



## OPEN ACCESS

## EDITED BY

Xiaowei Qi,  
Army Medical University, China

## REVIEWED BY

Ruo Wang,  
Shanghai Jiao Tong University, China  
Rong Mu,  
CSL Parkville, Australia

## \*CORRESPONDENCE

Zuowei Zhao  
✉ dmuzhaozuowei@163.com  
Man Li  
✉ man\_li@dmu.edu.cn

†These authors have contributed  
equally to this work and share  
first authorship

RECEIVED 10 October 2023

ACCEPTED 04 January 2024

PUBLISHED 25 January 2024

## CITATION

Hu Y, Chen F, Sun S, Xv L, Wang X, Wang M,  
Zhao S, Zhao Z and Li M (2024) mTOR  
inhibitor introduce disitamab vedotin  
(RC48-ADC) rechallenge microtubule-  
chemotherapy resistance in HER2-low  
MBC patients with PI3K mutation.  
*Front. Oncol.* 14:1312634.  
doi: 10.3389/fonc.2024.1312634

## COPYRIGHT

© 2024 Hu, Chen, Sun, Xv, Wang, Wang, Zhao,  
Zhao and Li. This is an open-access article  
distributed under the terms of the [Creative  
Commons Attribution License \(CC BY\)](#). The  
use, distribution or reproduction in other  
forums is permitted, provided the original  
author(s) and the copyright owner(s) are  
credited and that the original publication in  
this journal is cited, in accordance with  
accepted academic practice. No use,  
distribution or reproduction is permitted  
which does not comply with these terms.

# mTOR inhibitor introduce disitamab vedotin (RC48-ADC) rechallenge microtubule- chemotherapy resistance in HER2-low MBC patients with PI3K mutation

Ye Hu<sup>†</sup>, Fengxi Chen<sup>†</sup>, Siwen Sun<sup>†</sup>, Lingzhi Xv, Xueqing Wang,  
Meiling Wang, Shanshan Zhao, Zuowei Zhao\* and Man Li\*

Department of Oncology & Breast Surgery, The Second Hospital of Dalian Medical University,  
Dalian, China

This study aimed to explore the efficacy and potential mechanisms of rechallenge therapy with microtubule-targeting agents (MTAs) in patients with HER2-low metastatic breast cancer (MBC). We performed a systematic review to investigate the rechallenge treatment concept in the field of HER2-low MBC treatment and utilized a series of cases identified in the literature to illustrate the concept. Here we reported two clinical cases of HER2-low MBC patients whose disease progressed after prior treatment with MTAs such as docetaxel and vincristine. When rechallenged with disitamab vedotin ((RC48-antibody-drug conjugate (ADC), a monomethyl auristatin (MMAE) MTA)), both patients achieved a partial response and the final progression-free survival (PFS) was 13.5 and 9 months, respectively. Genomic profiling detected a PIK3CA H1047R mutation in the patients. The patients were treated with everolimus before being rechallenged with RC48, which may lead to a better response. This study further summarizes and analyzes the potential mechanism of the PI3K-AKT signaling pathway in MTA resistance and reveals that the PIK3CA H1047R mutation may be a potential molecular marker for the efficacy prediction of mTOR inhibitors, providing new insights and potential therapeutic strategies for the application of MTAs to MBC patients.

## KEYWORDS

HER2-low metastatic breast cancer, disitamab vedotin (RC48), microtubule targeting agents (MTAs), rechallenge, mTOR inhibitor, PIK3CA mutation

## 1 Introduction

Breast cancer (BC) is the most commonly diagnosed malignancy and has the second highest mortality rate globally among all cancers in women (1). Compared with other tumors, the slowing decline in mortality and the rising incidence of breast cancers are concerning (2). For metastatic breast cancer (MBC), therapeutic goals are prolonging life and symptom palliation. MBC remains incurable in virtually all affected patients (3), and a major contributor to this is the abrogation of the efficacy of chemotherapy owing to the emergence of drug resistance (4, 5). Consequently, additional targeted cancer control interventions are needed. Approximately 60% of human epidermal growth factor receptor 2 (HER2)-negative MBC express low levels of HER2 (HER2-low), defined as a score of 1+ on immunohistochemical (IHC) analysis or as an IHC score of 2+ with a negative result of fluorescence *in situ* hybridization (FISH). These patients have limited treatment options after progression during primary therapy and most commonly receive single-agent palliative chemotherapy. Multiple studies have shown that the objective response rate (ORR) is only 6%–17%, and the median PFS is only 1.6–2.8 months in patients with HER2-negative (including HER2 0, HER2 1+, and HER2 2+) MBC who received more than three lines of chemotherapy (6, 7).

MTAs, such as taxanes, have been a mainstay of BC therapy for decades. However, in patients who have previously used taxanes, the prognosis when reusing taxanes in the first line of relapse or metastasis within 2 years is poor. Furthermore, there are clinical and resistance issues that limit the efficacy of these drugs and pose a challenge to improving patient outcomes. In addition to switching to new treatments, rechallenge has attracted widespread attention. Rechallenge therapy is the reintroduction of the same therapy to a patient who has developed resistance after an interval of treatment (8).

Recent advances in ADC agents have led to the widespread clinical use of drugs that target cancer cells via specific antigens (e.g., HER2 (9–11)). The effectiveness of ADCs in prolonging the survival of MTA-resistant patients has attracted our attention. Trastuzumab deruxtecan (also known as T-DXd and DS-8201) is an ADC that has achieved great success in the treatment of HER2-positive BC. The DESTINY-Breast03 trial showed the superiority of DS-8201 in reducing the risk of progression or death in patients with HER2-positive MBC who had been previously treated with trastuzumab and taxane (12, 13). One of the reasons for the success is that DS-8201 consists of a topoisomerase I inhibitor as a payload, which does not develop cross-resistance with MTAs and has an effective bystander effect. RC48 consists of MMAE as a payload (14). MMAE exhibits a marked mitotic inhibitory effect by inhibiting tubulin polymerization (15, 16). Pre-clinical studies and two clinical studies revealed that RC48 demonstrated consistent efficacy in HER2-positive and HER2-low advanced BC (17–19). In 2021, the American Society of Clinical Oncology (ASCO) announced that RC48 can achieve good efficacy in patients with HER2-positive or HER2-low advanced BC who had received multiline treatments, with a median PFS of 4.0 months in the HER2-positive and 5.7 months in the HER2-low (20). Further studies are needed to

explore how MMAE-ADC agents are more effective at prolonging the prognosis of patients with HER2-low advanced BC compared with those with HER2-positive advanced BC.

As mentioned above, the role of rechallenge therapy in third-line or fourth-line settings is not clear, but rechallenge could be a possibility for patients who do not have any other valid treatment options (8, 21). Treatment may cause epigenetic alterations that induce chemoresistance, but altered therapy or intermittent therapy may restore the epigenetic features. On this basis, our study explored the rechallenge treatment concept in the field of HER2-negative MBC treatment and used a series of cases to illustrate. Furthermore, we suggest a new strategy that rapamycin (mTOR) inhibitors can be used as an intermittent therapy to prolong PFS in patients with HER2-negative MBC who receive MMAE-ADC agents after MTA progression, and the PIK3CA H1047R mutation may be a potential molecular marker.

## 2 Materials and methods

### 2.1 Search strategy

An electronic search was conducted using PubMed. The following search terms were used: breast cancer, HER2, rechallenge, microtubule-targeting agent, mTOR inhibitors, PI3K (phosphoinositide 3-kinase)-AKT pathway, PIK3CA mutation. The titles and abstracts of all remaining citations were reviewed, and irrelevant citations were discarded. The full text of potentially relevant studies was retrieved and evaluated. A manual search of the reference list of relevant reports was performed to identify any relevant studies missed by the search strategy. The evaluation of efficacy was evaluated using the Response Evaluation Criteria in Solid Tumors (RECIST) guidelines (version 1.1).

## 3 Results

We reported a series of cases of MBC patients in which RC48 was reintroduced after the progression of multiline standard-of-care microtubule inhibitors as interventional therapy with mTOR inhibitors and achieved good PFS (Figure 1). Additionally, five cases of MBC patients in MTA rechallenge therapy had undergone next-generation sequencing (NGS), and the mutational signature profiling is shown in Figure 2.

### 3.1 Case series: five failed cases in HER2-low MBC

Five patients (cases 3–7), who were diagnosed with HER2-low MBC, received multiline chemotherapy treatment, including taxanes, vinorelbine, etc. However, when these patients were given RC48 directly after MTA treatment, the disease developed within 1 or 2 months (Figure 1).

