Check for updates

OPEN ACCESS

EDITED BY Giulio Innamorati, University of Verona, Italy

REVIEWED BY Lucio Ildebrando Maria Cocco, University of Bologna, Italy Suzanne Scarlata, Worcester Polytechnic Institute, United States

*CORRESPONDENCE Zhou-Wei Xu Xzw0318@sina.com

RECEIVED 31 May 2023 ACCEPTED 13 July 2023 PUBLISHED 28 July 2023

CITATION

Wu Y-N, Su X, Wang X-Q, Liu N-N and Xu Z-W (2023) The roles of phospholipase C- β related signals in the proliferation, metastasis and angiogenesis of malignant tumors, and the corresponding protective measures. *Front. Oncol.* 13:1231875. doi: 10.3389/fonc.2023.1231875

COPYRIGHT

© 2023 Wu, Su, Wang, Liu and Xu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The roles of phospholipase $C-\beta$ related signals in the proliferation, metastasis and angiogenesis of malignant tumors, and the corresponding protective measures

Yu-Nuo Wu¹, Xing Su¹, Xue-Qin Wang¹, Na-Na Liu² and Zhou-Wei Xu^{2,3*}

¹Department of Clinical Medical, the First Clinical Medical College of Anhui Medical University, Hefei, Anhui, China, ²Department of Emergency Surgery, The First Affiliated Hospital of Anhui Medical University, Hefei, Anhui, China, ³Institute of Clinical Pharmacology, Anhui Medical University, Key Laboratory of Anti-inflammatory and Immune Medicine, Ministry of Education, Anhui Collaborative Innovation Center of Anti-inflammatory and Immune Medicine, Hefei, Anhui, China

PLC- β is widely distributed in eukaryotic cells and is the key enzyme in phosphatidylinositol signal transduction pathway. The cellular functions regulated by its four subtypes (PLC- β 1, PLC- β 2, PLC- β 3, PLC- β 4) play an important role in maintaining homeostasis of organism. PLC- β and its related signals can promote or inhibit the occurrence and development of cancer by affecting the growth, differentiation and metastasis of cells, while targeted intervention of PLC- β 1-PI3K-AKT, PLC- β 2/CD133, CXCR2-NHERF1-PLC- β 3, G α q-PLC- β 4-PKC-MAPK and so on can provide new strategies for the precise prevention and treatment of malignant tumors. This paper reviews the mechanism of PLC- β in various tumor cells from four aspects: proliferation and differentiation, invasion and metastasis, angiogenesis and protective measures.

KEYWORDS

PLC- β , cancer, proliferation, invasion, angiogenesis, protection

1 Introduction

PLC-β is widely distributed on the cytoplasmic membrane of eukaryotes. It belongs to βisoenzyme of phospholipase C, which is the key enzyme in the signal transduction pathway of phosphatidylinositol. Its regulated cellular function plays an important role in maintaining homeostasis of the organism. It is an effector enzyme of G protein-coupled receptors, GPCRs, with seven transmembrane hydrophobic regions, and molecules such as bradykinin, histamine, angiotensin II, M1 muscarinic-like receptors and α 1 adrenergic receptors activate PLC-β via Goαq of the pertussis toxin (PTX)-sensitive Gq subfamily, while M2 and M4 muscarinic-like receptors activate PLC- β *via* G $\beta\gamma$ of the PTX-sensitive Go/Gi subfamily (1). In the process of recognizing different subunits of G-protein, the C-terminal and N-terminal PH domain of PLC- β play an important role. The action point of PLC- β and G α q extends at the C-terminal of the enzyme to form the tailer sequence, while the action point of PLC- β and $\beta\gamma$ dimer is located at the N-terminal PH domain, which is the key step of exerting PLC- β 's biological effect. The basic signal pathway mediated by PLC- β is: transmembrane receptor \rightarrow G-protein \rightarrow PLC- β \rightarrow DAG and IP3 (2). In cell signal transduction, phosphatidylinositol-4,5-bisphosphate (PIP2) can be hydrolyzed by the activated PLC- β to generate the second messenger diglyceride (DAG) and inositol-1,4,5-triphosphate (IP3). Among them, IP3 diffuses freely and binds to IP3-specific receptors, which leads to the release of intracellular Ca²⁺, while DAG activates protein kinase C (PKC) together with Ca²⁺ (Figure 1) (3).

PLC-β is not only distributed on the cytoplasmic membrane, but also in the nucleus and cytoplasm. PLC-β1 consist of two subtypes, 150-kDa PLC-β1a and 140-kDa PLC-β1b, which are detectable both in cytosolic and nuclear fractions (4, 5). Several studies have revealed that nuclear PLC-β1 signaling played a direct role in G1 progression by means of a specific target, i.e. cyclin D3/cdk4 (4, 6). There's also research suggesting that nuclear PLC-β1 is a positive regulator in process of myoblast differentiation (7). Besides nuclear function, PLC-β1 has a cytosolic population that can drive the differentiation of rat pheochromocytoma cells (PC12 cells) by regulating component 3 of RISC activity (C3PO). Studies suggest that we can control the differentiation of cultured neuronal cells by down-regulating the levels of cytosolic PLC- β 1 or by driving it to the membrane with stimulation of Gaq (8). Additionally, we found that reducing PLC- β 1 levels increases the rate of proliferation in cells of neuronal lineage. In the early stages of differentiation, PLC- β 1 binds and inhibits cyclindependent kinase 16 (CDK16) to promote proliferation. The bifunctional effects of PLC- β 1 on differentiation and proliferation is also seen in the cultured human neuronal line SK-N-SH (9).

There are four subtypes of PLC- β , namely PLC- β 1, PLC- β 2, PLC- β 3 and PLC- β 4. They are widely distributed in human brain, breast, liver, pancreas and vascular smooth muscle tissues, and their regulated cellular functions play an important role in maintaining homeostasis (1, 2). PLC- β is involved in the process of cell proliferation, differentiation and apoptosis, and is closely related to various biological activities of tumors. In this paper, the role of PLC- β in several aspects of tumor proliferation, metastasis, angiogenesis and protective measures will be discussed separately.

2 Proliferation and differentiation

2.1 ERK/MAPK signal

Mitogen activated protein kinases (MAPKs) is a kind of serine/ threonine protein kinases. Its related pathways can be activated by



extracellular stimuli such as growth factors, inflammation, hormones, neurotransmitters, environment and cell stress, and then induce a wide range of intracellular reactions (10). Extracellular signal regulated kinase (ERK) was first discovered and widely studied in MAPK pathway. Mutation or abnormal activation of MAPK/ERK signaling pathway was found in more than half of cancers. ERK mainly includes five subtypes ERK1~ERK5. Among them, ERK1 and ERK2 are the two subtypes that have been deeply studied at present, and ERK1/2 plays a key role in cancer proliferation and metastasis (11, 12).

The expression of PLC- β 1 in hepatocellular carcinoma (HCC) is significantly higher than that in adjacent tissues, and it is closely related to tumor stage. The results show that the high expression of PLC-β1 in cells can stimulate the phosphorylation of ERK1/2 pathway, while ERK inhibitor can inhibit the promotion effect of PLC-B1 on the growth of hepatocellular carcinoma cells. Therefore, PLC-B1 can exert carcinogenic activity by activating ERK signal in HCC cells (13). In addition, insulin-like growth factor 1 (IGF-1) induces the phosphorylation of nuclear PLC-B1 while activating nuclear ERK, and then starts the inositol phosphate cycle, which may be the loop mechanism of regulation between PLC- β 1 and ERK (14, 15). miRNA-205 can also activate liver cancer stem cell subsets and maintain the characteristics of liver cancer stem cells by regulating the upstream target PLC-B1, which is closely related to the recurrence and metastasis of liver cancer (16). The expression of PLC-B1 in HCC tissues is significantly higher than that in paracancer tissues and is closely related to tumor staging. There was a positive correlation between PLC-B1 expression and advanced tumor stage, that is, PLC-β1 expression was significantly correlated with tumor T stage, and had no significant correlation with other clinical features. This suggests that high PLC-B1 expression contributes to the progression of HCC. At the same time, patients with positive tumor PLC-B1 expression had a lower OS rate than the patients with negative PLC-B1 expression. According to the above, the expression of PLC-β1 in HCC is an important prognostic factor for survival and T stage (17).

