Check for updates

OPEN ACCESS

EDITED BY Zheng Wang, Shanghai Jiao Tong University, China

REVIEWED BY Pai-Sheng Chen, National Cheng Kung University, Taiwan Yuqi Yang, St. John's University, United States

*CORRESPONDENCE Zhiqun Cao Caozhiqun@163.com

[†]These authors have contributed equally to this work and share first authorship

RECEIVED 05 July 2023 ACCEPTED 06 September 2023 PUBLISHED 27 September 2023

CITATION

Zhou J, Xiang H and Cao Z (2023) Dual mechanism of Let-7i in tumor progression. *Front. Oncol.* 13:1253191. doi: 10.3389/fonc.2023.1253191

COPYRIGHT

© 2023 Zhou, Xiang and Cao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Dual mechanism of Let-7i in tumor progression

Jiapei Zhou^{1†}, Hongjie Xiang^{2†} and Zhiqun Cao^{1*}

¹First Clinical Medical College, Shandong University of Traditional Chinese Medicine, Jinan, China, ²Department of Traditional Chinese Medicine, The First Affiliated Hospital of Shandong First Medical University & Shandong Provincial Qianfoshan Hospital, Jinan, China

Let-7i regulates tumors primarily by binding to the 3' untranslated region (3' UTR) of mRNA, which indirectly regulates post-transcriptional gene expression. Let-7i also has an epigenetic function via modulating DNA methylation to directly regulate gene expression. Let-7i performs a dual role by inducing both the promotion and inhibition of various malignancies, depending on its target. The mechanism of Let-7i action involves cancer cell proliferation, migration, invasion, apoptosis, epithelial-mesenchymal transition, EV transmission, angiogenesis, autophagy, and drug resistance sensitization. Let-7i is closely related to cancer, and hence, is a potential biomarker for the diagnosis and prognosis of various cancers. Therapeutically, it can be used to promote an anti-cancer immune response by modifying exosomes, thus exerting a tumor-suppressive effect.

KEYWORDS

Let-7i, cancer, promoter, inhibitor, dual mechanism

1 Introduction

MicroRNA (miRNA) refers to short non-coding RNA with a length of 19-25 nucleotides that functions as a conservative post-transcriptional regulator of gene expression. It recruits Argonaute proteins to form the RNA-induced silencing complex (RISC), which regulates RNA via base-complementary pairing. The combination of miRNA and RISC can inhibit mRNA translation without destroying the stability of mRNA as well as silence unwanted genetic material and transcripts (1, 2). When the miRNA and mRNA involved are entirely complementary, the complex can also mediate mRNA degradation to inhibit transcription (3), thereby regulating the production of the resulting protein. In addition, it has been found that mature miRNAs have the ability to enter the nucleus, directly combine with the original components of gene promoter regions, and contribute to the regulation of non-classical gene transcription (4). MiRNA is involved in almost all biological processes, including cell growth, proliferation, differentiation, metabolism, and the development of organisms (5). Each miRNA binds to hundreds of different mRNAs, and miRNA controls more than half of human protein-coding genes (6). Therefore, the dysregulation of miRNA expression is closely related to the occurrence of various diseases, including cancer (7).

MiRNA biogenesis requires a series of sequential processing events. First, miRNA is transcribed as long primary transcripts (pri-miRNA). This pri-miRNA is subsequently

trimmed to 70-nucleotide (nt) pre-miRNAs in the nucleus. Then, the trimmed pre-miRNA is exported to the cytoplasm and synergized by Dicer and Drosha, which are both members of the RNase III superfamily of bidentate nucleases. This cleavage event yields mature miRNA molecules that are approximately 22 nt in length (8–11).

Let-7 was first found in the nematode and identified as a key developmental regulator (12). It is one of the two first known microRNAs (the other one being Lin-4) and the first known human miRNA. The Let-7 family is often present in multiple copies in a genome (13). To distinguish between its multiple subtypes, a letter is placed after Let-7 to represent its various sequences, while numbers at the end of the name indicate that the same sequence exists in multiple genomic locations (13, 14). There are 10 mature Let-7 family sequences in humans that arise from 13 precursor sequences and function in similar ways (13).

Let-7 expression is reportedly downregulated in several human cancers, including esophageal, lung, and breast cancers. As a tumor suppressor, Let-7 miRNA targets various oncogenic molecules (including RAS, HMGA 2, and cell cycle and apoptosis regulators) and exerts its anti-tumor effect by preventing proliferation, promoting apoptosis, inhibiting angiogenesis, and reducing immune surveillance (15–17).

Small differences in the sequence of Let-7 can alter the affinity for its target sequences, thereby resulting in differences in its function or employed mechanism (18). The expression of different family members also varies significantly between tumors. Most Let-7i family members exert an anti-tumor effect to function as tumor suppressors (19), but interestingly, recent studies have found that Let-7i may also act as an oncogene to promote the occurrence and development of cancer (20, 21). Let-7i has been shown to have tumor-suppressive as well as tumor-promoting properties simultaneously. To clarify the specific mechanisms differentiating between the tumor-suppressing and tumorpromoting roles of Let-7i, the current Review summarizes previous studies to provide guidance for further targeted precision therapies in a clinical setting.

2 Tumor suppressor function

Let-7i has been widely recognized and studied as a tumor suppressor. Its mechanism of inhibiting tumor development involves not only the modulation of cell proliferation, metastasis, and changes in the tumor cells themselves (such as autophagy, apoptosis, and stem cell properties) but also changes in the tumor microenvironment, such as alterations to immunity and angiogenesis.

2.1 Regulation of malignant phenotypes: proliferation, migration, invasion, and apoptosis

Let-7i regulates gene expression to control the processes that underpin malignant phenotypes, such as tumor cell proliferation, migration, invasion, and apoptosis. Let-7i can regulate gene expression indirectly via the classical mRNA regulatory pathway or directly via the non-canonical epigenetic regulation pathway.

2.1.1 mRNA regulation

The regulation of protein levels by specifically binding to the mRNA 3' UTR is the classic mechanism employed by Let-7i. Let-7i reduces melanoma cell proliferation and metastasis by upregulating KISS1 expression (22), inhibits the proliferation and invasion of osteosarcoma by downregulating the expression of Aurora B (a member of the serine/threonine protein kinase family) (23), inhibits the survival, proliferation, and motility of gastric cancer cells by downregulating the expression of COL1A1 (24), and promotes the DDP-induced apoptosis of esophageal cancer cells by downregulating the expression of ABCC10 (25). In the process of suppressing the occurrence and development of colorectal cancer, Let-7i can not only specifically bind to serine protease (KLK6) mRNA to inhibit its transcription (26) but also inhibits the activity of the ERK signaling pathway by inhibiting the expression of CCND1 (27). In glioblastoma, a study by Xiaopeng Sun et al. found that Let-7i-5p could downregulate the levels of cyclin-dependent kinases (CDK2 and CDK4), cyclin A2, and BCL-2 by silencing GALE, thereby inducing cell cycle arrest and a reduction in proliferation (28). Furthermore, according to experiments by Lobna Elkhadragy, ERK3 and BMI1 are both highly expressed in head and neck cancer and BMI1 upregulates ERK3 by suppressing the expression of Let-7i, ultimately facilitating the migration of head and neck cancer cells (29). Therefore, we speculated that Let-7i can prevent head and neck cancer cells from migrating by reducing the activity of ERK3.

2.1.2 Epigenetic alterations

DNA, histones, non-histone proteins, and a small amount of RNA can all bind and interact with chromatin, which is a linear complex structure containing the genetic material of interphase cells (30). Epigenetics refers to heritable modifications of gene function that ultimately alter phenotype but do not entail changes to the DNA sequence itself. DNA methylation, histone modification, non-coding RNA regulation, and chromatin remodeling are all examples of epigenetic processes. Let-7i may play a tumor-regulating role by modulating epigenetics.

