T_2 -weighted image showed a mass with slightly high intensity in the rectum. **(B)** APT- T_2 merged image showed the mass with a mean APT SI of 2.8%. **(C–E)** D, D^* and D' maps showed the mass with values of $0.79 \times 10^{-3} \text{mm}^2/\text{s}$, $9.40 \times 10^{-3} \text{mm}^2/\text{s}$ and 0.17, respectively. **(F)** The mass showed low intensity ($0.93 \times 10^{-3} \text{mm}^2/\text{s}$) on the ADC map. **(G)** HE staining revealed poorly differentiated adenocarcinoma. ($\times 200$).

TABLE 2 | Clinical and pathological characteristics.

| Category | Gender (Male/ Female) | Relationship with peritoneal reflection (Above/ Across/Below) | Tumor location (Upper/Middle/ Lower) | Tumor longitudinal diameter ($\geq 50\text{mm}/<50\text{mm}$) | Tumor transverse diameter ($\geq 10\text{mm}/<10\text{mm}$) | Bowel circumferential involvement ($\geq 1/2/<1/2$) |
|--------------------------------|--------------------------|---|--|---|---|---|
| Pathological type | | | | | | |
| MC (n=17) | 9/8 | 3/6/8 | 5/7/5 | 10/7 | 14/3 | 15/2 |
| AC (n=93) | 60/33 | 15/28/50 | 32/41/20 | 36/57 | 70/23 | 74/19 |
| WHO grade(AC) | | | | | | |
| G1 (n=2) | 1/1 | 0/1/1 | 1/1/0 | 0/2 | 0/2 | 0/2 |
| G2 (n=67) | 46/21 | 10/18/39 | 13/41/13 | 22/45 | 50/17 | 57/10 |
| G3 (n=24) | 17/7 | 3/7/14 | 4/15/5 | 10/14 | 21/3 | 21/3 |
| T stage(AC) | | | | | | |
| pT1 (n=5) | 3/2 | 0/0/5 | 0/2/3 | 2/3 | 3/2 | 3/2 |
| pT2 (n=33) | 25/8 | 3/8/22 | 4/24/5 | 17/16 | 25/8 | 24/9 |
| pT3 (n=47) | 32/15 | 11/13/23 | 14/19/14 | 16/31 | 39/8 | 42/5 |
| pT4 (n=8) | 3/5 | 0/5/3 | 2/4/2 | 5/3 | 7/1 | 8/0 |
| N stage(AC) | | | | | | |
| pN0 (n=64) | 40/24 | 10/14/40 | 11/39/14 | 33/31 | 53/11 | 54/10 |
| pN1 (n=17) | 12/5 | 1/8/8 | 3/9/5 | 11/6 | 13/4 | 15/2 |
| pN2 (n=12) | 7/5 | 2/5/5 | 4/7/1 | 7/5 | 9/3 | 11/1 |
| Perineural invasion (AC) | | | | | | |
| Positive (n=23) | 14/9 | 1/5/17 | 3/13/7 | 12/11 | 20/3 | 21/2 |
| Negative (n=70) | 49/21 | 12/21/37 | 15/41/14 | 35/35 | 52/18 | 58/12 |
| Lymphovascular invasion(AC) | | | | | | |
| Positive (n=26) | 15/11 | 4/9/13 | 5/13/8 | 16/10 | 20/6 | 21/5 |
| Negative (n=67) | 47/20 | 10/17/40 | 13/42/12 | 33/34 | 53/14 | 57/10 |
| EMVI(AC) | | | | | | |
| Positive (n=25) | 14/11 | 3/8/14 | 7/12/6 | 14/11 | 23/2 | 25/0 |
| Negative (n=68) | 49/19 | 10/18/40 | 10/43/15 | 33/35 | 51/17 | 54/14 |

MC, mucinous adenocarcinoma; AC, common adenocarcinoma.

