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GLI1-altered mesenchymal tumor involving the parietal pleura: case report and literature review

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GLI1-altered mesenchymal tumors represent a rare category of soft tissue tumors that have recently been incorporated into the classification of head and neck soft tissue tumors in the fifth edition of the World Health Organization (WHO) classification. However, their precise nature remains undefined, and they have yet to be assigned an ICD code. These tumors are predominantly located in the head and neck region and display distinctive pathological morphology and molecular characteristics. We present the first documented case of a GLI1-altered mesenchymal tumor occurring in the pleura. Microscopic examination revealed that the tumor was composed of ovoid-to-round and vaguely epithelioid cells, as well as a few spindle cells, all exhibiting a uniform morphology and organized in a nested and reticular arrangement, accompanied by a rich capillary network in the stroma. Immunohistochemical staining demonstrated positivity for CD56, S-100, and SMA. Next-generation sequencing (NGS) revealed a PTCH1-GLI1 fusion. Based on the morphological and immunophenotypic characteristics, molecular studies confirmed the diagnosis of a GLI1-altered mesenchymal tumor. At the 15-month follow-up, the patient was alive. We conducted a review of all cases of recurrence and metastasis, concluding that this type of tumor has a distinct propensity to metastasize to the lungs. The tumor exhibits malignant potential, and factors such as its occurrence outside the head and neck region, high-grade histological morphology, active mitosis (>5/10HPF), necrosis and PTCH1-GLI1 fusion are all considered potential risk factors.

KEYWORDS

GLI1, mesenchymal tumor, *PTCH1-GLI1* fusion, parietal pleura, malignant potential, risk factors

Introduction

In 2004, Dahlen et al. collected a unique cohort of soft tissue tumors located in the tongue (3 cases), stomach (1 case), and lower leg (1 case). These tumors were morphologically characterized by a lobulated structure composed of oval and short spindle cells, enriched with a network of thin-walled vessels. Immunohistochemical analysis revealed positivity for SMA and Laminin, while electron microscopy observations indicated differentiation features consistent with pericytes. Molecular results demonstrated the presence of the ACTB::GLI1 fusion. No recurrence or metastasis was observed during a median follow-up period of 24 months. Based on these findings, the researchers classified this group of tumors as benign tumors within the lineage of perivascular cell tumors and designated them as "Pericytoma with t(7;12)" (1). Subsequently, additional reports emerged documenting cases at various anatomical sites and identifying different GL11 fusion partner genes, with some cases exhibiting recurrence and metastasis. In the fifth edition of the WHO Classification of Head and Neck Tumors, this tumor type was included and renamed GLI1-altered mesenchymal tumors. Due to its broad morphological and immunohistochemical spectrum and the absence of a clear differentiation pathway, it is currently classified as a tumor with uncertain differentiation. This report presents the first documented case of a GLI1-altered mesenchymal tumor located in the pleura (2).

Case presentation

The patient is a 34-year-old female who was admitted to the thoracic surgery department of our hospital on September 17, 2023, after the detection of a pleural nodule during a physical examination conducted more than a year earlier. During her hospitalization, a chest CT scan revealed a well-defined nodule in the left pleura, measuring approximately 22 mm in length, which was likely of pleural origin (Figure 1A). After ruling out any surgical contraindications, a resection of the left pleural nodule was performed on September 22, 2023.

Pathological examination revealed a nodular object measuring $25 \times 18 \times 15$ mm, characterized as solid cystic with a gray-red coloration. Microscopic observation showed clearly defined tumor boundaries (Figure 1B), with the fibrocystic wall tissue and solid areas displaying nested (Figure 1C), reticulated (Figure 1D), and nodular patterns. The tumor cells were composed of monomorphic ovoid-to-round and vaguely epithelioid cells (Figure 1E), as well as a few spindle cells (Figure 1F). The cytoplasm ranged from paleeosinophilic to clear. The nuclei were round to oval, with fine chromatin, and the nucleoli were either inconspicuous or small, with occasional mitotic figures observed (< 1/2 mm²). The tumor cells were interspersed within a well-developed arborizing capillary network, and no significant necrosis was detected. Immunohistochemical staining results showed diffuse positivity for CD56 (Figure 1G) and focal