Melanoma is one of the highly malignant epidermal cancers. It was found that PLC-B2 was highly expressed in melanoma tissues, and its level was significantly up-regulated in human melanoma cell lines. This can affect the related biological functions of melanoma, while interfering with PLC-B2 can significantly inhibit cell viability and promote cell apoptosis (18). Ras-Raf-ERK pathway can promote cell growth, division and differentiation, and participate in cell cycle regulation, wound healing, tissue repair, cell migration and integrin signal transduction. Ras mutations are the oncogenic form in more than 15% of cancers, and the B-Raf gene (which is a member of the Raf family) is mutated in 66% of malignant melanomas. Ras can activate Raf/MEK signal in turn to up-regulate the phosphorylation level of ERK1/2, and induce the development of cancer cells (19). The mutual regulation of Ras and antioncogene p53 can affect the process of cell proliferation, apoptosis, movement, inflammatory reaction and angiogenesis, while the overexpression of PLC-B2 can significantly reduce the mRNA and protein level of p53. MAPK signal can regulate the expression of Bax, Bcl-2 and caspase-3 genes, among which Bax protein promotes apoptosis while Bcl-2 protein inhibits apoptosis, and caspase-3 is the key executor of apoptosis. It can significantly enhance the expression of caspase-3 and Bax genes and reduce the activity of Bcl-2 by interfering with PLC- β 2. Therefore, downregulation of PLC- β 2 can inhibit the activation of Ras/Raf/MAPK pathway and promote the apoptosis of melanoma cells, so it is considered that PLC- β 2 may be a potential target gene for controlling the proliferation of melanoma cells (Figure 2) (18).

2.2 CSF1-P2RY6 and Stat5/SHP-1 pathways

PLC-β3 is related to a specific subgroup of leukemia, namely chronic myelomonocytic leukemia (CMML). It is found that PLC-β3 (-/-) mice can cause CMML-like diseases (20), which are characterized by immature and stunted granulocytes. Generally, after colony stimulating factor 1 (CSF1) binds to the receptor in hematopoietic cells, CSF1 receptor (CSF1R) triggers the activation of P2Y6 receptor (P2RY6, a GPCR in P2 receptor family), which in turn activates PLC-β3 and induces autophagy and monocyte differentiation. However, in CMML cells, the synthesis and secretion of a large number of defensin α 1 (DEFA1) and defensin α 3 (DEFA3) can antagonize P2RY6, thus inhibiting the monocyte differentiation induced by CSF1 and PLC-β3. Therefore, PLC-β3 knockout in hematopoietic cells can effectively repress CSF1-mediated monocyte differentiation through calcium signaling pathway (21).

In addition, the over-activation of signal transducer and activator of transcription 5 (Stat5) can also lead to CMML. Src homology region 2 domain-containing phosphatase 1 (SHP-1) is a non-receptor protein tyrosine phosphatase mainly expressed in hematopoietic cells and epithelial cells. It can dephosphorylate Stat5 to inhibit the activity of Stat5, and the existence of PLC- β 3 can inhibit the malignant reaction related to Stat5 by enhancing the function of SHP-1. The same transformation mechanism has been found in other human lymphoid and myeloid malignancies (20, 22).

2.3 Wnt/Ca²⁺ signal

Cutaneous squamous cell carcinoma (CSCC) is a malignant tumor originating from epidermal keratinocytes or adnexal cells, which has a high incidence. Once CSCC metastasizes, it is difficult to treat and its prognosis is poor. Wnt5a (a member of Wnt protein family) can participate in the non-classical Wnt pathway through Wnt/Ca²⁺ pathway. It was found that the expression of PLC- β 4 gene in this pathway decreased in cancer, but there was no significant difference in Wnt5a mRNA level. WNT5a may activate PLC- β 4 by binding frizzled 4 (FZD4), and initiate the non-classical Wnt pathway to play an antioncogene-like role in CSCC. Therefore, PLC- β 4 can inhibit the proliferation and invasion of CSCC cells, and its high expression can also repress the growth of tumor cells and CSCC formation in nude mice (23).

2.4 WNK1-PI4KIIIa and TRPC6-NFATc1 pathways

The imbalance of Ca²⁺ signal is one of the important characteristics of cancer progression. Lysine-deficient protein



kinase 1 (WNK1) is the main regulator of renal ion transport, and regulates Ca²⁺ signal by stimulating phosphatidylinositol 4-kinase IIIa (PI4KIIIa) to activate Gαq coupled receptor and PLC-β signal pathway. The expression of WNK1 is directly related to the nuclear grade of clear cell carcinoma of kidney (ccRCC). Studies have shown that tumor tissues with higher Fuhrman nuclear grade express a high level of WNK1, while WNK1 protein is rarely detected in lower grade tumors. This indicates that the overexpression of WNK1 may be closely related to the pathological development of ccRCC. Functional experiments showed that WNK1 combined with PI4KIIIa activated Ca²⁺ influx mediated by transient receptor potential channel 6 (TRPC6), and Gaq coupled receptor and PLC-β signal played an important role in it (24, 25). Both the functional acquired mutation and overexpression of TRPC6 can lead to kidney diseases, such as focal and segmental glomerulosclerosis, fibrosis and renal cell carcinoma (26-29). The nuclear factor of activated T-cells cytoplasmic 1 (NFATc1) can be repressed by inhibiting the activation of TRPC6 mediated by WNK1, thus reducing the proliferation and migration activity of ccRCC cells. Therefore, TRPC6-NFATc1 pathway mediated by WNK1-PI4KIIIa through PLC- β signal may play a key role in the tumor growth of ccRCC (30), and this mechanism can provide a potential new target for the treatment of renal cell carcinoma in the future.

2.5 LPA

Lysophosphatidic acid (LPA) regulates PLC- β 1 and PLC- β 2 in different ways to enhance the proliferation and migration of intestinal epithelial cell (IEC), thus promoting wound healing and

the recovery of intestinal epithelial barrier. The interaction between Gaq and PLC- β 2 induced by LPA binding to LPA receptor 1 accelerated the migration of IEC cells. While LPA's activity of promoting IEC cell proliferation is PLC- β 1-dependent, involving the translocation of Gaq to the nucleus, where it interacts with PLC- β 1 to change the process of cell cycle (31). In addition, LPA, through its homologous receptors (LPAR1-LPAR6), can produce a variety of cellular responses with the cooperation of PLC- β signals (32, 33), for example, LPAR2 stimulates the proliferation and migration of colon cancer cells, while the deletion of LPAR2 can inhibit the progress of colon cancer (34, 35).

2.6 CXC chemokine/CXCR2 and CXCL5 signals

CXC chemokine and its homologous receptor CXC chemokine receptor 2 (CXCR2) play a key role in tumor growth and angiogenesis. It has been proved that blocking the biological axis of CXC chemokine/CXCR2 can reduce the risk of malignant tumors (36–39). CXCR2 is expressed in many pancreatic ductal adenocarcinoma (PDAC) cell lines (40–43), and it is mainly involved in enhancing the proliferation and viability of cancer cells through autocrine or paracrine action (40, 43, 44). CXCR2, PLC-β3 and Na⁺/H⁺ exchanger regulatory factor-1 (NHERF1) form macromolecular complexes on the plasma membrane of pancreatic cancer cells, which functionally couples the signal cascade mediated by CXC chemokine and PLC-β3 (45). The formation of CXCR2-NHERF1-PLC-β3 complex is mediated by PDZ domain in NHERF1. Interruption of PDZ-mediated interaction can eliminate CXCR2 and PLC-β3 signals, thus inhibiting the

10.3389/fonc.2023.1231875

proliferation of PANC-1 cells (human pancreatic cancer cells). This result has also been found in animal models of PDAC xenotransplantation. In addition, it was found that the destruction of CXCR2-NHERF1-PLC- β 3 complex can significantly inhibit the progression of pancreatic malignant tumors *in vitro* and *in vivo* (45). Therefore, CXC chemokine/CXCR2 signaling is an important mechanism for the occurrence and development of many malignant tumors including pancreatic cancer (46–50).

Pancreatic intraepithelial neoplasia (PanIN) is considered to be the precursor of pancreatic cancer. Pathologically, PanIN lesions are classified as dysregulated ductal epithelium progressing from PanIN-1 to PanIN-3 (51, 52). Both PanIN-2 and PanIN-3 are high-grade lesions that represent the initial steps toward invasive cancer. The expression of CXCL5 in PanIN-2 and PanIN-3 was significantly higher than that of PanIN-1, indicating that CXCL5 gradually increased in the progression of precancerous PanIN lesions to invasive cancer. Patients with pancreatic cancer and high CXCL5 expression who also underwent cancerectomy had significantly lower survival rates than those without overexpression. These results indicated that CXCL5 expression was negatively correlated with tumor differentiation, clinical stage and survival rate (44).

