Check for updates

OPEN ACCESS

EDITED BY Frank A. Orlando, University of Florida, United States

REVIEWED BY Dechao Feng, University College London, United Kingdom Hamidullah Khan, University of Wisconsin-Madison, United States Xiaobin Gu, First Affiliated Hospital of Zhengzhou University, China Ikhwan Rinaldi, RSUPN Dr. Cipto Mangunkusumo, Indonesia

*CORRESPONDENCE Fangzhen Cai ☑ caifz@fjmu.edu.cn Wei Zhuang ☑ 612032@fjmu.edu.cn Pu Zhang ☑ 18981960971@163.com

[†]These authors have contributed equally to this work

RECEIVED 23 January 2024 ACCEPTED 01 May 2024 PUBLISHED 23 May 2024

CITATION

He Q, Wei C, Cao L, Zhang P, Zhuang W and Cai F (2024) Blood cell indices and inflammation-related markers with kidney cancer risk: a large-population prospective analysis in UK Biobank. *Front. Oncol.* 14:1366449. doi: 10.3389/fonc.2024.1366449

COPYRIGHT

© 2024 He, Wei, Cao, Zhang, Zhuang and Cai. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Blood cell indices and inflammation-related markers with kidney cancer risk: a largepopulation prospective analysis in UK Biobank

Qingliu He^{1,2†}, Chengcheng Wei^{2,3†}, Li Cao^{4†}, Pu Zhang^{5*}, Wei Zhuang^{1*} and Fangzhen Cai^{1*}

¹Department of Urology, The Second Affiliated Hospital of Fujian Medical University, Quanzhou, China, ²Department of Urology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, Hubei, China, ³Department of Urology, The First Affiliated Hospital of Chongqing Medical University, Chongqing, China, ⁴Department of Orthopaedic, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, Hubei, China, ⁵Department of Urology, Sichuan Provincial People's Hospital, School of Medicine, University of Electronic Science and Technology of China, Chengdu, China

Background: Kidney cancer is a prevalent malignancy with an increasing incidence worldwide. Blood cell indices and inflammation-related markers have shown huge potential as biomarkers for predicting cancer incidences, but that is not clear in kidney cancer. Our study aims to investigate the correlations of blood cell indices and inflammation-related markers with kidney cancer risk.

Methods: We performed a population-based cohort prospective analysis using data from the UK Biobank. A total of 466,994 participants, free of kidney cancer at baseline, were included in the analysis. The hazard ratios (HRs) and 95% confidence intervals (Cls) for kidney cancer risk were calculated using Cox proportional hazards regression models. Restricted cubic spline models were used to investigate nonlinear longitudinal associations. Stratified analyses were used to identify high-risk populations. The results were validated through sensitivity analyses.

Results: During a mean follow-up of 12.4 years, 1,710 of 466,994 participants developed kidney cancer. The Cox regression models showed that 13 blood cell indices and four inflammation-related markers were associated with kidney cancer incidence. The restricted cubic spline models showed non-linear relationships with kidney cancer. Finally, combined with stratified and sensitivity analyses, we found that the mean corpuscular hemoglobin concentration (MCHC), red blood cell distribution width (RDW), platelet distribution width (PDW), systemic immune-inflammation index (SII), and product of platelet count and neutrophil count (PPN) were related to enhanced kidney cancer risk with stable results.

Conclusion: Our findings identified that three blood cell indices (MCHC, RDW, and PDW) and two inflammation-related markers (SII and PPN) were independent risk factors for the incidence of kidney cancer. These indexes may serve as potential predictors for kidney cancer and aid in the development of targeted screening strategies for at-risk individuals.

KEYWORDS

kidney cancer, blood cell indices, inflammation-related markers, UK Biobank, prospective analysis

1 Introduction

As one of the most common urogenital malignancies, kidney cancer has garnered increased attention due to its rising incidence and significant mortality rates, accounting for 5% of all cancer cases and holding the sixth position among the most prevalent cancers in men (1-4). Kidney cancer, also known as renal cancer, is a disease that is becoming more common globally, with over 400,000 new cases reported each year. The mortality rate for kidney cancer is also high, with around 175,000 fatalities occurring annually worldwide (5-8). Despite its frequency, kidney cancer remains a complex and heterogeneous disease, characterized by challenges in early detection, limited treatment options, and an inadequate understanding of its underlying mechanisms (9). Several wellestablished risk factors have been identified, including age, sex, smoking, obesity, and hypertension (10, 11). However, numerous other factors that are suspected of elevating the risk of kidney cancer require further investigation (10).

Blood indices, such as the mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), and platelet count, have served as valuable indicators of hematological alterations that could potentially reflect systemic body state and tumor progression (12-15). These biomarkers provide valuable insights into the underlying pathophysiological processes and interactions within the tumor microenvironment (16, 17). Notably, kidney cancer patients frequently exhibit anemia, with up to 35% of cases demonstrating decreased levels of hemoglobin (HGB), hematocrit (HCT), MCV, and MCH due to the weak activity of erythropoietin (EPO) and abnormal iron metabolism (18, 19). For the red blood cell and platelet systems, preoperative reductions in HGB, HCT, MCV, and MCH have been linked to an increased predisposition for early recurrence and progression of kidney cancer (20, 21). Additionally, a decrease in the average size of platelets has emerged as an independent predictor of tumor-specific mortality (18). While these findings highlight the significance of abnormal hemograms in the progression and prognosis of kidney cancer, little attention is focused on the occurrence of kidney cancer (22, 23).

Inflammation plays an important role in the incidence and progression of cancer (24, 25). Systemic inflammation is usually

assessed through various biochemical or hematological markers routinely measured in common blood tests or as ratios from these measurements, which are called inflammation-related markers, including the systemic immune-inflammation index (SII), neutrophil-lymphocyte ratio (NLR), platelet-lymphocyte ratio (PLR), and product of platelet count and neutrophil count (PPN) (26–30). While increasing evidence indicates that these markers served as prognostic markers in newly diagnosed cancer patients with various malignancies, only a few studies have focused on the risk of cancer incidence (24, 31–33). As an immunogenic tumor, kidney cancer has a strongly evident interaction with inflammatory mediators (34). However, the association between inflammationrelated markers and kidney cancer is still unclear.

In this paper, we aim to examine a wide range of blood cell indices and inflammation-related markers to determine their independent contributions to kidney cancer risk. Using a comprehensive prospective study design, we analyzed a large dataset comprising kidney cancer patients and matched controls, thereby providing valuable indexes into the development of this malignancy.

2 Methods

2.1 Study design

The UK Biobank is a large-scale prospective study designed to provide valuable resources for investigating the causes of various diseases. Comprehensive information regarding the study procedure and data collection can be accessed online or through relevant literature sources (https://www.ukbiobank.ac.uk/media/ gnkeyh2q/study-rationale.pdf) (35). In brief, the study included participants who were registered with the NHS and lived within a 40-km radius of a UK Biobank assessment center. An initial invitation was extended to approximately 9.2 million individuals (36). Between 2006 and 2010, over 500,000 men and women aged between 40 and 69 years old consented to participate in the cohort and visited one of the 22 assessment centers located in England, Wales, and Scotland. The UK Biobank study was approved by the National Health and Social Care Information Management Board and the North West Multicenter Research Ethics Committee (11/ NW/0382). This approval ensures that the study was conducted in compliance with ethical guidelines and that the rights and privacy of the participants were protected.

2.2 Population selection

This study was performed following the guidelines and regulations outlined in UKB application number 61083. A total of 502,401 participants were included in our analysis. Initially, we excluded 32,677 participants who had missing blood cell indices. Subsequently, we eliminated 397 participants who had kidney cancer at baseline and were lacking critical covariate data (N = 482). Ultimately, the primary analysis consisted of 466,994 participants (Figure 1).

