Check for updates

OPEN ACCESS

EDITED BY Huang He, Tianjin University, China

REVIEWED BY

Hongwei Yao, Beijing Friendship Hospital, China Li Zhang, University of Minnesota Twin Cities, United States

*CORRESPONDENCE Dongxin Tang 🔀 tangdongxin1204@sina.com

RECEIVED 26 May 2023 ACCEPTED 07 August 2023 PUBLISHED 23 August 2023

CITATION

Wang X, Pan L, Wang F, Long F, Yang B and Tang D (2023) Interventional effects of oral microecological agents on perioperative indicators of colorectal cancer: a meta-analysis. *Front. Oncol.* 13:1229177. doi: 10.3389/fonc.2023.1229177

COPYRIGHT

© 2023 Wang, Pan, Wang, Long, Yang and Tang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Interventional effects of oral microecological agents on perioperative indicators of colorectal cancer: a meta-analysis

Xueyan Wang¹, Lijun Pan², Feiqing Wang³, Fengxi Long⁴, Bing Yang⁵ and Dongxin Tang⁶*

¹The First College of Clinical Medicine, Guizhou University of Traditional Chinese Medicine, Guiyang, Guizhou, China, ²Department of Medical Affairs, The First Affiliated Hospital of Guizhou University of Traditional Chinese Medicine, Guiyang, Guizhou, China, ³Research Laboratory, The First Affiliated Hospital of Guizhou University of Traditional Chinese Medicine, Guiyang, Guizhou, China, ⁴Development Planning Division, Guizhou University of Traditional Chinese Medicine, Guiyang, Guizhou, China, ⁵Student Management Office, The First Affiliated Hospital of Guizhou University of Traditional Chinese Medicine, Guiyang, Guizhou, China, ⁶Dean's Office, The First Affiliated Hospital of Guizhou University of Traditional Chinese Medicine, Guiyang, Guizhou, China

Purpose: To investigate the efficacy of the application of microecological agents in patients with perioperative colorectal cancer.

Methods: The seven electronic databases including PubMed, Cochrane Library, Excerpt Medica Database (Embase), Web of Science (WOS), Chinese Biomedical Literature Database (CBM), China National Knowledge Infrastructure (CNKI), and Wan-fang Database were systematically searched for eligible studies from 2000 to February 2023.

Results: A total of 38 randomized controlled clinical trials were included in this study, with a total of 1765 patients in the microecological preparation group and 1769 patients in the control group. All data were analyzed using Review Manager 5.4 and R 4.2.2 software. Meta-analysis showed that in the perioperative period of colorectal cancer, the microecological agents group reduced patients' adverse drug reactions, improved intestinal flora with Lactobacillus (SMD, 3.0858, [2.0197; 4.1520], p< 0. 0001), Bifidobacterium (SMD, 2.1551, [1.6145; 2.6956], p< 0.0001) and Escherichia coli (SMD, -1.1393, [-1.6247; -0.6538], p< 0.0001); protection of intestinal mucosal barrier function, endotoxin (SMD, -2.6850 [-4.1399; -1.2301], p=0.0003), DAO (SMD, -2.5916, [-3.4694; -1.7137], p<0.0001) and plasma D-lactate (SMD, -5.4726, [-9.8901; -1.0551], p= 0.0152), reduced inflammatory response, IL-6 (SMD, -3.1279 [-5.7706; -0.4852], p=0.0204) and CRP (SMD, -3.9698 [-7.6296; -0.3100], p=0.0335); improved the immune function of the organism, CD4+ (SMD, 1.5817 [1.0818; 2.0817], p< 0.0001), CD4+/CD8+ (SMD, 1.2938 [0.9693; 1.6183] p< 0.0001) and IgG (SMD, 1.1376 [0.2993; 1.9759] p=0.0078), improved short-term clinical efficacy, ORR (RR, 1.5105 [1.2306; 1.8541], p< 0.0001) and DCR (RR, 0.3896 [0.2620; 0.5795], p< 0.0001).

Conclusion: By increasing the number of beneficial flora such as *Lactobacillus* and *Bifidobacterium* and decreasing the number of harmful flora such as *Escherichia coli*, the micro-ecological preparation group is beneficial in improving the ecological dysregulation in colorectal cancer patients receiving different treatments in the perioperative period. The microecological preparation group was able to reduce many types of adverse drug reactions, such as infections and gastrointestinal discomfort, compared to the control group. The microecological agents also reduced inflammatory responses, decreased the increase in harmful metabolites, enhanced patients' immune function, protected intestinal mucosal barrier function, and improved short-term clinical outcomes.

Systematic review registration: https://inplasy.com/inplasy-2023-4-0051/, identifier INPLASY202340051.

KEYWORDS

colorectal cancer, perioperative period, microecological agents, meta-analysis, intestinal flora

1 Introduction

Colorectal cancer(CRC) is one of the top three causes of cancer deaths worldwide, and the number of cases and deaths are on the rise, and the incidence rate among young people (20-49 years old) has increased significantly, with CRC ranking third in incidence rate and second in mortality rate in 2020 (1, 2).

Microorganisms play a crucial role in human health and disease development, colonizing various parts of our body (3-5), and having different types of crosstalk with various organs, but the highest numbers are found in the intestine (6). Gut microbes interact with the immune system, providing signals to promote the maturation of immune cells and the normal development of immune function (7, 8), which in turn is a major force in the regulation of cancer. Studies have shown that the occurrence of CRC is closely related to disorders of the intestinal microbiota (9).

CRC patients have significant ecological dysbiosis in their intestinal flora, and the various treatments that CRC patients receive during the perioperative period can cause changes in intestinal flora, and intestinal flora disorders can cause a series of adverse effects including increased intestinal inflammatory responses and harmful metabolites. In addition to preoperative mechanical bowel preparation, chemotherapy, radiotherapy, antibiotics and acid suppressants, CRC surgery itself and the stress response to surgery may also affect the intestinal flora and cause significant changes in the intestinal flora structure, which may affect postoperative recovery, short-term complications and longterm oncologic outcomes (10). In recent years, microecological preparations have been successfully used to improve the intestinal microbiota for the treatment of CRC and to mitigate treatmentmediated side effects (11). A large number of probiotic bacteria, their metabolites and other prebiotic components have been shown to influence CRC incidence and mediate intestinal immunity, while they also exhibit anti-inflammatory properties (12, 13). Gut microbial metabolites, which are very important regulators of the interaction between the gut microbiota and the host immune system (14), are abundant and include short chain fatty acids (SCFAs), tryptophan metabolites, vitamins and bile acids. These metabolites have different functions, e.g. Clostridium difficile bacteria, Bifidobacterium bifidum, Streptococcus and Lactobacillus in the gut produce SCFAs that can modulate intestinal immune function by binding to G protein-coupled receptors (GPCRs), and inhibiting the activity of histone deacetylases (HDACs) (15). However, bound bile acids, such as glycochenodeoxycholic acid and glycodeoxycholic acid, promotes tumorigenesis by stimulating cancer cell growth and increasing IL-6 expression (16, 17). And oral microecological agents, not only targeting systemic immunity, are also adept at managing mucosal immunity, thus addressing the inability of systemic immunity to affect the mucosal layer in the colon (18, 19). Microecological preparation is a general term for a class of cultures (live bacteria, dead bacteria or their metabolites) that can effectively participate in the establishment of intestinal micro-ecological balance, promote the growth of normal flora and inhibit the proliferation of pathogens after ingestion by animals, which can improve the health status and growth performance of the organism (20). According to their material composition and mode of action, micro-ecological agents can be divided into three categories: probiotics, prebiotics and synbiotics (21).

In this study, we conducted a systematic evaluation and metaanalysis of intestinal flora alterations, intestinal mucosal barrierrelated factors, immune function-related indices, inflammatory factors, clinical efficacy and adverse effects produced after intervention with microecological agents in the perioperative period of CRC to provide a basis for the involvement of microecological agents in the perioperative treatment of CRC.

2 Methods

The protocol for this systematic review was registered on INPLASY (unique ID number) and is available in full at inpla sy.com (https://inplasy.com/inplasy-2023-4-0051/).