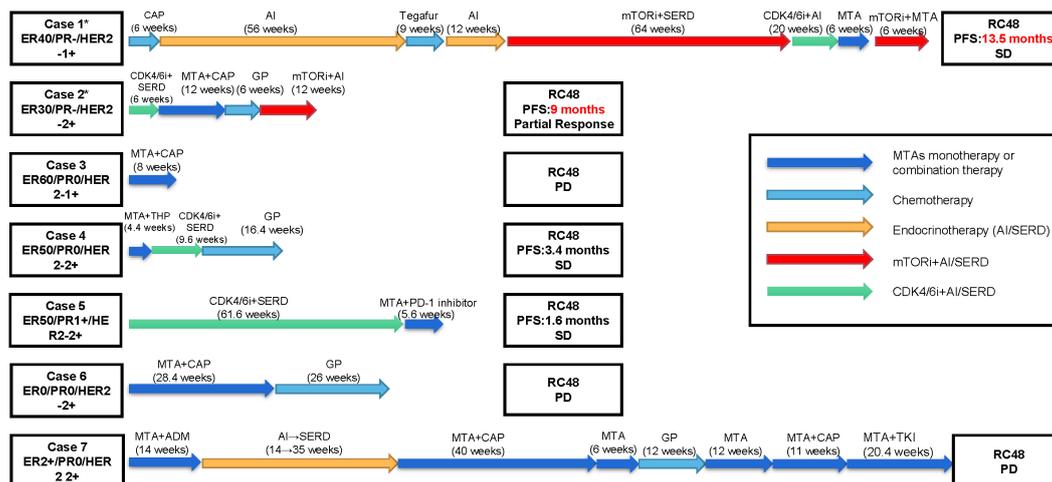


FIGURE 1

Patients' overview. Clinical timelines for the seven cases of metastatic breast cancer patients in which RC48 was reintroduced after the progression of multiline standard-of-care microtubule inhibitors. Patients' histories are shown from advanced first-line treatment until RC48 reintroduction. The arrows represent distinct therapies and durations. The asterisk represents patients carrying PIK3CA H1047R mutation. ER, estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor receptor 2; SD, stable disease; PD, progressive disease; CAP, capecitabine; GP, gemcitabine + cisplatin/carboplatin; THP, docetaxel + trastuzumab + pertuzumab; ADM, doxorubicin; MTA, microtubule-targeting agent; AI, aromatase inhibitor; SERD, selective estrogen receptor downregulator; mTORi, mammalian target of rapamycin inhibitors; CDK4/6i, cyclin 4- and 6-dependent kinase inhibitors; PD-1 inhibitor, programmed cell death 1 inhibitor; TKI, tyrosine kinase inhibitor.

### 3.2 Case 1: a 58-year-old woman with HER2-low MBC

In May 2011, the patient underwent radical mastectomy for left BC with a pathologic diagnosis of invasive ductal carcinoma of the left breast (pT2N2M0). The IHC analysis confirmed that the patient was estrogen receptor (ER)-negative, progesterone receptor (PR)-negative, and HER2-0. After the operation, adjuvant chemotherapy and radiotherapy and adjuvant endocrine standard therapy were taken in 5 years. However, the patient developed bone metastasis in 2016, which was diagnosed as ER 40%, PR-negative, and HER2 1+. After local radiotherapy and a multiline line of endocrine therapy (ET), the disease was poorly controlled, and a liver malignancy was diagnosed by PET/CT in 2018. Thus, the patient received two cycles of vinorelbine (MTA) chemotherapy. However, the tumor markers increased continuously, accompanied by enlargement of the liver lesions (Figure 3A). After the failure of previous treatments, a genomic analysis was performed and the PIK3CA H1047R mutation was identified (Figure 2). The patient was started on everolimus plus eribulin/utidelson for 9–10-line therapy. Subsequently, the patient received RC48 for 15 cycles (2 mg/kg, ivgtt, 14 days/cycle), after which her condition stabilized and the PFS reached 13.5 months. This case suggested that mTOR inhibitors as intermittent therapy can reverse the resistance of microtubule inhibitors.

### 3.3 Case 2: a 58-year-old woman with HER2-low MBC

In 2018, the patient received a modified radical mastectomy for left BC. At the time of initial diagnosis, the postoperative pathology was invasive BC (pT1N0M0). The IHC analysis found that the cancer was ER 70%, PR 30%, and HER2-negative. After adjuvant chemotherapy and adjuvant ET, sternal metastasis occurred in February 2021, which was identified as BC metastasis by bone biopsy, and the IHC analysis confirmed that the cancer was ER 30%, PR-negative, HER2 2+, FISH no amplification, and Ki67 30%. Following local radiotherapy and multiline ET, the disease remained poorly controlled and was confirmed as a liver malignancy by PET/CT; consequently, the patient received two cycles of vinorelbine chemotherapy. The patient received advanced first-line ET with abemaciclib plus fulvestrant. Liver metastasis developed after two cycles, and the curative effect was evaluated as progressive disease (PD).

Subsequently, docetaxel (MTA) combined with capecitabine chemotherapy was administered until the liver lesion progressed after four cycles. Genomic profiling was performed and detected a PIK3CA H1047R mutation (Figure 2). The liver metastases continued to deteriorate after two cycles of gemcitabine combined with a carboplatin regimen and 1 month of ET with exemestane plus everolimus. However, when the patient received RC48 for four cycles, the intrahepatic lesions significantly reduced in number and size, achieving a partial response (Figure 3B). The final PFS was 9 months.

	Case 1	Case 2	Case 3	Case 4	Case 5
PIK3CA	H1047R	H1047R		E542K	
FGFR1	E545K E365K IGR-FGFR1 fusion				
FGFR2		M391L			
FGF19				E81*	
MAP2K1	E203K				
MEF2B	Splice				
PREX2	R577*				
ZNF703				H59D	
IGF1R		R40H			
IGF2	P98L				
JAK1	F413V				
KMT2C				T376Gfs*22	
KMT2A	S2562F				
RNF43	A182T				
DPYD		Q413K	V1024L		
TP53		N247I	R306*	S90Lfs*59	
PGR		R869C			
CYP2A6		E197Q			
CYP2C19					
ERBB2				R340Q	
XRCC2					V160L
APC			L629F		
GATA3				*445Yext*28	
CDK12				S363*	
CCN6			G171V		
CHD4				Q675*	
MYC				E257K	
ABCB1				E972Q	
CDH1		CDH1-IGR fusion			
CTCF				E492K	
ESR1				S341L	
FAT1				E2500Q	
FOXL2				E118Q	
RHBDF2				P347H	
SDHD				A61G	
SMARCB1				M2131I	
SRC				S190L	
TPMT				D221H	
NKX2-1		H349Y			
NF1					W2204*
BRAF					D594N
PTPN11					E76A
GRM3					A401D
PLK1					G81A
TOP2A					R740P
PDE11A			Q29*		
AURKA			R189W		
FANCD2			H1042Y		
NTRK2			G726Wfs*51		

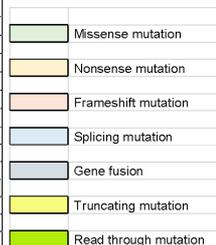


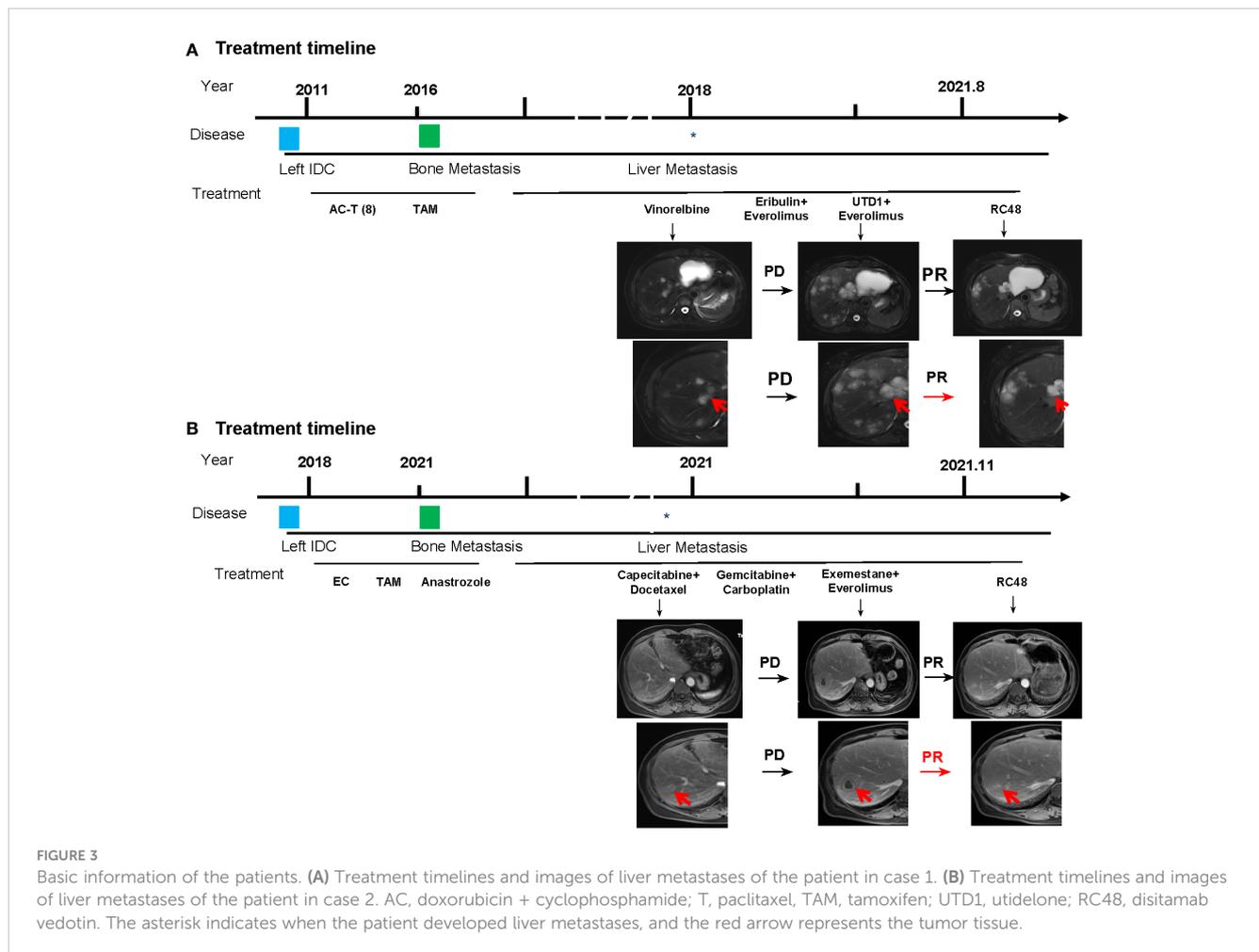
FIGURE 2  
Mutational profiling of breast cancer cases. Single-nucleotide variants are depicted across the five metastatic breast cancer patients. The asterisk indicates that this site mutates into a terminator so that the stop codon is encoded in advance, producing a truncated protein.