Let-7i acts on histone lysine demethylase to achieve tumor suppression through structural modification. In esophageal cancer, KDM5B, a histone 3 lysine 4 (H3K4) methylation regulator (31), can be downregulated by Let-7i to encourage the tri-methylation of H3K4 (H3K4me3) in the promoter region, consequently promoting the expression of the tumor suppressor SOX17 (32). Overexpressed SOX17 can then silence the tumor promoter GREB1, thereby reducing the proliferation and invasion of esophageal cancer cells and exerting anti-tumor efficacy in vitro (32, 33). In lung cancer, Let-7i enhances DCLK1 expression by interacting with endogenous KDM3A, allowing KDM3A to bind to the promoter region of DCLK1 and removing histone H3K9me2 (34). The enhancement of DCLK1 expression promotes the expression of FXYD3, which reduces the ability of lung cancer cells to proliferate, migrate, and invade, thereby exerting its anti-tumor effect. The above mechanisms have been confirmed in vitro and in vivo (34). Yawen Liu et al. suggested that Lin28B upregulates the level of TET3 by blocking Let-7i, while TET3 catalyzes the conversion of 5methylcytosine to 5-hydroxymethylcytosine, resulting in DNA depletion and pancreatic cell carcinoma (35). Through a feedback mechanism, TET3 and Let-7i can also promote the expression of Lin28B (35, 36).

High mobility group proteins A1 and 2 (HMGA1, 2) are members of the HMGA family that are a class of non-histone chromatin structural proteins with no transcriptional activity, which primarily regulate transcription by altering DNA conformation (37). HMGA1 exerts its tumor-regulating effects via multiple pathways, including DNA phosphorylation, acetylation, and methylation (37). Qin et al. illustrated that by targeting HMGA1, Let-7i suppressed the malignant phenotype of bladder cancer cell lines T24 and 5637 (38). According to Ravindresh Chhabra's study, Let-7i-5p overexpression and SOX2 silencing could both decrease the number of spheroids formed in the cervical cancer cell lines HeLa and CaSki, while HMGA2 and SOX2 expression were significantly reduced in CaSki following Let-7i-5p overexpression (39). HMGA2 has been shown to induce SOX2; therefore, we speculate that Let-7i-5p can disrupt the stem cell phenotype, alter the conformation of DNA, and downregulate SOX2 expression by targeting HMGA2 expression (39-41). Figure 1 shows a schematic diagram illustrating the way in which Let-7i regulates epigenetics.

2.2 Tumor microenvironment pathway regulation

2.2.1 Epithelial-mesenchymal transition and mesenchymal phenotype

Epithelial-mesenchymal transition (EMT) is the process by which epithelial cells lose polarity and transform into motile mesenchymal cells, acquiring a mesenchymal phenotype (42). This process mediates tumor metastasis by blocking connections between cells, reorganizing the cytoskeleton, altering cell shape, and encoding gene expression to enhance cell motility, migration, and invasion. In addition, EMT promotes stem cell likeness and plays a key role in the processes of treatment resistance, embryonic development, and organ fibrosis (42–44). Epithelial cadherin (Ecadherin) degradation is a fundamental mechanistic feature that deconstructs intercellular links and induces EMT, leading to tumor metastasis (42).

Hypoxia is a common feature of the tumor microenvironment. It can activate hypoxia-inducible factor-1 α (HIF-1 α) to further regulate the expression levels of Nur77 (45) and TWIST1 (46). Nur77 is a distinct nuclear receptor, the low expression of which can cause E-cadherin to be downregulated, causing more dispersed colonies to form and triggering EMT and typical mesenchymal morphology (45). Let-7i-5p plays a key role in this process. Under



hypoxia, Nur77 interacts with p63 to specifically inhibit Dicer, which affects the maturation of Let-7i-5p from precursor (pre)-Let-7i, resulting in a decrease in Let-7i-5p levels (45). Let-7i-5p binds p110 α mRNA on the 3' untranslated region (UTR) and promotes its degradation, while low expression of Let-7i-5p reduces the degradation of PI3K-p110 α to increase its level and activates the Akt signaling pathway. Additionally, the low expression of Let-7i-5p affects the phosphorylation of downstream mTORC1 and its target proteins p70S6K and 4E-BP1, thereby inducing colorectal cancer (CRC) EMT (45). Figure 2 provides an intuitive illustration of the above mechanism.

TWIST1, a basic helix-loop-helix (bHLH) transcription factor, is a master regulator of EMT (46–48). It is regulated by HIF-1α, regulates BMI1 levels, and cooperates with BMI1 to inhibit E-cadherin expression to induce EMT (49). Let-7i expression can be co-repressed by TWIST1 and BMI1 simultaneously, while low Let-7i levels can increase cell invasiveness. Let-7i downregulation changes the morphology of head and neck squamous cell carcinoma (HNSCC) OECM-1 cells, causing them to adopt an elongated shape with pseudopodia protrusions, which promotes the interstitial cell pattern, ultimately increasing their capacity to move and invade. In addition, downregulating Let-7i increases the expression of NEDD9 and DOCK3, activates RAC1, drives

interstitial movement, and further enhances the invasive phenotype (50).

Moreover, in human glioma cells, Yuan et al. confirmed that Let-7i directly targets IKBKE (inhibitor of nuclear factor kappa-B kinase subunit epsilon) to upregulate E-cadherin expression and suppress EMT (51). In endometrial cancer cells, Let-7i is expressed at low levels under the control of DICER1, and low levels of Let-7i have been found to downregulate the expression of EZH2 to affect the methylation of histone H3 at arginine 27 as well as total H3 acetylation, thereby inhibiting the expression of E-cadherin and encouraging EMT (52). In head and neck squamous cell carcinoma (HNSCC), Let-7i inhibits MBP4 to alter cell morphology, turn slender cells round, and decrease interstitial movement, ultimately preventing local invasion (53).

2.2.2 Extracellular vesicles

Extracellular vesicles (EVs), including exosomes and shed microvesicles (sMVs), mediate intercellular trafficking and are crucial for enabling bidirectional communication between cells and the microenvironment at both the paracrine and systemic levels (54). Studies have repeatedly demonstrated the close connection between EVs and the emergence of cancer. EVs transport a wide range of molecules from donor cells to recipient



Frontiers in Oncology

cells, including proteins (such as oncoproteins and oncopeptides), RNAs (such as microRNA and mRNA), DNA fragments, and lipids; this process profoundly alters the phenotype of the tumor microenvironment (54–56). Adeleh Taghi Khani et al. confirmed by *in vivo* and *in vitro* experiments that Let-7i can be delivered by the intercellular delivery system-EV, exerting its tumor suppressive effect in breast cancer cells (57). Experiments conducted by Jiefeng Liu et al. demonstrated that Let-7i inhibits the malignant phenotype of lung cancer through EV transport (34). Additionally, results from a study by Deyi Xiao et al. suggest that Let-7i may act on LIN28B and HMGA2 to alter the expression of EMT markers, thereby inhibiting exosome-mediated melanocyte invasion by suppressing EMT-like effects (58).

3 Tumor promoter function

Although Let-7i is widely recognized as a tumor suppressor, increasingly more studies in recent years have found that Let-7i also has a tumor-promoting effect. Moreover, it appears to promote tumor growth and development through different pathways in different tumors.