2.1 Eligibility criteria and outcome measures

According to the PICOS acronym (22), the inclusion criteria were as follows:

Participants (P): ① All cases included in the study must have pathologically confirmed CRC, and no metastases to the liver or other sites ② No microecological agents, antibiotics or laxatives within 1 month prior to surgery, have an indication for surgery and undergo radical CRC surgery ③ Approved by the hospital ethics committee, the patient and family understand and are informed, voluntarily participate in this study and sign the informed consent form ④ No restrictions by gender, race or country were found.

Intervention(I): Randomized controlled clinical trial of oral microecological preparations in the perioperative period for colorectal cancer and the content of the microecological preparations is not limited.

Comparison(C): On the basis of the control group, patients in the test group received oral microecological preparations.

Outcomes(O): Clinical efficacy and safety of microecological agents.

Study design(S): Randomized controlled clinical trials.

Exclusion criteria (1): non-randomized controlled trials (2) unclear dose and periodicity of microecological agents (3) incomplete test results (4) lack of sufficient data.

The primary outcome included two efficacy measures: (I) changes in intestinal flora: mainly involving changes in the numbers of Lactobacillus, Bifidobacterium, Escherichia coli, and Enterococcus faecalis; and (II) adverse drug reactions, assessed by detecting hematologic toxicity (leukopenia), gastrointestinal reactions (nausea, vomiting, diarrhea, flatulence), infections (pulmonary, abdominal, urinary, intestinal, incisional), and anastomotic fistulas.

Secondary outcome indicators included four efficacy measures: (i) short-term clinical efficacy ORR, DCR, short-term clinical efficacy according to the World Health Organization (WHO) criteria and Response Evaluation Criteria in Solid Tumors (RECIST), short-term tumor remission including complete remission (CR), partial remission (PR), stable disease remission (SD), progressive disease remission (PD), ORR, disease control rate ORR was defined as the sum of CR and PR, and DCR was the sum of CR, PR, and SD; (ii) immune function indicators CD4+,CD8+, CD4+/CD8+, and IgA, IgG; (iii) intestinal mucosal barrier detection indicators endotoxin, Diamine oxidase (DAO), plasma D-lactate; (iv) inflammatory factors IL-6, TNF- α , CRP.

2.2 Search strategy and study selection

Literature search in both international (Cochrane Library, PubMed, EMBASE, and Web of Science) and Chinese (CBM, CNKI, and Wan-fang Database) databases will be systematically searched for eligible studies from 2000 to February 2023, were independently conducted by two researchers. First, the MeSH database was searched by entering Colorectal Cancer, Intestinal flora, Randomized controlled clinical trials in turn, and then by searching for the terms ((Colorectal Neoplasm or Colorectal Tumor or Colorectal Cancer or Colorectal Carcinoma) AND (Intestinal flora or Gastrointestinal Microbiome or Gut Microbiota or Gastrointestinal Microbial Community or Intestinal Microbiome)) AND (Controlled Clinical Trials, Randomized or Randomized controlled trials or Clinical Studies) or (((Colorectal Neoplasm or Colorectal Tumor or Colorectal Cancer or Colorectal Carcinoma) AND (Intestinal flora or Gastrointestinal Microbiome or Gut Microbiota or Gastrointestinal Microbial Community or Intestinal Microbiome)) AND (Controlled Clinical Trials, Randomized or Randomized controlled trials or Clinical Studies)) AND (Probiotic or Probiotics) for screening. Two investigators independently screened titles and abstracts and then read the full text of the relevant literature to confirm inclusion, and any discrepancies were discussed with a third investigator.

2.3 Data extraction

The following study and participant characteristics were extracted for this study, including first author, year of publication, study type, sample size, mean age of participants, drug type, drug intervention dose and duration, and outcome indicators. Any disagreements were resolved by consensus.

2.4 Quality assessment and evidence level

The quality of the studies was assessed by the Cochrane risk of bias tool Review Manager 5.4. Included studies were assessed at three levels, including low, unclear, and high risk of bias. The review criteria covered seven areas, including random sequence generation, allocation concealment, blinding of investigators sequence generation, allocation concealment, blinding of participants and staff, blinding of participants and staff for outcome assessment, blinding for outcome assessment, incomplete outcome data, selective reporting, and other sources of bias. Sources of bias.

2.5 Statistical analyses

Statistical analyses were performed using Review Manager 5.4 and R 4.2.2 software. The outcomes were mainly represented by risk ratio

(RR) and standardized mean difference (SMD) with its 95% CIs. Twotailed p< 0.05 was considered to be statistically significant. Cochrane's Q test and I² statistics were used to assess heterogeneity between studies; $p \leq 0.1$ or I² > 50% indicated the presence of statistical heterogeneity, and a random-effects model was used to calculate the results when statistical heterogeneity was not present, and a fixedeffects model (common effects model) was used when statistical heterogeneity was not present. Publication bias was tested using funnel plot tests when more than 10 studies reported the same results. Sensitivity analyses were performed by removing one study at a time from the pooled analysis to explore the effect of individual studies on the pooled results. Subgroup analysis was performed according to whether or not combined chemotherapy was administered.

3 Results

3.1 Literature search and study characteristics

A total of 405 papers were initially retrieved, and after screening titles and abstracts, 139 papers were entered for full-text reading, and 38 studies with a total of 1765 patients in the microecological preparation group and 1769 in the control group were finally included for metaanalysis (Figure 1). 38 studies were randomized controlled clinical studies, and their characteristics are shown in Table S1.

3.2 Methodological bias of the included studies

The method of random assignment was clearly described in all 38 studies, suggesting that there was no selection bias in all included samples. Blinding of investigators and subjects was explicitly mentioned 1 in some studies and not specifically described in others, suggesting possible implementation bias and measurement bias. All data were complete and did not appear to be selectively reported. Other biases are unclear, and the characteristics and quality of all included studies are shown in Figure S1.

3.3 Intestinal flora

30 (23–52) reported alterations in intestinal flora (Table 1; Figures 3, S2). *Lactobacillus* ($I^2 = 96.8$), *Bifidobacterium* ($I^2 = 95.3$), *Escherichia coli* ($I^2 = 94.4$) and *Enterococcus faecalis* ($I^2 = 98.1$) were statistically heterogeneous, so the random- effects model was used for all four data. Meta-analysis showed that the microecological agents in the *Lactobacillus* (SMD, 3.0858, [2.0197; 4.1520], p< 0.0001) in the microecological preparation group, both in combination with chemotherapy (SMD, 3.09, [2.02; 4.15], p< 0.0001) and without chemotherapy (SMD, 3.75, [1.78; 5.73], p< 0.0001), improved better



than the control group. The same conclusion was found for *Bifidobacterium* (SMD, 2.1551, [1.6145; 2.6956], p< 0.0001) and *E. coli* (SMD, -1.1393, [-1.6247; -0.6538], p< 0.0001). No statistically significant results were found for both groups in *Enterococcus faecalis* (SMD, -0.7515, [-1.6823; 0.1792], p=0.1135>0.05).

3.4 Adverse drug reactions

A total of 29 trials (24–28, 30–32, 34–39, 41–45, 47, 49–51, 53– 58) reported adverse drug reactions (Table 2; Figure 2), with moderate heterogeneity in nausea ($I^2 = 47.1$). The results showed that compared to the control group, the microecological agent group showed pulmonary infection (RR, 0.3499, [0.1865; 0.6564], p=0.0011), incisional infection (RR, 0.3896, [0.2620; 0.5795], p<0.0001), leukocytopenia (RR, 0.5684, [0.4228; 0.7642], p=0.0002), nausea (RR, 0.5679, [0.4294; 0.7511], p<0.0001),

TABLE 1 Results of meta-analysis of intestinal flora.

Outcomes	Trials	SM	SMD,95% CI	l ² (%)	Q	р	PB
Lactobacillus	28	REM	3.0858, [2.0197; 4.1520]	96.8	< 0.0001	< 0.0001	Yes
Bifidobacterium	30	REM	2.1551, [1.6145; 2.6956]	95.3	< 0.0001	< 0.0001	Yes
Escherichia coli	29	REM	-1.1393, [-1.6247; -0.6538]	94.4	< 0.0001	< 0.0001	Yes
Enterococcus faecalis	24	REM	-0.7515, [-1.6823; 0.1792]	98.1	< 0.0001	0.1135	Yes

Forest of all results are in Figure 2.