positivity for S-100 (Figure 1H) and SMA, while AE1/AE3, Synaptophysin, Chromogranin A, CD34, Desmin, Calretinin, WT-1, and TTF1 all yielded negative results. The Ki-67 labeling index was approximately 2%. FISH analysis revealed GLI1 gene break-apart signals, indicating positive evidence for GLI1 rearrangement, primarily presenting as single green atypical signals (Figure 1I); the abnormal GL11 signals were present in 60% of the cells. Targeted RNA sequencing revealed the presence of a PTCH1-GLI1 fusion (Figure 2). The identified fusion in this case consists of the 3' end of exon 1 (5' UTR region) of PTCH1 and the 5' end of exon 5 of GL11, preserving the FOXP coiled-coil domain (FOXP-CC) as well as the DNA-binding zinc finger domains (C2H2 Zn fingers) of GLI1. These morphological and molecular findings support the diagnosis of a GLI1-altered (PTCH1-GLI1 fusion) mesenchymal tumor. During a 15-month follow-up time, the patient did not develop recurrent and metastatic disease.

Discussion

GLI1-altered mesenchymal tumors represent a rare category of neoplasms, with only 90 cases documented in the English literature to date (Supplementary Table). These tumors predominantly affect young and middle-aged adults, with a median age of onset at 40 years, and they exhibit no significant gender predisposition. Approximately 33% (30/90 cases) of the lesions occurred in the head and neck region, followed by the limbs, especially the thighs, which was basically consistent with previous studies (3–5). Less common sites include the stomach (1, 6), intestines (7, 8), ovaries (9, 10), uterus (11, 12), lungs (13), kidneys (11), and bone (14, 15). In this report, we present the first documented case of a *GLI1*-altered mesenchymal tumor arising in the pleura. The clinical manifestations of these tumors vary by location, with the majority of cases exhibiting slow and painless growth.

GLI1-altered mesenchymal tumors display complex and variable morphological structures and cytological features. These tumors typically present as lobulated formations, with tumor cells arranged in sheets, nests, and occasionally in a sieve-like, reticular, microcystic, pseudoglandular, papillary and fascicular patterns (16). The cellular morphology is relatively uniform, primarily characterized by epithelioid, round, oval, or spindle. The cytoplasm can be eosinophilic or clear, while the cell nuclei are round, featuring fine chromatin and small, pinpoint nucleoli. In some instances, highgrade morphology is observed, which is characterized by pleomorphic spindle cells and focal epithelioid cells (12). The number of nuclear divisions varies from 0 to 5 per mm², with a few cases exhibiting counts as high as 40 per 10 high-power fields (HPF) (12, 17). The protrusion of tumor cells into the vascular space is a common phenomenon. Additionally, there are rich dendritic vascular networks present in the stroma, and in certain cases, mesenchymal mucosis or hyalinosis may be observed (4, 5). Rare instances of necrosis have also been noted (18). The case presented demonstrates a typical histological structure, cellular characteristics, and a rich stroma of dendritic vascular networks. GLI1-altered mesenchymal tumors exhibit non-specific immunohistochemical characteristics. Most cases demonstrate expression of CD56, while

Abbreviations: WHO, World Health Organization; FISH, Fluorescence *in situ* hybridization; CT, Computed Tomography; HPF, High-power fields; NGS, Next generation sequencing.



FIGURE 1

Clinical and pathological findings. (A) Chest CT revealed a nodule on the left parietal pleura with well-defined boundaries. (B) Microscopically, the tumor also exhibited clear boundaries, with mesothelial cells visible at the periphery (4x). (C) The tumor cells were arranged in a nested pattern (10x). (D) Reticulated pattern (4x). (E) Monomorphic round cells (40x). (F) Focal spindling (40x). (G) Tumors are immunoreactive for CD56 (10x). (H) Focal expression of S-100 (10x). (I) FISH revealed positive *GL11* gene break-apart signals, primarily manifesting as single green atypical signals (100x).