2.3 Blood cell indices

UKB comprises a comprehensive collection of blood cell indices extracted from biological samples. Using a hematology analyzer, exclusive count data were acquired, and other parameters were calculated accordingly (details at https://biobank.ctsu.ox.ac.uk/ crystal/ukb/docs/haematology.pdf). All indices included in this study were extracted from the initial visit and then classified as "red blood cell," "immature red blood cell," "white blood cell," and "inflammation-related index." The red blood cell category included RBC, HCT, MCV, MCH, MCHC, and RDW. The immature RBC category included the reticulocyte count, reticulocyte percentage, nucleated RBC, nucleated RBC percentage, MRV, MSCV, and IRF. The white blood cell category included WBC, basophil count (BASO), basophil percentage, eosinophil count (EO), eosinophil percentage, monocyte count, monocyte percentage, neutrophil count (NEUT), neutrophil percentage, lymphocyte count (LYMPH), lymphocyte percentage, MPV, platelet distribution width, and platelet (PLT) and high light scatter reticulocyte percentage. For a more complete examination of the connection between "inflammation-related marker" and the incidence of kidney cancer, SII, PLR, NLR, and PPN were calculated. PLT, NEUT, and LYMPH concentrations were measured at 1,000 cells/ µL. PLT was multiplied by (NEUT/LYMPH) to arrive at SII. The PLR was computed by dividing the PLT by the LYMPH. NLR was determined using NEUT/LYMPH. PPN was determined by multiplying PLT by NEUT. All analytes within the dataset had identical detection limits. None of the results fell below the threshold for detection. Supplementary Table S1 of the Supplementary Materials shows a list of indicators with their corresponding abbreviations and data fields.



2.4 Covariates

We utilized the baseline touch-screen questionnaire to evaluate a number of potential confounding variables: age (years), sex, race (white, mixed, Asian, black, etc.), household income, lifestyle behavior (smoking status and alcohol drinker status), and body mass index (BMI). The questionnaire yielded the codes a degree (college or university degree) or no degree for education. The Townsend Deprivation Index is a composite measure of deprivation based on nonhome ownership, non-car ownership, unemployment, and domestic overcrowding that indicates the socioeconomic status of the participant. Adapted from the American Heart Association Guidelines, a balanced diet score was defined as adhering to four or five of the following components: (1) total fruit consumption of 4.5 pieces per week, (2) total vegetable consumption of 4.5 servings per week (three tablespoons of vegetable considered as one serving), (3) total fish consumption of two servings per week, (4) processed meat consumption at two times per week, and (5) red meat consumption at five times per week. The comorbidity variables included hypertension (Field 6150) and diabetes (Field 2443).

2.5 Kidney cancer diagnosis

This study's results were C64 (kidney malignant tumor, excluding renal pelvis). The International Classification of Diseases (ICD) classification system was utilized to record the diagnoses. The incident disease in this study was determined by the primary or secondary diagnoses from hospital admission data or by the primary or secondary causes after the baseline data collection. The participants were observed from the time of baseline until the date of the first diagnosis or on December 31, 2021.

2.6 Statistical analysis

The mean (standard deviation, SD) and number (percentage) for continuous variables were obtained. Figure 1 illustrates the study design flowchart. Blood cell indices were log-transformed and normalized to Z scores [Z = (value - mean)/SD] prior to Cox analysis so that the hazard ratio (HR) represents the effect per SD increment. Model 1 represented the unadjusted model, which did not consider any covariates. Model 2, the minimally adjusted model, accounted for gender and age as potential confounding variables. Model 3, the fully adjusted model, considered a comprehensive set of covariates including sex, age, race, qualification, BMI, smoking, alcohol consumption, health diet score, household income, diabetes, and hypertension. The potential for nonlinearity was investigated using restricted cubic spline models fitted to Cox models and adjusted for covariates as in model 3. Potential nonlinearity was examined by comparing the linear model to the model containing both linear and cubic spline terms using the likelihood ratio test. Then, we performed stratified analyses to estimate the potential modification effects according to age (\leq 60 and >60 years), sex (male or female), race (white, mixed, Asian, black, and others), smoking status (current, former, and never), alcohol drinker status (current, former, and never), qualifications (with college or university degree or none), diabetes (yes and no), hypertension (yes and no), and BMI (\leq 25 and >25). By modeling the cross-product terms of the stratifying variables with each blood cell index, interaction *P* values were examined. In the sensitivity analysis, we first applied models 1 and 2 to test the robustness of our findings. Second, we used quantile regression for further analysis. Third, we used the first repeat assessment visit of blood cell index data in 2012 and conducted Cox regression analysis to support the stability of the results.

All statistical analyses were carried out using R version 4.2.1. All *P* values listed below were adjusted.

3 Results

3.1 Baseline characteristics

Baseline characteristics stratified by kidney cancer status are shown in Table 1. A total of 466,994 kidney cancer-free participants from UKB were included in the primary analyses. Overall, the mean age of the participants was $56.5 (\pm 8.1)$ years; 214,089 (45.8%) were male patients and 252,905 (54.2%) were female patients. During a mean follow-up time of 12.4 years, 1,710 participants developed kidney cancer. Among them, 1,094 (64.0%) participants were male patients, and 616 (36.0%) participants were female patients.

3.2 Blood cell indices and kidney cancer risk

In the RBC category, decreased levels of MCV [HR 0.90, 95% confidence interval (CI): 0.86–0.95, P < 0.0001] and MCHC (HR 0.91, 95% CI: 0.876–0.96, P < 0.0006), indicative of an anemia state, were associated with higher kidney cancer incidence in the fully adjusted models (model 3). RDW, indicative of the heterogeneity of erythrocytes, was positively associated with kidney cancer risk (HR 1.10, 95% CI: 1.05–1.14, P < 0.0001). U-shaped relationships were discovered between RBC (P for nonlinearity = 0.0007; Figure 2), HCT (P for nonlinearity <0.0002; Figure 2), MCV (P for nonlinearity = 0.0227; Figure 2), and MCHC (P for nonlinearity = 0.0115; Figure 2) with incident kidney cancer.

In the immature RBC category, decreased reticulocyte maturation parameters, including NRBC (HR 0.88, 95% CI: 0.78–0.99, P = 0.0391), NRBC% (HR 0.88, 95% CI: 0.78–0.99, P = 0.0391), MRV (HR 0.94, 95% CI: 0.90–0.99, P = 0.0180), and MSCV (HR 0.88, 95% CI: 0.83–0.92, P < 0.0001), were significantly associated with a higher kidney cancer risk. MRV (P for nonlinearity = 0.00051) had a U-shaped association with kidney cancer risk. MSCV had a linear association with kidney cancer risk, rapidly reaching the lowest risk at ≈80 and becoming flat thereafter.

In the WBC category, increased levels of WBC (HR 1.04, 95% CI: 1.03–1.05, P < 0.0001), NEUT (HR 1.14, 95% CI: 1.11–1.17, P < 0.0001)

TABLE 1 Baseline characteristics of UK Biobank participants by incident kidney cancer incidence.

