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Strategies to overcome myeloid cell induced immune suppression in the tumor microenvironment

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Cancer progression and metastasis due to tumor immune evasion and drug resistance is strongly associated with immune suppressive cellular responses, particularly in the case of metastatic tumors. The myeloid cell component plays a key role within the tumor microenvironment (TME) and disrupts both adaptive and innate immune cell responses leading to loss of tumor control. Therefore, strategies to eliminate or modulate the myeloid cell compartment of the TME are increasingly attractive to non-specifically increase anti-tumoral immunity and enhance existing immunotherapies. This review covers current strategies targeting myeloid suppressor cells in the TME to enhance anti-tumoral immunity, including strategies that target chemokine receptors to deplete selected immune suppressive myeloid cells and relieve the inhibition imposed on the effector arms of adaptive immunity. Remodeling the TME can in turn improve the activity of other immunotherapies such as checkpoint blockade and adoptive T cell therapies in immunologically "cold" tumors. When possible, in this review, we have provided evidence and outcomes from recent or current clinical trials evaluating the effectiveness of the specific strategies used to target myeloid cells in the TME. The review seeks to provide a broad overview of how myeloid cell targeting can become a key foundational approach to an overall strategy for improving tumor responses to immunotherapy.

KEYWORDS

cancer, immune suppresion, tumor associate macrophages (TAM), myeloid derived suppressor cells (MDSC), tumor associated neutrophils (TAN), cancer immune therapy, tumor microenvironment, dendritc cells

1 Introduction

Local immune suppression and dysregulation are common features of cancer and are closely associated with tumor metastasis and resistance to therapy. The interaction between cancer and the host immune system is a key factor in determining tumor control or progression (1-3). Tumor infiltrating leukocytes, particularly monocytes, myeloid derived

suppressor cells (MDSCs) and neutrophils create a tumor microenvironment (TME) that is inhospitable to effector cells such as CD4 and CD8 T cells and NK cells (4-8). Myeloid lineage cells such as dendritic cells (DCs), tumor associated macrophages (TAMs) and MDSCs can serve a dichotomous role within the TME, though in general they are largely immune suppressive (9-12). These myeloid cells can promote tumor growth by exerting immune suppressive pressure, including secreted cytokines and growth factors promoting angiogenesis, direct cellular signaling or recruitment of Tregs and other immune suppressive cells such as TAMs, MDSCs, tumor associated neutrophils (TANs) and DCs (13). Myeloid cells in the TME can also assume a tumoricidal phenotype, as is the case with activated M1 macrophages producing free radicals and cytokines that stimulate the activation of effector T cells (14), or antigen presenting DCs that promote the expansion and activation of effector CD4 and CD8 T cells.

The net outcome of the dynamic interplay in the TME is determined in part by secreted factors and cell signaling from tumor and stromal cells and by the resident immune cells within the TME, which perpetuate either a suppressive or stimulatory immune landscape (1, 4, 10, 12). Targeting of myeloid immune suppressor cells to reduce or eliminate their immune suppressive impacts on adaptive immunity can turn the tide between cancer and the host's immunity, thereby increasing tumor control and improving the efficacy of other treatments. In this review we summarize past and current strategies including relevant clinical trials that target myeloid cells in the TME as cancer immunotherapy strategies. Although this is not intended to be a fully comprehensive review of all strategies and trials, the goal is to emphasize that the myeloid cell component of the TME presents many opportunities for development of new immune based therapeutics.

2 Immune suppressive myeloid cells, origins, and key functions

2.1 Origin and differentiation of immune suppressor cells

Immune suppressor cells in the tumor microenvironment can be characterized by their cell type of origin. Myeloid derived suppressor cells (MDSC) are comprised of both neutrophil derived MDSC (PMN-MDSC) and monocyte derived MDSC (M-MDSC) with potent immune suppressive activity (15). Tumor associated macrophages (TAM) are derived from inflammatory monocytes recruited from the bloodstream in response to chemokines produced by tumor cells and the tumor stroma, including also myeloid cells themselves, and can be clearly distinguished phenotypically and functionally from MDSC (16) (17). The distinction between tumor associated neutrophils (TAN) and PMN-MDSC is somewhat more complicated, in that they share many phenotypic characteristics (18). Tolerogenic DCs are dendritic cells exposed to polarizing cytokines and surface molecules secreted by tumor cells and stromal cells within the tumor microenvironment (9) This population of immune suppressive DC suppress effector T cell responses, thereby contributing to an overall immune suppressed and hostile environment for infiltrating T cells.

The immune suppressive TAMs, DCs, TANs and MDSCs are recruited to the tumor by a variety of cellular and soluble factors within the tumor milieu, where they suppress effector functions of T cells and NK cells (Figure 1). The various mechanisms employed by immune suppressive myeloid cells become potential targets for new immunotherapies designed to reprogram the TME. Among the mechanisms employed by TAMs and MDSCs to suppress effector T



cells and NK cells include upregulated expression of immune suppressive checkpoint molecules such as PD-L1 (19, 20), VISTA (21, 22), and B7-H3) (21, 23–25). Other mechanisms include secretion of immune suppressive cytokines such as IL10 and TGF β , and VEGF (6, 26).

2.2 Function of TAMs in the tumor microenvironment

Immunologically "cold" tumors evade immune surveillance through a variety of mechanisms. Down regulation of tumor associated antigens (TAA) (27, 28), maintaining inflammation leading to immune exhaustion (6, 26) and increasing angiogenesis to tumor sites (29, 30) are all methods used by tumors to persist and metastasize while evading detection by the immune system.

There is strong clinical evidence linking TAMs to cancer immune suppression. For example, the density of TAMs infiltrating tumors is strongly correlated with poor overall survival in many breast, ovarian, bladder, gastric, thyroid and colorectal cancers (31). TAMs have a relatively short half-life and are therefore must be replaced continuously by inflammatory monocytes recruited from the bloodstream primarily in response to the chemokine CCL2, produced by tumor cells, tumor fibroblasts, and by myeloid cells (32). This dependence of TAMs on continuous monocyte replacement opens a window of opportunity for therapeutic intervention and depletion of TAMS.

Once within the tumor, the differentiation of monocytes to TAMs is guided by either pro-inflammatory or anti-inflammatory factors produced within the TME. The overwhelming majority of TAMs in most tumors exist in a state that most closely resembles that of what has been defined experimentally as M2 polarization, which results in a macrophage that is generally immune suppressive and tumor growth and metastasis promoting (3). The M2 polarization state of TAMS is driven by a a diverse array of cytokines (eg, IL-10, TGF-b), chemokines (CXCL4, CCL5), growth factors (VEGF, M-CSF) and by local tumor hypoxia (33). Tumor cells can also directly contribute metabolically to M2 polarization by secretion of lactic acid and hypoxia-inducible factor (HIF1a) (34). Tumor cells also co-opt TAM signaling to promote tumor growth locally, and to become more invasive for generating metastases. For example, tumor secretion of TNF- α induces the chemokines CCL2 and CCL8 by TAMs which recruits additional CCR2+ monocytes to the TME (35). In another example, CCL8 produced by TAMs also upregulate tumor cell secretion of colony stimulating factor 1 (CSF-1) which is crucial to macrophage and DC survival and differentiation through signaling via CSF-1R (36, 37). One of the most important consequences of the accumulation of TAMs is the impact on T cell effector functions. For example, TGF- β signaling drives CD4 T cell differentiation towards immune suppressive Th2 and Treg phenotypes (38). TGF- β signaling also suppresses the effector functions of CD8 T cells and NK cells and decreases migration of DCs into the tumor tissues (9). Within the TME, T cells responding to TAM secreted factors exhibit upregulated expression of immune suppressive immune checkpoint molecules such as PD-1, CTLA-4, Lag3, and TIM3 (39). The expression of the ligand for PD-1 (PDL-1) is often higher on TAMs than it is on tumor cells, and PDL-1 signaling to TAM directly can reduce their ability to phagocytose tumor cells (40).

Metabolically, TAMs can reprogram the TME by producing enzymes that directly alter T cell signaling, or deplete necessary amino acids needed for T cell survival and proliferation. For example, TAM production of arginase 1 (Arg-1) leads to the depletion of L-arginine in turn leading to dysfunction of tumor infiltrating lymphocytes by TCR ζ chain downregulation (1, 12, 41). In another example, TAM (and tumor cells) can overproduce the enzyme indoleamine dehydrogenase (IDO), which depletes the TME of tryptophan, a necessary amino acid for T cell survival (42).

2.3 Function of immune suppressive DCs in the tumor microenvironment

Like TAMs, DCs in the TME exist primarily in an immune suppressive state and by inactivating effector T cells can promote more rapid tumor growth and metastasis (9). Immature DCs that reside in the TME recognize tumor cells and the products of tumor cell necrosis through damage-associated molecular patterns (DAMPs) which induce DC phagocytosis and processing of tumor antigens. This process matures DCs to serve their primary role as antigen presenting cells, and stimulates migration to lymph nodes, and ultimately leading to activation or inactivation of both CD4 and CD8 effector T cells (43). Classically differentiated DCs secrete proinflammatory cytokines such as IL-12 to activate IFNy producing T cells and NK cells, which drives differentiation of Th1 T cells and activated CD8+ cytotoxic T cells (44). However, in the TME most DCs exist in an immature state and become toleragenic DCs (tDCs) following sustained exposure to cytokines such as VEGF, IL10 and TGF β (9, 43). tDCs induce T cell anergy through checkpoint molecule signaling, including signaling via CD28 and PDL-1 to their cognate receptors CTLA-4 and PD-1 on T cells (45). tDCs also promote the generation of regulatory T cells (Tregs) from naïve CD4+ T cells by secretion of IL10 and TGF- β (43).

2.4 MDSCs in the tumor microenvironment

MDSCs are derived from immature monocytes and neutrophils, mobilized from the bone marrow in response to cytokines associated with chronic inflammation, including IL-3, GM-CSF, and G-CSF (46). The two primary populations of MDSC are defined as monocytic MDSC (M-MDSC) and neutrophilic MDSC (PMN-MDSC), which have both distinct and overlapping molecular and functional characteristics. For example, they exhibit distinct gene expression profiles, and unique immunologic functions such as production of arginase (PMN-MDSC) or reactive nitrogen or oxygen intermediates (M-MDSC) (47). Following their mobilization from the bone marrow and entry into the bloodstream, MDSC are recruited into tumor tissues in part by following chemokine gradients such as CCL2 and CXCL8, and other cytokines secreted by tumor cells and immune cells within the TME (48). Once in the TME, MDSC can be induced to undergo further differentiation to become more immune suppressive, by factors such as TGF-b produced by Tregs. MDSCs can also accumulate in secondary lymphoid tissues including the spleen and lymph nodes where they contribute to systemic immune suppression and further promote tumor progression (38, 49).

The expansion and differentiation of Tregs within the TME, is promoted in part by MDSC expression of tumor derived peptides on MHCI and MHCII molecules (50). As another mechanism of MDSC polarization, histamine released by mast cells binds to histamine receptor 1 on MDSCs inducing secretion of Arg-1 and nitric oxide synthase (iNOS) which inhibits T cell proliferation (51). Recent studies have correlated the abundance of MDSCs with poor prognosis and poor response to immune checking inhibitor (ICI) therapy in patients with various cancer types including breast, colorectal, lung and prostate cancers (52–54).

M-MDSC were originally defined in tumor-bearing mice as immature bone marrow derived cells that suppressed multiple T cell functions (55). The population of M-MDSC overall is considered to be more immune suppressive than PMN-MDSCs despite making up only 10-20% of the total MDSC population (56). Mechanisms of M-MDSC-mediated immune suppression include production of suppressive cytokines IL10 and TGF- β (4); They also promote T cell apoptosis by TCR- ζ chain downregulation through secretion of iNOS, arginase, and and reactive oxygen species (ROS) in mouse models. Secretion of iNOS also inhibits NK cells, thereby reducing antibody-dependent cell-mediated cytotoxicity (ADCC) (57). In a clinical setting, patients with non-small cell lung carcinoma (NSCLC), circulating tyrosine kinase receptor TIE2^{hi} expressing M-MDSCs were found to suppress antigen-specificT cell responses and their presence was linked to poor patient outcomes (58). In contrast, patients with NSCLC treated with anti-PD-1 checkpoint blockade that had lower frequencies of both M-MDSCs and PMN-MDSCs had longer overall progression free survival (59).

PMN-MDSCs comprise the majority of MDSC populations (60). These PMN-MDSC are metabolically distinctive from mature neutrophils and promote early tumor dissemination and establishment of the pre-metastatic niche in the lungs and other sites (61). They also migrate more effectively and exert significantly greater immune suppressive activity compared to normal neutrophils. Mechanisms by which PMN-MDSCs inhibit T cell function include reactive nitrogen intermediates in mice and dogs, and ROS in humans (60, 62). Patients with primary and metastatic lung cancers exhibited high numbers of tumor infiltrating PMN-MDSCs, which was associated with suppressed NK cell activation and cytolytic activity, thought to be mediated by both cell-cell contact with PMN-MDSC and production of soluble factors within the TME (54).

2.5 Immune modulative TANs in the tumor microenvironment

Immune suppressive TANs and PMN-MDSCs are recruited to the TME primarily by the chemokine CXCL8, which signals *via* the chemokine receptors CXCL1 and CXCL2 (63). Given their common origins in the bone marrow and their shared need for growth factors and cytokines such as G-CSF, IL-6, and IL-17, it is

somewhat difficult to definitely distinguish TANs from PMN-MDSCs (47, 63). Within tumor tissues, TANs are classified as either N1 or N2 populations, analogous to M1 and M2 macrophages (64). Populations of N1 TANs exhibit antitumor activity, whereas N2 TANs inhibit T cell proliferation and promote tumor growth (65). TGF- β secreted by tumor cells is one mechanism that polarizes TANs to the N2 phenotype (64, 66, 67). Within the TME, N2 TANs promote angiogenesis and play a role in establishing the pre-metastatic niche through secretion of VEGF and by expression of metalloproteinase-9 (MMP-9) which decreases the bioavailability of anti-angiogenic molecules (68, 69). High circulating numbers of N2 TANs have been associated with increased tumor metastatic progression, and genetic instability in tumors including melanoma and bronchioloalveolar carcinoma (70-72). Depletion of N2 TANs in animal models leads to increased numbers of effector CD8 T cells (73)' and promotes their infiltration into the tumor (63). Secretion of TGF- β and IL-10 by N2 TANs stimulates MDSC expansion, further augmenting the immune suppressive nature of the TME. Importantly, the mechanisms by which N2 TANs suppress tumor immunity may in many cases be distinct and different in mice versus humans (74).

In addition to suppressing T cells, both PMN-MDSC and N2 TANs produce neutrophil extracellular traps (NETs). These complex webs, comprised of extruded DNA molecules studded with chromatin and other nuclear proteins, can promote tumor metastasis by trapping migrating tumor cells within tumor blood vessels, and then facilitating the survival of these early metastatic tumor cells (75, 76). The NETS secreted by TANs and PMN-MDSC also interrupt the cytotoxic activities of CD8+ T cells and NK cells (77).

3 Therapeutic targeting of immune suppressive macrophages and MDSC

3.1 Direct depletion of myeloid cells (TAM, MDSC) in the TME

Depletion of immune suppressive myeloid cell populations within the TME is one method to overcome the immune suppressive pressure they exert, particularly given that these cells, especially TAMs, can be quite numerous in the TME, in some cases comprising over 50% of the entire tumor cell population (78). Below we provide examples of the multiple strategies designed to deplete TME populations of immune suppressive TAMs.

3.1.1 Colony-stimulating factor 1 receptor blockade

One approach that has been extensively investigated is TAM depletion *via* blocking signaling by the essential macrophage growth factor receptor colony-stimulating factor 1 receptor (CSF-1R). CSF-1R is expressed by TAMs and binds to the primary ligands CSF-1 and IL-34 (79). CSF-1R signaling is crucial to macrophage differentiation and survival (79, 80). The density of CSF-1R+ TAMs in tumors correlates with poor outcomes in many tumor types,

including colon adenocarcinoma, pancreatic cancer, classical Hodgkin lymphoma, leiomyosarcoma, hepatocellular carcinoma and breast cancer (81-86). CSF-1R is also expressed by other immune cells within the TME such as DC, MDSCs and neutrophils, and blocking CSF-1R signaling may therefore also deplete these cells in addition to TAMs (87). Disruption of CSF-1R signaling has been achieved by use of small molecules and monoclonal antibodies (87, 88). Multiple clinical trials are ongoing to evaluate the effects of CSF-1/CSF-1R blockade on TAM populations and tumor control in many both solid tumors and hematologic cancers (Table 1). To date clinical trials for CSF-1/ CSF-1R blockade have been completed in non-pancreatic neuroendocrine tumors (89), Hodgkin lymphoma (90), ovarian cancer (NCT03166891, NCT03901118), non-Hodgkin lymphoma (NCT03974243) and hepatocellular carcinoma (NCT03245190). The accumulated results from these trials indicates safety and tolerability of the CSF-1R inhibitors, but limited efficacy, suggesting either refined dosage or timing of CSF-1/CSF-1R blockade, or the need to employ with other combination therapies (91).

3.1.1.1 Pexidartinib (PLX3397, TURALIO)

The small molecule drug PLX3397 targets CSF1R signaling and reprograms intra-tumoral immune suppressive myeloid cells (92), and has been shown to convert immune suppressive M-DSCs to a more proinflammatory tumoricidal phenotype (93, 94). PLX3397was approved by the FDA in 2019 for use in the treatment of diffuse type tenosynovial giant cell tumors (dt-TGCT), a rare and often unresectable non-life-threatening cancer of the tendon sheath that is driven by CSF-1 expressing TAMs (95). CSF-1 activation in dt-TGCT leads to recruitment of CSF-1R+ macrophages which make up a large bulk of the tumor mass (96). This specific tumor type is well-suited for targeting by CSF-1/CSF-1R pathway blockade; and treatment with anti-CSF-1R antibodies has shown significant reduction of CSF-1R+ TAMs within tumor tissues (97). In a phase III double blind clinical trial, 14.8% of patients with unresectable dt-TGCT treated with PLX3397 had a complete response (CR) and 24.6% had a partial response (PR) per RECIST criteria compared to zero response in the placebo control group (98).

Current clinical trials are investigating the effectiveness of PLX3397 in multiple cancer types including melanoma, prostate cancer, recurrent glioblastoma multiforme (GBM) and hematological malignancies (99–101). Preclinical use of orally administered PLX3397 for the treatment of recurrent GBM in phase II trials did not show statistically significant improvement in progression free survival of patients compared to historical controls, and there were no partial or complete responses observed in their 38-patient cohort (92, 102). In a phase II trial with 20 patients with relapsed Hodgkin lymphoma, single agent PLX3397 treatment showed an objective overall response rate (ORR) of 5% (103). Thus, the value of CSF-1R inhibition alone for treatment of tumors such as GBM may be limited.

Trials investigating the use of PLX3397 in combination with other agents are ongoing breast cancer (NCT01042379) and

unresectable sarcomas and malignant peripheral nerve sheath tumors (NCT02584647). A Phase II trial in patients with advanced melanoma and other solid tumors in combination with PD-1 blocking antibody pembrolizumab (NCT02452424) was terminated early due to insufficient evidence of clinical efficacy (101).

3.1.1.2 Chiauranib (CS2164)

The small molecule drug chiauranib is a CSF-1R inhibitor that also selectively inhibits kinases related to angiogenesis, including VEGF, PDGFR, and c-kit (104). Chiauranib binds to the ATP site in VEGFR2 and inhibits kinase activity, as well as reducing phosphorylation of ERK1/2, thus decreasing expression of genes related to tumor angiogenesis. Chiauranib has shown efficacy in preclinical mouse models of hepatocellular carcinoma, colorectal cancer, and non-Hodgkin lymphoma (NHL) (105–107). Initial dose escalation trials demonstrated that 67% of patients achieved stable disease, with acceptable safety and tolerability (104). Current clinical trials are ongoing, with one phase II trial reported currently in the US (NCT05271292), evaluating chiauranib as a single agent to treat advanced solid malignant tumors.

3.1.1.3 Additional small molecule inhibitors of CSF-1R

Other small molecule CSF-1R inhibitors include ARRY-382, PLX7486, BLZ945 and JNJ-40346527 (edicotinib), and all are currently being evaluated in clinical trials for treatment of Hodgkin lymphoma (cHL) (87). A phase I study with ARRY382 for treatment of advanced solid tumors showed 15% stable disease with no objective responses observed out of 26 patients when administered in combination withpembrolizumab (NCT02880371). Phase I and II clinical studies of the drug JNJ-40346527 in patients with refractory Hodgkin lymphoma found that 11 of 20 patients (55.0%) had stable disease (SD) with progression free survival (PFS) times for all treated patients ranging from 2 days to 352 days (90).