DISCUSSION

The histopathologic type, tumor grade, T stage, N stage, perineural invasion, lymphovascular invasion, and EMVI

statuses are important prognostic factors for rectal cancer. In our study, we performed a comprehensive investigation of correlations of APT and IVIM parameters with rectal cancer prognostic factors, in comparison with results by DWI. Results

TABLE 3 | The comparison of APT SI, D, D*, f and ADC values in different groups of types.

| Groups | APT SI (%) | D ($\times 10^{-3}\text{mm}^2/\text{s}$) | D* ($\times 10^{-3}\text{mm}^2/\text{s}$) | f | ADC ($\times 10^{-3}\text{mm}^2/\text{s}$) |
|--------------------|-------------------|--|---|-------------------|--|
| Gross types | | | | | |
| Ulcerated (n=73) | 2.578 \pm 0.241 | 0.965 \pm 0.217 | 6.307 \pm 2.135 | 0.163 \pm 0.068 | 0.956 \pm 0.127 |
| Elevated (n=37) | 2.612 \pm 0.325 | 0.942 \pm 0.228 | 6.791 \pm 2.139 | 0.184 \pm 0.045 | 0.911 \pm 0.130 |
| P value | 0.185 | 0.107 | 0.218 | 0.195 | 0.136 |
| Histological types | | | | | |
| MC (n=17) | 3.192 \pm 0.661 | 1.153 \pm 0.238 | 7.017 \pm 2.579 | 0.150 \pm 0.073 | 1.535 \pm 0.203 |
| AC (n=93) | 2.333 \pm 0.471 | 0.792 \pm 0.173 | 6.989 \pm 2.711 | 0.212 \pm 0.033 | 0.986 \pm 0.124 |
| P value | 0.000 | 0.000 | 0.267 | 0.106 | 0.000 |

ADC, apparent diffusion coefficient.

TABLE 4 | The comparison of APT SI, D, D*, f and ADC values in different groups of AC.

| Groups | APT SI (%) | D ($\times 10^{-3}\text{mm}^2/\text{s}$) | D* ($\times 10^{-3}\text{mm}^2/\text{s}$) | f | ADC ($\times 10^{-3}\text{mm}^2/\text{s}$) |
|-------------------------|-------------------|--|---|-------------------|--|
| WHO grade | | | | | |
| Low-grade (n=69) | 2.226 \pm 0.347 | 0.842 \pm 0.148 | 7.193 \pm 2.913 | 0.225 \pm 0.141 | 1.004 \pm 0.129 |
| High-grade (n=24) | 2.688 \pm 0.438 | 0.777 \pm 0.178 | 6.361 \pm 1.877 | 0.171 \pm 0.100 | 0.929 \pm 0.085 |
| P value | 0.001 | 0.025 | 0.420 | 0.124 | 0.155 |
| T stage | | | | | |
| pT1-2 (n=38) | 2.417 \pm 0.318 | 0.772 \pm 0.193 | 6.837 \pm 2.502 | 0.214 \pm 0.158 | 0.983 \pm 0.129 |
| pT3-4 (n=55) | 2.276 \pm 0.335 | 0.806 \pm 0.158 | 7.094 \pm 2.865 | 0.211 \pm 0.116 | 0.989 \pm 0.121 |
| P value | 0.399 | 0.447 | 0.778 | 0.775 | 0.795 |
| N stage | | | | | |
| pN1-2 (n=29) | 2.279 \pm 0.366 | 0.770 \pm 0.175 | 6.689 \pm 2.015 | 0.189 \pm 0.090 | 0.952 \pm 0.110 |
| pN0 (n=64) | 2.236 \pm 0.413 | 0.841 \pm 0.160 | 7.127 \pm 2.575 | 0.222 \pm 0.149 | 1.002 \pm 0.127 |
| P value | 0.692 | 0.157 | 0.163 | 0.359 | 0.074 |
| Perineural invasion | | | | | |
| Positive (n=23) | 2.325 \pm 0.409 | 0.778 \pm 0.186 | 5.986 \pm 1.399 | 0.203 \pm 0.085 | 0.946 \pm 0.184 |
| Negative (n=70) | 2.335 \pm 0.491 | 0.841 \pm 0.111 | 7.294 \pm 2.938 | 0.215 \pm 0.145 | 0.998 \pm 0.131 |
| P value | 0.609 | 0.098 | 0.064 | 0.830 | 0.123 |
| Lymphovascular invasion | | | | | |
| Positive (n=26) | 2.354 \pm 0.443 | 0.787 \pm 0.169 | 6.660 \pm 2.092 | 0.201 \pm 0.081 | 0.968 \pm 0.101 |
| Negative (n=67) | 2.324 \pm 0.485 | 0.805 \pm 0.185 | 7.117 \pm 2.921 | 0.217 \pm 0.149 | 0.993 \pm 0.131 |
| P value | 0.294 | 0.799 | 0.689 | 0.847 | 0.389 |
| EMVI | | | | | |
| Positive (n=25) | 2.416 \pm 0.288 | 0.771 \pm 0.175 | 6.929 \pm 2.115 | 0.201 \pm 0.114 | 0.979 \pm 0.114 |
| Negative (n=68) | 2.306 \pm 0.315 | 0.858 \pm 0.151 | 7.009 \pm 2.891 | 0.216 \pm 0.140 | 0.988 \pm 0.127 |
| P value | 0.139 | 0.045 | 0.665 | 0.654 | 0.901 |