## 4 Discussion

In our study, the failed cases treated with MMAE-ADC agents after the progression of multiline MTA treatment indicated that the efficacy of treatment may be affected if the chemotherapy drug is the same as ADC-payload. This is mainly because they have the same anti-tumor effect, and the patients develop resistance after the progression of chemotherapy. In contrast to these cases, we identified two BC patients who received RC48 after MTA progression and achieved a better prognosis. These two patients had the PIK3CA H1047R mutation detected by genomic profiling and were treated with everolimus before starting the RC48 treatment. The PFS of these two patients treated with RC48 was 13.5 and 9 months, respectively. In addition, one patient achieved a continuous partial response after treatment with RC48. These two cases suggested that mTOR inhibitors as an interval treatment can promote the efficacy of MMAE-ADC agents after the progression of multiline MTA treatment in HER2-low BC. Meanwhile, this study provided new insights into the challenges faced by microtubule inhibitors, implying that mTOR inhibitors can be used as inducers to restore the therapeutic effects of microtubule inhibitors. We

hypothesize that mTOR inhibitors as inducers can effectively enhance the anti-tumor effect of MTAs in BC with HER2-low expression.

MTAs are widely used in clinical practice, and numerous studies have identified resistance to paclitaxel and docetaxel. The major structural subunit of microtubules is tubulin, which is composed of  $\alpha$ - and  $\beta$ -tubulin monomers that form a dimer and assemble onto the positive ends of the growing microtubule. In many cancer cell lines, MTA resistance results from alterations in microtubule dynamics and binding sites on the microtubules (22, 23). The mechanism of MTAs can be divided into three aspects: (1) disturb the spindle assembly: MTAs, such as taxol, impair the assembly/disassembly dynamic balance of microtubule, thereby activating the spindle assembly checkpoint (SAC) through unattached kinetochores (24); (2) influence on microtubule dynamics: The altered expression of microtubule proteins causes mitotic spindle abnormalities and cell cycle arrest, ultimately leading to genomic instability, which is considered a hallmark of a range of cancers (25–27). Analysis of clinical specimens has shown that, in many cancers, a high expression of several  $\beta$ -microtubulin isoforms is associated with aggressive clinical



manifestations, chemotherapy resistance, and poor patient prognosis in several cancers, including breast, colon, and renal cancers (28, 29); and (3) lead to mitotic arrest: The surface of the globular portion of microtubulin contains several pockets that act as intercalation sites for MTAs, thereby affecting the structure of microtubulin. Several studies have demonstrated that treatment with MTAs with this property leads to mitotic arrest and thus cell death (30, 31). The emergence of compensatory pathways leads to MAT resistance due to the alteration of some molecular features during tumor development. Restoring these resistance alterations will re-sensitize the cells to MTAs (32). In addition to our report, other studies have reported a relationship between the PI3K-AKT pathway and MTAs. This pathway is a vital oncogenic pathway and can induce tumor cell survival, proliferation, and metastasis (33–35). Several studies have demonstrated that low-dose PI3K-AKT inhibitors are effective in reversing paclitaxel resistance in different tumor types, including gastric, ovarian, lung, and prostate cancers (36–39). We further explored the mechanisms associated with these microtubule-targeted drug re-sensitization.

As we all know, tumor cells are often accompanied by abnormally active mitosis and disruption of spindle dynamics (40, 41). Many microtubules and related proteins consist of the basic structure of the mitotic spindle and are involved in the regulation of

mitotic spindle dynamics (42). MTAs inhibit cancer cell proliferation by disrupting the mitotic spindle (43, 44). It has been demonstrated that the activation of the PI3K-AKT signaling pathway is closely related to the stability of the microtubules and the mitotic spindle. Gris-Oliver and colleagues suggested that 64% of HER2-negative BC xenografts with resistance to eribulin (microtubule inhibitors) show PIK3CA, PIK3R1, or AKT1 mutation, indicating that PI3K pathway activation may induce resistance or early adaptation to microtubule inhibitors (45). According to the literature, we explored the potential molecular mechanisms by which mTOR inhibitors reverse MTA resistance by inhibiting the PI3K-AKT signaling pathway (Figure 4).

On the one hand, the interaction between PI3K-AKT mainly involves two metabolites: PIP2 (phosphatidylinositol-4,5-bisphosphate) and PIP3 (phosphatidylinositol-3,4,5-bisphosphate) (46, 47). PI3K is an intracellular phosphatidylinositol kinase. PI3K consists of a regulatory subunit (p85) and a catalytic subunit (p110) (48). When the ligand binds to the membrane receptor, the receptor activates p85 and recruits p110, which, in turn, catalyzes the production of PIP3 from PIP2 on the inner surface of the membrane (49–51). p110 $\alpha$  is activated at the initiation of mitosis, producing PIP3 in the mesocortex of metaphase cells to guarantee proper orientation of the spindle. Lipid phosphatases (e.g., PTEN)

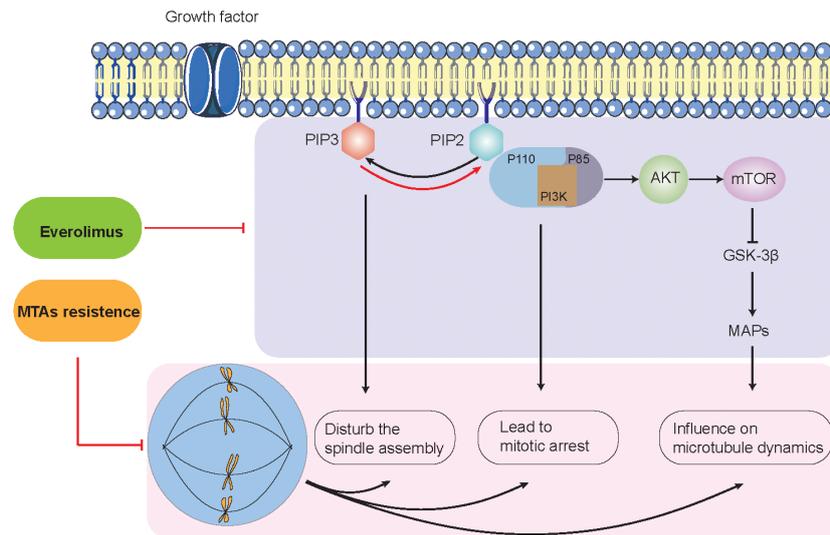


FIGURE 4  
Potential molecular mechanisms between PI3K/AKT signaling pathway and microtubule-targeting agent resistance.

can metabolize (dephosphorylate) PIP3 back to PIP2, thereby terminating the PI3K signaling pathway. In addition, PI3K-C2 $\alpha$ , a class II PI3K family member, interacts with the transformed acidic coiled-coil 3 (TACC3) and clathrin heavy chain (CHC) complexes to stabilize the kinetochore-microtubules formed by the spindle (52, 53). Due to the critical role of PI3K-AKT signaling in spindle formation, drug inhibitors such as LY-294002 and MK-2206 can lead to aberrant centrosome and mitotic spindle formation, thereby reversing MTA resistance and enhancing antitumor effects (52, 54, 55).

On the other hand, the PI3K-AKT-mTOR pathway has been implicated in the regulation of microtubule stability (35, 56–58). Onishi et al. showed that the addition of the pan-PI3K inhibitor LY-294002 destabilized microtubules in fibroblasts (59). Previous studies have shown that the localization of AKT to microtubules is important for sustaining AKT phosphorylation. Consistently, the introduction of a predominantly negative form of AKT into cells showed destabilized microtubules (57, 58). The regulation of microtubule dynamics by the PI3K-AKT pathway can be transduced by glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ). As previously mentioned, PI3K-AKT inhibits the activity of GSK-3 $\beta$  activity (60). The inhibition of PI3K-AKT signaling by LY-294002 induces the activation of GSK-3 $\beta$ , which, in turn, regulates a large repertoire of protein substrates, including microtubule-associated proteins (MAPs) (60). MAPs interact with tubulin dimers of the microtubule (e.g., MAP1, MAP2, MAP3, MAP4, and tau) (61). This interaction leads to microtubule stabilization and subsequent polymerization—for example, MAP4 decorates the microtubule network and participates in microtubule assembly and endosomal vesicle trafficking along microtubule tracks. Thapa et al. discovered that MAP4 is a binding partner of the p110 $\alpha$  catalytic subunit of PI3K $\alpha$  (62). The loss of MAP4 perturbs PI3K $\alpha$  recruitment along microtubule tracks and endosomes and disrupts PI3K $\alpha$  association with activated receptor complexes, which collectively contribute to

impaired PI3K $\alpha$  activation (62–64). A previous study revealed that LY-294002 did not induce cell death but resulted in the marked and selective enhancement of the induction of apoptosis by microtubule-destabilizing agents such as vincristine (60). The mechanism is that blockade of the PI3K-AKT pathway induces the activation of GSK-3 $\beta$ , which phosphorylates MAPs such as tau and thereby reduces their ability to bind and stabilize microtubules (64).