Characteristics	Overall (N = 466,994)	No kidney cancer (<i>N</i> = 465,284)	Kidney cancer (<i>N</i> =1,710)	P value
Mean follow-up duration (years) (SD)	12.4 ± 1.0	12.4 ± 0.9	6.5 ± 3.5	<0.001
Age, mean (SD), years	56.5 ± 8.1	56.5 ± 8.1	60.6 ± 6.5	<0.001
Gender (%)				
Male	214 089 (45.8)	212.995 (45.8)	1,094 (64.0)	
Female	252,905 (54.2)	252,289 (54.2)	616 (36.0)	
Race (%)				<0.001
White European	44,1207 (94.5)	439,551 (94.5)	1,656 (96.8)	
Mixed	2,717 (0.6)	2,711 (0.6)	6 (0.4)	
South Asian	8,798 (1.9)	8,777 (1.9)	21 (1.2)	
Black	7,073 (1.5)	7,061 (1.5)	12 (0.7)	
Others	7,199 (1.5)	7,184 (1.5)	15 (0.9)	
With college or university degree (%)	151,108 (32.4)	150,686 (32.4)	422 (24.7)	<0.001
BMI, mean (SD), kg/m ²	27.4 ± 4.8	27.4 ± 4.8	28.9 ± 4.9	<0.001
Smoking status (%)				<0.001
Current	48,996 (10.5)	48,757 (10.5)	239 (14.0)	
Former	161,651 (34.6)	160,930 (34.6)	721 (42.2)	
Never	254,506 (54.5)	253,763 (54.5)	743 (43.5)	
Missing	1,841 (0.4)	1,834 (0.4)	7 (0.4)	
Alcohol drinker status (%)				0.001
Current	429,274 (91.9)	427,711 (91.9)	1,563 (91.4)	
Former	16,682 (3.6)	16,604 (3.6)	78 (4.6)	
Never	20,387 (4.4)	20,325 (4.4)	62 (3.6)	
Missing	651 (0.1)	644 (0.1)	7 (0.4)	
Health diet score, mean (SD)	2.2 ± 0.9	2.2 ± 0.9	2.1 ± 0.9	<0.001
Household income (%)				<0.001
<18,000 (£)	19,707 (4.2)	19,638 (4.2)	69 (4.0)	
18,000-30,999 (£)	138,248 (29.6)	137,642 (29.6)	606 (35.4)	
31,000-51,999 (£)	101,721 (21.8)	101,297 (21.8)	424 (24.8)	
52,000-100,000 (£)	104,384 (22.4)	104,039 (22.4)	345 (20.2)	
>100,000 (£)	81,350 (17.4)	81,125 (17.4)	225 (13.2)	
"Do not know" or missing	21,584 (4.6)	21,543 (4.6)	41 (2.4)	
Health status (%)				
Diabetes history	24,210 (5.18%)	24,053 (5.17%)	157 (9.18%)	<0.001
Hypertension	111,708 (23.92%)	111,055 (23.87%)	653 (38.19%)	<0.001

For continuous variables, data are presented as mean (SD), and for categorical variables, data are presented as number (percentage). The level of education was classified as either higher (college/ university degree) or lower.



0.0001), and NEUT% (HR 1.17, 95% CI: 1.12–1.23, P < 0.0001) trended toward a higher kidney cancer risk, while increased levels of LYMPH% (HR 0.85, 95% CI: 0.81–0.89, P < 0.0001) showed protective effects against incident kidney cancer. Among platelet indices, we found

significant associations between kidney cancer incidence and PDW (HR 1.07, 95% CI: 1.02–1.12, P = 0.0054) and PLT (HR 1.12, 95% CI: 1.09–1.15, P < 0.001). U-shaped associations were also found between PLT (P for nonlinearity < 0.0001; Figure 2) and kidney cancer risk.

3.3 Inflammation-related markers and kidney cancer risk

We found that white blood indices were significantly positively associated with the incidence of kidney cancer. Thus, we hypothesize that inflammation status may be associated with kidney cancer. We developed four inflammation-related markers, including the systemic immune-inflammation index (SII), platelet-to-lymphocyte ratio (PLR), neutrophil-to-lymphocyte ratio (NLR), and the product of platelet count and neutrophil count (PPN). In the inflammation-related index category, increased SII (HR 1.02, 95% CI: 1.01–1.03, P < 0.0001), PLR (HR 1.02, 95% CI: 1.01–1.03, P < 0.0001), NLR (HR 1.04, 95% CI: 1.02–1.05, P < 0.0001), and PPN (HR 1.13, 95% CI: 1.11–1.16, P < 0.0001) trended toward a higher kidney cancer risk (Figure 3). U-shaped associations were also found between PLT (P for nonlinearity <0.0001; Figure 2) and kidney cancer risk. RCS models revealed a positive association between the inflammation-related index and kidney cancer risk (Figure 2). Then, we conducted

quantile regression between the inflammation-related markers and incident kidney cancer. We divided the inflammation-related index into four quartiles (Q1–Q4) (Table 2). We found that Q4 had a higher incidence of kidney cancer in all models, while *P* for trend was significantly different in model 3. In Q4, SII (HR 1.47, 95% CI: 1.29–1.68, *P* < 0.0001), PLR (HR 1.44, 95% CI: 1.26–1.64, *P* < 0.0001), NLR (HR 1.57, 95% CI: 1.36–1.81, *P* < 0.0001), and PPN (HR 1.55, 95% CI: 1.35–1.78, *P* < 0.0001) trended toward a higher kidney cancer risk compared with Q1 in model 3 (Table 2).

3.4 Stratified analyses

Based on possible effect modifiers and blood cell indices risk variables, HRs in stratified analyses were often in the same direction. In the RBC category, we found that the associations of MCV, MCHC, and RDW were stronger among older, male, white European, former smoker, current alcohol drinker, people with