S-100 expression varies, and some cases also express SMA, AE1/AE3, CD10, cyclin D1, and BCOR (3, 11). In instances of *GLI1* amplification, the expression of MDM2, CDK4, and STAT6 may also be detected (13). Other markers, such as CD34, ERG, Synaptophysin, Chromogranin A, desmin, calponin, and SOX10, are consistently negative. The positivity for CD56, S-100, and SMA in this case aligns with findings from previous studies. Notably, the immunohistochemical phenotype lacks specificity; therefore, reliance on positive immune responses to these relatively non-specific markers may result in misdiagnosis. Parrak et al. reported that GLI1 immunohistochemical expression has a sensitivity of 91% and a specificity of 98% for diagnosing *GLI1*-altered mesenchymal tumors. GLI1 expression is localized in the cytoplasm and nucleus or may be confined solely to the nucleus (19). In the absence of *GLI1* molecular testing, GLI1 immunohistochemistry can serve as a

valuable diagnostic adjunct, aiding in the differentiation of *GLI1*-altered mesenchymal tumors.

Molecular events associated with *GL11*-altered mesenchymal tumors include *GL11* gene fusion and amplification. A total of 76% (68/90 cases) were classified as fusion, which included one case of both amplification and fusion; however, the specific fusion site remained unclear (3). In the remaining three cases, GL11 fusion was identified through FISH, although the fusion partner was not determined. Notably, sixty-six percent (42/64 cases) of the identified fusion partners were *ACTB*, followed by *MALAT1*, *PTCH1*, and several rarer partners such as *APOD*, *DERA*, *SYT*, *NCOR2*, *PAMR1*, *DDIT3*, *KDM2B* and *TUBAIA* (3, 20). Occasionally, GL11 can also be fused with two genes simultaneously (21). These various fusion genes retain the FOXP coiled-coil domain and the DNA-binding zinc finger domain



located at the 3' end of the GLI1 gene. However, the promoter at the 5' end of the GLI1 gene is replaced by the highly active promoter of the fusion partner, which results in elevated expression of GLI1 and subsequent activation of downstream gene transcription, thereby promoting tumor development (22, 23). Notably, the fusion of GLI1 with different partner genes yields similar pathological morphology, immunohistochemical profiles (3). Additionally, approximately 26% (23/90 cases) demonstrated GLI1 gene amplification, often accompanied by the co-amplification of genes located near the 12q13-15 locus, including DDIT3, CDK4, MDM2, STAT6, and HMGA2. This amplification can lead to the overexpression of the corresponding proteins, which can be identified through immunohistochemical methods (13). In this case, GLI1 gene break-apart signals were successfully detected using FISH technology. NGS revealed a PTCH1-GLI1 fusion and the combined morphology consistently pointed to GLI1-altered mesenchymal tumor.

Surgery serves as the primary treatment for *GL11*-altered mesenchymal tumors, while inhibitors of the sonic hedgehog signaling pathway may offer potential targeted therapy for tumors exhibiting *GL11* activation (24, 25). Most *GL11*-altered mesenchymal tumors demonstrate an indolent clinical course following complete surgical resection. In this case, the patient remained stable at 15 months post-surgery. Due to the limited number of cases, the 5th edition of the WHO classification for Head and Neck tumors does not provide clear indications regarding

biological behavior, nor is there an associated ICD coding. We have compiled data from the English literature concerning *GL11*-altered mesenchymal tumors that have experienced recurrence and metastasis (Table 1).