CI, confidence interval; PB, Publication bias; SM, statistical method; SMD, Standardized mean difference.

vomiting (RR, 0.5679, [0.4294; 0.7511], p<0.0001), diarrhea (RR, 0.3895, [0.2932; 0.5174], p<0.0001), gastrointestinal distention (RR, 0.4512, [0.2980; 0.6833], p=0.0002), and anastomotic fistula (RR, 0.3630, [0.1780; 0.7403], p=0.0053) were at low risk. The results between the two groups for abdominal infection (RR, 0.3333, [0.0914; 1.2154], p=0.0960), urinary tract infection (RR, 0.6699, [0.2863; 1.5675], p=0.3557), and intestinal infection (RR, 1.0523, [0.3494; 3.1693], p=0.9278) were not statistically significant.

3.5 Short-term clinical efficacy

A total of 293 subjects from 3 trials (26, 29, 39) reported short-term clinical efficacy (Table 3; Figure 4). meta-analysis showed no heterogeneity in ORR and DCR results ($I^2 = 0$). Compared with the control group, the microecological preparation group had better ORR (RR, 1.5105 [1.2306; 1.8541], p< 0.0001) and DCR (RR, 0.3896 [0.2620; 0.5795], p< 0.0001).



FIGURE 2

Adverse drug reactions (A) Forest plot of lung infection analysis results. (B) Forest plot of incision infection analysis results. (C) Forest plot of leukopenia analysis results. (D) Forest plot of nausea analysis results. (E) Forest plot of vomiting analysis results. (F) Forest plot of diarrhea analysis results. (G) Forest plot of the results of the analysis of gastrointestinal distension. (H) Forest plot of the results of anastomotic fistula analysis. (I) Forest plot of the results of the analysis of abdominal infections. (J) Forest plot of urinary tract infection analysis results. (K) Forest plot of the results of the analysis of abdominal infections.



TABLE 2	Results of	meta-analysis	of adverse	reactions.
---------	------------	---------------	------------	------------

Outcomes	Trials	Microecological preparation group	Control group	SM	RR,95% CI	² (%)	Q	р	PB
		(Events/Total)	(Events/ Total)			(70)			
Lung infection	11	11/513	34/514	CEM	0.3499, [0.1865; 0.6564]	0.0	0.9776	0.0011	No
Incisional infection	13	31/704	80/703	CEM	0.3896, [0.2620; 0.5795]	0.0	1.0000	<0.0001	No
Abdominal infection	5	1/209	7/209	CEM	0.3333, [0.0914; 1.2154]	0.0	0.9344	0.0960	No
Urinary tract infection	5	8/282	12/284	CEM	0.6699, [0.2863; 1.5675]	0.0	0.8911	0.3557	No
Intestinal infection	3	5/156	5/162	CEM	1.0523, [0.3494; 3.1693]	0.0	0.4571	0.9278	No

(Continued)

Outcomes	Trials	Microecological preparation group (Events/Total)	Control group (Events/ Total)	SM	RR,95% CI	² (%)	Q	р	РВ
Anastomotic fistula	7	10/468	27/461	CEM	0.3630, [0.1780; 0.7403]	0.0	0.9218	0.0053	No
Lowered white blood cells	7	44/324	81/327	CEM	0.5684, [0.4228; 0.7642]	30.9	0.1922	0.0002	No
Nausea	10	92/409	171/414	REM	0.5679, [0.4294; 0.7511]	47.1	0.0485	<0.0001	Unclear
Vomiting	10	72/409	146/414	CEM	0.5062, [0.4033; 0.6353]	2.5	0.4163	<0.0001	No
Diarrhea	10	49/393	130/398	CEM	0.3895, [0.2932; 0.5174]	0.0	0.9802	<0.0001	No
Gastrointestinal distention	5	24/183	56/187	CEM	0.4512, [0.2980; 0.6833]	35.8	0.1828	0.0002	No

TABLE 2 Continued

Forest of all results are in Figure 4.

CI, confidence interval; CEM, common effects model; PB, Publication bias; REM, random- effects model; RR, relative ratio; SM, statistical method.

3.6 Intestinal mucosal barrier function

A total of 12 trials (25, 32–34, 36, 37, 42, 46, 49–51, 59) reported the detection of intestinal mucosal barrier-related factors (Table 4; Figures 5, S3), endotoxin ($I^2 = 96.7$), DAO ($I^2 = 90.7$) and plasma Dlactate ($I^2 = 97.9$) were statistically heterogeneous and therefore all were calculated using a random effects model. Meta-analysis results showed that compared to the control group, the microecological preparation group improved endotoxin (SMD, -2.6850[-4.1399; -1.2301], p=0.0003), DAO (SMD, -2.5916, [-3.4694; -1.7137], p<0.0001) and plasma D-lactate (SMD, -5.4726, [-9.8901; -1.0551], p= 0.0152) better.

Α	Experim	nental	С	ontrol				Weight	Weight
Study	Events	Total	Events	Total	Risk Ratio	RR	95%-CI	(common)	(random)
Yang Y 2022	39	60	23	60		- 1.70	[1.17; 2.46]	34.2%	29.9%
Xu TC 2022	27	40	19	40		1.42	[0.96; 2.10]	28.2%	27.0%
Liu D 2019	36	47	25	46		1.41	[1.04; 1.92]	37.6%	43.2%
Common effect mode	1	147		146		1.51	[1.23; 1.85]	100.0%	
Random effects mode						1.49	[1.22; 1.83]		100.0%
Heterogeneity: $I^2 = 0\%$, τ^2	= 0, <i>p</i> = 0.	72			0.5 1 2				
В									
	Experim			ontrol				Weight	Weight
Study	Events	Total	Events	Total	Risk Ratio	RR	95%-CI	(common)	(random)
-									
Yang Y 2022	57	60	52	60	+	1.10	[0.98; 1.23]	43.2%	53.8%
Yang Y 2022 Xu TC 2022	57 35	60 40					[0.98; 1.23] [0.94; 1.44]		53.8% 15.5%
•			30	40		- 1.17		24.9%	
Xu TC 2022	35 44	40	30 38	40		- 1.17 1.13	[0.94; 1.44]	24.9% 31.9%	15.5%
Xu TC 2022 Liu D 2019	35 44	40 47	30 38	40 46		- 1.17 1.13 1.13	[0.94; 1.44] [0.97; 1.32]	24.9% 31.9% 100.0%	15.5%
Xu TC 2022 Liu D 2019 Common effect model	35 44	40 47 147	30 38	40 46	0.8 1 1.25	- 1.17 1.13 1.13	[0.94; 1.44] [0.97; 1.32] [1.03; 1.23]	24.9% 31.9% 100.0%	15.5% 30.7%

Short-term clinical efficacy. (A) Forest plot of ORR analysis results. (B) Forest plot of DCR analysis results.

Outcomes	Trials	Microecological preparation group (Events/Total)	Control group (Events/Total)	SM	RR,95% CI	l ² (%)	Q	р	PB
ORR	3	102/147	67/146	CEM	1.5105, [1.2306; 1.8541]	0.0	0.7231	< 0.0001	No
DCR	3	136/147	120/146	CEM	0.3896, [0.2620; 0.5795]	0.0	1.0000	<0.0001	No

TABLE 3 Results of meta-analysis of Short-term clinical efficacy.

3.7 Immune function

A total of 19 trials (23, 24, 26–34, 38–40, 43, 48, 55, 59, 60) reported immune function-related indices (Table 5; Figures 6, S4), CD4+(I² = 90.5), CD8+ (I² = 94.3), CD4+/CD8+ (I² = 85.8), IgA (I² = 85.2), IgG (I² = 93.2) were statistically heterogeneous, so a random-effects model was used. Meta-analysis results showed that the microecological preparation group improved CD4+ (SMD, 1.5817 [1.0818; 2.0817], p< 0.0001) CD4+/CD8+ (SMD, 1.2938 [0.9693; 1.6183] p< 0.0001) and IgG (SMD, 1.1376[0.2993; 1.9759] p=0.0078) compared to the control group. The difference between the two groups for CD8+ (SMD, -0.6248[-1.1885; -0.0611] p=0.0298) and IgA (SMD, 0.4396 [-0.1487; 1.0279], p= 0.1430) were not statistically significant.