	Model 1			Model 2			Model 3		
Indices	HR (95% CI)	HR (95% CI)		HR (95% CI)		P value	HR (95% CI)		P valu
Red	_	i			i			i i	
RBC	1.22 (1.16, 1.28)	⊢ ∎	⊣ <0.0001*	1.06 (1.01, 1.12)	⊨ei	0.0299	1.03 (0.97, 1.08)		0.32
HCT	1.22 (1.17, 1.28)	H +	⊣ <0.0001*	0.99 (0.93, 1.04)	н е н	0.6324	0.95 (0.90, 1.01)	, ⊢⊕-#	0.090
MCV	0.98 (0.93, 1.02)	l ⊢⊕i	0.3196	0.90 (0.86, 0.94)	Her ¦	<0.0001*	0.90 (0.86, 0.95)	⊷	<0.000
MCHC	1.00 (0.95, 1.04)	н <mark>ф</mark> н	0.8445	0.92 (0.87, 0.96)	нен¦	0.0006*	0.91 (0.87, 0.96)	⊨∎⊣¦	0.000
RDW	1.13 (1.09, 1.17)	Her	<0.0001*	1.12 (1.08, 1.16)	¦ ⊨∎+	<0.0001*	1.10 (1.05, 1.14)	¦ ⊨ei	<0.000
Immature red		1						1	
RET	1.05 (1.03, 1.07)	-	<0.0001*	1.04 (1.02, 1.06)	i ei	0.0001*	1.02 (0.99, 1.05)	i e +	0.200
RET%	1.05 (1.02, 1.07)	H	<0.0001*	1.04 (1.02, 1.06)	-	0.0002*	1.02 (0.99, 1.06)	ke⊣	0.221
NRBC	0.86 (0.76, 0.97)	⊢•–-¦	0.0148*	0.87 (0.78, 0.98)	⊢∙⊷¦	0.0272*	0.88 (0.78, 0.99)	⊢•–{	0.039
NRBC%	0.86 (0.76, 0.97)	⊢•–-¦	0.0148*	0.87 (0.78, 0.98)	⊢ ● –-	0.0272*	0.88 (0.78, 0.99)	⊢ ∙ –i	0.039
MRV	1.00 (0.95, 1.05)	н <mark>н</mark>	0.9951	0.94 (0.89, 0.98)	Hen	0.0058*	0.94 (0.90, 0.99)	Heri	0.010
MSCV	0.90 (0.86, 0.95)	HeH I	<0.0001*	0.86 (0.82, 0.91)	HeH I	<0.0001*	0.88 (0.83, 0.92)	нн İ	<0.000
IRF	1.15 (1.09, 1.20)		<0.0001*	1.12 (1.07, 1.18)	i 🛶	<0.0001*	1.02 (0.97, 1.08)	He-I	0.379
White		ł			ł				
WBC	1.04 (1.03, 1.04)	¦•	<0.0001*	1.04 (1.03, 1.05)	l.	<0.0001*	1.04 (1.03, 1.05)		<0.000
BASO	1.01 (0.96, 1.05)	н ф и	0.7269	1.02 (0.98, 1.07)	¦ H ⊜ H	0.3644	0.99 (0.95, 1.05)	⊢ a ⊢i	0.840
BASO%	0.92 (0.86, 0.98)	⊢●⊣¦	0.0117*	0.95 (0.89, 1.01)	⊨e÷¦	0.0964	0.94 (0.88, 1.00)	⊢∙∔	0.066
EO	1.07 (1.04, 1.10)	i ei	<0.0001*	1.05 (1.01, 1.09)	¦⊷i	0.0115*	1.02 (0.98, 1.07)	i¦∎-i	0.295
EO%	1.01 (0.96, 1.05)	н р и	0.8059	0.98 (0.93, 1.03)	⊢∎¦i	0.3645	0.98 (0.93, 1.03)	⊢∎¦i	0.354
MONO	1.01 (1.01, 1.02)	+	<0.0001*	1.01 (1.00, 1.02)	+	0.002*	1.01 (1.00, 1.02)	i i	0.053
MONO%	1.02 (1.00, 1.03)	Þ	0.0153*	0.98 (0.96, 1.00)	•	0.0524	0.96 (0.91, 1.01)	⊢⊕∔	0.115
NEUT	1.15 (1.13, 1.17)		<0.0001*	1.16 (1.14, 1.18)		<0.0001*	1.14 (1.11, 1.17)		<0.000
NEUT%	1.24 (1.18, 1.30)	i H	➡ <0.0001*	1.19 (1.13, 1.25)	. ⊢•	┥ <0.0001*	1.17 (1.12, 1.23)	i ⊨•	┥ <0.000
LYMPH	1.02 (1.00, 1.04)	ie.	0.0145*	1.02 (1.01, 1.04)	ien i	0.0078*	1.02 (1.00, 1.04)	ler i	0.107
LYMPH%	0.97 (0.96, 0.97)	•	<0.0001*	0.98 (0.97, 0.98)		<0.0001*	0.85 (0.81, 0.89)	Here I	<0.000
MPV	0.99 (0.95, 1.04)	H	0.7339	1.01 (0.97, 1.06)	H e H	0.6066	1.00 (0.95, 1.05)	⊢ ⊷ ⊣	0.983
PDW	1.16 (1.11, 1.21)	H#H	<0.0001*	1.08 (1.03, 1.13)	¦ ⊨ ● ⊣	0.0008*	1.07 (1.02, 1.12)	¦+++	0.005
PLT	1.03 (0.98, 1.08)	I H ● H	0.2824	1.15 (1.10, 1.20)	¦ +•-	<0.0001*	1.13 (1.08, 1.19)		<0.000
High light scatter reticulocyte percentage	1.09 (1.07, 1.12)	i i i i i i i i i i i i i i i i i i i	<0.0001*	1.09 (1.06, 1.11)	l Hel	<0.0001*	1.03 (0.99, 1.08)	¦∙	0.144
Inflammation-related Markers		1						1	
SII	1.02 (1.01, 1.03)	þ	0.0007*	1.02 (1.01, 1.03)	þ	0.0006*	1.02 (1.01, 1.03)		<0.000
PLR	1.01 (1.00, 1.03)		0.0675	1.01 (1.00, 1.03)	•	0.014*	1.02 (1.01, 1.03)		0.000
NLR	1.05 (1.04, 1.06)	•	<0.0001*	1.04 (1.03, 1.05)	•	<0.0001*	1.04 (1.02, 1.05)	ie i	<0.000
PPN	1.15 (1.13, 1.18)	i 🖬	<0.0001*	1.14 (1.12, 1.16)	i 🖷	<0.0001*	1.13 (1.11, 1.16)	i 🖬	<0.000

FIGURE 3

Linear associations between blood cell indices and incident kidney cancer. *Model 1 was a non-adjusted model that adjusted for none. **Model 2 was a minimally adjusted model adjusted for gender and age. ***Model 3 was a fully adjusted model adjusted for gender, age, race, qualification, BMI, smoking, alcohol, health diet score, household income, diabetes, and hypertension. Exposures (excluding the inflammation-related markers) were log-transformed and standardized to the Z score so that the HR represents the predicted effect of a one-SD increment. Statistical significance at *P* < 0.05.

Inflammation- related index	Model 1 HR (95% CI)	Model 3 HR (95% CI)					
Systemic immune	-inflammation ir	idex (SII)					
Q1		Reference					
Q2	0.97 (0.85, 1.12)	1.02 (0.89, 1.17)	1.06 (0.92, 1.22)				
Q3	0.94 (0.82, 1.08)	1.01 (0.88, 1.16)	1.08 (0.93, 1.24)				
Q4	1.26 (1.11, 1.43)	1.33 (1.17, 1.52)	1.47 (1.29, 1.68)				
P trend	<0.001	< 0.001	<0.001				
Platelet-to-lymph	ocyte ratio (PLR))					
Q1	Reference						
Q2	0.96 (0.84, 1.10)	1.03 (0.90, 1.18)	1.10 (0.96, 1.26)				
Q3	0.93 (0.81, 1.07)	1.03 (0.90, 1.18)	1.14 (0.99, 1.31)				
Q4	1.13 (0.99, 1.29)	1.25 (1.10, 1.43)	1.44 (1.26, 1.64)				
P trend	0.068	<0.001	< 0.001				
Neutrophil-to-lym	phocyte ratio (N	ILR)					
Q1		Reference					
Q2	1.22 (1.05, 1.42)	1.17 (1.00, 1.36)	1.14 (0.98, 1.33)				
Q3	1.46 (1.26, 1.69)	1.34 (1.16, 1.55)	1.30 (1.12, 1.51)				
Q4	1.92 (1.67, 2.20)	1.61 (1.40, 1.86)	1.57 (1.36, 1.81)				
P trend	<0.001	<0.001	<0.001				
Product of platele	t count and neu	trophil count	(PPN)				
Q1	Reference						
Q2	1.12 (0.96, 1.30)	1.13 (0.98, 1.31)	1.08 (0.93, 1.25)				
Q3	1.34 (1.17, 1.55)	1.40 (1.22, 1.61)	1.29 (1.12, 1.48)				
Q4	1.62 (1.41, 1.86)	1.80 (1.57, 2.06)	1.55 (1.35, 1.78)				
P trend	< 0.001	< 0.001	< 0.001				

TABLE 2 Quantile regression between inflammation-related markers and incident kidney cancer.

Model 1 was a non-adjusted model that adjusted for none. Model 2 was a minimally adjusted model adjusted for gender and age. Model 3 was a fully adjusted model adjusted for gender, age, race, qualification, BMI, smoking, alcohol, health diet score, household income, diabetes, and hypertension.

college or university degree, no diabetes, hypertension, and BMI >25 participants. In the immature red category, MSCV and MRV showed significant associations among the above-mentioned specific groups, while NRBC and IRF were not remarkable. In the WBC category, we found that associations of WBC, NEUT, NEUT%, LYMPH%, and PDW had similar trends in both age and gender, while PLT had stronger associations in the older participants. For the inflammation-related index category, SII, PLR, NLR, and PPN had similar results to the RBC category (Supplementary Tables S2–S5).