2.7 Gαq-PIP2-PKC pathway

Gαq-PIP2-PKC is the core signal pathway of uveal melanoma (UM), in which PLC-β plays a key role (53). After PLC-β is activated by Gαq, phosphatidylinositol 4,5- bisphosphate (PIP2) is hydrolyzed into diacylglycerol (DAG) and inositol 1,4,5triphosphate (IP3) (54). DAG binds to and activates a variety of proteins including protein kinase C (PKC) and Ras guanyl nucleotide releasing proteins (RasGRPs) through its C1 domain (55). IP3 can increase intracellular Ca²⁺ level and activate Ca²⁺related signal pathways such as PKC signal. In UM microenvironment, PKC and Ras protein activator RasGRPs together activate MAPK signal, which is an important mechanism of inducing tumor cell growth. At present, it has been found that more than 90% UMs have mutations in Gαq structure, and PLC-β4 activation is the central node of carcinogenic signaling in Gαq mutated UMs (53).

3 Invasion and metastasis

3.1 Ang II/AT1R/CaM signal

Angiotensin II (Ang II), angiotensin II type 1 receptor (AT1R) and calmodulin (CaM) are important bioactive molecules that affect the formation and prognosis of HCC. Ang II can generate and upregulate the expression of AT1R in liver cancer tissues. After AT1R is coupled with PLC- β 1, it activates and promotes the change of CaM expression and conformation, thus regulating intracellular Ca²⁺ concentration and calcium-regulated kinase activity, resulting in a series of biological effects. Furthermore, PLC- β 1 siRNA was used to

transfect hepatocellular carcinoma cell lines, and it was found that the role of the above signal pathway was obviously weakened during the metastasis of HCC cells with low expression of PLC- β 1. Similar experimental results have also been confirmed in the mouse model of hepatocellular carcinoma, which indicates that PLC- β 1-related Ang II/AT1R/CaM signal is an important mechanism to promote the migration and invasion of HCC cells (3).

3.2 CD133, EMT markers and miR-146a

PLC-β2 is expressed in most breast tumors, and its level can affect the prognosis of patients (56). The role of PLC-β2 in different invasive breast tumors is related to molecules such as CD133 (glycosylated transmembrane protein), EMT markers and miR-146a (56–61). The expression of CD133 in breast cancer is significantly related to tumor stage, size, lymph node metastasis and sensitivity to neoadjuvant chemotherapy (60). EMT is the process of epithelial cells transforming into mesenchymal cells under certain physiological and pathological conditions, which can enhance the migration and invasion activity of cancer cells (62). MiR-146a is an antioncogene, which can inhibit the metastasis of tumor cells.

Triple-negative breast cancer (TNBC), which accounts for about 20% of all breast tumors, is a highly malignant tumor with a particularly poor prognosis. At present, there is no effective targeted therapy. In TNBC tumor-derived cells, the level of PLC- β 2 is positively correlated with the motility of tumor cells (61). It was found that TNBC cells with high CD133 expression have stronger metastatic potential, while PLC- β 2 level in highly invasive breast tumor-derived cells is negatively correlated with CD133 expression. That is to say, although PLC- β 2 can maintain the movement of breast tumor cells, it may also down-regulate the expression of CD133 and reduce the invasiveness of cells. Therefore, PLC- β 2 played a two-way regulating role in the transfer of TNBC (57, 61).

PLC-β2 in low-invasive breast tumor-derived cells can inhibit tumor development, but this protective mechanism is usually affected by hypoxia (57). Under hypoxia, the down-regulation of PLC-β2 is mainly related to the decrease of EMT marker E-cadherin or the up-regulation of CD133. However, the high expression of PLC-β2 in low-invasive tumor cells can also prevent the malignant progression related to hypoxia (59), that is, PLC-β2 can increase the level of E-cadherin in cells, block the up-regulation of CD133, and induce the adhesion of tumor cells to reduce their metastasis and invasion activity, thus preventing the progression of breast malignant tumors (Figure 3) (57).

The cancer cells of ductal carcinoma *in situ* (DCIS) are confined in the mammary duct and have not spread to the surrounding normal breast tissues. If they invade the surrounding matrix, they will develop into invasive ductal carcinoma (IDC). Up-regulation of PLC- β 2 expression in DCIS can counteract the increased metastatic potential of DCIS cells caused by hypoxia. Therefore, the expression level of PLC- β 2 may affect the malignant potential of DCIS, that is, the low expression of PLC- β 2 makes DCIS-derived cells easy to acquire invasion characteristics. MiR-146a is a tumor suppressor miRNA in DCIS and IDC, which can reduce the migration and invasion activity of cancer cells. The expression of PLC- β 2 in DCIS-derived cells is closely related to the imbalance of miR-146a. Considering that the down-regulation of miR-146a may be the basis for the ectopic appearance of PLC- β 2 in tumor cells (57).

Upregulated expression levels of PLC- $\beta 2$ were detected in different types of breast tumors. In addition, breast cancer cell lines with high invasive potential showed higher PLC- $\beta 2$ expression levels compared to less invasive breast cancer cell lines. The expression of PLC- $\beta 2$ was closely related to tumor grade, and the staining intensity increased from grade 1 to grade 3, among which grade 3 tumors showed the highest expression level of PLC- $\beta 1$, and the survival rate was lower than that of other grades. This suggests that PLC- β levels are strongly correlated with poor prognosis of breast cancer (56, 63).

3.3 PI(4,5)P2/Cofilin

The migration of tumor cells is the key step to complete the metastasis, and phosphatidylinositol-4,5-diphosphate (PI(4,5)P2) has been proved to be an important regulator of tumor cell migration, which affects the movement ability of cells by regulating actin (64). The expression of PLC- β 1 is up-regulated in highly invasive breast cancer cells, and it is related to metastasis and recurrence of breast cancer patients. It was found that PLC- β 1 could hydrolyze PI(4,5)P2 to produce IP3 and DAG, which targeted to reduce the level of PI(4,5) P2 in plasma membrane, which made PI(4,5)P2 binding protein cofilin released into cytoplasm from its inactive membrane association state, thereby increasing actin in breast cancer cells to promote cell migration activity. This suggests that the PI(4,5)P2/cofilin pathway is an important mode of PLC- β 1-induced cell metastasis (64).

3.4 PABPC1-PI3K-AKT pathway

Cholangiocarcinoma (CAA) is an epithelial malignant tumor originating from bile duct. High levels of PLC- β 1 were expressed in human CCA tissues and cell lines. PLC- β 1 can promote the proliferation activity of CCA cells by enhancing G1/S transition of cell cycle, and affect the movement and invasion ability of CCA cells. PLC- β 1 can regulate the phosphorylation of ERK and serine/ threonine kinase (AKT), and then activate phosphoinositide-3 kinase (PI3K)/AKT signal to induce CCA cells to undergo EMT. In addition, polyadenylation binding protein 1 (PABPC1) interacts with PLC- β 1 can further enhance the EMT process mediated by PI3K-AKT pathway, which is an important mechanism of early metastasis of tumor cells. While PLC- β 1 is the direct target of miR-26b-5p (antioncogene), which can act on PLC- β 1 and prevent CCA metastasis (65).

3.5 WNK1-TRPC6-NFATc1 pathway

WNK1 in ccRCC cells regulates Ca^{2+} signal by activating G α qcoupled receptor/PLC- β pathway, further mediates the activation of transient receptor potential cation channel 6 (TRPC6) and upregulates the level of NFATc1. This is related to the metastatic potential of ccRCC cells. It was found that inhibition of TRPC6/ NFATc1 signaling by cyclosporin A (CsA) or SKF96365 (calcium channel blocker) could reduce the migration activity of ccRCC cells. In addition, the invasiveness of ccRCC cells is inhibited by siRNA of WNK1 or TRPC6, while the transient overexpression of WNK1 or TRPC6 promotes cell invasion. Therefore, PLC- β related WNK1-TRPC6-NFATc1 pathway is the key mechanism to promote ccRCC transfer (30).



3.6 PI3K, mTOR and MAPK signals

Human colorectal tumors can express C-kit protooncogene, which has an important role in biological behaviors such as cell survival, proliferation, adhesion and chemotaxis (66). The pleiotropic function of C-kit is mainly mediated by molecular cascade of PI3K, PLC and MAPK (67–70). Existing studies have confirmed that PLC- β , mammalian target of rapamycin (mTOR) and MAPK-related signaling pathways are potential therapeutic targets for metastatic tumors (71, 72). In kidney and colon cancer cells, tumor invasion mediated by trefoil peptide family, thromboxane A2 and PAR-1 (thrombin receptor) is PLC- β or mTOR dependent (73–75). Therefore, PI3K, PLC- β , mTOR and MAPK signals affect the metastatic potential of cancer cells through their correlation with each other (76).

