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Hepatocellular carcinoma: pathogenesis, molecular mechanisms, and treatment advances

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Hepatocellular Carcinoma (HCC), a highly prevalent malignancy, poses a significant global health challenge. Its pathogenesis is intricate and multifactorial, involving a complex interplay of environmental and genetic factors. Viral hepatitis, excessive alcohol consumption, and cirrhosis are known to significantly elevate the risk of developing HCC. The underlying biological processes driving HCC are equally complex, encompassing aberrant activation of molecular signaling pathways, dysregulation of hepatocellular differentiation and angiogenesis, and immune dysfunction. This review delves into the multifaceted nature of HCC, exploring its etiology and the intricate molecular signaling pathways involved in its development. We examine the role of immune dysregulation in HCC progression and discuss the potential of emerging therapeutic strategies, including immune-targeted therapy and tumorassociated macrophage interventions. Additionally, we explore the potential of traditional Chinese medicine (TCM) monomers in inhibiting tumor growth. By elucidating the complex interplay of factors contributing to HCC, this review aims to provide a comprehensive understanding of the disease and highlight promising avenues for future research and therapeutic development.

KEYWORDS

hepatocellular carcinoma, immune-targeted therapy, molecular signaling pathways, macrophages, cirrhosis

1 Introduction

Liver diseases cause over 2 million deaths annually, accounting for 4% of all global deaths (1 in every 25 deaths) (1). Currently, liver diseases are the 11th leading cause of death, but liver-related mortality may have been underestimated (2).

Approximately two-thirds of all liver-related deaths occur in men (1, 3). The main causes of death are complications of cirrhosis and HCC. Liver diseases are diverse,

including hepatitis, alcoholic liver disease, metabolic dysfunctionassociated fatty liver disease(MAFLD), cirrhosis, and liver cancer. These diseases share common features such as varying degrees of inflammation and liver cell damage (4–6). In the early stages, symptoms may not be significant, but long-term accumulation can lead to cirrhosis or even liver cancer (HCC), ultimately resulting in death (7, 8).

HCC develops through a complex, multi-stage biological process. MAFLD, alcoholic liver disease, autoimmune hepatitis, hepatitis B, and hepatitis C (Figure 1) are all potential causes of HCC (9). Currently, the progression of HCC is often accompanied by genetic and epigenetic modifications, oxidative stress, inflammation, and immune involvement (10). Liver cancer stem cells (LCSC) play a critical role in cancer occurrence, metastasis, recurrence, and treatment resistance, affecting the dedifferentiation of mature hepatocytes and bile duct cells (11, 12). The loss of tumor suppressor proteins p53/p21 leads to the dedifferentiation of mature liver cells into progenitor-like cells, which further develop into HCC with gene mutations in the Wnt and Notch signaling pathways (12). The insulin-like growth factor (IGF) signaling pathway is involved in the occurrence, progression, and metastasis of HCC (13). Moreover, signaling pathways controlling growth factor receptors (such as FGFR, TGFA, EGFR, and IGFR), cytoplasmic intermediates (such as PI3K-AKT-mTOR, RAF/ERK/MAPK), and key cell differentiation pathways (such as Wnt-catenin, JAK/ STAT, Hippo, Hedgehog, and Notch) all influence the progression of HCC (14). In recent years, the greatest focus in cancer treatment has been on tumor-associated macrophages and liver cancer immunotherapy.

This review will provide a comprehensive understanding of the pathogenesis of liver diseases, current research advancements in cancer treatment, and future perspectives, offering clinical researchers a theoretical foundation and reference for potential therapeutic targets in treating liver cancer patients.



2 Pathogenesis of HCC

2.1 Alcoholic liver disease

Currently, 43% of the global population consumes alcohol, and excessive drinking is a significant risk factor for disease and death worldwide (15, 16). It is reported that alcohol increases the mortality rate related to liver disease by 260 times, cardiovascular disease by 3.2 times, and cancer by 5.1 times (17). About 35% of patients with alcohol use disorder (AUD) develop various forms of ALD (18). ALD often coexists with viral hepatitis and MAFLD. However, compared to liver diseases of other causes, alcoholic liver disease is more likely to progress to cirrhosis. Additionally, with the rising global prevalence of obesity and type 2 diabetes, alcoholrelated liver damage is increasing. Obesity and metabolic syndrome can synergistically exacerbate the severity of ALD at all stages (19).

Drinking alcohol also increases the risk of liver cancer for those who are overweight, obese, or have liver cirrhosis associated with MAFLD (20, 21). Without global interventions, ALD-related mortality is expected to rise significantly.

Alcohol metabolism in the body occurs through oxidative and non-oxidative pathways. The oxidative pathway involves alcohol dehydrogenase (ADH), microsomal cytochrome P450 enzymes (CYP450), and peroxidases.

2.2 Metabolic dysfunction-associated fatty liver disease

Currently, metabolic dysfunction-associated fatty liver disease (MAFLD) affects one-quarter of the global adult population (22, 23). MAFLD can also occur in individuals without obesity or metabolic syndrome, possibly due to certain metabolic disorders, such as insulin resistance(IR) or increased cardiovascular risk. Excess fatty acids lead to IR and liver steatosis, which eventually cause liver cell damage, inflammation, fibrosis, and other pathological changes due to oxidative stress and lipid peroxidation (24). The mechanisms of MAFLD pathology are varied, including oxidative stress, ER stress, and lipotoxicity (25).

2.2.1 Lipid accumulation

When energy intake exceeds consumption, the excess energy is stored as lipids in organs throughout the body (26–28). MAFLD is formed through ectopic lipid accumulation. The steatosis in MAFLD is triggered by the excessive synthesis of triglycerides (TG) in liver cells, with 60% of the substrates for synthesis coming from white adipose tissue (WAT), 26% from *de novo* lipogenesis (DNL), and 15% from high-fat and high-sugar diets (29, 30). Insulin has an anti-lipolytic effect, mediating the storage of TG in adipose tissue and promoting the esterification and storage of fatty acids (31). Therefore, IR is a key factor in MAFLD. In the IR state, insulin's anti-lipolytic function weakens, WAT is broken down, leading to a large release of free fatty acids (FFA) (32). Excess FFAs are then stored in the liver as TG, forming ectopic lipid deposits and leading to MAFLD (33). DNL is a key pathway for promoting lipid accumulation and is closely related to IR (34). DNL is regulated by sterol regulatory element-binding protein-1c (SREBP-1c) and carbohydrate response element-binding protein (ChREBP) (35, 36). IR activates SREBP-1c to promote DNL in liver cells (37, 38). Increased glucose concentrations activate ChREBP to regulate the expression of acetyl-CoA carboxylase (ACC) and fatty acid synthase (FAS), thus promoting DNL in liver cells (Figure 2) (39).

Dietary factors play a crucial role in the development of MAFLD (40, 41) A diet high in fats and sugars increases the expression of genes related to liver fibrosis, inflammation, ER stress, and adipocyte apoptosis (42). Animal models and human studies have shown that fructose has selective liver metabolism and triggers liver stress responses, including activating c-Jun N-terminal kinase (JNK) and IR, promoting liver fat accumulation, impairing fatty acid oxidation (FAO), and leading to liver inflammation and fibrosis (43, 44).

2.2.2 Oxidative stress

DNL converts excess carbohydrates into fatty acids, which are then esterified into triglycerides (TG) and stored in liver cells. When energy is insufficient, TG is used to supply energy through β -oxidation (45). However, an increase in FFA in the liver impairs β -oxidation and mitochondrial function, leading to oxidative stress (46).

Peroxisomes are the first enzymes in the fatty acid β -oxidation system. Peroxisome proliferator-activated receptor alpha (PPAR α) regulates the activity of three interconnected liver fatty acid oxidation systems: mitochondrial and peroxisomal β -oxidation and microsomal ω -oxidation pathways (47). Continuous activation of PPAR α can alleviate MAFLD by enhancing FAO and reducing ROS levels (48, 49). However, many studies have found that excessive activation of PPAR α leads to overconsumption of liver energy, disproportionately increasing H2O2 and triggering an inflammatory response (50).

MAFLD patients show damage to mitochondrial ultrastructure, reduced respiratory chain complex activity, and impaired ATP synthesis (51). Mitochondria play a crucial role in FAO and energy supply while producing large amounts of ROS (52). Mitochondrial dysfunction results from damage to the electron transport chain (ETC). Over-reduction of components of the mitochondrial respiratory chain leads to abnormal reactions between electrons and oxygen, increasing ROS (53). Moreover, ROS oxidize lipid deposits, releasing lipid peroxides that damage liver cells. In liver cells, ROS and lipid peroxides further disrupt the respiratory chain, directly or indirectly causing oxidative damage to the mitochondrial genome, which leads to more ROS production, creating a vicious cycle, ultimately leading to inflammation (Figure 3) (54).

2.2.3 Endoplasmic reticulum stress

ER stress represents a protective cellular reaction, triggering the unfolded protein response (UPR) in an effort to restore protein homeostasis. As lipid accumulation increases, ER stress results in a





buildup of unfolded proteins, triggering the UPR (55) The UPR is mediated by protein kinase RNA-like ER kinase (PERK), inositolrequiring enzyme 1 (IRE1), and activating transcription factor 6 (ATF6) (56), all of which regulate lipid storage in the liver (57). PERK-mediated phosphorylation of eukaryotic initiation factor 2α (eIF 2α) transiently reduces translation, while activating transcription factor 4 (ATF4) induces the expression of the gene CCAAT/enhancer-binding protein homologous protein (CHOP) (58). ATF6 and IRE1 promote the expression of X-box binding protein-1 (XBP1) and mediate inflammation through the JNK signaling pathway (59, 60). Additionally, IRE1 can directly activate JNK, which in turn activates TNF receptor-associated factor 2 (TRAF2), promoting apoptosis (61).

2.2.4 Lipotoxicity

Lipotoxicity refers to the toxic effects caused by the excessive deposition of lipids and their metabolites in non-adipose tissues (62). When the concentration of lipotoxic substances in liver cells exceeds the transport capacity of the liver, the damage to liver cells worsens, and The more that the disease advances, the more stages it will progress to. One of the reasons that IR will stem from the disease in MAFLD is due to a surge of FFA in plasma. Thus, the liver cells undergo an overload of FAO, which results in a great production of ROS, damage to mitochondria, ER stress, and the addition of inflammation. The lipotoxicity caused by lipid accumulation drives the further progression of the disease.

Liver parenchymal cells are mainly composed of hepatocytes. Hepatic stellate cells (HSCs) and Kupffer cells (KCs) are examples of non-parenchymal cells that also have a vital job in the development of NASH. Liver fibrosis is a major cause for the progression of NASH and usually stems from HSCs. It has been found that the activation of toll-like receptor 4 (TLR4) by lipotoxic substances promotes inflammation and fibrotic signaling in HSCs (63). KCs regulate inflammatory responses in the liver microenvironment and contribute to liver disease progression by secreting proinflammatory cytokines. In patients with NASH, elevated levels of oxidized LDL trigger inflammation in KCs (Figure 4) (64).