indicated that APT, IVIM, and DWI all can be used in differentiating between AC and MC. APT and IVIM can be used in differentiating grades of AC, and the combination of APT with IVIM could improve the diagnostic performance. DWI can't be used in differentiating grades of AC.

We observed that APT SIs were significantly higher in MC than AC. According to the literature, APT SI was mainly contributed by the endogenous cellular proteins and peptides and affected by intercellular pH environment. Otherwise, cell density, mucin and angiogenesis also have significant effects on APT SI (17, 29). MC is characterized by tumor cell hypersecretion, with more than 50% of mucus content in the tumor parenchyma (5), which may have contributed to the higher APT SIs. D is the pure diffusion coefficient representing pure molecular diffusion, D* is the pseudo-diffusion coefficient representing microperfusion related diffusion, while f is the perfusion fraction related to microcirculation. Our study also found that D and ADC values of MC were significantly higher than those of AC, which was in accordance with previous

research (30). Mucinous adenocarcinoma cells float on a layer of mucus in a relatively loose arrangement, which may decrease the cellularity and facilitate water molecule movement (31). The D* and f values showed no significant difference for distinguishing MC from AC, which may indicate the similar microperfusion component in these two types of lesions.

The histologic grade is an important prognostic factor for rectal adenocarcinoma. We demonstrated that the APT SIs of low-grade adenocarcinomas were significantly lower compared to those of high-grade adenocarcinomas, which was consistent with previous studies (25, 26). Therefore, APT weighted imaging may be helpful to identify the pathological grade of rectal cancer. Similar results have been reported in other tumors. For example, Sotirios et al. found that APT could differentiate low- from high-grade gliomas and predict the histopathological grade potentially (32). A study by Yin et al. demonstrated that APT SIs were significantly higher in prostate cancer than in benign prostatic hyperplasia and showed a strong correlation with the Gleason score (33). All these studies indicated that malignant tumors

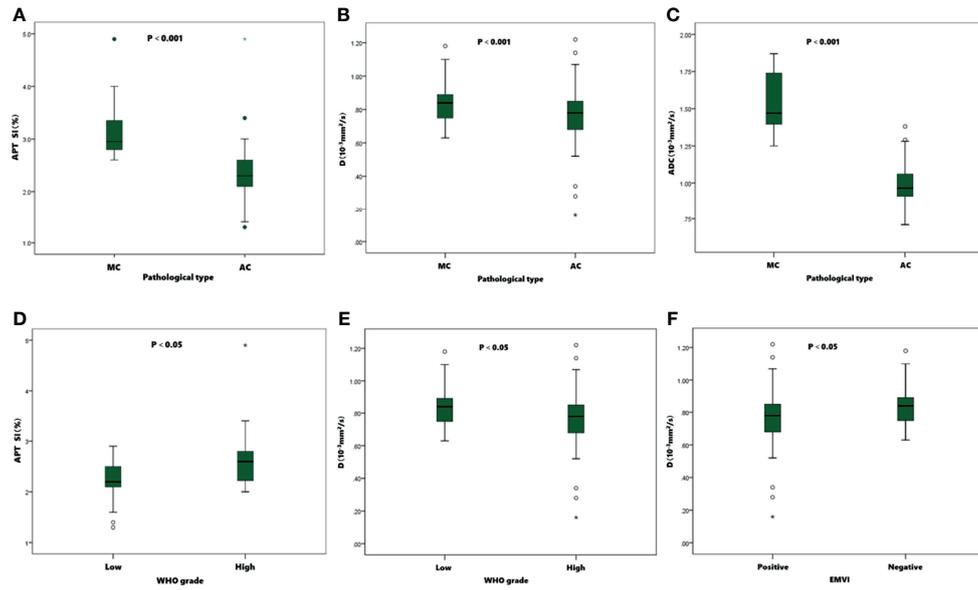


FIGURE 5 | Boxplots of APT SI, D and ADC values in different pathological types, WHO grades, and EMVI statuses of tumors. The APT SIs (A), D (B) and ADC (C) values were significantly higher in MC than in AC. In AC group, the APT SIs (D) were significantly lower in low-grade than in high-grade group, and the D values (E) were significantly higher than in high-grade group. The D values (F) were significantly lower in positive EMVI than negative EMVI.