3.1.1.4 Monoclonal antibodies targeting CSF-1 or CSF-R1

Monoclonal antibodies targeting CSF-1R in clinical development include emactuzumab, AMG820, IMC-CS4, cabiralizumab, MCS110 (lacnotuzumab) and PD036324 (Table 2). MCS110 and PD036324 target the CSF-1 (ligand) as opposed to the CSF-R1 receptor (87). Phase Ia/Ib trials with emactuzumab as either a single agent or in combination with paclitaxel in patients with metastatic solid tumors including mesothelioma, soft tissue sarcoma, ovarian, breast, pancreatic, endometrial cancer and dt-TGCT have been conducted. Study outcomes in the monotherapy group did not reveal any patients with objective tumor responses, with 13% of patients exhibiting SD. When administered in combination with paclitaxel, 7% of patients had PR with 43% showing SD (108). This study also demonstrated a significant reduction in the numbers of CSF-1R+ TAMs in both monotherapy and combination groups (101). The first human trial of AMG820 showed increased serum CSF-1 concentrations and decreased numbers of macrophages (109). Patients with relapsed or refractory advanced solid tumors treated with AMG820 experienced a 32% SD rate, while one patient with NSCLC

CSF-1/ CSF-1R blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
1	NCT03158103	A Study of MEK162 (Binimetinib) in Combination With Pexidartinib in Patients With Advanced Gastrointestinal Stromal Tumor (GIST)	Completed	No Results Available	Gastrointestinal Stromal Tumor (GIST)	MEK162 Pexidartinib	Phase 1	3
2	NCT02390752	Phase I Trial of Turalio (R) (Pexidartinib, PLX3397) in Children and Young Adults With Refractory Leukemias and Refractory Solid Tumors Including Neurofibromatosis Type 1 (NF1) Associated Plexiform Neurofibromas (PN)	Recruiting	No Results Available	Neurofibroma, Plexiform Precursor Cell Lymphoblastic Leukemia-Lymphoma Leukemia, Promyelocytic, Acute Sarcoma	TURALIO	Phase 1	54
3	NCT04635111	A Long-term Study Evaluating Hepatotoxicity Associated With TURALIO (Pexidartinib) Treatment	Recruiting	No Results Available	Hepatotoxicity Tenosynovial Giant Cell Tumor	TURALIO		30
4	NCT02371369	Phase 3 Study of Pexidartinib for Pigmented Villonodular Synovitis (PVNS) or Giant Cell Tumor of the Tendon Sheath (GCT- TS)	Completed	CR 24.2%, PR 29.7%	Pigmented Villonodular Synovitis Giant Cell Tumors of the Tendon Sheath Tenosynovial Giant Cell Tumor	Pexidartinib Placebo	Phase 3	120
5	NCT04526704	Study to Evaluate Discontinuation and Re- Treatment in Participants With Tenosynovial Giant Cell Tumor (TGCT) Previously Treated With Pexidartinib	Active, not recruiting	No Results Available	Tenosynovial Giant Cell Tumor	Pexidartinib	Phase 4	32
6	NCT02401815	CGT9486 (Formerly Known as PLX9486) as a Single Agent and in Combination With PLX3397 (Pexidartinib) or Sunitinib in Participants With Advanced Solid Tumors	Completed	No Results Available	Gastrointestinal Stromal Tumors	PLX9486 Pexidartinib Sunitinib	Phase 1 Phase 2	51
7	NCT01349036	A Phase 2 Study of PLX3397 in Patients With Recurrent Glioblastoma	Terminated	Surgical Cohort 1, Overall survival 76.9%. SD 23.1%, PD 76.9%. Non- Surgical Cohort 2 Overall survival 95.5%	Recurrent Glioblastoma	PLX3397	Phase 2	38

TABLE 1 US clinical trials in cancer using Colony-stimulating factor 1 receptor (CSF-1R) blockade as intervention.

(Continued)

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CSF-1/ CSF-1R blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
8	NCT02452424	A Combination Clinical Study of PLX3397 and Pembrolizumab To Treat Advanced Melanoma and Other Solid Tumors	Terminated	no CR in any dose escalation. PR up to 15.4% in Melanoma	Melanoma Non-small Cell Lung Cancer Squamous Cell Carcinoma of the Head and Neck Gastrointestinal Stromal Tumor (GIST) Ovarian Cancer	PLX3397 Biological: Pembrolizumab	Phase 1 Phase 2	78
9	NCT01790503	A Phase 1b/2 Study of PLX3397 + Radiation Therapy + Temozolomide in Patients With Newly Diagnosed Glioblastoma	Completed	CR + PR up to 18.2%, SD up to 54.5%	Patients With Newly Diagnosed Glioblastoma	PLX3397 Radiation: Radiation Therapy Temozolomide	Phase 1 Phase 2	65
10	NCT01525602	Safety Study of PLX3397 and Paclitaxel in Patients With Advanced Solid Tumors	Completed	Clinical benefit rate (CR, PR, or stable disease) 33~ 67%	Solid Tumors	PLX3397 Paclitaxel	Phase 1	74
11	NCT05271292	Chiauranib for Advanced Solid Malignant Tumors and Relapsed/Refractory SCLC.	Recruiting	No Results Available	Small-cell Lung Cancer Advanced Solid Malignant Tumor	Chiauranib	Phase 1 Phase 2	36
12	NCT01316822	A Study of ARRY-382 in Patients With Selected Advanced or Metastatic Cancers	Completed	No Results Available	Metastatic Cancer	ARRY-382, cFMS inhibitor; oral	Phase 1	26
13	NCT02880371	A Study of ARRY-382 in Combination With Pembrolizumab for the Treatment of Patients With Advanced Solid Tumors	Terminated	Phase 1b, 10.5% had confirmed PR, in phase 2, 3.7% with PDA had a PR lasting 2.4 months.	Advanced Solid Tumors	ARRY-382 Pembrolizumab	Phase 1 Phase 2	82
14	NCT01804530	Phase 1 Study of PLX7486 as Single Agent in Patients With Advanced Solid Tumors	Terminated	No Results Available	Solid Tumor Tumors of Any Histology With Activating Trk (NTRK) Point or NTRK Fusion Mutations Tenosynovial Giant Cell Tumor	PLX7486 TsOH	Phase 1	59
15	NCT02829723	A Study of BLZ945 Single Agent or BLZ945 in Combination With PDR001 in Advanced Solid Tumors	Terminated	No Results Available	Advanced Solid Tumors	BLZ945 PDR001	Phase 1 Phase 2	198
16	NCT03557970	JNJ-40346527 in Treating Participants With Relapsed or Refractory Acute Myeloid Leukemia	Terminated	55.0% SD, 40.0% PD. PFS for all treated patients ranged from 2 days to 352+ days.	Recurrent Acute Myeloid Leukemia Refractory Acute Myeloid Leukemia	Drug: Edicotinib Other: Pharmacokinetic Study		

CSF-1/ CSF-1R blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
17	NCT03177460	Daratumumab or FMS Inhibitor JNJ-40346527 Before Surgery in Treating Patients With High-Risk, Resectable Localized or Locally Advanced Prostate Cancer	Active, not recruiting	No Results Available	Prostate Adenocarcinoma Stage III Prostate Cancer AJCC v8 Stage IIIA Prostate Cancer AJCC v8 Stage IIIB Prostate Cancer AJCC v8 Stage IIIC Prostate Cancer AJCC v8 Testosterone Greater Than 150 ng/ dL	Biological: Daratumumab Drug: FMS Inhibitor JNJ- 40346527 Procedure: Radical Prostatectomy		

experienced a PR. All the agents in trials targeting CSF-R1 have generally been well-tolerated to date, suggesting that sustained CSF-1R blockade treatment for weeks to months is safe. However, to date none of the CSF-1 or CSF-1R targeted agents has demonstrated significant antitumor activity clinically (110).

3.1.2 Trabectedin as myeloid cell depleting chemotherapy

Trabectedin an alkaloid drug that binds a minor groove of DNA and blocks the cell cycle and DNA repair pathways (111). It has been shown to selectively reduce TAMs in tumors without affecting the infiltration of T cells (112). Treatment with trabectedin also inhibits local differentiation of monocytes into TAMs (113). Use of trabectedin in multiple preclinical animal tumor models demonstrated depletion of TAMs and reduction of tumor growth, suppression of angiogenesis, and reduced concentrations of IL6, CCL2 and CXCL8 (114). Current phase II clinical trials of trabectedin are ongoing for treatment of soft tissue sarcoma, bone tumors and small round-cell sarcomas, administered in combination with low-dose radiation therapy (NCT05131386). Trabectedin is FDA approved for treatment of unresectable or metastatic liposarcoma and leiomyosarcoma (115) (Table 3).

3.2 Chemokine receptor antagonists for monocyte and neutrophil migration inhibition

Chemokine receptor antagonists can reduce the infiltration of monocytes and MDSCs into the TME. The chemokine CCL2 binds to the receptor CCR2 expressed on inflammatory monocytes (116), which signals to circulating monocytes to promote extravasation from the vasculature and into inflamed tissues (32). Many tumors secrete large amounts of CCL2, thereby recruiting circulating inflammatory monocytes into tumor tissues where they then differentiate into M2 TAMs (32, 117, 118). CCL2 may also play a minor role in PMN-MDSC recruitment, though the primary chemokine driving TAN recruitment is CXCL8 (119). There have been numerous preclinical studies in rodent models assessing inhibitors of the CCL2-CCR2 axis using either small molecule CCL2 inhibitors or monoclonal antibodies, and most have demonstrated inhibition of tumor growth and/or decreased metastatic burden (120). In these models, CCL2-CCR2 signaling blockade has been shown to suppress tumor growth through multiple pathways including depletion of TAMs and M-MDSC and increasing infiltrating T cells (32, 118, 120).

3.2.1 CCR2 targeted antibodies

Carlumab (CNTO888) is a CCL2 neutralizing antibody that has been evaluated in multiple cancer models as either a single agent immunotherapy or in combination with chemotherapy (121). Preclinical mouse models evaluating carlumab have demonstrated increased IFN γ production by NK cells and antitumoral CD8+ T cells when combined with anticancer vaccines (122). Carlumab has demonstrated positive clinical responses when used in combination with chemotherapeutic drug docetaxel (123); phase II trials have been completed but Carlumab has since been discontinued (NCT00992186).

MLN1202 (plozalizumab) is a CCR2 blocking monoclonal antibody currently undergoing phase II clinical trials for treatment of metastatic bone cancer (NCT01015560). Results so far show that MLN1202 is relatively well tolerated with only 7.14% of patients experiencing severe adverse events (SAE). A phase I trial of MLN1202 in combination with nivolumab was terminated early due to serious adverse events (NCT02723006), which may suggest limited potential for MLN1202 as single or combined immunotherapy agent (124, 125) (Table 4).

3.2.2 CCL2 inhibitors

Bindarit is a small molecule drug that inhibits the synthesis of CCL2 and has been shown to induce tumor regression in preclinical studies by inhibiting TAM and MDSC infiltration of the TME in breast cancer, prostate cancer, and osteosarcoma animal models (126–129). A second CCL2 inhibitor mNOX-36 has been shown in a rat model of GBM to significantly inhibit tumor growth (130). The safety of mNOX-36 is currently being evaluated in Phase I trials (Table 4).

3.2.3 CCR2 inhibitors

RS 504393 is a small molecule CCR2 antagonist that has shown activity in blocking M-MDSCs and TAM recruitment into tumors following gemcitabine treatment in a mouse model of bladder cancer (131). Another CCR2 inhibitor (BMS CCR2 22), is a high

TABLE 2 US clinical trials in cancer using Monoclonal antibodies targeting CSF-1 or CDF-1R.

CSF-1/CSF- 1R mono- clonal anti- bodies	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
1	NCT05417789	Study of Emactuzumab for Tenosynovial Giant Cell Tumor (TGCT)	Active, not recruiting	No Results Available	TGCT	Drug: Emactuzumab Drug: Placebo	Phase 3	128
2	NCT03369964	A Study of Atezolizumab in Combination With an Immunotherapy Agent Investigated With or Without Anti- Cd20 Therapy in Patients With Relapsed or Refractory Non- Hodgkin Lymphoma	Withdrawn	No Results Available	Lymphoma, Non- Hodgkin	Drug: Atezolizumab Drug: Emactuzumab Drug: Obinutuzumab	Phase 1	0
3	NCT02760797	A Study of Emactuzumab and RO7009789 Administered in Combination in Participants With Advanced Solid Tumors	Completed	No Results Available	Neoplasms	Drug: Emactuzumab Drug: RO7009789	Phase 1	38
4	NCT02323191	A Study of Emactuzumab and Atezolizumab Administered in Combination in Participants With Advanced Solid Tumors	Completed	No Results Available	Solid Cancers	Drug: Atezolizumab Drug: Emactuzumab	Phase 1	221
5	NCT02923739	Paclitaxel and Bevacizumab With or Without Emactuzumab in Treating Patients With Platinum- Resistant Ovarian, Fallopian Tube, or Primary Peritoneal Cancer	Completed	No Results Available	Fallopian Tube Adenocarcinoma Fallopian Tube Clear Cell Adenocarcinoma Fallopian Tube Endometrioid Adenocarcinoma Fallopian Tube Mucinous Adenocarcinoma Fallopian Tube Serous Adenocarcinoma Fallopian Tube Serous Adenocarcinoma Cell Carcinoma Fallopian Tube Undifferentiated Carcinoma Fallopian Tube Undifferentiated Carcinoma Malignant Ovarian Brenner Tumor Ovarian Adenocarcinoma Ovarian Endometrioid Adenocarcinoma Ovarian Mucinous Adenocarcinoma Ovarian	Biological: Bevacizumab Biological: Emactuzumab Other: Laboratory Biomarker Analysis Drug: Paclitaxel Other: Pharmacological Study	Phase 2	9

CSF-1/CSF- 1R mono- clonal anti- bodies	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
					Carcinoma Ovarian Serous Adenocarcinoma Ovarian Transitional Cell Carcinoma Ovarian Undifferentiated Carcinoma Primary Peritoneal Serous Adenocarcinoma Recurrent Fallopian Tube Carcinoma Recurrent Ovarian Carcinoma Recurrent Primary Peritoneal Carcinoma			
6	NCT01494688	A Study of RO5509554 as Monotherapy and in Combination With Paclitaxel in Participants With Advanced Solid Tumors	Completed	No Results Available	Advanced Solid Tumors	Drug: Paclitaxel Drug: RO5509554	Phase 1	217
7	NCT01444404	A Study of AMG 820 in Subjects With Advanced Solid Tumors	Completed	No Results Available	Advanced Malignancy Advanced Solid Tumors	Drug: AMG 820	Phase 1	25
8	NCT02713529	Safety and Efficacy Study of AMG 820 and Pembrolizumab Combination in Select Advanced Solid Tumor Cancer	Completed	Objective Response Rate (ORR) up tp 5.3%, highest OS 75% at. 6 months and 41.7% at 12 months	Pancreatic Cancer Colorectal Cancer Non- Small Cell Lung Cancer	Biological: AMG820 and pembrolizumab	Phase 1 Phase 2	117
9	NCT01346358	A Study of IMC- CS4 in Subjects With Advanced Solid Tumors	Completed	No Results Available	Neoplasms	Biological: IMC- CS4	Phase 1	72
10	NCT03153410	Pilot Study With CY, Pembrolizumab, GVAX, and IMC- CS4 (LY3022855) in Patients With Borderline Resectable Adenocarcinoma of the Pancreas	Active, not recruiting	No Results Available	Pancreatic Cancer	Drug: Cyclophosphamide Drug: GVAX Drug: Pembrolizumab Drug: IMC-CS4	Early Phase 1	12
11	NCT02265536	A Study of LY3022855 In Participants With Breast or Prostate Cancer	Completed	No Results Available	Neoplasms Neoplasm Metastasis	Drug: LY3022855	Phase 1	36

CSF-1/CSF- 1R mono- clonal anti- bodies	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
12	NCT03697564	Nivolumab + Cabiralizumab + Gemcitabine in Patients With Stage IV Pancreatic Cancer Achieving Disease Control in Response to First- line Chemotherapy (GemCaN Trial).	Suspended	No Results Available	Pancreatic Cancer Stage IV	Drug: Gemcitabine Drug: Nivolumab 10 MG/ML Intravenous Solution [OPDIVO]]Drug: Cabiralizumab	Phase 2	40
13	NCT03502330	APX005M With Nivolumab and Cabiralizumab in Advanced Melanoma, Non- small Cell Lung Cancer or Renal Cell Carcinoma	Active, not recruiting	No Results Available	Advanced Melanoma Non-small Cell Lung Cancer Renal Cell Carcinoma	Drug: APX005M Drug: Cabiralizumab Drug: Nivolumab	Phase 1	42
14	NCT04848116	Neoadjuvant Targeting of Myeloid Cell Populations in Combination With Nivolumab in Head & Neck Cancer	Recruiting	No Results Available	Head and Neck Squamous Cell Carcinoma	Drug: Nivolumab Drug: HuMax-IL8 Drug: Cabiralizumab	Phase 2	24
15	NCT03927105	Nivolumab and the Antagonistic CSF-1R Monoclonal Antibody Cabiralizumab (BMS-986227) in Patients With Relapsed/ Refractory Peripheral T Cell Lymphoma	Active, not recruiting	2 paitients 4 month CR, 1NR,	Peripheral T Cell Lymphoma	Drug: Nivolumab Drug: cabiralizumab	Phase 2	4
16	NCT03431948	Stereotactic Body Radiotherapy (SBRT) Plus Immunotherapy for Cancer	Completed	No Results Available	Cancer	Drug: Nivolumab Drug: Cabiralizumab Drug: Urelumab Radiation: Stereotactic Body Radiation Therapy	Phase 1	60
17	NCT04050462	Nivolumab Combined With BMS-986253 in HCC Patients	Active, not recruiting	No Results Available	Hepatocellular Carcinoma	Drug: Nivolumab 240 mg IV every 2 weeks + Cabiralizumab 4 mg/kg IV every 2 weeks Drug: Nivolumab 240 mg IV every 2 weeks + BMS-986253 1200 mg IV every 2 weeks Drug: Nivolumab 240 mg IV every 2 weeks	Phase 2	23

CSF-1/CSF- 1R mono- clonal anti- bodies	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
18	NCT04331067	Neoadjuvant Nivolumab and Chemotherapy in Patients With Localized Triple- negative Breast Cancer	Recruiting	No Results Available	Triple Negative Breast Cancer	Drug: Paclitaxel Drug: Carboplatin Biological: Nivolumab Biological: Cabiralizumab Procedure: Tumor biopsy Procedure: Bone marrow Procedure: Blood draw	Phase 1 Phase 2	31
19	NCT02526017	Study of Cabiralizumab in Combination With Nivolumab in Patients With Selected Advanced Cancers	Completed	highest OS group 13 months , highest PFS 2.9 months	Advanced Solid Tumors Head and Neck Cancer Pancreatic Cancer Ovarian Cancer Renal Cell Carcinoma Malignant Glioma Non- small Cell Lung Cancer	Biological: Cabiralizumab Biological: Nivolumab	Phase 1	313
20	NCT02471716	Study of Cabiralizumab in Patients With Pigmented Villonodular Synovitis / Diffuse Type Tenosynovial Giant Cell Tumor	Completed	ORR up to 33%	Pigmented Villonodular Synovitis Tenosynovial Giant Cell Tumor	Biological: FPA008	Phase 1 Phase 2	66
21	NCT03336216	A Study of Cabiralizumab Given With Nivolumab With and Without Chemotherapy in Patients With Advanced Pancreatic Cancer	Active, not recruiting	No Results Available	Advanced Pancreatic Cancer	Biological: Cabiralizumab Drug: Nab- paclitaxel Drug: Onivyde Biological: Nivolumab Drug: Fluorouracil Drug: Gemcitabine Drug: Oxaliplatin Drug: Leucovorin Drug: Irinotecan Hydrochloride	Phase 2	202
22	NCT03335540	An Adaptive Study to Match Patients With Solid Tumors to Various Immunotherapy Combinations Based Upon a Broad Biomarker Assessment	Completed	No Results Available	Advanced Cancer	Biological: Nivolumab Biological: Relatlimab Biological: Cabiralizumab Biological: Ipilimumab Drug: IDO1 Inhibitor Radiation: Radiation Therapy	Phase 1	20
23	NCT03455764	MCS110 With BRAF/MEK Inhibition in Patients With Melanoma	Active, not recruiting	No Results Available	Melanoma	Drug: MCS110 Drug: Dabrafenib Drug: Trametinib	Phase 1 Phase 2	43
24	NCT01643850	MCS110 in Patients With Pigmented	Completed	decrease in tumor size	Pigmented Villonodular Synovitis PVNS Giant Cell Tumor of the Tendon Sheath GCCTS	Drug: MCS110 Drug: Placebo	Phase 2	36

CSF-1/CSF- 1R mono- clonal anti- bodies	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
		Villonodular Synovitis (PVNS)			Tenosynovial Giant Cell Tumor Localized or Diffused Type GCTS			
25	NCT02807844	Phase Ib/II Study of MCS110 in Combination With PDR001 in Patients With Advanced Malignancies	Completed	Clinical Benefit Rate up to 20%	Triple Negative Breast Cancer Pancreatic Carcinoma Melanoma Endometrial Carcinoma	Drug: MCS110 Drug: PDR001	Phase 1 Phase 2	141
26	NCT02435680	Efficacy Study of MCS110 Given With Carboplatin and Gemcitabine in Advanced Triple Negative Breast Cancer (TNBC)	Completed	PFS average 5.6 months, SD up tp 55.9%, ORR up to 37.5%	Advanced Triple Negative Breast Cancer (TNBC) With High TAMs	Drug: MCS110 Drug: carboplatin Drug: gemcitabine	Phase 2	50
27	NCT03742349	Study of Safety and Efficacy of Novel Immunotherapy Combinations in Patients With Triple Negative Breast Cancer (TNBC).	Active, not recruiting	No Results Available	Triple Negative Breast Cancer (TNBC)	Biological: spartalizumab Biological: LAG525 Drug: NIR178 Drug: capmatinib Biological: MCS110 Biological: canakinumab	Phase 1	64
29	NCT02554812	A Study Of Avelumab In Combination With Other Cancer Immunotherapies In Advanced Malignancies (JAVELIN Medley)	Active, not recruiting	No Results Available	Advanced Cancer	Drug: Avelumab Drug: Utomilumab Drug: PF-04518600 Drug: PD 0360324 Drug: CMP-001	Phase 1 Phase 2	398

affinity CCR2 antagonist that decreases TAM density as demonstrated in mouse metastatic hepatic cancer models. When combined with FOLFOX (folinic acid, fluorouracil oxaliplatin) chemotherapy regimine, administration of BMS CCR2 22 significantly increased efficacy and improved overall survival in mice with colon adenocarcinomas (117, 118). A third CCR2 antagonist, 747 is a natural product derived from the tree *Abies georgei* (132). The drug 747 is considered a selective CCR2 antagonist and has been shown to inhibit TAM recruitment and increase density of CD8+ tumor infiltrating lymphocytes as well as increase inflammatory cytokines such as IFN- γ in rodent mode. Treatment with 747 also increased tumor apoptosis when combined with sorafenib, a tyrosine kinase inhibitor, thereby potentiating antitumor activity by depleting TAMs (133).