2.3 Viral hepatitis

Viral hepatitis generally includes five types: hepatitis A, B, C, D, and E. When liver enzymes increase in the serum, the liver may be under viral attack. Cytotoxic T cells play a key role in virus clearance during the acute phase of hepatitis (65).

During viral infection, the virus activates cytotoxic T lymphocytes (CD8T cells), which produce virus-specific CD8T cells. Activated virus-specific CD8T cells differentiate into effector cytotoxic T lymphocytes, which specifically kill virus-infected cells, leading to liver damage and possibly progressing to liver cancer (66, 67).

Viral hepatitis includes both acute and chronic forms. In chronic infections, the body's immune response is impaired as the target virus persists. If the host immune response fails to clear the virus, it leads to immune evasion due to genetic mutations, and viral proteins suppress immune responses. Acute hepatitis can typically



resolve on its own if the liver cell damage is within recoverable limits. However, severe acute hepatitis may require medication to prevent progression to chronic hepatitis.

2.4 Cirrhosis

Chronic liver diseases progress to cirrhosis through an intermediate stage of liver fibrosis. Liver fibrosis occurs when ECM proteins (primarily cross-linked types I and III collagen) accumulate, replacing damaged normal tissue with fibrotic scars (68). Key mechanisms of liver fibrosis include chronic hepatocyte damage, epithelial or endothelial barrier injury, the release of inflammatory cytokines, recruitment of bone marrow-derived inflammatory cells, macrophage production of TGF- β , and activation of hepatic myofibroblasts that secrete type I collagen (COL1A1) to produce excessive ECM (69).

(a) HSCs reside in the liver's space of Disse and exist in a quiescent phenotype. Quiescent HSCs are the main storage site for vitamin A (70, 71), and maintaining this quiescent phenotype is critical (72). Upon liver injury, quiescent HSCs downregulate the expression of vitamin A, GFAP, and PPARγ, becoming activated (73).

Upon stimulation by fibrogenic mediators, HSCs upregulate α smooth muscle actin and other intracellular microfilaments of myofibroblasts. Activated HSCs migrate to the injury site and secrete ECM, forming fibrotic scars (74).

TGF- β is the most potent fibrogenic cytokine (75). COL1A1, COL1A2 proteins, Activin, and Pail genes are targets of TGF- β .

Additionally, IL-6 and IL-17 can induce COL1A1 transcription through the JAK-STAT3 signaling pathway (76). Connective tissue growth factor (CTGF) and IL-13 promote COL1A1 expression in activated HSCs via the TGF- β 1 pathway81. Research shows that inflammation is an essential factor for fibrosis; signals from damaged hepatocytes alone are insufficient to directly activate HSCs and cause fibrosis.

(b) Inflammation plays a crucial role in the pathogenesis of liver fibrosis. Neutrophils are often recruited to the injured liver as the first responders to clear apoptotic liver cells (77). Neutrophils release free DNA, which has a strong pro-inflammatory effect (78). Increased levels of neutrophil chemokines (IL-8, IL-18, IL-17, CCL3, CCL4, and CXCL2) accelerate liver fibrosis progression in mice (79, 80).

The activation of macrophages capable of producing TGF- β is a key factor in liver fibrosis. Kupffer cells are the main source of TGF- β , and they possess phagocytic and anti-inflammatory functions (81). Overexpression of myeloid TGF- β genes spontaneously induces fibrosis in target tissues and organs (including the liver), indicating that TGF- β is a crucial mediator of fibrosis. Furthermore, the deletion of IL-6, TNF, or IL-1 β genes reduces liver fibrosis progression, as these cytokines synergize with TGF- β (82, 83).

(c)Viral hepatitis (especially hepatitis B and C), alcoholic liver disease, and MAFLD can all lead to liver fibrosis and eventually cirrhosis. For example, ALD is a leading cause of cirrhosis and liver failure. ALD progresses from fatty degeneration to steatohepatitis, fibrosis, and HCC (84). Alcohol induces liver injury both directly (through toxic ethanol metabolites) and indirectly (via cytochrome P4502E1, an alcohol-metabolizing enzyme) (85). Alcohol activates SREBP1 or SREBP2-dependent fatty acid and cholesterol synthesis (84), leading to the accumulation of lipid droplets in the liver, the formation of Mallory bodies in ballooned hepatocytes, and alcohol-induced liver injury (86). Alcohol-induced hepatotoxic damage is associated with the upregulation of IL-8, IL-17, CXCL1, neutrophil infiltration, recruitment of bone marrow-derived or liver macrophages to the alcohol-injured liver, TGF- β secretion, and activation of myofibroblasts, which further produce excessive ECM proteins that contribute to liver fibrosis (86–88).

3 Hepatocellular carcinoma

HCC is a cancer that originates in the cells of the liver. Its pathogenesis involves multiple molecular defects, including cell cycle dysregulation, changes in DNA methylation, chromosomal instability, immune modulation, epithelial-to-mesenchymal transition, increases in HCC stem cells, and dysregulation of miRNAs (89). While the specific mechanisms driving HCC differ based on the underlying etiology, the usual progression involves liver injury, followed by chronic inflammation, fibrosis, cirrhosis, and ultimately HCC. The release of molecular mediators, including damage-associated molecular patterns (DAMPs) and pathogenassociated molecular patterns (PAMPs), by viral particles activates pattern recognition receptors (PRRs) present on immune cells. These PRRs encompass Toll-like receptors (TLRs), C-type lectin receptors, NOD-like receptors, and retinoic acid-inducible gene I (RIG-I)-like receptors, thereby triggering inflammation. This chronic inflammation can lead to fibrosis and eventually cirrhosis (90). Research into molecular mechanisms related to the development of liver cancer has the potential to identify therapeutic targets.

3.1 IGF pathway

Aberrant IGF signaling is critically involved in the pathogenesis and carcinogenic processes of HCC, especially in insulin resistancerelated HCC. Insulin and hyperinsulinemia promote the synthesis and bioactivity of IGF-1 and IGF-2, regulating energy-dependent growth processes (91).

IGF-1 has a higher affinity for IGF-1 R, which is associated with the development of precancerous lesions (92). The binding of IGF-1 to IGF-1 R can regulate stem cell pluripotency and differentiation, triggering cell proliferation, organ development, and tissue regeneration (93). Additionally, imbalances in IGF-1/IGF-1 R signaling can activate MAPK and c-JNK pathways, promoting HCC cell proliferation and inhibiting apoptosis. IGF-1 also promotes angiogenesis by increasing the production of VEGF (94). Plasma levels of IGF-2 are elevated in patients with obesity, cirrhosis, and HCC (95). During hepatocarcinogenesis, IGF-2 exerts various carcinogenic functions by binding to IGF-1 R, such as inhibiting apoptosis, promoting HCC cell proliferation and migration, and activating angiogenesis (96). Studies indicate that IRS-1 is an oncogene with higher expression levels in HCC (97). Hyperinsulinemia and increased IGF receptor activation lead to the phosphorylation of IRS-1, triggering the activation of multiple cytokine pathways, including the PI3 K/AKT/mTOR and MAPK cascades, which regulate the cell cycle and may potentially enhance tumor progression in HCC (98).

3.2 Wnt/ β -catenin pathway

Wnt/ β -catenin signaling pathway is one of the most important pathways for cell fate differentiation and the overall maintenance of liver metabolism and homeostasis (99). In patients with cirrhosis and HCC, Wnt activity is frequently overactivated. Abnormal activation of Wnt/ β -catenin signaling is a hallmark of various liver pathologies, playing a role in nearly every aspect of liver biology (100).

At the heart of the Wnt signaling cascade lies β -catenin, a protein produced from the CTNNB1 gene. The pathway centers on the interaction between Wnt ligands and the Frizzled/LRP coreceptor complex, leading to abnormal accumulation of β -catenin in the nucleus and the expression of multiple transcriptional targets, including genes responsible for proliferation (e.g., MYC), antiapoptosis (e.g., BIRC5), epithelial-mesenchymal transition (e.g., Snail), invasion (e.g., MMP), angiogenesis (e.g., VEGF), and inflammation (e.g., IL-6) (101). β-catenin also functions in cellcell adhesion as a component of adherens junctions (100). Hepatic stellate cells (HSCs) express several Wnt receptors, with components like Wnt 3a and Wnt 5a promoting HSC activation, which is crucial in fibrosis development and progression (102). Therefore, activation of the Wnt/β-catenin pathway regulates tissue development and regeneration, as well as HCC tumorigenicity and metastatic potential (99). Increasing evidence links Wnt/β-catenin to human inflammation (e.g., HBV and HCV) and metabolic dysfunction (103). It can regulate liver function by modulating Supplementary regulating cytokines like FAS, and the PPAR family (104). Additionally, Wnt/β -catenin pathway plays an essential role in HCC by mediating communication between the distinct components of the TME, such as immune cells, stem cells, and non-cellular constituents (105).

3.3 JAK/STAT pathway

As a key downstream signal transducer for numerous cytokines (such as IL-6) and growth factors (such as EGF), the JAK/STAT pathway exhibits dysregulation in inflammatory conditions and HCC.JAK and STAT regulate cell development, with persistent activation of STAT leading to harmful pathological effects (106, 107).

Disruption of the GH/JAK2/STAT5 signaling pathway, a result of inhibited growth hormone (GH) secretion (brought on by obesity, inflammation, and excessive glucose), results in increased lipid accumulation in the liver, further leading to MAFLD and subsequently HCC (108). STAT3 is strongly linked to liver injury, playing a significant role in the genesis of liver diseases; a common activator of STAT3 is IL-6. Activating the IL-6/JAK/STAT3 signaling cascade within the liver amplifies inflammation and immune responses, furthering the development of HCC (109).

3.4 PI3K/AKT pathway

Upon receptor binding by insulin and inflammation, the PI3K/ AKT pathway is activated, acting as an essential oncogenic mechanism controlling metabolism, cell growth, and survival. Inflammation is worsened by dysregulated PI3K/AKT signaling, which can then lead to type 2 diabetes mellitus and development of HCC. AKT maintains hepatic lipid homeostasis through regulation of lipid metabolism. The PI3K/AKT signaling process triggers the creation of genes for proteins and transcription factors that play a role in DNL, acetyl-CoA carboxylase α (ACC α) and SREBP1 for example (110, 111).

3.5 MAPK pathway

MAPK pathway comprises a family of mitogen-activated protein kinases, including stress-responsive MAPK, c-JNK, and p38 MAPK (112).