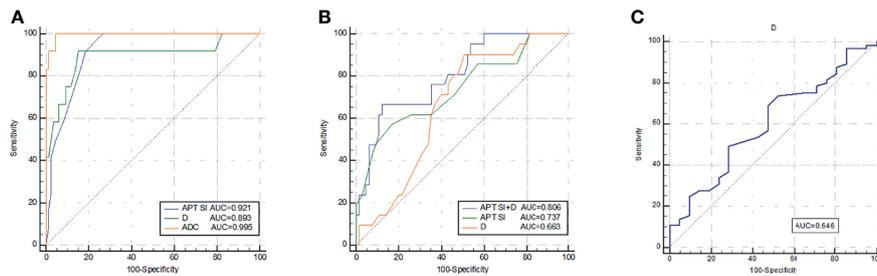


FIGURE 6 | ROC curves of APT SI, D, and ADC for discrimination between MC and AC (A); ROC curves of APT SI, D, and APT SI combined with D for discrimination between low- and high-grade AC (B); and the ROC curve of D for discrimination between positive EMVI and negative EMVI (C). All parameters were with significant differences between the groups.

TABLE 5 | Diagnostic performance of APT SI, D and ADC values with significant difference between groups.

| Category | P value | AUC (95% CI) | Threshold | Sensitivity (%) | Specificity (%) |
|------------------------|---------|---------------------|---|-----------------|-----------------|
| MC vs. AC | | | | | |
| APT SI | 0.000 | 0.921(0.895-0.978) | 2.650% | 91.7 | 81.6 |
| D | 0.000 | 0.893(0.767-1.000) | $0.930 \times 10^{-3} \text{mm}^2/\text{s}$ | 91.7 | 84.9 |
| ADC | 0.000 | 0.995(0.986-1.000) | $1.230 \times 10^{-3} \text{mm}^2/\text{s}$ | 100 | 95.4 |
| Low- vs. high grade AC | | | | | |
| APT SI | 0.001 | 0.737 (0.607-0.867) | 2.550% | 57.1 | 81.8 |
| D | 0.025 | 0.663(0.544-0.782) | $0.851 \times 10^{-3} \text{mm}^2/\text{s}$ | 48.5 | 80.5 |
| APT SI + D | 0.000 | 0.806(0.702-0.910) | N/A | 76.7 | 87.7 |
| EMVI (+) vs. (-) AC | | | | | |
| D | 0.045 | 0.646(0.511-0.780) | $0.785 \times 10^{-3} \text{mm}^2/\text{s}$ | 76.2 | 76.9 |

AUC, area under the curve; CI, confidence interval; APT SI +D, APT SI combined with D values.

commonly have significantly higher APT SIs compared to those of the benign or normal tissues, and the APT SI tends to increase as the pathological grade advanced (34, 35). The higher APT SIs in high-grade tumors can be due to the abundant proteins production, rapid cell proliferation and angiogenesis. Previous studies suggested that IVIM was helpful to assess tumor grades of intracranial tumors, solid soft-tissue tumors, HCC, and prostate cancer etc., and the D value was observed to be inversely correlated with the tumor grade (36–39). In present study, lower D values were observed in high-grade than those of low-grade rectal adenocarcinomas, which was in agreement with previous results (11, 40, 41). The D value, that represents the pure diffusion of free water molecules, was decreased with the increasing cellularity, tight cellular structure in high-grade tumors. The AUCs of APT and D for distinguishing low- from high-grade adenocarcinomas were 0.737 and 0.663 respectively, with moderate diagnostic performance. The AUC was raised to 0.806 by the combination of APT SI and D values, with 76.7% sensitivity and 87.7% specificity. The D^* and f values showed a trend of decreasing with increased tumor grades in our study but without statistically significance. Furthermore, previous studies showed D^* or f was negative correlated with tumor grade in rectal cancer (11, 30, 40, 41). A possible explanation is that tumor cells grow rapidly in high-grade tumor, leading to immature vascular structure and reduced microcirculation perfusion thus lower perfusion-related parameters, such as D^* and f values. The ADC values showed no significant differences for distinguishing tumor grade of AC in present study, which may be caused by the integrated effects of both diffusion and microperfusion.