Previous studies suggested that cleaved-PARP may be involved in the action of the PI3K-AKT inhibitors in the paclitaxel-resistant cell line (65, 66). PARP is a family of enzymes of poly (ADP-ribose) polymerases. In addition to mediating DNA damage response pathways, PARP is crucial in regulating genome stability through transcription and regulating the expression of oncogenes and tumor suppressor genes (67, 68). In the cases in this study, the administration of the combination of everolimus and ET or chemotherapy did not improve PFS and disease progression continued. However, after everolimus treatment, patients with HER2-negative BC who received RC48 achieved an improvement in PFS. These results indicate that mTOR inhibitors may act as inducers that support MMAE-ADC agent rechallenge in patients with MTA-resistant HER2-negative BC. HER2 and insulin receptor substrate (IRS) stimulate the RAS-ERK-RSK and AKT-mTOR pathways. Activation of mTOR signaling phosphorylates S6 kinase 1 (S6K) and eukaryotic initiation factor 4E-binding protein 1 (4E-BP1), which promote cancer growth and metastasis (69). The use of mTOR inhibitors and PI3K inhibitors to overcome resistance to current HER2 therapies is an active area of research (70). These may be the reasons for the re-sensitization of MTAs by mTOR inhibitors, and we still need more basic and clinical trials to verify this.

PIK3CA mutations are present in approximately 20% of HER2-positive BCs (71, 72). In patients with advanced hormone receptors (HR)+/HER2-negative BC, 28% of PIK3CA mutations were

identified in circulating tumor DNA (73, 74). The results from the SAFIR02 trial showed that PIK3CA mutations were associated with chemoresistance and poorer overall survival (OS) in ER+ HER2-negative BC (75). PIK3CA was the most frequent mutation observed, associated with an increased kinase activity of the PI3K pathway. In BC, the most frequent PIK3CA mutations lead to protein residue changes at the following positions of exon mutations: E542K (76–78), E545K (77, 78) in exon 9 and H1047R (77, 78) in exon 20, comprising approximately 78% of all PIK3CA mutations observed in breast tumors (79). Numerous studies have demonstrated that these hotspot mutations prompt transformation and tumorigenicity by inducing enhanced PI3K function and activating downstream signaling in the AKT-mTOR pathway (75, 80), with robust *in vitro* and *in vivo* transformation phenotypes (81–84). Different mutations at the same residues had different phenotypic activities depending on their mutation frequency. It has been reported that a higher frequency mutation (H1047R) showed a stronger tumorigenic phenotype compared to mutations of E542K, E536K, and E545K (85–88). In recent years, the relationship between gene mutations and response to mTOR inhibitors has received particular attention. Consistently, experimental studies have demonstrated that BCs with PIK3CA mutations are more sensitive to everolimus, and the IC50 value of the mTOR inhibitor is lower for H1047R than for E542K or E545K (89). The PIK3CA H1047R mutation was also reported to confer sensitivity to everolimus in early-phase clinical trials in many types of cancers (90, 91). The potential underlying mechanism is that the PIK3CA H1047R mutation is a stronger driver of tumor development than other types of PIK3CA mutations (92, 93).

Therefore, patients with PIK3CA H1047R mutations were more sensitive to the mTOR inhibitor everolimus (94). Based on the previous studies, among MBC patients who had been treated with everolimus, patients with the PIK3CA H1047R mutation had longer PFS compared with patients with wild-type or other mutant forms of PIK3CA ( $p < 0.050$ ) (95). It is worth noting that PIK3CA H1047R was detected in two cases with a favorable prognosis that was mentioned above (Figure 2). Both of them used mTOR inhibitors before MTA rechallenge treatment and showed longer PFS.

In summary, different mutations in PIK3CA lead to different sensitivities of tumors to everolimus, and patients with PIK3CA H1047R mutations were more sensitive to the mTOR inhibitor everolimus. This may be because the PIK3CA H1047R mutation is a stronger driver of tumor development than other types of PIK3CA mutations (94). More importantly, relevant clinical studies suggested that PIK3CA H1047R mutations may be a potent biomarker of sensitivity to everolimus in MBC (95–97). The combination of alpelisib, a selective PIK3CA inhibitor, with mTOR inhibitors has also shown a synergistic efficacy in PIK3CA-mutated (H1047R) hepatocellular carcinoma (98). These results still need further exploration in clinical trials.

In our study, we found that patients had other mutations in addition to PIK3CA. However, they were not associated with mTOR inhibitors and MTAs—for example, mutations in the fibroblast growth factor receptor (FGFR) 1/2 gene were associated with FGFR kinase inhibitor resistance (99, 100). Dihydropyrimidine dehydrogenase genotyping is related to the sensitivity of

fluoropyrimidines (5-fluorouracil, capecitabine, and tegafur) (99, 101). Sensitivity to combined RAF kinases (BRAF) and mitogen-activated protein kinase kinase (MEK) treatments is associated with co-mutations of mitogen-activated protein kinase kinase 1 (MAP2K1) and BRAF (102). There are still some meaningful variations that are worth exploring. In addition, tumor progression is, in essence, an evolutionary process, and different adaptive changes in molecular characteristics during tumor development can enable tumors to progress through tumor pathologic stages or transformation barriers (40, 103). According to the theory of “evolutionary selection”, molecular pathways that are activated or inactivated during tumor evolution may play important adaptive roles in promoting tumorigenesis, proliferation, and metastasis. This evolutionary theory could explain the molecular mechanisms that emerged during tumor evolution in the patient in our case after mTOR inhibitor treatment, and restoration of these alterations would re-sensitize cells in HER2-low MBC to MTAs.

Two-thirds of BC patients express HR and lack HER2 overexpression and/or amplification (104, 105). For these patients, the combination of ET with cyclin-dependent kinase (CDK)4/6 inhibitors exhibited significant survival benefits and is now the gold standard for HR+/HER2-negative MBC (106, 107). The choice of drug for use in clinical practice should be based on the response to prior treatment before progression. Previous studies revealed that when the disease progressed after the administration of microtubule inhibitors, rechallenging with the same drugs seems to be inadvisable. Conversely, few clinical studies have evaluated the role of rechallenge in BC (108, 109). We summarized eight reports about rechallenges in BC (Table 1), which highlight the clinical importance of paying more attention to rechallenges. Toulmonde et al. demonstrated docetaxel rechallenge as a second, third, fourth, or more line of chemotherapy in the metastatic setting. Among the 33 patients with disease assessed according to RECIST guidelines, 14 (42.5%) had a partial response and 11 (33.5%) had a stable disease >6 weeks. Globally, 55 patients (76%) obtained a benefit from the treatment. The median time to progress and OS were 5.7 months (95% CI: 5.0–6.3) and 10.2 months (95% CI: 8.6–11.8). This retrospective analysis supported the pragmatic strategy to retreat patients with MBC with docetaxel (111). Meanwhile, some trials with trastuzumab and CDK inhibitors have demonstrated the potential of rechallenge in clinical applications (113, 114, 116). In recent years, there have been many studies on ADC drugs in BC with HER2-low-expression, including a number of clinical trials (Table 2).

However, most clinical trials are still recruiting and further research results are anticipated. RC48 is the representative MMAE-ADC agent for BC treatment, showing consistent efficacy in posterior line HER2-low-expressing MBC. Collectively, the role of rechallenge therapy in the treatment of MBC has not yet been established—whether a MMAE-ADC agent is feasible as a choice after MTA resistance rechallenge in BC. Further studies are needed to verify this strategy.

In conclusion, our study provides a new insight that mTOR inhibitors, as an intermittent treatment, may induce sensitivity to MMAE-ADC agents in HER2-low-expressing BC, especially after

TABLE 1 Prior reports about rechallenge in breast cancer.

Regimen	No. of patients	Results	Subtype	References
Tamoxifen	MCF7 vs. MCF7-TAM12.5	Metastatic potential decreased; proliferation and clonogenicity increased	p-AuroraA/B upregulated	(110)
Docetaxel rechallenge	72	55 patients (76%) obtained a benefit from the treatment	MBC	(111)
Adjuvant anthracyclines vs. anthracycline rechallenge	70	ORR: 38%; CB: 71%; mOS: 16.5	MBC	(112)
(Neo) adjuvant trastuzumab–trastuzumab rechallenge vs. lapatinib	101 vs. 27	No significance between two groups	HER2+	(113)
Lapatinib-resistant vs. trastuzumab Trastuzumab-resistant vs. lapatinib rechallenge	74 50	CB: 32%	HER2+	(114)
Pembrolizumab vs. atezolizumab rechallenge	1	Excellent response without drug-related adverse effects	TNBC	(115)
CDK inhibitors	6	1 CR 2 PFS, 6 months 1 PFS, 10 months	HER2-negative MBC	(116)
Anthracyclines and taxanes vs. eribulin rechallenge	22	No significance in OS between two groups	MBC	(117)

ORR, objective response rate; CB, clinical benefit; mOS, median overall survival; PFS, progression-free survival.

TABLE 2 Clinical trials of antibody–drug conjugate (ADC) drugs in breast cancer with HER2-low expression.