In a cohort of 64 patients with follow-up data, 42% (27/64 cases) experienced recurrence and metastasis. Among these cases, 20% (13/64 cases) exhibited local recurrence, while 30% (19/64 cases) developed distant metastasis. Notably, of the patients with distant metastasis, 47% (9/19 cases) had metastasized to the lungs, including 2 patients who presented with bilateral lung metastases (7, 18). Soft tissue tumors typically metastasize through the bloodstream, and the lungs, characterized by an extensive capillary bed, provide a conducive environment for circulating tumor cells to stagnate. Furthermore, fibroblasts within the lungs may facilitate the extravasation and implantation of these circulating tumor cells by reshaping the local immune microenvironment (26). GLI1-altered mesenchymal tumors are distinguished by the protrusion of tumor cells into the vascular space, which, along with the aforementioned factors, contributes significantly to their propensity to metastasize to the lungs. Additionally, these tumors may also metastasize to lymph nodes, bone (27), liver (7), brain, intestines (11), and various soft tissue sites (28). The average age of recurrence and metastasis was 47 years, with no significant difference observed between males and females. Approximately 80% (20/25 cases) of the primary tumors were located outside the head and neck region, while the primary

TABLE 1 Summary of Recurrent and Metastatic GLI1-Altered Mesenchymal Tumors.

	Age/Sex	Site	Outcome (FU) (mo)	Pathologic Features	Mitoses	Necrosis	<i>GLI1</i> Alteration
Antonescu et al	34/F	Neck	LR, mets to LN and lung. AWD (80)	Round to epithelioid, myxoid stroma	1-5/10HPF	focal	PTCH1-GLI1
	30/F	Foot	LR, Inguinal LN met. AWD (21)	Round to epithelioid	1-5/10HPF	No	ACTB-GLI1
	79/F	Retroperitoneum	Inguinal LN mets	Round to epithelioid	1-5/10HPF	No	ACTB-GLI1
Agaram et al	39/M	Neck	LR, lung met. AWD (26)	Prominent spindle cell	>25/ 10HPF	Yes	GLI1 Amp
	51/F	Back	LR (16)	Round to epithelioid, focally increased atypia	>25/ 10HPF	Yes	GLI1 Amp
KerrDA et al	57/F	Tibia	Rib met, AWD (27)	Round to epithelioid	NA	NA	ACTB-GLI1
	62/M	Scapula	Lung (84) and soft tissue/bone (180) met, AWD	Round to epithelioid	NA	NA	ACTB-GLI1
Prall OWJ et al	73/M	Jejunum	Multiple mets, AWD (312)	Epithelioid and spindled	30/2mm ²	focal	MALAT-GLI1
Alwaqf RR et al	54/F	Ovary	Colonic mesentery met, AWD (49)	Epithelioid	39/10HPF	Yes	PTCH1-GLI1
Liu J et al	8/M	Mouth floor	LR (27 and 40)	Round to epithelioid	NA	No	ACTB-GLI1
	1.3/M	Elbow	LR (4)	Spindled to ovoid	NA	No	<i>GLI1</i> Amp and break-apart
Zhong H et al	56/M	Lingual	mets to LN and sacral (27). AWD (36)	Round to epithelioid, myxoedematous stroma	5/10HPF	focal	<i>GLI1</i> gene break-apart Signals (FISH)
Argani P et al	33/F	Renal pelvis	LR (25)	Ovoid-spindle cells	<1/10HPF	No	<i>GLI1</i> gene break-apart Signals (FISH)
	49/F	Uterine	LR (24) and mets to brain (29). DOD (36)	Round to epithelioid (met) and spindled (LR)	16/10HPF	Yes	GLI1 Amp
	88/F	Uterine	involving the rectal adventitia	Ovoid to spindle cell, myxoid stroma	15/10HPF	focal	GLI1 Amp
Punjabi LS et al	57/F	Uterine	LR, mets to LN and lung (11). DOD (18)	Sarcomatous, pleomorphic cell	40/10HPF	NA	PAMR1-GLI1
Machado I et al	65/M	Left knee subcutaneous	skin of left tibial (1). NED (8)	Ovoid to epithelioid and spindled, microcystic	2/10HPF	focal	GLI1 Amp
	66/M	chest	Recurrence; NED(1)	Ovoid cells, focal pleomorphism	8/10HPF	No	GLI1 amp
Kerr DA et al	40/F	Right thigh	bone and soft tissue met (77). NED (89)	Ovoid to epithelioid cells, myxohyaline stroma	6/2mm ²	No	ACTB-GLI1
	71/M	NA	lung met (bilateral). AWD (16)	Ovoid to epithelioid cells, myxoid stroma	1/2mm ²	No	ACTB-GLI1
	31/M	Right leg	bone met (12), lung met (54). AWD (54)	Ovoid to epithelioid cells, myxoidto collagenous stroma	6/2mm ²	No	ACTB-GLI1
	56/M	Thoracic vertebra T9	LR (18). AWD (21)	Ovoid to epithelioid cells, myxoid to collagenous stroma	<1/2mm ²	No	ACTB-GLI1
	26/F	T8-T9 Epidural tumor	LR (12, 105), lung met (36, 105). AWD (108)	Ovoid cells, myxoid stroma	2/2mm ²	No	ACTB-GLI1
	46/M	Right pelvis soft tissue	LR	Ovoid to round cells, solid to sieve-like, myxoid stroma	1/2mm ²	No	ACTB-GLI1