3.8 Inflammatory factors

A total of 12 trials (23, 24, 26, 28, 30, 32, 34, 38, 42, 44, 55, 56) reported on inflammatory factor-related indices (Table 6; Figures 7, S5). IL-6 ($I^2 = 95.6$), TNF- α ($I^2 = 94.4$), and CRP ($I^2 = 96.1$) were statistically heterogeneous, so a random-effects model was chosen. IL-6 (SMD, -3.1279[-5.7706; -0.4852], p=0.0204) and CRP (SMD, -3.9698[-7.6296; -0.3100], p=0.0335) were improved in the microecological preparation group compared to the control group. no statistical difference was found in TNF- α (SMD, -5.8744[-13.7876; 2.0388], p=0.1457) between the two groups.

3.9 Publication bias analysis

Funnel plots were used to examine Lactobacillus, Bifidobacterium, Escherichia coli, Enterococcus faecalis, Lung infection, Incisional infections, Nausea, Vomiting. Diarrhea, CD4+, CD8+, CD4+/CD8+, publication bias of TNF- α (Figure 8).

3.10 Sensitivity analysis

To assess the stability of the results. The meta-analysis of the remaining literature was combined after sequentially excluding one literature, and the changes in the combined results were observed to assess whether the results of the original meta-analysis were significantly changed by certain studies (Figure S6).

4 Discussion

In recent years, a growing number of studies have shown that microecological agents can be used to treat CRC and alleviate side effects due to treatment. Meta-analysis included 38 trials containing 3,234 patients to assess whether the addition of microecological agents is beneficial in improving outcome indicators in the perioperative period of CRC.

The development of CRC is strongly associated with disturbances in the gut microbiota. The data showed that the microecological preparation group was beneficial in improving the ecological dysbiosis brought about by various treatments received by CRC patients in the perioperative period, increasing the number of beneficial flora such as *Lactobacillus* and *Bifidobacterium*, while reducing harmful flora such as *E. coli*. However, under certain conditions, it may have the opposite effect. *Bifidobacteria* may play an important role in altering host metabolism during parasitic infections, thereby promoting the development of cholangiocarcinoma (CC) (61). In intrahepatic cholangiocarcinoma (ICC), *Lactobacillus* and *Alloscardovia* were positively correlated with taurocholanol deoxycholic acid. Plasma

TABLE 4 Results of meta-analysis of indicators related to intestinal mucosal barrier function.

Outcomes	Trials	SM	SMD,95% CI	l ² (%)	Q	р	PB
Endotoxin	7	REM	-2.6850, [-4.1399; -1.2301]	96.7	<0.0001	0.0003	Yes
DAO	5	REM	-2.5916, [-3.4694; -1.7137]	90.7	< 0.0001	< 0.0001	Yes
Plasma D-lactate	6	REM	-5.4726, [-9.8901; -1.0551]	97.9	< 0.0001	0.0152	Yes

		Exper	rimenta			Control	Standardised Mean			Weight	Weigh
Study	Total	Mear) Tota	l Mear	n SD	Difference	SMD	95%-CI	(common)	
Hou XH 2022	42	0.3	2 0.100	0 42	2 0.43	3 0.1200	: : 📼	-0.99	[-1.44; -0.53]	24.0%	14.59
Li YD 2021	60	0.03	3 0.010	0 60	0.09	9 0.0200		-3.77	[-4.37; -3.17]	13.7%	14.49
Zhang YJ 2015	60	0.03	3 0.010	0 60	0.09	9 0.0200		-3.77	[-4.37; -3.17]	13.7%	14.49
Zhang JW 2012	30	0.03	3 0.010	0 30	0.09	9 0.0100		-5.92	[-7.13; -4.72]	3.4%	13.49
Xu QW 2019	30	2.40	0.160	0 30	2.58	8 0.1900	·! 🗕	-1.01	[-1.55; -0.47]	17.0%	14.59
Chen W 2021	23	0.0	9 0.020	0 24	4 0.10	0.0300	i 🖶	-0.38	[-0.96; 0.19]	14.9%	14.49
Zhang YL 2020	50	4.48	8 0.850	0 47	7 8.68	3 1.6500		-3.20	[-3.81; -2.60]	13.4%	14.49
Common effect model	295	j		293	3		•	-2.13	[-2.35; -1.90]	100.0%	
Random effects model	1							-2.68	[-4.14; -1.23]		100.0
Heterogeneity: $l^2 = 97\%$, τ	² = 3.73	40, p <	0.01				-6 -4 -2 0 2 4 6				
в		Expo	rimenta			Control	Standardised Mean			Weight	Weigh
Study	Total	Mean) Total	Mean		Difference	SMD	95%-CI	(common)	
							2			()	(
Hou XH 2022	42	2.73	3 0.7000) 42	4.32	0.8100	; -	-2.08	[-2.62; -1.55]	21.7%	20.29
Liu F 2017	41	1.25	5 0.2000) 41	1.72	0.3600	2		[-2.10; -1.10]	24.9%	20.49
Dai AY 2016	39	2.74	4 0.7100) 39	4.36	0.8500			[-2.60; -1.50]	20.4%	20.19
Li YD 2021	60	2.66	6 0.5800) 60	4.53	0.5600		-3.26	[-3.81; -2.71]	20.5%	20.19
Zhang YL 2020	50	3.45	5 0.6500) 47	7.74	1.3500	• j		[-4.76; -3.35]	12.5%	19.29
Common effect model	232	2		229)		4 4	-2.44	[-2.69; -2.19]	100.0%	-
Random effects model								-2.59	[-3.47; -1.71]		100.0%
Heterogeneity: $l^2 = 91\%$, τ^2	² = 0.91	80, p <	0.01				-4 -2 0 2 4				
с											
	E	xperim	nental		Co	ontrol	Standardised Mean			Weight	Weigh
tudy 1	Fotal N	<i>l</i> lean	SD 1	Total N	lean	SD	Difference	SMD	95%-CI	(common)	(random
/ang JH 2020	42	0.62 0	0.5400	42	5.48 0	.8500	*	-6.76	[-7.89; -5.63]	5.6%	16.7%
iu F 2017	41	0.97 (0.1600	41	1.40 0	.2500		-2.03	[-2.57; -1.49]	24.6%	16.8%
	60	1.87 (60	6.75 0	.3200 -	- !! -		18.51; -14.24]	1.6%	16.29
i YD 2021	40	5.35 (0.7500	43	7.84 1	.4900		-2.09	[-2.62; -1.56]	25.3%	16.8%
	43			46	7.57 1	.3400	· · · · · · · · · · · · · · · · · · ·	-1.95	[-2.45; -1.45]	28.5%	16.8%
ong JH 2021	43 46	5.29 (0.9500	40							
)ong JH 2021 Vang Q 2021		5.29 (6.94 1			15.59 2	.7900		-4.02	[-4.72; -3.32]	14.4%	16.8%
i YD 2021 Jong JH 2021 Vang Q 2021 Ihang YL 2020 Common effect model Random effects model	46				15.59 2	2.7900		-2.80	[-4.72; -3.32] [-3.06; -2.53] [-9.89; -1.06]	14.4% 100.0%	16.8% - 100.0%

FIGURE 5

Intestinal mucosal barrier function. (A) Forest plot of endotoxin analysis results. (B) Forest plot of DAO analysis results. (C) Forest plot of plasma Dlactate analysis results.

TABLE 5	Results of	meta-analysis	of immune	function-related	indicators.
---------	------------	---------------	-----------	------------------	-------------

Outcomes	Trials	SM	SMD,95% CI	l ² (%)	Q	р	PB
CD4 ⁺	14	REM	1.5817, [1.0818; 2.0817]	90.5	< 0.0001	< 0.0001	Yes
CD8 ⁺	14	REM	-0.6248, [-1.1885; -0.0611]	94.3	< 0.0001	0.0298	Yes
CD4 ⁺ /CD8 ⁺	13	REM	1.2938, [0.9693; 1.6183]	85.8	< 0.0001	< 0.0001	Yes
IgA	4	REM	0.4396, [-0.1487; 1.0279]	85.2	0.0001	0.1430	Yes
IgG	5	REM	1.1376, [0.2993; 1.9759]	93.2	< 0.0001	0.0078	Yes



Immune function (A) Forest plot of CD4⁺ analysis results. (B) Forest plot of CD8⁺ analysis results. (C) Forest plot of CD4⁺/CD8⁺ analysis results. (D) Forest plot of IgA analysis results. (E) Forest plot of IgG analysis results.

tauroarsodeoxycholic acid was negatively correlated with Pseudomonas spp. and survival time, but positively correlated with vascular invasion (17, 62).