Target	ADC drugs	Tumors	Status	Combination	Trial phase	Results	NCT
HER2	DS8201	Third-line advanced HER2-low BC, post-chemo	Active, not recruiting	None	Phase III	PFS: 10.1 m OS: 23.9 m mPFS: 9.9 m	NCT03734029 (118)
		Third-line advanced HER2-low BC, chemo-naive	Recruiting	None	Phase III		NCT04494425
	SYD985	Neoadjuvant HER2-low BC		None	Phase II		NCT01042379 (119)
	ARX788	R/R Advanced HER2-low BC		None	Phase II		NCT05018676
	RC48	R/R Advanced HER2-low BC		None	Phase III		NCT04400695
	MRG002	R/R Advanced HER2+ BC		None	Phase II		NCT04924699
		R/R Advanced HER2 low BC		None	Phase II		NCT04742153
	A166	R/R Advanced HER2-expressing cancer		None	Phase II		NCT03602079
HER2-low advanced breast cancer				DS-8201a (single arm)		ORR: 37.0% 95%CI: 24.3%–51.3%	
Trop-2	DS-1062	Second-line advanced hormone receptor+/HER2- BC		None	Phase III		NCT05104866
		Third-line advanced hormone receptor+/HER2- BC		None	Phase III	ORR: 31.0% DOR: 7.4 m 95%CI: 4.4%–18.3% PFS: 6.8 m 95%CI: 4.6%–8.9%	NCT03901339 (120)
	IMMUN-132	Post-neoadjuvant HER2- BC		None	Phase III		NCT04595565 (121)
CD166	CX-2009	Post-neoadjuvant HER2- BC		None	Phase III		NCT04595565 (121)
HER3	U3-1402	Preoperative hormone receptor +/HER2- BC	Recruiting	None	Phase I	The primary endpoint is a CeTIL score after one single dose	NCT04610528 (122)

PFS, progression-free survival; mPFS, median progression free survival; OS, overall survival; ORR, objective response rate; DOR, duration of response; m, month.

disease progression following MTA multiline treatment. The NGS indicates that PIK3CA H1047R mutation may correlate to this better prognosis. Our data may help clinicians and patients make personalized decisions to try and rechallenge the ominous situation of metastatic disease to maximize efficacy and extend patient wellbeing for as long as possible.

## Author contributions

YH: Data curation, Writing – original draft, Writing – review & editing. FC: Data curation, Writing – original draft, Writing – review & editing. SS: Data curation, Writing – original draft, Writing – review & editing. LX: Methodology, Supervision, Writing – review & editing. XW: Supervision, Validation, Writing – review & editing. MW: Supervision, Validation, Writing – review & editing. SZ: Investigation, Visualization, Writing – review & editing. ZZ: Conceptualization, Funding acquisition, Writing – review & editing. ML: Conceptualization, Funding acquisition, Writing – review & editing.

## Funding

The author(s) declare financial support was received for research, authorship, and/or publication of this article. This work was supported by the National Natural Science Foundation of

China (Grant No. 81872156 to ML, No. 82274296 to ZZ). This work was also supported by the Dalian Medical University Interdisciplinary Research Cooperation Project Team Funding (Grant No. JCHZ2023009).

## Acknowledgments

The authors gratefully acknowledge the patients enrolled in this study.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## References

- Welch HG, Schwartz LM, Woloshin S. Are increasing 5-year survival rates evidence of success against cancer? *Jama* (2000) 283:2975–8. doi: 10.1001/jama.283.22.2975
- Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2022. *CA Cancer J Clin* (2022) 72(1):7–33. doi: 10.3322/caac.21708
- Waks AG, Winer EP. Breast cancer treatment: A review. *JAMA* (2019) 321:288–300. doi: 10.1001/jama.2018.19323
- Alfarouk KO, Stock C-M, Taylor S, Walsh M, Muddathir AK, Verduzco D, et al. Resistance to cancer chemotherapy: Failure in drug response from adme to p-gp. *Cancer Cell Int* (2015) 15:71. doi: 10.1186/s12935-015-0221-1
- Nikolaou M, Pavlopoulou A, Georgakilas AG, Kyrodimos E. The challenge of drug resistance in cancer treatment: A current overview. *Clin Exp Metastasis* (2018) 35:309–18. doi: 10.1007/s10585-018-9903-0
- Pivot X, Marmé F, Koenigsberg R, Guo M, Berrak E, Wolfé A. Pooled analyses of eribulin in metastatic breast cancer patients with at least one prior chemotherapy. *Ann Oncol* (2016) 27:1525–31. doi: 10.1093/annonc/mdw203
- Perez EA, Moreno-Aspitia A, Aubrey Thompson E, Andorfer CA. Adjuvant therapy of triple negative breast cancer. *Breast Cancer Res Treat* (2010) 120:285–91. doi: 10.1007/s10549-010-0736-z
- Tonini G, Imperatori M, Vincenzi B, Frezza AM, Santini D. Rechallenging therapy and treatment holiday: Different strategies in management of metastatic colorectal cancer. *J Exp Clin Cancer Res* (2013) 32:92. doi: 10.1186/1756-9966-32-92
- Ferraro E, Drago JZ, Modi S. Implementing antibody-drug conjugates (adcs) in her2-positive breast cancer: State of the art and future directions. *Breast Cancer research: BCR* (2021) 23:84. doi: 10.1186/s13058-021-01459-y
- Tarantino P, Carmagnani Pestana R, Corti C, Modi S, Bardia A, Tolane SM, et al. Antibody-drug conjugates: Smart chemotherapy delivery across tumor histologies. *CA: Cancer J Clin* (2022) 72:165–82. doi: 10.3322/caac.21705
- Fuentes-Antrás J, Genta S, Vijenthira A, Siu LL. Antibody-drug conjugates: In search of partners of choice. *Trends Cancer* (2023) 9:339–54. doi: 10.1016/j.trecan.2023.01.003
- Krop IE, LoRusso P, Miller KD, Modi S, Yardley D, Rodriguez G, et al. A phase ii study of trastuzumab emtansine in patients with human epidermal growth factor receptor 2-positive metastatic breast cancer who were previously treated with trastuzumab, lapatinib, an anthracycline, a taxane, and capecitabine. *J Clin Oncol* (2012) 30:3234–41. doi: 10.1200/jco.2011.40.5902
- Curigliano G, Dunton K, Rosenlund M, Janek M, Cathcart J, Liu Y, et al. Patient-reported outcomes and hospitalization data in patients with her2-positive metastatic breast cancer receiving trastuzumab deruxtecan or trastuzumab emtansine in the phase iii destiny-breast03 study. *Ann Oncol* (2023) 34:569–77. doi: 10.1016/j.annonc.2023.04.516
- Cortés J, Kim SB, Chung WP, Im SA, Park YH, Hegg R, et al. Trastuzumab deruxtecan versus trastuzumab emtansine for breast cancer. *New Engl J Med* (2022) 386:1143–54. doi: 10.1056/NEJMoa2115022
- Zhu Y, Zhu X, Wei X, Tang C, Zhang W. Her2-targeted therapies in gastric cancer. *Biochim Biophys Acta Rev Cancer* (2021) 1876:188549. doi: 10.1016/j.bbcan.2021.188549
- Li L, Xu MZ, Wang L, Jiang J, Dong LH, Chen F, et al. Conjugating mmae to a novel anti-her2 antibody for selective targeted delivery. *Eur Rev Med Pharmacol Sci* (2020) 24:12929–37. doi: 10.26355/eurrev\_202012\_24196
- Abdollahpour-Alitappeh M, Lotfinia M, Bagheri N, Sineh Sepehr K, Habibi-Anbouhi M, Kobarfard F, et al. Trastuzumab-monomethyl auristatin e conjugate exhibits potent cytotoxic activity *in vitro* against her2-positive human breast cancer. *J Cell Physiol* (2019) 234:2693–704. doi: 10.1002/jcp.27085
- Cullinane C, Fleming C, O'Leary DP, Hassan F, Kelly L, O'Sullivan MJ, et al. Association of circulating tumor DNA with disease-free survival in breast cancer: A systematic review and meta-analysis. *JAMA Netw Open* (2020) 3:e2026921. doi: 10.1001/jamanetworkopen.2020.26921
- Modi ND, Sorich MJ, Rowland A, McKinnon RA, Koczwara B, Wiese MD, et al. Predicting thrombocytopenia in patients with breast cancer treated with ado-trastuzumab emtansine. *Clin Breast Cancer* (2020) 20:e220–8. doi: 10.1016/j.clbc.2019.10.001
- Wang J, Liu Y, Zhang Q, Feng J, Fang J, Chen X, et al. Rc48-adc, a her2-targeting antibody-drug conjugate, in patients with her2-positive and her2-low expressing advanced or metastatic breast cancer: A pooled analysis of two studies. *J Clin Oncol* (2021) 39(15\_suppl):1022–2. doi: 10.1200/JCO.2021.39.15\_suppl.1022