3.5 Sensitivity analyses

In sensitivity analyses, we found that the association between the RBC category and kidney cancer risk lost significance in model 3, including MCV, MCHC, and RDW, while the association between RDW and kidney cancer risk remained significant in model 1 and model 2. For the immature RBC category, MRV maintained a significant difference and IRF regained a significant difference in model 3. For the WBC category, EO%, NEUT%, and PLT were significantly associated with kidney cancer risk. The inflammation-related markers SII and PPN had a stable association (Figure 4).

4 Discussion

To the best of our knowledge, this is a comprehensive prospective investigation of the connection between 27 blood cell indices and incident kidney cancer. The study utilized data from a substantial cohort of 466,994 participants from the UK Biobank, with an average follow-up period of 12.4 years. We confirmed that three blood cell indices (MCHC, RDW, and PDW) and two inflammation-related markers (SII and PPN) were associated with the risk of kidney cancer. However, further investigation is needed to elucidate the underlying mechanisms responsible for these associations. In the future, we can predict tumor risk by building disease prediction models based on blood cells. At the same time, it provides another strategy for population screening, and further detection of outliers in the blood cell index population could exclude the risk of tumor occurrence. Finally, our research can guide the direction of future basic research. Various exposure factors may promote the development or progression of tumors by mediating the circulation of blood cells.

The detection, classification, and function of circulating immune cells are helpful to further understand the complex mechanism of tumors. The changes in hematological indexes reflect the changes of the internal environment in the body, which may provide necessary conditions for the occurrence and development of tumors. Hypoxia plays an important role in cancer progression. Dynamin-related protein 1 (Drp1) was an important protein that controls the quality of mitochondria and cellular processes through alterations in its oligomeric structure and other modifications that may be associated with hematological index changes (37). Single-cell RNA sequencing could provide a new insight into the tumor immune microenvironment (38, 39). Emerging nano-/biotechnology could drive oncolytic virusactivated and combined cancer immunotherapy through hematological mediation, and some supramolecular biomaterials could also be applied for cancer immunotherapy (40-43). At present, more and more detection techniques based on multi-

Indices	Model 1			Model 2			Model 3		
	HR (95% CI)		P value	HR (95% CI)		Pvalue	HR (95% CI)		Pvalu
Red		1			1			1	
RBC	1.10 (0.86, 1.39)	⊢ ∙–−1	0.4492	0.93 (0.73, 1.20)	⊢•¦i	0.5868	0.91 (0.71, 1.17)	Let I	0.462
HCT	1.23 (0.97, 1.56)	⊨ •	0.0943	0.96 (0.74, 1.25)	⊢∎ <mark>i</mark> i	0.7754	0.95 (0.73, 1.23)	⊢•¦	0.687
MCV	1.20 (0.95, 1.51)	Ļ.	0.121	1.07 (0.84, 1.36)	⊢, ⊢,	0.5912	1.09 (0.86, 1.38)	⊢i•	0.5
MCHC	1.12 (0.94, 1.34)	i¦∙⊸i	0.1962	1.05 (0.85, 1.29)	⊢⊷	0.678	1.05 (0.87, 1.27)	⊢∲⊷⊣	0.587
RDW	1.20 (1.05, 1.37)	¦⊷⊷	0.007*	1.19 (1.02, 1.38)	⊢ •1	0.0268*	1.17 (1.00, 1.37)	⊢ •−1	0.055
Immature red									
RET	1.28 (1.06, 1.55)	i⊷⊷i	0.0119*	1.28 (1.04, 1.58)	¦ ⊢ ∙⊸i	0.0202*	1.20 (0.96, 1.51)	Ļ.	0.112
RET%	1.22 (1.05, 1.42)	⊷⊷-i	0.0103*	1.25 (1.06, 1.48)	⊷−	0.0079*	1.20 (0.99, 1.45)	⊢ •−-1	0.068
NRBC	1.00 (0.76, 1.31)	⊢∳→	0.9867	1.02 (0.78, 1.32)	⊢∔ −−1	0.9116	1.01 (0.78, 1.31)	⊢∔ i	0.95
MRV	1.37 (1.11, 1.68)	⊢ •−	H 0.0033*	1.25 (1.01, 1.55)	⊢ •−−1	0.0443*	1.27 (1.02, 1.57)	—	0.029
MSCV	1.04 (0.79, 1.35)	⊢ ⊨ ⊣	0.7947	0.96 (0.74, 1.26)	⊢ e ii	0.7942	1.02 (0.78, 1.34)	⊢ ∳ i	0.866
IRF	1.51 (1.17, 1.97)		0.0019*	1.47 (1.13, 1.91)	●	- 0.0041*	1.37 (1.04, 1.81)		- 0.027
White					1				
WBC	1.05 (0.97, 1.14)	l k a t	0.2183	1.05 (0.95, 1.16)	⊢ H e H	0.3662	1.03 (0.89, 1.19)	⊢ <mark>,</mark>	0.68
BASO	0.86 (0.63, 1.19)	Let I	0.372	0.86 (0.62, 1.20)	Let I	0.3839	0.84 (0.60, 1.17)	Let I	0.29
BASO%	0.69 (0.45, 1.08)	⊢ e _i	0.106	0.73 (0.47, 1.13)	⊢ • ∔·	0.1528	0.73 (0.48, 1.13)	L .	0.16
EO	0.90 (0.69, 1.17)	⊢∎¦-i	0.4337	0.85 (0.65, 1.12)	⊢•÷	0.2587	0.82 (0.62, 1.09)	⊢• i	0.16
EO%	0.73 (0.54, 1.00)	⊢ ● –ĺ	0.0507	0.71 (0.52, 0.97)	⊢∙−−¦	0.0309*	0.70 (0.51, 0.96)	⊢∙−-¦	0.026
MONO	1.06 (0.95, 1.18)	I H ∎ I	0.3249	1.00 (0.78, 1.27)	н е н	0.9908	0.96 (0.73, 1.26)	⊢ ≜ ⊣	0.77
NEUT	1.26 (1.10, 1.46)		0.0012*	1.21 (1.04, 1.41)	●	0.0165*	1.17 (0.99, 1.38)		0.073
NEUT%	1.50 (1.17, 1.91)	.	0.0011*	1.37 (1.07, 1.74)		H 0.0113*	1.37 (1.07, 1.75)	⊢	- 0.011
LYMPH	0.71 (0.37, 1.39)	⊢ ● ¦ → I	0.3233	0.87 (0.48, 1.58)	⊢ ●¦ →	0.6568	0.75 (0.39, 1.45)	⊢ ● − −	0.388
MPV	0.89 (0.70, 1.14)	⊢∎¦⊣	0.3618	0.90 (0.70, 1.15)	⊢ ● ¦ I	0.4019	0.87 (0.68, 1.12)	⊢ ●	0.27
PLT	1.09 (0.88, 1.36)	H•	0.4282	1.19 (1.04, 1.37)	⊨●1	0.0106*	1.21 (1.06, 1.39)		0.00
Inflammation-related Markers		1							
SII	1.10 (1.02, 1.19)	}ei	0.0104*	1.09 (1.01, 1.17)	}⊕i	0.0345*	1.09 (1.01, 1.18)	}⊷i	0.033
PLR	1.01 (0.89, 1.16)	⊢∳-i	0.8286	1.02 (0.90, 1.16)	ı.∳-i	0.7463	1.02 (0.91, 1.14)	i 🕌	0.719
NLR	1.02 (0.90, 1.15)	H H H	0.8055	1.01 (0.85, 1.21)	н і н	0.8936	1.01 (0.87, 1.17)	Let I	0.86
PPN	1.22 (1.06, 1.40)	। '⊢●-1	0.0062*	1.22 (1.10, 1.35)	, , ⊨⊕-i	0.0002*	1.24 (1.11, 1.38)	, , ⊢∎-1	0.000

FIGURE 4

Sensitivity analyses between blood cell indices and incident kidney cancer. Model 1 was a non-adjusted model that adjusted for none. Model 2 was a minimally adjusted model adjusted for gender and age. Model 3 was a fully adjusted model adjusted for gender, age, race, qualification, BMI, smoking, alcohol, health diet score, household income, diabetes, and hypertension. Exposures (excluding inflammation-related markers) were log-transformed and standardized to the Z score so that the HR represents the predicted effect of a one-SD increment. Statistical significance at *, P < 0.05.

omics have played an important role in the diagnosis, treatment, and identification of tumors. Our study highlights the possible role of immune-associated cells in the development of kidney cancer.