High activation of JNK is evident in HCC, which is related to the severity of liver histological activity and facilitates carcinogenesis (113). Increases of ROS, FFA, and TNF- α during chronic inflammation and obesity triggers activation of JNK in hepatocytes and macrophages, thus increasing production of inflammatory cytokines that drives inflammation, apoptosis, liver injury and fibrosis, and hepatic IR, thereby highlighting the metabolic effects of the JNK pathway (114). Macrophage overactivation of JNK is important for pro-inflammatory differentiation and tissue infiltration, while JNK1 deficiency within macrophages prevents hepatic IR, JNK directly contributes to reducing fatty acid oxidation and increasing the potential of steatosis by inhibiting hepatic PPAR α and other genes that it targets. Activation of apoptotic proteins, such as Bcl-2-L-11, BAD, and Bcl-2-L-4, result in the initiation of lipotoxicity and apoptosis through the function of JNK (115).

p38 α/β MAPK stimulates generation of hepatocytes by activating pro-apoptotic genes, such as PEPCK, G6Pase, and PGC-1 α . Activation of p38 α MAPK has been recently shown to drive ER stress and IR, accelerating the development of NASH (112); MAFLD patients who are obese display increased levels of p38 α MAPK, which leads to HCC (116).

3.6 AMPK pathway

An intracellular energy sensor is the AMPK pathway, also known as AMP-activated protein kinase, which plays an essential role in maintaining energy homeostasis while also taking part in various biological processes. Activation of AMPK increases if there is nutrient deprivation; however, it decreases if there is chronic inflammation and MAFLD (117). In order to combat liver injury and fibrosis, increasing AMPK activity is a viable therapeutic plan. The loss of AMPK activity would exacerbate liver injury and fibrosis. The prevention of HSC activation, proliferation, and migration can improve liver fibrosis by activating AMPK, in addition to reducing fibrotic stimuli and inhibiting the expression of fibrotic genes (118). Cell proliferation is regulated through AMPK's inhibition of mTOR signaling (119).

3.7 NF-κB and Toll-like pathway

Key inflammatory pathways involved in HCC are the NF- κ B and Toll-like receptor (TLR) pathways (120, 121). Chronic inflammation, which is a product of saturated fatty acids, activates pro-inflammatory pathways in adipocytes and macrophages using a mechanism dependent on TLR4 (115). After TLR signaling, transcription factors like NF- κ B and AP-1 are activated, increasing secretion of inflammatory cytokines like IL-6, IL-1 β , and TNF- α . This increase in pro-inflammatory cytokines that occur in hepatocytes will lead to insulin resistance, liver cell damage, and also the progression of MAFLD, NASH, and HCC. The gut microbiota's dependence on TLRs is an important characteristic when looking at the relationship between inflammation and obesity. Also, mice without TLR5 have a distinct gut microbiota profile, showing susceptibility to metabolic syndrome (122).

NF-κB is a transcription factor that is crucial in the processes of inflammation, immunity, cell proliferation, and how liver injury, fibrosis, and HCC occurs (123). IKKα/IKKβ, a complex that directly activates NF-κB, is associated with downstream gene expression of TLRs and cytokines. Broadly, NF-κB has many responsibilities in various cellular compartments; it has been seen to affect hepatocyte survival, inflammation that occurs in KCs, and also the survival, inflammation, and activation of HSCs (124). If NF-κB regulates HSC survival, it will also promote the induction and secretion of inflammatory chemokines such as CCL2 and CCL3. However, NF-κB has a protective effect in the liver. Significant inhibition of NF-κB has been shown to cause hepatocyte apoptosis (125).

3.8 p53 pathway

Integrating cellular stress responses, metabolism, and cell cycle regulation, the tumor suppressor gene p53 is a key regulatory element in both liver homeostasis and dysfunction (103, 126). Moderate and transient p53 activation inhibits both liver lipid accumulation and inflammation under normal conditions. However, during cellular stress stemming from inflammation or NASH, excessive p53 activation can trigger IR, lipid accumulation, inflammation, and oxidative stress through various mechanisms, thereby increasing the risk of HCC (127, 128).

Elevated p53 levels can worsen the release of pro-inflammatory cytokines, which in turn contributes to metabolic abnormalities

facilitating HCC initiation and progression (129, 130). For instance, systemic IR is triggered by p53 activation during hyperlipidemia or excessive caloric intake. The formation of white and brown adipose tissue is critically regulated by p53, functioning as a suppressor of adipocyte pre-differentiation (131). Within AT, NF- κ B signaling leads to the expression of pro-inflammatory adipokines following p53 activation, resulting in hepatic steatosis, IR, and inflammation. Conversely, reducing p53 activity can diminish inflammation and ease hepatic steatosis (132). Further, as a major positive regulator of lipid metabolism in hepatocytes, p53 plays a role in lipotoxicity-mediated NASH progression (133). Increased hepatocyte apoptosis, driven by p53 activation, contributes to liver fibrosis, and eliminating p53 completely negates this fibrotic phenotype, indicating significant implications for HCC progression (134).

4 Current status and prospects of liver cancer treatment

Ranking at fifth most common is HCC, hepatocellular carcinoma, when looking at cancers worldwide. It is the cause of death for the third leading cause of cancer-related deaths (135). Currently, tumor resection is the most effective form of treatment for this cancer; however, in postoperative recovery, the tumor can have high recurrence (136). In addition, there are only two clinical drugs that specifically target HCC. Those drugs, sorafenib and lenvatinib, can extend overall survival by approximately 2-3 months (135). Therefore, new treatments for HCC are urgently needed, and macrophage-targeted therapy and immunotherapy for liver cancer have become research hotspots.

4.1 Liver cancer immunotherapy

Cancer immunotherapy triggers systemic and lasting antitumor responses, making it a promising option for treating HCC. Immune checkpoint inhibitors (ICIs) targeting cytotoxic Tlymphocyte antigen-4 (CTLA-4), programmed cell death protein-1 (PD-1), or its ligand programmed cell death-ligand 1 (PD-L1) have demonstrated therapeutic benefit in HCC 144. Beyond ICIs, adoptive cell therapy, chimeric antigen receptor (CAR)-modified immune cells, engineered cytokines, and therapeutic cancer vaccines represent increasingly viable immunotherapy approaches in clinical settings (136, 137). The challenges and future directions in this research field are discussed below.

Immune Checkpoint Inhibitors (ICIs)

Expressed on immune cells, a collection of molecules known as immune checkpoints finely regulate the level of immune activation. An important function of these checkpoints is to prevent autoimmune reactions, a condition characterized by the immune system attacking the body's own healthy cells (138). Tumor cells can over-activate immune checkpoints, leading to immune system dysfunction. ICIs can relieve this inhibition, reactivating immune cells to attack and destroy cancer cells. ICIs are monoclonal antibodies designed to disrupt the interaction between immune checkpoint proteins and their respective ligands. By blocking T-cell inactivation and reinvigorating immune recognition and attack, ICIs amplify the anti-tumor immune response. Common ICI targets currently include PD-1, PD-L1, and CTLA-4 (139). PD-1 is found on the surface of most immune cell types, predominantly on activated T cells, NK cells, regulatory T cells (Tregs), myeloid-derived suppressor cells (MDSCs), monocytes, and dendritic cells (DCs). This protein can bind to its ligands PD-L1 and PD-L2, which are expressed in many tumors, including HCC, transmitting inhibitory signals to T cells and inducing immune evasion by tumor cells (140).

Acting as a transmembrane receptor on T cells, CTLA-4 is expressed mainly on dendritic cells and activated T cells. It participates in the negative regulation of the immune response after the B7 molecule binds to it. The B7 molecule ligand can be bound by both CTLA-4 and CD28 (141). Compared to CD28, the affinity for the ligand is 20-100 times higher in CTLA-4. If the ligand binds to it, then it inhibits cell proliferation, stops the production of cytokines, and prevents cell cycle progression. Competitively, CTLA-4 blocks CD28 for the B7-1/B7-2 ligand; because of this, CD28 co-stimulation is unable to work.(Figure 5).

4.2 Anti-tumor cell therapy

ACT uses immune cells from the patient or healthy donors to combat cancer and has become a viable option for cancer treatment (142). Compared to targeted drugs, ACT can be activated and replicated within the body, producing a lasting anti-tumor effect (143).

Traditional immune cell therapies, such as CIK (cytokineinduced killer) cells, involve culturing a patient's immune cells outside the body and reinfusing them to target and kill tumor cells. The key components of CIK cells are NKT cells, natural killer NK cells, and cytotoxic T lymphocytes (CTLs). Leveraging adhesion molecules, CIK cells recognize tumor cells and induce lysis independent of major histocompatibility complex (MHC) restriction (144). However, traditional immune cell therapy lacks specificity, limiting its effectiveness. The key to enhancing immune cell-mediated tumor killing lies in improving the immune cells' ability to recognize tumors.

Most tumor cells express certain tumor-specific or tumorassociated antigens (TSA or TAA). By combining the "antigen recognition domain (scFv)" of antibodies that recognize these tumor antigens with components that promote T-cell proliferation, and transducing them into the patient's T cells using gene transfer, the T cells are made to express a CAR. Once the patient's T cells are "reprogrammed," they can produce large quantities of tumor-specific CAR-T cells upon contact with target cells, achieving specific killing of tumor cells. This is known as CAR-T cell therapy. By introducing a synthetically designed CAR molecule into T cells, CAR-T cells gain a new targeted activation



function, and once reinfused into the patient, these CAR-T cells are no longer MHC-restricted. Instead, they are activated by binding to target antigens, efficiently killing tumor cells. This process is illustrated in Figure 6.

While still under development (145), due to the complexity of solid tumors and their heterogeneity, finding specific targets for CAR-T therapy for liver cancer continues to be the focus of research. One research avenue looks towards using GPC3 (glypican-3) as a target in order to kill HCC (146). Using Tet-On inducible CD147-CAR-T cells has also shown promise in that these cells have successfully destroyed several HCC cell lines and inhibited the growth of cancer in xenograft models (147). However, all of these targets can be also found in other parts of the body leading to toxicity. The research moving forward lies in finding more specific antigens, as well as improving the efficacy and safety of CAR-T therapy.

Building on CAR-T therapy, research has also extended to CAR-NK immunotherapy (148, 149), TCR-T (150, 151), and other advanced approaches that continue to be explored and developed.

4.3 Macrophage-targeted therapy in liver cancer

Depending on the signals present in their microenvironment, macrophages can adopt different polarized states. Primarily, based on their activation state and function, they are grouped as classically activated, pro-inflammatory M1 macrophages, or selectively activated, anti-inflammatory M2 macrophages (152). The polarization of macrophages towards an M1 phenotype can be triggered by IFN- γ , LPS, or GM-CSF. These M1 macrophages, by secreting inflammatory cytokines like IL-1, contribute to inflammatory responses, defend against intracellular pathogens, and exhibit anti-tumor effects (153, 154).

In contrast, M2 macrophages are induced by IL-4, IL-13, and exhibit high expression of CD206, enhanced phagocytic capacity, and secrete anti-inflammatory cytokines like IL-10 and TGF- β , facilitating Th2 cell differentiation, immune regulation, repair functions, wound healing, angiogenesis, and promoting tumor progression (155).