3.7 LPA/AMPK/RhoA pathway

AMP-activated protein kinase (AMPK) is an energy-sensing kinase, which can regulate the motor activity of cells. LPA induced the activation of RAS homologous gene family member A (RhoA) through AMPK, and further activated ezrin-radixin-moesin (ERM) protein to restructure the cytoskeleton, which played an important role in accelerating the migration of ovarian cancer cells. The biological function of AMPK is regulated by Ca^{2+} signal related to PLC- β 3, and the ability of AMPK to promote cell migration in PLC- β 3 knockout cells decreases obviously. In addition, the application of PLC inhibitor (U73122) or specific CaMKK inhibitor (STO-609) can inhibit LPA-activated phosphorylated AMPK (77). Therefore, LPA/AMPK/RhoA signaling pathway is Ca²⁺-dependent in promoting tumor metastasis.

3.8 PLC- β 1a, PLC- β 1b and ERBB4 signals

PLC-β1 is abundantly expressed in many parts of the brain, including cerebral cortex, hippocampus, amygdala, lateral septum, olfactory bulb and other regions. The signal transduction pathway mediated by PLC-B1 is related to brain development and various neurological diseases (2, 78). PLC-B1 also exists in glioma cells (4, 79), which is divided into PLC-B1a and PLC-B1b. PLC-B1a is mainly located in the cytoplasm, while PLC-B1b is mainly located in the nucleus of the cells. There is evidence that when stimulated, PLC- β 1 in glioma cells will translocate into the nucleus (80). Glioma is the most common primary brain tumor, and pathological grade is the most important factor in determining the prognosis of patients. Glioma grade I was defined as the lowest invasive glioma, and grade IV was defined as the most invasive glioma type. Grades I/II and III/IV are also known as low-grade (LGG) and high-grade gliomas (HGG). Grade IV glioma, also known as glioblastoma multiforme (GBM), is the most common and malignant primary brain tumor type, accounting for 50-60% of all gliomas (81). Due to the complexity and heterogeneity of the tumor, this tumor will develop resistance to treatment and relapse quickly, and PLC-B1 is a potential prognostic factor. The expression of PLC-B1 is negatively correlated with the pathological grade of glioma, and it is a new characteristic gene in the molecular classification of high-grade glioma. Studies have confirmed that PLC-B1 gene expression is significantly reduced in all type IV gliomas (glioblastoma) compared to grade II and grade III gliomas. The data showed that the survival time of glioma patients with medium level of PLC-B1 expression was significantly longer than that of PLC-B1 down-regulated group. Compared with low-grade gliomas and healthy patients, the expression of PLC-B1 in glioblastoma samples was decreased. The experiment also confirmed that down-regulation of PLC-B1 expression in cells leads to an increase in cell migration and invasion, indicating the potential role of PLC-β1 in maintaining a normal or less invasive glioma phenotype. In addition, there is also a correlation between the expression of PLC- β 1 and glioma PN signature gene ERBB4. ERBB4 protein is a tyrosine protein kinase and a member of the epidermal growth factor receptor subfamily, which will promote the pathogenesis of glioma. In the central nervous system, typical ERBB4 signaling was associated with downstream PLC and PI3K-AKT activation. In the developing brain, cleaved ERBB4 protein plays an cricial role in regulating the timing of astrogenesis. Moreover, experiments showed that PLC-B1 signal intensity was consistent with ERBB4 in low-grade and high-grade glioma tumors (78, 81).

4 Angiogenesis

4.1 VEGF and GPCR signals

Endothelial cells are the key components of neovascularization. Therefore, the proliferation and migration ability of endothelial cells is very important for angiogenesis. The proliferation and chemotaxis of endothelial cells are driven by vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF). VEGF exerts its biological activity mainly by coupling VEGF receptor 1 (VEGFR1) and VEGF receptor 2 (VEGFR2). VEGF, especially VEGFR2, can induce phosphorylation of two serine residues on PLC- β 3, and then enhance intracellular Ca²⁺ signal (82). It is found that cell division cycle protein 42 (CDC42) may be the downstream target molecule of VEGFR2-PLC-β3 axis to promote endothelial cell migration. In PLC-B3 knock-down cells, the expression and activity of CDC42 decreased obviously, and it could inhibit the production of endothelial cells stimulated by VEGF, resulting in abnormal development and delayed growth of tumor blood vessels (82, 83). These results suggest that PLC- β 3 can be used as a therapeutic target to inhibit tumor angiogenesis.

PLC-β3 can also be activated by GPCRs, and participates in the regulation of angiogenesis of various malignant tumors. Endothelial differentiation G-protein coupled receptor 1 (EDG1) is a functional sphingosine-1-phosphate (S1P) receptor, which plays a vital role in the formation of vascular endothelial cells and the activation of PLC. It was found that EDG1 can simultaneously regulate several downstream signal pathways through Gi/o protein, including adenylate cyclase (AC) inhibition, Ca²⁺ mobilization, Ras-MAPK and PLC-β3 activation (84, 85). However, the precise molecular mechanism of PLC-β3 regulating tumor angiogenesis still needs further research and exploration (86, 87).

4.2 IGF2 signal

The entry of endothelial progenitor cells (EPCs) into the neovascular region is considered to be an important step in the formation of vascular network during embryonic development. In the pathological state of tumor, the effect of EPCs on neovascularization has been generally accepted (88, 89). Insulin-like growth factor 2 (IGF2) can promote tumor angiogenesis and lymphangiogenesis by increasing EPCs recruitment (90-92). In addition, IGF2 induced by ischemia and hypoxia participates in angiogenesis of human hepatocellular carcinoma (93), and significantly increases VEGF mRNA and protein levels in tumor tissues in a time-dependent manner (90). The biological function of IGF2 is mainly realized by signal transduction of G(i) protein coupled with IGF-2 receptor (IGF2R), and intracellular Ca2+ mobilization induced by PLC-B2 needs to be involved (Figure 4) (90). Therefore, controlling the IGF-2/IGF2R signal associated with PLC-B2 can be used as a new method to treat angiogenesis-dependent tumors.

5 Protective measures

5.1 PLC- β 1-related PI3K-AKT signal inhibition and PKC α signal activation

PLC- β 1 can promote the proliferation and motility of CCA cells, and the CCA patients with high expression of PLC- β 1 do not perform well in TNM staging, distant metastasis and survival

prognosis. PLC- β 1 also induced the resistance of CCA to gemcitabine combined with cisplatin, but this could be reversed by AKT inhibitor MK2206. It was found that PLC- β 1-PI3K-AKT signal axis was very important for CCA development and EMT, and AKT can be used as a therapeutic target to overcome the chemotherapy resistance of CCA patients with high PLC- β 1 expression, thus inhibiting the growth of tumor and obviously improving the postoperative survival of patients (65).

The pathogenesis of myelodysplastic syndrome (MDS) and erythropoiesis involve PLC-β1/protein kinase Cα (PKCα) signal transduction, especially nuclear PLC-B1 gene (21). PLC-B1 gene is located on the short arm of chromosome 20, and two splicing variants can be identified: PLC-B1a and PLC-B1b. Both subtypes have a nuclear localization sequence, but PLC-B1a also has a nuclear export sequence (NES). Therefore, PLC-β1a also exists in the cytoplasm, while PLC-β1b mainly exists in the nucleus (94).PLC-B1a is a negative regulator of erythroid differentiation, and which is reduced in erythropoietinresponder MDS patients and in normal hematopoietic stem cell progenitors induced to erythroid differentiation (95). Nuclear PLC-B1 specifically targets PKCo, which is related to the proliferation and differentiation of human erythroleukemia cells and primitive human erythroid cells. MDS patients with low evolutionary risk of acute myeloid leukemia (AML) and 5q chromosome deletion can be treated with lenalidomide. Lenalidomide is an immunomodulatory drug that can induce erythroid differentiation. It specifically increases the expression of PLC-β1 in cytoplasm, but it can also induce PKCα translocation to nucleus, thereby stimulating erythropoiesis (21). However, MDS patients with high risk of AML evolution usually need epigenetic therapy, which



aims to inhibit the proliferation of hematopoietic stem cells. The nuclear PLC-B1 of hematopoietic cells is related to epigenetics of MDS. MDS patients can show a specific single allele deletion of the PLC-B1 gene, which can lead to a higher probability of evolution into AML, and also lead to a decrease in nuclear PLC-B1 expression levels in MDS patients. PLC-B1 is a specific molecular target of azacytidine (AZA). AZA is a demethylating drug that can accelerate tumor cell division and death by producing cytotoxicity. The cells of MDS patients showed a high level of recruitment of specific myeloid transcription factors (MZF-1) associated with the PLC-B1 promoter during azacitidine treatment. In this case, both high-risk and low-risk MDS patients who responded positively to treatment showed an early increase in the expression of nuclear PLC-B1 and PKCo, accompanied by a decrease in the specificity of PLC-B1 promoter methylation and induction of normal myeloid differentiation, leading to the improvement of clinical symptoms and the differentiation of normal bone marrow (Figure 5) (21).