(Continued)

	Age/Sex	Site	Outcome (FU) (mo)	Pathologic Features	Mitoses	Necrosis	<i>GLI1</i> Alteration
	30/M	NA	lung met	Ovoid to epithelioid cells, pseudoglandular, myxoid matrix	<1/2mm ²	No	ACTB-GL11
	37/F	Right neck	LR	Ovoid to epithelioid cells, corded, myxoid stroma	2/2mm ²	No	PTCH1-GLI1
Yajuan J. Liu	35/M	chest wall (multiple)	regional metastasis (involve a lymph node); NED(10)	ovoid to spindled cells	Rare	No	TUBA1A-GLI1

TABLE 1 Continued

F, female; M, male; NA, not available; FU, follow up; mo, months; LR, local recurrence; LN, lymph node; met, metastasis; AWD, alive with disease; NED, no evidence; Amp, amplification.

sites for 2 patients with distant metastasis could not be determined. Microscopically, the tumor cells were predominantly oval to epithelioid, arranged in nests with interstitial mucosis, and the number of mitotic figures varied. Among the patients with available prognostic information, 12 exhibited more than 5 mitotic images per 10 HPF, of which 83% (10/12 cases) experienced relapses and metastases. Among the 15 patients with necrosis present within the tumor, up to 67% (10/15 cases) developed recurrence and metastasis. At the molecular level, we observed that 50% (3/6 cases) of PTCH1-GLI1 tumors recurred and metastasized, while the remaining two cases had a follow-up period of less than four months, which is insufficient to reliably assess prognosis. Among the cases that recurred and metastasized, one exhibited bland morphology with unremarkable nuclear division and necrosis, yet still experienced a recurrence. This finding suggests that alterations in the PTCH1-GLI1 gene may be associated with a poor prognosis. Although the recurrence and metastasis rate was as high as 42%, the mortality rate was only 3% (2/64 cases), with both fatal cases occurring in the uterus. The tumor exhibits active mitotic figures, and one case demonstrates a high-grade sarcomatoid histological morphology. These findings suggest that this tumor possesses aggressive biological potential, and it may be more appropriate to classify it as a GLI1-altered mesenchymal tumor with malignant potential. The external location of the tumor relative to the head and neck, along with high-grade histological morphology, active mitosis (>5/10HPF), necrosis, and PTCH1-GLI1 gene alteration are all significant risk factors for recurrence and metastasis. Among these factors, high-grade histological morphology is the most aggressive. The presence of such morphology indicates that the tumor can be classified as malignant. Additionally, there was one case that experienced a recurrence without having any of the aforementioned risk factors. This case is situated outside the head and neck region, presenting a risk of recurrence and metastasis; therefore, stringent follow-up observation is essential. The statistical analysis of prognostic data is complicated by the variability in follow-up periods, as some cases have a brief follow-up duration while others extend beyond ten years before recurrence or metastasis occurs. This disparity may introduce bias into the data, thereby compromising its accuracy. To achieve a more precise assessment of prognosis, a larger sample size and an extended follow-up period are imperative. In our study, both tumor location and molecular changes are identified as risk factors, and we will conduct close follow-up.