3.1 Classical pathways to modulating malignant phenotypes: proliferation, migration, invasion, and apoptosis

Let-7i promotes hepatocellular carcinoma (HCC) by targeting TSP1. By conducting in vitro experiments, Hee Doo Yang et al. found that Let-7i-5p rescued a range of tumor suppressive effects of HDAC6, while the ectopic expression of a Let-7i-5p antisense inhibitor (AS-Let-7i-5p) inhibited tumor cell proliferation, induced apoptosis, and prevented migration under chemotactic stimulation, revealing that Let-7i-5p promotes HCC (20). The thrombospondin-1 gene (THBS1) 3' UTR was cloned into a reporter vector and detected using an AS-Let-7i-5p dual-luciferase reporter assay, after which, there was an observed increase in the relative luciferase activity (20). In addition, we observed that AS-Let-7i-5p transfection increased thrombospondin-1 protein (TSP1) secretion in the conditioned medium of HCC cells. It has been proposed that Let-7i-5p can interact directly with the transcript 3' UTR to selectively regulate the expression of THBS1, thereby regulating TSP1 secretion. The inhibition of Let-7i-5p can upregulate the level of TSP1 and inhibit both tumor growth and invasion (20). Therefore, we hypothesized that Let-7i-5p may contribute to tumor growth by suppressing the expression of THBS1 and lowering TSP1 levels. In nasopharyngeal carcinoma (NPC), Let-7i-5p has been demonstrated to act not only as an oncogene to promote cancer but also as a valuable biomarker to evaluate its end stage, predict its recurrence, and predict its metastasis risk. Bo You et al. revealed that Let-7i-5p expression was upregulated in NPC and was significantly associated with clinical stage, recurrence, and metastasis. Patients with a higher ISH staining score exhibited higher Let-7i expression, while patients with higher Let-7i-5p expression displayed worse overall survival

(OS) and progression-free survival (DFS) rates (59). Simultaneously, the study confirmed the faciliatory effect of Let-7i-5p on the proliferation and migration of NPC cells through several in vitro experiments (59). Results obtained from luciferase gene assays showed that Let-7i-5p binds to the 3' UTR of ATG10 and ATG16L1, revealing the direct targeting effect on genes (59). Let-7i-5p promotes tumor cell proliferation and migration, while the knockdown of ATG10 and/or ATG16L1 abolished this effect, indicating that Let-7i-5p exerts its effect by controlling ATG10 and ATG16L1 (59). In renal clear cell carcinoma (ccRCC), Let-7i-5p is also highly expressed as an oncogene, and its expression level is strongly associated with the pathological stage. Experiments conducted by Yujie Liu et al. showed that the level of Let-7i differed significantly across different pathological stages and different AJCC stages, allowing it to be used as a prognostic marker for ccRCC (21). Meanwhile, the same research showed that Let-7i-5p can promote malignant phenotypes by directly targeting hyaluronan-binding protein 4 (HABP4) (21). HABP4 is a nuclear and cytoplasmic regulatory protein involved in the regulation of gene expression at the transcriptional and mRNA levels. Additionally, it regulates the cell cycle and apoptosis to modulate cell proliferation (60). Downregulating the level of HABP4 to regulate the cell cycle may therefore be the mechanism by which Let-7i promotes ccRCC (21).

Interestingly, Let-7i can promote or suppress hepatocellular carcinoma growth by acting on different targets. Let-7i promotes HCC proliferation and invasion by upregulating the expression of THBS1 and TSP1. Conversely, it also inhibits the malignant phenotype of HCC cells via multiple pathways. A study by Injie Omar Fawzy et al. indicated that Let-7i can inhibit the viability and colony-forming ability of HCC cells either by directly targeting IGF1R or by indirectly reducing IGF1R expression via regulating the expression of insulin-like growth factor 2-mRNA-binding proteins (IGF2BP) 1, 2, and 3 (61). Alternatively, Let-7i can mediate the downregulation of the apoptosis protein Bcl-xL, thereby inhibiting HCC (62). Figure 3 features a schematic diagram that summarizes the classical mechanism of action of Let-7i.

3.2 Angiogenesis and extracellular vesicles

Angiogenesis in tumor tissue is an important prerequisite for rapid tumor proliferation. Tumor tissue blood vessels originate from the pre-existing vasculature and serve as a source of nutrients and oxygen for the tumor cells to ensure their rapid proliferation. The development of vascular architecture in the tumor microenvironment depends on the coordination of pro- and antiangiogenic factors (63). Hee Doo Yang et al. treated HCC cells with AS-Let-7i-5p and rTSP1, finding that the *in vitro* development of microtubule cells was noticeably suppressed. This effect was successfully rescued by combining the treatment with the TSP1 antibody C-terminal domain to the CD47 receptor (3F352). The research elucidated the following mechanism: the downregulation of Let-7i-5p levels mediates TSP1 binding to the cell surface receptor CD47 to exert anti-angiogenic activity (20). We thus



concluded that Let-7i contributes to the promotion of angiogenesis during the development of tumors.

In a study by Hee Doo Yang et al. (20), exosomes were isolated and purified from HCC cell culture medium, from which Let-7i-5p was detected by qPCR, and donor cell fractions were analyzed. The results of the study showed that Let-7i-5p was mainly present in the exosomes but not in the donor cells of HCC cells. The exosomes were then fluorescently labeled with PKH67 dye and incubated with the receptor system. Measurements found that the expression of Let-7i-5p in the receptor cells was significantly enhanced, suggesting that in HCC, Let-7i-5p facilitates communication between liver cancer cells and normal cells via exosomes, subsequently promoting the malignant transformation of cells and the development of cancer (20).

3.3 Regulation of autophagy

Autophagy is an intracellular degradation process that fuses autophagosomes and lysosomes by the action of various autophagy genes. It hydrolyzes damaged organelles and macromolecules by hydrolases (64). Autophagy plays a complex dual role in tumors, not only by inducing programmed death to eliminate tumor cells but also by promoting cancer cell-stroma communication to promote tumorigenesis and development, supporting tumor growth in a nutrient-limited environment (65, 66). Cancer autophagy is affected by factors such as nutrient availability, microenvironmental stress, and the immune system (65). Numerous studies have documented how miRNAs regulate autophagy and how autophagy affects tumor progression (67, 68).

In NPC, Bo You et al. found that the transfection of NPC cells with a Let-7i-5p inhibitor could inhibit their proliferation and migration ability via autophagy (59). The research also found that silencing the expression of Let-7i-5p induced LC3 aggregation and increased the number of both yellow fluorescent autophagosomes and red fluorescent autolysosomes in the autophagic flux assay, indicating that Let-7i-5p can inhibit the formation of the autophagy phagosome and inhibit the autophagic flux of NPC cells (59). Furthermore, after knocking out Let-7i-5p, western blot showed that the expression levels of the autophagy marker LC3-II and the autophagy-related gene ATG5 were significantly increased, while the level of the autophagy substrate p62 was decreased (59). The modulation of autophagy by Let-7i was also observed in non-small cell lung cancer (NSCLC). The transfection of a Let-7i-5p inhibitor into NSCLC cells resulted in an increase in the LC3-II/LC3-I ratio and an increase in the number of autophagosomes, while p62 levels were decreased, suggesting that Let-7i-5p negatively regulates autophagy (69). Taken together, Let-7i-5p exerts a tumorigenic role in NPC through the inhibition of autophagy activity (59).

3.4 Regulation of immune escape

As an important part of the immune system, innate immunity is the first line of defense against infection and malignant cell transformation (70). Macrophages can act as antigen-presenting cells (APCs) in innate immunity, processing and cross-presenting antigens to T cells to activate adaptive immunity (71). In addition, macrophages have the ability to mediate phagocytosis, involving multiple cell processes such as target cell recognition, phagocytosis, and lysosomal digestion, which are essential for the programmed clearance of damaged and foreign cells (72). Phagocytosis depends on the relative expression of pro- and anti-phagocytic signals on target cells. Tumor cells have been shown to evade macrophage phagocytosis by expressing anti-phagocytic signals, including CD200 and CD47 (73).