The expression level of phosphoinositide-specific phospholipase $C\beta1(PI-PLC\beta1)$ in MDS was lower than that in normal tissue. The expression level of PI-PLC $\beta1$ in high-risk group was lower than that in low-risk group in different types of MDS. The results indicated that the down-regulated expression of PI-PLC $\beta1$ reflected the progression of MDS disease and could be used as a prognostic indicator of MDS (96).

5.2 PLC- β 2/CD133 and PLC- β 2/miR-146a signals

Tumor stem cell marker CD133 can predict the sensitivity of breast cancer to neoadjuvant chemotherapy drugs (61), and its level is closely related to the invasiveness of TNBC. Therefore, targeting this surface antigen may be beneficial to the treatment of TNBC (60). The regulation mechanism of PLC-β2 and CD133 in TNBC has been described above, which proves that PLC-B2 can downregulate the expression of CD133 and reduce the metastatic potential of cells. Therefore, the intervention of PLC-B2/CD133 signal can be used as a new therapy to prevent the progression of invasive breast tumors (60). It is found that the negative correlation between PLC-B2 and miR-146a in primary DCIS cannot be detected in IDC, which indicates that the change of PLC-B2/miR-146a expression level in DCIS may constitute a molecular risk factor for IDC. Generally speaking, the down-regulation of miR-146a in DCIS without the increase of PLC-B2 will lead to the risk of malignant progression of DCIS. Evaluating the change of its level is helpful to identify whether patients have relapse tendency. Therefore, this signal pathway may become an important target for the prevention and treatment of DCIS in the future (57).



5.3 ORP4L/G α q/11/PLC- β 3 complex and CXCR2-NHERF1-PLC- β 3 complex

AML is characterized by the rapid growth of abnormal cells accumulated in bone marrow and blood. Leukemia patients often contain leukemia stem cells (LSCs), which can produce leukemia cells and are the main cause of AML recurrence. ORP4L belongs to the oxysterol binding protein-related protein family (ORPs). It is selectively expressed in LSCs and is essential for the survival of LSCs. It was found that ORP4L transported PLC- β 3 out of the nucleus of leukemia cells, and regulated PIP2 metabolism and downstream Ca²⁺ balance by forming ORP4L/G α q/11/PLC- β 3 complex, further regulating pyruvate dehydrogenase activity on mitochondria to provide oxidative phosphorylation energy for leukemia cells. This signal mechanism provided a potential intervention target for the treatment of leukemia such as AML (97).

NHERF1-mediated functional coupling of CXCR2 and PLC- β 3 will form a macromolecular complex, which is essential for CXCR2 signal transduction and malignant progression of pancreatic cancer. Studies have confirmed that the decomposition of CXCR2-NHERF1-PLC- β 3 macromolecules *in vitro* and *in vivo* can inhibit the growth and metastasis of pancreatic tumors. Therefore, destroying this CXCR2 complex may become an effective treatment strategy for pancreatic cancer (45).

5.4 G α q-PLC- β 4-PKC-MAPK and Wnt5a-PLC- β 4-Ca²⁺ pathways

Gαq-PLC-β4-PKC-MAPK pathway is the core signal that affects the occurrence and development of UM (53). IP1 is a stable metabolite of the second messenger IP3, which is produced when PLC-β is activated by Gαq (98). The output intensity of PLCβ signal can be detected by the content of IP1 in cells. YM-254890 (Gαq/11 protein inhibitor) can selectively inhibit IP1 production in a dose-dependent manner, and then block PLC-β4 and downstream PKC/MAPK pathway, which inhibits the proliferation of UM cells (53). Therefore, it is possible to improve the prognosis of UM patients by selecting molecules in Gαq-PLC-β4-PKC-MAPK signaling pathway for targeted therapy.

The non-classical Wnt pathway composed of Wnt5a-PLC- β 4-Ca²⁺ can reduce the proliferation and migration activity of CSCC cells, and inhibit the tumorigenicity of nude mice. Therefore, initiating the non-classical Wnt pathway is considered to play an antioncogene-like role in CSCC (23). Wnt5a can activate PLC- β 4 by binding FZD4, which is the key to the activation of this signal pathway, and it also provides a new exploration direction for CSCC therapy.

6 Conclusion

In recent years, the regulation mechanism of PLC- β related signal pathways and its role in malignant tumors have been widely studied. They believe that PLC- β is a promising new target for the treatment of malignant tumors, and may be an important enzyme related to the diagnosis and prognosis of malignant tumors. In this paper, the regulation modes of PLC- β and its upstream and downstream signals are summarized from the aspects of proliferation and differentiation, metastasis and invasion, angiogenesis and protective measures of malignant tumors. It is found that PLC- β plays a dual regulatory role in promoting and inhibiting the development of tumors through corresponding subtypes associated with ERK/MAPK, Wnt/Ca²⁺, G α q-PIP2-PKC, Ang II/AT1R/CaM and so on. Therefore, on the one hand, we optimistically predict that targeted intervention based on precision can open up a new clinical application direction for tumor treatment. However, on the other hand, we must realize that the safety and reliability of this treatment still depend on the accumulation of knowledge and the progress of technology.

Author contributions

Manuscript writing: Y-NW. Literature collection: Y-NW, XS and X-QW. Figure design and painting: N-NL and Z-WX. Revision: Z-WX. All authors contributed to the article and approved the submitted version.

Funding

This work has been supported by the Key Project of Natural Science Research of Universities in Anhui Province (2022AH051190), the Open Fund of Key Laboratory of Antiinflammatory and Immune Medicine, Ministry of Education, P.R. China (Anhui Medical University) (KFJJ-2020-02) and the School Fund of Anhui Medical University (2020xkj177).

Acknowledgments

The authors are thankful to all staff members of Department of Emergency Surgery, The First Affiliated Hospital of Anhui Medical University, for their help in instructing the work.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher. 1. Liu LR, Xia SH. Progress in the structure of phospholipase C and mechanisms of its activation. Acta Academiae Medicinae CPAPF (2006) 02):175-8.

2. Bai J, Luo SQ, Ji QS. Phospholipase C- β research progress. Chem Life (2000) 02):53–5.

3. Xu ZW, Liu NN, Wang XY, Ding BC, Zhang HF, Li Y, et al. Effect of PLC-β1/ CaM signaling pathway mediated by AT1R on the occurrence and development of hepatocellular carcinoma. *Cancer Cell Int* (2021) 21(1):1–21. doi: 10.1186/s12935-021-02261-8

4. Faenza I, Matteucci A, Manzoli L, Billi AM, Aluigi M, Peruzzi D, et al. A role for nuclear phospholipase Cbeta 1 in cell cycle control. *J Biol Chem* (2000) 275(39):30520–4. doi: 10.1074/jbc.M004630200

5. Bahk YY, Lee YH, Lee TG, Seo J, Ryu SH, Suh PG. Two forms of phospholipase C-beta 1 generated by alternative splicing. *J Biol Chem* (1994) 269(11):8240–5. doi: 10.1016/S0021-9258(17)37185-5

6. Bavelloni A, Faenza I, Cioffi G, Piazzi M, Parisi D, Matic I, et al. Proteomic-based analysis of nuclear signaling: PLCbeta1 affects the expression of the splicing factor SRp20 in Friend erythroleukemia cells. *Proteomics* (2006) 6(21):5725–34. doi: 10.1002/pmic.200600318

7. Faenza I, Bavelloni A, Fiume R, Lattanzi G, Maraldi NM, Gilmour RS, et al. Upregulation of nuclear PLCbeta1 in myogenic differentiation. *J Cell Physiol* (2003) 195 (3):446–52. doi: 10.1002/jcp.10264

8. Garwain O, Pearce KM, Jackson L, Carley S, Rosati B, Scarlata S. Stimulation of the Gaq/phospholipase C β 1 signaling pathway returns differentiated cells to a stem-like state. FASEB J (2020) 34(9):12663–76. doi: 10.1096/fj.201902668R

9. Garwain O, Valla K, Scarlata S. Phospholipase C β 1 regulates proliferation of neuronal cells. FASEB J (2018) 32(5):2891–8. doi: 10.1096/fj.201701284R

10. Cheng TT, Wang C, Pan XH. Research progress of MAPKs signaling pathway in cancer. J Binzhou Med University (2021) 44(04):302–5.

11. Liu HB, Qing L. Research progress on the relationship between MAPK signal pathway system and gastric cancer. *Chin J Curr Adv Gen Surg* (2019) 22(12):995–997 +1002.