Among them, M2 TAMs contribute to enhancing the stem celllike properties of cancer cells in liver cancer (156, 157). They participate in the growth of tumor microvessels and lymphatic vessels by secreting VEGF (vascular endothelial growth factor) and EGF, promoting tumor cell proliferation (158). Additionally, they secrete IL-1, CSF-1, MMPs, etc., which facilitate tumor cell metastasis and invasion (159, 160). M2 TAMs are also involved in tumor immune evasion regulation by producing IL-10, PGE2, TGF- β , and they can promote tumor growth by regulating tumor cell metabolism (Figure 7) (161, 162).

As an important cell type that promotes tumor growth and metastasis, M2 TAMs can serve as crucial therapeutic targets. Drug development targeting tumor-associated macrophages can be approached in three ways: inhibiting the production of tumorassociated macrophages and promoting their exhaustion (163, 164); suppressing the recruitment of TAMs; and reprogramming TAMs to shift from a tumor-suppressive immune state to a tumorpromoting immune state (transforming M2 to M1) (165, 166).

Besides CAR-T cell therapy, clinical trials are exploring CAR-M therapy. Genetically engineered macrophages can target CD19 and CD22 antigens to find tumor cells (167). These CAR-M cells then eat the tumor cells, release chemicals to change the tumor's environment, present tumor antigens to T cells, and boost immune responses (168). Studies in solid tumors show macrophages effectively destroy tumor cells via SYK (spleen tyrosine kinase) (169). A viral vector, Ad5/f35,



can make macrophages stay in an M1 state in mice, improving T cell activity and stopping solid tumor growth. However, finding specific targets on liver cancer cells and engineering macrophages within the liver environment is hard due to the diversity of liver cancers.

Attention must also be paid to the potential off-target toxicity and immunogenicity of these treatments (170–172). As CAR-M technology continues to evolve, developing safer, more reliable, and more effective CAR-M approaches is essential for translating it into clinical practice. Whether combining CAR-M with CAR-T, multitarget kinase inhibitors, and ICIs can synergistically enhance tumor suppression remains an area for future research.

4.4 Traditional Chinese medicine in liver cancer treatment

In Traditional Chinese Medicine (TCM), liver cancer falls under concepts such as "jaundice" and "accumulation." In the early stages, liver cancer is often characterized by excess conditions, gradually progressing to a combination of excess and deficiency, and ultimately becoming deficiency with excess signs in advanced stages, closely related to pathological factors such as blood stasis and damp-heat toxins (173). The research value of TCM in treating liver cancer is of significant importance in modern pharmacology. For instance, ginsenoside Rb1 can induce apoptosis and inhibit tumor progression by mediating mitochondrial autophagy (174). Ginseng polysaccharides can induce apoptosis in liver cancer cells via the ERK pathway, potentially blocking tumor cells at the G0/G1 phase and inhibiting their proliferation (175). Additionally, interventions with different concentrations of Astragalus polysaccharides on SMMC-7721 cells may reduce tumor cell migration and invasion by inhibiting the activation of the JAK/STAT signaling pathway (176). Studies have also shown that polysaccharides from Half-leaf Mimosa can inhibit tumor growth in H22 tumor-bearing mice potentially downregulating the expression of VEGFA and further suppressing the VEGF signaling pathway to inhibit tumor angiogenesis (177),. Many other traditional Chinese medicines have significant research value in liver cancer treatment, particularly the combination of TCM formulas, which may yield synergistic effects in therapy.

With the increasing incidence of HCC, the hepatoprotective effects of TCM are becoming increasingly important. Compared to conventional drugs, TCM offers advantages such as wide availability, lower cost, greater stability, and fewer side effects. Furthermore, numerous studies have demonstrated the liver-protective effects of TCM extracts through anti-lipid peroxidation mechanisms. Both hesperidin and Bicyclol have shown promise in addressing hepatic steatosis. Hesperidin, in *in vitro* and *in vivo* settings, has been shown to alleviate steatosis by upregulating antioxidant levels through PI3K/AKT-Nrf2 and inhibiting NF-KB-mediated inflammation (178). Bicyclol, found as an extract from *Schisandra chinensis*, possesses a wide variety of pharmacological activities. Notably, Bicyclol lessens tetracycline-induced steatosis while also ameliorating hepatic lipid accumulation and physalin-induced steatosis (179).



Therapies derived from TCM present certain obstacles for treatment of HCC, yet the prevention and early management benefits are undeniable. Clinical trials have explored several drugs and treatment strategies but none have resulted in significant improvement. Sadly, this is also true of later-approved drugs due to mechanisms of drug resistance. One of the most promising potential tumor growth inhibitors, Ferroptosis, can impact HCC development and progression by manipulating intracellular iron levels and ROS (180). Ferroptosis, however, is understudied in human clinical trials, and is mainly investigated on animal models. The evidence for its mechanisms at the molecular level are relatively limited.

Whether it is possible to clearly distinguish ferroptosis from other forms of PCD during pathogenesis, and to carry out targeted prevention and treatment, is also worthy of further investigation.

5 Conclusion and prospects

The liver functions as an immunoregulatory organ, containing a rich array of adaptive immune cells that can suppress inflammation

to a certain extent (181, 182). The interactions within the TIME are intricate, dependent on the populations of immune cells present, and predictive of how well immunotherapies will function and how long patients will live. HCC is known to harbor TAMs, MDSCs, CAFs, TANs, TILs, DCs, and elements of the ECM within its TIME (183). Compared to other solid tumors, HCC faces a steep climb in effectively utilizing immunotherapy due to its immunosuppressive TIME. In HCC, nearly all cell subpopulations and an army of regulatory processes conspire to advance the tumor's malignancy.

Macrophage research and advances in immunotherapy have provided some inroads into managing liver cancer, yet the road to fully tackling this disease remains beset with many challenges. Objective response rates can still be too low and adverse treatment effects occur with discouraging frequency. To surmount these obstacles and realize personalized precision treatment plans for each liver cancer patient, there is a clear need to take a bird's eye view analysis, evaluate, and predict treatment outcomes, and explore new combination therapy techniques. Targeting liver-specific immune environment macrophages with more stable, safe, and effective immunotherapeutic methods, alongside traditional Chinese medicine, will further advance the treatment of liver cancer.

Author contributions

ZD: Writing – original draft, Writing – review & editing. LW: Writing – review & editing. JS: Writing – review & editing. LZ: Writing – review & editing. YT: Writing – review & editing. HT: Conceptualization, Writing – review & editing, Writing – original draft.

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References

1. Devarbhavi H, Asrani SK, Arab JP, Nartey YA, Pose E, Kamath PS. Global burden of liver disease: 2023 update. *J Hepatol.* (2023) 79:516–37. doi: 10.1016/j.jhep.2023.03.017

2. Griffin C, Agbim U, Ramani A, Shankar N, Kanwal F, Asrani SK. Underestimation of cirrhosis-related mortality in the medicare eligible population, 1999-2018. *Clin Gastroenterol Hepatol.* (2023) 21:223-25.e3. doi: 10.1016/j.cgh.2021.10.036

3. Asrani SK, Devarbhavi H, Eaton J, Kamath PS. Burden of liver diseases in the world. J Hepatol. (2019) 70:151-71. doi: 10.1016/j.jhep.2018.09.014

4. Baumgartner K, Cooper J, Smith A, St Louis J. Liver disease: cirrhosis. FP Essent. (2021) 511:36-43.

5. Garcia-Pagan JC, Francoz C, Montagnese S, Senzolo M, Mookerjee RP. Management of the major complications of cirrhosis: Beyond guidelines. *J Hepatol.* (2021) 75 Suppl 1:S135-s146. doi: 10.1016/j.jhep.2021.01.027

6. Tincopa MA, Loomba R. Non-invasive diagnosis and monitoring of nonalcoholic fatty liver disease and non-alcoholic steatohepatitis. *Lancet Gastroenterol Hepatol.* (2023) 8:660-70. doi: 10.1016/s2468-1253(23)00066-3

7. Huang DQ, Terrault NA, Tacke F, Gluud LL, Arrese M, Bugianesi E, et al. Global epidemiology of cirrhosis - aetiology, trends and predictions. *Nat Rev Gastroenterol Hepatol.* (2023) 20:388–98. doi: 10.1038/s41575-023-00759-2

8. Huang DQ, Mathurin P, Cortez-Pinto H, Loomba R. Global epidemiology of alcohol-associated cirrhosis and HCC: trends, projections and risk factors. *Nat Rev Gastroenterol Hepatol.* (2023) 20:37–49. doi: 10.1038/s41575-022-00688-6

9. Singal AG, Kanwal F, Llovet JM. Global trends in hepatocellular carcinoma epidemiology: implications for screening, prevention and therapy. *Nat Rev Clin Oncol.* (2023) 20:864–84. doi: 10.1038/s41571-023-00825-3

10. Pan M, Luo M, Liu L, Chen Y, Cheng Z, Wang K, et al. EGR1 suppresses HCC growth and aerobic glycolysis by transcriptionally downregulating PFKL. J Exp Clin Cancer Res. (2024) 43:35. doi: 10.1186/s13046-024-02957-5

11. Mishra L, Banker T, Murray J, Byers S, Thenappan A, He AR, et al. Liver stem cells and hepatocellular carcinoma. *Hepatology*. (2009) 49:318–29. doi: 10.1002/ hep.22704

12. Nio K, Yamashita T, Kaneko S. The evolving concept of liver cancer stem cells. *Mol Cancer.* (2017) 16:4. doi: 10.1186/s12943-016-0572-9

13. Waly Raphael S, Yangde Z, Yuxiang C. Hepatocellular carcinoma: focus on different aspects of management. *ISRN Oncol.* (2012) 2012:421673. doi: 10.5402/2012/421673

14. Wang Z, Chen K, Jia Y, Chuang JC, Sun X, Lin YH, et al. Dual ARID1A/ARID1B loss leads to rapid carcinogenesis and disruptive redistribution of BAF complexes. *Nat Cancer*. (2020) 1:909–22. doi: 10.1038/s43018-020-00109-0

Conflict of interest

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15. Mackowiak B, Fu Y, Maccioni L, Gao B. Alcohol-associated liver disease. J Clin Invest. (2024) 134. doi: 10.1172/jci176345

16. Wu X, Fan X, Miyata T, Kim A, Cajigas-Du Ross CK, Ray S, et al. Recent advances in understanding of pathogenesis of alcohol-associated liver disease. *Annu Rev Pathol.* (2023) 18:411–38. doi: 10.1146/annurev-pathmechdis-031521-030435

17. Hagström H, Thiele M, Roelstraete B, Söderling J, Ludvigsson JF. Mortality in biopsy-proven alcohol-related liver disease: a population-based nationwide cohort study of 3453 patients. *Gut.* (2021) 70:170–9. doi: 10.1136/gutjnl-2019-320446

18. Stein E, Cruz-Lemini M, Altamirano J, Ndugga N, Couper D, Abraldes JG, et al. Heavy daily alcohol intake at the population level predicts the weight of alcohol in cirrhosis burden worldwide. *J Hepatol.* (2016) 65:998–1005. doi: 10.1016/j.jhep.2016.06.018