A fourth selective CCR2 inhibitor (PF-04136309) has demonstrated antitumor activity in an orthotopic mouse model of pancreatic cancer (134). Phase Ib clinical trials in patients with pancreatic cancer evaluated treatment with PF-04136309 in combination with the chemotherapy regimen FOLFIRINOX (folinic acid, fluorouracil, irinotecan hydrochloride, and oxaliplatin) and demonstrated a 49% response rate, compare to no responding patients treated with FOLFIRINOX alone. In addition, administration of PF-04136309 in combination with FOLFIRINOX significantly decreased the numbers of CCR2+ monocytes in bone marrow samples, compared to FOLFIRINOX alone treated patients (135). A phase I study in patients with metastatic pancreatic cancer found that PF-04136309 given in combination with chemotherapy gemcitabine and nab-paclitaxel significantly decreased CD14+CCR2+ monocytes in circulation. However, the high incidence of pulmonary toxicity in patients treated with PF-04136309 led to a discontinuation of further clinical evaluation (136).

TABLE 3 US clinical trials using Trabectedin as myeloid cell depleting chemotherapy for cancer.

Myeloid targeted chemotherapy	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
1	NCT03886311	Talimogene Laherparepvec, Nivolumab and Trabectedin for Sarcoma	Recruiting	No Results Available	Sarcoma	Drug: Talimogene Laherparepvec 10000000 PFU/1 ML Injection Suspension [IMLYGIC] Drug: Nivolumab IV Soln 100 MG/10ML Drug: Trabectedin 0.25 MG/ 1 VIAL Intravenous Powder for Solution	Phase 2	40
2	NCT04535271	Metronomic Trabectedin, Gemcitabine, and Dacarbazine for Leiomyosarcoma	Recruiting	No Results Available	Leiomyosarcoma	Drug: Trabectedin	Phase 2	80
3	NCT04076579	Trabectedin in Combination With Olaparib in Advanced Unresectable or Metastatic Sarcoma	Active, not recruiting	No Results Available	Sarcoma Sarcoma Metastatic	Drug: Olaparib Drug: Trabectedin	Phase 2	29
4	NCT00072670	A Phase 2 Study of Trabectedin (Yondelis) in Adult Male Participants With Advanced Prostate Cancer	Completed	No Results Available	Prostate Cancer	Drug: Trabectedin	Phase 2	59
5	NCT03074318	Avelumab and Trabectedin in Treating Patients With Liposarcoma or Leiomyosarcoma That is Metastatic or Cannot Be Removed by Surgery	Terminated	up to 18.8% PR, 66.7% SD at 12 weeks, clinical benefit rate 56%. OS highest group average 416 days	Metastatic Leiomyosarcoma Metastatic Liposarcoma Unresectable Leiomyosarcoma Unresectable Liposarcoma	Drug: Avelumab Drug: Trabectedin	Phase 1 Phase 2	35
6	NCT03138161	SAINT: Trabectedin, Ipilimumab and Nivolumab as First Line Treatment for Advanced Soft Tissue Sarcoma	Recruiting	No Results Available	Advanced Soft Tissue Sarcoma Metastatic Soft Tissue Sarcoma	Drug: Trabectedin Drug: Ipilimumab Drug: Nivolumab	Phase 1 Phase 2	45
7	NCT00147212	ET 743 (Yondelis) in Men With Advanced Prostate Cancer	Completed	Prostate specific antigen (PSA) response rate 7/50	Prostate Cancer	Drug: ET 743	Phase 2	50

3.2.4 Dual CCR2/CCR5 inhibitor for myeloid cell targeting

BMS-813160 is a dual CCR2/CCR5 inhibitor which has been investigated in phase I and phase II trials as combination therapy

(137). In ongoing phase II clinical trials for treatment of NSCLC and hepatocellular cancer, BMS-813160 is being administered in combination with nivolumab and the anti-CXCL8 drug BMS-986253 (NCT04123379) (138). BMS-813260 is also being investigated in phase

TABLE 4 US clinical trials in cancer targeting CCR2 or CCR5 axis.

CCL2/ CCR2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
1	NCT01015560	S0916, MLN1202 in Treating Patients With Bone Metastases	Completed	7.14% SAE	Metastatic Cancer Unspecified Adult Solid Tumor, Protocol Specific	Drug: anti-CCR2 monoclonal antibody MLN1202 Genetic: polymorphism analysis Other: laboratory biomarker analysis	Phase 2	44
2	NCT02723006	Study to Evaluate the Safety, Tolerability, and Pharmacodynamics of Investigational Treatments in Combination With Standard of Care Immune Checkpoint Inhibitors in Participants With Advanced Melanoma	Terminated	up to 58.33% in arm 3 triple drug combo	Melanoma	Drug: TAK-580 Drug: TAK-202 Drug: vedolizumab Drug: nivolumab Drug: ipilimumab	Phase 1	22
3	NCT01413022	FOLFIRINOX Plus PF-04136309 in Patients With Borderline Resectable and Locally Advanced Pancreatic Adenocarcinoma	Completed	No Results Available	Pancreatic Neoplasms	Drug: Oxaliplatin Drug: Irinotecan Drug: Leucovorin Drug: Fluorouracil Other: laboratory biomarker analysis Other: flow cytometry Other: immunohistochemistry staining method Other: pharmacological study Drug: PF-04136309	Phase 1	44
4	NCT02732938	Ph1b/2 Study of PF- 04136309 in Combination With Gem/Nab-P in First- line Metastatic Pancreatic Patients	Terminated	PFS not reported, 11/ 17 SAE in arm 1b combination treatment	Metastatic Pancreatic Ductal Adenocarcinoma	Drug: PF-04136309 Drug: Nab-paclitaxel Drug: Gemcitabine	Phase 2	22
CCR2/ CCR5 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
1	NCT03184870	A Study of BMS- 813160 in Combination With Chemotherapy or Nivolumab in Participants With Advanced Solid Tumors	Active, not recruiting	No Results Available	Colorectal Cancer Pancreatic Cancer	Drug: BMS-813160 Biological: Nivolumab Drug: Nab-paclitaxel Drug: Gemcitabine Drug: 5-fluorouracil (5- FU) Drug: Leucovorin Drug: Irinotecan	Phase 1 Phase 2	332
2	NCT04123379	Neoadjuvant Nivolumab With CCR2/5-inhibitor or Anti-IL-8) for Non- small Cell Lung Cancer (NSCLC) or Hepatocellular Carcinoma (HCC)	Recruiting	No Results Available	Non-small Cell Lung Cancer Hepatocellular Carcinoma	Drug: Nivolumab Drug: BMS-813160 Drug: BMS-986253	Phase 2	50
3	NCT02996110	A Study to Test Combination Treatments in People With Advanced Renal Cell Carcinoma	Completed	ORR up tp 17.4%, PFS at 24 Weeks up to 46.8% in arm5	Advanced Cancer	Biological: Nivolumab Biological: Ipilimumab Biological: Relatlimab Drug: BMS-986205 Drug: BMS-813160	Phase 2	182

CCL2/ CCR2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
4	NCT03767582	Trial of Neoadjuvant and Adjuvant Nivolumab and BMS- 813160 With or Without GVAX for Locally Advanced Pancreatic Ductal Adenocarcinomas.	Recruiting	No Results Available	Locally Advanced Pancreatic Ductal Adenocarcinoma (PDAC) Pancreatic Ductal Adenocarcinoma	Radiation: Stereotactic Body Radiation (SBRT) Drug: Nivolumab Drug: CCR2/CCR5 dual antagonist Drug: GVAX	Phase 1 Phase 2	30
5	NCT03496662	BMS-813160 With Nivolumab and Gemcitabine and Nab- paclitaxel in Borderline Resectable and Locally Advanced Pancreatic Ductal Adenocarcinoma (PDAC)	Active, not recruiting	SAE up to 68.00% in dose expantion	Pancreatic Ductal Adenocarcinoma	Drug: BMS-813160 Drug: Nivolumab Drug: Gemcitabine Drug: Nab- paclitaxel Procedure: Biopsy Procedure: Peripheral blood	Phase 1 Phase 2	40

II trials for pancreatic ductal carcinoma and colorectal cancer, administered in combination with either nivolumab or chemotherapy (139) (Table 4).

3.2.5 Repurposed angiotensin receptor antagonists for CCR2 inhibition

Losartan, a type 1 angiotensin II receptor (AT1R) blocker (ARB), has been found to exert off-target activity as a potent, non-competitive CCR2 antagonist (140). In a mouse syngeneic breast cancer model, losartan suppressed lung metastatic tumor burden significantly (141). In this model, the reduced metastatic burden was associated with a significant decrease in CD11b+/Ly6C + monocytes recruited to the lungs (140). In studies in a dog model of metastatic osteosarcoma, the combination high dose losartan (10mg/kg PO BID) with the non-selective tyrosine kinase inhibitor toceranib demonstrated a response rate (PR) of 25% and clinical benefit rate of 50% (142). A similar phase I clinical trial is underway for pediatric osteosarcoma using the combination of losartan with the non-selective tyrosine kinase inhibitor sunitinib (NCT03900793). There are also multiple other clinical trials currently evaluating losartan in combination with radiation therapy and chemotherapy or immunotherapy. A phase II clinical trial of losartan in combination with nivolumab is currently underway in patients with localized pancreatic cancer (NCT03563248). In addition, losartan is being evaluated in combination with radiation therapy and chemotherapy in pancreatic cancer (NCT03563248, NCT04106856). A recent study also indicates that losartan treatment can reduce cerebral edema following immunotherapy in a rodent GBM model (143) (Table 5).

3.2.6 CXCR1 blockade to deplete PMN-MDSC and TANs

The chemokine CXCL8, which signals *via* CXCR1 and CXCR2, triggers the recruitment of PMN-MDSC and TANs into the TME (144). The receptors CXCR1 and CXCR2 are primarily expressed on neutrophils (145). CXCR1 is very selective for CXCL8, whereas

CXCR2 also binds other chemokines. Signaling by CXCR1 and CXCR2 are major mechanisms for recruiting neutrophils and PMN-MDSC into the TME which then differentiate into TANs or PMN-MDSCs (146). High expression of CXCL8 by tumors has been correlated with poor prognosis in many tumor types (147). Thus, CXCR1 and CXCR2 antagonists have been evaluated as strategies to deplete the TME of immune suppressive N2 TANs and PMN-MDSC (48, 148) (Table 6).

3.2.6.1 CXCR1 antagonist navarixin

The selective CXCR1 antagonist navarixin was originally developed for treatment of chronic obstructive pulmonary disease (COPD), asthma and psoriasis (149). A current phase II clinical trial of navarixin in combination with pembrolizumab is underway in patients with either PD-1 positive refractory NSCLC, castration resistant prostate cancer, or microsatellite stable colorectal cancer (NCT03473925) (150).

3.2.6.2 CXCR1 antagonist reparixin

Reparixin is a small molecule dual antagonist of both CXCR1 and CXCR2 (151, 152). Reparixin was originally evaluated as a drug to prevent graft rejection for pancreatic islet cells (153). *In vitro* studies with reparixin in thyroid cancer found that it also exhibits direct anti-tumor activity (154). In a phase I clinical trial in patients with HER-2 negative metastatic breast cancer, reparixin was well tolerated in combination with paclitaxel chemotherapy (155). However, phase II double blinded clinical trials in triple negative breast cancer patients demonstrated no improvement reparixin in combination with paclitaxel exhibited no additional clinical benefit compared to treatment with paclitaxel alone (NCT02370238) (156).

3.2.6.3 CXCR1/2 antagonist ladarixin

Ladarixin, like reparixin is a dual CXCR11/2 antagonist (157). Preclinical evaluation of ladarixin demonstrated significant activity in a mouse model of pancreatic ductal adenocarcinomaimproved activity compared to either agent

alone (148, 158). In an animal model of uveal melanoma administration of ladarixin repolarized TAMs to a M1 phenotype and inhibited tumor cell migration (157). Ladarixin has been used in clinical trials for diabetes, however clinical trials for cancers have not been reported.

3.2.6.4 CXCR2 antagonist AZD5069

AZD5069 is a highly selective small molecule antagonist of CXCR2 receptors that has been shown to inhibit neutrophil migration in patients with COPD (159). It is currently in clinical trials to deplete TANs in the TME in patients with metastatic pancreatic ductal adenocarcinoma and in relapsed metastatic squamous cell carcinoma of the head and neck in combination with ICI (160, 161). In addition, AZD5069 is being evaluated in combination with the androgen receptor antagonist enzalutamide in phase I/II trials in patients with metastatic castration resistant prostate cancer (mCRPC) in the UK (NCT03177187). The combination treatment was well tolerated with no dose limiting toxicities observed. The study observed that 2 out of 15 patients experienced a PR and 10 of 15 patients experienced SD, with responses lasting 2-16 months. Another trial demonstrated that AZD5069 has antitumor activity and depleted TAN density in patients with mCRPC (162).

3.2.6.5 Dual CXCR1/2 antagonist SX-682

SX-682 is another dual CXCR1/2 antagonist, which in rodent models of head and neck cancer have demonstrated suppression of PMN-MDSC accumulation and enhanced tumor infiltration with adoptively transferred NK cells (163, 164). SX-682 is currently being tested in phase I clinical trials in combination with ICI for metastatic melanoma (NCT03161431), and in phase II trials for pancreatic cancer, lung cancer, colon and rectal adenocarcinoma (NCT05604560, NCT05570825, NCT04599140).

3.3 CXCR4 blockade to inhibit tumor angiogenesis and metastases

Signaling by the chemokine receptor CXCR4 after binding the chemokine CXCL12 (SCF-1) triggers increased tumor proliferation, survival, and chemotaxis (165). Notably, CXCR4 is overexpressed by many different types of cancers, where it plays a role in tumor metastasis, and also a critical role in mobilizing and recruiting MDSC from bone marrow. Blockade of the CXCR4 signaling is hypothesized to not only decrease tumor angiogenesis but also decrease the number of cancer stem cells and increase mobilization and recruitment of effector T cells into the TME (166) (Table 7).

TABLE 5 US clinical trials using Repurposed angiotensin receptor antagonists for CCR2 inhibition in cancer.

Losartan CCL2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
1	NCT01821729	Proton w/ FOLFIRINOX- Losartan for Pancreatic Cancer	Unknown status	SAE 30.61%	Pancreatic Cancer	Drug: FOLFIRINOX Drug: Losartan Radiation: Proton Beam Radiation	Phase 2	50
2	NCT04106856	Losartan and Hypofractionated Rx After Chemo for Tx of Borderline Resectable or Locally Advanced Unresectable Pancreatic Cancer (SHAPER)	Recruiting	No Results Available	Borderline Resectable Pancreatic Adenocarcinoma Locally Advanced Pancreatic Ductal Adenocarcinoma Locally Advanced Unresectable Pancreatic Adenocarcinoma Stage II Pancreatic Cancer AJCC v8 Stage IIA Pancreatic Cancer AJCC v8 Stage IIB Pancreatic Cancer AJCC v8 Stage III Pancreatic Cancer AJCC v8	Radiation: Hypofractionated Radiation Therapy Drug: Losartan Drug: Losartan Potassium Other: Quality-of-Life Assessment Other: Questionnaire Administration	Phase 1	20
3	NCT05077800	FOLFIRINOX + 9-Ing-41 + Losartan In Pancreatic Cancer	Recruiting	No Results Available	Pancreatic Adenocarcinoma Pancreatic Adenocarcinoma Metastatic	Drug: FOLFIRNINOX Drug: Losartan Drug: 9-ING-41	Phase 2	70
4	NCT05365893	PHL Treatment in Pancreatic Cancer	Recruiting	No Results Available	Pancreatic Ductal Adenocarcinoma	Combination Product: Paricalcitol, Hydroxychloroquine, Losartan Other: Neoadjuvant therapy	Early Phase 1	20

Losartan CCL2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
						and surgery only (Control)		
5	NCT01234922	Benazepril Hydrochloride, Lisinopril, Ramipril, or Losartan Potassium in Treating Hypertension in Patients With Solid Tumors	Terminated	Protocol was closed early due to slow accrual, no SAE observed	Hypertension Unspecified Adult Solid Tumor, Protocol Specific	Drug: lisinopril Drug: losartan potassium Other: laboratory biomarker analysis Drug: benazepril hydrochloride Drug: ramipril	Phase 2	6
6	NCT01276613	Tissue Pharmacokinetics of Intraoperative Gemcitabine in Resectable Adenocarcinoma of the Pancreas	Terminated	No Results Available	Pancreatic Cancer	Drug: Gemcitabine Drug: Losartan	Early Phase 1	18
7	NCT04539808	NeoOPTIMIZE: Early Switching of mFOLFIRINOX or Gemcitabine/ Nab-Paclitaxel Before Surgery for the Treatment of Resectable, Borderline Resectable, or Locally-Advanced Unresectable Pancreatic Cancer	Recruiting	No Results Available	Borderline Resectable Pancreatic Carcinoma Locally Advanced Unresectable Pancreatic Adenocarcinoma Resectable Pancreatic Ductal Adenocarcinoma Stage 0 Pancreatic Cancer AJCC v8 Stage I Pancreatic Cancer AJCC v8 Stage IA Pancreatic Cancer AJCC v8 Stage IB Pancreatic Cancer AJCC v8 Stage III Pancreatic Cancer AJCC v8 Stage IV Pancreatic Cancer AJCC v8	Drug: Capecitabine Drug: Fluorouracil Drug: Irinotecan Hydrochloride Drug: Losartan Potassium Drug: Losartan Potassium Radiation Therapy Procedure: Resection	Phase 2	60
8	NCT05607017	Losartan in Prevention of Radiation- Induced Heart Failure	Not yet recruiting	No Results Available	Breast Cancer Myocardial Fibrosis Radiation-Induced Fibrosis	Drug: Losartan Radiation: Radiation Therapy	Early Phase 1	10
9	NCT03563248	Losartan and Nivolumab in Combination With FOLFIRINOX and SBRT in Localized Pancreatic Cancer	Active, not recruiting	No Results Available	Pancreatic Cancer	Drug: FOLFIRINOX Drug: Losartan Drug: Nivolumab Radiation: SBRT Procedure: Surgery	Phase 2	168
10	NCT03864042	Pharmacokinetic Drug-drug Interaction Study of Encorafenib and Binimetinib on Probe Drugs in Patients With BRAF V600- mutant Melanoma or Other Advanced Solid Tumors	Active, not recruiting	No Results Available	Advanced Solid Tumors Metastatic Melanoma	Drug: losartan Drug: dextromethorphan Drug: caffeine Drug: omeprazole Drug: midazolam Drug: rosuvastatin Drug: bupropion immediate release (IR) Drug: encorafenib Drug: binimetinib Drug: modafinil	Phase 1	56