12. Zhang DT, Jiang SL. Mechanisms of drug resistance mediated by MAPK/ERK signaling pathway in lung cancer. *J Chin Oncol* (2022) 28(03):233–7.

13. Chu Y, Zhou S, Wang LZ, An TZ, Jiang TP, Song J, et al. The correlation of upregulated expression of PLC β 1 with hepatocellular carcinoma cell proliferationand prognosi. *J Interventional Radiology* (2018) 27(01):29–34.

14. Xu A, Suh PG, Marmy-Conus N, Pearson RB, Seok OY, Cocco L, et al. Phosphorylation of nuclear phospholipase C beta1 by extracellular signal-regulated kinase mediates the mitogenic action of insulin-like growth factor I. *Mol Cell Biol* (2001) 21(9):2981–90. doi: 10.1128/MCB.21.9.2981-2990.2001

15. Zhang HF, Gao X, Wang X, Chen X, Huang Y, Wang LB, et al. The mechanisms of renin-angiotensin system in hepatocellular carcinoma: From the perspective of liver fibrosis, HCC cell proliferation, metastasis and angiogenesis, and corresponding protection measures. *BioMed Pharmacother* (2021) 141:111868. doi: 10.1016/j.biopha.2021.111868

16. Zhao J, Xu G, Qian YW, Li YW. Down-regulation of miR-205 promotes stemness of hepatocellular carcinoma cells by targeting PLCβ1 and increasing CD24 expression. *Neoplasma* (2015) 62(4):567–73. doi: 10.4149/neo_2015_068

17. Li J, Zhao X, Wang D, He W, Zhang S, Cao W, et al. Up-regulated expression of phospholipase C, β 1 is associated with tumor cell proliferation and poor prognosis in hepatocellular carcinoma. *Onco Targets Ther* (2016) 9:1697–706.

18. Zhang H, Xie T, Shui Y, Qi Y. Knockdown of PLCB2 expression reduces melanoma cell viability and promotes melanoma cell apoptosis by altering Ras/Raf/ MAPK signals. *Mol Med Rep* (2020) 21:420-8.

19. Molina JR, Adjei AA. The ras/raf/MAPK pathway. J Thorac Oncol (2006) 1 (1):7–9. doi: 10.1016/S1556-0864(15)31506-9

20. Xiao W, Hong H, Kawakami Y, Kato Y, Wu D, Yasudo H, et al. Tumor suppression by phospholipase C- β 3 via SHP-1-mediated dephosphorylation of Stat5. Cancer Cell (2009) 16(2):161–71. doi: 10.1016/j.ccr.2009.05.018

21. Ratti S, Mongiorgi S, Rusciano I, Rusciano I, Manzoli L, Follo MY. Glycogen synthase kinase-3 and phospholipase C-beta signalling: roles and possible interactions in myelodysplastic syndromes and acute myeloid leukemia. *Biochim Biophys Acta* (*BBA)-Molecular Cell Res* (2020) 1867(4):118649. doi: 10.1016/j.bbamcr.2020.118649

22. Xiao W, Ando T, Wang HY, Wang HY, Kawakami Y, Kawakami T, et al. Lynand PLC- β 3-dependent regulation of SHP-1 phosphorylation controls Stat5 activity and myelomonocytic leukemia-like disease. *Blood* (2010) 116(26):6003–13. doi: 10.1182/blood-2010-05-283937

23. Li FJ. The role of WNT/Ca²⁺ pathway and the DNA methylation of ID4 regulated by UVB in cutaneous squamous cell carcinoma. Peking Union Medical College. doi: 10.27648/d.cnki.gzxhu.2019.000269

24. Nilius B, Owsianik G, Voets T, Owsianik G, Voets T, Peters JA, et al. Transient receptor potential cation channels in disease. *Physiol Rev* (2007) 87(1):165–217. doi: 10.1152/physrev.00021.2006

25. An SW, Cha SK, Yoon J, Chang S, Ross EM, Huang CL, et al. WNK1 promotes PIP2 synthesis to coordinate growth factor and GPCR-Gq signaling. *Curr Biol* (2011) 21(23):1979–87. doi: 10.1016/j.cub.2011.11.002

26. Winn MP, Conlon PJ, Lynn KL, Farrington MK, Creazzo T, Hawkins AF, et al. A mutation in the TRPC6 cation channel causes familial focal segmental glomerulosclerosis. *Science* (2005) 308(5729):1801–4. doi: 10.1126/science.1106215

27. Reiser J, Polu KR, Möller CC, Kenlan P, Altintas MM, Wei C, et al. TRPC6 is a glomerular slit diaphragm-associated channel required for normal renal function. *Nat Genet* (2005) 37(7):739–44. doi: 10.1038/ng1592

28. Kim JH, Xie J, Hwang KH, Wu YL, Oliver N, Eom M, et al. Klotho may ameliorate proteinuria by targeting TRPC6 channels in podocytes. *J Am Soc Nephrology* (2017) 28(1):140–51. doi: 10.1681/ASN.2015080888

29. Song J, Wang Y, Li X, Shen Y, Yin M, Guo Y, et al. Critical role of TRPC6 channels in the development of human renal cell carcinoma. *Mol Biol Rep* (2013) 40 (8):5115–22. doi: 10.1007/s11033-013-2613-4

30. Kim JH, Hwang KH, Eom M, Kim M, Park EY, Jeong Y, et al. WNK1 promotes renal tumor progression by activating TRPC6-NFAT pathway. *FASEB J* (2019) 33 (7):8588–99. doi: 10.1096/fj.201802019RR

31. Lee SJ, Leoni G, Neumann PA, Chun J, Nusrat A, Yun CC, et al. Distinct phospholipase C- β isozymes mediate lysophosphatidic acid receptor 1 effects on intestinal epithelial homeostasis and wound closure. *Mol Cell Biol* (2013) 33 (10):2016–28. doi: 10.1128/MCB.00038-13

32. Choi JW, Herr DR, Noguchi K, Yung YC, Lee CW, Mutoh T, et al. LPA receptors: subtypes and biological actions. *Annu Rev Pharmacol Toxicol* (2010) 50:157–86. doi: 10.1146/annurev.pharmtox.010909.105753

33. Mutoh T, Rivera R, Chun J. Insights into the pharmacological relevance of lysophospholipid receptors. *Br J Pharmacol* (2012) 165(4):829–44. doi: 10.1111/j.1476-5381.2011.01622.x

34. Lee SJ, Ritter SL, Zhang H, Shim H, Hall RA, Yun CC. MAGI-3 competes with NHERF-2 to negatively regulate LPA2 receptor signaling in colon cancer cells. *Gastroenterology* (2011) 140(3):924–34. doi: 10.1053/j.gastro.2010.11.054

35. Lin S, Lee SJ, Shim H, Chun J, Yun CC. The absence of LPA receptor 2 reduces the tumorigenesis by Apc Min mutation in the intestine. *Am J Physiology-Gastrointestinal Liver Physiol* (2010) 299(5):G1128–38. doi: 10.1152/ajpgi.00321.2010

36. Frick VO, Rubie C, Wagner M, Graeber S, Grimm H, Kopp B, et al. Enhanced ENA-78 and IL-8 expression in patients with malignant pancreatic diseases. *Pancreatology* (2008) 8(4-5):488–97. doi: 10.1159/000151776

37. Matsuo Y, Raimondo M, Woodward TA, Wallace MB, Gill KR, Tong Z, et al. CXC-chemokine/CXCR2 biological axis promotes angiogenesis in *vitro* and in *vivo* in pancreatic cancer. *Int J Cancer* (2009) 125(5):1027–37. doi: 10.1002/ijc.24383

38. Matsuo Y, Ochi N, Sawai H, Yasuda A, Takahashi H, Funahashi H, et al. CXCL8/IL-8 and CXCL12/SDF-1alpha co-operatively promote invasiveness and angiogenesis in pancreatic cancer. *Int J Cancer* (2009) 124(4):853–61. doi: 10.1002/ ijc.24040

39. Wente MN, Keane MP, Burdick MD, Friess H, Büchler MW, Ceyhan GO, et al. Blockade of the chemokine receptor CXCR2 inhibits pancreatic cancer cell-induced angiogenesis. *Cancer Lett* (2006) 241(2):221–7. doi: 10.1016/j.canlet.2005.10.041

40. Takamori H, Oades ZG, Hoch OC, Burger M, Schraufstatter IU. Autocrine growth effect of IL-8 and GROalpha on a human pancreatic cancer cell line, Capan-1. *Pancreas* (2000) 21(1):52–6. doi: 10.1097/00006676-200007000-00051