19. Raynard B, Balian A, Fallik D, Capron F, Bedossa P, Chaput JC, et al. Risk factors of fibrosis in alcohol-induced liver disease. *Hepatology*. (2002) 35:635–8. doi: 10.1053/jhep.2002.31782

20. Hart CL, Morrison DS, Batty GD, Mitchell RJ, Davey Smith G. Effect of body mass index and alcohol consumption on liver disease: analysis of data from two prospective cohort studies. *BMJ*. (2010) 340:c1240. doi: 10.1136/bmj.c1240

21. Ascha MS, Hanouneh IA, Lopez R, Tamimi TA, Feldstein AF, Zein NN. The incidence and risk factors of hepatocellular carcinoma in patients with nonalcoholic steatohepatitis. *Hepatology*. (2010) 51:1972–8. doi: 10.1002/hep.23527

22. Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology*. (2016) 64:73–84. doi: 10.1002/hep.28431

23. Ludwig J, Viggiano TR, Mcgill DB, Oh BJ. Nonalcoholic steatohepatitis: Mayo Clinic experiences with a hitherto unnamed disease. *Mayo Clin Proc.* (1980) 55:434–8. doi: 10.1016/S0025-6196(24)00530-5

24. Basaranoglu M, Basaranoglu G, Sentürk H. From fatty liver to fibrosis: a tale of "second hit. World J Gastroenterol. (2013) 19:1158–65. doi: 10.3748/wjg.v19.i8.1158

25. Buzzetti E, Pinzani M, Tsochatzis EA. The multiple-hit pathogenesis of nonalcoholic fatty liver disease (NAFLD). *Metabolism*. (2016) 65:1038–48. doi: 10.1016/ j.metabol.2015.12.012

26. Byrne CD, Targher G. Ectopic fat, insulin resistance, and nonalcoholic fatty liver disease: implications for cardiovascular disease. *Arterioscler Thromb Vasc Biol.* (2014) 34:1155–61. doi: 10.1161/atvbaha.114.303034

27. Byrne CD. Ectopic fat, insulin resistance and non-alcoholic fatty liver disease. Proc Nutr Soc. (2013) 72:412-9. doi: 10.1017/s0029665113001249

28. Takamura T, Misu H, Ota T, Kaneko S. Fatty liver as a consequence and cause of insulin resistance: lessons from type 2 diabetic liver. *Endocr J.* (2012) 59:745–63. doi: 10.1507/endocrj.ej12-0228

29. Heeren J, Scheja L. Metabolic-associated fatty liver disease and lipoprotein metabolism. *Mol Metab.* (2021) 50:101238. doi: 10.1016/j.molmet.2021.101238

30. MaChado MV, Cortez-Pinto H. Non-alcoholic fatty liver disease: what the clinician needs to know. *World J Gastroenterol.* (2014) 20:12956–80. doi: 10.3748/ wjg.v20.i36.12956

31. Tanase DM, Gosav EM, Costea CF, Ciocoiu M, Lacatusu CM, Maranduca MA, et al. The intricate relationship between type 2 diabetes mellitus (T2DM), insulin resistance (IR), and nonalcoholic fatty liver disease (NAFLD). *J Diabetes Res.* (2020) 2020:3920196. doi: 10.1155/2020/3920196

32. Haemmerle G, Lass A, Zimmermann R, Gorkiewicz G, Meyer C, Rozman J, et al. Defective lipolysis and altered energy metabolism in mice lacking adipose triglyceride lipase. *Science*. (2006) 312:734–7. doi: 10.1126/science.1123965

33. Samuel VT, Shulman GI. Mechanisms for insulin resistance: common threads and missing links. *Cell.* (2012) 148:852–71. doi: 10.1016/j.cell.2012.02.017

34. Luukkonen PK, Qadri S, Ahlholm N, Porthan K, Männistö V, Sammalkorpi H, et al. Distinct contributions of metabolic dysfunction and genetic risk factors in the pathogenesis of non-alcoholic fatty liver disease. *J Hepatol.* (2022) 76:526–35. doi: 10.1016/j.jhep.2021.10.013

35. Ferré P, Foufelle F. Hepatic steatosis: a role for de novo lipogenesis and the transcription factor SREBP-1c. *Diabetes Obes Metab.* (2010) 12 Suppl 2:83–92. doi: 10.1111/j.1463-1326.2010.01275.x

36. Song Z, Xiaoli AM, Yang F. Regulation and metabolic significance of *de novo* lipogenesis in adipose tissues. *Nutrients.* (2018) 10:1383–403. doi: 10.3390/nu10101383

37. Dong Q, Majumdar G, O'meally RN, Cole RN, Elam MB, Raghow R. Insulininduced de novo lipid synthesis occurs mainly via mTOR-dependent regulation of proteostasis of SREBP-1c. *Mol Cell Biochem.* (2020) 463:13–31. doi: 10.1007/s11010-019-03625-5

38. Liu L, Yang M, Lin X, Li Y, Liu C, Yang Y, et al. Modulation of hepatic sterol regulatory element-binding protein-1c-mediated gene expression contributes to Salacia oblonga root-elicited improvement of fructose-induced fatty liver in rats. *J Ethnopharmacol.* (2013) 150:1045–52. doi: 10.1016/j.jep.2013.10.020

39. Lawitz EJ, Coste A, Poordad F, Alkhouri N, Loo N, Mccolgan BJ, et al. AcetylcoA carboxylase inhibitor GS-0976 for 12 weeks reduces hepatic *de novo* lipogenesis and steatosis in patients with nonalcoholic steatohepatitis. *Clin Gastroenterol Hepatol.* (2018) 16:1983–91.e3. doi: 10.1016/j.cgh.2018.04.042

40. Sakurai Y, Kubota N, Yamauchi T, Kadowaki T. Role of insulin resistance in MAFLD. Int J Mol Sci. (2021) 22:4156–82. doi: 10.3390/ijms22084156

41. Fan JG, Kim SU, Wong VW. New trends on obesity and NAFLD in Asia. J Hepatol. (2017) 67:862–73. doi: 10.1016/j.jhep.2017.06.003

42. Basaranoglu M, Basaranoglu G, Sabuncu T, Sentürk H. Fructose as a key player in the development of fatty liver disease. *World J Gastroenterol.* (2013) 19:1166–72. doi: 10.3748/wjg.v19.i8.1166

43. Jensen T, Abdelmalek MF, Sullivan S, Nadeau KJ, Green M, Roncal C, et al. Fructose and sugar: A major mediator of non-alcoholic fatty liver disease. *J Hepatol.* (2018) 68:1063–75. doi: 10.1016/j.jhep.2018.01.019

44. Tappy L, Lê KA. Does fructose consumption contribute to non-alcoholic fatty liver disease? *Clin Res Hepatol Gastroenterol.* (2012) 36:554–60. doi: 10.1016/ j.clinre.2012.06.005

45. Ameer F, Scandiuzzi L, Hasnain S, Kalbacher H, Zaidi N. De novo lipogenesis in health and disease. *Metabolism.* (2014) 63:895–902. doi: 10.1016/j.metabol.2014.04.003

46. Farzanegi P, Dana A, Ebrahimpoor Z, Asadi M, Azarbayjani MA. Mechanisms of beneficial effects of exercise training on non-alcoholic fatty liver disease (NAFLD): Roles of oxidative stress and inflammation. *Eur J Sport Sci.* (2019) 19:994–1003. doi: 10.1080/17461391.2019.1571114

47. Pyper SR, Viswakarma N, Yu S, Reddy JK. PPARalpha: energy combustion, hypolipidemia, inflammation and cancer. *Nucl Recept Signal.* (2010) 8:e002. doi: 10.1621/nrs.08002

48. Chyau CC, Wang HF, Zhang WJ, Chen CC, Huang SH, Chang CC, et al. Antrodan alleviates high-fat and high-fructose diet-induced fatty liver disease in C57BL/6 mice model via AMPK/Sirt1/SREBP-1c/PPARγ Pathway. *Int J Mol Sci.* (2020) 21:360–78. doi: 10.3390/ijms21010360

49. Kim B, Woo MJ, Park CS, Lee SH, Kim JS, Kim B, et al. Hovenia dulcis extract reduces lipid accumulation in oleic acid-induced steatosis of hep G2 cells via activation of AMPK and PPAR α /CPT-1 pathway and in acute hyperlipidemia mouse model. *Phytother Res.* (2017) 31:132–9. doi: 10.1002/ptr.5741

50. Gao Q, Jia Y, Yang G, Zhang X, Boddu PC, Petersen B, et al. PPAR α -deficient ob/ob obese mice become more obese and manifest severe hepatic steatosis due to decreased fatty acid oxidation. *Am J Pathol.* (2015) 185:1396–408. doi: 10.1016/j.ajpath.2015.01.018

51. Pessayre D, Mansouri A, Fromenty B. Nonalcoholic steatosis and steatohepatitis. V. Mitochondrial dysfunction in steatohepatitis. *Am J Physiol Gastrointest Liver Physiol.* (2002) 282:G193–9. doi: 10.1152/ajpgi.00426.2001

52. Pessayre D, Berson A, Fromenty B, Mansouri A. Mitochondria in steatohepatitis. Semin Liver Dis. (2001) 21:57–69. doi: 10.1055/s-2001-12929

53. Shen GX. Oxidative stress and diabetic cardiovascular disorders: roles of mitochondria and NADPH oxidase. *Can J Physiol Pharmacol.* (2010) 88:241-8. doi: 10.1139/y10-018

54. Serviddio G, Sastre J, Bellanti F, Viña J, Vendemiale G, Altomare E. Mitochondrial involvement in non-alcoholic steatohepatitis. *Mol Aspects Med.* (2008) 29:22–35. doi: 10.1016/j.mam.2007.09.014

55. Li W, Cao T, Luo C, Cai J, Zhou X, Xiao X, et al. Crosstalk between ER stress, NLRP3 inflammasome, and inflammation. *Appl Microbiol Biotechnol.* (2020) 104:6129–40. doi: 10.1007/s00253-020-10614-y

56. Gong J, Wang XZ, Wang T, Chen JJ, Xie XY, Hu H, et al. Molecular signal networks and regulating mechanisms of the unfolded protein response. *J Zhejiang Univ Sci B*. (2017) 18:1–14. doi: 10.1631/jzus.B1600043

57. Gentile CL, Frye M, Pagliassotti MJ. Endoplasmic reticulum stress and the unfolded protein response in nonalcoholic fatty liver disease. *Antioxid Redox Signal.* (2011) 15:505–21. doi: 10.1089/ars.2010.3790

58. Nasiri-Ansari N, Nikolopoulou C, Papoutsi K, Kyrou I, Mantzoros CS, Kyriakopoulos G, et al. Empagliflozin attenuates non-alcoholic fatty liver disease (NAFLD) in high fat diet fed ApoE((-/-)) mice by activating autophagy and reducing ER stress and apoptosis. *Int J Mol Sci.* (2021) 22:818–39. doi: 10.3390/ijms22020818