Tumor stage is closely related to prognosis for rectal adenocarcinoma. In present study, postoperative pathological stage was used to retrospectively analyze the correlation of APT and IVIM parameters with tumor stages. Parameters derived from APT and IVIM showed no significant differences between pT1-2 and pT3-4 stages, or between pN1-2 and pN0 stages. These results were inconsistent with previous studies which showed that APT SIs were higher in advanced T stage and lymph node metastasis (25, 26). However, T1 or T4 stage cases were absent in previous studies, and the positive rate of lymph node metastasis was higher in previous studies than our study, which may cause the selection bias of the sample. Sun et al. observed that D and D^* showed a trend of decreasing with the increasing of tumor clinical stages and lymph node metastasis in rectal cancer (40). The parameters derived from APT and IVIM might exhibit more aggressive biologic behavior, further study is needed to evaluate the significance.

EMVI refers to the presence of tumor infiltration in the vessels outside the muscularis propria, and it is an independent prognostic factor of rectal cancer. Positive EMVI exhibits more local recurrence, more distant metastasis, and more tumor-related death (42). Although Chen et al. suggested that APT SIs were higher in EMVI-positive than in EMVI-negative cases (26), our study showed no significant difference of APT SI in EMVI involvement. We considered that the inconsistent results might be related to the different positive rate of EMVI status (26.9% in present study while 50.8% in the previous study). In addition, the D value was observed to be lower in the positive

EMVI group than in the negative group in this study, while Wei et al. identified that D value was lower in microvascular invasion (MVI)-positive than in MVI-negative HCCs (43). The decreased D value may be because tumor emboli or clusters of cancer cells restrict the diffusion of water molecules. The AUC for distinguishing EMVI involvement using the D value was 0.646 with moderate diagnostic significance. The high-resolution T₂WI images should be combined to improve the diagnosis accuracy of EMVI, which was considered positive if vessel wall irregularity, abnormal extension, suspected the empty signal was replaced by tumor tissue with intermediate signal intensity. Additionally, perineural invasion and lymphovascular invasion are prognostic factors for rectal cancer associated with recurrence, metastasis, and postoperative adjuvant therapy. No significant difference of parameters derived from APT and IVIM were found in groups with and without different types of structure invasion in our study, which may be because the tumor microenvironment reflected by APT or IVIM parameters is insufficient to cause significant changes in perineural and lymphovascular invasion.

The present study has some limitations. First, patients with locally advanced rectal cancer received neoadjuvant therapy were excluded, potentially causing selection bias. Second, only 2 types of rectal adenocarcinomas were collected, and further studies with abundant cases are needed to be explored. Third, the choice of different ROIs may also lead to differences in results due to tumor heterogeneity. Furthermore, this study did not analyze the correlation of APT and IVIM parameters with immunohistochemical indicators or gene expression. In the future, collection of complete data for more in-depth research is needed.

CONCLUSION

APT SI and D values can be used in discriminating between MC and AC, slightly inferior to ADC. The APT SI and D values were helpful to differentiate the low- and high-grade of AC, and the combination of APT SI with D values could improve the diagnostic performance. The D value can help determine EMVI status for AC patients. However, it is still debatable whether APT or IVIM can help distinguish stage, perineural invasion, and lymphovascular invasion. In conclusion, APT and IVIM were helpful to assess the prognostic factors related to rectal adenocarcinoma, including histopathological type, tumor grade and EMVI status.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

JL: manuscript preparation, literature research, data acquisition, statistical analysis, and manuscript editing. LL: study design, manuscript preparation, data analysis, and

manuscript revising. XG: manuscript preparation and literature research. SL and JC: study conception and design, manuscript review, and guarantor of integrity of the entire study. All authors contributed to the article and approved the submitted version.