Losartan CCL2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
11	NCT03900793	Losartan + Sunitinib in Treatment of Osteosarcoma	Recruiting	No Results Available	Osteosarcoma	Drug: Losartan Drug: Sunitinib	Phase 1	41
12	NCT01199978	Hearing Outcomes Using Fractionated Proton Radiation Therapy for Vestibular Schwannoma	Active, not recruiting	No Results Available	Vestibular Schwannoma Acoustic Neuroma	Radiation: Fractionated proton radiation Drug; Losartan	Phase 2	30
13	NCT03878524	Serial Measurements of Molecular and Architectural Responses to Therapy (SMMART) PRIME Trial	Recruiting	No Results Available	Accelerated Phase Chronic Myelogenous Leukemia, BCR-ABL1 Positive Anatomic Stage IV Breast Cancer AJCC v8 Anemia Ann Arbor Stage III Hodgkin Lymphoma Ann Arbor Stage III Non- Hodgkin Lymphoma Ann Arbor Stage IV Hodgkin Lymphoma Atypical Chronic Myeloid Leukemia, BCR-ABL1 Negative Blast Phase Chronic Myelogenous Leukemia, BCR-ABL1 Positive Castration-Resistant Prostate Carcinoma Chronic Phase Chronic Myelogenous Leukemia, BCR-ABL1 Positive Castration-Resistant Prostate Carcinoma Chronic Phase Chronic Myelogenous Leukemia, BCR-ABL1 Positive Hematopoietic and Lymphoid System Neoplasm Locally Advanced Pancreatic Adenocarcinoma Metastatic Breast Carcinoma Metastatic Malignant Solid Neoplasm Metastatic Pancreatic Adenocarcinoma Myelodysplastic/ Myeloproliferative Neoplasm With Ring Sideroblasts and Thrombocytosis Myelodysplastic/ Myelogroliferative Neoplasm, Unclassifiable Primary Myelofibrosis Recurrent Acute Lymphoblastic Leukemia Recurrent Acute Iymphoolysic Leukemia Recurrent Chronic Myelogenous Leukemia Recurrent Chronic Myelogenous Leukemia Recurrent Hematologic Malignancy Recurrent Hodgkin Lymphoma Recurrent Hematologic Malignancy Recurrent Hodgkin Lymphoma	Drug: Abemaciclib Drug: Afatinib Biological: Bevacizumab Drug: Bicalutamide Procedure: Biospecimen Collection Drug: Bortezomib Drug: Cabozantinib Drug: Cabozantinib Drug: Cabozantinib Drug: Cabozantinib Drug: Cabozantinib Drug: Cabozantinib Drug: Copanlisib Drug: Copanlisib Drug: Dabrafenib Drug: Dabrafenib Drug: Datrafenib Drug: Dacomitinib Drug: Dacomitinib Drug: Datrafenib Drug: Dasatinib Drug: Dasatinib Drug: Enasidenib Drug: Enzalutamide Drug: Enzalutamide Drug: Enzalutamide Drug: Enzalutamide Drug: Enzalutamide Drug: Enzalutamide Drug: Enzalutamide Drug: Enzalutamide Drug: Indelalisib Drug: Idelalisib Drug: Leverolimus Drug: Levatinib Biological: Ipilimumab Drug: Levatinib Drug: Lordatinib Drug: Losartan Drug: Neratinib Biological: Nivolumab Drug: Panobinostat Biological: Pembrolizumab Biological: Pembrolizumab Biological: Pentuzumab Drug: Panobinostat Biological: Pentuzumab Drug: Ponatinib Otug: Regorafenib Drug: Regorafenib Drug: Rugorafenib Drug: Rugoraf	Phase 1	40

Losartan CCL2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
					Neoplasm Recurrent Myeloproliferative Neoplasm Recurrent Non- Hodgkin Lymphoma Recurrent Plasma Cell Myeloma Recurrent Small Lymphocytic Lymphoma Refractory Acute Lymphoblastic Leukemia Refractory Acute Myeloid Leukemia Refractory Chronic Lymphocytic Leukemia Refractory Chronic Myelogenous Leukemia, BCR-ABL1 Positive Refractory Chronic Myelomonocytic Leukemia Refractory Hematologic Malignancy Refractory Hodgkin Lymphoma Refractory Malignant Solid Neoplasm Refractory Myelodysplastic Syndrome Refractory Myelodysplastic/ Myeloproliferative Neoplasm Refractory Non- Hodgkin Lymphoma Refractory Plasma Cell Myeloma Refractory Primary Myelofibrosis Refractory Small Lymphocytic Lymphoma Stage II Pancreatic Cancer AJCC v8 Stage III Pancreatic Cancer AJCC v8 Stage IV Prostate Cancer AJCC v8 Unresectable Pancreatic Adenocarcinoma	Sirolimus Drug: Sorafenib Drug: Sunitinib Drug: Trametinib Biological: Trastuzumab Emtansine Drug: Tretinoin Drug: Vemurafenib Drug: Venetoclax Drug: Vismodegib Drug: Vorinostat		

3.3.1 AMD3100

AMD3100 (plerixafor) is currently the only FDA approved CXCR4 inhibitor. This drug was initially developed for treatment and prevention of HIV, but has now also been approved for treatment of non-Hodgkin lymphoma (NHL) and multiple myeloma (MM) (167, 168). Use of AMD3100 in combination with the anti-VEGFR2 antibody ramucirumab in a mouse model of colorectal cancer significantly reduced recruitment of immune suppressive monocytes, as the study demonstrated that depletion of immune suppressive Ly6C^{low} monocytes by CXCR4 blockade was associated with enhanced treatment efficacy of ramucirumab (169, 170). Administration of AMD3100 was also associated with increased CD8+ T cell infiltration and synergistic activity when combined with ICI (171) Use of AMD3100 in NHL and MM suppressed tumor growth and metastasis and was associated with converting Tregs to a Th1 phenotype and enhancing CD8+ T cell infiltration (172).

Another mechanism of AMD3100 antitumor activity was to block CXCR4+ tumor cells from interacting with CXCL12 produced by cancer associated fibroblasts (173). Use of AMD3100 in combination with ICI in patients with microsatellite unstable pancreatic or colorectal cancer demonstrated enhanced B cell and T cell antitumor responses (174). Clinical trials evaluating AMD3100 include phase II trials for metastatic pancreatic cancer, phase I and II trials for glioma, and phase I and II trials for hematopoietic malignancies. The proposed mechanism targeted in these trials is to sensitize the TME to chemotherapy by blocking the CXCR4 and CXCR/2 axes (175). Other applications of AMD3100 is as a hematopoietic stem cell (HSC) mobilizing agent (typically in combination with G-CSF) for hematopoietic stem cell transplantation (176).

3.3.2 BPRCX807

BPRCX807 is a selective CXCR4 antagonist that has shown activity in mouse models of hepatocellular carcinoma (177). In these models BPRCX807 prevented tumor cell migration and limited the development of metastases. Another activity of BPRCX807 is to reprogram immune suppressive TAMs to a more an immunostimulatory M1 phenotype, while at the same time promoting CD8+ T cell infiltration into tumors (177). Early

TABLE 6 US clinical trials using CXCR1 antagonists for cancer treatment.

CXCR1/2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
Navarixin	NCT03473925	Efficacy and Safety Study of Navarixin (MK-7123) in Combination With Pembrolizumab (MK-3475) in Adults With Selected Advanced/ Metastatic Solid Tumors (MK- 7123-034)	Completed	ORR up to 3.9%, PFS up to 17.5 mo in NSCLC, OS up to 13.0 mo.	Solid Tumors Non-small Cell Lung Cancer Castration Resistant Prostate Cancer Microsatellite Stable Colorectal Cancer	Drug: Navarixin Biological: Pembrolizumab	Phase 2	107
Reparixin	NCT02001974	Pilot Study to Evaluate Reparixin With Weekly Paclitaxel in Patients With HER 2 Negative Metastatic Breast Cancer (MBC)	Completed	Clinical Benefit Rate (CBR) up to 56.5% in group 3 combination treatment. 6mo PFS 25.0%.	Metastatic Breast Cancer	Drug: Paclitaxel +Reparixin	Phase 1	33
	NCT01861054	Pilot Study to Evaluate Safety & Biological Effects of Orally Administered Reparixin in Early Breast Cancer	Terminated	5% SAE due to post op infection	Breast Cancer	Drug: Reparixin	Phase 2	20
	NCT02370238	A Double-blind Study of Paclitaxel in Combination With Reparixin or Placebo for Metastatic Triple- Negative Breast Cancer	Completed	non placebo group SD 15/57, CR 1, PR 15/57	Metastatic Breast Cancer	Drug: paclitaxel Drug: Reparixin Drug: placebo	Phase 2	194
AZD5069	NCT02499328	Study to Assess MEDI4736 With Either AZD9150 or AZD5069 in Advanced Solid Tumors & Relapsed Metastatic Squamous Cell Carcinoma of Head & Neck	Active, not recruiting	SAE 0 ~ 64.29% in dose escalation	Advanced Solid Tumors & Metastatic Squamous Cell Carcinoma of the Head and Neck	Drug: AZD9150 Drug: MEDI4736 Drug: AZD5069 Drug: tremelimumab (treme)	Phase 1 Phase 2	340
	NCT02583477	Phase Ib/II Study of MEDI4736 Evaluated in Different Combinations in Metastatic Pancreatic Ductal Carcinoma	Completed	Dose- Limiting Toxicities (DLT) up to 33.3%, SAE up to 80.00% in cohort 2, study was terminated by sponsor	Metastatic Pancreatic Ductal Adenocarcinoma	Drug: MED14736 in combination with nab- paclitaxel and gemcitabine Drug: MED14736 in combination with AZD5069	Phase 1 Phase 2	23
SX-682	NCT05604560	A Neoadjuvant Study of Tislelizumab and SX-682 for	Not yet recruiting	No Results Available	Pancreatic Cancer	Drug: Tislelizumab Drug: SX-682	Phase 2	25

CXCR1/2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
		Resectable Pancreas Cancer						
	NCT04574583	Phase I/II Trial Investigating the Safety, Tolerability, Pharmacokinetics, Immune and Clinical Activity of SX-682 in Combination With BinTrafusp Alfa (M7824 or TGF-beta "Trap"/ PD-L1) With CV301 TRICOM in Advanced Solid Tumors (STAT)	Active, not recruiting	No Results Available	Metastatic Cancer Solid Tumors	Drug: SX-682 Drug: M7824 Biological: MVA- BN-CV301 Biological: FPV- CV301	Phase 1 Phase 2	12
	NCT05570825	SX-682 With Pembrolizumab for the Treatment of Metastatic or Recurrent Stage IIIC or IV Non- Small Cell Lung Cancer	Recruiting	No Results Available	Metastatic Lung Non-Small Cell Carcinoma Recurrent Lung Non-Small Cell Carcinoma Stage IIIC Lung Cancer AJCC v8 Stage IV Lung Cancer AJCC v8	Procedure: Biopsy Procedure: Biospecimen Collection Procedure: Computed Tomography Drug: CXCR1/2 Inhibitor SX-682 Procedure: Magnetic Resonance Imaging Biological: Pembrolizumab Procedure: Positron Emission Tomography	Phase 2	30
	NCT04599140	SX-682 and Nivolumab for the Treatment of RAS-Mutated, MSS Unresectable or Metastatic Colorectal Cancer, the STOPTRAFFIC-1 Trial	Recruiting	No Results Available	Metastatic Colon Adenocarcinoma Metastatic Colorectal Carcinoma Metastatic Rectal Adenocarcinoma Stage III Colon Cancer AJCC v8 Stage III Rectal Cancer AJCC v8 Stage III Rectal Cancer AJCC v8 Stage IIIB Rectal Cancer AJCC v8 Stage IIIC Colon Cancer AJCC v8 Stage IIIC Rectal Cancer AJCC v8 Stage IV Colon Cancer AJCC v8 Stage IV Rectal Cancer AJCC v8 Stage IV Rectal Cancer AJCC v8 Stage IVA Colon Cancer AJCC v8 Stage IVA Rectal Cancer AJCC v8 Stage IVB Colon Cancer AJCC v8 Stage IVB Rectal Cancer AJCC v8 Stage IVC Colon Cancer AJCC v8 Stage IVC Rectal Cancer AJCC v8 Stage IVB Colon Cancer AJCC v8 Stage IVB Rectal Cancer AJCC v8 Stage IVC Colon Cancer AJCC v8 Stage IVC Rectal Cancer AJCC v8	Drug: CXCR1/2 Inhibitor SX-682 Biological: Nivolumab	Phase 1 Phase 2	53

22

CXCR1/2 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
					Adenocarcinoma Unresectable Rectal Adenocarcinoma			
	NCT04477343	A Study to Evaluate the Safety and Tolerability of SX-682 in Combination With Nivolumab as a Maintenance Therapy in Patients With Metastatic Pancreatic Ductal Adenocarcinoma	Recruiting	No Results Available	Pancreatic Ductal Adenocarcinoma Pancreatic Cancer	Drug: SX-682 Drug: Nivolumab Injectable Product	Phase 1	20
	NCT03161431	SX-682 Treatment in Subjects With Metastatic Melanoma Concurrently Treated With Pembrolizumab	Recruiting	No Results Available	Melanoma Stage III Melanoma Stage IV	Drug: SX-682 Biological: Pembrolizumab	Phase 1	77
	NCT04245397	SX-682 Treatment in Subjects With Myelodysplastic Syndrome Who Had Disease Progression or Are Intolerant to Prior Therapy	Recruiting	No Results Available	Myelodysplastic Syndromes	Drug: SX-682	Phase 1	64

mouse studies provide support for further investigation of CXCR4 blockade as a combination agent along with ICI (172).

4 Metabolic reprogramming to target myeloid suppressor cells

4.1 IDO inhibitors

The enzyme indoleamine 2,3-dioxygenase 1 (IDO1) converts the essential amino acid tryptophan (Trp) to kynurenine (Kyn), thereby leading to an overall depletion of this critical amino acid within the TME and tumor draining lymph nodes (178). Overexpression of IDO1 is considered an important driver of tumor associated immune suppression and a key to establishing immune tolerance of cancer antigens (179, 180). High intratumoral IDO1 expression is correlated with poor prognosis in melanoma, ovarian cancer, colorectal cancer, and lung cancers (181, 182). In ovarian cancer, high IDO1 expression also correlates with increased drug resistance (183). A high ratio of tryptophan to kynurenine in blood is also associated with a poorer prognosis in some cancers (184–187).

High levels of IDO1 expression by cancer cells can also drive MDSC expansion (188); Moreover, MDSCs also overexpress IDO1, triggering a positive feedback loop that reinforces and sustains the

immune suppressive TME (189). Local depletion of tryptophan by IDO leads to cell cycle arrest and apoptosis of effector T cells in tumor tissues (190). In addition, IDO1 positive MDSCs also contribute to T cell exhaustion through IL-6 secretion. The local buildup of kynurenine concentrations within the TME also triggers deleterious alterations in the metabolic properties of tumor infiltrating T cells and converts CD4 effector cells to Tregs (190– 193). Evaluation of IDO inhibitors in preclinical models demonstrated a reduction of IDO1+ MDSCs within the TME and measurable reduction of Kyn concentrations (178). Taken together, these properties make IDO1 a promising target for reversing immune suppression through metabolic reprogramming of the TME.

4.1.1 Epacadostat as an IDO synthesis inhibitor

Van den Eynde et al. summarized the many clinical trials evaluating epacadostat up to 2020 and in their paper discussed why the outcomes of these trials have been largely negative. The majority of these trials have evaluated epacadostat in combination with checkpoint blockade (CTLA4, PD-L1 or PD-1) and have to date failed to demonstrate any meaningful clinical benefit. It was concluded therefore that epacadostat did not improve ICI, as confirmed in at least 12 clinical trials (194, 195). Due to these poor results, remaining clinical trials with epacadostat have been withdrawn, downsized or suspended.

TABLE 7 US clinical trials using CXCR4 targeting drugs for cancer treatment.

CXCR4 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
1	NCT04177810	Plerixafor and Cemiplimab in Metastatic Pancreatic Cancer	Recruiting	No Results Available	Metastatic Pancreatic Cancer	Drug: Cemiplimab Drug: Plerixafor	Phase 2	21
2	NCT01610999	Pilot Study of Lymphoid Tumor Microenvironmental Dysruption Prior to Autologous Stem Cell Transplantation	Terminated	No Results Available	Chronic Lymphocytic Leukemia Lymphoma Multiple Myeloma	Drug: Plerixafor	Phase 1	7
3	NCT03240861	Genetically Engineered PBMC and PBSC Expressing NY-ESO- 1 TCR After a Myeloablative Conditioning Regimen to Treat Patients With Advanced Cancer	Recruiting	No Results Available	HLA-A*0201 Positive Cells Present Locally Advanced Malignant Neoplasm NY-ESO- 1 Positive Unresectable Malignant Neoplasm Sarcoma	Other: 18F-FHBG Biological: Aldesleukin Drug: Busulfan Biological: Cellular Therapy Procedure: Computed Tomography Biological: Filgrastim Drug: Fludarabine Procedure: Leukapheresis Drug: Plerixafor Procedure: Positron Emission Tomography	Phase 1	12
4	NCT01977677	Plerixafor After Radiation Therapy and Temozolomide in Treating Patients With Newly Diagnosed High Grade Glioma	Completed	1/3 (33.33%) SAE at Plerixafor 200 mcg/kg/ Day	Adult Ependymoblastoma Adult Giant Cell Glioblastoma Adult Glioblastoma Adult Medulloblastoma Adult Mixed Glioma Adult Oligodendroglial Tumors Adult Pineoblastoma Adult Supratentorial Primitive Neuroectodermal Tumor (PNET)	Radiation: radiation therapy Drug: temozolomide Drug: plerixafor Other: laboratory biomarker analysis Other: pharmacological study	Phase 1 Phase 2	30
5	NCT00512252	AMD3100 Plus Mitoxantrone, Etoposide and Cytarabine in Acute Myeloid Leukemia	Completed	CR up to 47%, 1 yr Relapse-free Survival 42.9%	Leukemia, Myeloid, Acute	Drug: AMD3100 Drug: Mitoxantrone Drug: Etoposide Drug: Cytarabine	Phase 1 Phase 2	52
6	NCT00669669	O6-Benzylguanine- Mediated Tumor Sensitization With Chemoprotected Autologous Stem Cell in Treating Patients With Malignant Gliomas	Terminated	response rate 9.1%, no SAE	Glioblastoma Gliosarcoma	Radiation: 3- Dimensional Conformal Radiation Therapy Procedure: Autologous Hematopoietic Stem Cell Transplantation Drug: Carmustine Biological: Filgrastim Procedure: In Vitro- Treated Peripheral Blood Stem Cell Transplantation Radiation: Intensity- Modulated Radiation Therapy Other: Laboratory Biomarker Analysis Drug: O6-	Phase 1 Phase 2	12

CXCR4 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
						Benzylguanine Drug: Plerixafor Radiation: Proton Beam Radiation Therapy Drug: Temozolomide		
7	NCT01160354	Plerixafor and Clofarabine in Frontline Treatment of Elderly Patients With Acute Myelogenous Leukemia (AML)	Terminated	CR 35.7%, PR 7.1% (Plerixafor 400 mcg/kg + Clofarabine),	Acute Myelogenous Leukemia	Drug: Plerixafor Drug: Clofarabine	Phase 1 Phase 2	22
8	NCT01352650	Decitabine and Plerixafor in Elderly Acute Myeloid Leukemia (AML)	Completed	No Results Available	Acute Myeloid Leukemia	Drug: plerixafor Drug: decitabine	Phase 1	71
9	NCT01027923	IV Plerixafor With Mitoxantrone Etoposide and Cytarabine for Acute Myeloid Leukemia (AML)	Terminated	No Results Available	Leukemia, Myeloid, Acute	Drug: Plerixafor Drug: Mitoxantrone Drug: Etoposide Drug: Cytarabine	Phase 1	6
10	NCT00943943	Granulocyte-colony Stimulating Factor (G-CSF) and Plerixafor Plus Sorafenib for Acute Myelogenous Leukemia (AML) With FLT3 Mutations	Completed	No Results Available	Acute Myelogenous Leukemia Leukemia	Drug: G-CSF Drug: Plerixafor Drug: Sorafenib	Phase 1	33
11	NCT05088356	Reduced Intensity Allogeneic HCT in Advanced Hematologic Malignancies w/T- Cell Depleted Graft	Recruiting	No Results Available	Allogeneic Hematopoietic Cell Transplantation (HCT) Advanced Hematologic Malignancies Acute Leukemia Chronic Myelogenous Leukemia Myelodysplastic Syndromes Myeloproliferative Disorders	Drug: Purified regulatory T-cells (Treg) plus CD34+ HSPC Drug: Fludarabine Drug: Melphalan Device: CliniMACS CD34 Reagent System Drug: Tacrolimus Drug: Cyclophosphamide Drug: Plerixafor Drug: Filgrastim granulocyte colony-stimulating factor (G-CSF) or equivalent	Phase 1	24
12	NCT00906945	Chemosensitization With Plerixafor Plus G-CSF in Acute Myeloid Leukemia	Completed	45 day CR 30%, Relapse Free-survival Rate 75% at 2 yrs. SAE 2/3 (66.67%) at dose level 2	Leukemia, Myeloid, Acute	Drug: G-CSF Drug: Plerixafor Drug: Mitoxantrone Drug: Etoposide Drug: Cytarabine	Phase 1 Phase 2	39
13	NCT00903968	Combination Plerixafor (AMD3100)and Bortezomib in Relapsed or Relapsed/Refractory Multiple Myeloma	Completed	SD up to 100% in dose level 1 and 5. Time to Progression (TTP) 12.6	Multiple Myeloma	Drug: Plerixafor Drug: bortezomib Drug: Dexamethasone	Phase 1 Phase 2	58

CXCR4 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
				mo, Duration of Response phase 2 (DOR) 12.9 mo				
14	NCT01696461	A Phase II Study Evaluating the Safety and Efficacy of Subcutaneous Plerixafor	Completed	No Results Available	Related Donors Donating PBSC to a Family Member Acute Myelogenous Leukemia Acute Lymphoblastic Leukemia Myelodysplastic Syndrome Chronic Myelogenous Leukemia Non- Hodgkin's Lymphoma Hodgkin's Disease Chronic Lymphocytic Leukemia	Drug: Plerixafor	Phase 2	128
15	NCT00990054	Study of Plerixafor Combined With Cytarabine and Daunorubicin in Patients With Newly Diagnosed Acute Myeloid Leukemia	Completed	No Results Available	Acute Myeloid Leukemia	Drug: Plerixafor	Phase 1	36
16	NCT03746080	Whole Brain Radiation Therapy With Standard Temozolomide Chemo- Radiotherapy and Plerixafor in Treating Patients With Glioblastoma	Recruiting	No Results Available	Glioblastoma Glioblastoma With Primitive Neuronal Component Gliosarcoma Malignant Glioma Oligodendroglial Component Present	Drug: Plerixafor Drug: Temozolomide Radiation: Whole-Brain Radiotherapy (WBRT) Radiation: Radiation Therapy	Phase 2	20
17	NCT01339039	Plerixafor (AMD3100) and Bevacizumab for Recurrent High- Grade Glioma	Terminated	No Results Available	High Grade Glioma: Glioblastoma (GBM) High Grade Glioma: Gliosarcoma Anaplastic Astrocytoma (AA) Anaplastic Oligodendroglioma (AO) Mixed Anaplastic Oligoastrocytoma (AOA)	Drug: Plerixafor Drug: Bevacizumab Procedure: Surgery	Phase 1	26
18	NCT01373229	Lenalidomide + Plerixafor in Previously Treated Chronic Lymphocytic Leukemia (CLL)	Completed	PFS 11 mo, OS 5.5 mo, SAE 93.33%	Leukemia, Lymphocytic, Chronic, B-Cell	Drug: Lenalidomide + Plerixafor (+ Rituximab)	Phase 1	21
19	NCT01065129	Plerixafor and Granulocyte Colony- stimulating Factor	Completed	No Results Available	Myelodysplastic Syndromes	Drug: G-CSF Drug: Plerixafor Drug: Azacitidine	Phase 1	28

CXCR4 blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
		(G-CSF) in Combination With Azacitidine for the Treatment of Myelodysplastic Syndrome (MDS)						
20	NCT00694590	Study of AMD3100 (Plerixafor) and Rituximab in Patients With Chronic Lymphocytic Leukemia or Small Lymphocytic Lymphoma	Completed	No Results Available	Chronic Lymphocytic Leukemia (CLL) Small Lymphocytic Lymphoma (SLL)	Drug: plerixafor	Phase 1	24
21	NCT01319864	POETIC Plerixafor as a Chemosensitizing Agent for Relapsed Acute Leukemia and MDS in Pediatric Patients	Completed	No Results Available	Relapsed/Refractory AML Relapsed/ Refractory ALL Secondary AML/ MDS Acute Leukemia of Ambiguous Lineage AML ALL	Drug: Plerixafor Dose Escalation	Phase 1	20

4.1.2 Navoximod (GDC-0919)

Navoximod has been evaluated clinically as a monotherapy or in combination with atezolizumab (NCT02048709, NCT02471846, NCT05469490, and these trials demonstrated that the navoximod was well tolerated and decreased plasma Kyn concentrations in a dose dependent manner (181). However, there was no clear tumor response benefit in the navoximod combination therapy arm when compared to treatment with atezolizumab alone (196).