CRC surgery itself and the stress response to surgery can affect patients' postoperative recovery as well as short-term complications. Compared with the control group, the microecological preparation group was able to reduce many types of adverse drug reactions such as infections and gastrointestinal discomfort. The intestinal microflora can influence the efficacy and adverse effects of chemotherapeutic drugs by regulating the body's immune response (63, 64), regulating the body's hormone levels (65, 66), regulating the body's metabolic levels (67, 68), and regulating the metabolism and transport of chemotherapeutic drugs (69-72). Therefore, appropriate supplementation of probiotics, prebiotics or synbiotics by micro-ecological means is beneficial to regulate the homeostasis of the intestinal microflora and thus reduce the adverse effects of chemotherapeutic drugs.

At the same time, the microecological agents also reduced the inflammatory response, decreased the increase of harmful metabolites, enhanced the immune function of patients, and improved short-term

TABLE 6	Results of	meta-analysis	of indicators	related to	inflammatory	factors.
IT OLL O	110301103 01	inicia anatysis	or marcators	1010100 10	maanmacory	1000010.

Outcomes	Trials	SM	SMD,95% CI	l ² (%)	Q	р	PB
IL-6	8	REM	-3.1279, [-5.7706; -0.4852]	95.6	< 0.0001	0.0204	Yes
TNF-α	10	REM	-5.8744, [-13.7876; 2.0388]	94.4	< 0.0001	0.1457	Yes
CRP	8	REM	-3.9698, [-7.6296; -0.3100]	96.1	< 0.0001	0.0335	Yes

Study	Total	Experin		Total	Mean	Control SD		Standard	ised Mea rence	n	SMD	٥	5% CI	Weight (common)	Weig (randor
Study	Total	mean	30	Total	mean	30		Dille	Tence		3110	5	570-01	(common)	francio
Zhao BF 2022	50	119.39 2	2.3400	48	145.43	21.2500		· · · · · · · · · · · · · · · · · · ·			-1.18	[-1.62;	-0.75]	16.1%	12.7
Song G 2022	45	11.83	0.8400	45	16.56	1.2500		*			-4.40	[-5.18;	-3.63]	5.0%	12.6
Wang WX 2022	93	21.12	2.4300	93	27.51	3.3400		•			-2.18	[-2.54;	-1.81]	22.5%	12.7
Xiao S 2022	30	72.12	1.1500	30	87.49	1.1900	\rightarrow				-12.96	[-15.41; -	10.52]	0.5%	11.5
Dong JH 2021	43	72.05 1	5.0600	43	87.64	17.3700					-0.95	[-1.40;	-0.50]	15.0%	12.7
Wang Q 2021	46	71.74 1	4.0500	46	90.27	18.8300					-1.11	[-1.55;	-0.67]	15.4%	12.7
Chen YL 2021	40	13.23	2.8500	39	18.48	2.7600		-			-1.85	[-2.38;	-1.32]	10.6%	12.6
Zhang YX 2020	47	13.56	3.2900	47	18.66	4.2700		2			-1.33	[-1.78;	-0.88]	14.9%	12.7
Common effect model	394			391							-1.67	[-1.84;	-1.501	100.0%	
Random effects model	004							-				[-5.77;			100.0
Heterogeneity: $l^2 = 96\%$, τ^2	- 1/ 20	62 n < 0 (N1					<u>г т Т</u>	 		۳	[-5.11]	-0.45]		100.0
Theterogeneity. 7 = 50 %, t	- 14.30	υz, μ ~ υ.ι					-15 -1	10 -5	05	10	15				
В															
		Experin				Control		Standardis						Weight	
Study	Tota	l Mean	SD	Total	Mean	SD		Differe	ence		SMD	959	%-CI (common) (random
Zhao BF 2022	50	0 10.54	4.2200	48	22.52	7.3700					-1.99 [-2.48; -1	.50]	10.4%	10.1%
Song G 2022	4	5 42.75	4.6700	45	48.65	5.0300					-1.21	-1.66; -0	.75]	12.1%	10.1%
Li XW 2018	3	5 38.23	7.2500	35	47.12	9.1800					-1.06	-1.57; -0	.56]	9.8%	10.1%
Wang WX 2022	93	3 23.12	4.2000	93	31.32	5.4300		•				-2.02; -1		21.9%	10.1%
Xiao S 2022	30	0 14.82	0.1500	30	22.79	0.1900 -					-45.96 [-	54.46; -37	.45]	0.0%	9.1%
Yang Y 2022	6	2.98	0.4400	60	4.69	0.5300						-4.06; -2		7.5%	10.1%
Dong JH 2021	43	3 14.87	4.0600	43	22.49	3.5600		0				-2.50; -1		9.1%	10.1%
Wang Q 2021	40				23.05							-2.94; -1		8.4%	10.1%
Chen YL 2021	40		0.7200	39		0.9400						-1.98; -0		9.8%	10.1%
Zhang YX 2020	4				15.78							-2.15; -1	-	11.0%	10.1%
Common effect mode	489	•		486							-1.81	-1.97; -1	661	100.0%	
Random effects mode		-						-				13.79; 2	-		100.0%
Heterogeneity: 1 ² = 94%,		.2454, p <	< 0.01				-40	-20 0	20	40	-0101 [10.10, 1			100107
с							-40	-20 0	20	40					
C		Experin	nental			Control		Standardi	sed Mear	1				Weight	Weig
Study	Total	Mean	SD	Total	Mean	SD		Diffe	ence		SMD	9	5%-CI	(common)	(randon
Zhao BF 2022	50	51.39	7.3200	50	82.18	11.9200					-3.09	[-3.68; -	2.50]	12.3%	12.6
Li XW 2018	35	7.13	1.2600	35	9.82	1.6900					-1.78	[-2.34; -	1.23]	13.6%	12.7
Xu QW 2019	30	20.20	4.4000	30	46.70	5.3000		*			-5.37	[-6.48; -	4.26]	3.4%	12.5
Xiao S 2022	30	21.65	1.1300	30	42.42	1.2000	<u> </u>				-17.59	-20.88; -1	4.30]	0.4%	11.5
Dong JH 2021	43	21.41 1	0.2200	43	42.08	12.1900		•			-1.82	[-2.33; -	1.32]	16.5%	12.7
Shen WT 2021	30	2.24	1.9500	30	4.47	2.4500		•			-0.99	[-1.53; -	0.46]	14.6%	12.7
Wang Q 2021	46		9.2400	46		11.1600		•				[-2.15; -	-	18.6%	12.7
Chen YL 2021	40			39		2.4000						[-1.14; -	-	20.5%	12.7
Common effect model	304			303							-1.77	[-1.98;	1.57]	100.0%	
Random effects model								-				[-7.63;	-		100.0
2	2 - 07 4	752 n < 0	01					1			٦		- 1		
Heterogeneity: /2 = 96%, t	= 21.41						-20	-10 () 1(20				

clinical outcomes. The harmful metabolites of the gut flora, such as ammonia, phenols and p-cresol, are involved in the development and progression of cancer through chronic inflammation and DNA damage (73, 74). For example, high levels of lipopolysaccharide (LPS), entering the bloodstream can cause a number of severe pathophysiological responses, including fever, coagulation and shock, by disrupting the host's immune, complement and coagulation systems (75). Primary bile acids enter the large intestine and are converted by intestinal bacteria into secondary bile acids, a class of metabolites with pro-cancer effects that can promote tumour development by stimulating oxidative stress (e.g. reactive oxygen species and reactive nitrogen species), inducing cellular DNA damage and activating EGFR and NF-KB (76-78).

5 Conclusion

The subtle interactions between the intestinal flora and human physiology can influence multiple aspects of health. Microbialepithelial interactions can maintain intestinal barrier function, modulate resistance to infection and intestinal immune function, and maintain host metabolism.

CRC is one of the top three causes of cancer deaths worldwide, and surgery is the primary treatment for colorectal cancer. However, trauma, disturbance of normal intestinal flora, decreased intestinal mucosal barrier function, increased systemic inflammation, decreased immune function, and also the risk of postoperative infection may occur after surgery (79).