REFERENCES

- Siegel RL, Miller KD, Goding Sauer A, Fedewa SA, Butterly LF, Anderson JC, et al. Colorectal Cancer Statistics, 2020. *CA Cancer J Clin* (2020) 70:145–64. doi: 10.3322/caac.21601
- Wong MCS, Huang J, Lok V, Wang J, Fung F, Ding H, et al. Differences in Incidence and Mortality Trends of Colorectal Cancer Worldwide Based on Sex, Age, and Anatomic Location. *Clin Gastroenterol Hepatol* (2021) 19:955–966.e61. doi: 10.1016/j.cgh.2020.02.026
- Glynn-Jones R, Wyrwicz L, Tiret E, Brown G, Rödel C, Cervantes A, et al. Rectal Cancer: ESMO Clinical Practice Guidelines for Diagnosis, Treatment and Follow-Up. *Ann Oncol* (2017) 28:22–40. doi: 10.1093/annonc/mdx224
- Wilkinson N. Management of Rectal Cancer. *Surg Clin North Am* (2020) 100:615–28. doi: 10.1016/j.suc.2020.02.014
- Reynolds IS, O'Connell E, Fichtner M, McNamara DA, Kay EW, Prehn JHM, et al. Mucinous Adenocarcinoma of the Colon and Rectum: A Genomic Analysis. *J Surg Oncol* (2019) 120:1427–35. doi: 10.1002/jso.25764
- McCawley N, Clancy C, O'Neill BD, Deasy J, McNamara DA, Burke JP. Mucinous Rectal Adenocarcinoma Is Associated With a Poor Response to Neoadjuvant Chemoradiotherapy: A Systematic Review and Meta-Analysis. *Dis Colon Rectum* (2016) 59:1200–8. doi: 10.1097/DCR.0000000000000635
- Nougaret S, Reinhold C, Mikhael HW, Rouanet P, Bibeau F, Brown G. The Use of MR Imaging in Treatment Planning for Patients With Rectal Carcinoma: Have You Checked the “DISTANCE”? *Radiology* (2013) 268:330–44. doi: 10.1148/radiol.13121361
- Horvat N, Carlos Tavares Rocha C, Clemente Oliveira B, Petkovska I, Gollub MJ. MRI of Rectal Cancer: Tumor Staging, Imaging Techniques, and Management. *Radiographics* (2019) 39:367–87. doi: 10.1148/rg.2019180114
- Schurink NW, Lambregts DMJ, Beets-Tan RGH. Diffusion-Weighted Imaging in Rectal Cancer: Current Applications and Future Perspectives. *Br J Radiol* (2019) 92:20180655. doi: 10.1259/bjr.20180655
- Zhu L, Pan Z, Ma Q, Yang W, Shi H, Fu C, et al. Diffusion Kurtosis Imaging Study of Rectal Adenocarcinoma Associated With Histopathologic Prognostic Factors: Preliminary Findings. *Radiology* (2017) 284:66–76. doi: 10.1148/radiol.2016160094
- Geng Z, Zhang Y, Yin S, Lian S, He H, Li H, et al. Preoperatively Grading Rectal Cancer With the Combination of Intravoxel Incoherent Motions Imaging and Diffusion Kurtosis Imaging. *Contrast Media Mol Imaging* (2020) 2020:2164509. doi: 10.1155/2020/2164509
- Bakke KM, Grovik E, Meltzer S, Negård A, Holmedal SH, Mikalsen LTG, et al. Comparison of Intravoxel Incoherent Motion Imaging and Multiecho Dynamic Contrast-Based MRI in Rectal Cancer. *J Magn Reson Imaging* (2019) 50:1114–24. doi: 10.1002/jmri.26740
- Le Bihan D. What Can We See With IVIM MRI? *Neuroimage* (2019) 187:56–67. doi: 10.1016/j.neuroimage.2017.12.062
- Ma G, Xu XQ, Zhu LN, Jiang JS, Su GY, Hu H, et al. Intravoxel Incoherent Motion Magnetic Resonance Imaging for Assessing Parotid Gland Tumors: Correlation and Comparison With Arterial Spin Labeling Imaging. *Korean J Radiol* (2021) 22:243–52. doi: 10.3348/kjr.2020.0290
- Song XL, Wang L, Ren H, Wei R, Ren JL, Niu J. Intravoxel Incoherent Motion Imaging in Differentiation Borderline From Malignant Ovarian Epithelial Tumors: Correlation With Histological Cell Proliferation and Vessel Characteristics. *J Magn Reson Imaging* (2020) 51:928–35. doi: 10.1002/jmri.26885
- Lecler A, Duron L, Zmuda M, Zuber K, Bergès O, Putterman M, et al. Intravoxel Incoherent Motion (IVIM) 3 T MRI for Orbital Lesion Characterization. *Eur Radiol* (2021) 31:14–23. doi: 10.1007/s00330-020-07103-1