4.2 Repurposed beta blockers as MDSC depleting agents

In addition to stimulation of cortisol release, chronic stress from inflammation in cancer is associated with prolonged activation of the sympathetic nervous system (53). Chronic adrenergic activation and release of catecholamines, primarily norepinephrine (Nor), has been associated with MDSC mobilization from the bone marrow and acquisition of greater immune suppressive properties, leading to both systemic and local immune suppression (197, 198). A consequence of increased Nor concentrations is higher concentrations of both MDSCs and TAMs in tumor tissues. For example, activation of β 2-adrenergic receptor (β -AR) signaling was shown to upregulate STAT3 and NFk-b signaling pathways which drive development of immune suppressive MDSC and TAMs (199). Activation of β -AR signaling has been shown to polarize macrophages to an immunosuppressive M2 phenotype in a rodent breast cancer model (200, 201). Adrenergic signaling in tumor cells themselves can also be triggered by tumor hypoxia (202).

4.2.1 Propranolol as non-selective β -blocker

Use of non-selective beta blockers such as propranolol have been investigated for their ability to reprogram immune suppressive cells within the TME (200, 203). Blocking β -AR signaling by MDSCs with propranolol can prevent their mobilization from the bone marrow (204). In addition, propranolol treatment reprograms MDSCs to a less immune suppressive state by blocking STAT3 signaling (205). This effect has been demonstrated in rodent models, where treatment with propranolol reduces MDSC mobilization and accumulation within the TME, accompanied by inhibition of tumor growth and metastasis (206). In rodent models, treatment with propranolol blocked the accumulation of M2 macrophages in metastatic breast cancer and inhibited metastases (53). In a spontaneous melanoma mouse model, propranolol treatment significantly reduced intratumoral accumulation of neutrophils, immune suppressive inflammatory (CD11c-Ly6C^{hi}Ly6G-) macrophages and DCs in both the primary tumor and metastatic lesions (207). Multiple rodent studies and recent clinical trials in dogs and human patients have demonstrated the ability of propranolol 3008 treatment to improve responses to radiation therapy for glioma, 3009 breast cancer, and pancreatic cancer (200, 208) (Table 8).

In a phase II trial in patients with metastatic breast cancer it was found that in tumor tissues from propranolol treated patients there was upregulated expression of genes associated with classical dendritic cells and an increase in M1 macrophage polarization, along with an increase in CD69+ activated TAMs (209). Phase I trials of propranolol with pembrolizumab in patients with metastatic and locally advanced melanoma showed encouraging responses and the combination therapy to be well tolerated (210). In the USA there are currently 17 trials investigating propranolol in

TABLE 8 US clinical trials using propranolol as cancer treatment.

β-AR blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
1	NCT01847001	Study of Propranolol in Newly Diagnosed Breast Cancer Patients Undergoing Neoadjuvant Chemotherapy	Completed	Propranolol + Neoadjuvant Chemotherapy SAE 10%	Locally Advanced Malignant Neoplasm Breast Cancer	Drug: Propranolol Other: DOT imaging Drug: Paclitaxel[Drug: Nab-paclitaxel]Drug: Trastuzumab[Drug: Pertuzumab[Drug: Doxorubicin]Drug: Cyclophosphamide Procedure: Surgery Drug: Premedication Drug: Anti-nausea therapy[Drug: Pegfilgrastim	Phase 2	10
2	NCT01308944	Therapeutic Targeting of Stress Factors in Ovarian Cancer Patients	Completed	No Results Available	Invasive Epithelial Ovarian Cancer Primary Peritoneal Carcinoma Fallopian Tube Cancer	Drug: Propranolol	Phase 1	24
3	NCT02165683	Use of Propranolol to Reduce FDG Uptake in Brown Adipose Tissue in Pediatric Cancer Patients PET Scans	Completed	No Results Available	Pediatric Cancer	Drug: Propranolol	Phase 1	10
4	NCT01902966	Feasibility - Beta Adrenergic Blockade (BB) in Cervical Cancer (CX)	Terminated	dose escalation 40 mg by mouth twice a day, SAE 20%	Cervical Cancer	Drug: Propranolol Behavioral: Diary Behavioral: Relaxation Audio Recording Behavioral: Questionnaires	Not Applicable	6
5	NCT04848519	Propranolol Hydrochloride and Pembrolizumab for the Treatment of Recurrent or Metastatic Urothelial Cancer	Recruiting	No Results Available	Recurrent or Metastatic Urothelial Cancer	Drug: Pembrolizumab Drug: Propranolol Hydrochloride	Phase 2	25
6	NCT03152786	Propranolol Hydrochloride in Treating Patients With Prostate Cancer Undergoing Surgery	Suspended	No Results Available	Prostate Carcinoma	Other: Laboratory Biomarker Analysis Drug: Propranolol Hydrochloride Other: Questionnaire Administration Other: Survey Administration	Phase 2	50
7	NCT05651594	Propranolol in Combination With Pembrolizumab and Standard Chemotherapy for the Treatment of Unresectable Locally Advanced or Metastatic Esophageal or Gastroesophageal Junction Adenocarcinoma	Recruiting	No Results Available	Unresectable Locally Advanced or Metastatic Esophageal or Gastroesophageal Junction Adenocarcinoma	Procedure: Biopsy Procedure: Biospecimen Collection Procedure: Computed Tomography Drug: Fluorouracil Drug: Leucovorin Drug: Oxaliplatin Biological: Pembrolizumab Drug: Propranolol Hydrochloride Other: Questionnaire Administration	Phase 2	40

β-AR blockade	NCT Number		Status	Study Results	Conditions	Interventions	Phases	Enrollment
8	NCT01504126	Propranolol Hydrochloride and Chemotherapy in Treating Patients With Ovarian, Primary Peritoneal, or Fallopian Tube Cancer	Completed	No Results Available	Ovarian, Primary Peritoneal, or Fallopian Tube Cancer	Drug: Chemotherapy Drug: Propranolol Hydrochloride Other: Quality-of-Life Assessment Procedure: Therapeutic Conventional Surgery	Early Phase 1	32
9	NCT04682158	Propranolol With Standard Chemoradiation for Esophageal Adenocarcinoma	Recruiting	No Results Available	Esophageal Adenocarcinoma	Drug: Carboplatin Radiation: 3 Dimensional Conformal Radiation Therapy Drug: Propranolol Radiation: Intensity Modulated Radiation Therapy Drug: Paclitaxel	Phase 2	60
10	NCT03384836	Propranolol Hydrochloride and Pembrolizumab in Treating Patients With Stage IIIC-IV Melanoma That Cannot Be Removed by Surgery	Recruiting	No Results Available	Stage IIIC Cutaneous Melanoma AJCC v7 Stage IV Cutaneous Melanoma AJCC v6 and v7	Other: Laboratory Biomarker Analysis Biological: Pembrolizumab Drug: Propranolol Hydrochloride	Phase 1 Phase 2	47
11	NCT00967226	Propranolol Versus Prednisolone for Treatment of Symptomatic Hemangiomas	Terminated	Propranolol SAE 1/11 (9.09%)	Hemangioma of Infancy	Drug: propranolol Drug: Prednisolone	Phase 2	19
12	NCT05312255	Non- chemotherapeutic Interventions for the Improvement of Quality of Life and Immune Function in Patients With Multiple Myeloma	Recruiting	No Results Available	Plasma Cell Myeloma Recurrent Plasma Cell Myeloma Refractory Plasma Cell Myeloma Smoldering Plasma Cell Myeloma	Behavioral: Behavioral Intervention Drug: Beta-Adrenergic Antagonist Drug: Propranolol Other: Quality-of-Life Assessment Other: Questionnaire Administration Other: Resistance Training Other: Short-Term Fasting	Not Applicable	150
13	NCT01074437	Corticosteroids With Placebo Versus Corticosteroids With Propranolol Treatment of Infantile Hemangiomas (IH)	Terminated	Has Results	Hemangioma	Drug: Prednisolone (Corticosteroid) Drug: Propranolol Drug: Placebo	Phase 2	9
14	NCT05479123	Assessing the Impact of Dosage Frequency of Propranolol on Sleep Patterns in Patients With Infantile Hemangiomas	Recruiting	No Results Available	Infantile Hemangioma	Drug: Propranolol three times a day Drug: Propranolol twice a day Drug: Timolol	Phase 4	174
15	NCT01056341	Study to Demonstrate the	Completed	Propranolol 3mg/kg/d 6	Infantile Hemangioma	Drug: Propranolol Drug: Placebo	Phase 2 Phase 3	512

β-AR blockade	NCT Number	Title	Status	Study Results	Conditions	Interventions	Phases	Enrollment
		Efficacy and Safety of Propranolol Oral Solution in Infants With Proliferating Infantile Hemangiomas Requiring Systemic Therapy		Months, 60.4% resolution. SAE 4.95% for Propranolol 3mg/kg/day for 6 months				
16	NCT01265576	Study of Sorafenib With or Without VT-122 in Patients With Hepatocellular Carcinoma (HCC)	Unknown status	No Results Available	НСС	Drug: Sorafenib Drug: VT-122 (propranolol plus etodolac) Drug: Placebo	Phase 2	20

breast cancer, cervical cancer, prostate, esophageal, infantile hemangioma and hepatocellular carcinoma. Further studies are warranted to elucidate the clinical benefit of propranolol as a repurposed immunotherapy for TME reprogramming (Table 8).

4.3 Tyrosine kinase inhibitors

Tyrosine kinase inhibitors (TKIs), especially early generation non-specific TKIs such as sunitinib, have been shown to alter the immune suppressive TME, in part by reprogramming TAMs from M2 to M1 phenotypes, by reducing total TAM infiltrates and by blocking the accumulation of MDSCs and TANs (211-213). Tyrosine kinase receptors are extremely diverse family of receptors and there are >40 FDA approved TKI drugs. These TKIs are categorized according to the main receptor targeting sites which include, anaplastic lymphoma kinase (ALK), epidermal growth factor receptor (EGFR), FMS-like tyrosine kinase 3 (FLT3), vascular endothelial growth factor (VEGFR), and tropomyosin receptor kinase (TRK) (214). The positive clinical benefits observed when TKIs are combined with ICI indicate that the TKI impact on the TME is substantial and complementary to ICI therapy. The list of multi-target TKIs is quite extensive, therefore few are selected here for discussion to illustrate their potential as immunotherapy drugs.

4.3.1 VEGFR targeted TKIs

4.3.1.1 Sunitinib (SU011248, Sutent)

Sunitinib is a small molecule inhibitor that targets multiple kinases, with inhibitory effects against signaling by VEGFR, PDGFR and c-kit (215). It is an FDA approved agent for treatment of renal cell carcinoma and gastrointestinal stromal tumors (216). To date there are currently 270 US trials of sunitinib to treat, alone or in combination, many different cancers, including breast, hepatic, lung, and renal cancers. Early generation, multi-function TKIs such as sunitinib have been shown to exert impressive immune modulatory effects (217, 218). For example, sunitinib has been shown to deplete both MDSC and Tregs, in part by inhibiting STAT3 signaling (219); and in clinical trials positive responses to treatment with sunitinib has been associated with Treg depletion (220, 221).

4.3.1.2 Sorafenib (Nexavar, BAY 43-9006)

Sorafenib is another small molecule multi-kinase inhibitor, which in hepatocellular carcinoma has shown clinical benefit and antitumoral activity that is associated with immune remodeling of the TME (222). For example, treatment with sorafenib has been reported to selectively decrease Tregs numbers without impacting effector T cell numbers (223). Sorafenib has been shown to regulate the differentiation DCs in the TME (224) and to repolarize M2 TAMs to an M1 phenotype through inhibition of miR-101 expression and reduction of TGF- β secretion. Sorafenib has also been reported to induce secretion of pro-inflammatory cytokines such as IL-12 by TAMs (225, 226) and to decrease expression of PD-L1 on MDCS and plasmacytoid DCs (227, 228). There are 430 clinical trials registered in the US using sorafenib in cancer patients ranging from phase I to phase IV clinical trials, with many focused on renal cell carcinoma.

4.3.1.3 Lenvatinib (E7080, Lenvima)

Lenvatinib is another multitarget TKI that has shown in phase III trials clinical benefit as reflected by significantly increased overall survival times in patients with hepatocellular carcinoma (227). There are currently 128 registered lenvatinib clinical trials in the USA, with multiple phase I through phase III trials for treatment of thyroid cancer, renal cell carcinoma, hepatocellular carcinoma and melanoma. The antitumor activity of lenvatinib is also heavily linked to its anti-angiogenic properties (229). In addition, Lenvatinib has been shown to reduce TAMs and increase IFNγ secreting CD8 effector T cells in tumor tissues in a mouse model of colon carcinoma (230).

4.3.2 EGFR targeted TKI

EGFR targeted TKIs disrupt the immune suppressive TME by several mechanism including blocking cancer cell migration and nutrient delivery through targeting of endothelial cells and suppressing pericyte coverage (231). Highly proliferative cancer stem cells also express EGFR and can be inhibited by EGFR targeted TKIs (232). For example, EGFR-mutated NSCLC is known to be
especially sensitive to treatment with EGFR TKIs (233); and these
TKIS are therefore often a first line treatment for this cancer (233,
234). Many EGFR TKI drugs have been developed, and first-greater activity
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234). Many EGFR TKI drugs have been developed, and firstgeneration drugs such as gefitinib, erlotinib, and afatinib are approved for the treatment of EGFR mutated NSCLC. Currently third generation EGFR TKI drugs are being investigated as monotherapy and in combination with chemotherapy (213). There are currently around 8 FDA approved EGFR targeted TKI (235) with over 1200 total clinical trials in the US ranging from phase I to phase IV.

5 Future opportunities for myeloid cells as targets in cancer immunotherapy

Many different strategies targeting immune suppressor cells within the TME to reverse or ameliorate immune suppression have been evaluated. To date, the most successful strategies have been those targeting MDSCs, including the use of multi-function TKIs and repurposed beta blockers. For reprogramming TAMs, the most studied targets to date have been CSF-R1 inhibitors, either as biologics or targeted agents, though clinical responses to date have not been impressive (201, 236, 237). Other strategies have been even less successful, including the use of arginase and IDO inhibitors to reprogram metabolic pathways used by TAMs and tumor cells (194, 238). In the future, the most successful rational strategies will likely employ drugs and biologics targeting multiple different complementary pathways of tumor immune evasion, to block non-redundant mechanisms and pathways. Such combination strategies may also include creative uses of radiation therapy to enhance tumor immunogenicity, while MDSC or inflammatory monocyte targeted drugs can be used to relieve radiation induced inflammatory responses. Other gains will undoubtedly be realized when newer drugs and biologics with

References

1. Labani-Motlagh A, Ashja-Mahdavi M, Loskog A. The tumor microenvironment: A milieu hindering and obstructing antitumor immune responses. *Front Immunol* (2020), 940. doi: 10.3389/fimmu.2020.00940

2. Talaat IM, Kim B. A brief glimpse of a tangled web in a small world: Tumor microenvironment. *Front Med* (2022) 9:1002715. doi: 10.3389/fmed.2022.1002715

3. Barnestein R, Galland L, Kalfeist L, Ghiringhelli F, Ladoire S, Limagne E. Immunosuppressive tumor microenvironment modulation by chemotherapies and targeted therapies to enhance immunotherapy effectiveness. *Oncoimmunology* (2022) 11:2120676. doi: 10.1080/2162402x.2022.2120676

4. Gabrilovich DI, Ostrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. *Nat Rev Immunol* (2012) 12:253–68. doi: 10.1038/nri3175

5. Pramanik A, Bhattacharyya S. Myeloid derived suppressor cells and innate immune system interaction in tumor microenvironment. *Life Sci* (2022) 305:120755. doi: 10.1016/j.lfs.2022.120755

6. Jiang Y, Li Y, Zhu B. T-Cell exhaustion in the tumor microenvironment. Cell Death Dis (2015) 6:e1792. doi: 10.1038/cddis.2015.162

7. Lin Y, Xu J, Lan H. Tumor-associated macrophages in tumor metastasis: Biological roles and clinical therapeutic applications. *J Hematol Oncol* (2019), 76. doi: 10.1186/s13045-019-0760-3 greater activity or more specific targeting of myeloid cell pathways enter the clinic. Thus, it is likely that we will see greater use of myeloid targeted agents as part of a more comprehensive strategy and platform for cancer immunotherapy (235).