In hepatocellular carcinoma (HCC), TSP1 can prevent the interaction between CD47 and SIRP α , disrupt the "don't eat me" signal between hepatoma cells and macrophages, and prevent immune escape (20). SIRP α is a signal-regulating protein that is mainly expressed on the surface of myeloid cells such as macrophages. It binds to the transmembrane protein CD47 and is activated to initiate a signal transduction cascade, resulting in the inhibition of phagocytosis (74). It has been reported that cell migration ability was significantly inhibited following the treatment of HCC cells with a Let-7i-5p antisense inhibitor and recombinant TSP1, whereas combined treatment with 3F352 rescued these responses, suggesting the existence of an autocrine/ paracrine TSP1-CD47 mechanism in HCC cells (20). Then, co-cultured mouse peritoneal macrophages and HCC cells were treated

with a Let-7i-5p antisense inhibitor and recombinant TSP1, and consequently, an increase in the phagocytic index and enhanced macrophage phagocytic activity were observed. This suggests that TSP1 can compete with SIRP α for CD47, convert the CD47-SIRP interaction between HCC and macrophages into the CD47-TSP1 interaction, activate the "eat me" signal, and restart macrophage phagocytosis. However, Let-7i could target and downregulate the levels of TSP1, inhibiting the competitive binding of TSP1, which suppressed the immune response, mediated immune escape, and encouraged the development of tumors (20).

Figure 4 illustrates the mechanisms by which Let-7i suppresses immunological response, promotes angiogenesis, inhibits autophagy, and delivers via exosomes.

4 Diagnostic and prognostic biomarkers

Although there is an extensive body of research on cancer and a deep understanding of its development, numerous challenges remain regarding its diagnosis. Most cancers occur insidiously but develop rapidly, and when diagnosed, they are often already at an advanced stage, which is greatly related to an untimely diagnosis. Therefore, it is of great significance to improve the diagnostic methods, establish convenient, accurate, and efficient diagnostic biomarkers, detect lesions in a timely manner, and follow up and confirm diagnoses at an early stage.

In almost all cancer types, miRNA signatures are enriched for proteoglycan-related proteins. Proteoglycans are macromolecules



FIGURE 4

Mechanism diagram of Let-7i regulating autophagy, angiogenesis, immune response, and exosomes. Let-7i suppresses autophagy by reducing LC3 aggregation. Let-7i blocks the immune response and initiates immune evasion by downregulating TSP1. Additionally, Let-7i inhibits the antiangiogenic effects of TSP1. Furthermore, Let-7i transmits between cells through exosomes. that are major components of the extracellular matrix, and alterations in their expression correlate with the prognosis of malignant tumors (75, 76). Specific miRNA signatures regulate proteoglycan and stem cell pluripotency in the tumor microenvironment, which may have profound implications for early cancer detection.

Sathipati et al. (77) suggested that the recognition of Let-7i signatures by CancerSig miRNAs can be used as a basis for predicting the development and stage of various types of cancers, which can help in early cancer identification and stratification. In experiments conducted by Liang Li et al. (78), serum Let-7i was detected in preclinical HCC patients and has the potential to be used in the screening of CHB patients at high risk of developing HCC 6-12 months after the measurement of miRNAs. Cochetti (79) used Let-7i to differentiate patients with prostate cancer from those with benign prostatic hyperplasia and found that the expression level of Let-7i decreased with increasing malignancy of prostate cancer, which led to the suggestion that Let-7i may be a potential marker for high-risk disease. In addition, Let-7i was found to be a potential biomarker for smoking-associated pneumonia (80). The nucleotide diversity of Let-7i can also affect the risk of cervical cancer, head and neck cancer, and many other cancers by influencing Let-7i levels (81, 82).

The issue of tumor prognosis, which considers tumor recurrence and metastasis, is still the focus of attention in the prevention and treatment of malignant tumors, as it seriously affects the survival time and quality of life of affected patients. Therefore, there is an urgent need to identify a biological marker to monitor the effect of tumor treatment and determine prognosis to intervene early, adjust the treatment plan in time, and select the optimal treatment. Let-7i has been extensively studied as a candidate prognostic biomarker for clinical applications. For pancreatic neuroendocrine tumors, Let-7i predicts metabolic aggressiveness and contributes to pancreatic neuroendocrine tumor (PanNET) stratification by peptide receptor radionuclide therapy (PRRT) (83). Let-7i was found to be significantly associated with hepatitis infection and overall survival in patients with hepatocellular carcinoma and was an independent factor in the development of hepatocellular carcinoma (HCC) in patients with chronic hepatitis B (CHB) and chronic hepatitis C (CHC) (84). Let-7i is an early predictor of HCC development after antiviral therapy, and circulating Let-7i levels can be used for the early surveillance of CHB and CHC with HCC risk and as a non-invasive biomarker to predict the risk of hepatocellular carcinoma after antiviral therapy in patients with chronic hepatitis B and C (84). Moreover, Let-7i is associated with poorer overall cancer survival (OS) and could be a potential biomarker for prognostic survival in individuals with tumors (85). Let-7i has been demonstrated to be a good predictor of overall survival (OS) in metastatic renal cancer (86), recurrencefree survival (RFS) in oral cancer (87), progression-free survival (PFS) in advanced ovarian cancer (88), and liver metastasis-free survival (HFS) in colorectal cancer (89). Let-7i not only predicts OS in gastric cancer but also predicts the sensitivity of gastric cancer to chemotherapy (90). Similarly, Let-7i can be predictive of chemotherapy resistance in ovarian and breast cancer cells (88).

The potential biomarker role of Let-7i renders it relevant for clinical studies. A proportional risk model for COX has been developed using the expression of miRNAs, including Let-7i, to robustly predict the high and low risk of distant metastasis in nasopharyngeal carcinoma patients (91).

5 Clinical target

Owing to the important role of Let-7i in tumorigenesis and development, research on its utility as a therapeutic target is progressively expanding. Let-7i can enable the cell-cell delivery of Let-7i via exosomes (20). Additionally, it can effectively induce dendritic cell (DC) maturation, which plays a key role in generating an anti-tumor immune response. Based on this, Let-7i-modified exosomes have emerged as a primary therapeutic direction, and this technology can be administered by either intramuscular or intraperitoneal injection to target DCs and promote their maturation as well as enhance the proliferation of T-cells and regulate the release of cytokines, thus exerting a powerful anti-tumor response through enhancing the immune response and remodeling the tumor microenvironment (57, 92). Let-7i-modified exosomes also hold promise in the development of a novel cell-free vaccine for cancer therapy (92).

In addition, Let-7i has been extensively studied for its ability to enhance the sensitivity of cancer cells to chemotherapeutic drugs (93). Let-7i can inhibit the transcription of lncRNA XIST and downregulate the expression of XIST (94). LncRNA XIST has been shown to confer chemoresistance to cancer cells via a variety of pathways, including improved DNA repair and apoptosis regulation (93). Therefore, the downregulation of lncRNA XIST can reduce the proliferation and anti-apoptotic ability of lung adenocarcinoma (LAD) cells, enhance LAD sensitivity to cisplatin, and improve the drug resistance of cancer cells (94). Yan-Ling Ren et al. (95) confirmed that propofol is not only useful as an intravenous anesthetic but also exerts non-anesthetic effects by interacting with various signaling pathways, thereby participating in the regulation of various human malignant tumors. Propofol can reduce HOXA11-AS expression and upregulate Let-7i to regulate the expression of ABCC10 and alleviate the resistance of colon cancer to chemotherapy (95). Nenghui Liu et al. illustrated that a MUC1 aptamer-Let-7i chimera can enhance the sensitivity of epithelial ovarian cancer cells to paclitaxel by downregulating the expression levels of cyclin D1, cyclin D2, Dicer 1, and PGRMC1 (96).