Probiotics have antitumor activity by a variety of mechanisms. The most common probiotic flora are two genera of Lactobacilli and Bifidobacteria, which are naturally present in the human digestive system. For example, antioxidants produced by Lactobacilli are able to fight against angiogenic factors, reduce DNA damage, reduce inflammation and tumor size, and inhibit the expression of tumorspecific proteins and polyamine components (80). In addition, prebiotics are fermentable components present in foods that alter the composition and activity of the intestinal microbiota and promote host health. One of the most commonly used prebiotics is resistant starch, which increases the biological activity of a wide range of probiotic bacteria, especially bifidobacteria, and modifies the immune response (81). Prebiotics are organic substances that are not digested or absorbed by the host, but can selectively promote the metabolism and proliferation of beneficial bacteria in the body, thereby improving the health of the host. Commonly used prebiotics include Fructo oligosaccharide, xylo-oligosaccharides and inulin. Studies have shown that Fructo oligosaccharide can reduce the number and activity of carcinogenic enzymes and regulate the body's immune capacity, and the short-chain fatty acids and lactic acid it ferments to produce in the colon can reduce intestinal pH and ammonia concentration, which is conducive to the reduction and inhibition of intestinal spoilage substances (82) Xylo-oligosaccharides can inhibit the invasion of exogenous pathogenic bacteria, improve the body's immune response and protect the barrier function of the intestinal mucosa (83).

Our study found that in the perioperative period of CRC, a more effective treatment regimen in the microecological agent group was accompanied by reduced adverse drug reactions in patients, improved intestinal flora, improved short-term clinical outcomes, enhanced body immune function, and reduced inflammatory responses. According to the World Health Organization, appropriate doses of probiotics are beneficial to human health. Proper consumption of microecological agents, such as probiotics or prebiotics, may be a promising way to prevent and treat CRC.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

Author contributions

Conceptualization: XW. Methodology: XW. Formal analysis and investigation: XW and LP. Writing—original draft preparation: XW. Writing—review and editing: XW, FW. Funding acquisition: DT and BY. Resources: XW. Supervision: DT and FL. All authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

10.3389/fonc.2023.1229177

Funding

This work was financially supported by National Natural Science Foundation of China (nos. 82260957, 82274610, 81860819, 81960818), the Key Research on "TCM Modernization" of National Key Research and Development Plan (2019YFC171250401), Guizhou Traditional Chinese Medicine Tumor Inheritance and Science and Technology Innovation Talent Base (No. Deaf leader-[2018] No. 3), Guizhou high-level innovative talent training plan (100 levels) (no. Yankehe Talents [2016] no. 4032), TCM graduate school workstation (No. Teaching and research JYSZ-[2014]018), The Big Health Science and Technology Cooperation Project of the First Affiliated Hospital of Guizhou University of Traditional Chinese Medicine (No. Building branch [2019] 9-2, No. Building branch [2019] 9-2-35, No. Building branch [2019] 9-2-30), Guizhou Province Traditional Chinese Medicine Oncology Inheritance and Scientific and Technological Innovation Talent Team (Qian Kehe Platform Talents [2020]5013), Guizhou Provincial Science and Technology Plan Project (Qianke Foundation ZK[2022] 487, Qianke Foundation ZK[2022] 498, Qianke Foundation [2021] 095).

References

1. Sharma R, Abbasi-Kangevari M, Abd-Rabu R, Abidi H, Abu-Gharbieh E, Acuna JM, et al. Global, regional, and national burden of colorectal cancer and its risk factors, 1990–2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet Gastroenterol Hepatol* (2022) 7(7):627–47. doi: 10.1016/S2468-1253(22)00044-9

2. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: Cancer J Clin* (2021) 71(3):209–49. doi: 10.3322/caac.21660

3. Neugent ML, Hulyalkar NV, Nguyen VH, Zimmern PE, De Nisco NJ. Advances in understanding the human urinary microbiome and its potential role in urinary tract infection. *mBio* (2020) 11(2):e00218-20. doi: 10.1128/mBio.00218-20

4. Sender R, Fuchs S, Milo R. Are we really vastly outnumbered? Revisiting the ratio of bacterial to host cells in humans. *Cell* (2016) 164(3):337-40. doi: 10.1016/j.cell.2016.01.013

5. de Vos WM, Tilg H, Van Hul M, Cani PD. Gut microbiome and health: mechanistic insights. *Gut* (2022) 71(5):1020-32. doi: 10.1136/gutjnl-2021-326789

6. Lloyd-Price J, Mahurkar A, Rahnavard G, Crabtree J, Orvis J, Hall AB, et al. Strains, functions and dynamics in the expanded Human Microbiome Project. *Nature* (2017) 550(7674):61–6. doi: 10.1038/nature23889

7. de Groot P, Nikolic T, Pellegrini S, Sordi V, Imangaliyev S, Rampanelli E, et al. Faecal microbiota transplantation halts progression of human new-onset type 1 diabetes in a randomised controlled trial. *Gut* (2021) 70(1):92–105. doi: 10.1136/ gutjnl-2020-322630

8. Kootte RS, Levin E, Salojärvi J, Smits LP, Hartstra AV, Udayappan SD, et al. Improvement of insulin sensitivity after lean donor feces in metabolic syndrome is driven by baseline intestinal microbiota composition. *Cell Metab* (2017) 26(4):611– 9.e6. doi: 10.1016/j.cmet.2017.09.008

9. Tilg H, Adolph TE, Gerner RR, Moschen AR. The intestinal microbiota in colorectal cancer. *Cancer Cell* (2018) 33(6):954–64. doi: 10.1016/j.ccell.2018.03.004

10. Wu Y, Wang T. Progress of research on the effect of perioperative treatment of colorectal cancer on intestinal flora. *Shandong Med J* (2022) 62(10):5. doi: 10.3969/j.issn.1002-266X.2022.10.023

11. Bhandari A, Woodhouse M, Gupta S. Colorectal cancer is a leading cause of cancer incidence and mortality among adults younger than 50 years in the USA: a SEER-based analysis with comparison to other young-onset cancers. J Invest Medicine: Off Publ Am Fed Clin Res (2017) 65(2):311–5. doi: 10.1136/jim-2016-000229

12. Singh S, Singh M, Gaur S. Probiotics as multifaceted oral vaccines against colon cancer: A review. *Front Immunol* (2022) 13:1002674. doi: 10.3389/fimmu.2022.1002674

13. Khani S, Hosseini HM, Taheri M, Nourani MR, Imani Fooladi AA. Probiotics as an alternative strategy for prevention and treatment of human diseases: a review. *Inflammation Allergy Drug Targets* (2012) 11(2):79–89. doi: 10.2174/187152812800392832

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2023.1229177/ full#supplementary-material

14. Levy M, Thaiss CA, Elinav E. Metabolites: messengers between the microbiota and the immune system. Genes Dev (2016) 30(14):1589–97. doi: 10.1101/gad.284091.116

15. Hao X, Lv H, Zhang Q, Liu H, Yi F. Progress in the study of gut microbial metabolites mediating intestinal immunity. *Feed Res* (2023) 1–16.

16. Özdirik B, Müller T, Wree A, Tacke F, Sigal M. The role of microbiota in primary sclerosing cholangitis and related biliary Malignancies. *Int J Mol Sci* (2021) 22 (13):6975. doi: 10.3390/ijms22136975

17. Jia X, Lu S, Zeng Z, Liu Q, Dong Z, Chen Y, et al. Characterization of gut microbiota, bile acid metabolism, and cytokines in intrahepatic cholangiocarcinoma. *Hepatol (Baltimore Md)* (2020) 71(3):893–906. doi: 10.1002/hep.30852

18. Shah T, Baloch Z, Shah Z, Cui X, Xia X. The intestinal microbiota: impacts of antibiotics therapy, colonization resistance, and diseases. *Int J Mol Sci* (2021) 22 (12):6597. doi: 10.3390/ijms22126597

19. Durán-Lobato M, Niu Z, Alonso MJ. Oral delivery of biologics for precision medicine. Advanced materials (Deerfield Beach Fla) (2020) 32(13):e1901935. doi: 10.1002/adma.201901935

20. Guarner F, Schaafsma GJ. Probiotics. Int J Food Microbiol (1998) 39(3):237-8. doi: 10.1016/S0168-1605(97)00136-0

21. Yu L, Ma L, Du Y, Liu S, Ping Y, Hu Y, et al. Advances in microecological preparations. *Chin J Microecolog* (2012) 24(01):84–6.

22. Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA Statement. *Open medicine: peer-reviewed independent open-access J* (2009) 3(3):e123–30. doi: 14306/renhyd.18.3.114

23. Xiao S. Analysis of the significance of oxaliplatin in combination with Bifidobacterium trisporium capsules on immune function in postoperative colorectal cancer patients. *Modern Drug Appl China* (2022) 16(13):3. doi: 10.14164/j.cnki.cn11-5581/r.2022.13.039

24. Zhao B. A randomized controlled study of compound probiotics on postoperative rehabilitation of patients with colorectal cancer. [master's thesis]. Guilin Medical College (2022) 9-19. doi: 10.27806/d.cnki.gglyx.2022.000026

25. Hou X. Study on the effect of Bifidobacterium bifidum triptans capsules on postoperative intestinal mucosal function in colorectal cancer patients. *Sci Tech Inf Gansu* (2022) 003):051. doi: 10.3969/j.issn.1672-6375.2022.03.025

26. Yang Y, He Y, Jia F, Li E, Zhi J. Effects of Bifidobacterium triple viable capsule combined with FOLFOX regimen on intestinal micro-ecology and anti-tumor immune response after colorectal cancer surgery. *J Hunan Normal University(Medical Sciences)* (2022) 19(01):95–8.

27. Han Z, Wang J, Li H, Wang J, Sun Y. Effect of bifidobacterium combined with adjuvant chemotherapy on intestinal flora and immune function in patients with colorectal cancer. *Med Innovation China* (2022) 19(01):29–33.

28. Song G. Influence of hypodermic long-standing vaccum drainage combined with microecological preparation on the incision infected rate and enteric flora change of colorectal cancer patients underwent surgery. *Chin J Coloproctology* (2022) 42(1):3.

29. Xu T. Effect of bifdobacterium triptans combined with rapid rehabilitation care on intestinal flora and CD4 CD8 CD4 _CD8 levels in postoperative patients with colorectal cancer. *Primary Care Forum* (2022) 026):026. doi: 10.19435/j.1672-1721.2022.26.042

30. Wang W, Xie J, Wang Y. Bifidobacterium tetrakis tablets in combination with enteral nutrition emulsion Rexin in postoperative colon cancer. *Pract Clin J Integrated Traditional Chin Western Med* (2022) 22(14):4. doi: 10.13638/j.issn.1671-4040.2022.14.007

31. Li M. Influence of bifibacterium tripletvaccine combined with XELOX scheme of chemotherapy on postoperative T-cell subpopulation and intestinal flora of colorectal cancer patients. *Chin J Coloproctology* (2021) 41(3):3.

32. Dong J, Cheng X, Yang Z, Li M, Yu K. Curative efficacy of bifidobacterium triple viable capsule combined with oxaliplatin in treatment of postoperative colorectal cancer and its effects on intestinal flora and immune function. *Med J West China* (2019) 31(3):5.

33. Chen W. Effect of probiotics combind with enteral nutrition on postoperative chemotherapy in patients with colorectal cancer. [master's thesis]. Hubei Medical College (2022) 10-22. doi: 10.27913/d.cnki.ghyby.2021.000038

34. Wang Q, Gong L, Zheng H. Effects of live Clostridium butyricum tablets on intestinal flora balance, toxic and side effects, and immune inflammatory indexes in colorectal cancer patients on postoperative FOLFOX4 chemotherapy. *World Chin J Digestology* (2021) 29(08):435–42. doi: 10.11569/wcjd.v29.i8.435

35. Liu Y. Relationship between intestinal microbial community and prognosis in patients with advanced colorectal cancer chemotherapy. *Chin J Microecology* (2021) 33 (08):958–61 + 66. doi: 10.13381/j.cnki.cjm.202108019

36. Li Y, Wu C, Li H. The status of intestinal flora and the treatment of Bifidobacterium after laparoscopic operation in patients with colon cancer. J Hunan Normal University(Medical Sciences) (2021) 18(2):4.

37. Wang J, Wu L. Effect of laparoscopic surgery combined with microecological agents on intestinal flora and intestinal barrier function in patients with colorectal cancer. *Chin J Microecology* (2020) 32(03):298–301 + 5. doi: 10.13381/j.cnki.cjm.202003012

38. Zhang Y, Zhou X, Gao X. Effect of probiotic intervention on postoperative chemotherapy complications, prognosis and intestinal flora in patients with colon cancer. *Modern Interventional Diagnosis Treat Gastroenterology* (2020) 25(06):768–71.

39. Liu D, Su W, Wu Z. Effect of Bifidobacterium trivium combined with XELOX chemotherapy on intestinal flora and immune function after colorectal cancer surgery. *China Med Herald* (2019) 16(32):119–22.

40. Cheng P. Effect of oral probiotic preparations on intestinal flora in patients after radical surgery for colon cancer. *Health Prot Promotion* (2019) 345(08):31–2. doi: CNKI:SUN:YSXD.0.2019-08-026

41. Liu H, Liu Y, Peng R, Zuo W, Gou J, Li Y, et al. Analysis of the effect of probiotics on intestinal flora in patients undergoing chemotherapy for colon cancer. *World Latest Med Inf* (2019) 19(45):56. doi: 10.19613/j.cnki.1671-3141.2019.45.027

42. Xu Q, Xu P, Cen Y, Li W. Effects of preoperative oral administration of glucose solution combined with postoperative probiotics on inflammation and intestinal barrier function in patients after colorectal cancer surgery. *Oncol Lett* (2019) 18 (1):694–8. doi: 10.3892/ol.2019.10336

43. Yu S. Effects of preoperative nutritional support combined with intestinal probiotic intervention on patients with colorectal cancer. *Med J Natl Defending Forces Southwest China* (2018) 28(09):837–9.

44. Li X. Influence of perioperative supplement of Bifid Triple Viable Capsules on inflammatory reactions and intestinal microecology in patients with colorectal cancer. *Chin J Microecolog* (2018) 30(01):75–8. doi: 10.13381/j.cnki.cjm.201801019

45. Zhan Y. Effect of probiotics on intestinal flora in patients with colon cancer undergoing chemotherapy. *China Pract Med* (2018) 13(32):105-7. doi: 10.14163/j.cnki.11-5547/r.2018.32.057

46. Liu F. Influence of Bifid Triple Viable Capsules on intestinal flora and intestinal mucosal barrier function of patients with post-operation diarrhea after colostomy. *Chin J Microecolog* (2017) 29(07):814–7. doi: 10.13381/j.cnki.cjm.201707016

47. Sun C, He Y, He R. Study on the clinical effect of preoperative application of microecological agents for colorectal cance. *Electronic J Clin Med Literature* (2017) 4 (01):34–5. doi: 10.16281/j.cnki.jocml.2017.01.020

48. Zuo J, Liu G, Yuan K. Effect of Bifidobacterium bifidum supplementation with Lactobacillus rhamnosus triplex on intestinal flora after intestinal surgery. *China Modern Med* (2017) 24(05):25–7.

49. Dai A. Influence of Bifid Triple Viable Capsules on intestinal flora and intestinal permeability after surgery for colorectal cancer. *Chin J Microecolog* (2016) 28(04):425–8. doi: 10.13381/j.cnki.cjm.201604013

50. Zhang Y, Zhou Z, Huang R. Analysis of the effect of preoperative intestinal flora supplementation with Bifidobacterium trisporus after colorectal cancer surgery. *Strait Pharm J* (2015) 27(10):155–6.