59. Urano F, Wang X, Bertolotti A, Zhang Y, Chung P, Harding HP, et al. Coupling of stress in the ER to activation of JNK protein kinases by transmembrane protein kinase IRE1. *Science*. (2000) 287:664–6. doi: 10.1126/science.287.5453.664

60. Das M, Sabio G, Jiang F, Rincón M, Flavell RA, Davis RJ. Induction of hepatitis by JNK-mediated expression of TNF-alpha. *Cell.* (2009) 136:249–60. doi: 10.1016/j.cell.2008.11.017

61. Ozcan U, Cao Q, Yilmaz E, Lee AH, Iwakoshi NN, Ozdelen E, et al. Endoplasmic reticulum stress links obesity, insulin action, and type 2 diabetes. *Science*. (2004) 306:457–61. doi: 10.1126/science.1103160

62. Marra F, Svegliati-Baroni G. Lipotoxicity and the gut-liver axis in NASH pathogenesis. J Hepatol. (2018) 68:280–95. doi: 10.1016/j.jhep.2017.11.014

63. Musso G, Cassader M, Paschetta E, Gambino R. Bioactive lipid species and metabolic pathways in progression and resolution of nonalcoholic steatohepatitis. *Gastroenterology.* (2018) 155:282–302.e8. doi: 10.1053/j.gastro.2018.06.031

64. Hendrikx T, Walenbergh SM, Hofker MH, Shiri-Sverdlov R. Lysosomal cholesterol accumulation: driver on the road to inflammation during atherosclerosis and non-alcoholic steatohepatitis. *Obes Rev.* (2014) 15:424–33. doi: 10.1111/obr.12159

65. Walker CM, Feng Z, Lemon SM. Reassessing immune control of hepatitis A virus. *Curr Opin Virol.* (2015) 11:7–13. doi: 10.1016/j.coviro.2015.01.003

66. Schulte I, Hitziger T, Giugliano S, Timm J, Gold H, Heinemann FM, et al. Characterization of CD8+ T-cell response in acute and resolved hepatitis A virus infection. *J Hepatol.* (2011) 54:201–8. doi: 10.1016/j.jhep.2010.07.010

67. Zhou Y, Callendret B, Xu D, Brasky KM, Feng Z, Hensley LL, et al. Dominance of the CD4(+) T helper cell response during acute resolving hepatitis A virus infection. *J Exp Med.* (2012) 209:1481–92. doi: 10.1084/jem.20111906

68. Friedman SL. Liver fibrosis – from bench to bedside. J Hepatol. (2003) 38 Suppl 1:S38–53. doi: 10.1016/s0168-8278(02)00429-4

69. Bataller R, Brenner DA. Liver fibrosis. J Clin Invest. (2005) 115:209-18. doi: 10.1172/jci24282

70. Geerts A. History, heterogeneity, developmental biology, and functions of quiescent hepatic stellate cells. *Semin Liver Dis.* (2001) 21:311–35. doi: 10.1055/s-2001-17550

71. Senoo H, Kojima N, Sato M. Vitamin A-storing cells (stellate cells). *Vitam Horm.* (2007) 75:131–59. doi: 10.1016/s0083-6729(06)75006-3

72. She H, Xiong S, Hazra S, Tsukamoto H. Adipogenic transcriptional regulation of hepatic stellate cells. J Biol Chem. (2005) 280:4959–67. doi: 10.1074/jbc.M410078200

73. Fallowfield JA, Mizuno M, Kendall TJ, Constandinou CM, Benyon RC, Duffield JS, et al. Scar-associated macrophages are a major source of hepatic matrix metalloproteinase-13 and facilitate the resolution of murine hepatic fibrosis. *J Immunol.* (2007) 178:5288–95. doi: 10.4049/jimmunol.178.8.5288

74. Koyama Y, Wang P, Liang S, Iwaisako K, Liu X, Xu J, et al. Mesothelin/mucin 16 signaling in activated portal fibroblasts regulates cholestatic liver fibrosis. *J Clin Invest.* (2017) 127:1254–70. doi: 10.1172/jci88845

75. Xu F, Liu C, Zhou D, Zhang L. TGF- β /SMAD pathway and its regulation in hepatic fibrosis. J Histochem Cytochem. (2016) 64:157–67. doi: 10.1369/0022155415627681

76. Liu Y, Meyer C, Müller A, Herweck F, Li Q, Müllenbach R, et al. IL-13 induces connective tissue growth factor in rat hepatic stellate cells via TGF- β -independent Smad signaling. *J Immunol.* (2011) 187:2814–23. doi: 10.4049/jimmunol.1003260

77. Mridha AR, Wree A, Robertson A, Yeh MM, Johnson CD, Van Rooyen DM, et al. NLRP3 inflammasome blockade reduces liver inflammation and fibrosis in experimental NASH in mice. *J Hepatol.* (2017) 66:1037–46. doi: 10.1016/j.jhep.2017.01.022

78. Meier A, Chien J, Hobohm L, Patras KA, Nizet V, Corriden R. Inhibition of human neutrophil extracellular trap (NET) production by propofol and lipid emulsion. *Front Pharmacol.* (2019) 10:323. doi: 10.3389/fphar.2019.00323

79. Moles A, Murphy L, Wilson CL, Chakraborty JB, Fox C, Park EJ, et al. A TLR2/ S100A9/CXCL-2 signaling network is necessary for neutrophil recruitment in acute and chronic liver injury in the mouse. *J Hepatol.* (2014) 60:782–91. doi: 10.1016/ j.jhep.2013.12.005 80. Gehrke N, Nagel M, Straub BK, Wörns MA, Schuchmann M, Galle PR, et al. Loss of cellular FLICE-inhibitory protein promotes acute cholestatic liver injury and inflammation from bile duct ligation. *Am J Physiol Gastrointest Liver Physiol.* (2018) 314:G319–g333. doi: 10.1152/ajpgi.00097.2017

81. Karlmark KR, Weiskirchen R, Zimmermann HW, Gassler N, Ginhoux F, Weber C, et al. Hepatic recruitment of the inflammatory Gr1+ monocyte subset upon liver injury promotes hepatic fibrosis. *Hepatology*. (2009) 50:261–74. doi: 10.1002/hep.22950

82. Miura K, Kodama Y, Inokuchi S, Schnabl B, Aoyama T, Ohnishi H, et al. Tolllike receptor 9 promotes steatohepatitis by induction of interleukin-1beta in mice. *Gastroenterology.* (2010) 139:323–34.e7. doi: 10.1053/j.gastro.2010.03.052

83. Sudo K, Yamada Y, Moriwaki H, Saito K, Seishima M. Lack of tumor necrosis factor receptor type 1 inhibits liver fibrosis induced by carbon tetrachloride in mice. *Cytokine*. (2005) 29:236–44. doi: 10.1016/j.cyto.2004.11.001

84. Gao B, Bataller R. Alcoholic liver disease: pathogenesis and new therapeutic targets. *Gastroenterology*. (2011) 141:1572-85. doi: 10.1053/j.gastro.2011.09.002

85. Teschke R. Alcoholic liver disease: alcohol metabolism, cascade of molecular mechanisms, cellular targets, and clinical aspects. *Biomedicines*. (2018) 6:106–63. doi: 10.3390/biomedicines6040106

86. O'shea RS, Dasarathy S, Mccullough AJ. Alcoholic liver disease. *Hepatology*. (2010) 51:307-28. doi: 10.1002/hep.23258

87. Lucey MR, Mathurin P, Morgan TR. Alcoholic hepatitis. N Engl J Med. (2009) 360:2758-69. doi: 10.1056/NEJMra0805786

88. Friedman SL. Mechanisms of hepatic fibrogenesis. *Gastroenterology*. (2008) 134:1655-69. doi: 10.1053/j.gastro.2008.03.003

89. Ogunwobi OO, Harricharran T, Huaman J, Galuza A, Odumuwagun O, Tan Y, et al. Mechanisms of hepatocellular carcinoma progression. *World J Gastroenterol.* (2019) 25:2279–93. doi: 10.3748/wjg.v25.i19.2279

90. Villanueva A, Luedde T. The transition from inflammation to cancer in the liver. *Clin Liver Dis (Hoboken).* (2016) 8:89–93. doi: 10.1002/cld.578

91. Streba LA, Vere CC, Rogoveanu I, Streba CT. Nonalcoholic fatty liver disease, metabolic risk factors, and hepatocellular carcinoma: an open question. *World J Gastroenterol.* (2015) 21:4103–10. doi: 10.3748/wjg.v21.i14.4103

92. Scalera A, Tarantino G. Could metabolic syndrome lead to hepatocarcinoma via non-alcoholic fatty liver disease? *World J Gastroenterol.* (2014) 20:9217–28. doi: 10.3748/wjg.v20.i28.9217

93. Plaz Torres MC, Jaffe A, Perry R, Marabotto E, Strazzabosco M, Giannini EG. Diabetes medications and risk of HCC. *Hepatology*. (2022) 76:1880–97. doi: 10.1002/ hep.32439

94. Kaaks R, Lukanova A. Energy balance and cancer: the role of insulin and insulinlike growth factor-I. *Proc Nutr Soc.* (2001) 60:91–106. doi: 10.1079/pns200070

95. Wu J, Zhu AX. Targeting insulin-like growth factor axis in hepatocellular carcinoma. J Hematol Oncol. (2011) 4:30. doi: 10.1186/1756-8722-4-30

96. Breuhahn K, Longerich T, Schirmacher P. Dysregulation of growth factor signaling in human hepatocellular carcinoma. *Oncogene*. (2006) 25:3787-800. doi: 10.1038/sj.onc.1209556

97. Tanaka S, Mohr L, Schmidt EV, Sugimachi K, Wands JR. Biological effects of human insulin receptor substrate-1 overexpression in hepatocytes. *Hepatology*. (1997) 26:598–604. doi: 10.1002/hep.510260310

98. Siddique A, Kowdley KV. Insulin resistance and other metabolic risk factors in the pathogenesis of hepatocellular carcinoma. *Clin Liver Dis.* (2011) 15:281–96, vii-x. doi: 10.1016/j.cld.2011.03.007

99. Shree Harini K, Ezhilarasan D. Wnt/beta-catenin signaling and its modulators in nonalcoholic fatty liver diseases. *Hepatobiliary Pancreat Dis Int.* (2023) 22:333–45. doi: 10.1016/j.hbpd.2022.10.003

100. Russell JO, Monga SP. Wnt/ β -catenin signaling in liver development, homeostasis, and pathobiology. Annu Rev Pathol. (2018) 13:351–78. doi: 10.1146/ annurev-pathol-020117-044010