6 Article summary

By reviewing the role and mechanism of Let-7i in various tumors, we conclude that Let-7i not only plays a tumor suppressor role but also acts as an oncogenic factor to promote the occurrence and development of cancer. Let-7i employs multiple mechanisms of action across different cancers in a cancer-specific manner. Furthermore, for the same cancer cell, depending on the target, it also plays a different role in promotion and inhibition. Multiple processes underlying the cancer phenotype, including cancer cell growth, migration, invasion, apoptosis, stem celllikeness, epithelial-mesenchymal transition, EV transmission, angiogenesis, immune evasion, and autophagy, are all regulated by Let-7i. Furthermore, Let-7i is a potential biomarker for the diagnosis or prognosis of various diseases. Therapeutically, Let-7i can modulate the anti-cancer immune response by modifying exosomes while also contributing to the sensitivity of cancer cells to chemotherapeutic drugs to varying degrees. The mechanism of Let-7i is complex and detailed. Table 1 summarizes different aspects of the mechanism of Let-7i in this paper, providing a theoretical basis and reference for the future use of Let-7i as a clinical target in the treatment of cancer.

TABLE 1 Summary of the Let-7i mechanism.

Author contributions

The idea comes from ZC and JZ, the article is written by JZ, article modification is by HX, all authors reviewed the article. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by the National Natural Science Foundation of China (82104723); Natural Science Foundation of Shandong Province Project (ZR2021LZY019); Key Project Subject of Traditional Chinese Medicine Science and Technology Development in Shandong Province (2020Z14); Shandong Provincial Key Discipline

Cancer type	The role of Let-7i	Direct target	Expression status	Downstream pathways involved	Involved phenotype
Hepatocellular carcinoma	Promoter	THBS1	Downregulate	TSP1	Proliferation, migration, apoptosis, angiogenesis, immune escape
	Inhibitor	Bcl-xL	Downregulate		Proliferation, apoptosis
	Inhibitor	IGF1R	Downregulate		Proliferation, migration, Invasion
	Inhibitor	IGF2BPs	Downregulate	IGF1R	Proliferation, migration, Invasion
Nasopharyngeal carcinoma	Promoter	ATG10, ATG16L1	Downregulate		Proliferation, migration
Clear cell renal cell carcinoma	Promoter	HABP4	Downregulate		Proliferation, migration, invasion
Melanoma	Inhibitor	KISS1	Upregulate		Proliferation, migration,
Osteosarcoma	Inhibitor	Aurora B	Downregulate		Migrate, Invasion
Stomach cancer	Inhibitor	COL1A1	Downregulate		Proliferation, migration,
Esophageal cancer	Inhibitor	ABCC10	Downregulate		Apoptosis
		KDM5B	Downregulate	SOX17, GREB1	proliferation, migration, Invasion, apoptosis
Colorectal cancer	Inhibitor	KLK6	Downregulate	Caspase signaling pathway	Proliferation, migration, Invasion, apoptosis
		CCND1	Downregulate	ERK signaling pathway	Proliferation, migration, invasion
		p110α	Downregulate	Akt	migrate, Invasion
Glioblastoma	Inhibitor	GALE	Downregulate	CDK2, CDK4, BCL-2, Cyclin A2	Proliferation, migration, angiogenesis
		IKBKE	Downregulate		migrate, Invasion
Head and neck cancer	Inhibitor	ERK3	Downregulate		Migrate
		NEDD9, Dock3	Downregulate	RAC1	Invasion
		MBP4	Downregulate		Migrate
Lung cancer	Inhibitor	KDM3A	Downregulate	DCLK1, FXYD3	Proliferation, migration, Invasion
Lung adenocarcinoma	Inhibitor	XIST	Downregulate		Proliferation, apoptosis, drug resistance
Pancreatic cancer	Inhibitor	TET3	Downregulate		Proliferation, invasion
Bladder Cancer	Inhibitor	HMGA1	Downregulate		Proliferation, migration, Invasion
Cervical cancer	Inhibitor	HMGA2	Downregulate	SOX2	Cell stemness
Endometrial cancer	Inhibitor	EZH2	Downregulate		Invasion

Construction Project of Traditional Chinese Medicine (Clinical Discipline of Integrated Traditional Chinese and Western Medicine).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Lu TX, Rothenberg ME. MicroRNA. J Allergy Clin Immunol (2018) 141(4):1202– 7. doi: 10.1016/j.jaci.2017.08.034

2. Ha M, Kim VN. Regulation of microRNA biogenesis. *Nat Rev Mol Cell Biol* (2014) 15(8):509–24. doi: 10.1038/nrm3838

3. Lim LP, Lau NC, Garrett-Engele P, Grimson A, Schelter J M, Castle J, et al. Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs. *Nature* (2005) 433(7027):769–73. doi: 10.1038/nature03315

4. Guo H, Pu M, Tai Y, Chen Y, Lu H, Qiao J, et al. Nuclear miR-30b-5p suppresses TFEB-mediated lysosomal biogenesis and autophagy. *Cell Death differentiation* (2021) 28(1):320–36. doi: 10.1038/s41418-020-0602-4

5. Ameres SL, Zamore PD. Diversifying microRNA sequence and function. Nat Rev Mol Cell Biol (2013) 14(8):475-88. doi: 10.1038/nrm3611

6. Friedman RC, Farh KK, Burge CB, et al. Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res* (2009) 19(1):92–105. doi: 10.1101/gr.082701.108

7. Sayed D, Abdellatif M. MicroRNAs in development and disease. *Physiol Rev* (2011) 91(3):827-87. doi: 10.1152/physrev.00006.2010

 Lee Y, Jeon K, Lee JT, Kim S, Kim V N. MicroRNA maturation: stepwise processing and subcellular localization. *EMBO J* (2002) 21(17):4663–70. doi: 10.1093/emboj/cdf476

9. Lee Y, Ahn C, Han J, Choi H, Kim J, Yim J, et al. The nuclear RNase III Drosha initiates microRNA processing. *Nature* (2003) 425(6956):415–9. doi: 10.1038/ nature01957

10. Hutvágner G, McLachlan J, Pasquinelli AE, Bálint E, Tuschl T, Zamore PD. A cellular function for the RNA-interference enzyme Dicer in the maturation of the Let-7 small temporal RNA. *Sci (New York NY)* (2001) 293(5531):834–8. doi: 10.1126/science.1062961

11. Bernstein E, Caudy AA, Hammond SM, Hannon GJ. Role for a bidentate ribonuclease in the initiation step of RNA interference. *Nature* (2001) 409(6818):363–6. doi: 10.1038/35053110

12. Reinhart BJ, Slack FJ, Basson M, Pasquinelli AE, Bettinger JC, Rougvie AE, et al. The 21-nucleotide Let-7 RNA regulates developmental timing in Caenorhabditis elegans. *Nature* (2000) 403(6772):901–6. doi: 10.1038/35002607

13. Roush S, Slack FJ. The Let-7 family of microRNAs. Trends Cell Biol (2008) 18 (10):505-16. doi: 10.1016/j.tcb.2008.07.007

14. Lee H, Han S, Kwon CS, Lee D. Biogenesis and regulation of the Let-7 miRNAs and their functional implications. *Protein Cell* (2016) 7(2):100–13. doi: 10.1007/s13238-015-0212-y