21. Osumi H, Vecchione L, Keilholz U, Vollbrecht C, Alig AHS, von Einem JC, et al. Neoras wild-type in metastatic colorectal cancer: Myth or truth?-case series and review of the literature. *Eur J Cancer (Oxford England: 1990)* (2021) 153:86–95. doi: 10.1016/j.ejca.2021.05.010
22. Hemmat M, Castle BT, Odde DJ. Microtubule dynamics: Moving toward a multi-scale approach. *Curr Opin Cell Biol* (2018) 50:8–13. doi: 10.1016/j.jceb.2017.12.013
23. Yang C-PH, Horwitz SB. Taxol®: The first microtubule stabilizing agent. *Int J Mol Sci* (2017) 18(8):1733. doi: 10.3390/ijms18081733
24. Dominguez-Brauer C, Thu KL, Mason JM, Blaser H, Bray MR, Mak TW. Targeting mitosis in cancer: Emerging strategies. *Mol Cell* (2015) 60:524–36. doi: 10.1016/j.molcel.2015.11.006
25. Leandro-García LJ, Leskelä S, Landa I, Montero-Conde C, López-Jiménez E, Letón R, et al. Tumoral and tissue-specific expression of the major human beta-tubulin isotypes. *Cytoskeleton (Hoboken)* (2010) 67:214–23. doi: 10.1002/cm.20436
26. Kavallaris M, Kuo DY, Burkhart CA, Regl DL, Norris MD, Haber M, et al. Taxol-resistant epithelial ovarian tumors are associated with altered expression of specific beta-tubulin isotypes. *J Clin Invest* (1997) 100:1282–93. doi: 10.1172/JCI119642
27. Ferguson RE, Taylor C, Stanley A, Butler E, Joyce A, Harnden P, et al. Resistance to the tubulin-binding agents in renal cell carcinoma: No mutations in the class I beta-tubulin gene but changes in tubulin isotype protein expression. *Clin Cancer Res* (2005) 11:3439–45. doi: 10.1158/1078-0432.CCR-04-2049
28. Ohishi Y, Oda Y, Basaki Y, Kobayashi H, Wake N, Kuwano M, et al. Expression of beta-tubulin isotypes in human primary ovarian carcinoma. *Gynecol Oncol* (2007) 105:586–92. doi: 10.1016/j.ygyno.2007.01.044
29. Nicoletti MI, Valoti G, Giannakakou P, Zhan Z, Kim JH, Lucchini V, et al. Expression of beta-tubulin isotypes in human ovarian carcinoma xenografts and in a sub-panel of human cancer cell lines from the nci-anticancer drug screen: Correlation with sensitivity to microtubule active agents. *Clin Cancer Res* (2001) 7:2912–22.
30. Steinmetz MO, Protá AE. Microtubule-targeting agents: Strategies to hijack the cytoskeleton. *Trends Cell Biol* (2018) 28:776–92. doi: 10.1016/j.tcb.2018.05.001
31. Parker AL, Teo WS, McCarroll JA, Kavallaris M. An emerging role for tubulin isotypes in modulating cancer biology and chemotherapy resistance. *Int J Mol Sci* (2017) 18(7):1434. doi: 10.3390/ijms18071434
32. Das T, Anand U, Pandey SK, Ashby CR Jr., Assaraf YG, Chen ZS, et al. Therapeutic strategies to overcome taxane resistance in cancer. *Drug Resist Updat* (2021) 55:100754. doi: 10.1016/j.drup.2021.100754
33. Garcin C, Straube A. Microtubules in cell migration. *Essays Biochem* (2019) 63:509–20. doi: 10.1042/ebc20190016
34. Ganguly A, Yang H, Sharma R, Patel KD, Cabral F. The role of microtubules and their dynamics in cell migration. *J Biol Chem* (2012) 287:43359–69. doi: 10.1074/jbc.M112.423905
35. Fruman DA, Chiu H, Hopkins BD, Bagrodia S, Cantley LC, Abraham RT. The pi3k pathway in human disease. *Cell* (2017) 170:605–35. doi: 10.1016/j.cell.2017.07.029
36. Yasumizu Y, Miyajima A, Kosaka T, Miyazaki Y, Kikuchi E, Oya M. Dual pi3k/mtor inhibitor nvp-bez235 sensitizes docetaxel in castration resistant prostate cancer. *J Urol* (2014) 191:227–34. doi: 10.1016/j.juro.2013.07.101
37. Yang YL, Lee KT, Park HJ, Kim TJ, Choi YS, Shih Ie M, et al. Tectorigenin sensitizes paclitaxel-resistant human ovarian cancer cells through downregulation of the akt and nfkb pathway. *Carcinogenesis* (2012) 33:2488–98. doi: 10.1093/carcin/bgs302
38. Li J, Davies BR, Han S, Zhou M, Bai Y, Zhang J, et al. The akt inhibitor azd5363 is selectively active in pi3kca mutant gastric cancer, and sensitizes a patient-derived gastric cancer xenograft model with pten loss to taxotere. *J Trans Med* (2013) 11:241. doi: 10.1186/1479-5876-11-241
39. Ping YH, Lee HC, Lee JY, Wu PH, Ho LK, Chi CW, et al. Anticancer effects of low-dose 10-hydroxycamptothecin in human colon cancer. *Oncol Rep* (2006) 15:1273–9. doi: 10.3892/or.15.5.1273
40. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. *Cell* (2011) 144:646–74. doi: 10.1016/j.cell.2011.02.013
41. Moreno-Andrés D, Holl K, Antonin V. The second half of mitosis and its implications in cancer biology. *Semin Cancer Biol* (2023) 88:1–17. doi: 10.1016/j.semcancer.2022.11.013
42. Sun M, Jia M, Ren H, Yang B, Chi W, Xin G, et al. Numa regulates mitotic spindle assembly, structural dynamics and function via phase separation. *Nat Commun* (2021) 12:7157. doi: 10.1038/s41467-021-27528-6
43. Čermák V, Dostál V, Jelínek M, Libusová L, Kovář J, Rösel D, et al. Microtubule-targeting agents and their impact on cancer treatment. *Eur J Cell Biol* (2020) 99:151075. doi: 10.1016/j.jceb.2020.151075
44. Henriques AC, Ribeiro D, Pedrosa J, Sarmiento B, Silva PMA, Bousbaa H. Mitosis inhibitors in anticancer therapy: When blocking the exit becomes a solution. *Cancer Lett* (2019) 440-441:64–81. doi: 10.1016/j.canlet.2018.10.005
45. Gris-Oliver A, Ibrahim YH, Rivas MA, García-García C, Sánchez-Guixé M, Ruiz-Pace F, et al. Pi3k activation promotes resistance to eribulin in her2-negative breast cancer. *Br J Cancer* (2021) 124:1581–91. doi: 10.1038/s41416-021-01293-1
46. Czech MP. Pip2 and pip3: Complex roles at the cell surface. *Cell* (2000) 100:603–6. doi: 10.1016/S0092-8674(00)80696-0
47. Insall RH, Weiner OD. Pip3, pip2, and cell movement—similar messages, different meanings? *Dev Cell* (2001) 1:743–7. doi: 10.1016/S1534-5807(01)00086-7
48. Shen Q, Han Y, Wu K, He Y, Jiang X, Liu P, et al. Mrgprf acts as a tumor suppressor in cutaneous melanoma by restraining pi3k/akt signaling. *Signal Transduct Target Ther* (2022) 7:147. doi: 10.1038/s41392-022-00945-9
49. Hoxhaj G, Manning BD. The PI3K-AKT network at the interface of oncogenic signalling and cancer metabolism. *Nat Rev Cancer* (2015) 20(2):74–88. doi: 10.1038/s41568-019-0216-7
50. He Y, Sun MM, Zhang GG, Yang J, Chen KS, Xu WW, et al. Targeting PI3K/Akt signal transduction for cancer therapy. *Signal Transduct Target Ther* (2021) 6(1):425. doi: 10.1038/s41392-021-00828-5
51. Glaviano A, Foo ASC, Lam HY, Yap KCH, Jacot W, Jones RH, et al. PI3K/AKT/mTOR signaling transduction pathway and targeted therapies in cancer. *Mol Cancer* (2023) 22(1):138. doi: 10.1186/s12943-023-01827-6
52. Toyoshima F, Matsumura S, Morimoto H, Mitsushima M, Nishida E. Ptdins (3,4,5)p3 regulates spindle orientation in adherent cells. *Dev Cell* (2007) 13:796–811. doi: 10.1016/j.devcel.2007.10.014
53. Silió V, Redondo-Muñoz J, Carrera AC. Phosphoinositide 3-kinase  $\beta$  regulates chromosome segregation in mitosis. *Mol Biol Cell* (2012) 23:4526–42. doi: 10.1091/mbc.E12-05-0371
54. Liu X, Shi Y, Woods KW, Hessler P, Kroeger P, Wilsbacher J, et al. Akt inhibitor a-443654 interferes with mitotic progression by regulating aurora a kinase expression. *Neoplasia (New York NY)* (2008) 10:828–37. doi: 10.1593/neo.08408
55. Leonard MK, Hill NT, Bubulya PA, Kadakia MP. The pten-akt pathway impacts the integrity and composition of mitotic centrosomes. *Cell Cycle (Georgetown Tex)* (2013) 12:1406–15. doi: 10.4161/cc.24516
56. Fruman DA, Rommel C. Pi3k and cancer: Lessons, challenges and opportunities. *Nat Rev Drug Discov* (2014) 13:140–56. doi: 10.1038/nrd4204
57. Higuchi M, Masuyama N, Fukui Y, Suzuki A, Gotoh Y. Akt mediates rac/cdc42-regulated cell motility in growth factor-stimulated cells and in invasive pten knockout cells. *Curr Biol* (2001) 11:1958–62. doi: 10.1016/S0960-9822(01)00599-1
58. Sasaki AT, Chun C, Takeda K, Firtel RA. Localized ras signaling at the leading edge regulates pi3k, cell polarity, and directional cell movement. *J Cell Biol* (2004) 167:505–18. doi: 10.1083/jcb.200406177
59. Onishi K, Higuchi M, Asakura T, Masuyama N, Gotoh Y. The pi3k-akt pathway promotes microtubule stabilization in migrating fibroblasts. *Genes Cells* (2007) 12:535–46. doi: 10.1111/j.1365-2443.2007.01071.x
60. Fujiwara Y, Hosokawa Y, Watanabe K, Tanimura S, Ozaki K, Kohno M. Blockade of the phosphatidylinositol-3-kinase-akt signaling pathway enhances the induction of apoptosis by microtubule-destabilizing agents in tumor cells in which the pathway is constitutively activated. *Mol Cancer Ther* (2007) 6:1133–42. doi: 10.1158/1535-7163.Mct-06-0639
61. Stanton RA, Gernert KM, Nettles JH, Aneja R. Drugs that target dynamic microtubules: A new molecular perspective. *Medicinal Res Rev* (2011) 31:443–81. doi: 10.1002/med.20242
62. Semenova I, Ikeda K, Resaul K, Kraikivski P, Aguiar M, Gygi S, et al. Regulation of microtubule-based transport by map4. *Mol Biol Cell* (2014) 25:3119–32. doi: 10.1091/mbc.E14-01-0022
63. Thapa N, Chen M, Horn HT, Choi S, Wen T, Anderson RA. Phosphatidylinositol-3-oh kinase signalling is spatially organized at endosomal compartments by microtubule-associated protein 4. *Nat Cell Biol* (2020) 22:1357–70. doi: 10.1038/s41556-020-00596-4
64. Khwaja S, Kumar K, Das R, Negi AS. Microtubule associated proteins as targets for anticancer drug development. *Bioorganic Chem* (2021) 116:105320. doi: 10.1016/j.bioorg.2021.105320
65. Cocco S, Leone A, Roca MS, Lombardi R, Piezzo M, Caputo R, et al. Inhibition of autophagy by chloroquine prevents resistance to pi3k/akt inhibitors and potentiates their antitumor effect in combination with paclitaxel in triple negative breast cancer models. *J Transl Med* (2022) 20:290. doi: 10.1186/s12967-022-03462-z
66. Liu Z, Zhu G, Getzenberg RH, Veltri RW. The upregulation of pi3k/akt and map kinase pathways is associated with resistance of microtubule-targeting drugs in prostate cancer. *J Cell Biochem* (2015) 116:1341–9. doi: 10.1002/jcb.25091
67. Slade D. Parp and parg inhibitors in cancer treatment. *Genes Dev* (2020) 34:360–94. doi: 10.1101/gad.334516.119
68. Murai J, S.-y. N, Das BB, Renaud A, Zhang Y, Doroshow JH, et al. Trapping of parp1 and parp2 by clinical parp inhibitors. *Cancer Res* (2012) 72:5588–99. doi: 10.1158/0008-5472.CAN-12-2753
69. Holloway RW, Marignani PA. Targeting mtor and glycolysis in her2-positive breast cancer. *Cancers (Basel)* (2021) 13(12):2922. doi: 10.3390/cancers13122922
70. Pernas S, Tolaney SM. Her2-positive breast cancer: New therapeutic frontiers and overcoming resistance. *Ther Adv Med Oncol* (2019) 11:1758835919833519. doi: 10.1177/1758835919833519
71. Loibl S, Majewski I, Guarneri V, Nekljudova V, Holmes E, Bria E, et al. Pik3ca mutations are associated with reduced pathological complete response rates in primary her2-positive breast cancer: Pooled analysis of 967 patients from five prospective trials