The differential diagnosis includes: (1) Mesothelioma: This case occurred in the pleura, with chest CT suggesting a possible pleural origin. Classic epithelioid mesothelioma is characterized by papillary, trabecular, and solid arrangements, exhibiting marked cellular atypia and pleomorphism. It is positive for mesothelial markers in immunohistochemistry, including Calretinin, WT1, and D2-40. (2) Glomus tumor: Glomus tumors share morphological similarities with GLI1-altered mesenchymal tumors, but they diffusely express SMA and Caldesmon in histochemistry, with molecular alterations characterized by NOTCH gene fusion. (3) Neuroendocrine tumor: Comprised of morphologically uniform cells with rich stromal vasculature, these tumors can be confused with neuroendocrine tumors. Differentiation can be achieved through immunohistochemistry, as neuroendocrine tumors characteristically exhibit diffuse positivity for neuroendocrine markers such as CD56, Synaptophysin, and Chromogranin A. (4) Ectomesenchymal chondromyxoid tumor: This tumor typically occurs on the dorsal side of the tongue and exhibits morphology similar to GLI1-altered mesenchymal tumors characterized by interstitial mucinous change. Notably, tumor cells generally express GFAP and S-100, with the molecular alteration identified as the RREB1-MKL2 fusion. (5) Other GL11 gene-altered tumors include gastroblastoma, gastric plexiform fibromyxoma, and liposarcoma, each of which presents distinct morphological and histochemical features.

Conclusion

GLI1-altered mesenchymal tumors are rare neoplasms characterized by distinct morphological and molecular features, primarily found in the head and neck, although they can also occur in other anatomical locations. These tumors are associated with a significant risk of recurrence and metastasis; even those with benign cellular characteristics may exhibit metastatic behavior, particularly to the lungs. Key risk factors for recurrence and metastasis include tumor location outside the head and neck, high-grade histological morphology, active mitosis (>5/10HPF), necrosis, and PTCH1-GLI1 gene alteration. Notably, high-grade histological morphology correlates with the most aggressive behavior of these tumors. As a result, they possess a considerable potential for malignancy and may sometimes exhibit malignant biological characteristics. Therefore, close follow-up is essential, making accurate diagnosis and continuous monitoring of these tumors critically important.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

Ethics statement

The studies involving humans were approved by the Research Ethics Committee for Biomedical Research Involving Humans (Ethics approval number: K2024145); the Fourth Affiliated Hospital of School of Medicine, and International School of Medicine, International Institutes of Medicine, Zhejiang University. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Author contributions

YZ: Conceptualization, Data curation, Writing – original draft, Writing – review & editing, Investigation, Methodology, Project administration. BL: Supervision, Writing – review & editing. GW: Supervision, Writing – review & editing. YL: Data curation, Writing – review & editing. ZC: Conceptualization, Methodology, Project administration, Writing – review & editing.

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Conflict of interest

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Supplementary material

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References

1. Dahlén A, Fletcher CD, Mertens F, Fletcher JA, Perez-Atayde AR, Hicks MJ, et al. Activation of the GLI oncogene through fusion with the beta-actin gene (ACTB) in a group of distinctive pericytic neoplasms: pericytoma with t(7;12). *Am J Pathol.* (2004) 164:1645–53. doi: 10.1016/s0002-9440(10)63723-6

2. Jo VY, Demicco EG. Update from the 5th edition of the World Health Organization classification of head and neck tumors: soft tissue tumors. *Head Neck Pathol.* (2022) 16:87-100. doi: 10.1007/s12105-022-01425-w

3. Liu J, Mao R, Lao IW, Yu L, Bai Q, Zhou X, et al. GLI1-altered mesenchymal tumor: a clinicopathological and molecular analysis of ten additional cases of an emerging entity. *Virchows Arch.* (2022) 480:1087–99. doi: 10.1007/s00428-021-03224-0