Author contributions

JC, LC, SD collection and assembly of data, conception design and manuscript writing and revision. JC and LC are equal contribution first authors. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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8. Shaul ME, Fridlender ZG. Tumour-associated neutrophils in patients with cancer. Nat Rev Clin Oncol (2019) 16:601-20. doi: 10.1038/s41571-019-0222-4

9. Shurin GV, Ma Y, Shurin MR. Immunosuppressive mechanisms of regulatory dendritic cells in cancer. *Cancer Microenviron* (2013) 6:159–67. doi: 10.1007/s12307-013-0133-3

10. Wei SY. Yin-yang regulating effects of cancer-associated genes, proteins, and cells: An ancient Chinese concept in vogue in modern cancer research. *Biosci Trends* (2017) 11:612–8. doi: 10.5582/bst.2017.01259

11. Clappaert EJ, Murgaski A, Van Damme H, Kiss M, Laoui D. Diamonds in the rough: Harnessing tumor-associated myeloid cells for cancer therapy. *Front Immunol* (2018) 9:2250. doi: 10.3389/fimmu.2018.02250

12. Cendrowicz E, Sas Z, Bremer E, Rygiel TP. The role of macrophages in cancer development and therapy. *Cancers (Basel)* (2021), 1946. doi: 10.3390/cancers13081946

13. Ostrand-Rosenberg S. Immune surveillance: A balance between protumor and antitumor immunity. *Curr Opin Genet Dev* (2008) 18:11-8. doi: 10.1016/j.gde.2007.12.007

14. Aminin D, Wang Y-M. Macrophages as a "weapon" in anticancer cellular immunotherapy. Kaohsiung J Med Sci (2021) 37:749-58. doi: 10.1002/kjm2.12405

15. Veglia F, Sanseviero E, Gabrilovich DI. Myeloid-derived suppressor cells in the era of increasing myeloid cell diversity. *Nat Rev Immunol* (2021) 21:485–98. doi: 10.1038/s41577-020-00490-y

16. Davidov V, Jensen G, Mai S, Chen S-H, Pan P-Y. Analyzing one cell at a TIME: Analysis of myeloid cell contributions in the tumor immune microenvironment. *Front Immunol* (2020) 11. doi: 10.3389/fimmu.2020.01842

 Movahedi K, Laoui D, Gysemans C, Baeten M, Stangé G, Van den Bossche J, et al. Different tumor microenvironments contain functionally distinct subsets of macrophages derived from Ly6C(high) monocytes. *Cancer Res* (2010) 70:5728–39. doi: 10.1158/0008-5472.CAN-09-4672

18. Bronte V, Brandau S, Chen S-H, Colombo MP, Frey AB, Greten TF, et al. Recommendations for myeloid-derived suppressor cell nomenclature and characterization standards. *Nat Commun* (2016) 7:12150. doi: 10.1038/ncomms12150

19. Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T, Minato N. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. *Proc Natl Acad Sci U S A* (2002) 99:12293–7. doi: 10.1073/pnas.192461099

20. Shi L, Chen S, Yang L, Li Y. The role of PD-1 and PD-L1 in T-cell immune suppression in patients with hematological malignancies. *J Hematol Oncol* (2013) 6:74. doi: 10.1186/1756-8722-6-74

21. Wang L, Rubinstein R, Lines JL, Wasiuk A, Ahonen C, Guo Y, et al. VISTA, a novel mouse ig superfamily ligand that negatively regulates T cell responses. *J Exp Med* (2011) 208:577–92. doi: 10.1084/jem.20100619

22. ElTanbouly MA, Croteau W, Noelle RJ, Lines JL. VISTA: a novel immunotherapy target for normalizing innate and adaptive immunity. *Semin Immunol* (2019) 42:101308. doi: 10.1016/j.smim.2019.101308

23. Miyamoto T, Murakami R, Hamanishi J, Tanigaki K, Hosoe Y, Mise N, et al. B7-H3 suppresses antitumor immunity *via* the CCL2-CCR2-M2 macrophage axis and contributes to ovarian cancer progression. *Cancer Immunol Res* (2022) 10:56–69. doi: 10.1158/2326-6066.Cir-21-0407

24. Sun Y, Wang Y, Zhao J, Gu M, Giscombe R, Lefvert AK, et al. B7-H3 and B7-H4 expression in non-small-cell lung cancer. *Lung Cancer* (2006) 53:143–51. doi: 10.1016/j.lungcan.2006.05.012

25. Ni L, Dong C. New checkpoints in cancer immunotherapy. *Immunol Rev* (2017) 276:52–65. doi: 10.1111/imr.12524

26. Zhang Z, Liu S, Zhang B, Qiao L, Zhang Y, Zhang Y. T Cell dysfunction and exhaustion in cancer. *Front Cell Dev Biol* (2020) 8:17. doi: 10.3389/fcell.2020.00017

27. Hicklin DJ, Marincola FM, Ferrone S. HLA class I antigen downregulation in human cancers: T-cell immunotherapy revives an old story. *Mol Med Today* (1999) 5:178–86. doi: 10.1016/s1357-4310(99)01451-3

28. Campoli M, Chang CC, Ferrone S. HLA class I antigen loss, tumor immune escape and immune selection. *Vaccine* (2002) 20 Suppl 4:A40–5. doi: 10.1016/s0264-410x(02)00386-9

29. Pluda JM. Tumor-associated angiogenesis: Mechanisms, clinical implications, and therapeutic strategies. *Semin Oncol* (1997) 24:203–18.

30. Veikkola T, Alitalo K. VEGFs, receptors and angiogenesis. Semin Cancer Biol (1999) 9:211-20. doi: 10.1006/scbi.1998.0091

31. Zhang Q-W, Liu L, Gong C-Y, Shi H-S, Zeng Y-H, Wang X-Z, et al. Prognostic significance of tumor-associated macrophages in solid tumor: A meta-analysis of the literature. *PloS One* (2012) 7:e50946. doi: 10.1371/journal.pone.0050946

32. Jin JK, Lin JT, Xu AK, Lou JA, Qian C, Li XM, et al. CCL2: An important mediator between tumor cells and host cells in tumor microenvironment. *Front Oncol* (2021) 11:722916. doi: 10.3389/fonc.2021.722916

33. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: Tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol* (2002) 23:549–55. doi: 10.1016/S1471-4906(02)02302-5

34. Keeley T, Costanzo-Garvey DL, Cook LM. Unmasking the many faces of tumorassociated neutrophils and macrophages: Considerations for targeting innate immune cells in cancer. *Trends Cancer* (2019) 5:789–98. doi: 10.1016/j.trecan.2019.10.013

35. Cassetta L, Fragkogianni S, Sims AH, Swierczak A, Forrester LM, Zhang H, et al. Human tumor-associated macrophage and monocyte transcriptional landscapes reveal cancer-specific reprogramming, biomarkers, and therapeutic targets. *Cancer Cell* (2019) 35:588–602.e10. doi: 10.1016/j.ccell.2019.02.009

36. Green CE, Liu T, Montel V, Hsiao G, Lester RD, Subramaniam S, et al. Chemoattractant signaling between tumor cells and macrophages regulates cancer cell migration, metastasis and neovascularization. *PloS One* (2009) 4:e6713. doi: 10.1371/journal.pone.0006713

37. Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. *Nat Rev Cancer* (2004) 4:71–8. doi: 10.1038/nrc1256

38. Yang L, Huang J, Ren X, Gorska AE, Chytil A, Aakre M, et al. Abrogation of TGF beta signaling in mammary carcinomas recruits gr-1+CD11b+ myeloid cells that promote metastasis. *Cancer Cell* (2008) 13:23–35. doi: 10.1016/j.ccr.2007.12.004

39. De Sanctis F, Adamo A, Cane S, Ugel S. Targeting tumour-reprogrammed myeloid cells: the new battleground in cancer immunotherapy. *Semin Immunopathology* (2022) 26:1–24. doi: 10.1007/s00281-022-00965-1

40. Hartley GP, Chow L, Ammons DT, Wheat WH, Dow SW. Programmed cell death ligand 1 (PD-L1) signaling regulates macrophage proliferation and activation. *Cancer Immunol Res* (2018) 6:1260–73. doi: 10.1158/2326-6066.CIR-17-0537

41. Rodriguez PC, Zea AH, DeSalvo J, Culotta KS, Zabaleta J, Quiceno DG, et al. L-arginine consumption by macrophages modulates the expression of CD3ζ chain in T lymphocytes. *J Immunol* (2003) 171:1232–9. doi: 10.4049/jimmunol.171.3.1232

42. Hasan MN, Capuk OA-O, Patel SA-O, Sun D. The role of metabolic plasticity of tumor-associated macrophages in shaping the tumor microenvironment immunity. *Cancers (Basel)* (2022) 14(14):3331. doi: 10.3390/cancers14143331

43. Wculek SK, Cueto FJ, Mujal AM, Melero I, Krummel MF, Sancho D. Dendritic cells in cancer immunology and immunotherapy. *Nat Rev Immunol* (2020) 20:7–24. doi: 10.1038/s41577-019-0210-z

44. Bottcher JP, Reis e Sousa C. The role of type 1 conventional dendritic cells in cancer immunity. *Trends Cancer* (2018) 4:784–92. doi: 10.1016/j.trecan.2018.09.001

45. Rowshanravan B, Halliday N, Sansom DM. CTLA-4: A moving target in immunotherapy. *Blood* (2018) 131:58–67. doi: 10.1182/blood-2017-06-741033

46. Wu YZ, Yi M, Niu MK, Mei Q, Wu KM. Myeloid-derived suppressor cells: An emerging target for anticancer immunotherapy. *Mol Cancer* (2022) 21. doi: 10.1186/s12943-022-01657-y

47. Peranzoni E, Zilio S, Marigo I, Dolcetti L, Zanovello P, Mandruzzato S, et al. Myeloid-derived suppressor cell heterogeneity and subset definition. *Curr Opin Immunol* (2010) 22:238-44. doi: 10.1016/j.coi.2010.01.021

48. Han ZJ, Li YB, Yang LX, Cheng HJ, Liu X, Chen H. Roles of the CXCL8-CXCR1/ 2 axis in the tumor microenvironment and immunotherapy. *Molecules* (2021) 27:137. doi: 10.3390/molecules27010137

49. Youn JI, Nagaraj S, Collazo M, Gabrilovich DI. Subsets of myeloid-derived suppressor cells in tumor-bearing mice. *J Immunol* (2008) 181:5791–802. doi: 10.4049/jimmunol.181.8.5791

50. Centuori SM, Trad M, LaCasse CJ, Alizadeh D, Larmonier CB, Hanke NT, et al. Myeloid-derived suppressor cells from tumor-bearing mice impair TGF- β -induced differentiation of CD4+ CD25+ FoxP3+ tregs from CD4+ CD25- FoxP3- T cells. J leukocyte Biol (2012) 92:987–97. doi: 10.1189/jlb.0911465

51. Martin RK, Saleem SJ, Folgosa L, Zellner HB, Damle SR, Nguyen GKT, et al. Mast cell histamine promotes the immunoregulatory activity of myeloid-derived suppressor cells. *J Leukocyte Biol* (2014) 96:151–9. doi: 10.1189/jlb.5A1213-644R

52. Sieminska I, Baran J. Myeloid-derived suppressor cells as key players and promising therapy targets in prostate cancer. *Front Oncol* (2022) 12:862416. doi: 10.3389/fonc.2022.862416

53. An JL, Feng LF, Ren JL, Li YF, Li GR, Liu C, et al. Chronic stress promotes breast carcinoma metastasis by accumulating myeloid-derived suppressor cells through activating beta-adrenergic signaling. *Oncoimmunology* (2021) 10:2004659. doi: 10.1080/2162402x.2021.2004659

54. Yang Z, Guo J, Weng L, Tang W, Jin S, Ma W. Myeloid-derived suppressor cellsnew and exciting players in lung cancer. *J Hematol Oncol* (2020) 13:10. doi: 10.1186/ s13045-020-0843-1

55. Bennett JA, Rao VS, Mitchell MS. Systemic bacillus calmette-guerin (BCG) activates natural suppressor cells. *Proc Natl Acad Sci U S A* (1978) 75:5142–4. doi: 10.1073/pnas.75.10.5142

56. Condamine T, Mastio J, Gabrilovich DI. Transcriptional regulation of myeloidderived suppressor cells. *J Leukoc Biol* (2015) 98:913–22. doi: 10.1189/jlb.4RI0515-204R

57. Groth C, Hu X, Weber R, Fleming V, Altevogt P, Utikal J, et al. Immunosuppression mediated by myeloid-derived suppressor cells (MDSCs) during tumour progression. *Br J Cancer* (2019) 120:16–25. doi: 10.1038/s41416-018-0333-1

58. Lauret Marie Joseph E, Laheurte C, Jary M, Boullerot L, Asgarov K, Gravelin E, et al. Immunoregulation and clinical implications of ANGPT2/TIE2(+) m-MDSC signature in non-small cell lung cancer. *Cancer Immunol Res* (2020) 8:268–79. doi: 10.1158/2326-6066.CIR-19-0326

59. Koh J, Kim Y, Lee KY, Hur JY, Kim MS, Kim B, et al. MDSC subtypes and CD39 expression on CD8(+) T cells predict the efficacy of anti-PD-1 immunotherapy in patients with advanced NSCLC. *Eur J Immunol* (2020) 50:1810–9. doi: 10.1002/eji.202048534

60. Tumino N, Besi F, Martini S, Di Pace AL, Munari E, Quatrini L, et al. Polymorphonuclear myeloid-derived suppressor cells are abundant in peripheral blood of cancer patients and suppress natural killer cell anti-tumor activity. *Front Immunol* (2022) 12:803014. doi: 10.3389/fmmu.2021.803014

61. Patel S, Fu S, Mastio J, Dominguez GA, Purohit A, Kossenkov A, et al. Unique pattern of neutrophil migration and function during tumor progression. *Nat Immunol* (2018) 19:1236–47. doi: 10.1038/s41590-018-0229-5

62. Youn JI, Collazo M, Shalova IN, Biswas SK, Gabrilovich DI. Characterization of the nature of granulocytic myeloid-derived suppressor cells in tumor-bearing mice. *J Leukoc Biol* (2012) 91:167–81. doi: 10.1189/jlb.0311177

63. Fridlender ZG, Albelda SM. Tumor-associated neutrophils: Friend or foe? *Carcinogenesis* (2012) 33:949–55. doi: 10.1093/carcin/bgs123

64. Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, et al. Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer Cell* (2009) 16:183–94. doi: 10.1016/j.ccr.2009.06.017

65. Masucci MT, Minopoli M, Carriero MV. Tumor associated neutrophils. their role in tumorigenesis, metastasis, prognosis and therapy. *Front Oncol* (2019) 9. doi: 10.3389/fonc.2019.01146

66. Kargl J, Zhu XD, Zhang HJ, Yang GHY, Friesen TJ, Shipley M, et al. Neutrophil content predicts lymphocyte depletion and anti-PD1 treatment failure in NSCLC. *JCI Insight* (2019) 4:e130850. doi: 10.1172/jci.insight.130850

67. Emmons TR, Giridharan T, Singel KL, Khan ANH, Ricciuti J, Howard K, et al. Mechanisms driving neutrophil-induced T-cell immunoparalysis in ovarian cancer. *Cancer Immunol Res* (2021) 9:790–810. doi: 10.1158/2326-6066.CIR-20-0922

68. Piccard H, Muschel RJ, Opdenakker G. On the dual roles and polarized phenotypes of neutrophils in tumor development and progression. *Crit Rev Oncol Hematol* (2012) 82:296–309. doi: 10.1016/j.critrevonc.2011.06.004

 Deryugina EI, Zajac E, Juncker-Jensen A, Kupriyanova TA, Welter L, Quigley JP. Tissue-infiltrating neutrophils constitute the major *in vivo* source of angiogenesisinducing MMP-9 in the tumor microenvironment. *Neoplasia* (2014) 16:771–88. doi: 10.1016/j.neo.2014.08.013

70. Schaider H, Oka M, Bogenrieder T, Nesbit M, Satyamoorthy K, Berking C, et al. Differential response of primary and metastatic melanomas to neutrophils attracted by IL-8. *Int J Cancer* (2003) 103:335–43. doi: 10.1002/ijc.10775

71. Haqqani AS, Sandhu JK, Birnboim HC. Expression of interleukin-8 promotes neutrophil infiltration and genetic instability in mutatect tumors. *Neoplasia* (2000) 2:561–8. doi: 10.1038/sj.neo.7900110

72. Bellocq A, Antoine M, Flahault A, Philippe C, Crestani B, Bernaudin JF, et al. Neutrophil alveolitis in bronchioloalveolar carcinoma: Induction by tumor-derived interleukin-8 and relation to clinical outcome. *Am J Pathol* (1998) 152:83–92.

73. Li P, Lu M, Shi J, Hua L, Gong Z, Li Q, et al. Dual roles of neutrophils in metastatic colonization are governed by the host NK cell status. *Nat Commun* (2020) 11:4387. doi: 10.1038/s41467-020-18125-0

74. Eruslanov EB, Singhal S, Albelda SM. Mouse versus human neutrophils in cancer: A major knowledge gap. *Trends Cancer* (2017) 3:149–60. doi: 10.1016/j.trecan.2016.12.006

75. Teijeira A, Garasa S, Migueliz I, Cirella A, Melero I. Cxcr1 and Cxcr2 chemokine receptor agonists produced by tumors induce neutrophil extracellular traps that interfere with immune cytotoxicity. *J Immunother Cancer* (2020) 8:A453–A. doi: 10.1136/jitc-2020-SITC2020.0755

76. Teijeira A, Garasa S, Ochoa MC, Villalba M, Olivera I, Cirella A, et al. IL8, neutrophils, and NETs in a collusion against cancer immunity and immunotherapy. *Clin Cancer Res* (2021) 27:2383–93. doi: 10.1158/1078-0432.Ccr-20-1319

77. Zhang Y, Wang CX, Yu MX, Zhao XY, Du JW, Li YY, et al. Neutrophil extracellular traps induced by activated platelets contribute to procoagulant activity in patients with colorectal cancer. *Thromb Res* (2019) 180:87–97. doi: 10.1016/j.thromres.2019.06.005

78. Pathria P, Louis TL, Varner JA. Targeting tumor-associated macrophages in cancer. *Trends Immunol* (2019) 40:310–27. doi: 10.1016/j.it.2019.02.003

79. Chitu V, Stanley ER. Colony-stimulating factor-1 in immunity and inflammation. Curr Opin Immunol (2006) 18:39–48. doi: 10.1016/j.coi.2005.11.006

80. Stanley ER, Chitu V. CSF-1 receptor signaling in myeloid cells. Cold Spring Harbor Perspect Biol (2014) 6:a021857. doi: 10.1101/cshperspect.a021857

81. Wang X, Zhang J, Hu B, Qian F. High expression of CSF-1R predicts poor prognosis and CSF-1R(high) tumor-associated macrophages inhibit anti-tumor immunity in colon adenocarcinoma. *Front Oncol* (2022) 12:850767. doi: 10.3389/ fonc.2022.850767

82. Candido JB, Morton JP, Bailey P, Campbell AD, Karim SA, Jamieson T, et al. CSF1R(+) macrophages sustain pancreatic tumor growth through T cell suppression and maintenance of key gene programs that define the squamous subtype. *Cell Rep* (2018) 23:1448–60. doi: 10.1016/j.celrep.2018.03.131

83. Koh YW, Park C, Yoon DH, Suh C, Huh J. CSF-1R expression in tumorassociated macrophages is associated with worse prognosis in classical Hodgkin lymphoma. *Am J Clin Pathol* (2014) 141:573-83. doi: 10.1309/AJCPR92TDDFARISU

84. Espinosa I, Beck AH, Lee CH, Zhu S, Montgomery KD, Marinelli RJ, et al. Coordinate expression of colony-stimulating factor-1 and colony-stimulating factor-1-related proteins is associated with poor prognosis in gynecological and nongynecological leiomyosarcoma. *Am J Pathol* (2009) 174:2347–56. doi: 10.2353/ajpath.2009.081037

85. Jia JB, Wang WQ, Sun HC, Zhu XD, Liu L, Zhuang PY, et al. High expression of macrophage colony-stimulating factor-1 receptor in peritumoral liver tissue is associated with poor outcome in hepatocellular carcinoma after curative resection. *Oncologist* (2010) 15:732–43. doi: 10.1634/theoncologist.2009-0170

86. Lin EY, Pollard JW. Tumor-associated macrophages press the angiogenic switch in breast cancer. *Cancer Res* (2007) 67:5064–6. doi: 10.1158/0008-5472.CAN-07-0912

87. Cannarile MA, Weisser M, Jacob W, Jegg AM, Ries CH, Ruttinger D. Colonystimulating factor 1 receptor (CSF1R) inhibitors in cancer therapy. *J Immunother Cancer* (2017) 5:53. doi: 10.1186/s40425-017-0257-y

 Denny WA, Flanagan JU. Small-molecule CSF1R kinase inhibitors; Review of patents 2015-present. Expert Opin Ther Pat (2021) 31:107–17. doi: 10.1080/13543776.2021.1839414

89. Lu X, Yan S, Koral KA, Chen Z. Surufatinib for the treatment of advanced extrapancreatic neuroendocrine tumors. *Expert Rev Anticancer Ther* (2021) 21:917–26. doi: 10.1080/14737140.2021.1944110

90. von Tresckow B, Morschhauser F, Ribrag V, Topp MS, Chien C, Seetharam S, et al. An open-label, multicenter, phase I/II study of JNJ-40346527, a CSF-1R inhibitor, in patients with relapsed or refractory Hodgkin lymphoma. *Clin Cancer Res* (2015) 21:1843–50. doi: 10.1158/1078-0432.CCR-14-1845

91. Holmgaard RB, Brachfeld A, Gasmi B, Jones DR, Mattar M, Doman T, et al. Timing of CSF-1/CSF-1R signaling blockade is critical to improving responses to CTLA-4 based immunotherapy. Oncoimmunology (2016) 5:e1151595. doi: 10.1080/2162402X.2016.1151595

92. Yan D, Kowal J, Akkari L, Schuhmacher AJ, Huse JT, West BL, et al. Inhibition of colony stimulating factor-1 receptor abrogates microenvironment-mediated therapeutic resistance in gliomas. *Oncogene* (2017) 36:6049–58. doi: 10.1038/ onc.2017.261

93. Erkes DA, Rosenbaum SR, Field CO, Chervoneva I, Villanueva J, Aplin AE. PLX3397 inhibits the accumulation of intra-tumoral macrophages and improves bromodomain and extra-terminal inhibitor efficacy in melanoma. *Pigment Cell Melanoma Res* (2020) 33:372–7. doi: 10.1111/pcmr.12845

94. Mok S, Koya RC, Tsui C, Xu J, Robert L, Wu L, et al. Inhibition of CSF-1 receptor improves the antitumor efficacy of adoptive cell transfer immunotherapy. *Cancer Res* (2014) 74:153–61. doi: 10.1158/0008-5472.CAN-13-1816

95. Benner B, Good L, Quiroga D, Schultz TE, Kassem M, Carson WE, et al. Pexidartinib, a novel small molecule CSF-1R inhibitor in use for tenosynovial giant cell tumor: A systematic review of pre-clinical and clinical development. *Drug Des Devel Ther* (2020) 14:1693–704. doi: 10.2147/DDDT.S253232

96. Somerhausen N, van den Rijn M. Tenosynovial giant cell tumour, diffuse type. WHO classification tumours soft Tissue Bone (2013) 4:102–3.