101. Perugorria MJ, Olaizola P, Labiano I, Esparza-Baquer A, Marzioni M, Marin JJG, et al. Wnt-β-catenin signalling in liver development, health and disease. *Nat Rev Gastroenterol Hepatol.* (2019) 16:121–36. doi: 10.1038/s41575-018-0075-9

102. Myung SJ, Yoon JH, Gwak GY, Kim W, Lee JH, Kim KM, et al. Wnt signaling enhances the activation and survival of human hepatic stellate cells. *FEBS Lett.* (2007) 581:2954–8. doi: 10.1016/j.febslet.2007.05.050

103. Zhang C, Yang M. Current options and future directions for NAFLD and NASH treatment. Int J Mol Sci. (2021) 22:7571–90. doi: 10.3390/ijms22147571

104. Zhou H, Zhu X, Yao Y, Su Y, Xie J, Zhu M, et al. TMEM88 modulates lipid synthesis and metabolism cytokine by regulating Wnt/ β -catenin signaling pathway in non-alcoholic fatty liver disease. *Front Pharmacol.* (2021) 12:798735. doi: 10.3389/fphar.2021.798735

105. Wang K, Qiu X, Zhao Y, Wang H, Chen L. The Wnt/β-catenin signaling pathway in the tumor microenvironment of hepatocellular carcinoma. *Cancer Biol Med.* (2021) 19:305–18. doi: 10.20892/j.issn.2095-3941.2021.0306

106. Lokau J, Schoeder V, Haybaeck J, Garbers C. Jak-Stat signaling induced by interleukin-6 family cytokines in hepatocellular carcinoma. *Cancers (Basel).* (2019) 11:1704–21. doi: 10.3390/cancers11111704

107. Dodington DW, Desai HR, Woo M. JAK/STAT - emerging players in metabolism. Trends Endocrinol Metab. (2018) 29:55–65. doi: 10.1016/j.tem.2017.11.001

108. Kaltenecker D, Themanns M, Mueller KM, Spirk K, Suske T, Merkel O, et al. Hepatic growth hormone - JAK2 - STAT5 signalling: Metabolic function, nonalcoholic fatty liver disease and hepatocellular carcinoma progression. *Cytokine*. (2019) 124:154569. doi: 10.1016/j.cyto.2018.10.010

109. Yu H, Lee H, Herrmann A, Buettner R, Jove R. Revisiting STAT3 signalling in cancer: new and unexpected biological functions. *Nat Rev Cancer*. (2014) 14:736–46. doi: 10.1038/nrc3818

110. Calvisi DF, Wang C, Ho C, Ladu S, Lee SA, Mattu S, et al. Increased lipogenesis, induced by AKT-mTORC1-RPS6 signaling, promotes development of human hepatocellular carcinoma. *Gastroenterology.* (2011) 140:1071–83. doi: 10.1053/j.gastro.2010.12.006

111. Huang X, Liu G, Guo J, Su Z. The PI3K/AKT pathway in obesity and type 2 diabetes. Int J Biol Sci. (2018) 14:1483–96. doi: 10.7150/ijbs.27173

112. Alshehade S, Alshawsh MA, Murugaiyah V, Asif M, Alshehade O, Almoustafa H, et al. The role of protein kinases as key drivers of metabolic dysfunction-associated fatty liver disease progression: New insights and future directions. *Life Sci.* (2022) 305:120732. doi: 10.1016/j.lfs.2022.120732

113. Noureddin M, Rinella ME. Nonalcoholic Fatty liver disease, diabetes, obesity, and hepatocellular carcinoma. *Clin Liver Dis.* (2015) 19:361–79. doi: 10.1016/j.cld.2015.01.012

114. Lawan A, Bennett AM. Mitogen-activated protein kinase regulation in hepatic metabolism. *Trends Endocrinol Metab.* (2017) 28:868–78. doi: 10.1016/j.tem.2017.10.007

115. Hirsova P, Ibrabim SH, Gores GJ, Malhi H. Lipotoxic lethal and sublethal stress signaling in hepatocytes: relevance to NASH pathogenesis. *J Lipid Res.* (2016) 57:1758–70. doi: 10.1194/jlr.R066357

116. Zhang X, Fan L, Wu J, Xu H, Leung WY, Fu K, et al. Macrophage p38 α promotes nutritional steatohepatitis through M1 polarization. *J Hepatol.* (2019) 71:163–74. doi: 10.1016/j.jhep.2019.03.014

117. Kumar V, Xin X, Ma J, Tan C, Osna N, Mahato RI. Therapeutic targets, novel drugs, and delivery systems for diabetes associated NAFLD and liver fibrosis. *Adv Drug Delivery Rev.* (2021) 176:113888. doi: 10.1016/j.addr.2021.113888

118. Zhao P, Saltiel AR. From overnutrition to liver injury: AMP-activated protein kinase in nonalcoholic fatty liver diseases. *J Biol Chem.* (2020) 295:12279–89. doi: 10.1074/jbc.REV120.011356

119. Woods A, Williams JR, Muckett PJ, Mayer FV, Liljevald M, Bohlooly YM, et al. Liver-specific activation of AMPK prevents steatosis on a high-fructose diet. *Cell Rep.* (2017) 18:3043–51. doi: 10.1016/j.celrep.2017.03.011

120. Li W, Xiao J, Zhou X, Xu M, Hu C, Xu X, et al. STK4 regulates TLR pathways and protects against chronic inflammation-related hepatocellular carcinoma. *J Clin Invest.* (2015) 125:4239–54. doi: 10.1172/jci81203

121. Yu J, Shen J, Sun TT, Zhang X, Wong N. Obesity, insulin resistance, NASH and hepatocellular carcinoma. *Semin Cancer Biol.* (2013) 23:483–91. doi: 10.1016/j.semcancer.2013.07.00

122. Baker RG, Hayden MS, Ghosh S. NF-κB, inflammation, and metabolic disease. *Cell Metab.* (2011) 13:11-22. doi: 10.1016/j.cmet.2010.12.008

123. Wu L, Xie Y. Effect of NF-κB on the pathogenic course of non-alcoholic fatty liver disease. *Zhong Nan Da Xue Xue Bao Yi Xue Ban.* (2017) 42:463–7. doi: 10.11817/ j.issn.1672-7347.2017.04.016

124. Luedde T, Schwabe RF. NF- κB in the liver-linking injury, fibrosis and hepatocellular carcinoma. Nat Rev Gastroenterol Hepatol. (2011) 8:108–18. doi: 10.1038/nrgastro.2010.213

125. De Gregorio E, Colell A, Morales A, Marí M. Relevance of SIRT1-NF-κB axis as therapeutic target to ameliorate inflammation in liver disease. *Int J Mol Sci.* (2020) 21:3858–82. doi: 10.3390/ijms21113858

126. Haberl EM, Weiss TS, Peschel G, Weigand K, Köhler N, Pauling JK, et al. Liver lipids of patients with hepatitis B and C and associated hepatocellular carcinoma. *Int J Mol Sci.* (2021) 22:5297–315. doi: 10.3390/ijms22105297

127. Yan Z, Miao X, Zhang B, Xie J. p53 as a double-edged sword in the progression of non-alcoholic fatty liver disease. *Life Sci.* (2018) 215:64–72. doi: 10.1016/ j.lfs.2018.10.051

128. Tomita K, Teratani T, Suzuki T, Oshikawa T, Yokoyama H, Shimamura K, et al. p53/p66Shc-mediated signaling contributes to the progression of non-alcoholic steatohepatitis in humans and mice. *J Hepatol.* (2012) 57:837–43. doi: 10.1016/j.jhep.2012.05.013

129. Zwezdaryk K, Sullivan D, Saifudeen Z. The p53/adipose-tissue/cancer nexus. Front Endocrinol (Lausanne). (2018) 9:457. doi: 10.3389/fendo.2018.00457

130. Krstic J, Reinisch I, Schupp M, Schulz TJ, Prokesch A. p53 functions in adipose tissue metabolism and homeostasis. *Int J Mol Sci.* (2018) 19:2622–43. doi: 10.3390/ ijms19092622

131. Strycharz J, Drzewoski J, Szemraj J, Sliwinska A. Is p53 involved in tissuespecific insulin resistance formation? *Oxid Med Cell Longev.* (2017) 2017:9270549. doi: 10.1155/2017/9270549 132. Shimizu I, Yoshida Y, Katsuno T, Tateno K, Okada S, Moriya J, et al. p53induced adipose tissue inflammation is critically involved in the development of insulin resistance in heart failure. *Cell Metab*. (2012) 15:51–64. doi: 10.1016/j.cmet.2011.12.006

133. Krstic J, Galhuber M, Schulz TJ, Schupp M, Prokesch A. p53 as a dichotomous regulator of liver disease: the dose makes the medicine. *Int J Mol Sci.* (2018) 19:921–44. doi: 10.3390/ijms19030921

134. Kodama T, Takehara T, Hikita H, Shimizu S, Shigekawa M, Tsunematsu H, et al. Increases in p53 expression induce CTGF synthesis by mouse and human hepatocytes and result in liver fibrosis in mice. *J Clin Invest.* (2011) 121:3343–56. doi: 10.1172/jci44957

135. Casadei-Gardini A, Scartozzi M, Tada T, Yoo C, Shimose S, Masi G, et al. Lenvatinib versus sorafenib in first-line treatment of unresectable hepatocellular carcinoma: An inverse probability of treatment weighting analysis. *Liver Int.* (2021) 41:1389–97. doi: 10.1111/liv.14817

136. Lee HW, Cho KJ, Park JY. Current status and future direction of immunotherapy in hepatocellular carcinoma: what do the data suggest? *Immune Netw.* (2020) 20:e11. doi: 10.4110/in.2020.20.e11

137. Kole C, Charalampakis N, Tsakatikas S, Vailas M, Moris D, Gkotsis E, et al. Immunotherapy for hepatocellular carcinoma: A 2021 update. *Cancers (Basel)*. (2020) 12:2859–87. doi: 10.3390/cancers12102859

138. He X, Xu C. Immune checkpoint signaling and cancer immunotherapy. Cell Res. (2020) 30:660-9. doi: 10.1038/s41422-020-0343-4

139. Zongyi Y, Xiaowu L. Immunotherapy for hepatocellular carcinoma. Cancer Lett. (2020) 470:8–17. doi: 10.1016/j.canlet.2019.12.002

140. Okazaki T, Honjo T. PD-1 and PD-1 ligands: from discovery to clinical application. Int Immunol. (2007) 19:813-24. doi: 10.1093/intimm/dxm057

141. Liu Y, Zheng P. Preserving the CTLA-4 checkpoint for safer and more effective cancer immunotherapy. *Trends Pharmacol Sci.* (2020) 41:4–12. doi: 10.1016/j.tips.2019.11.003

142. Laskowski T, Rezvani K. Adoptive cell therapy: Living drugs against cancer. J Exp Med. (2020) 217. doi: 10.1084/jem.20200377

143. Rosenberg SA, Restifo NP. Adoptive cell transfer as personalized immunotherapy for human cancer. *Science*. (2015) 348:62-8. doi: 10.1126/ science.aaa4967