- Zhou J, Heo HY, Knutsson L, van Zijl PCM, Jiang S. APT-Weighted MRI: Techniques, Current Neuro Applications, and Challenging Issues. *J Magn Reson Imaging* (2019) 50:347–64. doi: 10.1002/jmri.26645
- Wu Y, Chen Y, Zhao Y, Yang S, Zhao J, Zhou J, et al. Direct Radiofrequency Saturation Corrected Amide Proton Transfer Tumor MRI at 3T. *Magn Reson Med* (2019) 81:2710–9. doi: 10.1002/mrm.27562
- Law BKH, King AD, Ai QY, Poon DMC, Chen W, Bhatia KS, et al. Head and Neck Tumors: Amide Proton Transfer MRI. *Radiology* (2018) 288:782–90. doi: 10.1148/radiol.2018171528
- He YL, Li Y, Lin CY, Qi YF, Wang X, Zhou HL, et al. Three-Dimensional Turbo-Spin-Echo Amide Proton Transfer-Weighted MRI for Cervical Cancer: A Preliminary Study. *J Magn Reson Imaging* (2019) 50:1318–25. doi: 10.1002/jmri.26710
- Qamar S, King AD, Ai QH, Mo FKF, Chen W, Poon DMC, et al. Pre-Treatment Amide Proton Transfer Imaging Predicts Treatment Outcome in Nasopharyngeal Carcinoma. *Eur Radiol* (2020) 30:6339–47. doi: 10.1007/s00330-020-06985-5
- Li B, Sun H, Zhang S, Wang X, Guo Q. The Utility of APT and IVIM in the Diagnosis and Differentiation of Squamous Cell Carcinoma of the Cervix: A Pilot Study. *Magn Reson Imaging* (2019) 63:105–13. doi: 10.1016/j.jmri.2019.08.020
- Jia F, Wu B, Yan R, Li L, Wang K, Han D. Prediction Model for Intermediate-Stage Hepatocellular Carcinoma Response to Transarterial Chemoembolization. *J Magn Reson Imaging* (2020) 52:1657–67. doi: 10.1002/jmri.27189
- Nishie A, Asayama Y, Ishigami K, Ushijima Y, Takayama Y, Okamoto D, et al. Amide Proton Transfer Imaging to Predict Tumor Response to Neoadjuvant Chemotherapy in Locally Advanced Rectal Cancer. *J Gastroenterol Hepatol* (2019) 34:140–6. doi: 10.1111/jgh.14315
- Li L, Chen W, Yan Z, Feng J, Hu S, Liu B, et al. Comparative Analysis of Amide Proton Transfer MRI and Diffusion-Weighted Imaging in Assessing P53 and Ki-67 Expression of Rectal Adenocarcinoma. *J Magn Reson Imaging* (2020) 52:1487–96. doi: 10.1002/jmri.27212
- Chen W, Li L, Yan Z, Hu S, Feng J, Liu G, et al. Three-Dimension Amide Proton Transfer MRI of Rectal Adenocarcinoma: Correlation With Pathologic Prognostic Factors and Comparison With Diffusion Kurtosis Imaging. *Eur Radiol* (2021) 31:3286–96. doi: 10.1007/s00330-020-07397-1
- Nishie A, Takayama Y, Asayama Y, Ishigami K, Ushijima Y, Okamoto D, et al. Amide Proton Transfer Imaging Can Predict Tumor Grade in Rectal Cancer. *Magn Reson Imaging* (2018) 51:96–103. doi: 10.1016/j.jmri.2018.04.017
- Togao O, Keupp J, Hiwatashi A, Yamashita K, Kikuchi K, Yoneyama M, et al. Amide Proton Transfer Imaging of Brain Tumors Using a Self-Corrected 3d Fast Spin-Echo Dixon Method: Comparison With Separate B0 Correction. *Magn Reson Med* (2017) 77:2272–9. doi: 10.1002/mrm.26322
- Ray KJ, Simard MA, Larkin JR, Coates J, Kinchesh P, Smart SC, et al. Tumor pH and Protein Concentration Contribute to the Signal of Amide Proton Transfer Magnetic Resonance Imaging. *Cancer Res* (2019) 79:1343–52. doi: 10.1158/0008-5472.CAN-18-2168
- Lu B, Yang X, Xiao X, Chen Y, Yan X, Yu S. Intravoxel Incoherent Motion Diffusion-Weighted Imaging of Primary Rectal Carcinoma: Correlation With Histopathology. *Med Sci Monit* (2018) 24:2429–36. doi: 10.12659/msm.908574
- Wen Z, Chen Y, Yang X, Lu B, Liu Y, Shen B, et al. Application of Magnetic Resonance Diffusion Kurtosis Imaging for Distinguishing Histopathologic Subtypes and Grades of Rectal Carcinoma. *Cancer Imaging* (2019) 19:8. doi: 10.1186/s40644-019-0192-x
- Sotirios B, Demetriou E, Topriceanu CC, Zakrzewska Z. The Role of APT Imaging in Gliomas Grading: A Systematic Review and Meta-Analysis. *Eur J Radiol* (2020) 133:109353. doi: 10.1016/j.ejrad.2020.109353