4. Xu B, Chang K, Folpe AL, Kao YC, Wey SL, Huang HY, et al. Head and neck mesenchymal neoplasms with GLI1 gene alterations: A pathologic entity with distinct histologic features and potential for distant metastasis. *Am J Surg Pathol.* (2020) 44:729–37. doi: 10.1097/PAS.00000000001439

5. Antonescu CR, Agaram NP, Sung YS, Zhang L, Swanson D, Dickson BC. A distinct Malignant epithelioid neoplasm with GL11 gene rearrangements, frequent S100 protein expression, and metastatic potential: expanding the spectrum of pathologic entities with ACTB/MALAT1/PTCH1-GL11 fusions. *Am J Surg Pathol.* (2018) 42:553–60. doi: 10.1097/PAS.00000000001010

6. Castro E, Cortes-Santiago N, Ferguson LM, Rao PH, Venkatramani R, López-Terrada D. Translocation t(7;12) as the sole chromosomal abnormality resulting in ACTB-GLI1 fusion in pediatric gastric pericytoma. *Hum Pathol.* (2016) 53:137–41. doi: 10.1016/j.humpath.2016.02.015

7. Prall OWJ, McEvoy CRE, Byrne DJ, Iravani A, Browning J, Choong DY, et al. A Malignant neoplasm from the jejunum with a MALAT1-GL11 fusion and 26-year survival history. *Int J Surg Pathol.* (2020) 28:553–62. doi: 10.1177/1066896919900548

8. Jessurun J, Orr C, McNulty SN, Hagen CE, Alnajar H, Wilkes D, et al. GLI1-rearranged enteric tumor: expanding the spectrum of gastrointestinal neoplasms with GLI1 gene fusions. *Am J Surg Pathol.* (2023) 47:65–73. doi: 10.1097/PAS.000000000001950

9. Koh NWC, Seow WY, Lee YT, Lam JCM, Lian DWQ. Pericytoma with t(7;12): the first ovarian case reported and a review of the literature. *Int J Gynecol Pathol.* (2019) 38:479–84. doi: 10.1097/PGP.00000000000542

10. Hui L, Bai Q, Yang W, Xiaoyu T, Yao Q, Zhou X, et al. GLI1-rearranged mesenchymal tumor in the ovary. *Histopathology.* (2022) 81:688–92. doi: 10.1111/his.14785

11. Argani P, Boyraz B, Oliva E, Matoso A, Gross J, Fridman E, et al. GLI1 gene alterations in neoplasms of the genitourinary and gynecologic tract. *Am J Surg Pathol.* (2022) 46:677–87. doi: 10.1097/PAS.000000000001844

12. Punjabi LS, Goh CHR, Sittampalam K. Expanding the spectrum of GLI1-altered mesenchymal tumors-A high-grade uterine sarcoma harboring a novel PAMR1::GLI1 fusion and literature review of GLI1-altered mesenchymal neoplasms of the gynecologic tract. *Genes Chromosomes Cancer.* (2023) 62:107–14. doi: 10.1002/gcc.2309

13. Agaram NP, Zhang L, Sung YS, Singer S, Stevens T, Prieto-Granada CN, et al. GLI1-amplifications expand the spectrum of soft tissue neoplasms defined by GLI1 gene fusions. *Mod Pathol.* (2019) 32:1617–26. doi: 10.1038/s41379-019-0293-x

14. Bridge JA, Sanders K, Huang D, Nelson M, Neff JR, Muirhead D, et al. Pericytoma with t(7;12) and ACTB-GLI1 fusion arising in bone. *Hum Pathol.* (2012) 43:1524–9. doi: 10.1016/j.humpath.2012.01.019

15. Kerr DA, Pinto A, Subhawong TK, Wilky BA, Schlumbrecht MP, Antonescu CR, et al. Pericytoma With t(7;12) and ACTB-GLI1 Fusion: Reevaluation of an Unusual Entity and its Relationship to the Spectrum of GLI1 Fusion-related Neoplasms. *Am J Surg Pathol.* (2019) 43:1682–92. doi: 10.1097/PAS.00000000001360