97. Cassier PA, Italiano A, Gomez-Roca CA, Le Tourneau C, Toulmonde M, Cannarile MA, et al. CSF1R inhibition with emactuzumab in locally advanced diffuse-type tenosynovial giant cell tumours of the soft tissue: A dose-escalation and dose-expansion phase 1 study. *Lancet Oncol* (2015) 16:949–56. doi: 10.1016/S1470-2045(15)00132-1

98. Tap WD, Gelderblom H, Palmerini E, Desai J, Bauer S, Blay JY, et al. Pexidartinib versus placebo for advanced tenosynovial giant cell turnour (ENLIVEN): A randomised phase 3 trial. *Lancet* (2019) 394(10197):478–87. doi: 10.1016/S0140-6736(19)30764-0

99. Rao R, Han R, Ogurek S, Xue CB, Wu LM, Zhang LG, et al. Glioblastoma genetic drivers dictate the function of tumor-associated macrophages/microglia and responses to CSF1R inhibition. *Neuro-Oncology* (2022) 24:584–97. doi: 10.1093/neuonc/noab228

100. Wesolowski R, Sharma N, Reebel L, Rodal MB, Peck A, West BL, et al. Phase ib study of the combination of pexidartinib (PLX3397), a CSF-1R inhibitor, and paclitaxel in patients with advanced solid tumors. *Ther Adv Med Oncol* (2019) 11:1758835919854238. doi: 10.1177/1758835919854238

101. Wainberg ZA, Eisenberg PD, Sachdev JC, Weise AM, Kaufman DR, Hutchinson M, et al. Phase 1/2a study of double immune suppression blockade by combining a CSF1R inhibitor (pexidartinib/PLX3397) with an anti PD-1 antibody (pembrolizumab) to treat advanced melanoma and other solid tumors. *J Clin Oncol* (2016) 34:TPS465–TPS. doi: 10.1200/jco.2016.34.4_suppl.tps465

102. Butowski N, Colman H, De Groot JF, Omuro AM, Nayak L, Wen PY, et al. Orally administered colony stimulating factor 1 receptor inhibitor PLX3397 in recurrent glioblastoma: An ivy foundation early phase clinical trials consortium phase II study. *Neuro Oncol* (2016) 18:557–64. doi: 10.1093/neuonc/nov245

103. Moskowitz CH, Younes A, de Vos S, Bociek RG, Gordon LI, Witzig TE, et al. CSF1R inhibition by PLX3397 in patients with relapsed or refractory Hodgkin lymphoma: results from a phase 2 single agent clinical trial. *Blood* (2012) 120:1638. doi: 10.1182/blood.V120.21.1638.1638

104. Sun Y, Yang L, Hao X, Liu Y, Zhang J, Ning Z, et al. Phase I dose-escalation study of chiauranib, a novel angiogenic, mitotic, and chronic inflammation inhibitor, in patients with advanced solid tumors. *J Hematol Oncol* (2019) 12:9. doi: 10.1186/s13045-018-0695-0

105. Zhou Y, Fu C, Kong Y, Pan D, Wang Y, Huang S, et al. Antitumor and immunomodulatory effects of a novel multitarget inhibitor, CS2164, in mouse hepatocellular carcinoma models. *Anticancer Drugs* (2019) 30:909–16. doi: 10.1097/cad.00000000000000791

106. Yin H, Xie J, Jiang P, Jiang X, Duan D, Qi J, et al. Chiauranib selectively inhibits colorectal cancer with KRAS wild-type by modulation of ROS through activating the p53 signaling pathway. *Am J Cancer Res* (2020) 10:3666–85.

107. Deng M, Zhao H, Chen Q, Zhao J, Shi Y, Yu L, et al. CS2164 suppresses acute myeloid leukemia cell growth *via* inhibiting VEGFR2 signaling in preclinical models. *Eur J Pharmacol* (2019) 853:193–200. doi: 10.1016/j.ejphar.2019.03.041

108. Gomez-Roca CA, Italiano A, Le Tourneau C, Cassier PA, Toulmonde M, D'Angelo SP, et al. Phase I study of emactuzumab single agent or in combination with paclitaxel in patients with advanced/metastatic solid tumors reveals depletion of immunosuppressive M2-like macrophages. *Ann Oncol* (2019) 30:1381–92. doi: 10.1093/annonc/mdz163

109. Papadopoulos KP, Gluck L, Martin LP, Olszanski AJ, Tolcher AW, Ngarmchamnanrith G, et al. First-in-Human study of AMG 820, a monoclonal anti-Colony-Stimulating factor 1 receptor antibody, in patients with advanced solid tumors. *Clin Cancer Res* (2017) 23:5703–10. doi: 10.1158/1078-0432.CCR-16-3261

110. Lin CC. Clinical development of colony-stimulating factor 1 receptor (CSF1R) inhibitors. J Immunother Precis Oncol (2021) 4:105–14. doi: 10.36401/JIPO-20-32

111. Grosso F, Jones RL, Demetri GD, Judson IR, Blay J-Y, Le Cesne A, et al. Efficacy of trabectedin (ecteinascidin-743) in advanced pretreated myxoid liposarcomas: A retrospective study. *Lancet Oncol* (2007) 8:595–602. doi: 10.1016/S1470-2045(07)70175-4

112. Germano G, Frapolli R, Belgiovine C, Anselmo A, Pesce S, Liguori M, et al. Role of macrophage targeting in the antitumor activity of trabectedin. *Cancer Cell* (2013) 23:249–62. doi: 10.1016/j.ccr.2013.01.008

113. Allavena P, Belgiovine C, Digifico E, Frapolli R, D'Incalci M. Effects of the antitumor agents trabectedin and lurbinectedin on immune cells of the tumor microenvironment. *Front Oncol* (2022) 12:851790. doi: 10.3389/fonc.2022.851790

114. Allavena P, Signorelli M, Chieppa M, Erba E, Bianchi G, Marchesi F, et al. Antiinflammatory properties of the novel antitumor agent yondelis (trabectedin): inhibition of macrophage differentiation and cytokine production. *Cancer Res* (2005) 65:2964–71. doi: 10.1158/0008-5472.CAN-04-4037

115. Barone A, Chi DC, Theoret MR, Chen H, He K, Kufrin D, et al. FDA Approval summary: Trabectedin for unresectable or metastatic liposarcoma or leiomyosarcoma following an anthracycline-containing regimen. *Clin Cancer Res* (2017) 23:7448–53. doi: 10.1158/1078-0432.CCR-17-0898

116. O'Connor T, Heikenwalder M. CCL2 in the tumor microenvironment. *Tumor Microenvironment: Role Chemokines - Pt B* (2021) 1302:1–14. doi: 10.1007/978-3-030-62658-7_1

117. Grossman JG, Nywening TM, Belt BA, Panni RZ, Krasnick BA, DeNardo DG, et al. Recruitment of CCR2(+) tumor associated macrophage to sites of liver metastasis confers a poor prognosis in human colorectal cancer. *Oncoimmunology* (2018) 7: e1470729 doi: 10.1080/2162402X.2018.1470729

118. Xu M, Wang Y, Xia R, Wei Y, Wei X. Role of the CCL2-CCR2 signalling axis in cancer: Mechanisms and therapeutic targeting. In: *Cell proliferation*. John Wiley and Sons Inc (2021), e13115.

119. Oo MW, Kawai H, Takabatake K, Tomida S, Eguchi T, Ono K, et al. Resident stroma-secreted chemokine CCL2 governs myeloid-derived suppressor cells in the tumor microenvironment. *JCI Insight* (2022) 7:e148960. doi: 10.1172/jci.insight.148960

120. Qin R, Ren WH, Ya GQ, Wang B, He J, Ren SX, et al. Role of chemokines in the crosstalk between tumor and tumor-associated macrophages. *Clin Exp Med* (2022) 10:590941. doi: 10.1007/s10238-022-00888-z

121. Loberg RD, Ying C, Craig M, Day LL, Sargent E, Neeley C, et al. Targeting CCL2 with systemic delivery of neutralizing antibodies induces prostate cancer tumor regression *In vivo. Cancer Res* (2007) 67:9417–24. doi: 10.1158/0008-5472.Can-07-1286

122. Zhu X, Fujita M, Snyder LA, Okada H. Systemic delivery of neutralizing antibody targeting CCL2 for glioma therapy. *J Neurooncol* (2011) 104:83–92. doi: 10.1007/s11060-010-0473-5

123. Pienta KJ, Machiels JP, Schrijvers D, Alekseev B, Shkolnik M, Crabb SJ, et al. Phase 2 study of carlumab (CNTO 888), a human monoclonal antibody against CCchemokine ligand 2 (CCL2), in metastatic castration-resistant prostate cancer. *Invest New Drugs* (2013) 31:760–8. doi: 10.1007/s10637-012-9869-8

124. Vela M, Aris M, Llorente M, Garcia-Sanz JA, Kremer L. Chemokine receptorspecific antibodies in cancer immunotherapy: achievements and challenges. *Front Immunol* (2015) 6:12. doi: 10.3389/fimmu.2015.00012

125. Yumimoto K, Sugiyama S, Mimori K, Nakayama KI. Potentials of c-c motif chemokine 2-C-C chemokine receptor type 2 blockers including propagermanium as anticancer agents. *Cancer Sci* (2019) 110:2090–9. doi: 10.1111/cas.14075

126. Mora E, Guglielmotti A, Biondi G, Sassone-Corsi P. Bindarit: an antiinflammatory small molecule that modulates the NFkappaB pathway. *Cell Cycle* (2012) 11:159–69. doi: 10.4161/cc.11.1.18559

127. Steiner JL, Davis JM, McClellan JL, Guglielmotti A, Murphy EA. Effects of the MCP-1 synthesis inhibitor bindarit on tumorigenesis and inflammatory markers in the C3(1)/SV40Tag mouse model of breast cancer. *Cytokine* (2014) 66:60–8. doi: 10.1016/ j.cyto.2013.12.011

128. Zollo M, Di Dato V, Spano D, Martino D, Liguori L, Marino N, et al. Targeting monocyte chemotactic protein-1 synthesis with bindarit induces tumor regression in prostate and breast cancer animal models. *Clin Exp metastasis* (2012) 29:585–601. doi: 10.1007/s10585-012-9473-5

129. Chen P, Zhou J, Li J, Zhang Q, Zuo Q. TIPE1 suppresses osteosarcoma tumor growth by regulating macrophage infiltration. *Clin Trans Oncol* (2019) 21:334–41. doi: 10.1007/s12094-018-1927-z

130. Cho HR, Kumari N, Thi Vu H, Kim H, Park C-K, Choi SH. Increased antiangiogenic effect by blocking CCL2-dependent macrophages in a rodent glioblastoma model: Correlation study with dynamic susceptibility contrast perfusion MRI. *Sci Rep-Uk* (2019) 9:1–12. doi: 10.1038/s41598-019-47438-4

131. Mu X-Y, Wang R-J, Yao Z-X, Zheng Z, Jiang J-T, Tan M-Y, et al. RS 504393 inhibits m-MDSCs recruiting in immune microenvironment of bladder cancer after gemcitabine treatment. *Mol Immunol* (2019) 109:140–8. doi: 10.1016/j.molimm.2019.02.014

132. Wu X, Singh R, Hsu DK, Zhou Y, Yu S, Han D, et al. A small molecule CCR2 antagonist depletes tumor macrophages and synergizes with anti–PD-1 in a murine model of cutaneous T-cell lymphoma (CTCL). *J Invest Dermatol* (2020) 140:1390–400.e4. doi: 10.1016/j.jid.2019.11.018

133. Yao W, Ba Q, Li X, Li H, Zhang S, Yuan Y, et al. A natural CCR2 antagonist relieves tumor-associated macrophage-mediated immunosuppression to produce a therapeutic effect for liver cancer. *EBioMedicine* (2017) 22:58–67. doi: 10.1016/j.ebiom.2017.07.014

134. Farina S, Yang H, Tu GH, Gamelin EC, Lin JC, Wang C, et al. Abstract LB-194: Targeting tumor associated myeloid cells with CCR2 inhibitor PF-04136309 enhances gemcitabine/paclitaxel and doxorubicin anti-tumor activity. *Cancer Res* (2017) 77:LB-194-LB-. doi: 10.1158/1538-7445.AM2017-LB-194

135. Wang-Gillam A, Nywening TM, Sanford DE, Lockhart AC, Suresh R, Tan BR, et al. Phase IB study of FOLFIRINOX plus PF-04136309 in patients with borderline

resectable and locally advanced pancreatic adenocarcinoma (PC). Am Soc Clin Oncol (2015) 338. doi: 10.1200/jco.2015.33.3_suppl.338

136. Noel M, O'Reilly EM, Wolpin BM, Ryan DP, Bullock AJ, Britten CD, et al. Phase 1b study of a small molecule antagonist of human chemokine (CC motif) receptor 2 (PF-04136309) in combination with nab-paclitaxel/gemcitabine in first-line treatment of metastatic pancreatic ductal adenocarcinoma. *Investigational New Drugs* (2020) 38:800-11. doi: 10.1007/s10637-019-00830-3

137. Cherney RJ, Anjanappa P, Selvakumar K, Batt DG, Brown GD, Rose AV, et al. BMS-813160: A potent CCR2 and CCR5 dual antagonist selected as a clinical candidate. ACS Medicinal Chem Letters (2021) 12:1753–8. doi: 10.1021/acsmedchemlett.1c00373

138. Venturini N, Marron T, Casanova-Acebes M, Mandeli J, Doroshow D, Lucas N, et al. 629 neoadjuvant nivolumab combined with CCR2/5 inhibitor or anti-IL-8 antibody in non-small cell lung cancer and hepatocellular carcinoma. *BMJ Specialist Journals* (2022).

139. Le D, Gutierrez ME, Saleh M, Chen E, Mallick AB, Pishvaian MJ, et al. Abstract CT124: A phase lb/II study of BMS-813160, a CC chemokine receptor (CCR) 2/5 dual antagonist, in combination with chemotherapy or nivolumab in patients (pts) with advanced pancreatic or colorectal cancer. *Cancer Res* (2018) 78:CT124–CT. doi: 10.1158/1538-7445.AM2018-CT124

140. Regan DP, Coy JW, Chahal KK, Chow L, Kurihara JN, Guth AM, et al. The angiotensin receptor blocker losartan suppresses growth of pulmonary metastases *via* AT1R-independent inhibition of CCR2 signaling and monocyte recruitment. *J Immunol* (2019) 202:3087–102. doi: 10.4049/jimmunol.1800619

141. Coulson R, Liew SH, Connelly AA, Haines L, Palmer E, Kurihara JN, et al. The angiotensin receptor blocker, losartan, inhibits mammary tumor development and progression to invasive carcinoma. *Oncotarget* (2017) 8:18640–56. doi: 10.18632/ oncotarget.15553

142. Regan DP, Chow L, Das S, Haines L, Palmer E, Kurihara JN, et al. Losartan blocks osteosarcoma-elicited monocyte recruitment, and combined with the kinase inhibitor toceranib, exerts significant clinical benefit in canine metastatic osteosarcoma. *Clin Cancer Res* (2022) 28:662–76. doi: 10.1158/1078-0432.Ccr-21-2105

143. Datta M, Chatterjee S, Perez EM, Gritsch S, Roberge S, Duquette M, et al. Losartan controls immune checkpoint blocker-induced edema and improves survival in glioblastoma mouse models. *Proc Natl Acad Sci* (2023) 120:e2219199120. doi: 10.1073/pnas.2219199120

144. Ha H, Debnath B, Neamati N. Role of the CXCL8-CXCR1/2 axis in cancer and inflammatory diseases. *Theranostics* (2017) 7:1543–88. doi: 10.7150/thno.15625

145. SenGupta S, Hein LE, Parent CA. The recruitment of neutrophils to the tumor microenvironment is regulated by multiple mediators. *Front Immunol* (2021) 12. doi: 10.3389/fimmu.2021.734188

146. Yuen KC, Liu LF, Gupta V, Madireddi S, Keerthivasan S, Li CF, et al. High systemic and tumor-associated IL-8 correlates with reduced clinical benefit of PD-L1 blockade. *Nat Med* (2020) 26:693-+. doi: 10.1038/s41591-020-0860-1

147. Fousek K, Horn LA, Palena C. Interleukin-8: A chemokine at the intersection of cancer plasticity, angiogenesis, and immune suppression. *Pharmacol Therapeut* (2021) 219. doi: 10.1016/j.pharmthera.2020.107692

148. Che JX, Song R, Chen BH, Dong XW. Targeting CXCR1/2: The medicinal potential as cancer immunotherapy agents, antagonists research highlights and challenges ahead. *Eur J Med Chem* (2020) 185:111853. doi: 10.1016/j.ejmech.2019.111853

149. Porter DW, Bradley M, Brown Z, Canova R, Charlton S, Cox B, et al. The discovery of potent, orally bioavailable pyrazolo and triazolopyrimidine CXCR2 receptor antagonists. *Bioorganic Medicinal Chem Letters* (2014) 24:72–6. doi: 10.1016/j.bmcl.2013.11.074

150. Ronchetti I., Boubaker NS, Barba M, Vici P, Gurtner A, Piaggio G. Neutrophil extracellular traps in cancer: Not only catching microbes. *J Exp Clin Cancer Res* (2021) 40:1–9. doi: 10.1186/s13046-021-02036-z

151. Bizzarri C, Pagliei S, Brandolini L, Mascagni P, Caselli G, Transidico P, et al. Selective inhibition of interleukin-8-induced neutrophil chemotaxis by ketoprofen isomers. *Biochem Pharmacol* (2001) 61:1429–37. doi: 10.1016/S0006-2952(01)00610-4

152. Allegretti M, Bertini R, Cesta MC, Bizzarri C, Di Bitondo R, Di Cioccio V, et al. 2-arylpropionic CXC chemokine receptor 1 (CXCR1) ligands as novel noncompetitive CXCL8 inhibitors. *J medicinal Chem* (2005) 48:4312–31. doi: 10.1021/jm049082i

153. Citro A, Cantarelli E, Maffi P, Nano R, Melzi R, Mercalli A, et al. CXCR1/2 inhibition enhances pancreatic islet survival after transplantation. *J Clin Invest* (2012) 122:3647–51. doi: 10.1172/JCI63089

154. Liotti F, De Pizzol M, Allegretti M, Prevete N, Melillo RM. Multiple anti-tumor effects of reparixin on thyroid cancer. *Oncotarget* (2017) 8:35946. doi: 10.18632/ oncotarget.16412

155. Schott AF, Goldstein LJ, Cristofanilli M, Ruffini PA, McCanna S, Reuben JM, et al. Phase ib pilot study to evaluate reparixin in combination with weekly paclitaxel in patients with HER-2–negative metastatic breast CancerReparixin and weekly paclitaxel in metastatic breast cancer. *Clin Cancer Res* (2017) 23:5358–65. doi: 10.1158/1078-0432.CCR-16-2748

156. Goldstein LJ, Mansutti M, Levy C, Chang JC, Henry S, Fernandez-Perez I, et al. A randomized, placebo-controlled phase 2 study of paclitaxel in combination with reparixin compared to paclitaxel alone as front-line therapy for metastatic triple-negative breast cancer (fRida). *Breast Cancer Res Treat* (2021) 190:265–75. doi: 10.1007/s10549-021-06367-5

157. Kemp DM, Pidich A, Larijani M, Jonas R, Lash E, Sato T, et al. Ladarixin, a dual CXCR1/2 inhibitor, attenuates experimental melanomas harboring different molecular defects by affecting malignant cells and tumor microenvironment. *Oncotarget* (2017) 8:14428–42. doi: 10.18632/oncotarget.14803

158. Piro G, Carbone C, Agostini A, Esposito A, De Pizzol M, Novelli R, et al. CXCR1/2 dual-inhibitor ladarixin reduces tumour burden and promotes immunotherapy response in pancreatic cancer. *Br J Cancer* (2022) 128:331–41. doi: 10.1038/s41416-022-02028-6

159. Nicholls DJ, Wiley K, Dainty I, MacIntosh F, Phillips C, Gaw A, et al. Pharmacological characterization of AZD5069, a slowly reversible CXC chemokine receptor 2 antagonist. *J Pharmacol Exp Ther* (2015) 353:340–50. doi: 10.1124/jpet.114.221358

160. Steele CW, Karim SA, Leach JD, Bailey P, Upstill-Goddard R, Rishi L, et al. CXCR2 inhibition profoundly suppresses metastases and augments immunotherapy in pancreatic ductal adenocarcinoma. *Cancer Cell* (2016) 29:832–45. doi: 10.1016//i.ccell.2016.04.014

161. Anderson EM, Thomassian S, Gong J, Hendifar A, Osipov A. Advances in pancreatic ductal adenocarcinoma treatment. *Cancers* (2021) 13:5510. doi: 10.3390/ cancers13215510

162. Guo C, Sharp A, Vogl U, Colombo I, Stathis A, Jain S, et al. 454O a phase (Ph) I/II trial of the CXCR2 antagonist AZD5069 in combination with enzalutamide (ENZA) in patients (pts) with metastatic castration resistant prostate cancer (mCRPC). *Ann Oncol* (2022) 33:S745. doi: 10.1016/j.annonc.2022.07.583

163. Greene S, Robbins Y, Mydlarz WK, Huynh AP, Schmitt NC, Friedman J, et al. Inhibition of MDSC trafficking with SX-682, a CXCR1/2 inhibitor, enhances NK-cell immunotherapy in head and neck cancer ModelsMyeloid cell inhibition enhances NK cellular immunotherapy. *Clin Cancer Res* (2020) 26:1420–31. doi: 10.1158/1078-0432.CCR-19-2625

164. Sun L, Clavijo PE, Robbins Y, Patel P, Friedman J, Greene S, et al. Inhibiting myeloid-derived suppressor cell trafficking enhances T cell immunotherapy. *JCI Insight* (2019) 4:e126853. doi: 10.1172/jci.insight.126853