It is widely reported that MCHC plays an important role in regulating the development and progression of several cancers, such as non-small cell lung cancer (44), prostate cancer (45), oral squamous cell carcinoma, and head and neck cancers (46). We found that a decreased level of MCHC was significantly correlated with a higher kidney cancer incidence at first. MCHC is an important index of anemia, and anemia is hypothesized to be an independent adverse prognostic indicator in patients with kidney cancer (20, 47). Behind anemia, the accompanying hypoxia may play a critical role. Hypoxia, a condition in which tissues are oxygen-deprived, upregulates the expression of hypoxia-inducible factor (HIF), which then induces hundreds of genes in an HIFdependent manner to encode proteins that play key roles in numerous aspects of cancer biology, such as proliferation, cell survival, epithelial-to-mesenchymal transition (EMT), angiogenesis, invasion, and metastasis (48). Consequently, we infer that MCHC impacts kidney cancer risk, possibly through anemia and HIF pathways (49).

RDW, which reflects erythrocyte volume and size heterogeneity, is regarded as a strong and independent risk factor for death in the general population and is expected to be an effective indicator (50). In addition, it is generally recognized that RDW is a risk factor for various diseases, such as cardiovascular disease, venous thromboembolism, diabetes, and so on (50). Actually, we found that RDW was positively associated with kidney cancer risk, which supports RDW as a potential diagnosis marker for kidney cancer.

Platelets have been reported to influence cancer progression, metastasis, and angiogenesis in multiple ways, such as protecting tumor cells from high shear forces in the blood circulation and leukocyte attack and binding to C-type lectin-like receptor 2 (CLEC-2) to facilitate hematogenous cancer metastasis and cancer-associated thrombosis (12, 51). It is well documented that a higher platelet count is linked with shorter disease-specific survival in kidney cancer (52). PDW reflects the variation of platelet volume in the blood. The results demonstrated that elevated PDW is linked with a high kidney cancer incidence, which demonstrated that PDW might be an effective risk factor for kidney cancer.

While previous research discovered that the inflammation marker SII was connected with kidney cancer risk (53), the

association of PPN and kidney cancer has not been clear. Our paper showed that both SII and PPN were related to kidney cancer risk. Inflammatory conditions can exist in some types of cancer prior to the development of a malignant transformation. In other types of cancer, an oncogenic alteration causes an inflammatory microenvironment which improves cancer cell growth (54, 55). However, inflammatory conditions can exist in some types of cancer prior to the development to a malignant transformation, and chronic inflammation is generally acknowledged as an independent risk factor for the development of a majority of cancers (56). Another study suggests that inflammatory pathways can promote kidney cancer cell growth and immune evasion (34), which may provide theoretical bases for inflammation markers acting as risk factors for kidney cancer.

Several studies have shown a strong association between SII and kidney cancer. A study showed that SIRI and SII indexes show a moderate efficiency to show metastases in RCC (57). A large multicenter longitudinal study showed that SII increases the risk of total and cause-specific mortality among patients with chronic kidney disease, including kidney cancer (58). SII could also predict the survival of patients with renal cell cancer treated with nivolumab (59). Now, most articles have studied the relationship between SII and renal cancer progression and response to treatment. However, the incidence of kidney cancer and inflammation-related markers still requires further research.

There are several advantages to this study. First, this is a comprehensive examination of the association between blood indices and incident renal cancer. Second, owing to the large sample size, lengthy follow-up period, and extensive measurement of covariates, our study is longitudinal and possesses distinct advantages. Third, we summarized the mechanisms underpinning the association between blood indices and kidney cancer, which provides strong evidence for clinical application.

There are inevitably some limitations and deficiencies. Because these blood indices may be associated with health status and are susceptible to other factors such as comorbidities, socioeconomic status, and poor health, it is not feasible for this analysis to rule out detection bias. Then, the identification of UKB is determined by the ICD classification. Regrettably, the ICD categorization system relies on morphology rather than pathology, making it impossible to differentiate the relative risk of various blood cells for different forms of kidney cancer based on their pathological characteristics. In addition, the majority of UKB participants are European, which necessitated the use of stratified analyses that limited the generalizability of our results.

Additional limitations can be due to the fact that the pathogenesis of renal cancer is still unclear, and many factors influence blood cell indices. Hypertension and diabetes were the main risk factors of kidney cancer that could affect blood cell indices. Cigarette smoking and obesity are also likely to affect kidney cancer incidence trends and also included in the covariates (60–62). However, inadequate adjustment of covariates may lead to a bias in this study. Thus, establishing a larger cohort associated with renal cancer will help further explore the role of circulating immune cells. The simultaneous use of high-throughput

sequencing and more detailed case data could also help further understand the specific mechanisms of kidney cancer progression. Establishing a kidney cancer-related cohort based on a diverse population is necessary to improve generalizability.

5 Conclusion

This study identified three blood cell indices and two inflammation-related markers as dependent risk factors for kidney cancer incidence. As these indexes could be obtained through routine blood tests, they would be useful in large-scale screening or primary care settings to help discover individuals who might benefit from early screening or targeted prevention strategies for kidney cancer. Additional research is necessary to further illustrate the underlying mechanisms by which these blood cell indices and inflammatoryrelated markers are associated with kidney cancer risk.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

Ethics statement

This research was conducted under the UK Biobank application 61083. The UK Biobank study was approved by the National Health and Social Care Information Management Board and the North West Multicenter Research Ethics Committee (11/NW/0382). The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

QH: Conceptualization, Data curation, Formal analysis, Methodology, Software, Visualization, Writing – original draft, Writing – review & editing. CW: Conceptualization, Data curation, Formal analysis, Methodology, Writing – review & editing. LC: Validation, Writing – review & editing. PZ: Validation, Writing – review & editing. WZ: Conceptualization, Funding acquisition, Methodology, Supervision, Writing – review & editing. FC: Conceptualization, Funding acquisition, Methodology, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This research was funded by the Natural Science Foundation of Fujian Province (No.2020J01204, No.2021J01276 and No.2022J01273); The joint funds for the innovation of science and technology, Fujian

province (No.2023Y9228 and No.2023Y9243); The Quanzhou City Science & Technology Program of China, No.2023C008YR.

Acknowledgments

All authors express much thanks to Haomin Yang for his technical guidance and data support. CW is much thankful to Dr. Qi Xiong for her support and help.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Turajlic S, Swanton C, Boshoff C. Kidney cancer: the next decade. J Exp Med. (2018) 215:2477–9. doi: 10.1084/jem.20181617

2. Johansson ME, Axelson H. Kidney cancer. Semin Cancer Biol. (2013) 23:1-2. doi: 10.1016/j.semcancer.2012.05.006

3. Miller KD, Goding Sauer A, Ortiz AP, Fedewa SA, Pinheiro PS, Tortolero-Luna G, et al. Cancer statistics for hispanics/latinos, 2018. *CA Cancer J Clin.* (2018) 68:425–45. doi: 10.3322/caac.21494

4. Bahadoram S, Davoodi M, Hassanzadeh S, Bahadoram M, Barahman M, Mafakher L. Renal cell carcinoma: an overview of the epidemiology, diagnosis, and treatment. *G Ital Nefrol.* (2022) 39:3–18.

5. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* (2021) 71:209–49. doi: 10.3322/caac.21660

6. Motzer RJ, Jonasch E, Agarwal N, Alva A, Baine M, Beckermann K, et al. Kidney cancer, version 3.2022, nccn clinical practice guidelines in oncology. J Natl Compr Cancer Netw JNCCN. (2022) 20:71–90. doi: 10.6004/jnccn.2022.0001

7. Siegel RL, Giaquinto AN, Jemal A. Cancer statistics, 2024. CA: Cancer J Clin. (2024) 74:12–49. doi: 10.3322/caac.21820

8. Cirillo L, Innocenti S, Becherucci F. Global epidemiology of kidney cancer. Nephrol Dialysis Transplant Off Publ Eur Dialysis Transplant Assoc - Eur Renal Assoc. (2024) 10:1–9. doi: 10.1093/ndt/gfae036

9. Owens B. Kidney cancer. Nature. (2016) 537:S97. doi: 10.1038/537S97a

10. Capitanio U, Bensalah K, Bex A, Boorjian SA, Bray F, Coleman J, et al. Epidemiology of renal cell carcinoma. *Eur Urol.* (2019) 75:74-84. doi: 10.1016/j.eururo.2018.08.036

11. Stewart GD, Klatte T, Cosmai L, Bex A, Lamb BW, Moch H, et al. The multispeciality approach to the management of localised kidney cancer. *Lancet*. (2022) 400:523–34. doi: 10.1016/S0140-6736(22)01059-5

12. Schlesinger M. Role of platelets and platelet receptors in cancer metastasis. J Hematol Oncol. (2018) 11:125. doi: 10.1186/s13045-018-0669-2

13. Yang L, Huang S, Zhang Z, Liu Z, Zhang L. Roles and applications of red blood cell-derived extracellular vesicles in health and diseases. *Int J Mol Sci.* (2022) 23:5927. doi: 10.3390/ijms23115927

14. Naderi-Meshkin H, Ahmadiankia N. Cancer metastasis versus stem cell homing: role of platelets. J Cell Physiol. (2018) 233:9167–78. doi: 10.1002/jcp.26937

15. Martins Castanheira N, Spanhofer AK, Wiener S, Bobe S, Schillers H. Uptake of platelets by cancer cells and recycling of the platelet protein cd42a. *J Thromb Haemost.* (2022) 20:170–81. doi: 10.1111/jth.15543

 Guan X, Sun Y, Zhang C. Lncrnas in blood cells: roles in cell development and potential pathogenesis in hematological Malignancies. *Crit Rev Oncol/Hematol.* (2022) 180:103849. doi: 10.1016/j.critrevonc.2022.103849

17. Alonso-Moreda N, Berral-González A, de la Rosa E, González-Velasco O, Sánchez-Santos JM, De Las Rivas J. Comparative analysis of cell mixtures deconvolution and gene signatures generated for blood, immune and cancer cells. *Int J Mol Sci.* (2023) 24:10765. doi: 10.3390/ijms241310765

18. Prokopowicz G, Życzkowski M, Nowakowski K, Bogacki R, Bryniarski P, Paradysz A. Basic parameters of blood count as prognostic factors for renal cell carcinoma. *BioMed Res Int.* (2016) 2016:8687575. doi: 10.1155/2016/8687575

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2024.1366449/ full#supplementary-material

19. Dowd AA, Ibrahim FI, Mohammed MM. Renal cell carcinoma as a cause of iron deficiency anemia. *Afr J Urol.* (2014) 20:25–27. doi: 10.1016/j.afju.2013.11.001

20. Choi Y, Park B, Kim K, Jeong BC, Seo SI, Jeon SS, et al. Erythrocyte sedimentation rate and anaemia are independent predictors of survival in patients with clear cell renal cell carcinoma. *Br J Cancer*. (2013) 108:387–94. doi: 10.1038/bjc.2012.565

21. Fernández Gómez JM, Jalón Monzón A, Alvarez Múgica M, García Rodríguez J, Miranda Aranzubía O, González Alvarez RC. Significance of anemia as an independent prognostic factor in patients with renal cell carcinoma. *Med Clin (Barc)*. (2009) 133:407–13. doi: 10.1016/j.medcli.2009.05.038

22. Rhodes EC, Parikh SP, Bhattacharyya N. Severe autoimmune hemolytic anemia with renal neoplasm. *Pediatr Surg Int.* (2014) 30:243–4. doi: 10.1007/s00383-013-3321-9

23. Hutterer GC, Krieger D, Mrsic E, Pohlmann K, Bezan A, Stojakovic T, et al. Preoperative leucocytosis, thrombocytosis and anemia as potential prognostic factors in non-metastatic renal cell carcinoma. *Anticancer Res.* (2015) 35:3463–9.

24. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature*. (2008) 454:436-44. doi: 10.1038/nature07205

25. Diakos CI, Charles KA, McMillan DC, Clarke SJ. Cancer-related inflammation and treatment effectiveness. *Lancet Oncol.* (2014) 15:e493–503. doi: 10.1016/s1470-2045(14)70263-3

26. Ma J, Li K. Systemic immune-inflammation index is associated with coronary heart disease: A cross-sectional study of Nhanes 2009-2018. *Front Cardiovasc Med.* (2023) 10:1199433. doi: 10.3389/fcvm.2023.1199433

27. Ye C, Yuan L, Wu K, Shen B, Zhu C. Association between systemic immuneinflammation index and chronic obstructive pulmonary disease: A population-based study. *BMC Pulmonary Med.* (2023) 23:295. doi: 10.1186/s12890-023-02583-5

28. Ye Z, Hu T, Wang J, Xiao R, Liao X, Liu M, et al. Systemic immuneinflammation index as a potential biomarker of cardiovascular diseases: A systematic review and meta-analysis. *Front Cardiovasc Med.* (2022) 9:933913. doi: 10.3389/ fcvm.2022.933913

29. Hu X, Shao YX, Yang ZQ, Dou WC, Xiong SC, Li X. Preoperative systemic immune-inflammation index predicts prognosis of patients with non-metastatic renal cell carcinoma: A propensity score-matched analysis. *Cancer Cell Int.* (2020) 20:222. doi: 10.1186/s12935-020-01320-w

30. Vural S, Muhtaroğlu A, Güngör M. Systemic immune-inflammation index: A new marker in differentiation of different thyroid diseases. *Medicine*. (2023) 102: e34596. doi: 10.1097/md.00000000034596

31. Islas-Vazquez L, Aguilar-Cazares D, Galicia-Velasco M, Rumbo-Nava U, Meneses-Flores M, Luna-Rivero C, et al. Il-6, nlr, and sii markers and their relation with alterations in cd8+ T-lymphocyte subpopulations in patients treated for lung adenocarcinoma. *Biology*. (2020) 9:376. doi: 10.3390/biology9110376

32. Zhang L, Liu X, Yang R, Yang Y, Chen X. The diagnostic value of the systemic immune-inflammation index for venous thromboembolism in lung cancer patients: A retrospective study. *Mediators Inflamm.* (2022) 2022:9215311. doi: 10.1155/2022/9215311

33. Holub K, Biete A. Impact of systemic inflammation biomarkers on the survival outcomes of cervical cancer patients. *Clin Trans Oncol Off Publ Fed Spanish Oncol Societies Natl Cancer Institute Mexico*. (2019) 21:836–44. doi: 10.1007/s12094-018-1991-4

34. de Vivar Chevez AR, Finke J, Bukowski R. The role of inflammation in kidney cancer. *Adv Exp Med Biol.* (2014) 816:197–234. doi: 10.1007/978-3-0348-0837-8_9