33. Yin H, Wang D, Yan R, Jin X, Hu Y, Zhai Z, et al. Comparison of Diffusion Kurtosis Imaging and Amide Proton Transfer Imaging in the Diagnosis and Risk Assessment of Prostate Cancer. *Front Oncol* (2021) 11:640906. doi: 10.3389/fonc.2021.640906
34. Kamitani T, Sagiyama K, Togao O, Yamasaki Y, Hida T, Matsuura Y, et al. Amide Proton Transfer (APT) Imaging of Parotid Tumors: Differentiation of Malignant and Benign Tumors. *Eur J Radiol* (2020) 129:109047. doi: 10.1016/j.ejrad.2020.109047
35. Xu Z, Ke C, Liu J, Xu S, Han L, Yang Y, et al. Diagnostic Performance Between MR Amide Proton Transfer (APT) and Diffusion Kurtosis Imaging (DKI) in Glioma Grading and IDH Mutation Status Prediction at 3 T. *Eur J Radiol* (2021) 134:109466. doi: 10.1016/j.ejrad.2020.109466
36. Wu H, Zhang S, Liang C, Liu H, Liu Y, Mei Y, et al. Intravoxel Incoherent Motion MRI for the Differentiation of Benign, Intermediate, and Malignant Solid Soft-Tissue Tumors. *J Magn Reson Imaging* (2017) 46:1611–8. doi: 10.1002/jmri.25733
37. Kikuchi K, Hiwatashi A, Togao O, Yamashita K, Kamei R, Momosaka D, et al. Intravoxel Incoherent Motion MR Imaging of Pediatric Intracranial Tumors: Correlation With Histology and Diagnostic Utility. *Am J Neuroradiol* (2019) 40:878–84. doi: 10.3174/ajnr.A6052
38. Sokmen BK, Sabet S, Oz A, Server S, Namal E, Dayangac M, et al. Value of Intravoxel Incoherent Motion for Hepatocellular Carcinoma Grading. *Transplant Proc* (2019) 51:1861–6. doi: 10.1016/j.transproceed.2019.02.027
39. Cui Y, Li C, Liu Y, Jiang Y, Yu L, Liu M, et al. Differentiation of Prostate Cancer and Benign Prostatic Hyperplasia: Comparisons of the Histogram Analysis of Intravoxel Incoherent Motion and Monoexponential Model With in-Bore MR-Guided Biopsy as Pathological Reference. *Abdom Radiol (NY)* (2020) 45:3265–77. doi: 10.1007/s00261-019-02227-5
40. Sun H, Xu Y, Song A, Shi K, Wang W. Intravoxel Incoherent Motion MRI of Rectal Cancer: Correlation of Diffusion and Perfusion Characteristics With Prognostic Tumor Markers. *Am J Roentgenol* (2018) 210:W139–47. doi: 10.2214/AJR.17.18342
41. Cheng Y, Jiang H, Wang H, Tang Q, Liu T. Application of Field-Of-View Optimized and Constrained Undistorted Single Shot (FOCUS) With Intravoxel Incoherent Motion (IVIM) in 3T in Locally Advanced Rectal Cancer. *Dis Markers* (2021) 2021:5565902. doi: 10.1155/2021/5565902
42. Zhang XY, Wang S, Li XT, Wang YP, Shi YJ, Wang L, et al. MRI of Extramural Venous Invasion in Locally Advanced Rectal Cancer: Relationship to Tumor Recurrence and Overall Survival. *Radiology* (2018) 289:677–85. doi: 10.1148/radiol.2018172889
43. Wei Y, Huang Z, Tang H, Deng L, Yuan Y, Li J, et al. IVIM Improves Preoperative Assessment of Microvascular Invasion in HCC. *Eur Radiol* (2019) 29:5403–14. doi: 10.1007/s00330-019-06088-w

Conflict of Interest: Author LL was employed by Philips Healthcare.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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