15. Johnson CD, Esquela-Kerscher A, Stefani G, Byrom M, Kelnar K, Ovcharenko D, et al. The Let-7 microRNA represses cell proliferation pathways in human cells. *Cancer Res* (2007) 67(16):7713–22. doi: 10.1158/0008-5472.CAN-07-1083

16. Johnson SM, Grosshans H, Shingara J, Byrom M, Jarvis R, Cheng A, et al. RAS is regulated by the Let-7 microRNA family. *Cell* (2005) 120(5):635–47. doi: 10.1016/j.cell.2005.01.014

17. Wang T, Wang G, Hao D, Liu X, Wang D, Ning N, et al. Aberrant regulation of the LIN28A/LIN28B and Let-7 loop in human Malignant tumors and its effects on the hallmarks of cancer. *Mol Cancer* (2015) 14:125. doi: 10.1186/s12943-015-0402-5

18. Chirshev E, Oberg KC, Ioffe YJ, Unternaehrer JJ. Let-7 as biomarker, prognostic indicator, and therapy for precision medicine in cancer. *Clin Trans Med* (2019) 8(1):24. doi: 10.1186/s40169-019-0240-y

19. Chirshev E, Suzuki T, Wang H, Nguyen A, Hojo N, Sanderman L, et al. Let-7i reduces aggressive phenotype and induces BRCAness in ovarian cancer cells. *Cancers* (2021) 13(18):4617. doi: 10.3390/cancers13184617

20. Yang HD, Kim HS, Kim SY, Na M J, Yang G, Eun J W, et al. HDAC6 suppresses let-7i-5p to elicit TSP1/CD47-mediated anti-tumorigenesis and phagocytosis of hepatocellular carcinoma. *Hepatol (Baltimore Md)* (2019) 70(4):1262–79. doi: 10.1002/hep.30657

21. Liu Y, Hu X, Hu L, Xu C, Liang X. Let-7i-5p enhances cell proliferation, migration and invasion of ccRCC by targeting HABP4. *BMC Urol* (2021) 21(1):49. doi: 10.1186/s12894-021-00820-9

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

22. Alkafaji HA, Raji A, Rahman HS, Zekiy A O, Adili A, Jalili M, et al. Upregulation of KISS1 as a novel target of Let-7i in melanoma serves as a potential suppressor of migration and proliferation in *vitro. J Cell Mol Med* (2021) 25(14):6864– 73. doi: 10.1111/jcmm.16695

23. Zhang GM, Long XH, Liu JM, Zhu L B, Chen X Y, Huang S H, et al. Let-7i inhibits the Malignant phenotype of osteosarcoma cells by targeting Aurora-B. *Mol Med Rep* (2015) 12(3):3543-8. doi: 10.3892/mmr.2015.3798

24. Shi Y, Duan Z, Zhang X, Zhang X, Wang G, Li F. Down-regulation of the Let-7i facilitates gastric cancer invasion and metastasis by targeting COL1A1. *Protein Cell* (2019) 10(2):143–8. doi: 10.1007/s13238-018-0550-7

25. Wu K, Yang Y, Zhao J, Zhao S. BAG3-mediated miRNA Let-7g and Let-7i inhibit proliferation and enhance apoptosis of human esophageal carcinoma cells by targeting the drug transporter ABCC10. *Cancer Lett* (2016) 371(1):125–33. doi: 10.1016/j.canlet.2015.11.031

26. Song J, Wang L, Ma Q, Yang Y, Yang Z, Wang B, et al. Let-7i-5p inhibits the proliferation and metastasis of colon cancer cells by targeting kallikrein-related peptidase 6. Oncol Rep (2018) 40(3):1459-66. doi: 10.3892/or.2018.6577

27. Tu F, Li M, Chen Y, Chu H, Wang S, Hai L, et al. Let-7i-3p inhibits the cell cycle, proliferation, invasion, and migration of colorectal cancer cells *via* downregulating CCND1. *Open Med (Warsaw Poland)* (2022) 17(1):1019–30. doi: 10.1515/med-2022-0499

28. Sun X, Xue H, Xiong Y, Yu R, Gao X, Qian M, et al. GALE promotes the proliferation and migration of glioblastoma cells and is regulated by miR-let-7i-5p. *Cancer Manage Res* (2019) 11:10539-54. doi: 10.2147/CMAR.S221585

29. Elkhadragy L, Chen M, Miller K, Yang MH, Long W. A regulatory BMI1/Let-7i/ ERK3 pathway controls the motility of head and neck cancer cells. *Mol Oncol* (2017) 11 (2):194–207. doi: 10.1002/1878-0261.12021

30. van Steensel B. Chromatin: constructing the big picture. $E\!MBO~J$ (2011) 30 (10):1885–95. doi: 10.1038/emboj.2011.135

31. Xhabija B, Kidder BL. KDM5B is a master regulator of the H3K4-methylome in stem cells, development and cancer. *Semin Cancer Biol* (2019) 57:79–85. doi: 10.1016/j.semcancer.2018.11.001

32. Yang Y, Li W, Wei B, Wu K, Liu D, Zhu D, et al. MicroRNA let-7i inhibits histone lysine demethylase KDM5B to halt esophageal cancer progression. *Mol Ther Nucleic Acids* (2020) 22:846–61. doi: 10.1016/j.omtn.2020.09.012

33. Tan DS, Holzner M, Weng M, Srivastava Y, Jauch R. SOX17 in cellular reprogramming and cancer. *Semin Cancer Biol* (2020) 67(Pt 1):65–73. doi: 10.1016/j.semcancer.2019.08.008

34. Liu J, Feng Y, Zeng X, He M, Gong Y, Liu Y. Extracellular vesicles-encapsulated Let-7i shed from bone mesenchymal stem cells suppress lung cancer via KDM3A/ DCLK1/FXYD3 axis. J Cell Mol Med (2021) 25(4):1911–26. doi: 10.1111/jcmm.15866

35. Liu Y, Wang D, Zhou M, Chen H, Wang H, Min J, et al. The KRAS/Lin28B axis maintains stemness of pancreatic cancer cells *via* the Let-7i/TET3 pathway. *Mol Oncol* (2021) 15(1):262–78. doi: 10.1002/1878-0261.12836

36. Subramanian M, Francis P, Bilke S, Li X L, Hara T, Lu X, et al. A mutant p53/Let-7i-axis-regulated gene network drives cell migration, invasion and metastasis. *Oncogene* (2015) 34(9):1094–104. doi: 10.1038/onc.2014.46

37. Wang L, Zhang J, Xia M, Liu C, Zu X, Zhong J. High mobility group A1 (HMGA1): structure, biological function, and therapeutic potential. *Int J Biol Sci* (2022) 18(11):4414–31. doi: 10.7150/ijbs.72952

38. Qin MM, Chai X, Huang HB, Feng G, Li X N, Zhang J, et al. Let-7i inhibits proliferation and migration of bladder cancer cells by targeting HMGA1. *BMC Urol* (2019) 19(1):53. doi: 10.1186/s12894-019-0485-1

39. Chhabra R. Let-7i-5p, miR-181a-2-3p and EGF/PI3K/SOX2 axis coordinate to maintain cancer stem cell population in cervical cancer. *Sci Rep* (2018) 8(1):7840. doi: 10.1038/s41598-018-26292-w

40. Chien CS, Wang ML, Chu PY, Chang YL, Liu WH, Yu CC, et al. Lin28B/let-7 regulates expression of oct4 and sox2 and reprograms oral squamous cell carcinoma

cells to a stem-like state. Cancer Res (2015) 75(12):2553-65. doi: 10.1158/0008-5472.CAN-14-2215

41. Lee YS, Dutta A. The tumor suppressor microRNA Let-7 represses the HMGA2 oncogene. *Genes Dev* (2007) 21(9):1025–30. doi: 10.1101/gad.1540407

42. Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. *Nat Rev Mol Cell Biol* (2014) 15(3):178–96. doi: 10.1038/nrm3758

43. Gupta PB, Onder TT, Jiang G, Tao K, Kuperwasser C, Weinberg RA, et al. Identification of selective inhibitors of cancer stem cells by high-throughput screening. *Cell* (2009) 138(4):645–59. doi: 10.1016/j.cell.2009.06.034

44. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, et al. The epithelialmesenchymal transition generates cells with properties of stem cells. *Cell* (2008) 133 (4):704–15. doi: 10.1016/j.cell.2008.03.027

45. Shi Z, To SKY, Zhang S, Deng S, Artemenko M, Zhang M, et al. Hypoxiainduced Nur77 activates PI3K/Akt signaling *via* suppression of Dicer/Let-7i-5p to induce epithelial-to-mesenchymal transition. *Theranostics* (2021) 11(7):3376–91. doi: 10.7150/thno.52190

46. Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, et al. Direct regulation of TWIST by HIF-1alpha promotes metastasis. *Nat Cell Biol* (2008) 10 (3):295–305. doi: 10.1038/ncb1691

47. Furlong EE, Andersen EC, Null B, White KP, Scott MP. Patterns of gene expression during Drosophila mesoderm development. *Sci (New York NY)* (2001) 293 (5535):1629–33. doi: 10.1126/science.1062660

48. Yang J, Mani SA, Donaher JL, Ramaswamy S, Itzykson RA, Come C, et al. Twist, a master regulator of morphogenesis, plays an essential role in tumor metastasis. *Cell* (2004) 117(7):927–39. doi: 10.1016/j.cell.2004.06.006

49. Yang MH, Hsu DS, Wang HW, Wang HJ, Lan HY, Yang WH, et al. Bmil is essential in Twist1-induced epithelial-mesenchymal transition. *Nat Cell Biol* (2010) 12 (10):982–92. doi: 10.1038/ncb2099

50. Yang WH, Lan HY, Huang CH, Tai SK, Tzeng CH, Kao SY, et al. RAC1 activation mediates Twist1-induced cancer cell migration. *Nat Cell Biol* (2012) 14 (4):366–74. doi: 10.1038/ncb2455

51. Tian Y, Hao S, Ye M, Zhang A, Nan Y, Wang G, et al. MicroRNAs Let-7b/i suppress human glioma cell invasion and migration by targeting IKBKE directly. *Biochem Biophys Res Commun* (2015) 458(2):307–12. doi: 10.1016/j.bbrc.2015.01.105

52. Li B, Lu W, Qu J, Zhang Y, Wan X. DICER1 regulates endometrial carcinoma invasion *via* histone acetylation and methylation. *J Cancer* (2017) 8(6):933–9. doi: 10.7150/jca.17435

53. Yang WH, Lan HY, Tai SK, Yang MH. Repression of bone morphogenetic protein 4 by Let-7i attenuates mesenchymal migration of head and neck cancer cells. *Biochem Biophys Res Commun* (2013) 433(1):24–30. doi: 10.1016/j.bbrc.2013.02.076

54. Xu R, Rai A, Chen M, Suwakulsiri W, Greening DW, Simpson RJ, et al. Extracellular vesicles in cancer - implications for future improvements in cancer care. *Nat Rev Clin Oncol* (2018) 15(10):617–38. doi: 10.1038/s41571-018-0036-9

55. Luga V, Zhang L, Viloria-Petit AM, Ogunjimi AA, Inanlou MR, Chiu E, et al. Exosomes mediate stromal mobilization of autocrine Wnt-PCP signaling in breast cancer cell migration. *Cell* (2012) 151(7):1542–56. doi: 10.1016/j.cell.2012.11.024

56. Fang T, Lv H, Lv G, Li T, Wang C, Han Q, et al. Tumor-derived exosomal miR-1247-3p induces cancer-associated fibroblast activation to foster lung metastasis of liver cancer. *Nat Commun* (2018) 9(1):191. doi: 10.1038/s41467-017-02583-0

57. Khani AT, Sharifzad F, Mardpour S, Hassan ZM, Ebrahimi M. Tumor extracellular vesicles loaded with exogenous Let-7i and miR-142 can modulate both immune response and tumor microenvironment to initiate a powerful anti-tumor response. *Cancer Lett* (2021) 501:200–9. doi: 10.1016/j.canlet.2020.11.014

58. Xiao D, Barry S, Kmetz D, Egger M, Pan J, Rai SN, et al. Melanoma cell-derived exosomes promote epithelial-mesenchymal transition in primary melanocytes through paracrine/autocrine signaling in the tumor microenvironment. *Cancer Lett* (2016) 376 (2):318–27. doi: 10.1016/j.canlet.2016.03.050

59. You B, Zhang P, Gu M, Yin H, Fan Y, Yao H, et al. Let-7i-5p promotes a Malignant phenotype in nasopharyngeal carcinoma *via* inhibiting tumor-suppressive autophagy. *Cancer Lett* (2022) 531:14–26. doi: 10.1016/j.canlet.2022.01.019

60. Saito Â, Souza EE, Costa FC, Meirelles GV, Gonçalves KA, Santos MT, et al. Human regulatory protein ki-1/57 is a target of SUMOylation and affects PML nuclear body formation. J Proteome Res (2017) 16(9):3147–57. doi: 10.1021/acs.jproteome.7b00001

61. Fawzy IO, Hamza MT, Hosny KA, Esmat G, Abdelaziz AI. Abrogating the interplay between IGF2BP1, 2 and 3 and IGF1R by Let-7i arrests hepatocellular carcinoma growth. *Growth factors (Chur Switzerland)* (2016) 34(1-2):42–50. doi: 10.3109/08977194.2016.1169532

62. Wu L, Wang Q, Yao J, Jiang H, Xiao C, Wu F. MicroRNA Let-7g and Let-7i inhibit hepatoma cell growth concurrently *via* downregulation of the anti-apoptotic protein B-cell lymphoma-extra large. *Oncol Lett* (2015) 9(1):213–8. doi: 10.3892/ol.2014.2706

63. Abdalla AME, Xiao L, Ullah MW, Yu M, Ouyang C, Yang G. Current challenges of cancer anti-angiogenic therapy and the promise of nanotherapeutics. *Theranostics* (2018) 8(2):533–48. doi: 10.7150/thno.21674

64. Kaur J, Debnath J. Autophagy at the crossroads of catabolism and anabolism. Nat Rev Mol Cell Biol (2015) 16(8):461–72. doi: 10.1038/nrm4024

65. Amaravadi R, Kimmelman AC, White E. Recent insights into the function of autophagy in cancer. *Genes Dev* (2016) 30(17):1913–30. doi: 10.1101/gad.287524.116

66. Levy JMM, Towers CG, Thorburn A. Targeting autophagy in cancer. Nat Rev Cancer (2017) 17(9):528–42. doi: 10.1038/nrc.2017.53

67. Yin S, Jin W, Qiu Y, Fu L, Wang T, Yu H. Solamargine induces hepatocellular carcinoma cell apoptosis and autophagy *via* inhibiting LIF/miR-192-5p/CYR61/Akt signaling pathways and eliciting immunostimulatory tumor microenvironment. *J Hematol Oncol* (2022) 15(1):32. doi: 10.1186/s13045-022-01248-w

68. Yu T, Guo F, Yu Y, Sun T, Ma D, Han J, et al. Fusobacterium nucleatum promotes chemoresistance to colorectal cancer by modulating autophagy. *Cell* (2017) 170(3):548-63.e16. doi: 10.1016/j.cell.2017.07.008