51. Zhang JW, Du P, Gao J, Yang BR, Fang WJ, Ying CM. Preoperative probiotics decrease postoperative infectious complications of colorectal cancer. *Am J Med Sci* (2012) 343(3):199–205. doi: 10.1097/MAJ.0b013e31823aace6

52. Liu Z, Qin H, Yang Z, Xia Y, Liu W, Yang J, et al. Randomised clinical trial: the effects of perioperative probiotic treatment on barrier function and post-operative infectious complications in colorectal cancer surgery - a double-blind study. *Alimentary Pharmacol Ther* (2011) 33(1):50–63. doi: 10.1111/j.1365-2036.2010.04492.x

53. Huang F, Li S, Chen W, Han Y, Yao Y, Yang L, et al. Postoperative probiotics administration attenuates gastrointestinal complications and gut microbiota dysbiosis caused by chemotherapy in colorectal cancer patients. *Nutrients* (2023) 15(2):356. doi: 10.3390/nu15020356

54. Li Y. Effects of xylooligosaccharide on improving quality of life in colorectal cancer patients receiving chemotherapy by regulating gut microbiota. [master's thesis]. Jiangnan University (2022). 10-16 doi: 10.27169/d.cnki.gwqgu.2022.001569

55. Shen W, Cai G, Luo Y, Li Z, Lin K, Gu H, et al. Effect of microecological agents on gastrointestinal and inflammatory responses in patients undergoing chemotherapy for colorectal cancer. J Shantou Univ Med College (2021) 34(4):4. doi: 10.13401/j.cnki.jsumc.2021.04.005

56. Chen Y, Zhang Y, Che Z. Effect of intestinal microecological intervention on complications after operation and during chemotherapy in patients with colon cancer. *J Clin Med Pract* (2021) 25(20):73–7.

57. Kotzampassi K, Stavrou G, Damoraki G, Georgitsi M, Basdanis G, Tsaousi G, et al. A four-probiotics regimen reduces postoperative complications after colorectal surgery: A randomized, double-blind, placebo-controlled study. *World J Surg* (2015) 39 (11):2776–83. doi: 10.1007/s00268-015-3071-z

58. Aisu N, Tanimura S, Yamashita Y, Yamashita K, Maki K, Yoshida Y, et al. Impact of perioperative probiotic treatment for surgical site infections in patients with colorectal cancer. *Exp Ther Med* (2015) 10(3):966–72. doi: 10.3892/etm.2015.2640

59. Zhang Y, Chen Q, Liu Y. Effect of microecological agents on early colorectal cancer after radical resection. *Chin J Microecolog* (2020) 32(10):1167–72. doi: 10.13381/j.cnki.cjm.202010012

60. Xie X, He Y, Li H, Yu D, Na L, Sun T, et al. Effects of prebiotics on immunologic indicators and intestinal microbiota structure in perioperative colorectal cancer patients. *Nutr* (*Burbank Los Angeles County Calif)* (2019) 61:132–42. doi: 10.1016/j.nut.2018.10.038

61. Ketpueak T, Thiennimitr P, Apaijai N, Chattipakorn SC, Chattipakorn N. Association of chronic opisthorchis infestation and microbiota alteration on tumorigenesis in cholangiocarcinoma. *Clin Trans gastroenterology.* (2020) 12(1): e00292. doi: 10.14309/ctg.000000000000292

62. Rao B, Ren T, Wang X, Wang H, Zou Y, Sun Y, et al. Dysbiosis in the human microbiome of cholangiocarcinoma. *Front Physiol* (2021) 12:715536. doi: 10.3389/fphys.2021.715536

63. Viaud S, Saccheri F, Mignot G, Yamazaki T, Daillère R, Hannani D, et al. The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. *Sci (New York NY)* (2013) 342(6161):971–6. doi: 10.1126/science.1240537

64. Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. *Sci (New York NY)* (2013) 342(6161):967–70. doi: 10.1126/science.1240527

65. Sfanos KS, Markowski MC, Peiffer LB, Ernst SE, White JR, Pienta KJ, et al. Compositional differences in gastrointestinal microbiota in prostate cancer patients treated with androgen axis-targeted therapies. *Prostate Cancer Prostatic Dis* (2018) 21 (4):539–48. doi: 10.1038/s41391-018-0061-x

66. Pernigoni N, Zagato E, Calcinotto A, Troiani M, Mestre RP, Calì B, et al. Commensal bacteria promote endocrine resistance in prostate cancer through androgen biosynthesis. *Sci* (*New York NY*) (2021) 374(6564):216–24. doi: 10.1126/science.abf8403

67. He Y, Fu L, Li Y, Wang W, Gong M, Zhang J, et al. Gut microbial metabolites facilitate anticancer therapy efficacy by modulating cytotoxic CD8(+) T cell immunity. *Cell Metab* (2021) 33(5):988–1000.e7. doi: 10.1016/j.cmet.2021.03.002

68. Geng HW, Yin FY, Zhang ZF, Gong X, Yang Y. Butyrate suppresses glucose metabolism of colorectal cancer cells via GPR109a-AKT signaling pathway and enhances chemotherapy. *Front Mol Biosci* (2021) 8:634874. doi: 10.3389/fmolb.2021.634874

69. Yan A, Culp E, Perry J, Lau JT, MacNeil LT, Surette MG, et al. Transformation of the anticancer drug doxorubicin in the human gut microbiome. *ACS Infect diseases* (2018) 4(1):68–76. doi: 10.1021/acsinfecdis.7b00166

70. Okuda H, Nishiyama T, Ogura K, Nagayama S, Ikeda K, Yamaguchi S, et al. Lethal drug interactions of sorivudine, a new antiviral drug, with oral 5-fluorouracil prodrugs. *Drug Metab Dispos* (1997) 25(2):270–3.

71. Scott TA, Quintaneiro LM, Norvaisas P, Lui PP, Wilson MP, Leung KY, et al. Host-microbe co-metabolism dictates cancer drug efficacy in *C. elegans. Cell* (2017) 169 (3):442–56.e18. doi: 10.1016/j.cell.2017.03.040

72. García-González AP, Ritter AD, Shrestha S, Andersen EC, Yilmaz LS, Walhout AJM. Bacterial metabolism affects the *C. elegans* response to cancer chemotherapeutics. *Cell* (2017) 169(3):431–41.e8. doi: 10.1016/j.cell.2017.03.046

73. Le Gall G, Guttula K, Kellingray L, Tett AJ, Ten Hoopen R, Kemsley EK, et al. Metabolite quantification of faecal extracts from colorectal cancer patients and healthy controls. *Oncotarget* (2018) 9(70):33278–89. doi: 10.18632/oncotarget.26022

74. Marchesi JR, Adams DH, Fava F, Hermes GD, Hirschfield GM, Hold G, et al. The gut microbiota and host health: a new clinical frontier. *Gut* (2016) 65(2):330–9. doi: 10.1136/gutjnl-2015-309990

75. Raetz CR, Whitfield C. Lipopolysaccharide endotoxins. Annu Rev Biochem (2002) 71:635–700. doi: 10.1146/annurev.biochem.71.110601.135414

76. Yoshimoto S, Loo TM, Atarashi K, Kanda H, Sato S, Oyadomari S, et al. Obesityinduced gut microbial metabolite promotes liver cancer through senescence secretome. *Nature* (2013) 499(7456):97–101. doi: 10.1038/nature12347 77. Goodwin AC, Destefano Shields CE, Wu S, Huso DL, Wu X, Murray-Stewart TR, et al. Polyamine catabolism contributes to enterotoxigenic Bacteroides fragilisinduced colon tumorigenesis. *Proc Natl Acad Sci USA*(2011) 108(37):15354–9. doi: 10.1073/pnas.1010203108

78. Azcárate-Peril MA, Sikes M, Bruno-Bárcena JM. The intestinal microbiota, gastrointestinal environment and colorectal cancer: a putative role for probiotics in prevention of colorectal cancer? *Am J Physiol Gastrointestinal Liver Physiol* (2011) 301 (3):G401–24. doi: 10.1152/ajpgi.00110.2011

79. McMillan DC, Canna K, McArdle CS. Systemic inflammatory response predicts survival following curative resection of colorectal cancer. *Br J Surg* (2003) 90(2):215–9. doi: 10.1002/bjs.4038

80. Eslami M, Yousefi B, Kokhaei P, Hemati M, Nejad ZR, Arabkari V, et al. Importance of probiotics in the prevention and treatment of colorectal cancer. J Cell Physiol (2019) 234(10):17127–43. doi: 10.1002/jcp.28473

81. Ranadheera R, Baines S, Adams M. Importance of food in probiotic efficacy. Food Res Int (2010) 43(1):1-7. doi: 10.1016/j.foodres.2009.09.009

82. Zhong Y, Chen H. Oligofructose: A new approach to bowel cancer prevention and treatment. *Sci Technol Food Industry* (2014) 35(17):28–31.

83. Li Y. Regulation of intestinal flora by oligo-xylulose improves the quality of survival during chemotherapy in colorectal cancer patients. Jiangnan University. (2022).