41. Hidaka H, Ishiko T, Furuhashi T, Kamohara H, Suzuki S, Miyazaki M, et al. Curcumin inhibits interleukin 8 production and enhances interleukin 8 receptor expression on the cell surface: impact on human pancreatic carcinoma cell growth by autocrine regulation. *Cancer* (2002) 95(6):1206–14. doi: 10.1002/cncr.10812

42. Kuwada Y, Sasaki T, Morinaka K, Kitadai Y, Mukaida N, Chayama K, et al. Potential involvement of IL-8 and its receptors in the invasiveness of pancreatic cancer cells. *Int J Oncol* (2003) 22(4):765–71. doi: 10.3892/ijo.22.4.765

43. Miyamoto M, Shimizu Y, Okada K, Kashii Y, Higuchi K, Watanabe A. Effect of interleukin-8 on production of tumor-associated substances and autocrine growth of human liver and pancreatic cancer cells. *Cancer Immunol Immunother* (1998) 47 (1):47–57. doi: 10.1007/s002620050503

44. Li A, King J, Moro A, Guan X, Castelvetere MP, Oblak JJ, et al. Overexpression of CXCL5 is associated with poor survival in patients with pancreatic cancer. *Am J Pathol* (2011) 178(3):1340–9. doi: 10.1016/j.ajpath.2010.11.058

45. Wang S, Wu Y, Hou Y, Wang L, Zheng L, Xie K, et al. CXCR2 macromolecular complex in pancreatic cancer: a potential therapeutic target in tumor growth. *Trans Oncol* (2013) 6(2):216–25. doi: 10.1593/tlo.13133

46. Xie K, Wei D, Huang S. Transcriptional anti-angiogenesis therapy of human pancreatic cancer. *Cytokine Growth factor Rev* (2006) 17(3):147–56. doi: 10.1016/ j.cytogfr.2006.01.002

47. Xiong HQ, Abbruzzese JL, Lin E, Wang L, Zheng L, Xie K, et al. NF- κ B activity blockade impairs the angiogenic potential of human pancreatic cancer cells. *Int J Cancer* (2004) 108(2):181–8. doi: 10.1002/ijc.11562

48. Shi Q, Abbruzzese JL, Huang S, Fidler IJ, Xiong Q, Xie K, et al. Constitutive and inducible interleukin 8 expression by hypoxia and acidosis renders human pancreatic cancer cells more tumorigenic and metastatic. *Clin Cancer Res* (1999) 5(11):3711–21.

49. Zhu VF, Yang J, Lebrun DG, Li M. Understanding the role of cytokines in Glioblastoma Multiforme pathogenesis. *Cancer Letters* (2012) 316(2):139–50. doi: 10.1016/j.canlet.2011.11.001

50. Waugh DJ, Wilson C. The interleukin-8 pathway in cancer. Clin Cancer Res (2008) 14(21):6735-41. doi: 10.1158/1078-0432.CCR-07-4843

51. Hruban RH, Adsay NV, Albores-Saavedra J, Compton C, Garrett ES, Goodman SN, et al. Pancreatic intraepithelial neoplasia: a new nomenclature and classification system for pancreatic duct lesions. *Am J Surg Pathol* (2001) 25(5):579–86. doi: 10.1097/00000478-200105000-00003

52. Takaori K, Kobashi Y, Matsusue S, Matsui K, Yamamoto T. Clinicopathological features of pancreatic intraepithelial neoplasias and their relationship to intraductal papillary-mucinous tumors. *J Hepatobiliary Pancreat Surg* (2003) 10(2):125–36. doi: 10.1007/s00534-003-0756-8

53. Ma J, Weng L, Bastian BC, Chen X. Functional characterization of uveal melanoma oncogenes. *Oncogene* (2021) 40(4):806–20. doi: 10.1038/s41388-020-01569-5

54. Rhee SG, Bae YS. Regulation of phosphoinositide-specific phospholipase C isozymes. J Biol Chem (1997) 272(24):15045-8. doi: 10.1074/jbc.272.24.15045

55. Teixeira C, Stang SL, Zheng Y, Beswick NS, Stone JC. Integration of DAG signaling systems mediated by PKC-dependent phosphorylation of RasGRP3. *Blood* (2003) 102(4):1414–20. doi: 10.1182/blood-2002-11-3621

56. Bertagnolo V, Benedusi M, Querzoli P, Pedriali M, Magri E, Brugnoli F, et al. PLC-β2 is highly expressed in breast cancer and is associated with a poor outcome: a study on tissue microarrays. *Int J Oncol* (2006) 28(4):863–72. doi: 10.3892/ijo.28.4.863

57. Bertagnolo V, Grassilli S, Volinia S, Al-Qassab Y, Brugnoli F, Vezzali F, et al. Ectopic expression of PLC- β 2 in non-invasive breast tumor cells plays a protective role against malignant progression and is correlated with the deregulation of miR-146a. *Mol Carcinogenesis* (2019) 58(5):708–21. doi: 10.1002/mc.22964

58. Bertagnolo V, Benedusi M, Brugnoli F, Lanuti P, Marchisio M, Querzoli P, et al. Phospholipase C- β 2 promotes mitosis and migration of human breast cancer-derived cells. *Carcinogenesis* (2007) 28(8):1638–45. doi: 10.1093/carcin/bgm078

59. Brugnoli F, Grassilli S, Al-Qassab Y, Capitani S, Bertagnolo V. PLC- β 2 is modulated by low oxygen availability in breast tumor cells and plays a phenotype dependent role in their hypoxia-related malignant potential. *Mol Carcinogenesis* (2016) 55(12):2210–21. doi: 10.1002/mc.22462

60. Brugnoli F, Grassilli S, Lanuti P, Marchisio M, Al-Qassab Y, Vezzali F, et al. Upmodulation of PLC- β 2 reduces the number and malignancy of triple-negative breast tumor cells with a CD133+/EpCAM+ phenotype: a promising target for preventing progression of TNBC. *BMC Cancer* (2017) 17(1):617. doi: 10.1186/s12885-017-3592-y

61. Brugnoli F, Grassilli S, Piazzi M, Palomba M, Nika E, Bavelloni A, et al. In triple negative breast tumor cells, PLC-B2 promotes the conversion of CD133high to CD133low phenotype and reduces the CD133-related invasiveness. *Mol Cancer* (2013) 12:165. doi: 10.1186/1476-4598-12-165

62. Zou D, Cui L, Ma MY, Xiao H, Song AL. Research progression of long non-coding RNA NEAT1 in breast cancer. J Modern Oncol (2022) 30(19):3607–10.

63. Cai S, Sun PH, Resaul J, Shi L, Jiang A, Satherley LK, et al. Expression of phospholipase C isozymes in human breast cancer and their clinical significance. *Oncol Rep* (2017) 37(3):1707–15. doi: 10.3892/or.2017.5394

64. Sengelaub CA, Navrazhina K, Ross JB, Halberg N, Tavazoie SF. PTPRN 2 and PLC β 1 promote metastatic breast cancer cell migration through PI (4, 5) P2-dependent actin remodeling. *EMBO J* (2016) 35(1):62–76. doi: 10.15252/embj.201591973

65. Liang S, Guo H, Ma K, Li X, Wu D, Wang Y, et al. A PLCB1-PI3K-AKT signaling axis activates EMT to promote cholangiocarcinoma progression. *Cancer Res* (2021) 81(23):5889–903. doi: 10.1158/0008-5472.CAN-21-1538

66. Linnekin D. Early signaling pathways activated by c-Kit in hematopoietic cells. Int J Biochem Cell Biol (1999) 31(10):1053–74. doi: 10.1016/S1357-2725(99)00078-3

67. Hemesath TJ, Price ER, Takemoto C, Badalian T, Fisher DE. MAP kinase links the transcription factor Microphthalmia to c-Kit signalling in melanocytes. *Nature* (1998) 391(6664):298–301. doi: 10.1038/34681

68. Chian R, Young S, Danilkovitch-Miagkova A, Rönnstrand L, Leonard E, Ferrao P, et al. Phosphatidylinositol 3 kinase contributes to the transformation of hematopoietic cells by the D816V c-Kit mutant. *Blood* (2001) 98(5):1365–73. doi: 10.1182/blood.V98.5.1365

69. Plo I, Lautier D, Casteran N, Dubreuil P, Arock M, Laurent G, et al. Kit signaling and negative regulation of daunorubicin-induced apoptosis: role of phospholipase Cgamma. *Oncogene* (2001) 20(46):6752–63. doi: 10.1038/sj.onc.1204877

70. Ning ZQ, Li J, Arceci RJ. Signal transducer and activator of transcription 3 activation is required for Asp(816) mutant c-Kit-mediated cytokine-independent survival and proliferation in human leukemia cells. *Blood* (2001) 97(11):3559–67. doi: 10.1182/blood.V97.11.3559