144. Cappuzzello E, Vigolo E, D'accardio G, Astori G, Rosato A, Sommaggio R. How can Cytokine-induced killer cells overcome CAR-T cell limits. *Front Immunol.* (2023) 14:1229540. doi: 10.3389/fimmu.2023.1229540

145. June CH, Sadelain M. Chimeric antigen receptor therapy. N Engl J Med. (2018) 379:64–73. doi: 10.1056/NEJMra1706169

146. Gao H, Li K, Tu H, Pan X, Jiang H, Shi B, et al. Development of T cells redirected to glypican-3 for the treatment of hepatocellular carcinoma. *Clin Cancer Res.* (2014) 20:6418–28. doi: 10.1158/1078-0432.Ccr-14-1170

147. Zhang RY, Wei D, Liu ZK, Yong YL, Wei W, Zhang ZY, et al. Doxycycline inducible chimeric antigen receptor T cells targeting CD147 for hepatocellular carcinoma therapy. *Front Cell Dev Biol.* (2019) 7:233. doi: 10.3389/fcell.2019.00233

148. Sun H, Sun C, Tian Z, Xiao W. NK cells in immunotolerant organs. Cell Mol Immunol. (2013) 10:202–12. doi: 10.1038/cmi.2013.9

149. Tseng HC, Xiong W, Badeti S, Yang Y, Ma M, Liu T, et al. Efficacy of anti-CD147 chimeric antigen receptors targeting hepatocellular carcinoma. *Nat Commun.* (2020) 11:4810. doi: 10.1038/s41467-020-18444-2

150. Watanabe K, Nishikawa H. Engineering strategies for broad application of TCR-T- and CAR-T-cell therapies. *Int Immunol.* (2021) 33:551–62. doi: 10.1093/ intimm/dxab052

151. Spear TT, Callender GG, Roszkowski JJ, Moxley KM, Simms PE, Foley KC, et al. TCR gene-modified T cells can efficiently treat established hepatitis C-associated hepatocellular carcinoma tumors. *Cancer Immunol Immunother*. (2016) 65:293–304. doi: 10.1007/s00262-016-1800-2

152. Li Z, Wu T, Zheng B, Chen L. Individualized precision treatment: Targeting TAM in HCC. *Cancer Lett.* (2019) 458:86–91. doi: 10.1016/j.canlet.2019.05.019

153. Ivashkiv LB. IFNγ: signalling, epigenetics and roles in immunity, metabolism, disease and cancer immunotherapy. *Nat Rev Immunol.* (2018) 18:545–58. doi: 10.1038/ s41577-018-0029-z

154. Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdt S, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity.* (2014) 41:14–20. doi: 10.1016/j.immuni.2014.06.008

155. Mantovani A, Marchesi F, Malesci A, Laghi L, Allavena P. Tumour-associated macrophages as treatment targets in oncology. *Nat Rev Clin Oncol.* (2017) 14:399–416. doi: 10.1038/nrclinonc.2016.217

156. Wei R, Zhu WW, Yu GY, Wang X, Gao C, Zhou X, et al. S100 calcium-binding protein A9 from tumor-associated macrophage enhances cancer stem cell-like properties of hepatocellular carcinoma. *Int J Cancer.* (2021) 148:1233-44. doi: 10.1002/ijc.33371

157. Ye Y, Guo J, Xiao P, Ning J, Zhang R, Liu P, et al. Macrophages-induced long noncoding RNA H19 up-regulation triggers and activates the miR-193b/MAPK1 axis and promotes cell aggressiveness in hepatocellular carcinoma. *Cancer Lett.* (2020) 469:310–22. doi: 10.1016/j.canlet.2019.11.001

158. Fujita N, Nishie A, Aishima S, Kubo Y, Asayama Y, Ishigami K, et al. Role of tumor-associated macrophages in the angiogenesis of well-differentiated hepatocellular carcinoma: pathological-radiological correlation. *Oncol Rep.* (2014) 31:2499–505. doi: 10.3892/or.2014.3138

159. Yuan H, Lin Z, Liu Y, Jiang Y, Liu K, Tu M, et al. Intrahepatic cholangiocarcinoma induced M2-polarized tumor-associated macrophages facilitate tumor growth and invasiveness. *Cancer Cell Int.* (2020) 20:586. doi: 10.1186/s12935-020-01687-w

160. Zhou Z, Wang P, Sun R, Li J, Hu Z, Xin H, et al. Tumor-associated neutrophils and macrophages interaction contributes to intrahepatic cholangiocarcinoma progression by activating STAT3. *J Immunother Cancer*. (2021) 9. doi: 10.1136/jitc-2020-001946

161. Vitale I, Manic G, Coussens LM, Kroemer G, Galluzzi L. Macrophages and metabolism in the tumor microenvironment. *Cell Metab.* (2019) 30:36–50. doi: 10.1016/j.cmet.2019.06.001

162. Jiang Y, Han Q, Zhao H, Zhang J. Promotion of epithelial-mesenchymal transformation by hepatocellular carcinoma-educated macrophages through Wnt2b/ β -catenin/c-Myc signaling and reprogramming glycolysis. J Exp Clin Cancer Res. (2021) 40:13. doi: 10.1186/s13046-020-01808-3

163. Azizi E, Carr AJ, Plitas G, Cornish AE, Konopacki C, Prabhakaran S, et al. Single-cell map of diverse immune phenotypes in the breast tumor microenvironment. *Cell.* (2018) 174:1293–308.e36. doi: 10.1016/j.cell.2018.05.060

164. Nalio Ramos R, Missolo-Koussou Y, Gerber-Ferder Y, Bromley CP, Bugatti M, Núñez NG, et al. Tissue-resident FOLR2(+) macrophages associate with CD8(+) T cell infiltration in human breast cancer. *Cell.* (2022) 185:1189–207.e25. doi: 10.1016/j.cell.2022.02.021

165. Hage C, Hoves S, Strauss L, Bissinger S, Prinz Y, Pöschinger T, et al. Sorafenib induces pyroptosis in macrophages and triggers natural killer cell-mediated cytotoxicity against hepatocellular carcinoma. *Hepatology*. (2019) 70:1280–97. doi: 10.1002/hep.30666

166. Zhu Y, Yang J, Xu D, Gao XM, Zhang Z, Hsu JL, et al. Disruption of tumourassociated macrophage trafficking by the osteopontin-induced colony-stimulating factor-1 signalling sensitises hepatocellular carcinoma to anti-PD-L1 blockade. *Gut.* (2019) 68:1653–66. doi: 10.1136/gutjnl-2019-318419

167. Klichinsky M, Ruella M, Shestova O, Lu XM, Best A, Zeeman M, et al. Human chimeric antigen receptor macrophages for cancer immunotherapy. *Nat Biotechnol.* (2020) 38:947–53. doi: 10.1038/s41587-020-0462-y

168. Mukhopadhyay M. Macrophages enter CAR immunotherapy. *Nat Methods*. (2020) 17:561. doi: 10.1038/s41592-020-0862-4

169. 30th annual meeting and associated programs of the society for immunotherapy of cancer (SITC 2015). *J Immunother Cancer.* (2015) 3 Suppl 2:O1-p453.

170. Wang S, Yang Y, Ma P, Zha Y, Zhang J, Lei A, et al. CAR-macrophage: An extensive immune enhancer to fight cancer. *EBioMedicine*. (2022) 76:103873. doi: 10.1016/j.ebiom.2022.103873

171. Kang M, Lee SH, Kwon M, Byun J, Kim D, Kim C, et al. Nanocomplexmediated *in vivo* programming to chimeric antigen receptor-M1 macrophages for cancer therapy. *Adv Mater.* (2021) 33:e2103258. doi: 10.1002/adma.202103258

172. Zhang W, Liu L, Su H, Liu Q, Shen J, Dai H, et al. Chimeric antigen receptor macrophage therapy for breast tumours mediated by targeting the tumour extracellular matrix. *Br J Cancer*. (2019) 121:837–45. doi: 10.1038/s41416-019-0578-3

173. Wei L, Wang Z, Jing N, Lu Y, Yang J, Xiao H, et al. Frontier progress of the combination of modern medicine and traditional Chinese medicine in the treatment of hepatocellular carcinoma. *Chin Med.* (2022) 17:90. doi: 10.1186/s13020-022-00645-0

174. Zhou Y, Wang Z, Ren S, Li W. Mechanism of action of protopanaxadiol ginsenosides on hepatocellular carcinoma and network pharmacological analysis. *Chin Herb Med.* (2024) 16:548–57. doi: 10.1016/j.chmed.2024.06.002

175. Jia H, Zhao B, Zhang F, Santhanam RK, Wang X, Lu J. Extraction, structural characterization, and anti-hepatocellular carcinoma activity of polysaccharides from Panax ginseng Meyer. *Front Oncol.* (2021) 11:785455. doi: 10.3389/fonc.2021.785455

176. He L, Xu K, Niu L, Lin L. Astragalus polysaccharide (APS) attenuated PD-L1mediated immunosuppression via the miR-133a-3p/MSN axis in HCC. *Pharm Biol.* (2022) 60:1710–20. doi: 10.1080/13880209.2022.2112963

177. Li L, Xu X, Wu L, Zhu H, He Z, Zhang B, et al. Scutellaria barbata polysaccharides inhibit tumor growth and affect the serum proteomic profiling of hepatoma H22-bearing mice. *Mol Med Rep.* (2019) 19:2254–62. doi: 10.3892/mmr.2019.9862

178. Li J, Wang T, Liu P, Yang F, Wang X, Zheng W, et al. Hesperetin ameliorates hepatic oxidative stress and inflammation via the PI3K/AKT-Nrf2-ARE pathway in

oleic acid-induced HepG2 cells and a rat model of high-fat diet-induced NAFLD. Food Funct. (2021) 12:3898–918. doi: 10.1039/d0fo02736g

179. Yao XM, Li Y, Li HW, Cheng XY, Lin AB, Qu JG. Bicyclol attenuates tetracyclineinduced fatty liver associated with inhibition of hepatic ER stress and apoptosis in mice. *Can J Physiol Pharmacol.* (2016) 94:1–8. doi: 10.1139/cjpp-2015-0074

180. Couri T, Pillai A. Goals and targets for personalized therapy for HCC. *Hepatol Int*. (2019) 13:125–37. doi: 10.1007/s12072-018-9919-1

181. Jenne CN, Kubes P. Immune surveillance by the liver. *Nat Immunol.* (2013) 14:996–1006. doi: 10.1038/ni.2691

182. Ringelhan M, Pfister D, O'connor T, Pikarsky E, Heikenwalder M. The immunology of hepatocellular carcinoma. *Nat Immunol.* (2018) 19:222-32. doi: 10.1038/s41590-018-0044-z

183. Ilyas SI, Wang J, El-Khoueiry AB. Liver cancer immunity. *Hepatology*. (2021) 73 Suppl 1:86–103. doi: 10.1002/hep.31416