16. Cloutier JM, Kerr DA. GLI1-altered mesenchymal tumors. Surg Pathol Clin. (2024) 17:13-24. doi: 10.1016/j.path.2023.06.004

17. Alwaqfi RR, Samuelson MI, Guseva NN, Ouyang M, Bossler AD, Ma D. PTCH1-GLI1 fusion-positive ovarian tumor: report of a unique case with response to tyrosine kinase inhibitor pazopanib. *J Natl Compr Canc Netw.* (2021) 19:998–1004. doi: 10.6004/ jnccn.2021.7058 18. Kerr DA, Cloutier JM, Margolis M, Mata DA, Rodrigues Simoes NJ, Faquin WC, et al. GL11-altered mesenchymal tumors with ACTB or PTCH1 fusion: A molecular and clinicopathologic analysis. *Mod Pathol.* (2024) 37:100386. doi: 10.1016/j.modpat.2023.100386

19. Parrack PH, Mariño-Enríquez A, Fletcher CDM, Hornick JL, Papke DJ Jr. GLI1 immunohistochemistry distinguishes mesenchymal neoplasms with GLI1 alterations from morphologic mimics. *Am J Surg Pathol.* (2023) 47:453–60. doi: 10.1097/PAS.000000000002018

20. Papke DJ Jr, Dickson BC, Oliveira AM, Sholl LM, Fletcher CDM. Distinctive nested glomoid neoplasm: clinicopathologic analysis of 20 cases of a mesenchymal neoplasm with frequent GLI1 alterations and indolent behavior. *Am J Surg Pathol.* (2023) 47:12–24. doi: 10.1097/PAS.00000000001979

21. Wang CY, Chu YL, Lin SC, Wu CC, Huang WC, Kuo CH. Cervical myelopathy and extensive body destruction caused by primary Gli1 fusion sarcoma. *Neuropathology*. (2024) 44:236–9. doi: 10.1111/neup.12957

22. Ichikawa D, Yamashita K, Okuno Y, Muramatsu H, Murakami N, Suzuki K, et al. Integrated diagnosis based on transcriptome analysis in suspected pediatric sarcomas. *NPJ Genom Med.* (2021) 6:49. doi: 10.1038/s41525-021-00210-y

23. Spans L, Fletcher CD, Antonescu CR, Rouquette A, Coindre JM, Sciot R, et al. Recurrent MALAT1-GL11 oncogenic fusion and GL11 up-regulation define a subset of plexiform fibromyxoma. *J Pathol.* (2016) 239:335–43. doi: 10.1002/path.4730

24. Zhong J, Sun Y, Wu S, Zhang T, Yang J, He Y, et al. The impact of the Hedgehog signal pathway on the tumor immune microenvironment of gastric adenocarcinoma by integrated analysis of scRNA-seq and RNA-seq datasets. *Funct Integr Genomics*. (2023) 23:258. doi: 10.1007/s10142-023-01187-w

25. Doheny D, Manore SG, Wong GL, Lo HW. Hedgehog signaling and truncated GLI1 in cancer. *Cells.* (2020) 9:2114. doi: 10.3390/cells9092114

26. Gerstberger S, Jiang Q, Ganesh K. Metastasis. Cell. (2023) 186:1564-79. doi: 10.1016/j.cell.2023.03.003

27. Zhong H, Xu C, Chen X, Guo X, Yang S. GLI1-altered epithelioid soft tissue tumor: A newly described entity with a predilection for the tongue. *Oral Surg Oral Med Oral Pathol Oral Radiol.* (2022) 134:e14–22. doi: 10.1016/j.0000.2021.10.007

28. MaChado I, Hosler GA, Traves V, Claramunt R, Sanmartín O, Santonja C, et al. Superficial GLI1-amplified mesenchymal neoplasms: Expanding the spectrum of an emerging entity which reaches the realm of dermatopathology. *J Cutan Pathol.* (2023) 50:487–99. doi: 10.1111/cup.14345