71. Guba M, von Breitenbuch P, Steinbauer M, Koehl G, Flegel S, Hornung M, et al. Rapamycin inhibits primary and metastatic tumor growth by antiangiogenesis: involvement of vascular endothelial growth factor. *Nat Med* (2002) 8(2):128–35. doi: 10.1038/nm0202-128

72. MacDonald TJ, Brown KM, LaFleur B, Peterson K, Lawlor C, Chen Y, et al. Expression profiling of medulloblastoma: PDGFRA and the RAS/MAPK pathway as

therapeutic targets for metastatic disease. Nat Genet (2001) 29(2):143–52. doi: 10.1038/ng731

73. Nguyen QD, Faivre S, Bruyneel E, Rivat C, Seto M, Endo T, et al. RhoA-and RhoD-dependent regulatory switch of G α subunit signaling by PAR-1 receptors in cellular invasion. FASEB J (2002) 16(6):565–76. doi: 10.1096/fj.01-0525com

74. Emami S, Le Floch N, Bruyneel E, Thim L, May F, Westley B, et al. Induction of scattering and cellular invasion by trefoil peptides in src-and RhoA-transformed kidney and colonic epithelial cells. *FASEB J* (2001) 15(2):351–61. doi: $10.1096/f_{\rm J}.00-0355$ com

75. Rodrigues S, Nguyen QD, Faivre S, Bruyneel E, Thim L, Westley B, et al. Activation of cellular invasion by trefoil peptides and src is mediated by cyclooxygenase-and thromboxane A2 receptor-dependent signaling pathways. *FASEB J* (2001) 15(9):1517–28. doi: 10.1096/fj.00-0802com

76. Attoub S, Rivat C, Rodrigues S, Van Bocxlaer S, Bedin M, Bruyneel E, et al. The c-kit tyrosine kinase inhibitor STI571 for colorectal cancer therapy. *Cancer Res* (2002) 62(17):4879–83.

77. Kim EK, Park JM, Lim S, Choi JW, Kim HS, Seok H, et al. Activation of AMPactivated protein kinase is essential for lysophosphatidic acid-induced cell migration in ovarian cancer cells. *J Biol Chem* (2011) 286(27):24036–45. doi: 10.1074/ jbc.M110.209908

78. Ratti S, Marvi MV, Mongiorgi S, Obeng EO, Rusciano I, Ramazzotti G, et al. Impact of phospholipase C β 1 in glioblastoma: a study on the main mechanisms of tumor aggressiveness. *Cell Mol Life Sci* (2022) 79(4):195. doi: 10.1007/s00018-022-04198-1

79. Bahk YY, Song H, Baek SH, Park BY, Kim H, Ryu SH, et al. Localization of two forms of phospholipase C-beta1, a and b, in C6Bu-1 cells. *Biochim Biophys Acta* (1998) 1389(1):76–80. doi: 10.1016/S0005-2760(97)00128-8

80. Aisiku OR, Runnels LW, Scarlata S. Identification of a novel binding partner of phospholipase $c\beta$ 1: translin-associated factor X. *PloS One* (2010) 5(11):e15001.

81. Lu G, Chang JT, Liu Z, Chen Y, Li M, Zhu JJ, et al. Phospholipase C beta 1: a candidate signature gene for proneural subtype high-grade glioma. *Mol Neurobiol* (2016) 53(9):6511–25. doi: 10.1007/s12035-015-9518-2

82. Bhattacharya R, Kwon J, Li X, Wang E, Patra S, Bida JP, et al. Distinct role of PLCbeta3 in VEGF-mediated directional migration and vascular sprouting. *J Cell Sci* (2009) 122(Pt7):1025–34. doi: 10.1242/jcs.041913

83. Ha JM, Baek SH, Kim YH, Jin SY, Lee HS, Kim SJ, et al. Regulation of retinal angiogenesis by phospholipase C- β 3 signaling pathway. *Exp Mol Med* (2016) 48(6): e240.

84. Okamoto H, Takuwa N, Gonda K, Okazaki H, Chang K, Yatomi Y, et al. EDG1 is a functional sphingosine-1-phosphate receptor that is linked *via* a Gi/o to multiple signaling pathways, including phospholipase C activation, Ca^{2+} mobilization, Ras-Mitogen-activated protein kinase activation, and adenylate cyclase inhibition. *J Biol Chem* (1998) 273(42):27104–10. doi: 10.1074/jbc.273.42.27104

85. Lee MJ, Thangada S, Claffey KP, Ancellin N, Liu CH, Kluk M, et al. Vascular endothelial cell adherens junction assembly and morphogenesis induced by. *Cell* (1999) 99(3):301–12. doi: 10.1016/S0092-8674(00)81661-X

86. Tatemoto K, Hosoya M, Habata Y, Fujii R, Kakegawa T, Zou MX, et al. Isolation and characterization of a novel endogenous peptide ligand for the human APJ receptor. *Biochem Biophys Res Commun* (1998) 251(2):471–6. doi: 10.1006/bbrc.1998.9489

87. Kasai A, Shintani N, Kato H, Matsuda S, Gomi F, Haba R, et al. Retardation of retinal vascular development in apelin-deficient mice. *Arterioscler Thromb Vasc Biol* (2008) 28(10):1717–22. doi: 10.1161/ATVBAHA.108.163402

88. Nolan DJ, Ciarrocchi A, Mellick AS, Jaggi JS, Bambino K, Gupta S, et al. Bone marrow-derived endothelial progenitor cells are a major determinant of nascent tumor neovascularization. *Genes Dev* (2007) 21(12):1546–58. doi: 10.1101/gad.436307

89. Gao D, Nolan DJ, Mellick AS, Bambino K, McDonnell K, Mittal V, et al. Endothelial progenitor cells control the angiogenic switch in mouse lung metastasis. *Science* (2008) 319(5860):195-8. doi: 10.1126/science.1150224

90. Maeng YS, Choi HJ, Kwon JY, Park YW, Choi KS, Min JK, et al. Endothelial progenitor cell homing: prominent role of the IGF2-IGF2R-PLCbeta2 axis. *Blood* (2009) 113(1):233–43. doi: 10.1182/blood-2008-06-162891

91. Lee OH, Bae SK, Bae MH, Lee YM, Moon EJ, Cha HJ, et al. Identification of angiogenic properties of insulin-like growth factor II in *in vitro* angiogenesis models. *Br J Cancer* (2000) 82(2):385–91. doi: 10.1054/bjoc.1999.0931

92. Björndahl M, Cao R, Nissen LJ, Clasper S, Johnson LA, Xue Y, et al. Insulin-like growth factors 1 and 2 induce lymphangiogenesis in vivo. *Proc Natl Acad Sci U.S.A.* (2005) 102(43):15593–8. doi: 10.1073/pnas.0507865102

93. Kim KW, Bae SK, Lee OH, Bae MH, Lee MJ, Park BC, et al. Insulin-like growth factor II induced by hypoxia may contribute to angiogenesis of human hepatocellular carcinoma. *Cancer Res* (1998) 58(2):348–51.

94. Ratti S, Ramazzotti G, Faenza I, Fiume R, Mongiorgi S, Billi AM, et al. Nuclear inositide signaling and cell cycle. *Adv Biol Regul* (2018) 67:1–6. doi: 10.1016/ j.jbior.2017.10.008

95. Follo MY, Mongiorgi S, Clissa C, Paolini S, Martinelli G, Martelli AM, et al. Activation of nuclear inositide signalling pathways during erythropoietin therapy in low-risk MDS patients. *Leukemia* (2012) 26(12):2474–82. doi: 10.1038/leu.2012.133

96. Xu P, Wang DJ, Wang JH, Dong JQ, Wang CY, Jiang YF, et al. Expression of PI-PLC β 1 in myelodysplastic syndromes and its significance. J Harbin Med Univ (2021) 55 (01):41–3.

97. Zhong W, Xu M, Li C, Zhu B, Cao X, Li D, et al. ORP4L extracts and presents PIP2 from plasma membrane for PLC β 3 catalysis: targeting it eradicates leukemia stem cells. *Cell Rep* (2019) 26(8):2166–2177.e9. doi: 10.1016/j.celrep. 2019.01.082

98. Trinquet E, Fink M, Bazin H, Grillet F, Maurin F, Bourrier E, et al. D-myoinositol 1-phosphate as a surrogate of D-myo-inositol 1,4,5-tris phosphate to monitor G protein-coupled receptor activation. *Anal Biochem* (2006) 358(1):126–35. doi: 10.1016/j.ab.2006.08.002