- investigating lapatinib and trastuzumab. *Ann Oncol* (2016) 27:1519–25. doi: 10.1093/annonc/mdw197
72. Dave B, Migliaccio I, Gutierrez MC, Wu MF, Chamness GC, Wong H, et al. Loss of phosphatase and tensin homolog or phosphoinositide-3 kinase activation and response to trastuzumab or lapatinib in human epidermal growth factor receptor 2-overexpressing locally advanced breast cancers. *J Clin Oncol* (2011) 29:166–73. doi: 10.1200/jco.2009.27.7814
73. Yi Z, Ma F, Li C, Chen R, Yuan L, Sun X, et al. Landscape of somatic mutations in different subtypes of advanced breast cancer with circulating tumor DNA analysis. *Sci Rep* (2017) 7:5995. doi: 10.1038/s41598-017-06327-4
74. Ahmad F, Badwe A, Verma G, Bhatia S, Das BR. Molecular evaluation of pik3ca gene mutation in breast cancer: Determination of frequency, distribution pattern and its association with clinicopathological findings in Indian patients. *Med Oncol (Northwood London England)* (2016) 33:74. doi: 10.1007/s12032-016-0788-y
75. Martínez-Sáez O, Chic N, Pascual T, Adamo B, Vidal M, González-Farré B, et al. Frequency and spectrum of pik3ca somatic mutations in breast cancer. *Breast Cancer Res* (2020) 22:45. doi: 10.1186/s13058-020-01284-9
76. Qiu W, Schönleben F, Li X, Ho DJ, Close LG, Manolidis S, et al. Pik3ca mutations in head and neck squamous cell carcinoma. *Clin Cancer Res* (2006) 12:1441–6. doi: 10.1158/1078-0432.Ccr-05-2173
77. Dogruluk T, Tsang YH, Espitia M, Chen F, Chen T, Chong Z, et al. Identification of variant-specific functions of pik3ca by rapid phenotyping of rare mutations. *Cancer Res* (2015) 75:5341–54. doi: 10.1158/0008-5472.Can-15-1654
78. Ng PK, Li J, Jeong KJ, Shao S, Chen H, Tsang YH, et al. Systematic functional annotation of somatic mutations in cancer. *Cancer Cell* (2018) 33:450–462.e10. doi: 10.1016/j.ccell.2018.01.021
79. The Cancer Genome Atlas Network. Comprehensive molecular portraits of human breast tumours. *Nature* (2012) 490:61–70. doi: 10.1038/nature11412
80. Arafeh R, Samuels Y. Pik3ca in cancer: The past 30 years. *Semin Cancer Biol* (2019) 59:36–49. doi: 10.1016/j.semcancer.2019.02.002
81. Samuels Y, Diaz LA Jr., Schmidt-Kittler O, Cummins JM, Delong L, Cheong I, et al. Mutant pik3ca promotes cell growth and invasion of human cancer cells. *Cancer Cell* (2005) 7:561–73. doi: 10.1016/j.ccr.2005.05.014
82. Isakoff SJ, Engelman JA, Irie HY, Luo J, Brachmann SM, Pearlman RV, et al. Breast cancer-associated pik3ca mutations are oncogenic in mammary epithelial cells. *Cancer Res* (2005) 65:10992–1000. doi: 10.1158/0008-5472.Can-05-2612
83. Kang S, Bader AG, Vogt PK. Phosphatidylinositol 3-kinase mutations identified in human cancer are oncogenic. *Proc Natl Acad Sci USA* (2005) 102:802–7. doi: 10.1073/pnas.0408864102
84. Hennessy BT, Smith DL, Ram PT, Lu Y, Mills GB. Exploiting the pi3k/akt pathway for cancer drug discovery. *Nat Rev Drug Discov* (2005) 4:988–1004. doi: 10.1038/nrd1902
85. Gymnopoulos M, Elsliger MA, Vogt PK. Rare cancer-specific mutations in pik3ca show gain of function. *Proc Natl Acad Sci USA* (2007) 104:5569–74. doi: 10.1073/pnas.0701005104
86. Ikenoue T, Kanai F, Hikiba Y, Obata T, Tanaka Y, Imamura J, et al. Functional analysis of pik3ca gene mutations in human colorectal cancer. *Cancer Res* (2005) 65:4562–7. doi: 10.1158/0008-5472.Can-04-4114
87. Hon WC, Berndt A, Williams RL. Regulation of lipid binding underlies the activation mechanism of class ia pi3-kinases. *Oncogene* (2012) 31:3655–66. doi: 10.1038/ncr.2011.532
88. Burke JE, Perisic O, Masson GR, Vadas O, Williams RL. Oncogenic mutations mimic and enhance dynamic events in the natural activation of phosphoinositide 3-kinase p110 $\alpha$  (pik3ca). *Proc Natl Acad Sci USA* (2012) 109:15259–64. doi: 10.1073/pnas.1205508109
89. Matthews DJ, O'Farrell M, James J, Giddens AC, Rewcastle GW, Denny WA. Abstract 4485: Preclinical characterization of pwt33597, a dual inhibitor of pi3-kinase alpha and mtor. *Cancer Res* (2011) 71:4485–5. doi: 10.1158/1538-7445.AM2011-4485
90. Janku F, Wheler JJ, Naing A, Falchook GS, Hong DS, Stepanek VM, et al. Pik3ca mutation h1047r is associated with response to pi3k/akt/mtor signaling pathway inhibitors in early-phase clinical trials. *Cancer Res* (2013) 73:276–84. doi: 10.1158/0008-5472.Can-12-1726
91. Vasan N, Razavi P, Johnson JL, Shao H, Shah H, Antoine A, et al. Double pik3ca mutations in cis increase oncogenicity and sensitivity to pi3k $\alpha$  inhibitors. *Science* (2019) 366:714–23. doi: 10.1126/science.aaw9032
92. du Rusquec P, Blonz C, Frenel JS, Campone M. Targeting the pi3k/akt/mtor pathway in estrogen-receptor positive her2 negative advanced breast cancer. *Ther Adv Med Oncol* (2020) 12:1758835920940939. doi: 10.1177/1758835920940939
93. Weigelt B, Warne PH, Downward J. Pik3ca mutation, but not pten loss of function, determines the sensitivity of breast cancer cells to mtor inhibitory drugs. *Oncogene* (2011) 30:3222–33. doi: 10.1038/ncr.2011.42
94. Bader AG, Kang S, Vogt PK. Cancer-specific mutations in pik3ca are oncogenic in vivo. *Proc Natl Acad Sci USA* (2006) 103:1475–9. doi: 10.1073/pnas.0510857103
95. Yi Z, Ma F, Liu B, Guan X, Li L, Li C, et al. Everolimus in hormone receptor-positive metastatic breast cancer: Pik3ca mutation h1047r was a potential efficacy biomarker in a retrospective study. *BMC Cancer* (2019) 19:442. doi: 10.1186/s12885-019-5668-3
96. Mohseni M, Park BH. Pik3ca and kras mutations predict for response to everolimus therapy: Now that's rad001. *J Clin Invest* (2010) 120:2655–8. doi: 10.1172/JCI44026
97. Meric-Bernstam F, Akcakanat A, Chen H, Do K-A, Sangai T, Adkins F, et al. Pik3ca/pten mutations and akt activation as markers of sensitivity to allosteric mtor inhibitors. *Clin Cancer Res* (2012) 18:1777–89. doi: 10.1158/1078-0432.Ccr-11-2123
98. Xu H, Chen K, Shang R, Chen X, Zhang Y, Song X, et al. Alpelisib combination treatment as novel targeted therapy against hepatocellular carcinoma. *Cell Death Dis* (2021) 12:920. doi: 10.1038/s41419-021-04206-5
99. Goyal L, Kongpetch S, Crolley VE, Bridgewater J. Targeting fgfr inhibition in cholangiocarcinoma. *Cancer Treat Rev* (2021) 95:102170. doi: 10.1016/j.ctrv.2021.102170
100. Goyal L, Meric-Bernstam F, Hollebecque A, Valle JW, Morizane C, Karasic TB, et al. Futibatinib for fgfr2-rearranged intrahepatic cholangiocarcinoma. *N Engl J Med* (2023) 388:228–39. doi: 10.1056/NEJMoa2206834
101. White C, Scott RJ, Paul C, Ziolkowski A, Mossman D, Fox SB, et al. Dihydropyrimidine dehydrogenase deficiency and implementation of upfront dpdy genotyping. *Clin Pharmacol Ther* (2022) 112:791–802. doi: 10.1002/cpt.2667
102. Ye T, Zhang J-Y, Liu X-Y, Zhou Y-H, Yuan S-Y, Yang M-M, et al. The predictive value of map2k1/2 mutations on efficiency of immunotherapy in melanoma. *Front Immunol* (2021) 12:785526. doi: 10.3389/fimmu.2021.785526
103. Hausser J, Alon U. Tumour heterogeneity and the evolutionary trade-offs of cancer. *Nat Rev Cancer* (2020) 20:247–57. doi: 10.1038/s41568-020-0241-6
104. Pereira B, Chin SF, Rueda OM, Vollan HK, Provenzano E, Bardwell HA, et al. The somatic mutation profiles of 2,433 breast cancers refines their genomic and transcriptomic landscapes. *Nat Commun* (2016) 7:11479. doi: 10.1038/ncomms11479
105. Lopez G, Costanza J, Colleoni M, Fontana L, Ferrero S, Miozzo M, et al. Molecular insights into the classification of luminal breast cancers: The genomic heterogeneity of progesterone-negative tumors. *Int J Mol Sci* (2019) 20(3):510. doi: 10.3390/ijms20030510
106. Cardoso F, Paluch-Shimon S, Senkus E, Curigliano G, Aapro MS, André F, et al. 5th eso-esmo international consensus guidelines for advanced breast cancer (abc 5). *Ann Oncol* (2020) 31:1623–49. doi: 10.1016/j.annonc.2020.09.010
107. Piezzo M, Chiodini P, Riemma M, Cocco S, Caputo R, Cianniello D, et al. Progression-free survival and overall survival of cdk 4/6 inhibitors plus endocrine therapy in metastatic breast cancer: A systematic review and meta-analysis. *Int J Mol Sci* (2020) 21(17):6400. doi: 10.3390/ijms21176400
108. de Graffenried LA, Friedrichs WE, Russell DH, Donzis EJ, Middleton AK, Silva JM, et al. Inhibition of mtor activity restores tamoxifen response in breast cancer cells with aberrant akt activity. *Clin Cancer Res* (2004) 10:8059–67. doi: 10.1158/1078-0432.Ccr-04-0035
109. Shui R, Liang X, Li X, Liu Y, Li H, Xu E, et al. Hormone receptor and human epidermal growth factor receptor 2 detection in invasive breast carcinoma: A retrospective study of 12,467 patients from 19 chinese representative clinical centers. *Clin Breast Cancer* (2020) 20:e65–74. doi: 10.1016/j.clbc.2019.07.013
110. Chang YC, Cheung CHA, Kuo YL. Tamoxifen rechallenge decreases metastatic potential but increases cell viability and clonogenicity in a tamoxifen-mediated cytotoxicity-resistant subline of human breast mcf7 cancer cells. *Front Cell Dev Biol* (2020) 8:485. doi: 10.3389/fcell.2020.00485
111. Toulmonde M, Madranges N, Brouste V, Donamaria C, MacGrogan G, Durand M, et al. Docetaxel rechallenge after a first response in non-resistant metastatic breast cancer: Significant activity with manageable toxicity. *Breast Cancer Res Treat* (2012) 134:325–32. doi: 10.1007/s10549-012-2060-2
112. Trudeau ME, Clemons MJ, Provencher L, Panasci L, Yelle L, Rayson D, et al. Phase ii multicenter trial of anthracycline rechallenge with pegylated liposomal doxorubicin plus cyclophosphamide for first-line therapy of metastatic breast cancer previously treated with adjuvant anthracyclines. *J Clin Oncol* (2009) 27:5906–10. doi: 10.1200/jco.2009.22.7504
113. Blondeaux E, Ferreira AR, Poggio F, Puglisi F, Bighin C, Sottotetti F, et al. Clinical outcomes of patients with breast cancer relapsing after (neo)adjuvant trastuzumab and receiving trastuzumab rechallenge or lapatinib-based therapy: A multicentre retrospective cohort study. *ESMO Open* (2020) 5(4):e000719. doi: 10.1136/esmoopen-2020-000719
114. Araki K, Fukada I, Horii R, Takahashi S, Akiyama F, Iwase T, et al. Trastuzumab rechallenge after lapatinib- and trastuzumab-resistant disease progression in her2-positive breast cancer. *Clin Breast Cancer* (2015) 15:432–9. doi: 10.1016/j.clbc.2015.06.002
115. Feng D, Guan Y, Liu M, He S, Zhao W, Yin B, et al. Excellent response to atezolizumab after clinically defined hyperprogression upon previous treatment with pembrolizumab in metastatic triple-negative breast cancer: A case report and review of the literature. *Front Immunol* (2021) 12:608292. doi: 10.3389/fimmu.2021.608292
116. Fuentes-Antrás J, de Luna A, López de Sá A, Ocaña A, García-Sáenz J, Moreno F. Safety and efficacy of cyclin-dependent kinase inhibitor rechallenge following ribociclib-induced limiting hypertransaminasemia. *Breast (Edinburgh Scotland)* (2020) 54:160–3. doi: 10.1016/j.breast.2020.10.007