69. Li C, Chen L, Song W, Peng B, Zhu J, Fang L. DICER activates autophagy and promotes cisplatin resistance in non-small cell lung cancer by binding with Let-7i-5p. *Acta histochemica* (2021) 123(7):151788. doi: 10.1016/j.acthis.2021.151788

70. Dranoff G. Cytokines in cancer pathogenesis and cancer therapy. *Nat Rev Cancer* (2004) 4(1):11–22. doi: 10.1038/nrc1252

71. Iwasaki A, Medzhitov R. Control of adaptive immunity by the innate immune system. *Nat Immunol* (2015) 16(4):343–53. doi: 10.1038/ni.3123

72. Jutras I, Desjardins M. Phagocytosis: at the crossroads of innate and adaptive immunity. *Annu Rev Cell Dev Biol* (2005) 21:511-27. doi: 10.1146/annurev.cellbio.20.010403.102755

73. Chao MP, Majeti R, Weissman IL. Programmed cell removal: a new obstacle in the road to developing cancer. *Nat Rev Cancer* (2011) 12(1):58–67. doi: 10.1038/nrc3171

74. Logtenberg MEW, Scheeren FA, Schumacher TN. The CD47-SIRPα Immune checkpoint. *Immunity* (2020) 52(5):742–52. doi: 10.1016/j.immuni.2020.04.011

75. Theocharis AD, Skandalis SS, Tzanakakis GN, Karamanos NK. Proteoglycans in health and disease: novel roles for proteoglycans in Malignancy and their pharmacological targeting. *FEBS J* (2010) 277(19):3904–23. doi: 10.1111/j.1742-4658.2010.07800.x

76. Sofeu Feugaing DD, Götte M, Viola M. More than matrix: the multifaceted role of decorin in cancer. *Eur J Cell Biol* (2013) 92(1):1–11. doi: 10.1016/ j.ejcb.2012.08.004

77. Yerukala Sathipati S, Tsai MJ, Shukla SK, Ho SY. Artificial intelligence-driven pan-cancer analysis reveals miRNA signatures for cancer stage prediction. *HGG Adv* (2023) 4(3):100190. doi: 10.1016/j.xhgg.2023.100190

78. Li L, Chen J, Chen X, Tang J, Guo H, Wang X, et al. Serum miRNAs as predictive and preventive biomarker for pre-clinical hepatocellular carcinoma. *Cancer Lett* (2016) 373(2):234–40. doi: 10.1016/j.canlet.2016.01.028

79. Cochetti G, Poli G, Guelfi G, Boni A, Egidi MG, Mearini E. Different levels of serum microRNAs in prostate cancer and benign prostatic hyperplasia: evaluation of potential diagnostic and prognostic role. *OncoTargets Ther* (2016) 9:7545–53. doi: 10.2147/OTT.S119027

80. Huang J, Wu J, Li Y, Li X, Yang T, Yang Q, et al. Deregulation of serum microRNA expression is associated with cigarette smoking and lung cancer. *BioMed Res Int* (2014) 2014:364316. doi: 10.1155/2014/364316

81. Yerukala Sathipati S, Ho SY. Novel miRNA signature for predicting the stage of hepatocellular carcinoma. *Sci Rep* (2020) 10(1):14452. doi: 10.1038/s41598-020-71324-z

82. Liu J, Ni S. Association between genetic polymorphisms in the promoters of Let-7 and risk of cervical squamous cell carcinoma. *Gene* (2018) 642:256–60. doi: 10.1016/ j.gene.2017.11.038

83. Bocchini M, Tazzari M, Ravaioli S, Piccinini F, Foca F, Tebaldi M, et al. Circulating hsa-miR-5096 predicts (18)F-FDG PET/CT positivity and modulates somatostatin receptor 2 expression: a novel miR-based assay for pancreatic neuroendocrine tumors. *Front Oncol* (2023) 13:1136331. doi: 10.3389/ fonc.2023.1136331

84. Tsai YS, Huang CI, Tsai PC, Yeh ML, Huang CF, Hsieh MH, et al. Circulating Let-7 Family Members as Non-Invasive Biomarkers for Predicting Hepatocellular Carcinoma Risk after Antiviral Treatment among Chronic Hepatitis C Patients. *Cancers* (2022) 14(8):2023. doi: 10.3390/cancers14082023

85. Ding HX, Lv Z, Yuan Y, Xu Q. MiRNA polymorphisms and cancer prognosis: A systematic review and meta-analysis. *Front Oncol* (2018) 8:596. doi: 10.3389/ fonc.2018.00596

86. Du M, Giridhar KV, Tian Y, Tschannen MR, Zhu J, Huang CC, et al. Plasma exosomal miRNAs-based prognosis in metastatic kidney cancer. *Oncotarget* (2017) 8 (38):63703–14. doi: 10.18632/oncotarget.19476

87. Falzone L, Lupo G, La Rosa GRM, Crimi S, Anfuso CD, Salemi R, et al. Identification of novel microRNAs and their diagnostic and prognostic significance in oral cancer. *Cancers* (2019) 11(5):610. doi: 10.3390/cancers11050610

88. Yang N, Kaur S, Volinia S, Greshock J, Lassus H, Hasegawa K, et al. MicroRNA microarray identifies Let-7i as a novel biomarker and therapeutic target in human epithelial ovarian cancer. *Cancer Res* (2008) 68(24):10307–14. doi: 10.1158/0008-5472.CAN-08-1954

89. Coebergh van den Braak RRJ, Sieuwerts AM, Lalmahomed ZS, Smid M, Wilting SM, Bril SI, et al. Confirmation of a metastasis-specific microRNA signature in primary colon cancer. *Sci Rep* (2018) 8(1):5242. doi: 10.1038/s41598-018-22532-1

90. Liu K, Qian T, Tang L, Wang J, Yang H, Ren J. Decreased expression of microRNA Let-7i and its association with chemotherapeutic response in human gastric cancer. *World J Surg Oncol* (2012) 10:225. doi: 10.1186/1477-7819-10-225

91. Zhao L, Fong AHW, Liu N, Cho WCS. Molecular subtyping of nasopharyngeal carcinoma (NPC) and a microRNA-based prognostic model for distant metastasis. J Biomed Sci (2018) 25(1):16. doi: 10.1186/s12929-018-0417-5

92. Taghikhani A, Hassan ZM, Ebrahimi M, Moazzeni SM. microRNA modified tumor-derived exosomes as novel tools for maturation of dendritic cells. *J Cell Physiol* (2019) 234(6):9417–27. doi: 10.1002/jcp.27626

93. Zhou X, Ao X, Jia Z, Li Y, Kuang S, Du C, et al. Non-coding RNA in cancer drug resistance: Underlying mechanisms and clinical applications. *Front Oncol* (2022) 12:951864. doi: 10.3389/fonc.2022.951864

94. Sun J, Pan LM, Chen LB, Wang Y. LncRNA XIST promotes human lung adenocarcinoma cells to cisplatin resistance via Let-7i/BAG-1 axis. Cell Cycle (Georgetown Tex) (2017) 16(21):2100-7. doi: 10.1080/15384101.2017.1361071

95. Ren YL, Zhang W. Propofol promotes apoptosis of colorectal cancer cells via alleviating the suppression of lncRNA HOXA11-AS on miRNA Let-7i. Biochem Cell Biol = Biochimie biologie cellulaire (2020) 98(2):90–8. doi: 10.1139/bcb-2018-0235

96. Liu N, Zhou C, Zhao J, Chen Y. Reversal of paclitaxel resistance in epithelial ovarian carcinoma cells by a MUC1 aptamer-Let-7i chimera. *Cancer Invest* (2012) 30 (8):577-82. doi: 10.3109/07357907.2012.707265