165. Chatterjee S, Behnam Azad B, Nimmagadda S. The intricate role of CXCR4 in cancer. Adv Cancer Res (2014) 124:31-82. doi: 10.1016/b978-0-12-411638-2.00002-1

166. Luker GD, Yang J, Richmond A, Scala S, Festuccia C, Schottelius M, et al. At The bench: Pre-clinical evidence for multiple functions of CXCR4 in cancer. *J leukocyte Biol* (2021) 109:969–89. doi: 10.1002/JLB.2BT1018-715RR

167. Micallef IN, Stiff PJ, Nademanee AP, Maziarz RT, Horwitz ME, Stadtmauer EA, et al. Plerixafor plus granulocyte colony-stimulating factor for patients with non-Hodgkin lymphoma and multiple myeloma: Long-term follow-up report. *Biol Blood Marrow Transplant* (2018) 24:1187–95. doi: 10.1016/j.bbmt.2018.01.039

168. Uy GL, Rettig MP, Motabi IH, McFarland K, Trinkaus KM, Hladnik LM, et al. A phase 1/2 study of chemosensitization with the CXCR4 antagonist plerixafor in relapsed or refractory acute myeloid leukemia. *Blood* (2012) 119:3917–24. doi: 10.1182/ blood-2011-10-383406

169. Jung K, Heishi T, Incio J, Huang Y, Beech EY, Pinter M, et al. Targeting CXCR4dependent immunosuppressive Ly6Clow monocytes improves antiangiogenic therapy in colorectal cancer. *Proc Natl Acad Sci* (2017) 114:10455–60. doi: 10.1073/pnas.1710754114

170. Dmello RS, To SQ, Chand AL. Therapeutic targeting of the tumour microenvironment in metastatic colorectal cancer. *Int J Mol Sci* (2021) 22:2067. doi: 10.3390/ijms22042067

171. Feig C, Jones JO, Kraman M, Wells RJ, Deonarine A, Chan DS, et al. Targeting CXCL12 from FAP-expressing carcinoma-associated fibroblasts synergizes with anti-PD-L1 immunotherapy in pancreatic cancer. *Proc Natl Acad Sci U S A* (2013) 110:20212–7. doi: 10.1073/pnas.1320318110

172. Bule P, Aguiar SI, Aires-Da-Silva F, Dias JNR. Chemokine-directed tumor microenvironment modulation in cancer immunotherapy. *Int J Mol Sci* (2021) 22:9804. doi: 10.3390/ijms22189804

173. Shen B, Zheng M-Q, Lu J-W, Jiang Q, Wang T-H, Huang X-E. CXCL12-CXCR4 promotes proliferation and invasion of pancreatic cancer cells. *Asian Pacific J Cancer Prev* (2013) 14:5403–8. doi: 10.7314/APJCP.2013.14.9.5403

174. Fearon DT, Janowitz T. AMD3100/Plerixafor overcomes immune inhibition by the CXCL12–KRT19 coating on pancreatic and colorectal cancer cells. *Brit J Cancer* (2021) 125:149–51. doi: 10.1038/s41416-021-01315-y

175. Biasci D, Smoragiewicz M, Connell CM, Wang Z, Gao Y, Thaventhiran JE, et al. CXCR4 inhibition in human pancreatic and colorectal cancers induces an integrated immune response. *Proc Natl Acad Sci* (2020) 117:28960–70. doi: 10.1073/pnas.2013644117

176. Liu T, Li X, You S, Bhuyan SS, Dong L. Effectiveness of AMD3100 in treatment of leukemia and solid tumors: From original discovery to use in current clinical practice. *Exp Hematol Oncol* (2015) 5:19. doi: 10.1186/s40164-016-0050-5

177. Song J-S, Chang C-C, Wu C-H, Dinh TK, Jan J-J, Huang K-W, et al. A highly selective and potent CXCR4 antagonist for hepatocellular carcinoma treatment. *Proc Natl Acad Sci* (2021) 118:e2015433118. doi: 10.1073/pnas.2015433118

178. Le Naour J, Galluzzi L, Zitvogel L, Kroemer G, Vacchelli E. Trial watch: IDO inhibitors in cancer therapy. *OncoImmunology* (2020), 1777625. doi: 10.1080/2162402X.2020.1777625

179. Liu M, Wang X, Wang L, Ma X, Gong Z, Zhang S, et al. Targeting the IDO1 pathway in cancer: From bench to bedside. *J Hematol Oncol* (2018) 11:1–12. doi: 10.1186/s13045-018-0644-y

180. Li F, Zhang R, Li S, Liu J. IDO1: An important immunotherapy target in cancer treatment. *Int immunopharmacology* (2017) 47:70-7. doi: 10.1016/ j.intimp.2017.03.024

181. Jung KH, LoRusso P, Burris H, Gordon M, Bang YJ, Hellmann MD, et al. Phase I study of the indoleamine 2,3-dioxygenase 1 (IDO1) inhibitor navoximod (GDC-0919) administered with PD-L1 inhibitor (atezolizumab) in advanced solid tumors. *Clin Cancer Res* (2019) 25:3220–8. doi: 10.1158/1078-0432.CCR-18-2740

182. Nandre R, Verma V, Gaur P, Patil V, Yang XD, Ramlaoui Z, et al. IDO vaccine ablates immune-suppressive myeloid populations and enhances antitumor effects independent of tumor cell IDO status. *Cancer Immunol Res* (2022) 10:571–80. doi: 10.1158/2326-6066.Cir-21-0457

183. Niu N, Shen W, Zhong Y, Bast RCJr., Jazaeri A, Sood AK, et al. Expression of B7– H4 and IDO1 is associated with drug resistance and poor prognosis in high-grade serous ovarian carcinomas. *Hum pathology* (2021) 113:20–7. doi: 10.1016/j.humpath.2021.04.003

184. Suzuki Y, Suda T, Furuhashi K, Suzuki M, Fujie M, Hahimoto D, et al. Increased serum kynurenine/tryptophan ratio correlates with disease progression in lung cancer. *Lung cancer* (2010) 67:361–5. doi: 10.1016/j.lungcan.2009.05.001

185. Lee SH, Mahendran R, Tham SM, Thamboo TP, Chionh BJ, Lim YX, et al. Tryptophan-kynurenine ratio as a biomarker of bladder cancer. *BJU Int* (2021) 127:445–53. doi: 10.1111/bju.15205

186. Mandarano M, Orecchini E, Bellezza G, Vannucci J, Ludovini V, Baglivo S, et al. Kynurenine/tryptophan ratio as a potential blood-based biomarker in non-small cell lung cancer. *Int J Mol Sci* (2021) 22:4403. doi: 10.3390/ijms22094403

187. de Jong RA, Nijman HW, Boezen HM, Volmer M, Klaske A, Krijnen J, et al. Serum tryptophan and kynurenine concentrations as parameters for indoleamine 2, 3dioxygenase activity in patients with endometrial, ovarian, and vulvar cancer. *Int J Gynecologic Cancer* (2011) 21:1320–7. doi: 10.1097/IGC.0b013e31822017fb

188. Schafer CC, Wang Y, Hough KP, Sawant A, Grant SC, Thannickal VJ, et al. Indoleamine 2,3-dioxygenase regulates anti-tumor immunity in lung cancer by metabolic reprogramming of immune cells in the tumor microenvironment. *Oncotarget* (2016) 7:75407–24. doi: 10.18632/oncotarget.12249

189. Li F, Zhao Y, Wei L, Li S, Liu J. Tumor-infiltrating treg, MDSC, and IDO expression associated with outcomes of neoadjuvant chemotherapy of breast cancer. *Cancer Biol Ther* (2018) 19:695–705. doi: 10.1080/15384047.2018.1450116

190. Holmgaard RB, Zamarin D, Li Y, Gasmi B, Munn DH, Allison JP, et al. Tumorexpressed IDO recruits and activates MDSCs in a treg-dependent manner. *Cell Rep* (2015) 13:412–24. doi: 10.1016/j.celrep.2015.08.077

191. Xu K, Fu Y, Gao H, Bai M, Liu H, Duan Y. L-tryptophan activates the aryl hydrocarbon receptor and induces cell cycle arrest in porcine trophectoderm cells. *Theriogenology* (2021) 171:137–46. doi: 10.1016/j.theriogenology.2021.05.012

192. Yoshida J, Ishibashi T, Nishio M. G1 cell cycle arrest by amlodipine, a dihydropyridine Ca2+ channel blocker, in human epidermoid carcinoma A431 cells. *Biochem Pharmacol* (2007) 73:943–53. doi: 10.1016/j.bcp.2006.12.011

193. Labadie BW, Bao R, Luke JJ. Reimagining IDO pathway inhibition in cancer immunotherapy *via* downstream focus on the tryptophan–Kynurenine–Aryl hydrocarbon AxisTrp–Kyn–AhR immunotherapy. *Clin Cancer Res* (2019) 25:1462–71. doi: 10.1158/1078-0432.CCR-18-2882

194. Van den Eynde BJ, van Baren N, Baurain J-F. Is there a clinical future for IDO1 inhibitors after the failure of epacadostat in melanoma? *Annu Rev Cancer Biol* (2020) 4:241–56. doi: 10.1146/annurev-cancerbio-030419-033635

195. Komiya T, Huang CH. Updates in the clinical development of epacadostat and other indoleamine 2, 3-dioxygenase 1 inhibitors (IDO1) for human cancers. *Front Oncol* (2018) 8:423. doi: 10.3389/fonc.2018.00423

196. Jung KH, LoRusso P, Burris H, Gordon M, Bang Y-J, Hellmann MD, et al. Phase I study of the indoleamine 2, 3-dioxygenase 1 (IDO1) inhibitor navoximod (GDC-0919) administered with PD-L1 inhibitor (Atezolizumab) in advanced solid TumorsNavoximod and atezolizumab in advanced solid tumors. *Clin Cancer Res* (2019) 25:3220-8. doi: 10.1158/1078-0432.CCR-18-2740

197. Sloan EK, Priceman SJ, Cox BF, Yu S, Pimentel MA, Tangkanangnukul V, et al The sympathetic nervous system induces a metastatic switch in primary breast cancer. doi: 10.1158/0008-5472.CAN-10-0522

198. Qiao G, Chen M, Mohammadpour H, MacDonald CR, Bucsek MJ, Hylander BL, et al. Chronic adrenergic stress contributes to metabolic dysfunction and an exhausted phenotype in t cells in the tumor microenvironment. *Cancer Immunol Res* (2021) 9:651–64. doi: 10.1158/2326-6066.CIR-20-0445

199. Braadland PR, Ramberg H, Grytli HH, Taskén KA. β -adrenergic receptor signaling in prostate cancer. Front Oncol (2015) 4:375. doi: 10.3389/fonc.2014.00375

200. Borgatti A, Dickerson EB, Lawrence J. Emerging therapeutic approaches for canine sarcomas: Pushing the boundaries beyond the conventional. *Veterinary Comp Oncol* (2020) 18:9–24. doi: 10.1111/vco.12554

201. Fjaestad KY, Romer AMA, Goitea V, Johansen AZ, Thorseth ML, Carretta M, et al. Blockade of beta-adrenergic receptors reduces cancer growth and enhances the response to anti-CTLA4 therapy by modulating the tumor microenvironment. *Oncogene* (2022) 41:1364–75. doi: 10.1038/s41388-021-02170-0

202. Barathova M, Grossmannova K, Belvoncikova P, Kubasova V, Simko V, Skubla R, et al. Impairment of hypoxia-induced CA IX by beta-blocker propranolol–impact on progression and metastatic potential of colorectal cancer cells. *Int J Mol Sci* (2020) 21:8760. doi: 10.3390/ijms21228760

203. Jin M-Z, Jin W-L. The updated landscape of tumor microenvironment and drug repurposing. Signal transduction targeted Ther (2020) 5:1–16. doi: 10.1038/s41392-020-00280-x

204. Cao M, Huang W, Chen Y, Li G, Liu N, Wu Y, et al. Chronic restraint stress promotes the mobilization and recruitment of myeloid-derived suppressor cells through β -adrenergic-activated CXCL5-CXCR2-Erk signaling cascades. *Int J Cancer* (2021) 149:460–72. doi: 10.1002/ijc.33552

205. Iñigo-Marco I, Alonso MM. Destress and do not suppress: targeting adrenergic signaling in tumor immunosuppression. *J Clin Invest* (2019) 129:5086–8. doi: 10.1172/JCI133115

206. MacDonald C, Ministero S, Pandey M, Robinson D, Forti Hong E, Hylander B, et al. Comparing thermal stress reduction strategies that influence MDSC accumulation in tumor bearing mice. *Cell Immunol* (2021) 361:104285. doi: 10.1016/j.cellimm.2021.104285

207. Jean Wrobel L, Bod L, Lengagne R, Kato M, Prevost-Blondel A, Le Gal FA. Propranolol induces a favourable shift of anti-tumor immunity in a murine spontaneous model of melanoma. *Oncotarget* (2016) 7:77825–37. doi: 10.18632/oncotarget.12833

208. Ammons DT, Guth A, Rozental AJ, Kurihara J, Marolf AJ, Chow L, et al. Reprogramming the Canine Glioma Microenvironment with Tumor Vaccination plus Oral Losartan and Propranolol Induces Objective Responses. *Cancer Treat. Commun* (2022) 2:1657–67. doi: 10.1158/2767-9764.CRC-22-0388

209. Hiller JG, Cole SW, Crone EM, Byrne DJ, Shackleford DM, Pang JMB, et al. Preoperative β -blockade with propranolol reduces biomarkers of metastasis in breast cancer: A phase II randomized trial. Clin Cancer Res (2020) 26:1803–11. doi: 10.1158/1078-0432.CCR-19-2641

210. Gandhi S, Pandey MR, Attwood K, Ji W, Witkiewicz AK, Knudsen ES, et al. Phase I clinical trial of combination propranolol and pembrolizumab in locally advanced and metastatic melanoma: Safety, tolerability, and preliminary evidence of antitumor activity. *Clin Cancer Res* (2021) 27:87–95. doi: 10.1158/1078-0432.CCR-20-2381

211. Phan TT, Ho TT, Nguyen HT, Nguyen HT, Tran TB, Nguyen ST. The prognostic impact of neutrophil to lymphocyte ratio in advanced non-small cell lung cancer patients treated with EGFR TKI. *Int J Gen Med* (2018) 11:423–30. doi: 10.2147/IJGM.S174605

212. Yun NK, Rouhani SJ, Bestvina CM, Ritz EM, Gilmore BA, Tarhoni I, et al. Neutrophil-to-Lymphocyte ratio is a predictive biomarker in patients with epidermal growth factor receptor (EGFR) mutated advanced non-small cell lung cancer (NSCLC) treated with tyrosine kinase inhibitor (TKI) therapy. *Cancers* (2021) 13:1426. doi: 10.3390/cancers13061426

213. Tan C-S, Kumarakulasinghe NB, Huang Y-Q, Ang YLE, Choo JR-E, Goh B-C, et al. Third generation EGFR TKIs: Current data and future directions. *Mol Cancer* (2018) 17:1–14. doi: 10.1186/s12943-018-0778-0

214. Huang I, Jiang S, Shi Y. Tyrosine kinase inhibitors for solid tumors in the past 20 years (2001–2020). J Hematol Oncol (2020) 13:143. doi: 10.1186/s13045-020-00977-0

215. Mena AC, Pulido EG, Guillén-Ponce C. Understanding the molecular-based mechanism of action of the tyrosine kinase inhibitor: Sunitinib. *Anticancer Drugs* (2010) 21 Suppl 1:S3–11. doi: 10.1097/01.cad.0000361534.44052.c5

216. Abdel-Aziz AK, Abdel-Naim AB, Shouman S, Minucci S, Elgendy M. From resistance to sensitivity: Insights and implications of biphasic modulation of autophagy by sunitinib. *Front Pharmacol* (2017) 8. doi: 10.3389/fphar.2017.00718

217. Arora A, Scholar EM. Role of tyrosine kinase inhibitors in cancer therapy. J Pharmacol Exp Ther (2005) 315:971–9. doi: 10.1124/jpet.105.084145

218. London CA. Tyrosine kinase inhibitors in veterinary medicine. *Topics companion Anim Med* (2009) 24:106–12. doi: 10.1053/j.tcam.2009.02.002

219. Hao Z, Sadek I. Sunitinib: The antiangiogenic effects and beyond. *OncoTargets Ther* (2016) 9:5495. doi: 10.2147/OTT.S112242

220. Ko JS, Zea AH, Rini BI, Ireland JL, Elson P, Cohen P, et al. Sunitinib mediates reversal of myeloid-derived suppressor cell accumulation in renal cell carcinoma patients. *Clin Cancer Res* (2009) 15:2148–57. doi: 10.1158/1078-0432.CCR-08-1332

221. London C, Regan D, Chow L, Weishaar K, Gardner H, Thamm D, et al. 840 triple-drug oral immunotherapy targeting myeloid cells for treatment of metastatic osteosarcoma evaluated in spontaneous canine model. *J ImmunoTherapy Cancer* (2022) 10:A876–A. doi: 10.1136/jitc-2022-SITC2022.0840

222. Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc J-F, et al. Sorafenib in advanced hepatocellular carcinoma. *New Engl J Med* (2008) 359:378–90. doi: 10.1056/NEJMoa0708857

223. Cabrera R, Ararat M, Xu Y, Brusko T, Wasserfall C, Atkinson MA, et al. Immune modulation of effector CD4+ and regulatory T cell function by sorafenib in patients with hepatocellular carcinoma. *Cancer Immunol Immunother* (2013) 62:737–46. doi: 10.1007/s00262-012-1380-8

224. Chuang H-Y, Chang Y-F, Liu R-S, Hwang J-J. Serial low doses of sorafenib enhance therapeutic efficacy of adoptive T cell therapy in a murine model by improving tumor microenvironment. *PloS One* (2014) 9:e109992. doi: 10.1371/journal.pone.0109992

225. Keating GM. Sorafenib: A review in hepatocellular carcinoma. *Targeted Oncol* (2017) 12:243–53. doi: 10.1007/s11523-017-0484-7

226. Bruix J, Raoul J-L, Sherman M, Mazzaferro V, Bolondi L, Craxi A, et al. Efficacy and safety of sorafenib in patients with advanced hepatocellular carcinoma: subanalyses of a phase III trial. *J hepatology* (2012) 57:821–9. doi: 10.1016/j.jhep.2012.06.014

227. Kudo M, Finn RS, Qin S, Han K-H, Ikeda K, Piscaglia F, et al. Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. *Lancet* (2018) 391:1163–73. doi: 10.1016/S0140-6736(18)30207-1

228. Hatanaka T, Naganuma A, Kakizaki S. Lenvatinib for hepatocellular carcinoma: A literature review. *Pharmaceuticals* (2021) 14:36. doi: 10.3390/ph14010036

229. Capozzi M, De Divitiis C, Ottaiano A, von Arx C, Scala S, Tatangelo F, et al. Lenvatinib, a molecule with versatile application: From preclinical evidence to future development in anti-cancer treatment. *Cancer Manag Res* (2019) 11:3847–60. doi: 10.2147/cmar.S188316

230. Kato Y, Tabata K, Kimura T, Yachie-Kinoshita A, Ozawa Y, Yamada K, et al. Lenvatinib plus anti-PD-1 antibody combination treatment activates CD8+ T cells through reduction of tumor-associated macrophage and activation of the interferon pathway. *PloS One* (2019) 14:e0212513. doi: 10.1371/journal.pone.0212513

231. Tan H-Y, Wang N, Lam W, Guo W, Feng Y, Cheng Y-C. Targeting tumour microenvironment by tyrosine kinase inhibitor. *Mol Cancer* (2018) 17:43. doi: 10.1186/ s12943-018-0800-6

232. Xu Y, Afify SM, Du J, Liu B, Hassan G, Wang Q, et al. The efficacy of PI3K γ and EGFR inhibitors on the suppression of the characteristics of cancer stem cells. *Sci Rep* (2022) 12:347. doi: 10.1038/s41598-021-04265-w

233. Ramalingam SS, Vansteenkiste J, Planchard D, Cho BC, Gray JE, Ohe Y, et al. Overall survival with osimertinib in untreated, EGFR-mutated advanced NSCLC. *New Engl J Med* (2020) 382:41–50. doi: 10.1056/NEJMoa1913662

234. Nan X, Xie C, Yu X, Liu J. EGFR TKI as first-line treatment for patients with advanced EGFR mutation-positive non-small-cell lung cancer. *Oncotarget* (2017) 8:75712–26. doi: 10.18632/oncotarget.20095

235. Pottier C, Fresnais M, Gilon M, Jérusalem G, Longuespée R, Sounni NE. Tyrosine kinase inhibitors in cancer: Breakthrough and challenges of targeted therapy. *Cancers (Basel)* (2020) 12:731. doi: 10.3390/cancers12030731

236. Law AM, Valdes-Mora F, Gallego-Ortega D. Myeloid-derived suppressor cells as a therapeutic target for cancer. *Cells* (2020) 9:561. doi: 10.3390/ cells9030561

237. Ries CH, Cannarile MA, Hoves S, Benz J, Wartha K, Runza V, et al. Targeting tumor-associated macrophages with anti-CSF-1R antibody reveals a strategy for cancer therapy. *Cancer Cell* (2014) 25:846–59. doi: 10.1016/j.ccr.2014.05.016

238. Puthenveetil A, Dubey S. Metabolic reprograming of tumor-associated macrophages. Ann Transl Med (2020) 8:1030. doi: 10.21037/atm-20-2037