117. Ramaswami R, O’Cathail SM, Brindley JH, Silcocks P, Mahmoud S, Palmieri C. Activity of eribulin mesylate in heavily pretreated breast cancer granted access via the cancer drugs fund. *Future Oncol (London England)* (2014) 10:363–76. doi: 10.2217/fon.13.210
118. Modi S, Jacot W, Yamashita T, Sohn J, Vidal M, Tokunaga E, et al. Trastuzumab deruxtecan in previously treated her2-low advanced breast cancer. *New Engl J Med* (2022) 387:9–20. doi: 10.1056/NEJMoa2203690
119. Barbieri E, Rubino D, Hakim R, Fini A, Lenzi M, Zamagni C. Eribulin long-term response and rechallenge: Report of two clinical cases. *Future Oncol (London England)* (2017) 13:35–43. doi: 10.2217/fon-2016-0520
120. Bardia A, Mayer IA, Diamond JR, Morooso RL, Isakoff SJ, Starodub AN, et al. Efficacy and safety of anti-trop-2 antibody drug conjugate sacituzumab govitecan (immu-132) in heavily pretreated patients with metastatic triple-negative breast cancer. *J Clin Oncol* (2017) 35:2141–8. doi: 10.1200/jco.2016.70.8297
121. Marmè F, Hanusch C, Furlanetto J, Morris P, Link T, Denkert C, et al. 580 safety interim analysis (sia) of the phase iii postneoadjuvant scasia study evaluating sacituzumab govitecan (sg) in patients with primary her2-negative breast cancer (bc) at high relapse risk after neoadjuvant treatment. *Ann Oncol* (2022) 33:S148–9. doi: 10.1016/j.annonc.2022.03.074
122. Pascual T, Oliveira M, Ciruelos E, Bellet Ezquerro M, Saura C, Gavilá J, et al. Solti-1805 tot-her3 study concept: A window-of-opportunity trial of patritumab deruxtecan, a her3 directed antibody drug conjugate, in patients with early breast cancer. *Front Oncol* (2021) 11:638482. doi: 10.3389/fonc.2021.638482