35. Fry A, Littlejohns TJ, Sudlow C, Doherty N, Adamska L, Sprosen T, et al. Comparison of sociodemographic and health-related characteristics of UK Biobank participants with those of the general population. *Am J Epidemiol.* (2017) 186:1026–34. doi: 10.1093/aje/kwx246

36. Sudlow C, Gallacher J, Allen N, Beral V, Burton P, Danesh J, et al. UK Biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. *PloS Med.* (2015) 12:e1001779. doi: 10.1371/journal.pmed.1001779

37. Hao S, Huang H, Ma RY, Zeng X, Duan CY. Multifaceted functions of drp1 in hypoxia/ischemia-induced mitochondrial quality imbalance: from regulatory mechanism to targeted therapeutic strategy. *Military Med Res.* (2023) 10:46. doi: 10.1186/s40779-023-00482-8

38. Li PH, Kong XY, He YZ, Liu Y, Peng X, Li ZH, et al. Recent developments in application of single-cell rna sequencing in the tumour immune microenvironment and cancer therapy. *Military Med Res.* (2022) 9:52. doi: 10.1186/s40779-022-00414-y

39. Yu A, Fu J, Yin Z, Yan H, Xiao X, Zou D, et al. Continuous expression of interferon regulatory factor 4 sustains cd8(+) T cell immunity against tumor. *Res* (*Washington DC*). (2023) 6:271. doi: 10.34133/research.0271

40. Fang C, Xiao G, Wang T, Song L, Peng B, Xu B, et al. Emerging nano-/ biotechnology drives oncolytic virus-activated and combined cancer immunotherapy. *Res (Washington DC)*. (2023) 6:108. doi: 10.34133/research.0108

41. Liang H, Lu Q, Yang J, Yu G. Supramolecular biomaterials for cancer immunotherapy. *Res (Washington DC)*. (2023) 6:211. doi: 10.34133/research.0211

42. Ti D, Yan X, Wei J, Wu Z, Wang Y, Han W. Inducing immunogenic cell death in immuno-oncological therapies. *Chin J Cancer Res = Chung-kuo yen cheng yen chiu.* (2022) 34:1–10. doi: 10.21147/j.issn.1000-9604.2022.01.01

43. Qian Y, Yang T, Liang H, Deng M. Myeloid checkpoints for cancer immunotherapy. *Chin J Cancer Res = Chung-kuo yen cheng yen chiu.* (2022) 34:460–82. doi: 10.21147/j.issn.1000-9604.2022.05.07

44. Qu X, Zhang T, Ma H, Sui P, Du J. Lower mean corpuscular hemoglobin concentration is associated with unfavorable prognosis of resected lung cancer. *Future Oncol (London England).* (2014) 10:2149–59. doi: 10.2217/fon.14.121

45. Adris N, Chua ACG, Knuiman MW, Divitini ML, Trinder D, Olynyk JK. A prospective cohort examination of haematological parameters in relation to cancer death and incidence: the Busselton health study. *BMC Cancer*. (2018) 18:863. doi: 10.1186/s12885-018-4775-x

46. Bhattathiri VN. Relation of erythrocyte and iron indices to oral cancer growth. *Radiother Oncol.* (2001) 59:221–6. doi: 10.1016/s0167-8140(01)00326-7

47. Magera JS, Leibovich BC, Lohse CM, Sengupta S, Cheville JC, Kwon ED, et al. Association of abnormal preoperative laboratory values with survival after radical nephrectomy for clinically confined clear cell renal cell carcinoma. *Urology.* (2008) 71:278–82. doi: 10.1016/j.urology.2007.08.048

48. Chen T, Li K, Liu Z, Liu J, Wang Y, Sun R, et al. Wdr5 facilitates emt and metastasis of cca by increasing hif-1 α Accumulation in myc-dependent and

independent pathways. Mol Ther J Am Soc Gene Ther. (2021) 29:2134-50. doi: 10.1016/j.ymthe.2021.02.017

49. Lee W-C, Chiu C-H, Chu T-H, Chien Y-S. Wt1: the hinge between anemia correction and cancer development in chronic kidney disease. *Front Cell Dev Biol.* (2022) 10:876723. doi: 10.3389/fcell.2022.876723

50. Salvagno GL, Sanchis-Gomar F, Picanza A, Lippi G. Red blood cell distribution width: A simple parameter with multiple clinical applications. *Crit Rev Clin Lab Sci.* (2015) 52:86–105. doi: 10.3109/10408363.2014.992064

51. Suzuki-Inoue K. Platelets and cancer-associated thrombosis: focusing on the platelet activation receptor clec-2 and podoplanin. *Blood.* (2019) 134:1912–8. doi: 10.1182/blood.2019001388

52. Haemmerle M, Stone RL, Menter DG, Afshar-Kharghan V, Sood AK. The platelet lifeline to cancer: challenges and opportunities. *Cancer Cell.* (2018) 33:965–83. doi: 10.1016/j.ccell.2018.03.002

53. Nøst TH, Alcala K, Urbarova I, Byrne KS, Guida F, Sandanger TM, et al. Systemic inflammation markers and cancer incidence in the UK Biobank. *Eur J Epidemiol.* (2021) 36:841–8. doi: 10.1007/s10654-021-00752-6

54. Bower JE. Cancer-related fatigue-mechanisms, risk factors, and treatments. Nat Rev Clin Oncol. (2014) 11:597–609. doi: 10.1038/nrclinonc.2014.127

55. Shinko D, Diakos CI, Clarke SJ, Charles KA. Cancer-related systemic inflammation: the challenges and therapeutic opportunities for personalized medicine. *Clin Pharmacol Ther.* (2017) 102:599–610. doi: 10.1002/cpt.789

56. Candido J, Hagemann T. Cancer-related inflammation. J Clin Immunol. (2013) 33 Suppl 1:S79–84. doi: 10.1007/s10875-012-9847-0

57. Arı E, Köseoğlu H, Eroğlu T. Predictive value of siri and sii for metastases in rcc: A prospective clinical study. *BMC Urol.* (2024) 24:14. doi: 10.1186/s12894-024-01401-2

58. Lai W, Xie Y, Zhao X, Xu X, Yu S, Lu H, et al. Elevated systemic immune inflammation level increases the risk of total and cause-specific mortality among patients with chronic kidney disease: A large multi-center longitudinal study. *Inflammation Res Off J Eur Histamine Res Soc [et al]*. (2023) 72:149–58. doi: 10.1007/s00011-022-01659-y

59. De Giorgi U, Procopio G, Giannarelli D, Sabbatini R, Bearz A, Buti S, et al. Association of systemic inflammation index and body mass index with survival in patients with renal cell cancer treated with nivolumab. *Clin Cancer Res.* (2019) 25:3839–46. doi: 10.1158/1078-0432.Ccr-18-3661

60. Wang S, Lo Galbo MD, Blair C, Thyagarajan B, Anderson KE, Lazovich D, et al. Diabetes and kidney cancer risk among post-menopausal women: the Iowa women's health study. *Maturitas.* (2021) 143:190–6. doi: 10.1016/j.maturitas.2020.07.010

61. Zhang YB, Pan XF, Lu Q, Wang YX, Geng TT, Zhou YF, et al. Associations of combined healthy lifestyles with cancer morbidity and mortality among individuals with diabetes: results from five cohort studies in the USA, the Uk and China. *Diabetologia*. (2022) 65:2044–55. doi: 10.1007/s00125-022-05754-x

62. Chow WH, Dong LM, Devesa SS. Epidemiology and risk factors for kidney cancer. Nat Rev Urol. (2010) 7:245–57. doi: 10.1038/nrurol.2010.46